

HHS Public Access

Author manuscript Semin Cell Dev Biol. Author manuscript; available in PMC 2023 July 01.

Published in final edited form as:

Semin Cell Dev Biol. 2022 July ; 127: 121-132. doi:10.1016/j.semcdb.2021.08.006.

Multigenerational Epigenetic Inheritance: Transmitting information across generations

Nicholas O. Burton^{1,2,3}, Eric L. Greer^{4,5,6}

¹Centre for Trophoblast Research, Department of Physiology, Development and Neuroscience, University of Cambridge, Cambridge, CB2 3EG, UK

²Gurdon Institute, University of Cambridge, Cambridge, CB2 1QN, UK

³Present Address: Center for Epigenetics, Van Andel Institute, Grand Rapids, MI USA

⁴Division of Newborn Medicine, Boston Children's Hospital, 300 Longwood Avenue, Boston, MA 02115, USA

⁵Department of Pediatrics, Harvard Medical School, Boston MA 02115, USA

⁶Harvard Medical School Initiative for RNA Medicine, Boston MA 02115, USA

Abstract

Inherited epigenetic information has been observed to regulate a variety of complex organismal phenotypes across diverse taxa of life. This continually expanding body of literature suggests that epigenetic inheritance plays a significant, and potentially fundamental, role in inheritance. Despite the important role these types of effects play in biology, the molecular mediators of this non-genetic transmission of information are just now beginning to be deciphered. Here we provide an intellectual framework for interpreting these findings and how they can interact with each other. We also define the different types of mechanisms that have been found to mediate epigenetic inheritance and to regulate whether epigenetic information persists for one or many generations. Epigenetic inheritance is entering an exciting phase, where mechanistic understanding of how non-genetic information without permanently altering the genetic code. A more complete understanding of how and when epigenetic inheritance occurs will advance our understanding of numerous different aspects of biology ranging from how organisms cope with changing environments to human pathologies influenced by a parent's environment.

Declaration of interest: none

Correspondence should be addressed to Eric L. Greer (eric.greer@childrens.harvard.edu) or Nicholas O. Burton (nick.burton@vai.org).

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Declaration of interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Keywords

Transgenerational Epigenetic Inheritance; Intergenerational; DNA methylation; histone methylation; small RNAs; non-genetic; non-Mendellian; epigenetics

Introduction

Since the original observations of the inheritance of traits in pea plants by Gregor Mendel in 1865 [1], and their rediscovery in 1900 [2–4], a vast majority of all traits in eukaryotic organisms have been found to follow Mendelian inheritance patterns and are transmitted via DNA [5]. For almost equally long, but perhaps less appreciated, it has also been known that some traits do not follow Mendelian inheritance patterns and can be influenced by the environment or signaling pathways in previous generations. For example, original studies in 1909 by Mary Isabel McCracken [6], and in 1924 and 1925 by K. Watanabe [7] and Yositiro Umeya [8], respectively, reported that external temperature affected the ovaries of female silk moths (Bombyx mori) in a way that influenced whether her offspring would enter a hibernating diapause state. Later studies demonstrated that this heritable trait was controlled via a diapause hormone which is released from the mother's somatic cells (the suboesophageal ganglion) and transmitted via blood [9] to oocytes [10–12]. These studies of diapause in *B. mori* collectively described the first animal trait known to be controlled by somatic cell experiences in a previous generation. Of particular note, the experimental blood transfusions in *B. mori* by Yositiro Umeya [8] are conceptually identical to Francis Galton's early experiments on the inheritance of coat color in rabbits using blood transfusions [13]. Galton's experiment ultimately was influential in refuting Charles Darwin's theory of germules [14] and impacted the acceptance of August Weismann's "Weismann barrier" which suggested that information could not be transmitted from somatic cells to germ cells [15]. However, the conclusions reached by Umeya [8] and later confirmed by others [10-12]were the exact opposite of those reached by Galton and Weismann, and indicated that some traits were in fact controlled by factors present in the blood moving to germ cells.

The findings of McCracken, Watanabe, and Umeya never received the same level of scientific attention and interest as those of Galton and Weismann. Nonetheless, similar observations to those in *B. mori* of a parent's environment affecting traits in their offspring have since been observed in studies of diverse organisms ranging from nematodes to mammals [16]. These studies include numerous examples where a parent's exposure to particular environmental stresses can promote adaptive changes in offspring. These consistent observations of non-genetic inheritance across diverse evolutionary taxa suggest that such effects might represent a fundamental aspect of biology. Despite its potential importance in biology, non-genetic inheritance remains poorly understood and a better understanding of such effects and the mechanisms that mediate them could significantly advance our understanding of both biological and medical sciences.

To date, a growing body of literature has painted a complex picture of both the phenomena and mechanisms underlying non-Mendelian inheritance. Non-Mendelian effects have been reported to be transmitted anywhere from one to an indefinite number of generations; they

sometimes, but not always, occur in response to environmental stimuli; and they play an as-yet-unknown role in biology with estimates ranging from rare occurrences observed in organisms with short generation times [17] to pervasive effects that are potentially found in all eukaryotic species [18]. Because the molecular mechanisms underlying non-Mendelian inheritance are just beginning to be deciphered, a non-overlapping patchwork of mechanisms have been proposed even within a single species.

Here, we review the current state of research into multigenerational effects, encompassing both intergenerational effects (lasting 1–2 generations) and transgenerational effects (lasting 3+ generations), with a particular focus on the molecular mechanisms that mediate the transmission of such effects via germ cells. In addition, we highlight how different evolutionary pressures are likely to favor the evolution of intergenerational or transgenerational effects and how such pressures might explain why organisms may have evolved separate mechanisms to mediate each type of effect. We propose that this type of thinking and categorization of multigenerational effects might help avoid some of the problems that have led to significant confusion in the field, such as the term transgenerational having a different definition in studies performed in different species. Furthermore, this categorization will allow us to better estimate how common different types of effects might be, what molecular mechanisms might be most likely to underlie newly observed examples of non-Mendelian inheritance, and if the molecular mechanisms underlying one multigenerational effect are likely to mediate similar multigenerational effects in other organisms.

What are intergenerational and transgenerational effects?

In mammals, transgenerational effects, particularly those that occur in response to the environment, are defined as any phenotypic or molecular effect that persists for 3 or more generations through the female line or 2 or more generations through the male line [19]. By contrast, effects that only persist for 1 or 2 generations are for the most part referred to as intergenerational effects [19]. Whether a phenotype is intergenerational or transgenerational was originally determined by whether the genetic material for the subsequent generations was present at the time of exposure to the altered environment. This often differs between different species, so caution must be used to identify whether the germ cells were present at the time of exposure. The original distinction between these two terms lay in the fact that intergenerational effects could, in principle, be caused by the effects of the parent's environment/physiology directly on the developing embryo/ fetus or on germ cells but transgenerational effects could not be due to direct exposure. However, mechanistic investigations of multiple different intergenerational effects have since discovered mechanisms of intergenerational regulation that are not due to the direct effects of the environment on germ cells or F1 embryos [12, 20–23]. In some cases, these mechanisms are initiated and maintained using similar mechanisms as transgenerational effects such as the transmission of small RNA molecules via germ cells [24, 25]. Nonetheless these effects remain described as intergenerational effects. Thus, the currently used definition of intergenerational has evolved to refer mainly to the duration a phenotypic effect persists for rather than the potential mechanism by which the effect is mediated (Figure 1). By comparison, for a phenotype to be considered transgenerational, none of the

individuals genetic material can be present at the time of the environmental insult (Figure 1). Thus, transgenerational effects predominantly refer to phenotypic effects that persist for three or more generations. For the purposes of this review, and for comparing effects across species, we will define all effects that only persist for a single generation (parent-offspring effects) as intergenerational regardless of species or timing of germ cell development. We will also define all effects that persist three or more generations as transgenerational. Observations of multigenerational effects lasting for specifically two generations will be discussed on a case-by-case basis with considerations for species and line of transmission.

Why do organisms maintain non-genetic information across generations?

The question of why organisms transmit non-genetic information across generations has been asked since Alexander Brink first reported transgenerational epigenetic inheritance in maize in 1956 [26]. If we were to anthropomorphize evolution, epigenetic inheritance exists to transmit environmental information to the descendants without modifying the heritable material of DNA and therefore permit future generations to revert back to a "normal" state when the environmental insult has passed. In addition to the question of why information is maintained at all, there also remain significant questions related to why organisms maintain certain types of information for differing numbers of generations. A better understanding of these outstanding questions in the field is likely to significantly enhance our ability to compare different multigenerational effects across species. In an effort to advance this understanding, here we break known multigenerational effects into three distinct types of responses based on the duration of their inheritance – intergenerational effects, transient transgenerational effects, and permanent transgenerational effects.

Intergenerational effects

Intergenerational effects refer, most commonly, to the effects of a parent's environment on their offspring and can be both adaptive and deleterious from the perspective of the offspring. For the purposes of this review, we focus on adaptive intergenerational effects, as these are the most likely to be mediated by active mechanisms that evolved to mediate a heritable biological function rather than the toxic side effects of various biological or chemical insults.

Adaptive intergenerational effects are commonly observed throughout evolution and can lead to substantial and sometimes dramatic changes in organism development and physiology. For example, in the pea aphid *Acyrthosiphon pisum*, parental exposure to certain stresses can lead to the development of wings in offspring in addition to many additional behavioral and physiological changes [27–29]. The development of wings allows offspring to fly away from stressful conditions, but comes at the expense of fecundity [28]. Such trade-offs are commonly observed for intergenerational adaptive effects and likely explain why these forms of plasticity are only observed in response to specific environmental stimuli and are lost or erased within one generational adaptations to stress, including observations of trade-offs, have been observed in diverse taxa ranging from plants to mammals including *Arabidopsis* responses to pathogen infection [30], *C. elegans* responses to osmotic stress,

social environment, and multiple types of pathogen infections [22, 23, 31–33], *Daphnia* helmet formation in response to predators [34], the overwintering response of *Bombyx mori* [11], and the response of red squirrels to food and territory availability [35] among many other examples. Furthermore, intergenerational adaptive changes in response to nutrient stress, and their deleterious tradeoffs, have been hypothesized to underlie observations of fetal programming in humans and possibly contribute to multiple human metabolic pathologies including Type 2 diabetes [36].

Studies of the tradeoffs of intergenerational adaptations and mathematical modelling of intergenerational adaptive effects suggest that intergenerational adaptive effects are likely to evolve in any scenario where (1) the environment is variable, (2) a parent's environment is predictive of their offspring's future environment, (3) the benefits of the adaptation outweigh the costs [37]. These conditions are likely common for most organisms in response to one or more stresses, and thus intergenerational adaptive effects might similarly be common. In addition, despite the potential benefits of intergenerational adaptations to stress, the costs of these adaptations might promote their loss or active erasure when the environment changes, potentially explaining why most intergenerational adaptations do not persist for more than one generation [38].

Transgenerational effects

Like intergenerational effects, transgenerational effects can also be both adaptive and deleterious. However, unlike intergenerational effects, both adaptive and deleterious transgenerational effects are likely to be mediated by epigenetic mechanisms, as transgenerational effects cannot usually be due to the direct exposure of an organism to any particular stress or condition. Furthermore, experimental studies of transgenerational effects that there might be at least two different types of transgenerational effects that may or may not be mechanistically related. Here we describe these two types of transgenerational effects as permanent transgenerational effects that persist indefinitely and transient transgenerational effects that occur in response to a specific set of conditions but are ultimately lost after a varying number of generations.

Permanent transgenerational effects

Some of the original observations of transgenerational epigenetic inheritance come from studies of paramutation in maize [26]. Paramutation-like phenomena have since been described in several species and in each case represent the transmission of epigenetic information that can be maintained for an indefinite number of generations [39–42]. Studies of the molecular mechanisms underlying permanent transgenerational effects have almost exclusively identified epigenetic mechanisms that regulate the silencing of transposons, repetitive elements, and foreign DNA, and most known examples of paramutation involve the silencing of a transposon or repetitive element that indirectly affects the expression of a nearby gene [39–42]. Permanent transgenerational effects are likely to evolve in response to conditions that do not change or when a stress is permanently present by virtue of being integrated into the host genome, such as the presence of a transposon. These types of effects are unlikely to occur in response to environmental stimuli that can change between generations.

Transient transgenerational effects

In contrast to permanent transgenerational effects, transient transgenerational effects have been reported to persist for anywhere from three to approximately ten generations and are often reported in response to environmental stimuli or stresses [43–65]. Initial studies of the mechanisms underlying transient transgenerational effects that occur in response to the environment have identified similar mechanisms to those that regulate permanent transgenerational effects, such as small RNAs [41, 43, 44, 46, 47, 52, 58, 60, 66], DNA methylation [53], and histone modifications [59, 60]. The mechanisms that prevent transient transgenerational effects from persisting permanently in most organisms remain unclear, but recent work in C. elegans suggests that H3K9 methylation and the chromodomain protein HERI-1 antagonize the transgenerational inheritance of small RNA silencing and in the absence of either the putative histone methyltransferase MET-2 or HERI-1, animals can inherit a normally transient transgenerational effect permanently [51, 67]. Similarly, transgenerational inheritance of elevated H3K4me2 is also antagonized by regulators of H3K9 methylation [63, 68, 69]. Collectively, these results suggest that H3K9 methylation might play a role in preventing transient transgenerational epigenetic inheritance from becoming permanent silencing. It will be important, in future studies, to deduce whether there are common or unique mechanisms that set the transgenerational clock and prevent epigenetic effects from persisting indefinitely.

