RESEARCH ARTICLE

Long non‑coding RNA SOS1‑IT1 promotes endometrial cancer progression by regulating hypoxia signaling pathway

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Abstract

Endometrial cancer (EC) is one of the most common types of gynecological cancer. Hypoxia is an important clinical feature and regulates various tumor processes. However, the prognostic value of hypoxia-related lncRNA in EC remains to be further elucidated. Here, we aimed to characterize the molecular features of EC by the development of a classifcation system based on the expression profle of hypoxia-related lncRNA. Based on univariate Cox regression analysis, we identifed 17 hypoxiarelated lncRNAs signifcantly associated with overall survival. Next, the least absolute shrinkage and selection operator Cox regression model was utilized to construct a multigene signature in the TCGA EC cohort. The risk score was confrmed as an independent predictor for overall survival in multivariate Cox regression analysis and receiver operating characteristic (ROC) curve analysis. Besides, the survival time of EC patients in diferent risk group was signifcantly correlated to clinicopathologic factors, such as age, stage and grade. Furthermore, hypoxia-related lncRNA associated with the high-risk group were involved in various aspects of the malignant progression of EC via Gene Ontology, Kyoto Encyclopedia of Genes and Genomes pathway, and Gene Set Enrichment Analysis. Moreover, the risk score was closely correlated to immunotherapy response, microsatellite instability and tumor mutation burden. Finally, we select one hypoxia-related lncRNA SOS1-IT1 to validate its role in hypoxia and EC progression. Interestingly, we found SOS1-IT1 was overexpressed in tumor tissues, and closely correlated with clinicopathological parameters of EC. The expression level of SOS1-IT1 was signifcantly increased under hypoxia condition. Additionally, the important hypoxia regulatory factor HIF-1 α can directly bind SOS1-IT1 promoter region, and afect its expression level. In summary, this study established a new EC classifcation based on the hypoxiarelated lncRNA signature, thereby provide a novel sight to understand the potential mechanism of human EC development.

Keywords Endometrial cancer · SOS1-IT1 · Hypoxia · lncRNA · Prognostic prediction

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Abbreviations

- EC Endometrial cancer
- OS Overall survival
- ROC Receiver operating characteristic
- GESA Gene set enrichment analysis
- GO Gene ontology
- KEEG Kyoto encyclopedia of genes and genomes

Introduction

Endometrial cancer (EC) is one of the most common gynecological cancer in developed countries (Raglan et al. [2019](#page-17-0); Siegel et al. 2020).^{[1](#page-1-0)} EC has been considered as a global threat to women's health and well-being. Most EC patients are diagnosed at an early stage with a favorable prognosis. However, 20–30% of EC patients were diagnosed with a high-risk of recurrence and poor prognosis. EC patients often showed diferent prognoses and treatment responses, even if with the same degree of progression (Habiba et al. [2018](#page-17-2); McDonald and Bender [2019;](#page-17-3) Tsikouras et al. [2013](#page-17-4)). Therefore, effective EC biomarkers must be found to assess prognosis and identify EC patients with high-risk.

Hypoxia is a common phenomenon in solid tumors. Hypoxia is harmful to cancer cells, but it can promote cancer cells to adapt, thus promotes malignant progression (Jing et al. [2019](#page-17-5)). In the hypoxic microenvironment, tumor cells exhibit activating downstream genes to facilitate tumor growth, with strong aggressiveness and metastasis ability (Yang et al. [2020;](#page-17-6) Yuen and Wong [2020\)](#page-17-7). In response to hypoxia, the major component of hypoxia signaling pathways is hypoxia inducible factors-1 (HIF-1), which is a heterodimer composed of α and β subunits (Choudhry and Harris [2018](#page-17-8); Hajizadeh et al. [2019](#page-17-9)). HIF-1α mainly interacts with HIF-1 β subunit to form a stable HIF-1 complex in the nucleus. Under hypoxic conditions, the HIF-1 complex binds to the target gene's promoter to induce their transcription. Most of the HIF-1 complex dependent genes were associated with proliferation, epithelial to mesenchymal transition, angiogenesis, and metastasis (Albadari et al. [2019](#page-17-10); Semenza [2013](#page-17-11); Soni and Padwad [2017](#page-17-12)). However, due to the lack of efective biomarkers, how hypoxia leads to tumor progression remain to be further elucidated.

