



Role of the Gut in the Temporal Changes of β -Cell Function After Gastric Bypass in Individuals With and Without Diabetes Remission

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OBJECTIVE

The role of the gut in diabetes remission after Roux-en-Y gastric bypass (RYGB) is incompletely understood. We assessed the temporal change in insulin secretory capacity after RYGB, using oral and intravenous (IV) glucose, in individuals with type 2 diabetes.

RESEARCH DESIGN AND METHODS

Longitudinal, prospective measures of β -cell function were assessed after oral glucose intake and graded glucose infusion in individuals with severe obesity and diabetes studied at 0, 3 ($n = 29$), 12 ($n = 24$), and 24 ($n = 20$) months after RYGB. Data were collected between 2015 and 2019 in an academic clinical research center.

RESULTS

The decreases in body weight, fat mass, waist circumference, and insulin resistance after surgery (all $P < 0.001$ at 12 and 24 months) did not differ according to diabetes remission status. In contrast, both the magnitude and temporal changes in β -cell glucose sensitivity after oral glucose intake differed by remission status ($P = 0.04$): greater (6.5-fold; $P < 0.01$) and sustained in those in full remission, moderate and not sustained past 12 months in those with partial remission (3.3-fold; $P < 0.001$), and minimal in those not experiencing remission (2.7-fold; $P =$ not significant). The improvement in β -cell function after IV glucose administration was not apparent until 12 months, significant only in those in full remission, and only ~33% of that observed after oral glucose intake. Preintervention β -cell function and its change after surgery predicted remission; weight loss and insulin sensitivity did not.

CONCLUSIONS

Our data show the time course of changes in β -cell function after RYGB. The improvement in β -cell function after RYGB, but not changes in weight loss or insulin sensitivity, drives diabetes remission.

The role of the gut in diabetes remission after Roux-en-Y gastric bypass (RYGB) (1) is incompletely understood. The rapid postoperative calorie restriction, the

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decreased glucose toxicity, and the enhancement of the incretin effect all improve islet function, independent of weight loss (2–6); whereas insulin sensitivity improves as a function of weight loss (7). The recovery of β -cell function measured during an oral stimulus, observed after RYGB (5), is not seen after a purely restrictive surgery such as gastric banding, even after matched weight loss (8,9), highlighting the importance of gastrointestinal factors. Here, we sought to characterize temporal changes in β -cell function after both oral and an intravenous (IV) glucose challenges in individuals with variable states of type 2 diabetes (T2D) remission after RYGB. We hypothesized that after RYGB 1) the improvement in β -cell function after oral glucose intake will be greater in magnitude and occur sooner than after IV glucose administration; and 2) the improvement in body composition and in insulin sensitivity would not differ by diabetes remission status.

RESEARCH DESIGN AND METHODS

Participants

Individuals with obesity and T2D, scheduled to have RYGB at St Luke's Roosevelt Hospital, New York City, provided written informed consent before participating. Of the 36 consented individuals, 29 returned at 3 months (3M); 1 never had surgery, 2 had unplanned vertical sleeve gastrectomy, and 4 were lost to follow-up, 24 returned at 12 months (12M), and 20 at 24 months (24M) (see the CONSORT diagram in Supplementary Material).

Study Design

This study was a longitudinal prospective study of participants studied before and at 3M, 12M, and 24M after RYGB. All participants had T2D before surgery (10). After surgery, the same criteria (10) were used to define full remission ($HbA_{1c} < 5.7\%$ [39 mmol/mol], fasting glucose < 100 mg/dL, 120-min glucose test level < 140 mg/dL, not taking any diabetes medication); persistent diabetes ($HbA_{1c} \geq 6.5\%$ [48 mmol/mol], fasting glucose ≥ 126 mg/dL, and/or 120-min glucose-test level ≥ 200 mg/dL, with or without diabetes medication); and partial remission (HbA_{1c} 5.7%–6.4% [39 to 46 mmol/mol], fasting glucose 100–125 mg/dL and/or 120-min glucose-

test level 140–199 mg/dL, not taking diabetes medications). Diabetes status was assessed at each study time point. For prediction models of long-term diabetes remission, data from the latest available study time point postsurgery was used to divide participants into those in full remission (F-REM; $n = 5$), partial remission (P-REM; $n = 16$), and those not in remission (N-REM; $n = 8$). For all participants but five, who only came back at 3M, the last assessment was at 12M ($n = 24$) or 24M ($n = 20$). Before surgery, GLP-1 analogs, DPP-4 inhibitors, and thiazolidinediones were discontinued at least 2 months before the first study visit and replaced as needed by sulfonylureas or insulin. All oral antidiabetes medications were withheld for 3 days prior to each visit; the last shot of insulin was given 24 h before each study visit.

Surgery

All RYGB surgeries were done by the same bariatric team. The jejunum was divided 30 cm from the ligament of Treitz, anastomosed to a 30-mL proximal pouch, and re-anastomosed 150 cm distal to the gastrojejunostomy (2). Diet before or after surgery was neither monitored nor controlled for, and participants followed dietary recommendations from the bariatric team; none followed a liquid diet at the time of visits.

Experimental Procedures

Oral Glucose Tolerance Test

Participants underwent a 3-h oral glucose tolerance test (OGTT; 75 g of glucose in 222 mL noncarbonated drink) after a 12-h overnight fast at each visit. The glucose solution was consumed over 15 min; blood samples, collected in chilled EDTA tubes at 0, 15, 30, 45, 60, 90, 120, and 180 min using an antecubital IV catheter, were centrifuged at 4°C and stored at -80°C until assayed.