Unlike intergenerational responses to environmental stimuli, modelling of transient transgenerational responses to environmental stimuli has found that they are more likely to evolve when a parent's environment is not predictive of their offspring's environment [70]. While in most cases a parent's environment is highly likely to be predictive of their offspring's future environment, there are certain cases where the opposite is true. For example, in certain cyclical environments such as the changing of seasons, and in organisms with short generation times, information about a great-grandparent's environment could be more predictive of an individual's future environment, such as the coming of winter, than their parent's. Alternatively, in highly variable environments it could be more useful to average the environments of multiple previous generations rather than use the input of parental environment alone. Consistent with this hypothesis, several studies have demonstrated that the exposure of multiple successive generations to these pathogens [31, 55].

These differences in the pressures favoring intergenerational vs transient transgenerational effects are predominantly due to the costs of maintaining information about the environment across many generations and the potential costs of adapting to one particular stress which can come at the expense of being able to adapt to other stresses [32]. Transient transgenerational effects, particularly adaptive ones, need to avoid costs that occur in mismatched environments that do not match the stress that triggered the transgenerational effect, or have benefits to the organism that outweigh any potential costs. These differences in the pressures favoring transient transgenerational effects, when compared to intergenerational effects, raises the possibility that different molecular mechanisms might evolve to mediate these two types of multigenerational effects.

Consistent with the hypothesis that different evolutionary pressures favor intergenerational vs transgenerational effects, some organisms have evolved parallel mechanisms to elicit separate intergenerational or transgenerational responses to the same stimuli. For example, the nematode Caenorhabditis elegans can elicit both intergenerational and transgenerational responses to the presence of dsRNA [24, 25, 43, 44, 66, 71]. To accomplish these two separate types of inheritance, different mechanisms have evolved that are spatially separated in different tissues to mediate intergenerational vs transgenerational dsRNA-directed gene silencing. Specifically, the argonaute NRDE-3 mediates intergenerational gene silencing in somatic tissues for somatically expressed genes [25] while the argonaute HRDE-1 mediates transgenerational gene silencing in germ cells for genes expressed in germ cells [43]. The observations that these two argonautes function in different tissues to carry out intergenerational silencing in somatic cells and transgenerational silencing in germ cells in response to the same dsRNA stimuli suggests that C. elegans has evolved separate argonautes to mediate intergenerational vs transgenerational inheritance and that the tissues in which intergenerational and transgenerational silencing occur in might also be separate. It will be interesting, in future studies, to determine how specific small RNAs are selected to transmit information intergenerationally or transgenerationally. Is the inheritance of small RNAs simply dictated by when and where the small RNAs are expressed or are the specific small RNAs which are destined to be inherited marked by specific chemical modifications [72] or potentially sequestered into special subcellular compartments to be transmitted to descendants [73, 74]? Identifying these mechanistic determinants of small RNA inheritance will help to distinguish why the phenotypic consequences of some small RNAs are not inherited at all, some are intergenerationally inherited, and some are transgenerationally inherited.

Comparing the mechanisms that mediate multigenerational effects

Analyses of the mechanisms underlying multigenerational effects have often grouped intergenerational and transgenerational effects together. This is in part because similar general mechanisms can mediate both types of multigenerational effect, such as small RNAs and histone modifications. If, however, different evolutionary pressures favor the evolution of different types of multigenerational effects then further insight might be gained by comparing the mechanisms underlying these phenomena separately. Here we consider the mechanisms underlying intergenerational and transgenerational effects separately. We also examine the gaps that remain in our mechanistic understanding of these types of effects and to what extent similar mechanisms might underlie the many observations of intergenerational or transgenerational effects that have been observed across different species.

Mechanisms underlying intergenerational effects

Hormone signaling to oocytes

As was observed in *B. mori*, hormone signaling to oocytes has also been shown to play an important role in *C. elegans* response to osmotic stress. Specifically, parental exposure of animals to mild osmotic stress protects offspring from future exposure to osmotic stress [22]. This intergenerational adaptation is regulated by insulin-like signaling to

oocytes, with reduced signaling resulting in an increase in the expression of the glycerol-3phosphate dehydrogenase GPDH-2 in offspring [22]. GPDH-2 subsequently increases glycerol production which, in turn, promotes resistance to osmotic stress [22]. It will be interesting, in future studies, to elucidate the mechanisms by which insulin-like signaling to oocytes causes gene expression and metabolic changes. Nonetheless, the strong evolutionary conservation of insulin signaling throughout metazoans and recent observations in mammals demonstrating that maternal dietary stress can affect oocytes in a way that modifies insulin sensitivity and metabolism in offspring [75] suggest that intergenerational effects in other organisms might also be regulated by insulin signaling to oocytes. Identifying such mechanisms will be critical to our future understanding of how hormone signaling to oocytes can affect offspring, how common such effects might be, and if such effects play a role in human pathologies linked to maternal environment such as Type 2 diabetes.

Small RNA-based intergenerational mechanisms

RNAi silencing of a subset of genes can be passed from parents to offspring via germ cells in *C. elegans* [24, 25, 43, 44, 66, 71]. For almost all such genes that are expressed in somatic cells this silencing lasts for only a single generation [25, 71], suggesting that RNAi silencing in somatic tissues is inherited intergenerationally in *C. elegans*. Genetic studies of the factors that regulate the intergenerational inheritance of RNAi silencing found that some genes were required for the initiation of RNAi silencing, such as the RNA helicase RDE-1 and the RNA binding protein RDE-4 [24], while others were required for the maintenance of RNAi silencing, such as the 3'–5' exoribonuclase MUT-7 and the MUT-7 interacting protein RDE-2 [24, 76]. Later studies found that the nuclear RNAi pathway mediates the inheritance of RNAi silencing of genes expressed in somatic cells [25]. Collectively, these studies demonstrated that a dedicated NRDE-3 dependent mechanism has evolved to mediate the intergenerational inheritance of RNAi silencing in somatic cells [25].

In addition to RNAi inheritance, various tRNA fragments and miRNAs have also been reported to mediate the transmission of information about the environment from parents to offspring via germ cells in mammals [20, 21, 77–84]. The abundance of many of these RNAs, in particular tRNA fragments, have been shown to be responsive to different environmental conditions [20, 21, 83, 85]. Furthermore, injection of total sperm RNA or purified RNA fragments into either oocytes or zygotes recapitulates some of the effects of a father's environment on offspring [20, 21, 42, 82–84]. These findings provide strong evidence that such RNA populations mediate some of the effects of a parent's environment on offspring. However, the mechanisms by which these RNAs exert their effects on long-term offspring health and physiology remain unclear.

Histone modifications and DNA methylation-based intergenerational mechanisms

Early forays into *in vitro* fertilization in mice revealed that having two sets of only maternal or only paternal chromosomes were not viable [86–88], suggesting that each sex had to contribute one set of chromosomes for viability and that the chromosomes coming from different sexes were not identical. Subsequent discoveries of maternally (*Igf2*) and

paternally (*H19* and *Igf2t*) imprinted and C5-cytosine methylated (5mC) genes [89–95], and demonstration that the C5-cytosine methyltransferase DNMT1 was required for maintenance of imprinted gene expression [96] found that DNA cytosine methylation could mark, and epigenetically silence, gene expression of critical genes inherited from parents. The presence of 5mC in monoallelic imprinting control regions allows for consistent expression of critical genes from a single parental allele. The precise method by which DNA cytosine methylation at critical imprinting control regions, in a parent-of-origin specific manner, escapes the canonical epigenetic erasure that occurs upon fertilization is still being deciphered [97–99]. DNA cytosine methylation has also been demonstrated to regulate imprinting in *Arabidopsis* [100]

The demonstration that methylated histones can be retained through cell divisions [101] and that the polycomb complex, a complex responsible for methylating histone H3 on lysine 27 (H3K27) [102–105], can be retained on the chromatin through DNA replication [106, 107], has suggested potential mechanisms by which histone methylation could escape erasure or facilitate the immediate reapplication of histone modifications through cell divisions and potentially across generations. Similarly, mutational analyses combined with fluorescent labelling experiments in Drosophila, have demonstrated an asymmetric inheritance of histones in the germline [108, 109] which could provide mechanistic support for how histone modifications could transmit through cell divisions. Several recent reports have also demonstrated a DNA methylation independent imprinting which is driven by inheritance of maternal histone H3 lysine 27 trimethylation (H3K27me3) and brief histone H2A lysine 119 ubiquitination (H2AK119ub1) [110–115]. Interestingly, some elegant work in C. elegans and Drosophila has further supported the inheritance of H3K27me3 across generations [116, 117], suggesting that inherited histone methylation could be a conserved process of transmitting non-genetic information to descendants. This does not appear to be restricted to H3K27me3 as fluorescent labeling experiments have also demonstrated the inheritance of H3K36me3 from parents to their children in C. elegans [118]. While paternal inheritance of histones is rarer in mammals, due to the replacement of histones with the more compactable protamines, in *C. elegans* histones are retained in paternal sperm and have been suggested to retain high levels of H3K27me3 and H3K36me3 and lower levels of H3K4me3 [119]. This paternal inheritance of modified histories correlates with repression of a neuronal fate in early germ cells [120]. Whether modified histones or DNA themselves are the transmitted material or something else is transmitted to cause the altered reacquisition of histone methylation/acetylation remains to be determined.

Interestingly, there are examples where genetic deletion of the enzymes necessary for these DNA and histone modifications eliminate certain intergenerational phenotypes arising from environmental stresses raising the possibility that these altered histone modifications themselves are transmitted from parents to their children. The naïve progeny of *Arabidopsis* which are exposed to the pathogenic bacteria *Pseudomonas syringae* have reduced DNA methylation [121], decreased seed production, and increased resistance in response to alternative pathogenic bacteria [30]. Elimination of DNA cytosine methyltransferases *drm1*, *drm2*, and *cmt3* eliminates the inherited resistance to the alternative pathogenic bacteria [30]. Similarly, naïve progeny from *Arabidopsis* ancestors which were repeatedly exposed to elevated salinity display altered DNA methylation and increased resistance to hyperosmotic

stress. This increased resistance to elevated salt is dependent on the presence of both DNA demethylases and DNA methyltransferases [122], suggesting that intergenerational inheritance is dependent on inheriting the appropriate levels of DNA methylation.

Correlations of changes in DNA and histone modifications in response to parental diet span the evolutionary tree [123–127]. For instance, naïve children of Drosophila fathers fed a high-sugar diet display inherited obesity correlated with elevated H3K9me3 and H3K27me3 [127]. These heritable obesity phenotypes are dependent on the presence of the H3K9 and H3K27 methyltransferase machinery [127]. Similarly, Drosophila fed a high-fat diet have reduced cardiac function correlated with elevated H3K27me3 which is transmitted to naïve descendants [128]. Overexpression of the H3K27me3 demethylase dUTX or chemical inhibition of the H3K27 methyltransferase EzH2 protect against the heritable heart defects [128]. The importance of H3K27 methylation in intergenerational inheritance is further highlighted by experiments in genetically wildtype *M. musculus* descendants of the H3K27me3 demethylase Utx which display increased DNA methylation and increased susceptibility to cancer relative to wildtype descendants from wildtype parents [129]. Chromatin modifications can be elicited in different manners. Exposure of Drosophila to heat stress can activate the transcription factor dATF-2 which binds to the H3K9me3 binding protein HP1 [130]. Heat stress causes dATF-2 phosphorylation and disruption of heterochromatin which can be epigenetically inherited [130]. Interestingly, repeated generational exposures to elevated temperature dulled the epigenetic phenotype suggesting that repeated exposures to the same stimuli can alter transgenerational responses to stress. A similar observation of multigenerational adaptation to repeated exposures was detected from exposure of C. elegans to pathogens [55]. Together these data raise the exciting possibility that modified histones and DNA can transmit environmental information to their naïve descendants to regulate intergenerational phenotypes.

Maternal provisioning

There are many known mechanisms by which changes in maternal provisioning of resources to offspring result in changes in offspring phenotype. This includes many examples of adaptive effects such as the transfer of antibodies from mother's to offspring in mammals [131], changes in yolk provisioning affect several offspring phenotypes in *C. elegans* including starvation resistance [132, 133], and changes the deposition of hormones into eggs in birds [134]. In many cases, these adaptive intergenerational provisioning effects achieve the same biological goals as intergenerational effects that are transmitted via germ cells. A comprehensive discussion of all of the possible intergenerational effects mediated by changes in provisioning is outside the scope of this review, but we note that these types of effects likely play as large of a role in biology as effects transmitted via germ cells. In addition, and similar to intergenerational effects mediated via germ cells, the molecular mechanisms by which altered maternal provisioning can affect the long-term health and physiology of offspring remain poorly understood and future studies of these effects are likely to substantially advance our understanding of diverse aspects of biology.

Notable examples of intergenerational effects that occur via unknown mechanisms

An especially notable example of intergenerational effects in humans is the observation of many different changes in metabolism and physiology in children conceived during the Dutch Hunger Winter [135, 136] and the Great Chinese Famine [137, 138]. These observations fit broadly within the emerging field of fetal programming - studies of how the *in utero* environment affects long-term health and physiology [36]. Fetal programming has been linked to several major pathologies including Type 2 diabetes and cardiovascular disease and stressful environments appear to cause similar changes in offspring physiology in diverse species of mammals [139–144]. The mechanisms by which fetal programming occur are poorly understood but are likely related to the mechanisms that regulate intergenerational effects in other organisms. In support of this hypothesis, recent studies of a model of fetal programming in mice found that the effects of a mother's high-fat diet on offspring obesity and insulin resistance can be transmitted via oocytes [75]. Fetal programming involving both fetal adaptations to stressful maternal or in utero environments comes at the expense of long term health [36]. These observations resemble the tradeoffs observed in intergenerational adaptive effects observed in other non-mammalian organisms. Future studies of the mechanisms underlying fetal programming will be critical for our understanding of the role these intergenerational effects play in human disease and if they are evolutionarily related to intergenerational effects observed in non-mammalian systems.

