

# Limiting factors for milk production in dairy cows: perspectives from physiology and nutrition

Josef J. Gross<sup>†</sup> 

Veterinary Physiology, Vetsuisse Faculty, University of Bern, Bremgartenstrasse 109a, CH-3012 Bern, Switzerland

<sup>†</sup>Corresponding author: [josef.gross@vetsuisse.unibe.ch](mailto:josef.gross@vetsuisse.unibe.ch)

## Abstract

Milk production in dairy cows increases worldwide since many decades. With rising milk yields, however, potential limiting factors are increasingly discussed. Particularly, the availability of glucose and amino acids is crucial to maintain milk production as well as animal health. Limitations arise from feed sources, the rumen and digestive tract, tissue mobilization, intermediary metabolism and transport, and the uptake of circulating nutrients by the lactating mammary gland. The limiting character can change depending on the stage of lactation. Although physiological boundaries are prevalent throughout the gestation–lactation cycle, limitations are aggravated during the early lactation period when high milk production is accompanied by low feed intake and high mobilization of body reserves. The knowledge about physiological constraints may help to improve animal health and make milk production more sustainably. The scope of this review is to address contemporary factors related to production limits in dairy cows from a physiological perspective. Besides acknowledged physiological constraints, selected environmental and management-related factors affecting animal performance and physiology will be discussed. Potential solutions and strategies to overcome or to alleviate these constraints can only be presented briefly. Instead, they are thought to address existing shortcomings and to identify possibilities for optimization. Despite a scientific-based view on physiological limits, we should keep in mind that only healthy animals could use their genetic capacity and produce high amounts of milk.

## Lay Summary

With increasing milk yields in dairy cows, potential limiting factors are intensively discussed. The present review addresses physiological and nutritional constraints that are considered limiting for milk production. The limiting character can change depending on the stage of lactation. Especially after parturition, the limited availability of glucose and amino acids does not only restrict lactational performance but also immune function. Further limitations imposed by feed, intestinal absorption, intermediary metabolism, and nutrient uptake by the mammary gland are described. Moreover, the impact of environmental (heat stress, photoperiod length) and management-related factors (e.g., rearing intensity, dry-period length) on milk yield are elucidated. However, the physiological constraints addressed in this review give space for improvements. Of course, boundaries are set by the farming system, climate, etc. that cannot be overcome. Efforts in improving welfare, husbandry, feeding, and management are likely to further enhance milk production, but will simultaneously improve other traits like reproductive performance and animal health. The existing variation in metabolic adaptation to different environmental stimuli provides further potential for appropriately selecting cows fitting best to the respective conditions. However, increasing yearly milk yields must not be dismissed as driving forces worsening animal health. Only healthy animals can perform well and produce high amounts of milk.

**Key words:** limits, milk production, physiology, nutrition, dairy cow

**Abbreviations:** AA, amino acid; ADF, acid detergent fiber; AT, adipose tissue; BHB, beta-hydroxybutyrate; CP, crude protein; DMI, dry matter intake; FA, fatty acid; NDF, neutral detergent fiber; NEFA, nonesterified fatty acid; PPAR, peroxisome proliferator-activated receptor; SARA, subclinical rumen acidosis; TCA, tricarboxylic acid cycle; VFA, volatile fatty acid; VLDL, very low-density lipoprotein

## Introduction

Worldwide, yearly milk production in dairy cows increases almost linearly since many decades (FAOSTAT, 2021). Currently, there seems to be no plateau or end of lactation performance in sight (Pulina et al., 2020). Brito et al. (2021) summarized figures on current genetic trends and milk yield development of, for example, Canadian or U.S. dairy breeds. Continuous selection toward higher milk yield along with improvements in management, housing, feeding, and veterinary care resulted in high-yielding dairy cows with individuals producing more than 35,000 kg of milk per year (Hoard's Dairyman, 2020). Milk production is a function of the number and activity of secretory epithelial cells (Akers, 2000; Capuco et al., 2003; Boutinaud et al., 2004), where potential limiting factors may