Graded Glucose Infusion

Patients were admitted after a 12-h overnight fast; antecubital IV catheters were placed in each arm. If fasting glucose level was > 150 mg/dL (8.33 mmol/L), a 0.6 unit/kg bolus of IV regular insulin followed by a continuous infusion of 0.01 unit/kg/h was initiated until the blood glucose level reached ≤ 150 mg/dL, at which point the infu-

sion was stopped. The graded glucose infusion (GGI) started 60 min later (time 0) with an infusion of 20% dextrose at 2 mg/kg/min. Every 30 min, the infusion rate was increased to 4, 6, 8, and 12 mg/kg/min, with blood samples collected every 10 min. The order of OGTT and GGI was randomized, with at least 1 week separating the two tests.

Body Composition by Three-Dimensional Photonic Scanner

Fat mass and waist circumference were measured by the validated three-dimensional photonic scanner (Hamamatsu Photonics KK, Hamamatsu City, Shizuoka, Japan) (11,12). Seven participants had missing data at 12M or 24M and were omitted from the analysis.

Biomarkers

Plasma glucose level was determined by the glucose oxidase method with an Analox glucose analyzer (Lunenburg, MA). Plasma insulin and C-peptide levels were measured by radioimmunoassay (EMD Millipore, St Charles, MO, USA) in the Diabetes Research Center Biomarkers Core. Intra- and interassay coefficients of variance ranged from 3.4% to 7.4%.

Calculations

Total area under the curve (tAUC) and incremental area under the curve (iAUC) during the OGTT and GGI were calculated using the trapezoidal method (13). HOMA-IR was calculated as follows: (fasting insulin $\mu\text{U}/\text{mL} \times$ fasting glucose mmol/L)/22.5 (14). β -Cell function was assessed using the following: the prehepatic insulin secretion rates (ISRs), determined by C-peptide deconvolution using a two-compartment model (15); β -cell glucose sensitivity (BCGS) was calculated as the slope of the relationship between ISR and corresponding plasma glucose, using fasting glucose to peak glucose values (usually at 30 min) during the OGTT, and fasting glucose to highest glucose value (at 180 min) for the GGI; the early OGTT insulinogenic index (eIGI), calculated as $(\text{insulin}_{30 \text{ min}} - \text{insulin}_0 \text{ min}) / (\text{glucose}_{30 \text{ min}} - \text{glucose}_0 \text{ min})$ and the total insulinogenic index (tIGI), calculated as $\text{insulin}_{\text{tAUC}} / \text{glucose}_{\text{tAUC}}$. The disposition index (DI), calculated as the product of either BCGS after oral glucose challenge (O-BCGS) or BCGS after GGI (GGI-BCGS)

and 1/HOMA-IR. Fasting insulin clearance rate was calculated as following: fasting ISR/fasting insulin; and ICR during OGTT as $(ISR_{tAUC}/insulin_{tAUC}) - (V * [(insulin_{end\ time} - insulin_{start\ time})/insulin_{tAUC}])$, where V is the volume of distribution of insulin and estimated as 0.14 L/kg (16).

Nomenclature

Condition refers to the months when participants were studied: presurgery, 3M, 12M, and 24M after surgery; time, in minutes, refers to the time point at which blood samples were collected during OGTT or GGI. Groups were defined on the basis of diabetes remission status at the latest time point after surgery: F-REM, P-REM, and N-REM. All variables derived from OGTT are labeled with an *O*- (e.g., *O*-ISR for ISR calculated during the OGTT). Variables calculated during the GGI are labeled with the prefix GGI- (e.g., GGI-ISR).

Statistical Analysis

Continuous variables were tested for normality and log transformed if found to have a nonparametric distribution. ANOVA tests were used to discern differences of mean values between before and after surgery within and between remission groups.

The general linear model with repeated measure was used to study the effect of time since surgery (condition effect) and of remission status (condition × group effect) on changes in outcome variables and to study changes in metabolic and hormonal variables occurring during the OGTT and GGI glucose challenges (time effect).

Pearson and Spearman correlations and linear regression were used to determine the predictive value of baseline clinical characteristics, baseline β-cell function or its early change at 3M or total change at 12M and 24M, and percent weight loss on glucose control post-RYGB. Multinomial logistic regression was used to determine these variables' predictive potential on remission status at the latest time point. Data are expressed as mean ± SD except in figures where mean ± SEM are reported.

Power analysis was based on the change of β-cell function after RYGB in individuals in diabetes remission (4). We determined that eight participants would be needed to see a difference of BCGS

between pre- and post-RYGB of 1.74 with a SD of 1.22 (effect size $d = 1.43$ [i.e., $1.74/1.22$], and $\alpha = 0.05$).

RESULTS

Baseline Participant Characteristics

Participants were either Hispanic ($n = 22$) or Black ($n = 7$), predominantly women (79%), aged 42.9 ± 8.3 years (range 21–61 years), with known T2D duration of 7.7 ± 7.3 years (range 1 month to 26 years) (Supplementary Table 1). There were no baseline differences in participants' characteristics and no difference at 3M in weight, weight loss, and values for HbA_{1c} , HOMA-IR, fasting and 120-min glucose, *O*-glucose AUC, and *O*-BCGS between those who completed the study and participants who dropped out after the 3M study time point (data not shown).

There were no baseline differences in body composition and insulin resistance between remission groups (Table 1); however, as expected, preintervention indicators of β-cell function (i.e., *O*-BCGS, *O*-ISR, and GGI-ISR) were greatest in F-REM and P-REM, compared with N-REM ($P < 0.05$) (Supplementary Table 2).