Long non coding RNAs (lncRNAs) are a large class of heterogeneous transcripts with a length of more than 200 nucleotides, and their protein coding potential are limited. LncRNAs are widely transcribed in eukaryotic genomes, suggesting their important regulatory role in complex organisms (Bhan et al. [2017;](#page-17-13) Peng et al. [2017](#page-17-14)). Although only a small number of functional lncRNAs have been well characterized, more and more evidence showed that lncRNAs play a key role in controlling a large number of cancer-related cell processes, such as proliferation, migration, invasion, autophagy and stemness (Ding et al. [2021;](#page-17-15) Liu et al. [2019a,](#page-17-16) [2020](#page-17-17); Wang et al. [2020a\)](#page-17-18). A specifc group of lncRNAs are regulated by tumor microenvironment, such as hypoxia. The hypoxia responsive lncRNAs, such as NORAD,

RAB11B-AS1 and LncHIFCAR, may be the basis of cancer cell survival and disease progression (Li et al. [2017](#page-17-19); Niu et al. [2020;](#page-17-20) Shih et al. [2017\)](#page-17-21). MALAT1 was the frst reported lncRNA mediating hypoxia-induced pro-survival autophagy in endometriosis (Liu et al. [2019b\)](#page-17-22). However, there are few reports about the relationship between hypoxia related lncRNA and EC in recent years. Whether lncRNAs are involved in the response to hypoxia in EC, and the prognostic value of hypoxia-related lncRNA in EC remains to be further elucidated.

In this study, we frst downloaded the RNA expression profles of hypoxia-related lncRNA and corresponding clinical data of EC patients from public databases. The diferent lncRNA expression patterns detected among EC cases were identifed the candidate lncRNA biomarkers based on The Cancer Genome Atlas data (TCGA). Besides, we found that the hypoxia-related lncRNA could classify the EC patients with significantly different clinical and molecular characteristics. Finally, we focused on lncRNA SOS1-IT1, and found it was signifcantly upregulated in EC cells under hypoxic conditions. We evaluated its biological role and clinical signifcance in EC progression and revealed SOS1-IT1 is hypoxia-inducible and directly transactivated by HIF-1α.

Materials and methods

Data collection

The TCGA RNA sequencing dataset and corresponding clinicopathological characteristics information, such as age, grade, stage, radiation therapy, surgical approach and survival information were downloaded from TCGA database (<http://www.cancergenome.nih.gov/>). RNA-seq data from 550 tumor tissue samples and 35 normal samples were downloaded from TCGA. A total of 486 EC patients were included in the cluster and risk signature study by eliminating cases with missing complete clinicopathological characteristics information. As suggested, we have added the detail information of EC patient cohort in Table [1](#page-2-0).

Identifcation of diferentially expressed hypoxia‑related lncRNA

Based on the Spearman correlation analysis, lncRNAs related to the hypoxia were identified (Filter: $|r| > 0.5$ and *P*<0.001). The prognosis-related lncRNAs were screened by K–M survival analyses $(P < 0.01)$. Differentially expressed hypoxia-related lncRNA were identifed in EC from the TCGA datasets by using the EdgeR package in R statistical software. The signifcance criteria for determining diferentially expressed lncRNA were set as adjusted

¹ A non-peer reviewed version of this paper is available on the research square preprint server ([https://www.researchsquare.com/artic](https://www.researchsquare.com/article/rs-633056/v1) [le/rs-633056/v1\)](https://www.researchsquare.com/article/rs-633056/v1).

 P value < 0.05. Heatmaps were plotted by the "pheatmap" package.

Comprehensive analysis of interaction network

According to the correlation efficiency and probability cutoff value (Filter: $|r| > 0.5$ and $P < 0.001$), 677 lncRNAs were screened out as hypoxia-related lncRNAs. Then we explored the interaction network between hypoxia and hypoxia-related lncRNAs, by using the Cytoscape software.

Consensus clustering of hypoxia‑related lncRNA

Consensus clustering was utilized to separate EC patients into subgroups according to the expression of hypoxiarelated lncRNA. We classifed EC patients into diferent clusters using the "ConsensusClusterPlus" package in R statistical software. The cumulative distribution function (CDF) and consensus heatmap were used to evaluate the optimal k value. Due to the grouping was suboptimal when they were divided into more than 2 clusters, we divided the hypoxiarelated lncRNA into two groups based on their expression indices using $k=2$ as the optimal value.

The following shows the full code.

library(ConsensusClusterPlus) results = ConsensusClusterPlus(data, maxK=maxK, reps=50, pItem=0.8, pFeature=1, title=workDir, clusterAlg="km", distance="euclidean", seed=123456, plot="png") clusterNum=2 cluster=results[[clusterNum]][["consensusClass"]] write.table(cluster,file="cluster.txt",sep="\t",quote=F,col.names=F)

maxK=9

Generation of hypoxia‑related lncRNA signature

The expression data of hypoxia-related lncRNA in 550 EC and 35 normal tissues were analyzed with the Limma package and visualized as a heatmap. The "Glmnet" and "Survival" packages were used to perform LASSO regression analysis (Tibshirani [1997\)](#page-17-23). We also performed the univariate and multivariate Cox hazard analysis of clinicopathological characteristics information by "survival" package.