exert direct and indirect effects. The essential role of glucose and amino acids (AA) with regard to milk production is well recognized (Clark, 1975; Cant et al., 2002). However, how much milk can a dairy cow produce? Where are the physiological limits? These and similar questions did not arise recently, but already much earlier, when the scientific picture of the relationships between endocrine and metabolic processes and lactation outcome became more comprehensive (Bines and Hart, 1982). Already in the 1980s, researchers discussed if the cows' genetic capacity for milk production was reached and if further increases of milk yield would provoke the antagonism to fitness traits (Kennedy, 1984). Now, almost 40 years later, average milk production per cow more than doubled in many countries, and nowadays, performance level

Received December 16, 2021 Accepted February 10 2022.

© The Author(s) 2022. Published by Oxford University Press on behalf of the American Society of Animal Science. All rights reserved. For permissions, please e-mail: [journals.permissions@oup.com](mailto:journals.permissions@oup.com).

pushes us to investigate the issue when the overall capacity of the lactating dairy cow would be finally reached. Moreover, many dairy cows are suffering from health disorders that are primarily attributed to the high performance level and associated failures of metabolic and immunological adaptation, although the relationship between the occurrence of the so-called production diseases and high milk yield is not definitely clarified (Gröhn et al., 1995; Fleischer et al., 2001). The awareness and a deeper knowledge about physiological constraints may help to further improve animal health and produce milk more efficient and sustainable.

The scope of this review is to give in short an overview on various factors that are presently considered limiting milk production in dairy cows from a physiological perspective. The question how much milk a cow can actually produce cannot be answered here. Depending on the stage of lactation, the focus is set on factors that are currently considered as physiological and nutritional boundaries for milk production in general with respect to further negative impacts on animal health, reproductive performance, and environment. Besides these physiological constraints, selected exogenous environmental and management-related factors affecting animal performance and physiology are discussed. Figure 1 illustrates factors imposing limitations to milk production in dairy cows. However, this short review cannot entirely cover all physiological aspects in detail. Potential solutions and strategies to overcome or to alleviate physiological constraints can only be presented briefly in passing and are thought to address existing shortcomings and to identify possibilities for optimization. The implementation of targeted measures related to physiological limits, however, shall not primarily pursue the aim of further pushing milk production, though positive effects are very likely along with improvements of animal health, reproductive performance, etc. Despite a scientific-based view on physiological limits, we should keep in mind that only healthy animals feeling comfortable could use their genetic capacity and produce high amounts of milk.

## Feed Intake and Feed Sources

The digestive system of dairy cows is dominated by the rumen, where plant fibers (i.e., structural non-starch carbohydrates such as acid detergent fiber [ADF] and neutral

detergent fiber [NDF]) are degraded and used energetically by microbes. Furthermore, dietary crude protein (CP) sources and minerals (e.g., cobalt) support microbial growth. Shortly before parturition and particularly during the early lactation period, dry matter intake is not sufficient to cover the energetic and nutrient requirements of the cow. Furthermore, the limited capacity of rumen fill enables resumption of feed uptake only after rumen content was emptied. Depending on dietary composition, ruminal fermentation and passage rate may be limiting factors for initiating dry matter intake (DMI). On the other hand, dietary fiber degradation by rumen microbes takes time. Therefore, feed intake should be maximized whenever possible by the use of feed ingredients with a high palatability, but not lowering rumen passage rate and overall digestibility (Grummer et al., 2004).