Effect of Surgery on Diabetes Remission

As predicted, the use of all diabetes medications decreased after surgery. HbA_{1c} decreased by 1.65% at 3M and remained at this improved level up to 24M ($P < 0.0001$) (Supplementary Table 1). Although the decrease in HbA_{1c} was clinically relevant in all groups (−0.5% to −1.9%), it was statistically significant only in F-REM and P-REM (Table 1).

Effect of Surgery on Body Composition and Insulin Sensitivity

Body weight, BMI, waist and hip circumferences, and fat mass decreased after surgery (condition effect, $P < 0.001$) without differences by remission status (condition × group, $P =$ not significant). HOMA-IR decreased after surgery ($P < 0.0001$), without group effect (condition × group, $P = 0.49$) (Table 1). HOMA-IR was strongly associated with fat mass ($r = 0.45$; $P < 0.001$), waist circumference ($r = 0.49$; $P < 0.001$), and body weight ($r = -0.43$; $P = 0.001$) across conditions.

Post-RYGB Changes in Glycemia and β-Cell Function During OGTT

The postoperative improvement in glycemia during the OGTT (condition effect, $P < 0.001$) varied by remission status (condition × group × time, $P = 0.004$) (Fig. 1A). Improvement was greatest in F-REM, moderate in P-REM, and minimal in N-REM (Fig. 1B–D). For the entire cohort, compared with presurgery, the temporal pattern of change in β-cell function was an immediate and significant improvement at 3M, further increased at 12M, and returned to 3M levels at 24M (Table 2). Hypoglycemia (48–65 mg/dL) was documented during the OGTT in three of five participants with F-REM at 12M and 24M but was symptomatic in only one participant. The *O*-ISR (i.e., $tAUC$ -ISR and $iAUC$ -ISR) increased at all conditions after surgery, compared with presurgery (condition effect $P = 0.012$ and $P = 0.002$, respectively) (Table 2), without group differences (condition × group $P = 0.23$ and $P = 0.34$, respectively) (Supplementary Table 2), with peak increase observed at 12M (Table 2).

O-BCGS increased after surgery (condition effect $P < 0.001$) by 2.7-fold at 3M ($P = 0.008$) and by 3.8-fold at 12M ($P < 0.001$), and reverted to 3M values at 24M (2.8-fold, $P = 0.42$ vs. 3M) (Table 2). *O*-BCGS strongly correlated with 2-h postprandial glucose level (Supplementary Fig. 2). Both the magnitude and temporal changes in *O*-BCGS varied by remission status (condition × group, $P = 0.04$). They were greater (5.2- to 6.5-fold; $P < 0.01$) with a sustained increase in F-REM, moderate increase and not sustained past 12M in P-REM (2.3- to 3.3-fold, $P < 0.001$), and minimal and very transient in N-REM (0.8- to 2.7-fold; $P < 0.05$) (Fig. 2A). The overall trend and statistical significance (Supplementary Fig. 1), and the variation according to remission status, were similar for the *O*-DI (Table 2 and Supplementary Table 2).

Interestingly, the pattern of change was different for the IGI (both eIGI and tIGI). The improvement in eIGI after surgery (condition effect, $P < 0.01$) (Table 2), did not differ by group (condition × group $P = 0.85$) (Supplementary Table 2).

The changes in *O*-BCGS and *O*-DI were associated with fat mass ($r = 0.37$, $P = 0.001$; $r = -0.49$, $P < 0.001$, respectively) and waist circumference ($r =$

