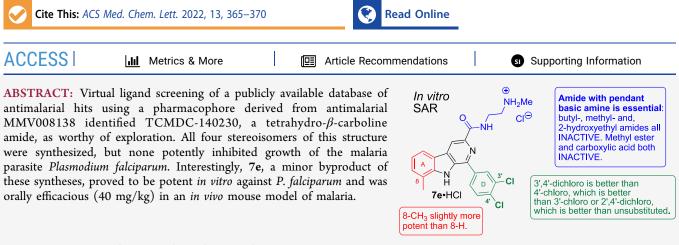


# Malaria Box-Inspired Discovery of N-Aminoalkyl- $\beta$ -carboline-3carboxamides, a Novel Orally Active Class of Antimalarials

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Worldwide, malaria is one of the deadliest diseases, causing more than 600,000 deaths in 2020.<sup>1</sup> In our search for new and safe drugs that can kill resistant strains of *Plasmodium falciparum* parasites, we were drawn to the 2-Cmethyl-D-erythritol-4-phosphate (MEP) pathway for synthesis of isoprenoid precursors, since this pathway is not present in humans.<sup>2</sup> Using the isopentenyl pyrophosphate (IPP) chemical rescue approach<sup>3</sup> to screen the 400-compound Malaria Box,<sup>4</sup> only MMV008138 (**1a**, Figure 1) was found to be an inhibitor of this pathway.<sup>5</sup> Other researchers determined that **1a** inhibits

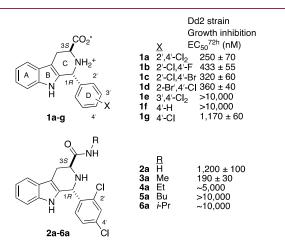


Figure 1. MEP pathway inhibiting antimalarials based on the MMV008138 (1a) scaffold.  $^{8-10}$ 

IspD, the third enzyme in the MEP pathway.<sup>6,7</sup> We subsequently synthesized and assayed dozens of analogs of **1a** that differed in stereochemistry and substitution of the B-, C-, and D-rings.<sup>8–10</sup> From these studies a clear picture emerged: (1R,3S)- stereochemistry, 2',4'-halogen disubstitution of the D-ring (e.g., **1a**–d), and a 3-carboxylic acid or methyl amide (e.g., **1a** and **3a**) were required for antimalarial potency. However, to date none of these potent *Pf* IspD inhibitors has shown oral efficacy in mice models of malaria.

Since 1a, as a member of the 400-compound Malaria Box, was selected from a much larger initial hit set (20K compounds with 3D7 strain half-maximal effective concentration (EC<sub>50</sub>) < 1  $\mu$ M),<sup>4</sup> we wondered if other structurally related compounds were present in the initial set. We thus performed virtual ligand screening (VLS) of the GlaxoSmithKline (GSK) portion of this larger collection (~13K compounds),<sup>11</sup> using a 3D pharmacophore derived from 1a and 92 close analogs. From this exercise the compound TCMDC-140230 emerged (Figure 2).

Several features of this compound stand out; in addition to the undisclosed stereochemistry, 3',4'-dichloro substitution and the large amide substituent were not suggestive of good

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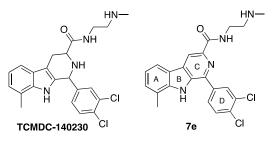


Figure 2. VLS hit TCMDC-140230 and derived lead 7e.

antimalarial potency within the established pharmacophore of 1a (Figure 1, cf. 1e vs 1a; 4a–6a vs 3a). As described below, the stereoisomers of tetrahydro- $\beta$ -carboline TCMDC-140230 proved to be dead ends, but the  $\beta$ -carboline analog 7e (tested as the HCl salt, Table 1) was found to have excellent antimalarial activity *in vitro* and *in vivo*.