Construction of hypoxia‑related lncRNA prognostic model

Prognosis-related lncRNA were constructed using multivariate cox regression. The risk score of each patient was calculated according to the predictive signature model. Using the median risk score as the cutoff point, EC patients were divided into the high-risk group and low-risk group. The prediction efficiency of the risk signature model was evaluated by the receiver operating characteristic (ROC) curve using R package survival ROC.

Functional enrichment analysis of the hypoxia‑related lncRNA

Functional enrichment analysis of the hypoxia-related lncRNA was conducted using DAVID, including biological **Fig. 1** The fow chart of the identifcation of EC hypoxiarelated lncRNA signature

functions, cellular components, and molecular functions. The Kyoto Encyclopedia of Genes and Genomes (KEGG) database was searched for signifcant pathways. We also performed Gene set enrichment analysis (GSEA) to identify the signifcantly alerted KEGG pathways between the high-risk and low-risk groups. The Java GSEA program were used and the gene set from Molecular Signatures Database was selected as the reference gene set. Biological processes with normalized P value < 0.05 and false discovery rate (FDR) q value < 0.05 were considered statistically significant.

Evaluation of tumor‑infltrating immune cells and the immune infltration level

The subpopulation of 22 immune cells in each tumor sample was explored by CIBERSORT algorithm in diferent clusters. The CIBERSORT results of samples with *P*<0.05 showed that the inferred fractions of immune cell populations were accurate and could be further analyzed. Furthermore, we analyzed the correlation between tumor immune cell infltrating and prognostic risk signature.

Tumor mutation burden assessment

The tumor mutation burden (TMB) for each organization was detected by the VarScan method, as calculated using the R package "maftools". According to the cutoff value of median TMB data, EC Patients were divided into high TMB group and low TMB group. We also analyzed the correlation between TMB and prognostic risk signature.

Cell culture

Human endometrial cancer cell lines Ishikawa was purchased from ATCC (American Type Culture Collection). The cells were grown in DMEM supplemented with 10% fetal bovine serum (FBS) and cultured at 37 °C with 5% $CO₂$. To assess the efficacy of hypoxia, EC cells were cultured under normoxia conditions to 70% confuence. Then the cells were cultured under consistent 1% O₂ hypoxia condition for 24 h. The concentration of $CoCl₂$ used in this study was 100 μM.

Gene knockdown and overexpression experiments

The knockdown of HIF-1 α in EC cells was performed using siRNA. The siRNA and negative control siRNA were purchased from RiboBio Co., Ltd (Guangzhou, China). The knockdown of SOS1-IT1 in EC cells was performed with lentiviruses carrying control shRNA or SOS1-IT1 shRNA, purchased from Genechem Co., Ltd. (Shanghai, China). The Flag-tagged SOS1-IT1 plasmid was transfected into EC cell

Typ

Fig. 2 Identifcation hypoxia-related lncRNA in EC. **a** The univariate Cox regression analysis of 17 hypoxia-related lncRNAs in EC. **b**, **c** The expression heatmaps (**b**) and box plot (**c**) of 17 hypoxia-

line Ishikawa cells, for SOS1-IT1 overexpression experiments. The Flag-tagged SOS1-IT1 expression plasmid was constructed by subcloning the SOS1-IT1 full-length complementary DNA (cDNA) fragments into $3 \times$ flag vector, and $3 \times$ flag empty vector was used as the relative control. All constructs were confrmed by DNA sequencing.

Total RNA extraction and REAL‑time PCR

The total RNA was extracted by TRIZOL reagent (Invitrogen) according to the manufacturer's instruction. The cDNA

related lncRNAs in diferent EC tumor and normal tissue samples. * *P*<0.05, ** *P*<0.01, ****P*<0.001

was obtained from the purifed RNA using a PrimeScript RT Reagent Kit (Takara). Real-time PCR was performed using the SYBR Premix Ex Taq (Takara) following the manufacturer's instructions. The results were normalized to β-actin gene.

Cell proliferation assays

The cell proliferation assays were determined using a commercial WST-1 assay kit according to the manufacturer's instructions as described before (Wan et al. [2018](#page-17-24) [2019a,](#page-17-25)

Fig. 3 Classifcation of EC based on hypoxia-related lncRNA. (**a**) ◂ The cumulative distribution function (CDF) plots show the cumulative distributive functions of the consensus matrix for k values (indicated by diferent colors) between 2 and 9. **b**, **c** Relative change in area under CDF curve (**b**) and tracking plot (**c**) according to various k values. (**d**) The heatmaps show the consensus clustering matrix for hypoxia-related lncRNA in the EC dataset for k=2. **e** Heatmap shows the association of clinic pathological features based on the two clusters. **f** The survival analysis of EC patients between cluster 1 and 2 in TCGA cohort

[b](#page-17-26)). First, 15 μl WST-1 reagent was added to the cells in the 96-well plates, and incubated for 2 h. Later, the OD value was measured in a microplate reader at 450 nm.