Table 1—Participants' characteristics by remission status

	Presurgery						3M			12M			24M		
	F-REM	P-REM	N-REM	F-REM	P-REM	N-REM	F-REM	P-REM	N-REM	F-REM	P-REM	N-REM	F-REM	P-REM	N-REM
Sex (men/women)	1/4	3/13	2/6	1/4	3/13	2/6	1/4	3/13	2/6	1/4	3/13	2/6	1/4	3/13	2/6
Age (years)	46.4 ± 6.23	40.6 ± 9.61	45.3 ± 5.42	46.4 ± 6.23	40.6 ± 9.61	45.3 ± 5.42	46.4 ± 6.23	40.6 ± 9.61	45.3 ± 5.42	46.4 ± 6.23	40.6 ± 9.61	45.3 ± 5.42	46.4 ± 6.23	40.6 ± 9.61	45.3 ± 5.42
Ethnicity (Hispanic/Black)	4/1	13/3	5/3	4/1	13/3	5/3	4/1	13/3	5/3	4/1	13/3	5/3	4/1	13/3	5/1
Oral medication (n)	3/5	12/16	7/8	0/5	6/16	8/8	0/5	6/16	8/8	0/5	6/16	8/8	0/5	6/16	3/6
Using insulin (n)	0/5	3/16	6/8	0/5	0/16	2/8	0/5	0/16	2/8	0/5	0/16	2/8	0/5	0/16	2/6
T2D duration (years) [‡]	1.48 ± 0.99 ^a	7.38 ± 7.95 ^b	12.2 ± 5.17 ^b	1.48 ± 0.99 ^a	7.38 ± 7.95 ^b	12.2 ± 5.17 ^b	1.46 ± 1.00 ^a	8.38 ± 8.47 ^b	13.9 ± 3.44 ^b	1.46 ± 1.00 ^a	8.38 ± 8.47 ^b	13.9 ± 3.44 ^b	1.96 ± 0.81	8.16 ± 8.46	11.2 ± 5.47
HbA _{1c} (%) [‡]	7.34 ± 0.85 ^a	7.68 ± 1.17 ^a	8.41 ± 0.88 ^a	5.56 ± 0.52 ^{a***}	5.96 ± 0.38 ^{a***}	7.39 ± 1.09 ^b	5.51 ± 0.19 ^{a***}	5.87 ± 0.29 ^{a***}	7.34 ± 1.25 ^b	5.42 ± 0.13 ^{a***}	5.97 ± 0.23 ^{a***}	7.86 ± 1.71 ^b	5.42 ± 0.13 ^{a***}	5.97 ± 0.23 ^{a***}	7.86 ± 1.71 ^b
(mmol/mol)	57	60	68	37	42	57	37	41	57	36	42	62	36	42	62
BMI (kg/m ²)	42.2 ± 4.29 ^a	43.3 ± 4.97 ^a	40.6 ± 4.10 ^a	34.1 ± 4.14 ^a	36.2 ± 4.68 ^{a**}	33.9 ± 3.94 ^{a*}	29.2 ± 5.22 ^{a**}	32.7 ± 5.12 ^{a***}	29.5 ± 5.16 ^{a**}	29.8 ± 6.82 ^{a*}	27.6 ± 5.47 ^{a***}	33.5 ± 6.02 ^{a***}	29.8 ± 6.82 ^{a*}	27.6 ± 5.47 ^{a***}	33.5 ± 6.02 ^{a***}
BMI (% loss)	19.16 ± 3.44	16.44 ± 3.76	16.61 ± 3.25	31.00 ± 7.67	24.84 ± 4.91	27.54 ± 9.66	33.1 ± 11.1	22.5 ± 9.76	29.9 ± 11.3	33.1 ± 11.1	22.5 ± 9.76	29.9 ± 11.3	33.1 ± 11.1	22.5 ± 9.76	29.9 ± 11.3
Weight (kg)	113 ± 12.2 ^a	118 ± 17.3 ^a	107 ± 25.9 ^a	91.2 ± 11.5 ^a	98.8 ± 14.8 ^{a**}	89.5 ± 22.4 ^a	78.1 ± 14.3 ^{a**}	88.6 ± 14.8 ^{a**}	79.0 ± 25.8 ^a	78.7 ± 22.2 ^{a*}	89.2 ± 17.1 ^{a***}	68.2 ± 16.9 ^{a*}	78.7 ± 22.2 ^{a*}	89.2 ± 17.1 ^{a***}	68.2 ± 16.9 ^{a*}
Weight loss (%)	19.2 ± 3.25 ^a	16.4 ± 3.8 ^a	16.61 ± 3.3 ^a	31.0 ± 7.6 ^a	24.8 ± 4.9 ^a	27.5 ± 9.6 ^a	33.1 ± 11.1 ^a	24.8 ± 4.9 ^a	27.5 ± 9.6 ^a	33.1 ± 11.1 ^a	24.8 ± 4.9 ^a	27.5 ± 9.6 ^a	33.1 ± 11.1 ^a	24.8 ± 4.9 ^a	27.5 ± 9.6 ^a
Fat mass (kg)	60.0 ± 10.8 ^a	58.2 ± 12.9 ^a	52.2 ± 8.8 ^a	43.8 ± 7.9 ^{a*}	46.7 ± 13.7 ^a	42.9 ± 8.2 ^a	40.3 ± 5.7 ^{a*}	39.5 ± 8.1 ^{a**}	33.2 ± 1.0 ^{a***}	36.8 ± 6.0 ^{a**}	43.2 ± 12.4 ^{a*}	32.4 ± 1.0 ^{a*}	36.8 ± 6.0 ^{a**}	43.2 ± 12.4 ^{a*}	32.4 ± 1.0 ^{a*}
Fat mass loss (%)	25.5 ± 13.6 ^a	20.2 ± 16.1 ^a	20.6 ± 11.6 ^a	33.8 ± 20.8 ^a	30.9 ± 15.3 ^a	31.5 ± 16.8 ^a	23.4 ± 13.5 ^a	30.9 ± 15.3 ^a	31.5 ± 16.8 ^a	23.4 ± 13.5 ^a	26.5 ± 13.8 ^a	39.7 ± 2.5 ^a	23.4 ± 13.5 ^a	26.5 ± 13.8 ^a	39.7 ± 2.5 ^a
Waist circumference (cm)	121 ± 5.7 ^a	125 ± 12.5 ^a	122 ± 12.3 ^a	103 ± 5.8 ^{a*}	111 ± 18.7 ^a	109 ± 13.4 ^a	96.7 ± 13.2 ^{a*}	105 ± 13.1 ^{a*}	86.1 ± 16.7 ^b	97.4 ± 11.2 ^{a*}	106 ± 15.7 ^{b*}	79.4 ± 12.5 ^{a**}	97.4 ± 11.2 ^{a*}	106 ± 15.7 ^{b*}	79.4 ± 12.5 ^{a**}
Hip circumference (cm) [‡]	133 ± 12.9 ^a	130 ± 9.6 ^a	120 ± 6.7 ^a	119 ± 11.4 ^a	118 ± 16.2 ^a	111 ± 5.6 ^{a*}	113 ± 9.6 ^a	114 ± 10.3 ^{a*}	98.2 ± 15.9 ^{a**}	115 ± 12.0 ^a	115 ± 11.2 ^a	91.5 ± 10.4 ^{a**}	115 ± 11.2 ^a	115 ± 11.2 ^a	91.5 ± 10.4 ^{a**}
Waist-to-hip ratio [‡]	0.91 ± 0.07 ^a	0.96 ± 0.06 ^a	1.00 ± 0.10 ^a	0.87 ± 0.06 ^a	0.95 ± 0.15 ^a	0.99 ± 0.11 ^a	0.85 ± 0.07 ^a	0.92 ± 0.06 ^a	0.87 ± 0.03 ^a	0.88 ± 0.03 ^a	0.93 ± 0.06 ^a	0.87 ± 0.04 ^a	0.88 ± 0.03 ^a	0.93 ± 0.06 ^a	0.87 ± 0.04 ^a
SBP (mmHg) [‡]	131 ± 17.1 ^a	126 ± 20.8 ^a	124 ± 12.4 ^a	119 ± 15.9 ^a	126 ± 20.4 ^a	133 ± 15.9 ^a	116 ± 21.2 ^a	124 ± 24.2 ^a	121 ± 12.2 ^a	119 ± 22.3 ^a	125 ± 21.2 ^a	114 ± 15.0 ^a	119 ± 22.3 ^a	125 ± 21.2 ^a	114 ± 15.0 ^a
DBP (mmHg)	77.0 ± 11.9 ^a	76.1 ± 15.4 ^a	78.8 ± 11.2 ^a	68.8 ± 12.2 ^a	76.2 ± 10.5 ^a	82.6 ± 5.7 ^a	67.2 ± 15.0 ^a	75.9 ± 11.3 ^a	76.5 ± 7.5 ^a	76.7 ± 10.0 ^a	76.2 ± 8.8 ^a	72.0 ± 9.7 ^a	76.7 ± 10.0 ^a	76.2 ± 8.8 ^a	72.0 ± 9.7 ^a
HOMA-IR [‡]	9.34 ± 2.91 ^a	13.5 ± 6.13 ^a	12.2 ± 8.49 ^a	2.29 ± 1.12 ^{a***}	3.61 ± 2.10 ^{a***}	5.07 ± 3.09 ^{a**}	1.98 ± 0.84 ^{a***}	2.33 ± 0.53 ^{a***}	4.11 ± 2.00 ^{b**}	1.58 ± 0.86 ^{a**}	2.41 ± 2.04 ^{a***}	2.99 ± 0.90 ^{a**}	1.58 ± 0.86 ^{a**}	2.41 ± 2.04 ^{a***}	2.99 ± 0.90 ^{a**}