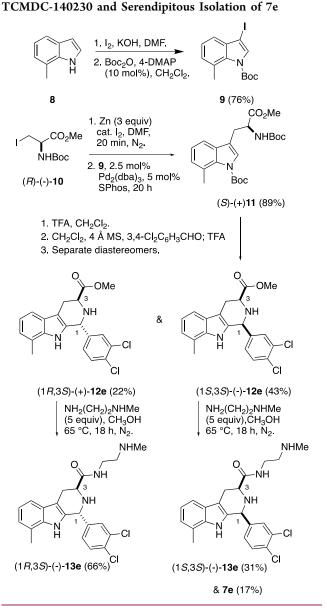
Table 1. In Vitro P. falciparum Growth Inhibition of 1a, 13e Stereoisomers, and 7e·HCl

Compound	$EC_{50} (nM)^a$	IPP rescue <sup>b</sup>
1a	$250 \pm 70^{c}$	100% @ 2.5 µM
(1R,3S)-(-)- <b>13e</b>	$1,950 \pm 240$	50% @ 5 µM
(1 <i>S</i> ,3 <i>S</i> )-(–)- <b>13e</b>	$1,300 \pm 100$	nd
(1 <i>S</i> ,3 <i>R</i> )-(+)- <b>13e</b>	3,690 ± 310	nd
(1R,3R)-(+)- <b>13e</b>	$1,430 \pm 70$	nd
7e·HCl	$108 \pm 7$	0% @ 2.5 μM

"Growth inhibition (72 h) of asexual blood stages determined using SYBR Green I assay. Values represent average  $\pm$  SEM from at least three independent biological replicates. The Dd2 strain is resistant to chloroquine, pyrimethamine, and mefloquine. <sup>b</sup>Recovery of parasite growth in the presence of 200  $\mu$ M IPP, at the indicated concentration of the test compound (Supporting Information); nd designates "not determined". <sup>c</sup>Reported previously.<sup>8</sup>

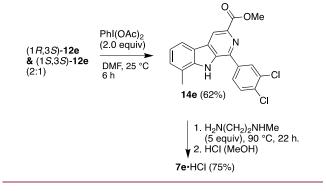
Since stereochemistry proved so crucial to the antimalarial activity of 1a, we undertook the enantioselective synthesis of the four stereoisomers of TCMDC-140230. 7-Methylindole 8 was converted to 9, which then underwent Negishi coupling with the organozinc reagent derived from N-Boc-iodoalanine methyl ester (R)-10,<sup>12</sup> affording the protected 7-methyltryptophan ester (S)-11. Deprotection, followed by Pictet-Spengler reaction<sup>8</sup> with 3,4-dichlorobenzaldhyde afforded (1R,3S)- and (1*S*,3*S*)-12e (Scheme 1). The relative configuration of the 12e diastereomers was assigned on the basis of the  ${}^{1}H{-}^{1}H$  coupling constants.<sup>13</sup> Conversion to the amides (1*R*,3*S*)- and (1S,3S)-13e was achieved with N-methylethylene diamine in MeOH at reflux under nitrogen. The remaining enantiomeric pair of diastereomers of 13e was then prepared from (S)-10 (Supporting Information). Interestingly, in the amidation of (1S,3S)- and (1R,3R)-12e, oxidized byproduct 7e was isolated in 14-17% yield (Scheme 1 and Supporting Information). To confirm its structure, 7e was independently prepared by oxidation of the mixture of the *trans-* and *cis*-esters 12e to  $\beta$ carboline 14e, using PhI(OAc)2<sup>14</sup> (Scheme 2). Amidation of 14e in neat amine and HCl treatment yielded 7e·HCl in 75% vield. With these compounds in hand, we examined their activity against asexual blood stage P. falciparum (Table 1).

None of the stereoisomers of tetrahydro- $\beta$ -carboline 13e potently inhibited the growth of *P. falciparum*. Since (1*R*,3*S*)-13e has the same configuration of 1a, the ability of IPP supplementation (200  $\mu$ M) to reverse growth inhibition was



Scheme 1. Synthesis of the Stereoisomers of VLS Hit

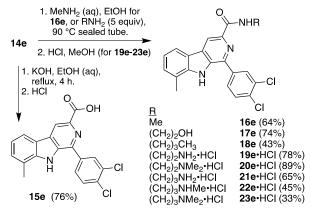
Scheme 2. Independent Synthesis of 7e·HCl



assessed; the 50% rescue seen at 5  $\mu$ M drug suggested this stereoisomer is a weak MEP pathway inhibitor. Fortuitously, the  $\beta$ -carboline amide 7e, isolated initially as a byproduct, did potently inhibit parasite growth with EC<sub>50</sub> = 108 ± 7 nM. Interestingly, supplementation with IPP did not rescue parasite growth. Thus, the antimalarial activity of 7e·HCl is not related to inhibition of the MEP pathway.

We then prepared a range of analogs of 7e that differed in the C3 carbonyl substituent (Scheme 3).

