





Sequencing and Analysis of the Entire Genome of the Mycoparasitic Bioeffector Fungus *Trichoderma asperelloides* Strain T 203 (Hypocreales)

Maggie Gortikov, a Zheng Wang, b Andrei S. Steindorff, c Igor V. Grigoriev, de Irina S. Druzhinina, e Jeffrey P. Townsend, b Oded Yarden

^aDepartment of Plant Pathology and Microbiology, The Robert H. Smith Faculty of Agriculture, Food and Environment, The Hebrew University of Jerusalem, Rehovot, Israel

^bDepartment of Biostatistics, Yale University, New Haven, Connecticut, USA

cU.S. Department of Energy Joint Genome Institute, Lawrence Berkeley National Laboratory, Berkeley, California, USA

^dDepartment of Plant and Microbial Biology, University of California, Berkeley, Berkeley, California, USA

eFungal Genomics Laboratory (FungiG), Nanjing Agricultural University, Nanjing, China

Department of Ecology and Evolutionary Biology, Yale University, New Haven, Connecticut, USA

ABSTRACT The filamentous mycoparasitic fungus *Trichoderma asperelloides* (Hypocreales, Ascomycota, Dikarya) strain T 203 was isolated from soil in Israel by the Ilan Chet group in the 1980s. As it has been the subject of laboratory, greenhouse, and field experiments and has been incorporated into commercial agricultural preparations, its genome has been sequenced and analyzed.

The mycoparasitic strain *Trichoderma asperelloides* T 203 (=TH 203), formerly identified as *Trichoderma asperellum* and prior to that *Trichoderma harzianum*, is a cryptic sister species to *T. asperellum* (1). Its fungal host range, host recognition traits, and mechanistic aspects of the mycoparasite interactions have been extensively studied (2–4). The strain has also been shown to confer transient repression of the plant immune response, followed by enhanced root colonization and eventual stimulation of plant growth and resistance to a wide range of adverse environmental conditions, including salt stress (5–8). The potential to use this strain as a biocontrol agent has been repeatedly examined under greenhouse and field conditions (9).

DNA was extracted from a freeze-dried, powdered culture of *T. asperelloides* T 203 (Fig. 1) grown in potato dextrose broth for 1 week, using a modified cetyltrimethylammonium bromide (CTAB)-based method, followed by chlorophorm:octanol extraction and isopropanol precipitation steps (10-12). The DNA yield and quality were assessed using the Synergy HTX multimode reader (Biotek, VT, USA) and verified by DNA electrophoresis. The sequencing library was built using the IDT Lotus DNA library prep kit (part number 10001074), full-length dual barcode adapters were ligated, and the quality of the fragments was checked using the high-sensitivity D5000 tapes (part number 5067-5592) on the Agilent TapeStation 4200 system and the KAPA library quantification kit (catalog number 07960298001), respectively. Genome sequencing was carried out on the Illumina NovaSeq S4 platform with 2×151 -bp reads. A total of 52 million raw reads were produced for the T. asperelloides samples. The raw reads (read length, 2 × 150 bp) were processed using Trim Galore v0.6.6 (https://github.com/FelixKrueger/ TrimGalore). An enriched set of mitochondrial reads was then extracted from the original input fastq reads by kmer matching using BBDuk in BBTools v38.44, using defaults, against the Trichoderma organelle contigs available at GenBank/NCBI. The matching reads were used to assemble the mitochondrial genome using SPAdes v3.15.2 (13). A similar methodology employing the UNITE ribosomal DNA (rDNA) database (14) was used

Editor Jason E. Stajich, University of California,

Copyright © 2022 Gortikov et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0

Address correspondence to Oded Yarden, oded.yarden@mail.huji.ac.il.

The authors declare no conflict of interest.

Received 12 December 2021 Accepted 31 January 2022 Published 17 February 2022 Gortikov et al.



FIG 1 Trichoderma asperelloides strain T 203 cultured for 1 week on potato dextrose agar (PDA) at 28° C.

to reassemble the rDNA from the filtered reads. Finally, an assembly of the target genome was generated using the resulting nonmitochondrial reads with SPAdes (13) using the following parameters: —phred-offset 33 —cov-cutoff auto –t 12 -m 32 —careful. The assembly size was 36,270,279 bp, with $100\times$ coverage. The assembly comprised 354 genomic scaffolds; the L_{50} value was 0.29 Mbp, the N_{50} value was 35 bp, and the GC content was 47.98%. The genome assembly was annotated using the JGl Annotation Pipeline (15), which combines several gene predictions and annotation methods with transcriptomics data and integrates the annotated genomes into MycoCosm (https://mycocosm.jgi.doe .gov), a Web-based fungal resource for comparative analysis (15). The completeness of the genome annotation was assessed using BUSCO v4.0.6 (16) using the hypocreales_odb10 database, resulting in 97.5% (single copy, 97.2%; duplicate, 0.3%) completeness.

The genome of *T. asperelloides* T 203 will contribute to the understanding of genome evolution within the genus *Trichoderma* as well as to the understanding of the comparative genome organization and diversity among strains of *T. asperelloides*, a species under continuous study for both fundamental science as well as commercial domestication.

Data availability. This whole-genome shotgun sequence of *T. asperelloides* T 203 has been deposited at DDBJ/ENA/GenBank under BioProject accession number PRJNA772304 with BioSample accession number JAJKFY000000000. The raw reads can be found under SRA accession number SRR17157108.