Mechanisms underlying transgenerational effects

Small RNA-based mechanisms

Observations of transgenerational epigenetic inheritance that are mediated by small RNAs have been reported in multiple different taxa including yeast, plants, and nematodes. In general, many observations of small RNA-based transgenerational epigenetic inheritance function to silence repetitive elements, transposons, and foreign DNA present in the genome (reviewed in [145–148]). For example, observations of paramutation in maize were one of the first known examples of transgenerational epigenetic inheritance [26], and it was later observed that paramutation in maize occurs due to the silencing of a tandem repeat sequences present in certain isolates of this species via a mechanism that requires small RNA biogenesis via RNA-dependent RNA polymerases [149–151]. Observations of paramutation have been reported in animals and were also found to silence transposable elements via a mechanism that depends on PIWI-interacting RNAs (piRNAs) [40]. These RNA-based silencing of repetitive and foreign elements largely represent permanent transgenerational effects.

Multiple studies have also reported transient transgenerational epigenetic effects in response to environmental stress that require small RNA-dependent machinery [45–50, 52, 55, 58, 60, 152]. These transient transgenerational effects have predominantly been reported in *C. elegans* and appear to converge on the PIWI-like argonaute PRG-1 [45, 58, 60], which is a core regulator of piRNA function in *C. elegans*, and/or the germline nuclear-localized argonaute protein HRDE-1 [47, 52, 152, 153] which functions to silence the expression of various retrotransposons, cryptic loci, foreign DNA and certain coding genes specifically in germ cells [43, 44, 66, 154, 155]. However, several other argonaute proteins have also

been proposed to regulate transgenerational effects in *C. elegans* including CSR-1 [66], PPW-1 [156, 157], WAGO-1 [66], and WAGO-4 [158]. These effects often also require the production of small RNAs by the RNA-dependent RNA polymerase RRF-1 [159].

Despite piRNA and endo-siRNA mediated silencing by PRG-1 and HRDE-1 frequent association with permanent transgenerational epigenetic silencing, recent studies suggest that PRG-1 and HRDE-1 might also mediate transient transgenerational silencing under certain conditions. For example, exogenous uptake of dsRNA [43, 160], transgenic expression of the Flock House Virus [46], starvation [45, 47], heat stress [152], and bacterial infection [58, 60] have all been reported to transgenerationally alter animal gene expression or physiology via PRG-1 and/or HRDE-1 dependent mechanisms. These transgenerational effects often persist for between 3 and 5 generations after which they reset via a mechanism dependent on the H3K9me1/2 methyltransferase MET-2 [51] and the chromodomain containing protein HERI-1 [67]. These findings suggest that MET-2 and HERI-1 might be dedicated molecular factors that separate the targets of PRG-1 and HRDE-1 into permanently silenced and transiently silenced targets.

Despite a growing body of literature indicating that small RNAs play an important role in regulating transient transgenerational epigenetic effects in *C. elegans* and yeast, there remain several outstanding questions about the mechanisms underlying these observations. For example, how certain environmental stresses initiate transgenerational epigenetic effects via small RNAs are poorly understood. In cases involving bacterial pathogens, *C. elegans* ingest dsRNAs produced by bacteria that in turn guide the silencing of *C. elegans* genes involved in a pathogen response [58]. However, the molecular mechanisms by which stresses such as heat or starvation might elicit similar small RNA-dependent changes is still unclear. Similarly, the extent to which these transgenerational epigenetic effects represent stress-specific responses or a general stress response and the extent to which similar small RNA-dependent transgenerational effects might occur in organisms lacking RNA-dependent RNA polymerases, such as humans, remains unknown. Future studies of these effects will be critical in elucidating the role small RNAs play in transgenerational effects more broadly.

Histone modification-based mechanisms

The realization that insufficient folate in the diet of pregnant women correlated with increased anemia, placental abruption, neural tube defects, and abortions [161] led to trials to supplement folate which substantially prevented neural tube defects [162, 163]. The success of these trials has led many countries to enrich grain products with folic acid, which has reduced the incidence of neural tube defects by 19–55% [164]. Folates' importance stems from its role in the one-carbon pathway where it is required to synthesize methionine which can be subsequently converted to S-adenosylmethionine which donates methyl groups to DNA, RNA, lipids, and proteins [165]. The importance of this pathway not only in intrauterine health for the offspring, but transgenerational viability was demonstrated by a hypomorphic mutation of a folate metabolism gene, *Mttr*, in mice. Loss of *Mttr* in ancestors was demonstrated to cause intrauterine growth retardation, developmental delay, and congenital malformations of the neural tube, heart and placenta for four generations of wild type descendants [166]. These experiments raised the possibility that insufficient

methylation of a variety of different substrates could have transgenerational consequences on the health of descendants.

A number of studies have pointed toward histone methylation playing an essential role in transgenerational epigenetic inheritance. Genetic mutation of enzymes that regulate H3K4 methylation have been revealed to elicit transgenerational epigenetic phenotypes independent of the mutation. For instance, mutation of an H3K4 trimethylation complex in *C. elegans* causes a ~20–30% extension in lifespan [167], which is transmitted for three generations to genetically wild-type descendants [64]. This H3K4 trimethylation complex regulates other transgenerational phenotypes in *C. elegans*, mutation of this complex causes a progressive sterility and transgenerational misregulation of genes [168– 170]. Additionally knock-out of the members of the H3K4 methyltransferase complex in *D. discoideum* eliminate the inheritance of active transcriptional states [171]. Conversely, knockout of the H3K4me1/2 demethylase *spr-5* in *C. elegans* causes a transgenerational progressive decline in fertility [172] and a transgenerational extension in lifespan [173]. In mammals, overexpression of the human H3K4me1/me2 demethylase, LSD1, in mice results in heritable effects on development and survival [174]. Collectively, these studies point to the importance of H3K4 methylation for regulating transgenerational epigenetic inheritance.

In addition to H3K4 methylation, several other histone methylation marks have been linked to transgenerational effects in multiple different species. For example, in C. elegans a heat stress can elicit transgenerational effects on gene expression [175], and this effect is associated with decreased heritable levels of H3K9me3 [59]. Interestingly, deletion of the putative H3K9 methyltransferase, SET-25, eliminates the transgenerational change in gene expression of a heat shock inducible transgenerational reporter [59]. H3K27me3 has also been implicated in transgenerational inheritance in Drosophila. A series of studies have created transgenic lines linking a region where both H3K4 and H3K27 methyltransferases bind to lacZ reporters and drivers of eye color [176]. Using this reporter line the authors have demonstrated that transient activation of this reporter can be transgenerationally inherited corresponding with heritably altered H3K4me3, H3K27me3, and H3 and H4 acetylation levels that are dependent on the H3K27 methyltransferase machinery [176-178]. These heritable histone modifications are not restricted to methylation. Due to the prevalence of preacetylated histones in the pool of free histones, histone acetylation is a prime candidate for transmitting non-genetic information. Indeed, increases in histone acetylation correlate with an intergenerational adaptation to maternal sensing of pheromones in A. friebergensis [179]. Therefore, both traditionally activating histone methylation marks, such as H3K4me and histone acetylation, and traditionally repressive histone methylation marks, including H3K9me3 and H3K27me3, have been correlated with changes in heritable epigenetic information.

Complicating the mechanistic understanding of the consequence of manipulating histone modifying enzymes is the fact that none of these epigenetic cues exist in isolation, they all communicate with each other to reinforce an epigenetic signature. Therefore identifying what the heritable cues are and what is the cause of each phenotype is complicated. For instance, elimination of the H3K4me1/me2 demethylase *spr-5* in *C. elegans* causes a transgenerational accumulation of H3K4me2 but a transgenerational decline in H3K9me3

and increase in H3K36me3 [63], and manipulation of the H3K9me and H3K36me regulatory machinery can eliminate or exacerbate the transgenerational phenotypes of *spr-5* mutant worms [63, 68, 180]. Similarly the transgenerational effects on lifespan of the H3K4 trimethyltransferase complex can be eliminated by genetic manipulation of the H3K9me2 methylation machinery [69]. In mice, overexpression of the H3K4me1/me2 demethylase LSD1 causes an increase in H3K4me3 in the first generation descendants [181]. These examples highlight how difficult it is to distinguish causal non-genetic cues transmitted across generations from those which are subsequently altered in response to the initial change.

DNA methylation-based mechanisms

While DNA methylation can be inherited through cell divisions [182] and is essential for maintenance of imprinting, as discussed above, it's role in transgenerational epigenetic inheritance is less well delineated in animals. In mammals, under basal conditions, while the majority of DNA cytosine methylation is erased between embryonic day 8.5 and 13.5 [183, 184], intracisternal A particle retrotransposons (IAPs) retain 5mC [185, 186]. Interestingly an IAP inserted upstream of the agouti gene to produce viable yellow (A^{vy}) mutations result in yellow coat color, increased tumor incidence, and adult onset obesity [187] which can be regulated and inherited in an epigenetic manner through the maternal line retention of 5mC [188]. Another IAP retrotransposon inserted upstream of axin-fused (Axin^{fu}) that is regulated by DNA cytosine methylation causes kinked tails in mice, has been shown to transmit the 5mC state and kinked tail epigenetically through both the paternal and maternal lineages [189]. While these examples of DNA methylation regulating transgenerational epigenetic phenomena in mammals are relatively rare under basal conditions [190], it still remains to be determined whether extreme environmental manipulations, which can elicit transgenerational epigenetic inheritance phenotypes, and which have been shown to alter epigenetic alleles [191–194], can cause DNA methylation to be retained transgenerationally in regions other than IAPs.

While cytosine methylation is relatively prevalent in metazoans DNA N6-adenine methylation (6mA) [195, 196] is an epigenetic modified base which is much more prevalent in bacteria and protists [197]. A number of recent reports have suggested that this rarer DNA methylation occurs in metazoans [198–204]. However, a number of groups have had difficulty detecting this rare modification in metazoans [205–208]. This discrepancy could be due to the relative scarcity of 6mA in metazoans, the sensitivity of detection methods, and that its importance might only be revealed under specific circumstances [207, 209]. Interestingly, 6mA has been shown to increase in response to specific stresses and tracks with transgenerational epigenetic inheritance [53, 198, 210]. Whether 6mA exists and plays an important role in transmitting non-genetic information across generations will have to be further tested through site specific directed methylation and demethylation of adenines.

Contrary to the relatively rare DNA cytosine methylation retention in animals, DNA methylation retention in plants to regulate transgenerational epigenetic silencing of transposable elements is relatively prevalent [211–218]. The prevalence of DNA methylation transgenerational epigenetic inheritance in plants is presumably due to the limited DNA

methylation reprograming that occurs [219]. Therefore, different organisms might rely more heavily on one epigenetic mode over another depending on which epigenetic cues are more prevalent or less tightly erased upon fertilization. For example small RNAs, which are essential for the basic immune response in *C. elegans* [220, 221], and therefore have an outsized importance, have been repeatedly identified in regulating transgenerational epigenetic inheritance in *C. elegans* while, DNA methylation, which is much more prevalent in plants and is not actively erased upon fertilization, plays a consistent role in regulating epigenetic inheritance in plants.

Crosstalk of epigenetic cues

As we mentioned earlier, chromatin modifications do not exist in isolation and oftentimes communicate with each other to reinforce non-genetic signatures. This modification cross talk is also prevalent in the communication between different epigenetic inheritance substrates. For instance, the RNAi machinery has been demonstrated to physically interact with histone modifying enzymes and binding proteins to help reinforce a repressed chromatin state across virtually all species tested [155, 222–230]. The RNAi machinery is required for the establishment and/or maintenance of heterochromatin characterized by H3K9 methylation in *S. pombe* [223, 224, 226], *C. elegans* [155], *A. thaliana* [222], and mammals [225] and H3K27me3 in *C. elegans* [230] and mice [227]. Reciprocally, studies in *S. pombe* and *C. elegans* have demonstrated that the establishment of H3K9 methylation by small RNAs is required for the long-term maintenance of RNAi silencing, especially across generations [44, 231]. These findings suggest that small RNAs and H3K9 methylation interact, as feedback loops, to maintain silencing at certain genomic loci. However, in *D. melanogaster* Dicer 2 and Argonaute 2 are associated with euchromatic loci [229] suggesting that there might be some species specific divergence with these crosstalk pathways.

Similarly to RNAi and histone modifications, DNA methylation is also important for directing histone methylation and vice versa, through the physical interaction between modifying enzymes and the alternative epigenetic modification [232–237]. Furthermore, many other signaling and metabolic pathways might also interact with these classical epigenetic pathways to mediate multigenerational effects in ways that are as-yet-unknown. For example, recent studies have found that differential deposition of lipids into germ cells can trigger transgenerational changes in histone methylation patterns [238]. These feedback loops are important for maintaining a variety of transgenerational phenotypes [60, 230] and thus make it more difficult to identify the initiating signal, what is transmitted from parents to their children, and what is mediating the phenotypic consequences. It is likely that these intercommunicating networks are essential for most, if not all, epigenetic inheritance paradigms, and that perturbance of any node of this network will elicit disruptions in other interconnected epigenetic pathways.