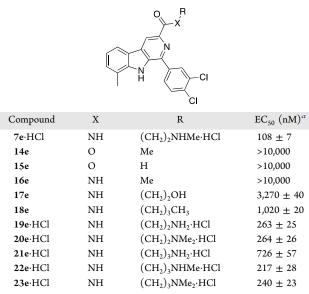
## Scheme 3. Synthesis of Acid and Amide Variants of 7e



Hydrolysis of 14e afforded carboxylic acid 15e; amidation of 14e yielded neutral amides 16e-18e and amides 19e-23e, which resemble 7e in having a pendant basic amine. These were tested for growth inhibition of *P. falciparum*, and a clear pattern emerged (Table 2). As can be seen, ester 14e and acid

 Table 2. In Vitro P. falciparum Growth Inhibition of Acid

 and Amide Variants of 7e

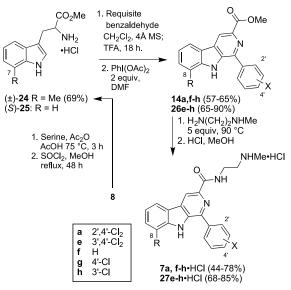


<sup>*a*</sup>Growth inhibition (72 h) of asexual blood stages determined using SYBR Green I assay. Values represent average  $\pm$  SEM from at least three independent biological replicates. The Dd2 strain is resistant to chloroquine, pyrimethamine, and mefloquine.

15e did not inhibit growth at the highest concentration tested, and only amides with a pendant basic amine (7e and 19e - 23e) had submicromolar potency. Within the limited set of compounds explored, it appears that the *N*-(2-methylamino)-ethyl amide (i.e., 7e) confers the best antimalarial potency.

We thus prepared a set of analogs of 7e·HCl that differed in the 8-substituent of the A-ring and in the substitution of the D-ring (Scheme 4).

## Scheme 4. Synthesis of A- and D-Ring Variants of 7e•HCl



Racemic 7-methyl tryptophan methyl ester  $(\pm)$ -24 was prepared from 8 by adapting a literature method;<sup>15</sup> it and (S)-25 were subjected to Pictet–Spengler reaction with the requisite benzaldehydes; oxidation with PhI(OAc)<sub>2</sub> yielded the corresponding  $\beta$ -carboline methyl esters 14a,f–h and 26e–h. Amidation and treatment with methanolic HCl afforded the desired A- and D-ring variants of 7e·HCl. The antimalarial potencies of these compounds and 7e·HCl are shown in Table 3.

Table 3. Antimalarial Potencies of 7e•HCl and Its A-/D-Ring Variants

8			
	NHMe•HCI		
	Q	NH NH	
	R R	N X	
Compound	R	Х	$EC_{50} (nM)^a$
7a·HCl	CH <sub>3</sub>	2',4'-Cl <sub>2</sub>	$583 \pm 91$
7e∙HCl	CH <sub>3</sub>	3',4'-Cl <sub>2</sub>	$108 \pm 7$
7f·HCl	CH <sub>3</sub>	Н	$4,000 \pm 900$
7 <b>g</b> ∙HCl	CH <sub>3</sub>	4'-Cl	$274 \pm 12$
7h·HCl	CH <sub>3</sub>	3'-Cl	$342 \pm 7$
27e·HCl	Н	3',4'-Cl <sub>2</sub>	$227 \pm 33$
27f·HCl	Н	Н	2,500-5,000
27g·HCl	Н	4'-Cl	496 ± 60
27h·HCl	Н	3'-Cl	641 ± 11

<sup>*a*</sup>Growth inhibition (72 h) of asexual blood stages determined using SYBR Green I assay. Values represent average  $\pm$  SEM from at least three independent biological replicates. The Dd2 strain is resistant to chloroquine, pyrimethamine, and mefloquine.

As can be seen,  $3',4'-Cl_2$  substitution of the D-ring (i.e., 7e-HCl and 27e-HCl) affords the best potency in both the R = CH<sub>3</sub> and R= H series. Unsubstituted analogs 7f-HCl and 27f-HCl are approximately 10-fold less potent than 7e-HCl and 27e-HCl. Single Cl-substitutions in the 4'- or 3'-positions (e.g., 7g,h-HCl /27g,h-HCl) recover some potency relative to the

unsubstituted analogs 7f·HCl/27f·HCl but are less potent than  $3',4'-Cl_2$  analogs 7e·HCl/27e·HCl. Compound 7a·HCl provides one example of  $2',4'-Cl_2$  disubstitution (cf. *Pf*IspD inhibitor 1a), but it is less potent than 7e·HCl. Lastly, in this small series it appears that 8-methyl substitution may confer some advantage for *in vitro* potency (cf. 7e·HCl and 27e·HCl).