Data are given as mean ± SD. Different letter superscripts indicate statistically significant differences between subgroups by one-way ANOVA post hoc multiple comparison tests at $P < 0.05$. For within-group differences from presurgery, * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$. †Not normally distributed; statistical analysis was done with log-transformed data. DPB, diastolic blood pressure; SBP, systolic blood pressure.

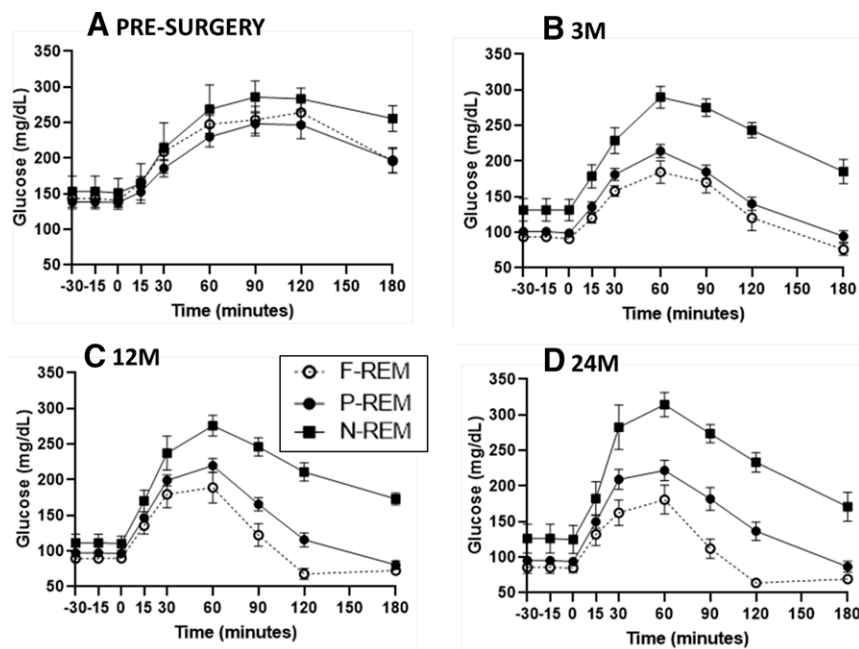


Figure 1—Changes in glycemia during the OGTT at (A) presurgery, (B) 3M, (C) 12M, and (D) 24M in F-REM (open circles), P-REM (black circles), and N-REM (black squares). Overall condition \times group \times time effect by general linear model test of within-subjects contrasts, $P = 0.004$.

-0.41 , $P < 0.001$; $r = -0.55$, $P < 0.001$, respectively) across conditions, and, but only for *O*-DI, with absolute and percent weight loss ($r = 0.34$, $P = 0.004$; $r = 0.33$, $P = 0.007$, respectively).

Change in β -Cell Function During GGI After Surgery

Seven participants at the presurgery condition, and only one at 3M, required a small insulin dose (bolus with or without insulin infusion) because of elevated glucose prior to initiating the GGI. Glu-

cose changes during GGI are presented in Supplementary Figure 4. Compared with *O*-BCGS, the increase in GGI-BCGS after surgery was of much smaller magnitude and statistically nonsignificant: 1.1-fold at 3M, 1.8-fold at 12M, and 1.7-fold at 24M (condition effect $P = 0.188$; condition \times group effect, $P = 0.440$) (Fig. 2B and Table 2). GGI-BCGS increased significantly only in F-REM at 12M ($P = 0.020$) and 24M ($P = 0.002$) compared with presurgery (Fig. 2B and Supplementary Table 2).