Dual‑luciferase reporter assays

For the luciferase reporter experiment, the SOS1-IT1 wild type (WT) or mutant vectors were used. The Ishikawa cells were seeded in six-well plates and transfected with relative plasmids or siRNA for 48 h. The Dual-Luciferase Reporter Assay System was used to assess the activities of frefy luciferase and Renilla luciferase sequentially. Then the relative luciferase activities were calculated, and control cells were used for normalization.

Statistical analysis

Diferences between survival curves were generated by the Kaplan–Meier method and compared by log-rank tests. The multivariate analysis was performed using the cox proportional hazard model. For comparisons of two groups, a t-test was used. R studio package was used for all statistical analysis. All statistical tests were only considered statistically significant when $p < 0.05$ was achieved.

Results

Identifcation hypoxia‑related lncRNA in EC

To better understand the important role of hypoxiarelated lncRNA in oncogenesis and progression, we frst downloaded from the hallmark gene sets of hypoxia including 200 genes from Molecular Signatures Database. RNA-seq data from 550 tumor tissue samples and 35 normal samples were downloaded from TCGA. A flow chart was developed to systematically describe our study $(Fig. 1)$ $(Fig. 1)$. According to the correlation efficiency and probability cut-off value (Filter: $|r| > 0.5$ and $P < 0.001$), 677 lncRNAs were screened out as hypoxia-related lncRNA. In order to investigate the prognostic value of these hypoxiarelated lncRNA in EC, univariate Cox regression analysis was performed based on the expression levels of these lncRNA in TCGA database. As a result, we found that 17 out of the 677 lncRNAs were signifcantly associated with overall survival $(p < 0.05)$ (Fig. [2](#page-4-0)a). The expression values of 17 hypoxia-related lncRNAs were extracted from EC patients. The expression profle of 17 prognostic associated hypoxia-related lncRNA was showed in the heatmap and box plot (Fig. [2](#page-4-0)b, c). As shown in Fig. [1](#page-3-0)c, 17 hypoxiarelated lncRNAs were signifcantly abnormally expressed in EC tissues samples. Compared to normal tissue samples, lncRNAs AL021578.1, AP000593.3, LINC01843, SOS1- IT1 and NRAV were up-regulated, while AC079921.1, AC119674.1, AC114947.2, AC011466.1, CFAP58-DT, LINC01614, AC244517.7, TPM1-AS, AL035530.2, AC007786.2, AP003096.1 and AC020910.5 were downregulated in EC tumor tissues. In addition, based on the co-expression relationship, we also built an interaction network based on hypoxia-related lncRNAs and the corresponding hypoxia associated genes (Fig. [2](#page-4-0)d).

Classifcation of EC based on hypoxia‑related lncRNA

In order to analysis the consensus cluster of hypoxia-related lncRNA in EC, we used the common clusterplus package to identify the diferent groups of hypoxia-related lncRNA based on their co-expression patterns in EC tissues from the TCGA database. Due to the grouping was suboptimal when they were divided into more than 2 clusters, we divided the hypoxia-related lncRNA into two groups based on their expression indices using $k=2$ as the optimal value (Fig. [3a](#page-6-0)–d). Next, we analyzed the relationship between

Fig. 4 Prognostic value of hypoxia-related lncRNA and construction ◂ of a risk signature predicting prognosis. **a** The risk scores for all EC patients in entire cohort are plotted in ascending order, and marked as high-risk (red) and low-risk (blue), as divided by the threshold. **b** Survival days of EC patients were showed in ascending order of risk parameters. **c**, **d** The forest plot of univariate (**c**) and multivariate (**d**) Cox regression analysis in the cohorts. **e** Heatmap shows the association of risk scores and clinic pathological features based on the six lncRNA risk signature. **f** The Kaplan–Meier curves of OS in the highrisk and the low-risk groups stratifed by the hypoxia-related lncRNA signature in the cohorts. **g** The ROC analysis of OS for the signature and the clinicopathologic parameters. **h** The nomogram for predicting proportion of patients with 1-, 3‐ or 5‐year OS

these two clusters and the clinicopathological characteristics of EC patients. We found that consensus clustering could make signifcant diferences in the clinical and molecular characteristics of the two EC clusters (Fig. [3e](#page-6-0)). Cluster 1 patients were strikingly correlated with stage, grade, fustat and disease type by Chi-square test. Besides, compared with patients in cluster 2, EC patients in cluster 1 showed a shorter overall survival time (Fig. [3](#page-6-0)f).