Since 7e·HCl emerged as the best compound, we evaluated it for potency against a number of different strains of *P*. *falciparum* (Table 4).

Table 4. In Vitro Antimalarial and ADME-Tox Data, Physicochemical Parameters, and In Vivo PK Data for 7e• HCl

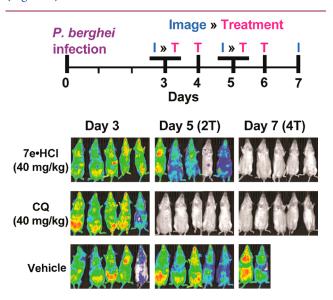
Entry	Assay	Value
1	Dd2 $EC_{50}^{72h} (nM)^{a,b}$	$108 \pm 7$
2	$3D7 EC_{50}^{72h} (nM)^{a,c}$	$53 \pm 7$
3	Dd2-KAE609 <sup>R</sup> $EC_{50}^{72h}$ (nM) <sup><i>a</i>,<i>d</i></sup>	$101 \pm 5$
4	W2 $EC_{50}^{72h}$ (nM) <sup><i>a</i>,<i>e</i></sup>	$100 \pm 8$
5	$4 \text{G EC}_{50}^{72\text{h}} (\text{nM})^{a,f}$	$110 \pm 17$
6	$3D7 {}^{3}H-H IC_{50} {}^{48h} (nM)^{g}$	$70 \pm 10$
7	$3D7 \text{ LDH IC}_{50}^{72h} (nM)^{h}$	117 ± 11
8	HEK293 CC <sub>50</sub> (nM)	32,000 ± 4,000
9	hPHep $CC_{50}^{96h}$ (nM) <sup>i</sup>	8,500 ± 1,900
10	E. coli MIC (nM)	>250,000
11	hERG IC <sub>50</sub> (nM)	510
12	Mini-Ames Panel	negative
13	MW (g/mol, free base)	427
14	tPSA (Å <sup>2</sup> )	65.5
15	Log D (pH 7.4) <sup><i>j</i></sup>	3.3
16	PBS Solubility ( $\mu$ M, pH 7.4)	9.5
17	PPB (mouse)	99.49%
18	Mouse plasma stability	100% at 1 h
19	Mouse microsomal $t_{1/2}$ (min)	110
20	$Cl_{int}$ (mouse $\mu g/mL/min$ )	12.6
21	%F $(rat)^{k}$	40%
22	$C_{\max}$ (nM, oral 40 mg/kg, rat) <sup>k</sup>	800 ± 25
23	Rat plasma $t_{1/2}$ (h) <sup>k</sup>	8
24	$V_{\rm dss} \ ({ m L/kg, rat})^k$	21.8

<sup>*a*</sup>Growth inhibition determined using the SYBR Green I assay for all  $EC_{50}$  determinations. Values represent average ± SEM from at least three independent biological replicates. <sup>*b*</sup>Resistant to chloroquine (CQ), pyrimethamine (PY), and mefloquine (MQ). <sup>*c*</sup>Sensitive to CQ, PY, and MQ. <sup>*d*</sup>KAE609-resistant strain, courtesy of E. Winzeler. <sup>*e*</sup>Resistant to CQ, quinine, PY, cycloguanil, and sulfadoxine. <sup>*f*</sup>Dihydroartemisinin- and CQ-resistant field strain, courtesy of D. E. Kyle. <sup>*g*</sup>[<sup>3</sup>H]hypoxanthine incorporation assay. <sup>*h*</sup>Lactate dehydrogenase activity. <sup>*i*</sup>Primary human hepatocytes. <sup>*j*</sup>Experimentally determined. <sup>*k*</sup>See Supporting Information for details.