Conclusions and Future Directions

While we have mostly focused on DNA methylation, histone methylation, and small noncoding RNA as potential carriers of heritable non-genetic information other mechanisms of inheritance clearly exist. We have not been able to give sufficient attention to the inheritance

of prions and other maternally inherited proteins as well as the microbiome which have both been extensively investigated. Additionally other putative carriers of non-genetic information which have not been studied but could readily transmit epigenetic information across generations include lipids, which are frequently used as cell to cell and organism to organism communication molecules and other RNAs which have diverse roles in virtually every aspect of biology. Epigenetic inheritance is in an exciting era of both discovery of new inter and transgenerational epigenetic inheritance phenomena as well as at the cusp of identifying the molecular mechanisms underlying these amazing and complex biological processes. Time will tell whether these diverse epigenetic cues coalesce into a common epigenetic signaling mechanism or whether a network of independent and sometimes interconnecting nodes of epigenetic information can be transmitted from ancestors to their descendants.

Acknowledgement:

We apologize for literature omitted owing to space limitations.

Funding:

this work was supported by the National Institutes of Health [grant numbers DP2AG055947 and R21HG010066 to E.L.G]; and the Next Generation Fellowship from the Centre for Trophoblast Research [to N.O.B.].

References

- Mendel GJ, Versuche über Pflanzen-Hybriden, Verhandlungen des naturforschenden Vereines in Brünn IV (1865) 3–47.
- [2]. Correns C., Untersuchungen ueber die Xenien bei Zea mays, Berichte der Deutschen Botanischen Gesellschaft 18 (1900) 158–168.
- [3]. de Vries H., Sur la fécondation hybride de l'albumen, Biologisches Zentralblatt 20 (1900) 129– 130.
- [4]. Tschermak-Seysenegg E.v., Ueber künstliche Kreuzung bei Pisum sativum, Berichte der Deutschen Botanischen Gesellschaft 18 (1900) 232–239.
- [5]. Avery OT, Macleod CM, McCarty M., Studies on the Chemical Nature of the Substance Inducing Transformation of Pneumococcal Types : Induction of Transformation by a Desoxyribonucleic Acid Fraction Isolated from Pneumococcus Type Iii, J Exp Med 79(2) (1944) 137–58. [PubMed: 19871359]
- [6]. McCracken I., Heredity of the race-characters univoltinism and bivoltinism in the silkworm (Bombyx mori). A case of non-Mendelian inheritance, Journal of Experimental Zoology 7(4) (1909) 747–764.
- [7]. Watanabe K., Studies on the voltinism in the silkworm, Bombyx mori, Bull. seric. Exp. Sta 6 (1924) 411–455.
- [8]. Umeya Y., On the experiments of ovarian transplanta-tion and blood transfusion in the silkworm with special ref-erence to the alternation of voltinism, Jpn. J. Genet 3 (1925) 155–182.
- [9]. Hasegawa K., The Diapause Hormone of the Silkworm, Bombyx mori, Nature 179(4573) (1957) 1300–1301.
- [10]. Yamashita O, Hasegawa K., Studies on the mode of action of the diapause hormone in the silkworm, Bombyx mori L, The journal of sericultural science of Japan 33 (1964) 115–123.
- [11]. Yamashita O., Hormonal and metabolic control of egg diapause of the silkworm, Bombyx mori (Lepidoptera : Bombycidae), Entomol. Gen 7 (1981) 195–211.
- [12]. Ikeda M, Su Z.-h., Saito H, Imai K, Sato Y, Isobe M, Yamashita O., Induction of embryonic diapause and stimulation of ovary trehalase activity in the silkworm, Bombyx mori, by synthetic diapause hormone, Journal of Insect Physiology 39(10) (1993) 889–895.

- [13]. Galton F., Experiments in Pangenesis, by Breeding from Rabbits of a Pure Variety, into whose Circulation Blood Taken from other Varieties had Previously Been Largely Transfused, Proc. Roy. Soc 19 (1871) 393–410.
- [14]. Darwin C., The variation of animals and plants under domestication, 0. Judd1868.
- [15]. Weismann A., The germ-plasm: a theory of heredity, Scribner's1893.
- [16]. Perez MF, Lehner B., Intergenerational and transgenerational epigenetic inheritance in animals, Nat Cell Biol 21(2) (2019) 143–151. [PubMed: 30602724]
- [17]. Uller T, Nakagawa S, English S., Weak evidence for anticipatory parental effects in plants and animals, Journal of Evolutionary Biology 26(10) (2013) 2161–2170. [PubMed: 23937440]
- [18]. Mousseau TA, Fox CW, The adaptive significance of maternal effects, Trends in Ecology & Evolution 13(10) (1998) 403–407. [PubMed: 21238360]
- [19]. van Otterdijk SD, Michels KB, Transgenerational epigenetic inheritance in mammals: how good is the evidence?, The FASEB Journal 30(7) (2016) 2457–2465. [PubMed: 27037350]
- [20]. Chen Q, Yan M, Cao Z, Li X, Zhang Y, Shi J, Feng GH, Peng H, Zhang X, Zhang Y, Qian J, Duan E, Zhai Q, Zhou Q., Sperm tsRNAs contribute to intergenerational inheritance of an acquired metabolic disorder, Science (New York, N.Y 351(6271) (2016) 397–400. [PubMed: 26721680]
- [21]. Sharma U, Conine CC, Shea JM, Boskovic A, Derr AG, Bing XY, Belleannee C, Kucukural A, Serra RW, Sun F, Song L, Carone BR, Ricci EP, Li XZ, Fauquier L, Moore MJ, Sullivan R, Mello CC, Garber M, Rando OJ, Biogenesis and function of tRNA fragments during sperm maturation and fertilization in mammals, Science (New York, N.Y 351(6271) (2016) 391. [PubMed: 26721685]
- [22]. Burton NO, Furuta T, Webster AK, Kaplan REW, Baugh LR, Arur S, Horvitz HR, Insulin-like signalling to the maternal germline controls progeny response to osmotic stress, Nat Cell Biol 19(3) (2017) 252–257. [PubMed: 28166192]
- [23]. Willis AR, Zhao W, Sukhdeo R, Wadi L, El Jarkass HT, Claycomb JM, Reinke AW, A parental transcriptional response to microsporidia infection induces inherited immunity in offspring, Sci Adv 7(19) (2021).
- [24]. Grishok A, Tabara H, Mello CC, Genetic requirements for inheritance of RNAi in C. elegans, Science (New York, N.Y 287(5462) (2000) 2494–7. [PubMed: 10741970]
- [25]. Burton NO, Burkhart KB, Kennedy S., Nuclear RNAi maintains heritable gene silencing in Caenorhabditis elegans, Proceedings of the National Academy of Sciences of the United States of America 108(49) (2011) 19683–19688. [PubMed: 22106253]
- [26]. Brink RA, A Genetic Change Associated with the R Locus in Maize Which Is Directed and Potentially Reversible, Genetics 41(6) (1956) 872–89. [PubMed: 17247669]
- [27]. Vellichirammal NN, Gupta P, Hall TA, Brisson JA, Ecdysone signaling underlies the pea aphid transgenerational wing polyphenism, Proc Natl Acad Sci USA 114(6) (2017) 1419. [PubMed: 28115695]
- [28]. Zera AJ, Denno RF, PHYSIOLOGY AND ECOLOGY OF DISPERSAL POLYMORPHISM IN INSECTS, Annu. Rev. Entomol 42(1) (1997) 207–230. [PubMed: 15012313]
- [29]. Müller CB, Williams IS, Hardie J., The role of nutrition, crowding and interspecific interactions in the development of winged aphids, Ecological Entomology 26(3) (2001) 330–340.
- [30]. Luna E, Bruce TJ, Roberts MR, Flors V, Ton J., Next-generation systemic acquired resistance, Plant Physiol 158(2) (2012) 844–53. [PubMed: 22147520]
- [31]. Burton NO, Riccio C, Dallaire A, Price J, Jenkins B, Koulman A, Miska EA, Cysteine synthases CYSL-1 and CYSL-2 mediate C. elegans heritable adaptation to P. vranovensis infection, Nat Commun 11(1) (2020) 1741. [PubMed: 32269224]
- [32]. Burton NO, Willis AR, Fisher K, Braukmann F, Price J, Stevens L, Baugh LR, Reinke A, Miska EA, Intergenerational adaptations to stress are evolutionarily conserved, stressspecific, and have deleterious trade-offs, bioRxiv (2021) bioRxiv 2021.05.07.443118; doi: 10.1101/2021.05.07.443118
- [33]. Perez MF, Shamalnasab M, Mata-Cabana A, Valle SD, Olmedo M, Francesconi M, Lehner B., Neuronal perception of the social environment generates an inherited memory that controls the development and generation time of *C. elegans*, Curr Biol 31 (2021) 1–13. [PubMed: 33065012]

- [34]. Agrawal AA, Laforsch C, Tollrian R., Transgenerational induction of defences in animals and plants, Nature 401 (1999) 60–63.
- [35]. Dantzer B, Newman AE, Boonstra R, Palme R, Boutin S, Humphries MM, McAdam AG, Density triggers maternal hormones that increase adaptive offspring growth in a wild mammal, Science (New York, N.Y 340(6137) (2013) 1215–7.
- [36]. Langley-Evans SC, Developmental programming of health and disease, Proc Nutr Soc 65(1) (2006) 97–105. [PubMed: 16441949]
- [37]. Uller T., Developmental plasticity and the evolution of parental effects, Trends in Ecology & Evolution 23(8) (2008) 432–438. [PubMed: 18586350]
- [38]. Burton NO, Willis A, Fisher K, Braukmann F, Price J, Stevens L, Ryan Baugh L, Reinke A, Miska EA, Intergenerational adaptations to stress are evolutionarily conserved, stress-specific, and have deleterious trade-offs, bioRxiv (2021) 2021.05.07.443118.
- [39]. Chandler VL, Paramutation: From Maize to Mice, Cell 128(4) (2007) 641–645. [PubMed: 17320501]
- [40]. de Vanssay A, Bougé A-L, Boivin A, Hermant C, Teysset L, Delmarre V, Antoniewski C, Ronsseray S., Paramutation in Drosophila linked to emergence of a piRNA-producing locus, Nature 490(7418) (2012) 112–115. [PubMed: 22922650]
- [41]. Sapetschnig A, Sarkies P, Lehrbach NJ, Miska EA, Tertiary siRNAs mediate paramutation in C. elegans, PLoS genetics 11(3) (2015) e1005078–e1005078. [PubMed: 25811365]
- [42]. Rassoulzadegan M, Grandjean V, Gounon P, Vincent S, Gillot I, Cuzin F., RNA-mediated non-mendelian inheritance of an epigenetic change in the mouse, Nature 441(7092) (2006) 469–74.
 [PubMed: 16724059]
- [43]. Buckley BA, Burkhart KB, Gu SG, Spracklin G, Kershner A, Fritz H, Kimble J, Fire A, Kennedy S., A nuclear Argonaute promotes multigenerational epigenetic inheritance and germline immortality, Nature 489(7416) (2012) 447–51. [PubMed: 22810588]
- [44]. Ashe A, Sapetschnig A, Weick EM, Mitchell J, Bagijn MP, Cording AC, Doebley AL, Goldstein LD, Lehrbach NJ, Le Pen J, Pintacuda G, Sakaguchi A, Sarkies P, Ahmed S, Miska EA, piRNAs can trigger a multigenerational epigenetic memory in the germline of C. elegans, Cell 150(1) (2012) 88–99. [PubMed: 22738725]
- [45]. Ewe CK, Torres Cleuren YN, Flowers SE, Alok G, Snell RG, Rothman JH, Natural cryptic variation in epigenetic modulation of an embryonic gene regulatory network, Proc Natl Acad Sci USA 117(24) (2020) 13637. [PubMed: 32482879]
- [46]. Rechavi O, Minevich G, Hobert O., Transgenerational Inheritance of an Acquired Small RNA-Based Antiviral Response in C. elegans, Cell 147(6) (2011) 1248–1256. [PubMed: 22119442]
- [47]. Rechavi O, Houri-Ze'evi L, Anava S, Goh WS, Kerk SY, Hannon GJ, Hobert O., Starvation-Induced Transgenerational Inheritance of Small RNAs in C. elegans, Cell 158(2) (2014) 277–87. [PubMed: 25018105]
- [48]. Houri-Ze'evi L, Korem Y, Sheftel H, Faigenbloom L, Toker IA, Dagan Y, Awad L, Degani L, Alon U, Rechavi O., A Tunable Mechanism Determines the Duration of the Transgenerational Small RNA Inheritance in C. elegans, Cell 165(1) (2016) 88–99. [PubMed: 27015309]
- [49]. Houri-Zeevi L, Korem Kohanim Y, Antonova O, Rechavi O., Three Rules Explain Transgenerational Small RNA Inheritance in C. elegans, Cell 182(5) (2020) 1186–1197 e12. [PubMed: 32841602]
- [50]. Lev I, Gingold H, Rechavi O., H3K9me3 is required for inheritance of small RNAs that target a unique subset of newly evolved genes, eLife 8 (2019) e40448. [PubMed: 30869075]
- [51]. Lev I, Seroussi U, Gingold H, Bril R, Anava S, Rechavi O., MET-2-Dependent H3K9 Methylation Suppresses Transgenerational Small RNA Inheritance, Current Biology 27(8) (2017) 1138–1147. [PubMed: 28343968]
- [52]. Posner R, Toker IA, Antonova O, Star E, Anava S, Azmon E, Hendricks M, Bracha S, Gingold H, Rechavi O., Neuronal Small RNAs Control Behavior Transgenerationally, Cell 177(7) (2019) 1814–1826.e15. [PubMed: 31178120]
- [53]. Ma C, Niu R, Huang T, Shao LW, Peng Y, Ding W, Wang Y, Jia G, He C, Li CY, He A, Liu Y., N6-methyldeoxyadenine is a transgenerational epigenetic signal for mitochondrial stress adaptation, Nat Cell Biol 21(3) (2019) 319–327. [PubMed: 30510156]