Prognostic value of hypoxia‑related lncRNA and construction of a risk signature predicting prognosis

To identifed the most powerful prognostic hypoxia-related lncRNAs, the last absolute shrinkage and selection operator (LASSO) Cox regression analysis to the 17 prognosis-related lncRNAs was conducted. We constructed prognostic models using the multivariate Cox proportional hazards regression analysis and the coefficient of each independent prognostic gene were calculated. The risk score was estimated based on the coefficients from the LASSO results. According to the median risk score, EC patients were assigned into low-risk and high-risk groups. The distribution of the hypoxia-related risk signature in the TCGA dataset and survival status of EC patients in diferent groups were shown in Fig. [4a](#page-8-0), b. After adjusting for clinicopathological features such as age, grade, stage, radiation therapy, surgical approach, and disease type, we found that age, grade, stage, disease type and risk score were correlated with the OS of EC patients in univariate analysis, while multivariate COX regression analysis showed that age, grade, stage, radiation therapy, surgical approach and risk score were independent risk factors for the prognosis of EC patients (Fig. [4](#page-8-0)c, d, Table [2\)](#page-9-0).

The heatmap of these most signifcantly six lncRNAs were shown in Fig. [4e](#page-8-0), and we observed a strong correlation between the risk score and the clinicopathological characteristics such as stage, age and grade. The result of survival analysis showed that the high-risk group had signifcantly shorter survival time compared to low-risk group (Fig. [4f](#page-8-0)). The AUC value of the ROC curve in risk score for 5-year survival is 0.691, which is obviously higher than that of ROC in grade (0.662), radiation therapy (0.371), surgical approach (0.461) and disease type (0.569), but lower than stage (0.702) (Fig. [4](#page-8-0)g). In addition, the calibration plot revealed ideal agreements of these genes between the actual observations and 1-, 3- and 5-year OS predicted by the nomogram (Fig. [4h](#page-8-0)). In addition, we divided the EC cases in TCGA cohort with clinicopathological characteristics information into training and validation groups randomly (Figure S1). Consistent with the result in entire TCGA cohort, the result of survival analysis showed that the high-risk group had signifcantly shorter survival time compared to low-risk group, both in the training and validation. Besides, the AUC value of the ROC curve in risk score for 5-year survival is 0.720 and 0.653, in the training and validation cohort, respectively, which indicate its efectiveness as a prognostic biomarker in predicting overall survival of EC patients. These results indicated that the prognostic index based on hypoxia-related lncRNA has the potential to predict the survival of EC patients.

Relationships between the risk score and clinicopathologic factors

Next, we built a complete prognostic model based on the entire set. The stratification analysis was done according to the age, grade, stage, radiation therapy and surgical approach. EC patients were stratified into age ≤ 60 and > 60 subgroup, grade G1–G2 and G3–G4 subgroup, stage I–II and III–IV subgroup, surgical approach minimally invasive and open subgroup, radiation therapy No and Yes subgroup. For the EC patients in age>60 subgroup, grade G3–G4 subgroup, and stage III-IV subgroup, the survival time of patients was signifcantly shorter than that of patients in another group (Fig. [5](#page-10-0)a), and the average risk score was much higher (Fig. [5](#page-10-0)d, e). However, there is no diference between surgical approach and radiation therapy subgroup.

Table 2 Univariate and multivariate Cox regression analyses of hypoxia-related lncRNA risk signature

Besides, we found the survival time of patients in high-risk group was signifcantly shorter in age>60 subgroup, grade G3-G4 subgroup, stage III-IV subgroup, surgical approach minimally invasive subgroup, and radiation therapy No subgroup (Fig. $5b$, c).

Biological characteristics and pathway analysis of hypoxia‑related risky lncRNA

The EC patients in TCGA were divided into high-risk and low-risk groups based on the median risk score. Then, the lncRNA significantly upregulated (fold change> 1 and $P < 0.05$) or downregulated (fold change <-1 and $P < 0.05$) were selected for Gene Ontology (GO), Kyoto Encyclopedia of Genes and Genomes (KEEG) pathway, and Gene Set Enrichment Analysis (GESA). To elucidate the biological functions and pathways that were associated with the risk score, the hypoxia-related lncRNA between the high-risk and low-risk groups were used to perform GO enrichment and KEGG pathway analyses (Fig. [6](#page-11-0)a–d). As expected, hypoxia-related lncRNAs were enriched in several cancerrelated biological pathways, such as membrane potential regulation, cell–cell adhesion, and protein digestion signaling pathway, et al. The GSEA analysis results showed that diferent risk group was involved in multiple signifcantly enriched pathways, including cell cycle, wnt signaling, spliceosome, et al. (Fig. [6e](#page-11-0)). These results showed the two-risk group identifed based on the six hypoxia-related risky lncR-NAs were closely associated with the malignancy of EC.