Compound 7e·HCl had similar  $EC_{50}$  values against the susceptible 3D7 strain and a number of resistant strains of *P. falciparum* (Table 4, entries 1–5). Thus, 7e·HCl is apparently not subject to the resistance mechanisms of a number of antimalarials such as chloroquine, mefloquine, quinine, pyrimethamine, cycloguanil, sulfadoxazine, KAE609, and dihydroartemisinin. Furthermore, the antimalarial potency against the 3D7 strain was similar when assessed for DNA replication, purine biosynthesis, and metabolic activity (Table 4, entries 2, 6, and 7). 7e·HCl was also not generally cytotoxic, as demonstrated by 78- to 320-fold higher half-maximal cytotoxic concentration ( $CC_{50}$ ) values against mammalian cells and no effect at 250  $\mu$ M against human microbiome constituent *E. coli* (Table 4, entries 8–10). The compound

showed a hERG liability, but it was not mutagenic in a mini-Ames panel (Table 4, entries 11–12). The physicochemical properties of 7e·HCl are suggestive of acceptable oral bioavailability (Table 4, entries 13–15). The compound is highly plasma-protein-bound but has excellent plasma and microsomal stability and, consequently, low  $Cl_{int}$  (Table 4, entries 17–20). In rat, 7e·HCl has moderate oral bioavailability (40%) and a  $C_{max}$  of 800 ± 25 nM (at 40 mg/kg po); the long plasma half-life (8 h) and relatively high volume of distribution (21.8 L/kg) are also positive features for an antimalarial candidate (Table 4, entries 21–24).

With this data in hand, the *in vivo* antimalarial activity of 7e-HCl was assessed in a *P. berghei* model of murine malaria (Figure 3).



**Figure 3.** 7e·HCl (40 mg/kg/day) has oral efficacy in *P. berghei*infected mice. A transgenic *P. berghei* ANKA strain (l676m1cl1 line, PbGFP-Luccon) was used, that expresses a fusion GFP (mutant 3) and firefly luciferase (LucIAV). CQ is chloroquine. Note: three of the five vehicle-treated mice died before Day 7.

Mice were infected on Day 0 with 10<sup>3</sup> P. berghei-infected erythrocytes and treated for 4 days starting on Day 3 after inoculation with *P. berghei*. Mice were imaged (luminescence) before dosing on Days 3 and 5; final images were taken on Day 7. As can be seen in Figure 3, at 40 mg/kg (po), 7e·HCl substantially reduced parasitemia in 3-4 doses. At the same dose, chloroquine cleared infection in 1-2 doses. No signs of toxicity (lethargy, gait, weight loss, ruffled fur, and differences in behavior) were observed in any of the groups. Results were reproduced in two independent trials with independently synthesized samples of 7e-HCl at 40 mg/kg. Note that no reduction in parasitemia was seen at 20 mg/kg, using the same dosing/imaging protocol shown in Figure 3. Thus, further increases in in vitro protency will be needed. Nevertheless, to the best of our knowledge, this is the first example of oral antimalarial efficacy within the  $\beta$ -carboline (as opposed to the tetrahydro- $\beta$ -carboline) scaffold. In addition to its good druglike properties and low general toxicity, 7e·HCl has a number of other promising characteristics, which we will disclose in due course.

## ASSOCIATED CONTENT

### **Supporting Information**

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acsmedchemlett.1c00663.

Synthetic procedures, tabulations of analytical data, and NMR spectra of all tested compounds; *in vitro* growth inhibition procedures; details on the ADME-Tox assays, pharmacokinetics, and *in vivo* efficacy studies (PDF)

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### **Author Contributions**

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

#### Notes

The authors declare no competing financial interest.

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## REFERENCES

(1) World Malaria Report 2021. The World Health Organization, available at https://www.who.int/teams/global-malaria-programme/ reports/world-malaria-report-2021 (accessed 2021-12-13).

(2) Jomaa, H.; Wiesner, J.; Sanderbrand, S.; Altincicek, B.; Weidemeyer, C.; Hintz, M.; Türbachova, I.; Eberl, M.; Zeidler, J.; Lichtenthaler, H. K.; Soldati, D.; Beck, E. Inhibitors of the Nonmevalonate Pathway of Isoprenoid Biosynthesis as Antimalarial Drugs. *Science* **1999**, 285, 1573–1576.

(3) Yeh, E.; DeRisi, J. L. Chemical Rescue of Malaria Parasites Lacking an Apicoplast Defines Organelle Function in Blood-Stage *Plasmodium falciparum. PLoS. Biol.* **2011**, *9*, e1001138.

(4) Spangenberg, T.; Burrows, J. N.; Kowalczyk, P.; McDonald, S.; Wells, T. N. C.; Willis, P. The Open Access Malaria Box: A Drug Discovery Catalyst for Neglected Diseases. *PLoS One* **2013**, *8*, e62906.