- [54]. Webster AK, Jordan JM, Hibshman JD, Chitrakar R, Baugh LR, Transgenerational Effects of Extended Dauer Diapause on Starvation Survival and Gene Expression Plasticity in Caenorhabditis elegans, Genetics 210(1) (2018) 263–274. [PubMed: 30049782]
- [55]. Palominos MF, Verdugo L, Gabaldon C, Pollak B, Ortíz-Severín J, Varas MA, Chávez FP, Calixto A., Transgenerational Diapause as an Avoidance Strategy against Bacterial Pathogens in Caenorhabditis elegans, mBio 8(5) (2017) e01234–17. [PubMed: 29018118]
- [56]. Bozler J, Kacsoh BZ, Bosco G., Transgeneratonal inheritance of ethanol preference is caused by maternal NPF repression, eLife 8 (2019) e45391. [PubMed: 31287057]
- [57]. Akay A, Di Domenico T, Suen KM, Nabih A, Parada GE, Larance M, Medhi R, Berkyurek AC, Zhang X, Wedeles CJ, Rudolph KLM, Engelhardt J, Hemberg M, Ma P, Lamond AI, Claycomb JM, Miska EA, The Helicase Aquarius/EMB-4 Is Required to Overcome Intronic Barriers to Allow Nuclear RNAi Pathways to Heritably Silence Transcription, Developmental Cell 42(3) (2017) 241–255.e6. [PubMed: 28787591]
- [58]. Kaletsky R, Moore RS, Vrla GD, Parsons LR, Gitai Z, Murphy CT, elegans C interprets bacterial non-coding RNAs to learn pathogenic avoidance, Nature 586(7829) (2020) 445–451. [PubMed: 32908307]
- [59]. Klosin A, Casas E, Hidalgo-Carcedo C, Vavouri T, Lehner B., Transgenerational transmission of environmental information in C. elegans, Science (New York, N.Y 356(6335) (2017) 320–323.
- [60]. Moore RS, Kaletsky R, Murphy CT, Piwi/PRG-1 Argonaute and TGF-beta Mediate Transgenerational Learned Pathogenic Avoidance, Cell 177(7) (2019) 1827–1841 e12. [PubMed: 31178117]
- [61]. Barucci G, Cornes E, Singh M, Li B, Ugolini M, Samolygo A, Didier C, Dingli F, Loew D, Quarato P, Cecere G., Small-RNA-mediated transgenerational silencing of histone genes impairs fertility in piRNA mutants, Nat Cell Biol 22(2) (2020) 235–245. [PubMed: 32015436]
- [62]. Stassen JHM, López A, Jain R, Pascual-Pardo D, Luna E, Smith LM, Ton J., The relationship between transgenerational acquired resistance and global DNA methylation in Arabidopsis, Scientific reports 8(1) (2018) 14761–14761. [PubMed: 30283021]
- [63]. Greer EL, Beese-Sims SE, Brookes E, Spadafora R, Zhu Y, Rothbart SB, Aristizabal-Corrales D, Chen S, Badeaux AI, Jin Q, Wang W, Strahl BD, Colaiacovo MP, Shi Y., A histone methylation network regulates transgenerational epigenetic memory in C. elegans, Cell Reports 7(1) (2014) 113–26. [PubMed: 24685137]
- [64]. Greer EL, Maures TJ, Ucar D, Hauswirth AG, Mancini E, Lim JP, Benayoun BA, Shi Y, Brunet A., Transgenerational epigenetic inheritance of longevity in Caenorhabditis elegans, Nature 479(7373) (2011) 365–71. [PubMed: 22012258]
- [65]. Lee H-C, Gu W, Shirayama M, Youngman E, Conte D Jr., Mello CC, elegans C piRNAs mediate the genome-wide surveillance of germline transcripts, Cell 150(1) (2012) 78–87. [PubMed: 22738724]
- [66]. Shirayama M, Seth M, Lee HC, Gu W, Ishidate T, Conte D Jr., Mello CC, piRNAs initiate an epigenetic memory of nonself RNA in the C. elegans germline, Cell 150(1) (2012) 65–77. [PubMed: 22738726]
- [67]. Perales R, Pagano D, Wan G, Fields BD, Saltzman AL, Kennedy SG, Transgenerational Epigenetic Inheritance Is Negatively Regulated by the HERI-1 Chromodomain Protein, Genetics 210(4) (2018) 1287–1299. [PubMed: 30389807]
- [68]. Kerr SC, Ruppersburg CC, Francis JW, Katz DJ, SPR-5 and MET-2 function cooperatively to reestablish an epigenetic ground state during passage through the germ line, Proceedings of the National Academy of Sciences of the United States of America 111(26) (2014) 9509–14. [PubMed: 24979765]
- [69]. Lee TW, David HS, Engstrom AK, Carpenter BS, Katz DJ, Repressive H3K9me2 protects lifespan against the transgenerational burden of COMPASS activity in C. elegans, Elife 8 (2019).
- [70]. Uller T, English S, Pen I., When is incomplete epigenetic resetting in germ cells favoured by natural selection?, Proceedings of the Royal Society B: Biological Sciences 282(1811) (2015) 20150682.

- [71]. Fire A, Xu S, Montgomery MK, Kostas SA, Driver SE, Mello CC, Potent and specific genetic interference by double-stranded RNA in Caenorhabditis elegans, Nature 391(6669) (1998) 806– 811. [PubMed: 9486653]
- [72]. Shukla A, Yan J, Pagano DJ, Dodson AE, Fei Y, Gorham J, Seidman JG, Wickens M, Kennedy S., poly(UG)-tailed RNAs in genome protection and epigenetic inheritance, Nature 582(7811) (2020) 283–288. [PubMed: 32499657]
- [73]. Kawasaki I, Shim YH, Kirchner J, Kaminker J, Wood WB, Strome S., PGL-1, a predicted RNA-binding component of germ granules, is essential for fertility in C. elegans, Cell 94(5) (1998) 635–45. [PubMed: 9741628]
- [74]. Wan G, Fields BD, Spracklin G, Shukla A, Phillips CM, Kennedy S., Spatiotemporal regulation of liquid-like condensates in epigenetic inheritance, Nature 557(7707) (2018) 679– 683. [PubMed: 29769721]
- [75]. Huypens P, Sass S, Wu M, Dyckhoff D, Tschop M, Theis F, Marschall S, Hrabe de Angelis M, Beckers J., Epigenetic germline inheritance of diet-induced obesity and insulin resistance, Nature genetics 48(5) (2016) 497–9. [PubMed: 26974008]
- [76]. Tops BBJ, Tabara H, Sijen T, Simmer F, Mello CC, Plasterk RHA, Ketting RF, RDE-2 interacts with MUT-7 to mediate RNA interference in Caenorhabditis elegans, Nucleic Acids Res 33(1) (2005) 347–355. [PubMed: 15653635]
- [77]. Chen Q, Yan W, Duan E., Epigenetic inheritance of acquired traits through sperm RNAs and sperm RNA modifications, Nature Reviews Genetics 17(12) (2016) 733–743.
- [78]. Peng H, Shi J, Zhang Y, Zhang H, Liao S, Li W, Lei L, Han C, Ning L, Cao Y, Zhou Q, Chen Q, Duan E., A novel class of tRNA-derived small RNAs extremely enriched in mature mouse sperm, Cell Res 22(11) (2012) 1609–1612. [PubMed: 23044802]
- [79]. Sharma U, Sun F, Conine CC, Reichholf B, Kukreja S, Herzog VA, Ameres SL, Rando OJ, Small RNAs Are Trafficked from the Epididymis to Developing Mammalian Sperm, Dev Cell 46(4) (2018) 481–494 e6. [PubMed: 30057273]
- [80]. Yuan S, Schuster A, Tang C, Yu T, Ortogero N, Bao J, Zheng H, Yan W., Sperm-borne miRNAs and endo-siRNAs are important for fertilization and preimplantation embryonic development, Development (Cambridge, England) 143(4) (2016) 635–647.
- [81]. Conine CC, Moresco JJ, Gu W, Shirayama M, Conte D Jr., Yates JR 3rd, Mello CC, Argonautes Promote Male Fertility and Provide a Paternal Memory of Germline Gene Expression in C. elegans, Cell 155(7) (2013) 1532–44. [PubMed: 24360276]
- [82]. Conine CC, Sun F, Song L, Rivera-Pérez JA, Rando OJ, Small RNAs Gained during Epididymal Transit of Sperm Are Essential for Embryonic Development in Mice, Developmental Cell 46(4) (2018) 470–480.e3. [PubMed: 30057276]
- [83]. Gapp K, Jawaid A, Sarkies P, Bohacek J, Pelczar P, Prados J, Farinelli L, Miska E, Mansuy IM, Implication of sperm RNAs in transgenerational inheritance of the effects of early trauma in mice, Nat Neurosci 17(5) (2014) 667–669. [PubMed: 24728267]
- [84]. Rodgers AB, Morgan CP, Leu NA, Bale TL, Transgenerational epigenetic programming via sperm microRNA recapitulates effects of paternal stress, Proc Natl Acad Sci USA 112(44) (2015) 13699. [PubMed: 26483456]
- [85]. de Castro Barbosa T, Ingerslev LR, Alm PS, Versteyhe S, Massart J, Rasmussen M, Donkin I, Sjögren R, Mudry JM, Vetterli L, Gupta S, Krook A, Zierath JR, Barrès R., High-fat diet reprograms the epigenome of rat spermatozoa and transgenerationally affects metabolism of the offspring, Molecular metabolism 5(3) (2015) 184–197. [PubMed: 26977389]
- [86]. Surani MA, Barton SC, Development of gynogenetic eggs in the mouse: implications for parthenogenetic embryos, Science (New York, N.Y 222(4627) (1983) 1034–6.
- [87]. Surani MA, Barton SC, Norris ML, Development of reconstituted mouse eggs suggests imprinting of the genome during gametogenesis, Nature 308(5959) (1984) 548–50. [PubMed: 6709062]
- [88]. McGrath J, Solter D., Completion of mouse embryogenesis requires both the maternal and paternal genomes, Cell 37(1) (1984) 179–83. [PubMed: 6722870]

- [89]. Barlow DP, Stoger R, Herrmann BG, Saito K, Schweifer N., The mouse insulin-like growth factor type-2 receptor is imprinted and closely linked to the Tme locus, Nature 349(6304) (1991) 84–7. [PubMed: 1845916]
- [90]. DeChiara TM, Robertson EJ, Efstratiadis A., Parental imprinting of the mouse insulin-like growth factor II gene, Cell 64(4) (1991) 849–59. [PubMed: 1997210]
- [91]. Bartolomei MS, Zemel S, Tilghman SM, Parental imprinting of the mouse H19 gene, Nature 351(6322) (1991) 153–5. [PubMed: 1709450]
- [92]. Ferguson-Smith AC, Cattanach BM, Barton SC, Beechey CV, Surani MA, Embryological and molecular investigations of parental imprinting on mouse chromosome 7, Nature 351(6328) (1991) 667–70. [PubMed: 2052093]
- [93]. Bartolomei MS, Webber AL, Brunkow ME, Tilghman SM, Epigenetic mechanisms underlying the imprinting of the mouse H19 gene, Genes & development 7(9) (1993) 1663–73. [PubMed: 7690336]
- [94]. Ferguson-Smith AC, Sasaki H, Cattanach BM, Surani MA, Parental-origin-specific epigenetic modification of the mouse H19 gene, Nature 362(6422) (1993) 751–5. [PubMed: 8469285]
- [95]. Stoger R, Kubicka P, Liu CG, Kafri T, Razin A, Cedar H, Barlow DP, Maternal-specific methylation of the imprinted mouse Igf2r locus identifies the expressed locus as carrying the imprinting signal, Cell 73(1) (1993) 61–71. [PubMed: 8462104]
- [96]. Li E, Beard C, Jaenisch R., Role for DNA methylation in genomic imprinting, Nature 366(6453) (1993) 362–5. [PubMed: 8247133]
- [97]. Lee JT, Bartolomei MS, X-inactivation, imprinting, and long noncoding RNAs in health and disease, Cell 152(6) (2013) 1308–23. [PubMed: 23498939]
- [98]. Eckersley-Maslin MA, Alda-Catalinas C, Reik W., Dynamics of the epigenetic landscape during the maternal-to-zygotic transition, Nature reviews 19(7) (2018) 436–450.
- [99]. Tucci V, Isles AR, Kelsey G, Ferguson-Smith AC, Erice Imprinting G., Genomic Imprinting and Physiological Processes in Mammals, Cell 176(5) (2019) 952–965. [PubMed: 30794780]
- [100]. Gehring M, Bubb KL, Henikoff S., Extensive demethylation of repetitive elements during seed development underlies gene imprinting, Science (New York, N.Y 324(5933) (2009) 1447–51.
- [101]. Xu M, Long C, Chen X, Huang C, Chen S, Zhu B., Partitioning of histone H3-H4 tetramers during DNA replication-dependent chromatin assembly, Science (New York, N.Y 328(5974) (2010) 94–8.
- [102]. Cao R, Wang L, Wang H, Xia L, Erdjument-Bromage H, Tempst P, Jones RS, Zhang Y, Role of histone H3 lysine 27 methylation in Polycomb-group silencing, Science (New York, N.Y 298(5595) (2002) 1039–43.
- [103]. Czermin B, Melfi R, McCabe D, Seitz V, Imhof A, Pirrotta V., Drosophila enhancer of Zeste/ESC complexes have a histone H3 methyltransferase activity that marks chromosomal Polycomb sites, Cell 111(2) (2002) 185–96. [PubMed: 12408863]
- [104]. Kuzmichev A, Nishioka K, Erdjument-Bromage H, Tempst P, Reinberg D., Histone methyltransferase activity associated with a human multiprotein complex containing the Enhancer of Zeste protein, Genes & development 16(22) (2002) 2893–905. [PubMed: 12435631]
- [105]. Muller J, Hart CM, Francis NJ, Vargas ML, Sengupta A, Wild B, Miller EL, O'Connor MB, Kingston RE, Simon JA, Histone methyltransferase activity of a Drosophila Polycomb group repressor complex, Cell 111(2) (2002) 197–208. [PubMed: 12408864]
- [106]. Francis NJ, Follmer NE, Simon MD, Aghia G, Butler JD, Polycomb proteins remain bound to chromatin and DNA during DNA replication in vitro, Cell 137(1) (2009) 110–22. [PubMed: 19303136]
- [107]. Petruk S, Sedkov Y, Johnston DM, Hodgson JW, Black KL, Kovermann SK, Beck S, Canaani E, Brock HW, Mazo A., TrxG and PcG proteins but not methylated histones remain associated with DNA through replication, Cell 150(5) (2012) 922–33. [PubMed: 22921915]
- [108]. Tran V, Lim C, Xie J, Chen X., Asymmetric division of Drosophila male germline stem cell shows asymmetric histone distribution, Science (New York, N.Y 338(6107) (2012) 679–82. [PubMed: 23118191]