The risk score of hypoxia‑related lncRNA was associated with immunotherapy, microsatellite instability and tumor mutation burden in EC

The combination of anti-CTLA-4 and anti-PD-1 treatment could increase the proportion of activated CD8+cells and natural killer cells in the tumor microenvironment, and decrease the proportion of inhibitory immune cells, resulting in changes in the immune landscape, which achieved therapeutic efect in the mouse model and prolonged the tumor free survival (Boutros et al. [2016](#page-17-27)). After analyzing the difference in response to immunotherapy between diferent risk score group. We found that low-risk group tended to respond efectively to immunotherapy such as anti-PD-1 and anti-CTLA-4 therapy (Fig. [7a](#page-12-0)–d). High microsatellite instability (MSI-H) is associated with the response to immunotherapy treatment. Interestingly, we found that the hypoxia-related lncRNA risk score of EC patients with MSI-H was lower than that of EC patients with low MSI or microsatellite stability (MSS) (Fig. [7](#page-12-0)e, [f\)](#page-12-0). Tumor genome mutation leads to the production of new antigens that is benefcial to immunotherapy. Tumor mutation burden (TMB) is an important biomarker, which can be used to predict the response of a variety of tumors to PD-1/PD-L1 targeted immunotherapy. The box plot showed that there was a diference in mutation frequency between diferent risk score groups (Fig. [7](#page-12-0)g). We observed that the risk score was negatively correlated with TMB in correlation analysis $(R=-0.23, P=1.2e-07)$ (Fig. [7](#page-12-0)h). The EC patients were classifed into the high and low TMB group. Kaplan–Meier analysis showed High TMB

Fig. 5 Relationships between the risk score and clinicopathologic factors. **a** The outcome prediction of the hypoxia-related lncRNA signature in patients stratifed by age, grade, stage, surgical approach and radiation therapy in TCGA cohort. **b**–**e** The outcome (**b**, **c**) and

expression (d, e) in different age (≤ 60 and > 60), grade (G1–G2 and G3–G4), stage (I–II and III–IV), surgical approach (minimally invasive and open) and radiation therapy (No and Yes) group were analyzed

Fig. 6 Biological characteristics and pathway analysis of hypoxia-related risky lncRNA. **a**-**b** Gene Ontology (GO) and **c**, **d** Kyoto Encyclopedia of Genes and Genomes (KEEG) pathway analysis. **e** The Gene Set Enrichment Analysis (GESA),

Fig. 7 The risk score of hypoxia-related lncRNA was associated with immunotherapy, microsatellite instability and tumor mutation burden in EC. **a**–**d** The response to anti-CTLA-4 and anti-PD-1 treatment between diferent risk score group. **e**, **f** The relationship between hypoxia-related lncRNA risk score and microsatellite instability in EC patients. **g** The tumor mutation burden levels in diferent risk

score group. **h** The Pearson correlation coefficients between tumor mutation burden levels and risk scores. **i**, **j** Kaplan–Meier analysis showed the correlation between tumor mutation burden and EC prognosis (**i**), and the diference in survival rate according to risk score between diferent TMB subgroups (**j**)

Fig. 8 SOS1-IT1 is clinically relevant in EC and promotes EC cell ◂ growth. **a**, **b** The expression level of SOS1-IT1 in normal and tumor tissues of TCGA EC database. **c**–**f** The correlation analysis of SOS1- IT1 and clinicopathological parameters of EC, including the age, disease type, fustat and grade. **g** Kaplan–Meier analysis of SOS1- IT1 and the prognosis in EC patients. **h**, **i** The endometrial cancer cell line Ishikawa was knocked down with two diferent SOS1-IT1 shRNA (**h**), or transfected with SOS1-IT1 plasmid (**i**), and the expression level of SOS1-IT1 was detected by real-time PCR. **j**, **k** The cell growth was performed by WST-1 assays in SOS1-IT1 knocked down or overexpression in Ishikawa cells. ** *P*<0.01, ****P*<0.001

signifcantly correlated with better prognosis (Fig. [7i](#page-12-0)). Then we evaluated the synergistic effect of risk score on prognosis stratifcation. The survival analysis showed that there was signifcant diference in survival rate according to risk score between diferent TMB subgroups (Fig. [7j](#page-12-0)).

SOS1‑IT1 is clinically relevant in EC and promotes EC cell growth

SOS1-IT1 was the most correlated prognostic lncRNAs in this model. Therefore, we will further evaluate the role of SOS1-IT1 in EC to verify the hypoxia-related lncRNA model. To investigate the clinical signifcance of SOS1-IT1 in EC, we analyzed its expression and clinical relevance in TCGA EC database. As shown in Fig. [8a](#page-14-0), [b,](#page-14-0) SOS1-IT1 was overexpressed in tumor tissues. Analysis from clinical investigations suggested that the aberrant level of SOS1- IT1 was closely correlated with clinicopathological parameters of EC, including the age, disease type, fustat and grade (Fig. [8](#page-14-0)c–f). Kaplan–Meier analysis revealed that high expression of SOS1-IT1 was signifcantly associated with a poor prognosis in EC patients (Fig. [8](#page-14-0)g). To directly investigate the biological functions of SOS1-IT1 on EC cells, SOS1-IT1 was knocked down or overexpressed respectively in EC cell line Ishikawa cells. The real-time PCR analysis results indicated that the expression level of SOS1-IT1 was markedly decreased or increased in knocking down or overexpressing cells respectively (Fig. [8](#page-14-0)h, i). The downregulation of SOS1-IT1 signifcantly inhibited EC cell growth. In contrast, overexpressing SOS1-IT1 yielded the opposite results (Fig. $8i$, k). These results indicate that SOS1-IT1 may lead to increased EC aggressiveness.