Microbial fermentation of dietary carbohydrates provides the rumen host with volatile fatty acids (VFAs) that are mainly directed toward gluconeogenesis and milk fat synthesis (Aschenbach et al., 2010). High milk yields, particularly during early lactation, require high amounts of energy and nutrients. Relative to the energetic requirements for maintenance, the additional needs for milk production assumed at 50 to 60 kg/d of a 650-kg cow may amount to a four- to fivefold greater level (for calculations, see the recommendations for energy and nutrient supply in dairy cows, e.g., NRC, 2001). Classical types of roughage used as feedstuffs in dairy cow nutrition include grazed (e.g., pasture and rangeland) and preserved forms (e.g., silage, hay) besides crop residues and byproducts (e.g., straw, hulls from soy and sunflowers). Roughages are characterized by a relatively high fiber and rather low energy and nutrient content. Consequently, exclusive feeding of cows with even high-quality herbage and on well-managed pastures can energetically support only milk production levels up to around 30 kg/d (Bargo et al., 2003; Kolver, 2003), while greater yields are accompanied by considerable mobilization of body fat stores (see chapter below), or can be only achieved by supplementary concentrates (Zbinden et al., 2017). However, maintenance of rumen activity requires a minimum dietary fiber content in the diet, that is, physically effective ADF and NDF (Varga et al., 1998; Zebeli et al., 2012, and recommendations of NRC, 2001, and other feed evaluation systems, e.g., NorFor, 2011 or INRA, 2018). In contrast, too high amounts of starch-based concentrates

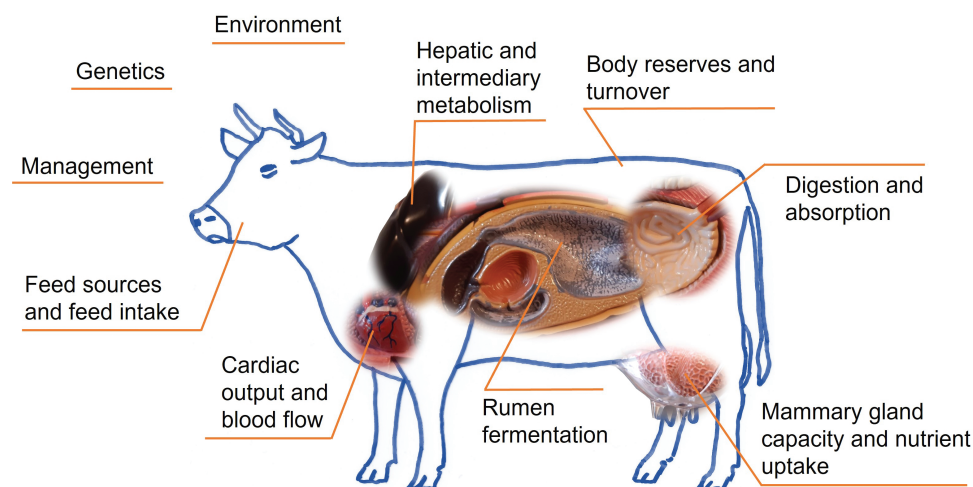


Figure 1. Factors related to limitations of milk production in dairy cows.