- [109]. Xie J, Wooten M, Tran V, Chen BC, Pozmanter C, Simbolon C, Betzig E, Chen X., Histone H3 Threonine Phosphorylation Regulates Asymmetric Histone Inheritance in the Drosophila Male Germline, Cell 163(4) (2015) 920–33. [PubMed: 26522592]
- [110]. Inoue A, Jiang L, Lu F, Suzuki T, Zhang Y., Maternal H3K27me3 controls DNA methylationindependent imprinting, Nature 547(7664) (2017) 419–424. [PubMed: 28723896]
- [111]. Inoue A, Jiang L, Lu F, Zhang Y., Genomic imprinting of Xist by maternal H3K27me3, Genes & development 31(19) (2017) 1927–1932. [PubMed: 29089420]
- [112]. Inoue A, Chen Z, Yin Q, Zhang Y., Maternal Eed knockout causes loss of H3K27me3 imprinting and random X inactivation in the extraembryonic cells, Genes & development 32(23– 24) (2018) 1525–1536. [PubMed: 30463900]
- [113]. Mei H, Kozuka C, Hayashi R, Kumon M, Koseki H, Inoue A., H2AK119ub1 guides maternal inheritance and zygotic deposition of H3K27me3 in mouse embryos, Nature genetics 53(4) (2021) 539–550. [PubMed: 33821003]
- [114]. Chen Z, Djekidel MN, Zhang Y., Distinct dynamics and functions of H2AK119ub1 and H3K27me3 in mouse preimplantation embryos, Nature genetics 53(4) (2021) 551–563. [PubMed: 33821005]
- [115]. Andergassen D, Smith ZD, Rinn JL, Meissner A., Diverse mechanisms for epigenetic imprinting in mammals, bioRxiv (2021) 10.1101/2021.04.30.442087
- [116]. Gaydos LJ, Wang W, Strome S., Gene repression. H3K27me and PRC2 transmit a memory of repression across generations and during development, Science (New York, N.Y 345(6203) (2014) 1515–8. [PubMed: 25237104]
- [117]. Zenk F, Loeser E, Schiavo R, Kilpert F, Bogdanovic O, Iovino N., Germ line-inherited H3K27me3 restricts enhancer function during maternal-to-zygotic transition, Science (New York, N.Y 357(6347) (2017) 212–216. [PubMed: 28706074]
- [118]. Kreher J, Takasaki T, Cockrum C, Sidoli S, Garcia BA, Jensen ON, Strome S., Distinct Roles of Two Histone Methyltransferases in Transmitting H3K36me3-Based Epigenetic Memory Across Generations in Caenorhabditis elegans, Genetics 210(3) (2018) 969–982. [PubMed: 30217796]
- [119]. Tabuchi TM, Rechtsteiner A, Jeffers TE, Egelhofer TA, Murphy CT, Strome S., Caenorhabditis elegans sperm carry a histone-based epigenetic memory of both spermatogenesis and oogenesis, Nat Commun 9(1) (2018) 4310. [PubMed: 30333496]
- [120]. Kaneshiro KR, Rechtsteiner A, Strome S., Sperm-inherited H3K27me3 impacts offspring transcription and development in C. elegans, Nat Commun 10(1) (2019) 1271. [PubMed: 30894520]
- [121]. Pavet V, Quintero C, Cecchini NM, Rosa AL, Alvarez ME, Arabidopsis displays centromeric DNA hypomethylation and cytological alterations of heterochromatin upon attack by pseudomonas syringae, Mol Plant Microbe Interact 19(6) (2006) 577–87. [PubMed: 16776291]
- [122]. Wibowo A, Becker C, Marconi G, Durr J, Price J, Hagmann J, Papareddy R, Putra H, Kageyama J, Becker J, Weigel D, Gutierrez-Marcos J., Hyperosmotic stress memory in Arabidopsis is mediated by distinct epigenetically labile sites in the genome and is restricted in the male germline by DNA glycosylase activity, Elife 5 (2016).
- [123]. Carone BR, Fauquier L, Habib N, Shea JM, Hart CE, Li R, Bock C, Li C, Gu H, Zamore PD, Meissner A, Weng Z, Hofmann HA, Friedman N, Rando OJ, Paternally induced transgenerational environmental reprogramming of metabolic gene expression in mammals, Cell 143(7) (2010) 1084–96. [PubMed: 21183072]
- [124]. Ng SF, Lin RC, Laybutt DR, Barres R, Owens JA, Morris MJ, Chronic high-fat diet in fathers programs beta-cell dysfunction in female rat offspring, Nature 467(7318) (2010) 963–6. [PubMed: 20962845]
- [125]. Radford EJ, Ito M, Shi H, Corish JA, Yamazawa K, Isganaitis E, Seisenberger S, Hore TA, Reik W, Erkek S, Peters A, Patti ME, Ferguson-Smith AC, In utero effects. In utero undernourishment perturbs the adult sperm methylome and intergenerational metabolism, Science (New York, N.Y 345(6198) (2014) 1255903. [PubMed: 25011554]
- [126]. Rando OJ, Simmons RA, I'm eating for two: parental dietary effects on offspring metabolism, Cell 161(1) (2015) 93–105. [PubMed: 25815988]

- [127]. Ost A, Lempradl A, Casas E, Weigert M, Tiko T, Deniz M, Pantano L, Boenisch U, Itskov PM, Stoeckius M, Ruf M, Rajewsky N, Reuter G, Iovino N, Ribeiro C, Alenius M, Heyne S, Vavouri T, Pospisilik JA, Paternal diet defines offspring chromatin state and intergenerational obesity, Cell 159(6) (2014) 1352–64. [PubMed: 25480298]
- [128]. Guida MC, Birse RT, Dall'Agnese A, Toto PC, Diop SB, Mai A, Adams PD, Puri PL, Bodmer R., Intergenerational inheritance of high fat diet-induced cardiac lipotoxicity in Drosophila, Nat Commun 10(1) (2019) 193. [PubMed: 30643137]
- [129]. Lesch BJ, Tothova Z, Morgan EA, Liao Z, Bronson RT, Ebert BL, Page DC, Intergenerational epigenetic inheritance of cancer susceptibility in mammals, Elife 8 (2019).
- [130]. Seong KH, Li D, Shimizu H, Nakamura R, Ishii S, Inheritance of stress-induced ATF-2dependent epigenetic change, Cell 145(7) (2011) 1049–61. [PubMed: 21703449]
- [131]. Boulinier T, Staszewski V., Maternal transfer of antibodies: raising immuno-ecology issues, Trends in Ecology & Evolution 23(5) (2008) 282–288. [PubMed: 18375011]
- [132]. Perez MF, Francesconi M, Hidalgo-Carcedo C, Lehner B., Maternal age generates phenotypic variation in Caenorhabditis elegans, Nature 552(7683) (2017) 106–109. [PubMed: 29186117]
- [133]. Jordan JM, Hibshman JD, Webster AK, Kaplan REW, Leinroth A, Guzman R, Maxwell CS, Chitrakar R, Bowman EA, Fry AL, Hubbard EJA, Baugh LR, Insulin/IGF Signaling and Vitellogenin Provisioning Mediate Intergenerational Adaptation to Nutrient Stress, Current biology : CB 29(14) (2019) 2380–2388.e5. [PubMed: 31280992]
- [134]. Groothuis TGG, Hsu B-Y, Kumar N, Tschirren B., Revisiting mechanisms and functions of prenatal hormone-mediated maternal effects using avian species as a model, Philosophical Transactions of the Royal Society B: Biological Sciences 374(1770) (2019) 20180115.
- [135]. Painter RC, Roseboom TJ, Bleker OP, Prenatal exposure to the Dutch famine and disease in later life: An overview, Reproductive Toxicology 20(3) (2005) 345–352. [PubMed: 15893910]
- [136]. Roseboom T, de Rooij S, Painter R., The Dutch famine and its long-term consequences for adult health, Early Human Development 82(8) (2006) 485–491. [PubMed: 16876341]
- [137]. Cheng Q, Trangucci R, Nelson KN, Fu W, Collender PA, Head JR, Hoover CM, Skaff NK, Li T, Li X, You Y, Fang L, Liang S, Yang C, He J.g., Zelner JL, Remais JV, Prenatal and early-life exposure to the Great Chinese Famine increased the risk of tuberculosis in adulthood across two generations, Proc Natl Acad Sci USA 117(44) (2020) 27549. [PubMed: 33077583]
- [138]. Li Y, He Y, Qi L, Jaddoe VW, Feskens EJM, Yang X, Ma G, Hu FB, Exposure to the Chinese Famine in Early Life and the Risk of Hyperglycemia and Type 2 Diabetes in Adulthood, Diabetes 59(10) (2010) 2400. [PubMed: 20622161]
- [139]. Hales CN, Barker DJP, Type 2 (non-insulin-dependent) diabetes mellitus: the thrifty phenotype hypothesis, Diabetologia 35(7) (1992) 595–601. [PubMed: 1644236]
- [140]. Simmons RA, Templeton LJ, Gertz SJ, Intrauterine Growth Retardation Leads to the Development of Type 2 Diabetes in the Rat, Diabetes 50(10) (2001) 2279. [PubMed: 11574409]
- [141]. Aiken CE, Ozanne SE, Transgenerational developmental programming, Human Reproduction Update 20(1) (2014) 63–75. [PubMed: 24082037]
- [142]. Petry CJ, Dorling MW, Pawlak DB, Ozanne SE, Hales CN, Diabetes in old male offspring of rat dams fed a reduced protein diet, Int J Exp Diabetes Res 2(2) (2001) 139–143. [PubMed: 12369717]
- [143]. Shankar K, Harrell A, Liu X, Gilchrist JM, Ronis MJJ, Badger TM, Maternal obesity at conception programs obesity in the offspring, American Journal of Physiology-Regulatory, Integrative and Comparative Physiology 294(2) (2008) R528–R538. [PubMed: 18032473]
- [144]. Sasson IE, Vitins AP, Mainigi MA, Moley KH, Simmons RA, Pre-gestational vs gestational exposure to maternal obesity differentially programs the offspring in mice, Diabetologia 58(3) (2015) 615–624. [PubMed: 25608625]
- [145]. Ernst C, Odom DT, Kutter C., The emergence of piRNAs against transposon invasion to preserve mammalian genome integrity, Nat Commun 8(1) (2017) 1411. [PubMed: 29127279]
- [146]. Parhad SS, Theurkauf WE, Rapid evolution and conserved function of the piRNA pathway, Open Biol 9(1) (2019) 180181–180181. [PubMed: 30958115]
- [147]. Saxe JP, Lin H., Small noncoding RNAs in the germline, Cold Spring Harb Perspect Biol 3(9) (2011) a002717–a002717. [PubMed: 21669983]