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was overexpressed in tumor tissues. Analysis from clinical investigations suggested that the aberrant level of SOS1- IT1 was closely correlated with clinicopathological parameters of EC, including the age, disease type, fustat and grade (Fig. [8](#page-14-0)c–f). Kaplan–Meier analysis revealed that high expression of SOS1-IT1 was signifcantly associated with a poor prognosis in EC patients (Fig. [8](#page-14-0)G). To directly investigate the biological functions of SOS1-IT1 on EC cells, SOS1-IT1 was knocked down or overexpressed respectively in EC cell line Ishikawa cells. The real-time PCR analysis results indicated that the expression level of SOS1-IT1 was markedly decreased or increased in knocking down or overexpressing cells respectively (Fig. [8h](#page-14-0), i). The downregulation of SOS1-IT1 signifcantly inhibited EC cell growth. In contrast, overexpressing SOS1-IT1 yielded the opposite results (Fig. [8j](#page-14-0)–k). These results indicate that SOS1-IT1 may lead to increased EC aggressiveness.

SOS1‑IT1 is upregulated under hypoxia and directly transactivated by HIF‑1α

To further confrm whether SOS1-IT1 was a functional efector of hypoxia in EC progression, we treated Ishikawa cells with hypoxia or its chemical inducer CoCl₂ for 24 h, and found the expression level of SOS1-IT1 was signifcantly increased (Fig. [9](#page-16-0)a). Besides, the expression level of SOS1-IT1 was signifcantly decreased after knocking down HIF-1 α , which is the main signaling pathway response component to hypoxia, both in normoxia and hypoxia condition (Fig. $9b$, c). After detecting of the upstream region (\sim 1 KB upstream) of SOS1-IT1 gene by promoter sequence analysis, we found a putative HIF-1 α binding site in the promoter region of SOS1-IT1 gene (Fig. [9d](#page-16-0)). We generated the mutant binding site of the reporter constructs, following with luciferase assays after transfection of diferent reporter constructs in EC cells. Under normoxia conditions, there was no much diference of luciferase activity in SOS1-IT1 wild type (WT) or mutant, as well as empty vector group. However, under hypoxic conditions, there was a nearly six-fold induction of luciferase activity in SOS1-IT1 WT construct. Mutation in SOS1-IT1 promoter decreased the luciferase activity to nearly basal levels caused by hypoxia (Fig. [9](#page-16-0)e). Besides, the Chromatin immunoprecipitation (ChIP) assays confrmed that HIF-1 α directly bind to the promoter region promoter of SOS1-IT1 (Fig. [9](#page-16-0)f). Next, we knocked down HIF-1 α , and found HIF-1 α suppression remarkably repressed the luciferase density in cells with WT promoter, but not in mutant group (Fig. [9](#page-16-0)g). Moreover, SOS1-IT1 silencing partially reduced EC cell proliferation ability by hypoxia condition or HIF-1 α overexpression (Fig. [9h](#page-16-0)–j). Totally, these results intensively indicated that SOS1-IT1 was upregulated under hypoxia and a direct transcriptional target of HIF-1α.

Discussion

Endometrial cancer is the sixth most common neoplasm in females, with rapidly increasing in the worldwide and causes ∼74,000 deaths per year (Siegel et al. [2020;](#page-17-1) Tsikouras

et al. [2013\)](#page-17-4). The TCGA networking group initially identifed four molecular subtypes with diferent prognosis based on genome somatic copy number changes, microsatellite instability and tumor mutation load (Suri and Arora [2015](#page-17-28)). There are more and more molecular targeted therapies and