(5) Bowman, J. D.; Merino, E. F.; Brooks, C. F.; Striepen, B.; Carlier, P. R.; Cassera, M. B. Antiapicoplast and Gametocytocidal Screening To Identify the Mechanisms of Action of Compounds within the Malaria Box. *Antimicrob. Agents Chemother.* **2014**, *58*, 811–819.

(6) Wu, W.; Herrera, Z.; Ebert, D.; Baska, K.; Cho, S. H.; DeRisi, J. L.; Yeh, E. A chemical rescue screen identifies a Plasmodium falciparum apicoplast inhibitor targeting MEP isoprenoid precursor biosynthesis. *Antimicrob. Agents Chemother.* **2015**, *59*, 356–364.

(7) Imlay, L. S.; Armstrong, C. M.; Masters, M. C.; Li, T.; Price, K. E.; Edwards, R. L.; Mann, K. M.; Li, L. X.; Stallings, C. L.; Berry, N. G.; O'Neill, P. M.; Odom, A. R. Plasmodium IspD (2-C-methyl-Derythritol 4-phosphate cytidyltransferase), an essential and druggable antimalarial target. ACS Infect. Dis. **2015**, *1*, 157–167.

(8) Yao, Z.-K.; Krai, P. M.; Merino, E. F.; Simpson, M. E.; Slebodnick, C.; Cassera, M. B.; Carlier, P. R. Determination of the active stereoisomer of the MEP pathway-targeting antimalarial agent MMV008138, and initial structure-activity studies. *Bioorg. Med. Chem. Lett.* **2015**, *25*, 1515–1519.

(9) Ghavami, M.; Merino, E. F.; Yao, Z.-K.; Elahi, R.; Simpson, M. E.; Fernández-Murga, M. L.; Butler, J. H.; Casasanta, M. A.; Krai, P. M.; Totrov, M. M.; Slade, D. J.; Carlier, P. R.; Cassera, M. B. Biological Studies and Target Engagement of the 2-C-Methyl-D-Erythritol 4-Phosphate Cytidylyltransferase (IspD)-Targeting Antimalarial Agent (1*R*,3*S*)-MMV008138 and Analogs. ACS Infect. Dis. **2018**, *4*, 549–559.

(10) Ding, S.; Ghavami, M.; Butler, J. H.; Merino, E. F.; Slebodnick, C.; Cassera, M. B.; Carlier, P. R. Probing the B- & C-rings of the antimalarial tetrahydro- $\beta$ -carboline MMV008138 for steric and conformational constraints. *Bioorg. Med. Chem. Lett.* **2020**, 30, 127520.

(11) Gamo, F.-J.; Sanz, L. M.; Vidal, J.; de Cozar, C.; Alvarez, E.; Lavandera, J.-L.; Vanderwall, D. E.; Green, D. V. S.; Kumar, V.; Hasan, S.; Brown, J. R.; Peishoff, C. E.; Cardon, L. R.; Garcia-Bustos, J. F. Thousands of chemical starting points for antimalarial lead identification. *Nature* **2010**, *465*, 305–310.

(12) Ross, A. J.; Lang, H. L.; Jackson, R. F. W. Much Improved Conditions for the Negishi Cross-Coupling of Iodoalanine Derived Zinc Reagents with Aryl Halides. *J. Org. Chem.* **2010**, 75, 245–248. (13) Cagašová, K.; Ghavami, M.; Yao, Z.-K.; Carlier, P. R. Questioning the  $\gamma$ -gauche effect: stereoassignment of 1,3-disubstituted-tetrahydro- $\beta$ -carbolines using 1H–1H coupling constants. Org. Biomol. Chem. **2019**, 17, 6687–6698.

(14) Kamal, A.; Tangella, Y.; Manasa, K. L.; Sathish, M.; Srinivasulu, V.; Chetna, J.; Alarifi, A. PhI(OAc)2-mediated one-pot oxidative decarboxylation and aromatization of tetrahydro- $\beta$ -carbolines: synthesis of norharmane, harmane, eudistomin U and eudistomin I. *Org. Biomol. Chem.* **2015**, *13*, 8652–8662.

(15) Konda-Yamada, Y.; Okada, C.; Yoshida, K.; Umeda, Y.; Arima, S.; Sato, N.; Kai, T.; Takayanagi, H.; Harigaya, Y. Convenient synthesis of 7' and 6'-bromo-D-tryptophan and their derivatives by enzymatic optical resolution using D-aminoacylase. *Tetrahedron* **2002**, *58*, 7851–7861.