- [148]. Claycomb JM, Ancient endo-siRNA pathways reveal new tricks, Current biology : CB 24(15) (2014) R703–15. [PubMed: 25093565]
- [149]. Stam M, Belele C, Dorweiler JE, Chandler VL, Differential chromatin structure within a tandem array 100 kb upstream of the maize b1 locus is associated with paramutation, Genes & development 16(15) (2002) 1906–1918. [PubMed: 12154122]
- [150]. Stam M, Belele C, Ramakrishna W, Dorweiler JE, Bennetzen JL, Chandler VL, The regulatory regions required for B' paramutation and expression are located far upstream of the maize b1 transcribed sequences, Genetics 162(2) (2002) 917–930. [PubMed: 12399399]
- [151]. Arteaga-Vazquez M, Sidorenko L, Rabanal FA, Shrivistava R, Nobuta K, Green PJ, Meyers BC, Chandler VL, RNA-mediated trans-communication can establish paramutation at the b1 locus in maize, Proc Natl Acad Sci USA 107(29) (2010) 12986. [PubMed: 20616013]
- [152]. Ni JZ, Kalinava N, Chen E, Huang A, Trinh T, Gu SG, A transgenerational role of the germline nuclear RNAi pathway in repressing heat stress-induced transcriptional activation in C. elegans, Epigenetics Chromatin 9 (2016) 3. [PubMed: 26779286]
- [153]. Spracklin G, Fields B, Wan G, Becker D, Wallig A, Shukla A, Kennedy S., The RNAi Inheritance Machinery of Caenorhabditis elegans, Genetics 206(3) (2017) 1403. [PubMed: 28533440]
- [154]. Ni JZ, Chen E, Gu SG, Complex coding of endogenous siRNA, transcriptional silencing and H3K9 methylation on native targets of germline nuclear RNAi in C. elegans, BMC Genomics 15(1) (2014) 1157. [PubMed: 25534009]
- [155]. Gu SG, Pak J, Guang S, Maniar JM, Kennedy S, Fire A., Amplification of siRNA in Caenorhabditis elegans generates a transgenerational sequence-targeted histone H3 lysine 9 methylation footprint, Nature genetics 44(2) (2012) 157–64. [PubMed: 22231482]
- [156]. Yigit E, Batista PJ, Bei Y, Pang KM, Chen CC, Tolia NH, Joshua-Tor L, Mitani S, Simard MJ, Mello CC, Analysis of the C elegans Argonaute family reveals that distinct Argonautes act sequentially during RNAi, Cell 127(4) (2006) 747–57. [PubMed: 17110334]
- [157]. Tijsterman M, Okihara KL, Thijssen K, Plasterk RHA, PPW-1, a PAZ/PIWI Protein Required for Efficient Germline RNAi, Is Defective in a Natural Isolate of C. elegans, Current Biology 12(17) (2002) 1535–1540. [PubMed: 12225671]
- [158]. Xu F, Feng X, Chen X, Weng C, Yan Q, Xu T, Hong M, Guang S., A Cytoplasmic Argonaute Protein Promotes the Inheritance of RNAi, Cell Reports 23(8) (2018) 2482–2494. [PubMed: 29791857]
- [159]. Sijen T, Fleenor J, Simmer F, Thijssen KL, Parrish S, Timmons L, Plasterk RH, Fire A., On the role of RNA amplification in dsRNA-triggered gene silencing, Cell 107(4) (2001) 465–76. [PubMed: 11719187]
- [160]. Alcazar RM, Lin R, Fire AZ, Transmission Dynamics of Heritable Silencing Induced by Double-Stranded RNA in Caenorhabditis elegans, Genetics 180(3) (2008) 1275–1288. [PubMed: 18757930]
- [161]. Hibbard BM, The Role of Folic Acid in Pregnancy; with Particular Reference to Anaemia, Abruption and Abortion, J Obstet Gynaecol Br Commonw 71 (1964) 529–42. [PubMed: 14194440]
- [162]. Group MVSR, Prevention of neural tube defects: results of the Medical Research Council Vitamin Study. MRC Vitamin Study Research Group, Lancet 338(8760) (1991) 131–7. [PubMed: 1677062]
- [163]. Czeizel AE, Dudas I., Prevention of the first occurrence of neural-tube defects by periconceptional vitamin supplementation, N Engl J Med 327(26) (1992) 1832–5. [PubMed: 1307234]
- [164]. Crider KS, Bailey LB, Berry RJ, Folic acid food fortification-its history, effect, concerns, and future directions, Nutrients 3(3) (2011) 370–84. [PubMed: 22254102]
- [165]. Champe PC, Harvey RA, Lippincott's Illustrated Reviews: Biochemistry 2nd edition, Lippincott Williams & Wilkins (1994).
- [166]. Padmanabhan N, Jia D, Geary-Joo C, Wu X, Ferguson-Smith AC, Fung E, Bieda MC, Snyder FF, Gravel RA, Cross JC, Watson ED, Mutation in Folate Metabolism Causes Epigenetic Instability and Transgenerational Effects on Development, Cell (2013).

- [167]. Greer EL, Maures TJ, Hauswirth AG, Green EM, Leeman DS, Maro GS, Han S, Banko MR, Gozani O, Brunet A., Members of the H3K4 trimethylation complex regulate lifespan in a germline-dependent manner in C. elegans, Nature 466(7304) (2010) 383–7. [PubMed: 20555324]
- [168]. Xiao Y, Bedet C, Robert VJ, Simonet T, Dunkelbarger S, Rakotomalala C, Soete G, Korswagen HC, Strome S, Palladino F., Caenorhabditis elegans chromatin-associated proteins SET-2 and ASH-2 are differentially required for histone H3 Lys 4 methylation in embryos and adult germ cells, Proceedings of the National Academy of Sciences of the United States of America 108(20) (2011) 8305–10. [PubMed: 21527717]
- [169]. Robert VJ, Mercier MG, Bedet C, Janczarski S, Merlet J, Garvis S, Ciosk R, Palladino F, The SET-2/SET1 histone H3K4 methyltransferase maintains pluripotency in the Caenorhabditis elegans germline, Cell Rep 9(2) (2014) 443–50. [PubMed: 25310986]
- [170]. Robert VJ, Knutson AK, Rechsteiner A, Yvert G, Strome S, Palladino F., The C. elegans SET-2 histone methyltransferase maintains germline fate by preventing progressive transcriptomic deregulation across generations, bioRxiv (2019) 10.1101/583799
- [171]. Muramoto T, Muller I, Thomas G, Melvin A, Chubb JR, Methylation of H3K4 Is required for inheritance of active transcriptional states, Current biology : CB 20(5) (2010) 397–406. [PubMed: 20188556]
- [172]. Katz DJ, Edwards TM, Reinke V, Kelly WG, A C. elegans LSD1 demethylase contributes to germline immortality by reprogramming epigenetic memory, Cell 137(2) (2009) 308–20. [PubMed: 19379696]
- [173]. Greer EL, Becker B, Latza C, Antebi A, Shi Y, Mutation of C elegans demethylase spr-5 extends transgenerational longevity, Cell Res 26(2) (2016) 229–38. [PubMed: 26691751]
- [174]. Siklenka K, Erkek S, Godmann M, Lambrot R, McGraw S, Lafleur C, Cohen T, Xia J, Suderman M, Hallett M, Trasler J, Peters AH, Kimmins S., Disruption of histone methylation in developing sperm impairs offspring health transgenerationally, Science (New York, N.Y 350(6261) (2015) aab2006. [PubMed: 26449473]
- [175]. Schott D, Yanai I, Hunter CP, Natural RNA interference directs a heritable response to the environment, Sci Rep 4 (2014) 7387. [PubMed: 25552271]
- [176]. Cavalli G, Paro R., The Drosophila Fab-7 chromosomal element conveys epigenetic inheritance during mitosis and meiosis, Cell 93(4) (1998) 505–18. [PubMed: 9604927]
- [177]. Cavalli G, Paro R., Epigenetic inheritance of active chromatin after removal of the main transactivator, Science (New York, N.Y 286(5441) (1999) 955–8. [PubMed: 10542150]
- [178]. Ciabrelli F, Comoglio F, Fellous S, Bonev B, Ninova M, Szabo Q, Xuereb A, Klopp C, Aravin A, Paro R, Bantignies F, Cavalli G., Stable Polycomb-dependent transgenerational inheritance of chromatin states in Drosophila, Nature genetics 49(6) (2017) 876–886. [PubMed: 28436983]
- [179]. Robles P, Turner A, Zuco G, Adams S, Paganopolou P, Winton M, Hill B, Kache V, Bateson C, Pires-daSilva A., Parental energy-sensing pathways control intergenerational offspring sex determination in the nematode Auanema freiburgensis, BMC Biol 19(1) (2021) 102. [PubMed: 34001117]
- [180]. Carpenter BS, Lee TW, Plott CF, Rodriguez JD, Brockett JS, Myrick DA, Katz DJ, Caenorhabditis elegans establishes germline versus soma by balancing inherited histone methylation, Development (Cambridge, England) 148(3) (2021).
- [181]. Lismer A, Dumeaux V, Lafleur C, Lambrot R, Brind'Amour J, Lorincz MC, Kimmins S., Histone H3 lysine 4 trimethylation in sperm is transmitted to the embryo and associated with diet-induced phenotypes in the offspring, Dev Cell 56(5) (2021) 671–686 e6. [PubMed: 33596408]
- [182]. Wigler M, Levy D, Perucho M., The somatic replication of DNA methylation, Cell 24(1) (1981) 33–40. [PubMed: 6263490]
- [183]. Hajkova P, Erhardt S, Lane N, Haaf T, El-Maarri O, Reik W, Walter J, Surani MA, Epigenetic reprogramming in mouse primordial germ cells, Mech Dev 117(1–2) (2002) 15–23. [PubMed: 12204247]
- [184]. Seki Y, Hayashi K, Itoh K, Mizugaki M, Saitou M, Matsui Y, Extensive and orderly reprogramming of genome-wide chromatin modifications associated with specification and early

development of germ cells in mice, Developmental biology 278(2) (2005) 440–58. [PubMed: 15680362]

- [185]. Lane N, Dean W, Erhardt S, Hajkova P, Surani A, Walter J, Reik W., Resistance of IAPs to methylation reprogramming may provide a mechanism for epigenetic inheritance in the mouse, Genesis 35(2) (2003) 88–93. [PubMed: 12533790]
- [186]. Popp C, Dean W, Feng S, Cokus SJ, Andrews S, Pellegrini M, Jacobsen SE, Reik W., Genomewide erasure of DNA methylation in mouse primordial germ cells is affected by AID deficiency, Nature 463(7284) (2010) 1101–5. [PubMed: 20098412]
- [187]. Duhl DM, Vrieling H, Miller KA, Wolff GL, Barsh GS, Neomorphic agouti mutations in obese yellow mice, Nature genetics 8(1) (1994) 59–65. [PubMed: 7987393]
- [188]. Morgan HD, Sutherland HG, Martin DI, Whitelaw E., Epigenetic inheritance at the agouti locus in the mouse, Nature genetics 23(3) (1999) 314–8. [PubMed: 10545949]
- [189]. Rakyan VK, Chong S, Champ ME, Cuthbert PC, Morgan HD, Luu KV, Whitelaw E., Transgenerational inheritance of epigenetic states at the murine Axin(Fu) allele occurs after maternal and paternal transmission, Proceedings of the National Academy of Sciences of the United States of America 100(5) (2003) 2538–43. [PubMed: 12601169]
- [190]. Kazachenka A, Bertozzi TM, Sjoberg-Herrera MK, Walker N, Gardner J, Gunning R, Pahita E, Adams S, Adams D, Ferguson-Smith AC, Identification, Characterization, and Heritability of Murine Metastable Epialleles: Implications for Non-genetic Inheritance, Cell 175(5) (2018) 1259–1271 e13. [PubMed: 30454646]
- [191]. Dolinoy DC, Weidman JR, Waterland RA, Jirtle RL, Maternal genistein alters coat color and protects Avy mouse offspring from obesity by modifying the fetal epigenome, Environ Health Perspect 114(4) (2006) 567–72. [PubMed: 16581547]
- [192]. Wolff GL, Kodell RL, Moore SR, Cooney CA, Maternal epigenetics and methyl supplements affect agouti gene expression in Avy/a mice, FASEB J 12(11) (1998) 949–57. [PubMed: 9707167]
- [193]. Cooney CA, Dave AA, Wolff GL, Maternal methyl supplements in mice affect epigenetic variation and DNA methylation of offspring, J Nutr 132(8 Suppl) (2002) 2393S–2400S. [PubMed: 12163699]
- [194]. Kaminen-Ahola N, Ahola A, Maga M, Mallitt KA, Fahey P, Cox TC, Whitelaw E, Chong S., Maternal ethanol consumption alters the epigenotype and the phenotype of offspring in a mouse model, PLoS Genet 6(1) (2010) e1000811. [PubMed: 20084100]
- [195]. Dunn DB, Smith JD, Occurrence of a new base in the deoxyribonucleic acid of a strain of Bacterium coli, Nature 175(4451) (1955) 336–7. [PubMed: 13235889]
- [196]. Dunn DB, Smith JD, The occurrence of 6-methylaminopurine in deoxyribonucleic acids, Biochem J 68(4) (1958) 627–36. [PubMed: 13522672]
- [197]. O'Brown ZK, Greer EL, N6-Methyladenine: A Conserved and Dynamic DNA Mark, Adv Exp Med Biol 945 (2016) 213–246. [PubMed: 27826841]
- [198]. Greer EL, Blanco MA, Gu L, Sendinc E, Liu J, Aristizabal-Corrales D, Hsu CH, Aravind L, He C, Shi Y, DNA Methylation on N(6)-Adenine in C. elegans, Cell 161(4) (2015) 868–78. [PubMed: 25936839]
- [199]. Zhang G, Huang H, Liu D, Cheng Y, Liu X, Zhang W, Yin R, Zhang D, Zhang P, Liu J, Li C, Liu B, Luo Y, Zhu Y, Zhang N, He S, He C, Wang H, Chen D, N(6)-methyladenine DNA modification in Drosophila, Cell 161(4) (2015) 893–906. [PubMed: 25936838]
- [200]. Koziol MJ, Bradshaw CR, Allen GE, Costa AS, Frezza C, Gurdon JB, Identification of methylated deoxyadenosines in vertebrates reveals diversity in DNA modifications, Nature structural & molecular biology 23(1) (2016) 24–30.
- [201]. Wu TP, Wang T, Seetin MG, Lai Y, Zhu S, Lin K, Liu Y, Byrum SD, Mackintosh SG, Zhong M, Tackett A, Wang G, Hon LS, Fang G, Swenberg JA, Xiao AZ, DNA methylation on N-adenine in mammalian embryonic stem cells, Nature (2016).
- [202]. Liu J, Zhu Y, Luo GZ, Wang X, Yue Y, Wang X, Zong X, Chen K, Yin H, Fu Y, Han D, Wang Y, Chen D, He C., Abundant DNA 6mA methylation during early embryogenesis of zebrafish and pig, Nat Commun 7 (2016) 13052. [PubMed: 27713410]