For the entire cohort, the GGI-DI increased significantly ($P = 0.002$ at 3M, $P < 0.001$ at 12 and 24M) with a maximum >8 -fold increase at 24M (Table 2 and Supplementary Fig. 1). This pattern of improvement differed from the *O*-DI, which peaked at 12M. GGI-DI improved significantly in F-REM and P-REM at 3M ($P < 0.05$ for both) and 12M ($P < 0.01$ and $P < 0.001$, respectively), and peaked at 24M ($P < 0.01$ and $P < 0.001$, respectively), but did not improve in N-REM (Supplementary Table 2 and Fig. 2B).

The changes in GGI-BCGS and GGI-DI across conditions were associated with fat mass ($r = -0.29$, $P = 0.012$; $r = -0.50$, $P < 0.00$, respectively), waist circumference (-0.34 , $P = 0.003$; -0.55 , $P < 0.001$, respectively), and weight loss ($r = 0.25$, $P = 0.039$; $r = 0.42$, $P < 0.001$, respectively).

Comparison of Oral vs IV-Derived β -Cell Function Biomarkers

GGI-BCGS was lower than *O*-BCGS at presurgery ($P < 0.001$) and remained lower at 3M ($P = 0.001$), 12M ($P < 0.001$), and 24M ($P = 0.20$) (Table 2 and Fig. 2). The absolute change in *O*-BCGS from pre- to postsurgery was more than 20 times greater than the change in GGI-BCGS at 3M (0.70 ± 0.77 vs. 0.03 ± 0.31 ; $P < 0.001$) and 40 times greater at 12M (1.21 ± 0.93 vs. 0.03 ± 0.44 ; $P < 0.001$). This large difference in the increase in BCGS after the oral versus the IV glucose challenge persisted but to a significantly lesser degree at 24M, with the change in *O*-BCGS at 24M only

Table 2—Change of β -cell function during oral and GGI challenges

	Presurgery	3M	12M	24M
<i>O</i> -ISR AUC (pmol/kg*180 min) [‡]	763 \pm 433	995 \pm 539	1,175 \pm 703*	1,165 \pm 758
<i>O</i> -ISR iAUC (pmol/kg*180 min) [‡]	406 \pm 302	718 \pm 473	868 \pm 557*	822 \pm 670*
<i>O</i> -BCGS (pmol/kg/min/mmol) [‡]	0.48 \pm 0.44	1.19 \pm 0.97**	1.68 \pm 1.19***	1.25 \pm 1.41
<i>O</i> -DI [‡]	0.05 \pm 0.05	0.47 \pm 0.52***	0.80 \pm 0.74***	0.65 \pm 0.80***
<i>O</i> -early IGI [‡]	79.51 \pm 128.3	140.8 \pm 161.6*	136.3 \pm 146.7	128.9 \pm 136.8
<i>O</i> -total IGI [‡]	47.44 \pm 66.18	64.27 \pm 81.09	55.99 \pm 53.78	50.48 \pm 59.49
GGI-ISR AUC (pmol/kg*180 min) [‡]	542 \pm 353	518 \pm 291	565 \pm 331	643 \pm 392
GGI-ISR iAUC (pmol/kg*180 min) [‡]	255 \pm 274	294 \pm 241	565 \pm 331	375 \pm 334
GGI-BCGS (pmol/kg/min/mmol) [‡]	0.30 \pm 0.31	0.33 \pm 0.32	0.54 \pm 0.57	0.52 \pm 0.57
GGI-DI [‡]	0.04 \pm 0.05	0.14 \pm 0.15**	0.25 \pm 0.28***	0.33 \pm 0.40***

Data given as mean \pm SD. Differences from presurgery by one-way ANOVA post hoc multiple comparison tests, * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$. [‡]Not normally distributed; statistical analysis was done with log-transformed data. Data were collected during the oral glucose challenge or during the GGI. iAUC, incremental area under the curve.

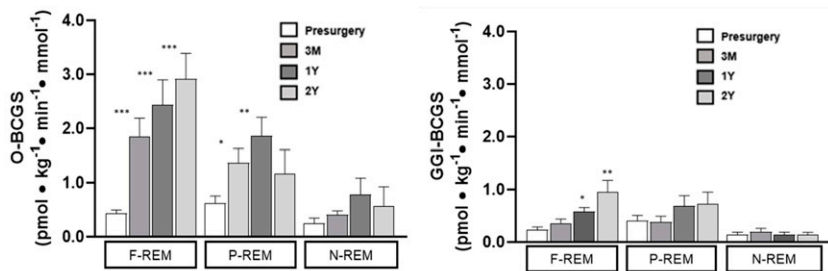


Figure 2—Change in *O*-BCGS and after GGI-BCGS according to diabetes remission. Mean \pm SEM. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$ vs. presurgery by one-way ANOVA with post hoc multiple comparison tests.

approximately four times greater (0.82 ± 1.38 vs. 0.22 ± 0.61 ; $P = 0.057$).