relative to the forage content (concentrate amount in the diet > 60%) offered in only few portions may result in (subclinical) rumen acidosis (SARA) as consequence of retarded rumination activity and lacking buffering substances from saliva (Humer et al., 2018; Neubauer et al., 2020). Rumen acidosis in turns reduces DMI and rumen passage rate. Especially in early lactating cows, a considerable risk for developing SARA emanates from the necessity of feeding high amounts of concentrates in relation to the low roughage intake to support the energy requirements of milk production. Supplementary concentrates (mainly cereals or byproducts from soybeans, rapeseed, cotton, etc.) contain high amounts of starch, protein, and fat, and are indispensable sources of additional energy and nutrients. Only the combination of different types of concentrates allows the formulation of balanced diets where nutrients can be utilized most efficiently and that can energetically support higher milk production levels compared to classical sole roughage diets. When feeding high amounts of cereal-based concentrates containing significant amounts of carbohydrates in non-rumen-protected form, the additional use of buffer substances may help (NRC, 2001) to reduce the risk of developing SARA. In general, high grain diets should include feed components that naturally show a low rumen degradability of starch (e.g., maize kernels vs. wheat). Similarly, protein-rich feedstuffs with a high degree of rumen-protection (e.g., beer draff, corn gluten) should be given preference over dietary CP sources that are rapidly degraded to ammonia in the rumen (e.g., grass). Overall and independent from feed ingredients, significant energy losses and a low efficiency are characteristic for rumen degradation processes of dietary components with the need of a subsequent resynthesis of, for example, glucose and proteins. However, diet formulation must ensure that sufficient energy and nitrogen are (synchronously) available for the rumen microbes not restricting their development and growth. Estimates on figures characterizing microbial protein synthesis in relation to energy supply can be found in NRC (2001). Otherwise, excessive CP is excreted via urine and feces, with concomitantly higher metabolic stress imposed by elevated plasma urea concentrations negatively affecting, for example, the immune system (Spek et al., 2013; Raboisson et al., 2014a). To sum up, microbial fermentation products from the rumen are alone not sufficient to cover the needs of high-yielding dairy cows, especially during early lactation. Therefore, nutrients by-passing the rumen play a significant role in current dairy cow feeding. For many years, the supply with microbial and undegraded feed protein was assumed to be sufficient in terms of quantity and AA pattern (Bines and Hart, 1982). Meanwhile, methionine, lysine, and histidine may be first limiting for milk production (Huhtanen et al. 2002) and are supplemented in rumen-protected form (Schwab and Broderick, 2017). Similarly, the limited capacity of microbial synthesis of B-vitamins and the need of an additional supplementation in high-yielding dairy cows are increasingly discussed (Seck et al., 2017).

### Digestive Tract and Nutrient Absorption

Earlier reports attested the rumen by-pass nutrients only a minor role in the supply of animal requirements (e.g., Bines and Hart, 1982). Of course, milk yields were much lower at that time compared with the current production levels. As stated in the previous chapter, due to restrictions of rumen digestive capacity and the need of higher dietary nutrient

and energy density as consequence of increased performance, high-quality and intact by-pass nutrients such as proteins, starch, and fat that are absorbed at the duodenum are becoming increasingly important to support the rising lactational needs during the first weeks of lactation. However, postruminal digestive and absorptive capacity can be assumed limited throughout the lactation period. For instance, excessive dietary starch might not be fully digested and absorbed at the duodenum, and therefore passes to the large intestine where it is subject to microbial fermentation (Owens et al., 1986). Although VFA production in the hindgut is similar to the rumen, the absorption rate of nutrients and amount of VFA produced in the hindgut are rather low, and the overall energetic efficiency of starch-based carbohydrate utilization after microbial breakdown is low (Harmon et al., 2004). However, detailed figures on the duodenal absorptive capacity of glucose, AA, and FAs are scarce. Figures on starch arrival at the duodenum and limiting factors for intestinal starch digestion in ruminants were reported by Owens et al. (1986) and Harmon et al. (2004). We can assume that current dairy cow diets containing a high proportion of concentrates (>60% in the total ration) fed along with maize silage provide even greater amounts of rumen-protected starch appearing at the duodenum as compared to older reports. Huntington et al. (2006) investigated the starch digestion in growing cattle, and estimated that up to 1.5-kg starch reached the duodenum based on earlier data published by Harmon et al. (2004). Further research is warranted to evaluate the current capacity of duodenal starch digestion and glucose absorption. Another way to alleviate energy shortages is the dietary supply with fat sources. Again, their application is commonly restricted to approximately 5 to 6% of the total diet to avoid a depression of fiber degradation and rumen passage (Rabiee et al., 2012; Weld and Armentano, 2017). Therefore, higher amounts of dietary fat should be provided in rumen-protected form. More details on the regulation and capacity of FA absorption were recently published by Bionaz et al. (2020). Besides a minimal and local use of some nutrients (e.g., glucose or short-chain fatty acid in enterocytes), the majority of absorbed nutrients is drained from the intestinal tissues by the portal vein and lymph system, transported to the liver, and distributed across the body. Physiological limitations in terms of transport capacities are currently not known for dairy cows.