- [203]. Liang Z, Shen L, Cui X, Bao S, Geng Y, Yu G, Liang F, Xie S, Lu T, Gu X, Yu H., DNA N(6)-Adenine Methylation in Arabidopsis thaliana, Dev Cell 45(3) (2018) 406–416 e3. [PubMed: 29656930]
- [204]. Hao Z, Wu T, Cui X, Zhu P, Tan C, Dou X, Hsu KW, Lin YT, Peng PH, Zhang LS, Gao Y, Hu L, Sun HL, Zhu A, Liu J, Wu KJ, He C., N(6)-Deoxyadenosine Methylation in Mammalian Mitochondrial DNA, Molecular cell 78(3) (2020) 382–395 e8. [PubMed: 32183942]
- [205]. Schiffers S, Ebert C, Rahimoff R, Kosmatchev O, Steinbacher J, Bohne AV, Spada F, Michalakis S, Nickelsen J, Muller M, Carell T., Quantitative LC-MS Provides No Evidence for m(6) dA or m(4) dC in the Genome of Mouse Embryonic Stem Cells and Tissues, Angew Chem Int Ed Engl 56(37) (2017) 11268–11271. [PubMed: 28371147]
- [206]. Liu B, Liu X, Lai W, Wang H., Metabolically Generated Stable Isotope-Labeled Deoxynucleoside Code for Tracing DNA N(6)-Methyladenine in Human Cells, Anal Chem 89(11) (2017) 6202–6209. [PubMed: 28471639]
- [207]. O'Brown ZK, Boulias K, Wang J, Wang SY, O'Brown NM, Hao Z, Shibuya H, Fady PE, Shi Y, He C, Megason SG, Liu T, Greer EL, Sources of artifact in measurements of 6mA and 4mC abundance in eukaryotic genomic DNA, BMC Genomics 20(1) (2019) 445. [PubMed: 31159718]
- [208]. Douvlataniotis K, Bensberg M, Lentini A, Gylemo B, Nestor CE, No evidence for DNA N (6)-methyladenine in mammals, Sci Adv 6(12) (2020) eaay3335. [PubMed: 32206710]
- [209]. Wang SY, Mao H, Shibuya H, Uzawa S, O'Brown ZK, Wesenberg S, Shin N, Saito TT, Gao J, Meyer BJ, Colaiacovo MP, Greer EL, The demethylase NMAD-1 regulates DNA replication and repair in the Caenorhabditis elegans germline, PLoS Genet 15(7) (2019) e1008252. [PubMed: 31283754]
- [210]. Wan QL, Meng X, Dai W, Luo Z, Wang C, Fu X, Yang J, Ye Q, Zhou Q., N(6)methyldeoxyadenine and histone methylation mediate transgenerational survival advantages induced by hormetic heat stress, Sci Adv 7(1) (2021).
- [211]. Martienssen R, Barkan A, Taylor WC, Freeling M., Somatically heritable switches in the DNA modification of Mu transposable elements monitored with a suppressible mutant in maize, Genes & development 4(3) (1990) 331–43. [PubMed: 2159936]
- [212]. Martienssen R, Baron A., Coordinate suppression of mutations caused by Robertson's mutator transposons in maize, Genetics 136(3) (1994) 1157–70. [PubMed: 8005422]
- [213]. Cubas P, Vincent C, Coen E., An epigenetic mutation responsible for natural variation in floral symmetry, Nature 401(6749) (1999) 157–61. [PubMed: 10490023]
- [214]. Mirouze M, Reinders J, Bucher E, Nishimura T, Schneeberger K, Ossowski S, Cao J, Weigel D, Paszkowski J, Mathieu O., Selective epigenetic control of retrotransposition in Arabidopsis, Nature 461(7262) (2009) 427–30. [PubMed: 19734882]
- [215]. Johannes F, Porcher E, Teixeira FK, Saliba-Colombani V, Simon M, Agier N, Bulski A, Albuisson J, Heredia F, Audigier P, Bouchez D, Dillmann C, Guerche P, Hospital F, Colot V., Assessing the impact of transgenerational epigenetic variation on complex traits, PLoS Genet 5(6) (2009) e1000530. [PubMed: 19557164]
- [216]. Becker C, Hagmann J, Muller J, Koenig D, Stegle O, Borgwardt K, Weigel D., Spontaneous epigenetic variation in the Arabidopsis thaliana methylome, Nature 480(7376) (2011) 245–9. [PubMed: 22057020]
- [217]. Schmitz RJ, Schultz MD, Lewsey MG, O'Malley RC, Urich MA, Libiger O, Schork NJ, Ecker JR, Transgenerational epigenetic instability is a source of novel methylation variants, Science (New York, N.Y 334(6054) (2011) 369–73. [PubMed: 21921155]
- [218]. Cortijo S, Wardenaar R, Colome-Tatche M, Gilly A, Etcheverry M, Labadie K, Caillieux E, Hospital F, Aury JM, Wincker P, Roudier F, Jansen RC, Colot V, Johannes F., Mapping the epigenetic basis of complex traits, Science (New York, N.Y 343(6175) (2014) 1145–8. [PubMed: 24505129]
- [219]. Calarco JP, Borges F, Donoghue MT, Van Ex F, Jullien PE, Lopes T, Gardner R, Berger F, Feijo JA, Becker JD, Martienssen RA, Reprogramming of DNA methylation in pollen guides epigenetic inheritance via small RNA, Cell 151(1) (2012) 194–205. [PubMed: 23000270]

- [220]. Schott DH, Cureton DK, Whelan SP, Hunter CP, An antiviral role for the RNA interference machinery in Caenorhabditis elegans, Proceedings of the National Academy of Sciences of the United States of America 102(51) (2005) 18420–4. [PubMed: 16339901]
- [221]. Ermolaeva MA, Schumacher B., Insights from the worm: the C. elegans model for innate immunity, Semin Immunol 26(4) (2014) 303–9. [PubMed: 24856329]
- [222]. Zilberman D, Cao X, Jacobsen SE, ARGONAUTE4 control of locus-specific siRNA accumulation and DNA and histone methylation, Science (New York, N.Y 299(5607) (2003) 716–9. [PubMed: 12522258]
- [223]. Noma K, Sugiyama T, Cam H, Verdel A, Zofall M, Jia S, Moazed D, Grewal SI, RITS acts in cis to promote RNA interference-mediated transcriptional and post-transcriptional silencing, Nature genetics 36(11) (2004) 1174–80. [PubMed: 15475954]
- [224]. Verdel A, Jia S, Gerber S, Sugiyama T, Gygi S, Grewal SI, Moazed D., RNAi-mediated targeting of heterochromatin by the RITS complex, Science (New York, N.Y 303(5658) (2004) 672–6. [PubMed: 14704433]
- [225]. Fukagawa T, Nogami M, Yoshikawa M, Ikeno M, Okazaki T, Takami Y, Nakayama T, Oshimura M., Dicer is essential for formation of the heterochromatin structure in vertebrate cells, Nat Cell Biol 6(8) (2004) 784–91. [PubMed: 15247924]
- [226]. Sugiyama T, Cam H, Verdel A, Moazed D, Grewal SI, RNA-dependent RNA polymerase is an essential component of a self-enforcing loop coupling heterochromatin assembly to siRNA production, Proceedings of the National Academy of Sciences of the United States of America 102(1) (2005) 152–7. [PubMed: 15615848]
- [227]. Ogawa Y, Sun BK, Lee JT, Intersection of the RNA interference and X-inactivation pathways, Science (New York, N.Y 320(5881) (2008) 1336–41. [PubMed: 18535243]
- [228]. Moazed D., Mechanisms for the inheritance of chromatin States, Cell 146(4) (2011) 510–8.[PubMed: 21854979]
- [229]. Cernilogar FM, Onorati MC, Kothe GO, Burroughs AM, Parsi KM, Breiling A, Lo Sardo F, Saxena A, Miyoshi K, Siomi H, Siomi MC, Carninci P, Gilmour DS, Corona DF, Orlando V., Chromatin-associated RNA interference components contribute to transcriptional regulation in Drosophila, Nature 480(7377) (2011) 391–5. [PubMed: 22056986]
- [230]. Mao H, Zhu C, Zong D, Weng C, Yang X, Huang H, Liu D, Feng X, Guang S., The Nrde Pathway Mediates Small-RNA-Directed Histone H3 Lysine 27 Trimethylation in Caenorhabditis elegans, Curr Biol 25(18) (2015) 2398–403. [PubMed: 26365259]
- [231]. Yu R, Wang X, Moazed D., Epigenetic inheritance mediated by coupling of RNAi and histone H3K9 methylation, Nature 558(7711) (2018) 615–619. [PubMed: 29925950]
- [232]. Fuks F, Hurd PJ, Deplus R, Kouzarides T., The DNA methyltransferases associate with HP1 and the SUV39H1 histone methyltransferase, Nucleic Acids Res 31(9) (2003) 2305–12. [PubMed: 12711675]
- [233]. Bartke T, Vermeulen M, Xhemalce B, Robson SC, Mann M, Kouzarides T., Nucleosomeinteracting proteins regulated by DNA and histone methylation, Cell 143(3) (2010) 470–84. [PubMed: 21029866]
- [234]. Johnson LM, Bostick M, Zhang X, Kraft E, Henderson I, Callis J, Jacobsen SE, The SRA methyl-cytosine-binding domain links DNA and histone methylation, Current biology : CB 17(4) (2007) 379–84. [PubMed: 17239600]
- [235]. Rajakumara E, Wang Z, Ma H, Hu L, Chen H, Lin Y, Guo R, Wu F, Li H, Lan F, Shi YG, Xu Y, Patel DJ, Shi Y., PHD finger recognition of unmodified histone H3R2 links UHRF1 to regulation of euchromatic gene expression, Molecular cell 43(2) (2011) 275–84. [PubMed: 21777816]
- [236]. Fuks F., DNA methylation and histone modifications: teaming up to silence genes, Current opinion in genetics & development 15(5) (2005) 490–5. [PubMed: 16098738]
- [237]. Rountree MR, Selker EU, DNA methylation and the formation of heterochromatin in Neurospora crassa, Heredity (Edinb) 105(1) (2010) 38–44. [PubMed: 20407471]
- [238]. Peng D, Wang C, Li K-L, Gan Z-X, Li Y-H, Wang H-W, Li Q-Y, Liu X-W, Sun H-Y, Jing Y-Y, Fang Q, Zhao Q, Zhang L, Chen H-H, Wei H-M, Sun J, Tang H-Y, Yang X-M, Chang J-F, Sun F, Jiang C-Z, Yuan H-B, Li W, Sun F-L, The Establishment of Transgenerational Epigenetic

Inheritance in the C. elegans Germline is Mediated by Lipid Metabolism, bioRxiv (2021) doi: 10.1101/2020.11.04.367854

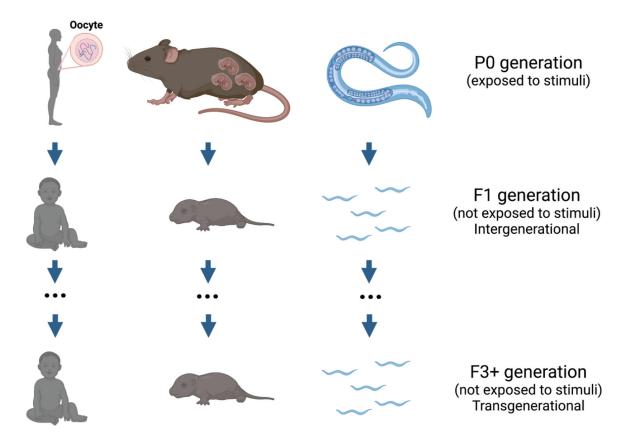


Figure 1: Distinction between inter and transgenerational phenotypes

Numerous different parental (P0) stresses can have multigenerational effects on offspring. Intergenerational effects represent any effect of parental stress on F1 progeny that either directly acts on or is communicated through P0 germ cells or developing F1 embryos *in utero*. By comparison, all effects that are initiated in the P0 generation and persist into the F3 (or later) generations are transgenerational effects. Effects that are initiated in the P0 generation and persist to the F2 generation are intergenerational if any germ cells of F1 animals have formed *in utero* when the initiating event/stress was present and transgenerational if no F1 germ cells have formed. These original distinctions between intergenerational and transgenerational effects in F2 progeny are still used as definitions in the literature irrespective of the mechanisms that mediate multigenerational effects in progeny, including cases where such effects might not be transmitted via germ cells. Figure created with BioRender.com.