Interestingly, the magnitude of the difference between *O*-BCGS and GGI-BCGS differed by remission status, being greater in F-REM compared with P-REM and N-REM. In F-REM, *O*-BCGS was larger than GGI-BCGS at all time points ($P < 0.05$ at presurgery, 12M, and 24M; $P < 0.01$ at 3M). In P-REM, *O*-BCGS was greater than GGI-BCGS at 3M and 12M ($P \leq 0.001$) but not at 24M. In N-REM, however, the difference between *O*-BCGS and GGI-BCGS was minimal and significant only at 12M ($P < 0.001$) (Supplementary Table 2). Although the *O*-DI and the GGI-DI did not differ significantly at presurgery, the magnitude of the increase after surgery was twice as great for *O*-DI (16-fold) than for GGI-DI (>8-fold) (Table 2 and Supplementary Fig. 1).

As noted, GLP-1 concentrations during the OGTT (peak and AUC) increased by two to three times at all time points after RYGB (data not shown). The magnitude (peak and AUC) of the GLP-1 postprandial release did not differ by diabetes remission and did not change overtime (data not shown).

Insulin Clearance

As we have reported previously (16), insulin clearance under fasting conditions increased significantly and continuously at 3M, 12M, and 24M, when it more than tripled the presurgical value ($P < 0.001$), with no difference by group (condition \times group effect, $P = 0.42$). The *O*-ICR also increased after surgery, maximally at 24M ($P < 0.001$) again but with no difference by group (condition \times group effect, $P = 0.10$) (Table 1 and Supplementary Table 1).

Predictors of Diabetes Remission and Glucose Control

Bivariate predictive models were constructed using a stepwise regression or multinomial logistic regression based on clinical (diabetes duration, insulin use) and β -cell function (*O*-BCGS, GGI-BCGS) predictors with $P < 0.05$ during univariate linear regression or likelihood ratio (LR) tests, respectively (Supplementary Tables 3 and 4). A single clinical and β -cell function variable was added to each model, for a maximum of two baseline, early change, and total change predictors at once.

In a multinomial regression model, presurgery diabetes duration with either baseline *O*-BCGS or GGI-BCGS were the best predictors of remission status (LR test, $P = 0.001$ and $P = 0.003$, respectively). When BCGS was added to a model with presurgery insulin use, insulin use fell out of the model (insulin use was not significant by LR tests), leaving *O*-BCGS and GGI-BCGS as strong predictors of remission status (LR test, $P = 0.003$ and $P = 0.013$, respectively) (Supplementary Table 5B). Baseline *O*-BCGS was not a significant predictor of long-term HbA_{1c} when paired with either insulin use or diabetes duration (*O*-BCGS not significant and fell out of the model; $P = 0.45$ and $P = 0.24$, respectively). The best predictors of postsurgery HbA_{1c} were presurgery insulin use, with the early change in GGI-BCGS ($P < 0.001$; $R^2=0.51$), and with both the early and long-term change in *O*-BCGS ($P < 0.001$, $R^2=0.58$; $P < 0.001$, $R^2=0.55$, respectively). Diabetes duration paired with either early or long-term change in *O*-BCGS also significantly predicted HbA_{1c} ($P = 0.002$, $R^2=0.43$; $P = 0.003$, $R^2=0.43$) (Supplementary Table 5A).

CONCLUSIONS

The following are the main findings of the present study in individuals with severe obesity and T2D: 1) presurgery β -cell function is a strong predictor of T2D remission; 2) the temporality and magnitude of the improvement in β -cell function after RYGB are highly associated with remission status; 3) the greater magnitude of the improvement in β -cell function after oral glucose compared with IV glucose is more pronounced in F-REM; and 4) weight loss, fat mass loss, and decreases in waist circumference and insulin resistance after surgery are not key determinants of diabetes remission.

Our data confirm that the high degree of insulin resistance, a hallmark of T2D, improved significantly postsurgery, despite participants remaining in the obesity range at 12M with a mean BMI of 31 kg/m^2 . However, neither presurgery nor the large decreases in body weight ($\sim 30\%$ at 1 and 2 years), fat mass, and waist circumference, and the resulting improvement in insulin sensitivity differ according to diabetes remission status. So although the weight loss-dependent improvement in insulin sensitivity after surgery (7) surely plays a role in overall metabolism, it does not differentiate patients in remission from N-REM and does not appear to be a key determinant of the diabetes remission phenotype.

Contrary to weight loss and insulin sensitivity, presurgery β -cell function and its temporal variation after RYGB emerged as the strongest predictor of the magnitude of the reversal of hyperglycemia, as shown previously (5,6). Not surprisingly, individuals with the worst β -cell function at presurgery had the longest known diabetes duration, were more likely to be using insulin, had higher HbA_{1c} values, and were the least likely to experience remission after surgery. Although insulin resistance did not differ between groups at presurgery, presurgery β -cell function, calculated both after an oral and IV glucose challenge, was significantly more impaired in the N-REM group than in the F-REM or P-REM groups. Moreover, the temporal changes of β -cell function assessed during the OGTT were vastly different according to remission status; in the F-REM, robust, immediate (3M) continuous

and sustained (24M) improvement occurred, whereas in the P-REM or N-REM groups, the lesser improvement was only transient (at 3M and/or 12M). These changes were independent of weight and fat-mass losses or change in insulin sensitivity, or even change in insulin clearance, which were all similar in all three groups.