### Body Reserves, Body Tissue Turnover, and Intermediary Metabolism

Besides dietary contributions to the overall energy and nutrient pool, body tissues (e.g., adipose tissue [AT], skeletal muscles, bones) represent a dynamic reservoir of various nutrients (e.g., minerals like calcium and phosphorus from bones, AA from skeletal muscle degradation). The amount of stored nutrients seems not to be a limiting factor per se. However, their turnover temporarily imposes physiological boundaries to the organism (Bauman and Currie, 1980; Drackley, 1999). For instance, several kilograms of calcium are stored in the skeleton. However, the abrupt increase of calcium requirements for milk production at onset of lactation exceeds the circulating calcium pool in plasma consisting of only a few grams (Goff, 2014). Consequently, the delayed endocrine adaptation (including calcitonin, parathyroid hormone, vitamin D, serotonin, etc.) to sufficiently provide calcium may result in hypocalcemia that is associated with reduced peak milk yield,

DMI, and an elevated risk for the development of further production diseases such as displaced abomasum, ketosis, and mastitis (Reinhardt et al., 2011; Venjakob et al., 2017). Approximately 2 to 3 d postpartum, a coordinated regulation of calcium homeostasis with mobilization, absorption, and excretion is widely achieved (Hernández-Castellano et al., 2017a,b; Kessler et al., 2018).

According to literature reports, the amount of mobilizable body fat (mainly stored in visceral and subcutaneous AT) accounts up to 120 kg (Chilliard, 1992; Raschka et al., 2016). For compensation of the negative energy balance (NEB) after parturition, AT lipids and skeletal muscle proteins are mobilized (Grummer, 1993; Drackley, 1999; Gross et al., 2011). For a more details on the physiology of AT, readers might refer to other excellent research articles and reviews providing more details (e.g., Mann et al., 2016; McNamara et al., 2016). However, overconditioning of cows during the preceding dry period must not be equated with a larger reservoir of body reserves. Instead, overconditioning adversely affects animal health after parturition as cows with a high body condition score substantially mobilize more fat tissues compared to leaner cows (Locher et al., 2015; Schuh et al., 2019). This aggravates metabolic load as lipolysis occurs at an excessive level likely resulting in the development of steatosis and ketosis (Bobe et al., 2004). Concomitantly and most relevant for physiological limits of milk production during early lactation, cows suffering from ketosis show a depression of DMI and a reduced milk production (Rukkwamsuk et al., 1999; Raboisson et al., 2014b). Furthermore, the characteristically elevated concentrations of nonesterified fatty acid (NEFA) and ketone bodies in first weeks of lactation (Gross et al., 2011) impair the animal's immune system (Suriyasathaporn et al., 1999; Sordillo et al., 2009). Therefore, it is not surprising that the highest incidence rates for metabolic and infectious diseases such as ketosis, fatty liver disease, mastitis, and lameness can be observed during this early lactation period (Ingvartsen, 2006; Ingvartsen and Moyes, 2013; Vergara et al., 2014; Brunner et al., 2019).

Low insulin and elevated bST concentrations trigger the lipolysis of triglycerides stored in adipocytes (Bines and Hart, 1982; Campbell and Scanes, 1988). Concomitantly, the high metabolic priority of the lactating mammary gland after parturition and the prevailing low insulin sensitivity of peripheral tissues widely inhibit a re-uptake of glucose and FA into AT (De Koster and Opsomer, 2013; Gross and Bruckmaier, 2019). Among other enzymes involved in lipolysis, hormone-sensitive lipase is activated (Greenberg et al., 2001; Locher et al., 2011; van der Kolk et al., 2017), and glycerol (used for gluconeogenesis) and FA are released. Hence, we assume that there are no rate-limiting steps in lipolysis and skeletal muscle breakdown that would restrict milk production. Instead, we are facing the problem that NEFA are excessively released after parturition (e.g., in overconditioned dairy cows, or cows with a severe NEB; Locher et al., 2015; Schuh et al., 2019). Released FA are primarily transported to the liver, but FA oxidation also occurs to a lower extent in skeletal muscles (Schäff et al., 2013). Increasing NEFA concentrations activate the key regulators of FA oxidation such as transcription factors of the peroxisome proliferator-activated receptor (PPAR) family that trigger further processes related to FA transport, uptake, and finally oxidation (Desvergne et al., 2006; van der Kolk et al., 2017). Here, carnitine is an essential substrate for the transport of long-chain FAs across the inner mitochondrial