ACKNOWLEDGMENTS

This research was funded by grant number 2018712 from the Binational Israel—U.S. Science Foundation—U.S. National Science Foundation (O.Y., Z.W., and J.P.T.). The work conducted by the U.S. Department of Energy Joint Genome Institute, a DOE Office of Science User Facility, is supported by the Office of Science of the U.S. Department of Energy under contract number DE-AC02-05CH11231.

REFERENCES

- Samuels GJ, Ismaiel A, Bon M-C, De Respinis S, Petrini O. 2010. Trichoderma asperellum sensu lato consists of two cryptic species. Mycologia 102:944–966. https://doi.org/10.3852/09-243.
- Elad Y, Chet I, Boyle P, Henis Y. 1983. Parasitism of *Trichoderma* spp. on *Rhizocto-nia solani* and *Sclerotium rolfsii*-scanning electron microscopy and fluorescence microscopy. Phytopathology 73:85–88. https://doi.org/10.1094/Phyto-73-85.

Volume 11 Issue 2 e00995-21 mra.asm.org **2**



- Inbar J, Chet I. 1994. A newly isolated lectin from the plant pathogenic fungus Sclerotium rolfsii: purification, characterization and its role in mycoparasitism. Microbiology 140:651–657. https://doi.org/10.1099/00221287-140-3-651.
- Viterbo A, Montero M, Ramot O, Friesem D, Monte E, Llobell A, Chet I. 2002. Expression regulation of the endochitinase chit36 from *Trichoderma asperellum (T. harzianum* T-203). Curr Genet 42:114–122. https://doi.org/10.1007/s00294-002-0345-4.
- Shoresh M, Yedidia I, Chet I. 2005. Involvement of jasmonic acid/ethylene signaling pathway in the systemic resistance induced in cucumber by *Tri*choderma asperellum T203. Phytopathology 95:76–84. https://doi.org/10 .1094/PHYTO-95-0076.
- Viterbo A, Landau U, Kim S, Chernin L, Chet I. 2010. Characterization of ACC deaminase from the biocontrol and plant growth-promoting agent Trichoderma asperellum T203. FEMS Microbiol Lett 305:42–48. https://doi.org/10.1111/j.1574-6968.2010.01910.x.
- Gal-Hemed I, Atanasova L, Komon-Zelazowska M, Druzhinina IS, Viterbo A, Yarden O. 2011. Marine isolates of *Trichoderma* as potential halotolerant agents of biological control for arid-zone agriculture. Appl Environ Microbiol 77:5100–5109. https://doi.org/10.1128/AEM.00541-11.
- Brotman Y, Landau U, Cuadros-Inostroza Á, Tohge T, Takayuki T, Fernie AR, Chet
 I, Viterbo A, Willmitzer L. 2013. *Trichoderma*-plant root colonization: escaping
 early plant defense responses and activation of the antioxidant machinery for
 saline stress tolerance. PLoS Pathog 9:e1003221. https://doi.org/10.1371/journal
 .ppat.1003221.
- Chet I. 1987. Trichoderma—application, mode of action, and potential as biocontrol agent of soilborne plant pathogenic fungi, p 137–160. In Chet I (ed), Innovative approaches to plant disease control. John Wiley, New York, NY.

- Doyle JJ, Doyle JL. 1987. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. Phytochem Bull 19:11–15.
- Ziv C, Gorovits R, Yarden O. 2008. Carbon source affects PKA-dependent polarity of *Neurospora crassa* in a CRE-1-dependent and independent manner. Fungal Genet Biol 45:103–116. https://doi.org/10.1016/j.fgb.2007.05.005.
- 12. Carter-House D, Stajich JE, Unruh S, Kurbessoian T. 2020. Fungal CTAB DNA extraction. protocols.io https://doi.org/10.17504/protocols.io.bhx8j7rw.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- 14. Köljalg U, Nilsson RH, Abarenkov K, Tedersoo L, Taylor AFS, Bahram M, Bates ST, Bruns TD, Bengtsson-Palme J, Callaghan TM, Douglas B, Drenkhan T, Eberhardt U, Dueñas M, Grebenc T, Griffith GW, Hartmann M, Kirk PM, Kohout P, Larsson E, Lindahl BD, Lücking R, Martín MP, Matheny PB, Nguyen NH, Niskanen T, Oja J, Peay KG, Peintner U, Peterson M, Pöldmaa K, Saag L, Saar I, Schüßler A, Scott JA, Senés C, Smith ME, Suija A, Taylor DL, Telleria MT, Weiss M, Larsson K-H. 2013. Towards a unified paradigm for sequence-based identification of fungi. Mol Ecol 22:5271–5277. https://doi.org/10.1111/mec.12481.
- Grigoriev IV, Nikitin R, Haridas S, Kuo A, Ohm R, Otillar R, Riley R, Salamov A, Zhao X, Korzeniewski F, Smirnova T, Nordberg H, Dubchak I, Shabalov I. 2014. MycoCosm portal: gearing up for 1000 fungal genomes. Nucleic Acids Res 42:D699–D704. https://doi.org/10.1093/nar/gkt1183.
- Seppey M, Manni M, Zdobnov EM. 2019. BUSCO: assessing genome assembly and annotation completeness, p 227–245. In Methods in molecular biology. Humana Press, Inc., New York, NY.

Volume 11 Issue 2 e00995-21 mra.asm.org **3**