We (2) and others (17) have shown the importance of the incretins in restoring insulin secretion during an oral glucose challenge after RYGB. We also previously showed that the recovery of the β -cell function measured during an isoglycemic IV glucose clamp was only partial when compared with individuals with severe obesity and normal glucose tolerance or with lean individuals, even in individuals with full clinical diabetes remission studied up to 3 years after RYGB (4,6). In the present study, therefore, we assessed β -cell function after both oral glucose and GGI. As expected, we observed a striking difference in the improvement of β -cell function according to the route of the glucose challenge, with a three to four times greater increase after oral than IV glucose administration. In addition, we show that the improvement in BCGS during the GGI was quite limited in magnitude, significant only in F-REM, and delayed, starting only 1 year postsurgery. This is contrary to the rapid recovery of β -cell function after oral glucose shown previously after 1 month (4,6) and here at 3 months postsurgery. We also observed striking differences in the temporal changes in β -cell function according to remission status. Regardless of the route of the glucose stimulus, participants in the F-REM group demonstrated a continuous and sustained improvement of their β -cell function, even after body weight stabilized between years 1 and 2. The substantial improvement in the β -cell function of P-REM, however, was only transient and not sustained beyond year 1, despite absence of weight regain.

Interestingly, the gap between BCGS after oral and IV glucose administration, or the greater *O*-BCGS compared with GGI-BCGS was associated with remission status; it was primarily observed in F-REM and P-REM, and less so in the N-REM. On the basis of past work (2), these data suggest a larger incretin effect in patients with diabetes remission. In F-REM, *O*-BCGS tended to

increase overtime, which suggest some temporality of the involvement of the gut in the recovery of β -cell function. The accelerated nutrient transit is the main mechanism by which postprandial GLP-1 release is enhanced after RYGB (18). Because nutrient transit is unlikely to change significantly over time after surgery, the observed temporal change in *O*-BCGS could be related to either some degree of intestinal adaptation or reprogramming (19) or to change in the sensitivity to insulinotropic hormones after RYGB observed in individuals without diabetes (20,21).

Our data hint at the mechanisms by which RYGB improves β -cell function. Calorie restriction (22) rapidly decreases fasting glucose and clearly plays a role after RYGB when food and calorie intake decrease significantly (23). The resulting removal of glucose toxicity can also enhance BCGS (24). Chronic hyperglycemia is toxic to the β -cell (25) and BCGS is a key determinant of postprandial glucose level. This close temporal bidirectional relationship of BCGS and postprandial glucose levels, illustrated in Supplementary Figure 2, was previously highlighted by Nannipieri et al. (5) in a similar cohort.

A third mechanism is the large and sustained decrease of peripheral insulin resistance, which decreases the workload of the β -cell. These three mechanisms—calorie restriction, decreased glucose toxicity, and reduced insulin resistance—should equally play a role in improving BCGS after oral and IV glucose challenges. However, the modest improvement in IV glucose-challenge BCGS after RYGB, which was significant only in individuals in the F-REM group, speaks to the minimal parts played by these mechanisms. It could also indicate alteration of BCGS to IV-administered glucose, observed by others in individuals after RYGB who did not have diabetes (26).

On the contrary, the role of the incretins is likely to be quite significant and explains the extraordinary temporal recovery of *O*-BCGS after RYGB, which was of significantly greater magnitude in patients in remission. The postprandial incretin release is immediate (2,17) and sustained after RYGB (6), largely due to the rerouting of nutrients to the lower part of the gut (18). In individuals with T2D, the blunted incretin effect on

insulin secretion is very rapidly recovered 1 month after RYGB (2). The contribution of endogenous GLP-1 to the recovery of BCGS shortly after RYGB was elegantly illustrated in an experiment using the GLP-1 receptor antagonist exendin9–39 at 1 week and 3 months after RYGB (17) or in cross-sectional cohorts (27,28) in individuals without diabetes or who were in full diabetes remission. By comparing patients in remission and N-REM, the present study sheds new light on the possible role of the incretins in diabetes remission: First, the *O*-BCGS improves significantly more in patients in remission than in N-REM, whereas weight loss and change in insulin sensitivity, and even circulating GLP-1 concentrations (data not shown), were identical in the three groups; second, the differential improvement of *O*-BCGS over GGI-BCGS is more marked in patients in remission than N-REM, suggesting a better incretin effect.

The best predictors of remission were presurgery BCGS with or without diabetes duration. The best predictors of long-term HbA_{1c} were presurgery insulin use or diabetes duration, paired with changes in BCGS resulting from the surgery. Neither weight loss, fat-mass loss, nor change in insulin sensitivity predicted remission. These data confirm the importance of β -cell functional reserve, as shown by others (5,29) and confirm previous data in larger cohorts, using various scores, based on presurgery clinical variables (30,31), with or without change in β -cell function postsurgery (32–34). Interestingly, in a different homogenous cohort of individuals with diabetes of short duration (<2 years), with good glycemic control and taking few medications (i.e., with good β -cell reserve), both weight loss and preintervention β -cell function contributed to remission status (6).

Our study has many strengths. Data were from a single bariatric center with standardized gastric bypass technique, longitudinal measures of β -cell function in individuals with and without diabetes remission were obtained, both oral and IV glucose stimuli were used concomitantly in the same participant, and up to 2-year follow-up was conducted. Some limitations need to be acknowledged, including the lack of a sleeve gastrectomy group, given that this is

now more frequently performed, and the relative short-term follow-up, because relapse is often observed at 5 years post-surgery (35).

In summary, our data confirm that presurgery β -cell functional reserve is a key determinant of diabetes remission postsurgery. The data also show the temporal variability in the improvement, or lack of, in β -cell function after RYGB. Finally, the greater recovery of β -cell function after oral versus IV glucose stimulus underlines the key role of gut factors (i.e., the incretins) to restore β -cell function in diabetes remission.

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