membrane, where FA are subject of  $\beta$ -oxidation after their activation with coenzyme A (Longo et al., 2016). For more information on the role of PPARs, carnitine, and  $\beta$ -oxidation, please, refer to reviews going more into detail (e.g., Desvergne et al., 2006; Ringseis and Eder, 2011; van der Kolk et al., 2017; Ringseis et al., 2018). However, the capacity of the liver to completely oxidize NEFA during the early lactation period is limited (Armentano et al., 1991). We can identify two major limiting factors: 1) limited availability of carnitine during and capacity of the carnitine shuttle system for importing FA, 2) limited capacity for  $\beta$ -oxidation and subsequent degradation of formed acetyl-CoA in the tricarboxylic acid (TCA) cycle. Carnitine is synthesized from lysine and methionine, i.e., two AAs that are currently considered first limiting for milk production as well. Research showed that the supplementation of l-carnitine could enhance  $\beta$ -oxidation and reduce hepatic triglyceride accumulation in the liver (Carlson et al., 2007; Ringseis et al., 2018). Limiting substrate for the TCA cycle capacity is oxaloacetate that is simultaneously required for gluconeogenesis (White, 2015). Glucogenic precursors such as propionate, and AAs such as alanine, cysteine, glycine, serine, and threonine can enter the TCA cycle and do further support gluconeogenesis (Aschenbach et al., 2010).

In recent years, there is increasing evidence that FA from lipolysis may impair insulin sensitivity in dairy cows (McFadden and Rico, 2019). Particularly, the role of ceramide in the regulation of energy expenditure and nutrient partitioning (via modulating insulin signaling and interaction with fibroblast growth factor 21 and adiponectin; McFadden and Rico, 2019; McFadden, 2020) requires further research. Davis et al. (2021) observed a positive correlation between plasma ceramide concentrations and milk yield.

Nevertheless, the excessively released FAs temporally exceed the hepatic capacity for oxidation during early lactation. Alternatively, abundant NEFA are re-esterified to triglycerides and remain in the liver. However, extreme lipid accumulation in the liver results in steatosis that is associated with low feed intake, milk yield, and reproductive performance (Bobe et al., 2004; Gross et al., 2013). Fatty liver develops particularly during early lactation when the hepatic export of triglycerides via very low-density lipoprotein (VLDL), whose transport capacity is low in dairy cows anyway, is additionally reduced (Gross et al., 2013; Kessler et al., 2014). Furthermore, plasma concentrations of cholesterol and lipoproteins such as low-density lipoprotein, high-density lipoprotein, and VLDL are low during early lactation, and despite the upregulation of rate-limiting and further enzymes involved in cholesterol homeostasis (Schlegel et al., 2012; Kessler et al., 2014; Gross et al., 2015a), the crucial role of cholesterol and apolipoproteins in hepatic triglyceride export could be speculated to be another limiting factor for milk production in dairy cow physiology.

A second alternative pathway eliminating abundant FA from circulation is the formation of ketone bodies such as acetone and beta-hydroxybutyrate (BHB). Hyperketonemia may result in subclinical and clinical ketosis with an associated depression of milk production, reduced reproductive success, and DMI (McArt et al., 2012). A reduction of DMI further aggravates the energy deficiency and enhances lipolysis of AT with the consequence of even more NEFA arriving at the liver and overwhelming the local oxidation capacities. Just as serious are the inhibiting effects of NEFA and BHB on immune cells (Suriyasathaporn et al., 1999).