Fig. 9 SOS1-IT1 is upregulated under hypoxia and directly transac-◂ tivated by HIF-1α. **a** The Ishikawa cells were treated with hypoxia or its chemical inducer CoCl₂ for 24 h, and the expression level of SOS1-IT1 was detected by real-time PCR. **b**–**c** HIF-1α was knocked down by two diferent HIF-1α small interfering RNA (siRNA), and the expression level was analyzed by western blot (**b**). The expression level of SOS1-IT1 was analyzed after knocking down HIF-1 α , both in normoxia and hypoxia condition (**c**). **d** Schematic illustration of the putative HIF-1α binding site in SOS1-IT1 gene promoter. **e** The dual-luciferase reporter assays were performed in Ishikawa cells transfected with diferent reporter constructs, both in normoxia and hypoxia condition. **f** The ChIP assays were performed using control IgG or anti-HIF-1 α antibody in Ishikawa cells. The binding of HIF-1α on SOS1-IT1 promoter was analyzed by real-time PCR. **G** The dual-luciferase reporter assays were performed in Ishikawa cells when knocking down HIF-1α with two diferent siRNAs. **h** The cell growth was performed by WST-1 assays after SOS1-IT1 knocked down in normoxia or hypoxia condition. **i** The Ishikawa cells were overexpressed with fag tagged HIF-1α, and the expression of HIF-1α protein was verified by western blot. The flag tagged HIF-1 α followed was detected by immunoblotting with an anti-fag antibody, and the actin protein act as internal reference. **j** The cell growth was performed by WST-1 assays after HIF-1 α overexpression with SOS1-IT1 knocked down. ** *P*<0.01, ****P*<0.001

diagnostic methods for EC (Morice et al. [2016\)](#page-17-29). However, the clinical efect of EC is still unsatisfactory. In recent years, many studies have shown that the lncRNA were closely associated with the prognosis of EC patients. In this study, we established a hypoxia-related lncRNA signature, which can efectively distinguish EC patients and predict their survival. To the best of our knowledge, this is the frst study to investigate the association of hypoxia-related lncRNA with prognostic features in EC patients.

The hypoxic microenvironment common to cancer cells emerges as an important factor for cancer progression (Jing et al. [2019\)](#page-17-5). Besides, hypoxia is an aggressive feature in EC, which can be used for treatments targeting biological changes related to hypoxia. Growing evidence indicates a close correlation between lncRNA and EC (Liu et al. [2019a](#page-17-16)). However, there is no systematic study on the correlation between hypoxia and EC. The rapid development of highthroughput gene sequencing technology has laid a foundation for the research of large cancer data (Prokop et al. [2018](#page-17-30)). A large number of genomic data were extracted from a single specimen to identify new prognostic and pharmacological biomarkers (Ballester et al. [2016](#page-17-31); Su et al. [2011](#page-17-32)). This selected risk prediction may be a more targeted and efective prognostic assessment for predicting positive clinical outcomes. Compared with other known prognostic indicators, it may be a more efective classifcation tool for EC patients.

In the current study, we built a hypoxia-related lncRNA signature. We found that age, grade, stage, disease type and risk score were correlated with the OS of EC patients, and were independent risk factors for the prognosis of EC patients. In order to understand the mechanism of these signature, we used GSEA to analyze the KEGG pathway between diferent risk groups. We found that the most signifcant enrichment pathways in the high-risk group were cell cycle and wnt signaling, which were reported associated with the poor prognosis of EC. This partly explained the molecular mechanism of the diference in prognosis between high-risk group and low-risk group. In addition, the risk score was closely related to immunotherapy response, microsatellite instability and TMB.

In order to further confrm the model built in bioinformatics analysis, we select SOS1-IT1 to validate its role in EC cell line. SOS1-IT1 was frst reported as the core lncRNA in the prognostic model of ivermectin-related three-lncRNA signature in ovarian cancer (Li and Zhan [2020\)](#page-17-33). It was also identifed as autophagy-related lncRNA in endometrial cancer (Wang et al. [2020b\)](#page-17-34). However, its detailed function and molecular mechanism in EC need to be further elucidated. In this study, we found SOS1-IT1 was overexpressed in tumor tissues, and closely correlated with clinicopathological parameters of EC, including the age, disease type, fustat and grade. Moreover, we detected SOS1-IT1 regulated EC cell growth by using knocked down and overexpression experiments. Interestingly, we also found SOS1-IT1 was a functional efector of hypoxia in EC progression. Its expression level was increased in hypoxia or its chemical inducer $CoCl₂$. Additionally, the important hypoxia regulatory factor HIF-1α can directly bind SOS1-IT1 promoter region, and afect its expression level. These results further indicate the relationship between SOS1-IT1 and hypoxia, which may increase EC aggressiveness.

In conclusion, we frst clarifed the clinical signifcance of hypoxia-related lncRNA signature in predicting the overall survival rate of EC patients. The hypoxia-related lncRNA was involved in the growth and progression of EC through diferent pathways. Consistent with previous studies, our fndings suggested that some hypoxia-related lncRNA can predict survival outcomes and monitor tumor progression, which increase the reliability of hypoxia-related lncRNA signature. It is also of great signifcance to reveal the potential molecular mechanism and roles of these lncRNA in other types of malignant tumors.

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Author contributions HYL, JHW, SHC contributed to the manuscript preparation and design of the study. QLF, HYL, JHW carried out the experiments and data analysis. JYL, JL provided their expertise. All authors reviewed the draft and approved the submitted version.

Declarations

Conflict of interest The authors have declared that no competing interest exists.

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