Recently, elevated concentrations of ketone bodies in plasma were shown to reduce glucose concentrations independently from the stage of lactation (Zarrin et al., 2013, 2017). Thus, the low glucose concentrations in early lactation are likely to further decline and impose a physiological boundary for milk production that is depending on this essential substrate.

Glycogenolysis and gluconeogenesis are directly related to circulating glucose levels. Whereas glycogen stores in liver and skeletal muscle are very limited, gluconeogenesis in dairy cows is of major importance besides duodenal absorption of glucose derived from the intestinal starch digestion (Aschenbach et al., 2010). Figures on glucose production and needs of high-yielding dairy cows can be found in the review paper of Aschenbach et al. (2010). Whereas lactate and glycerol appear significant substrates for gluconeogenesis during the NEB in early lactation, particularly propionate from the rumen fermentation and AAs (both from intestinal absorption and muscle breakdown) are of importance. Despite an upregulation of gluconeogenesis between late pregnancy and early lactation, glucose concentrations are at a low level during the very first weeks of lactation, where milk production is concomitantly increasing (Hötger et al., 2013). Given its essential role for lactose production and milk fat synthesis, glucose is undoubtedly a limiting factor for daily milk production. More than 85% of the glucose produced are directed to the mammary gland to support lactation (Bickerstaffe et al., 1974; Chaiyabutr et al., 1980). However, providing more glucose does not necessary increase daily milk yield (Al-Trad et al., 2009). Instead, other organs, tissues, and functional compartments that are concomitantly competing for glucose (e.g., the immune system, reproductive tissues) are benefitting from an improved glucose metabolism (Trevisi et al., 2012; Thompson-Crispi et al., 2014; Bradford et al., 2015). At the same time, lipolysis of AT is reduced and ketone body concentrations are lower when glucose concentrations are elevated (Grossen-Rösti et al., 2018; Malacco et al., 2020). Dietary supply with glucogenic precursors during the NEB as well as glucose sparing agents such as conjugated linoleic acids that depress milk fat synthesis and thus glucose output could ameliorate glucose shortages after parturition (Bauman et al., 2008; Larsen and Kristensen, 2013; Ma et al., 2015).

Skeletal muscle serves as an important source of AA for milk synthesis, particularly during its mobilization in early lactation. The balance between mobilization and regeneration of muscle tissue is controlled among other factors via the mammalian target of rapamycin and the ubiquitin–proteasome system (Sadri et al., 2016; Yang et al., 2020). Their interaction with the endocrine regulation of tissue turnover (e.g., by insulin) must be considered in the dynamic regulation of nutrient partitioning between the mammary gland and other peripheral tissues.

### Blood Flow and Nutrient Uptake by the Mammary Gland

Absorbed and de novo synthesized nutrients are transported and distributed via the blood stream to the target tissues including the mammary gland. Besides concentrations of substrates in blood, the mammary blood flow determines the supply of the mammary gland (Davis and Collier, 1985). It is noteworthy that cardiac output and blood flow are not constant throughout the lactation–gestation cycle. Substantial increases in both mammary blood flow and cardiac output

were observed about the time of parturition (Linzell, 1974; Götze et al., 2010) affecting the exchange of nutrients across the capillaries into the mammary gland for the subsequent synthesis of milk components (Prosser et al., 1996). In addition, vascularization of the mammary gland increases during pregnancy (Yasugi et al., 1989; Matsumoto et al., 1992; Djonov et al., 2001). Although the endocrine mechanisms involved in vascularization and blood flow are out of the scope of the present review, the use of bovine somatotropin is of practical relevance as it increases both cardiac output and mammary blood flow accompanied by an increase of milk production (Peel and Bauman, 1987). However, an elevated mammary blood flow does not causally increase daily milk yield (Lacasse and Prosser, 2003; Brown and Allen, 2013). Many other limiting factors as partly outlined here must be taken into account. As pointed out by Davis and Collier (1985) and Komatsu et al. (2005), the rate of nutrient transport into the mammary epithelial cells rather than delivery to the cells may be another rate-limiting step for milk production. Experimental data showed that elevated glucose concentrations in plasma did not linearly increase glucose entry rate into the mammary gland (Cant et al., 2002; Xiao and Cant, 2005). There are a number of specific transport systems enabling the uptake of AAs, glucose, or FAs into the mammary gland (Miller et al., 1991). For further details, readers are advised to refer to recent experimental and review papers (e.g., Zhao and Keating, 2007a, b; Gutgesell et al., 2009; Cant et al., 2018). Besides substrate availability, also kinetics and endocrine regulation of transporters depending on the physiological stage of the cow must be assumed to be limiting for milk synthesis. For instance, glucose transporters express a differential sensitivity toward endocrine mediators such as insulin manifesting in, for example, insulin-dependent and insulin-independent representatives (Zhao and Keating, 2007a, b). Furthermore, their expression within a certain tissue such as the mammary gland or AT changes with the lactational stage of the cow (Mattmiller et al., 2011; Jaakson et al., 2018; Karis et al., 2020). Consequently, overall tissue responsiveness and glucose utilization can be regulated and adjusted to the current needs and substrate availability (Gross et al., 2015b; Karis et al., 2020).

### Environmental Factors Limiting Milk Production

Independent from a balanced diet formulation, animal health, and genetic capacity, there are a number of environmental factors restricting milk production. In this article, two examples, the impact of heat stress and photoperiod shall be addressed only shortly as many excellent reviews on these topics are available (e.g., Kadzere et al., 2002; Dahl, 2008; Dahl et al., 2012; Roth, 2020). Climate change is an emerging challenge that aggravates heat stress conditions for dairy cows in many regions worldwide (Gauly and Ammer, 2020). Therefore, heat stress must be considered a major threat imposing limits to milk production as it decreases not only daily milk yield, but also DMI, animal health, and reproductive key figures (Roth, 2020). It should be noted that independent from the ambient temperature higher daily milk yields go along with more endogenous heat production originating from the digestive tract and metabolism. Certainly, the reduction of DMI and rumen fermentation due to heat stress can be considered the major drivers for reducing milk production as less nutrients can be

supplied for milk synthesis. Concomitantly with the decrease in DMI, a transient NEB arises that is accompanied by a mobilization of body reserves as described above. Cattle breeds with a greater heat tolerance (e.g., cross-bred cows) could be an alternative to keep dairying in regions with increasing heat stress days (Gaughan et al., 2010; Freitas et al., 2021). Furthermore, cooling devices or increasing dietary energy content via supplementary fat may be considered supporting means to alleviate thermal stress (Sammad et al., 2020; Levit et al., 2021). However, special attention must be paid to the use of dietary components accounting for the maintenance of rumen function, that is, avoidance of SARA, use of rumen-protected nutrients, etc.

During recent years, the impact of photoperiod length on milk production was extensively studied (Dahl et al., 2000, 2012; Lacasse and Petitclerc, 2021). Whereas long-day photoperiods had a persistent effect on daily milk production during established lactation, short-day photoperiods during the dry-period resulted in a greater milk production in the subsequent lactation (Dahl et al., 2000; Dahl, 2008). The underlying endocrine mechanisms have been elucidated during the last years (e.g., Dahl et al., 2012; Lacasse et al., 2014; Ponchon et al., 2017). Besides positive effects of daylight exposure on milk yield, calcium homeostasis around parturition is positively affected (Özçelik et al., 2017).

### Genetic Capacity and Variation of Metabolic Adaptation

Given the enormous differences in dairy cow feeding, housing, and management conditions, it is hardly possible to determine the maximum genetic capacity for yearly milk production of dairy cows. Cows of breeds intensively selected toward milk production such as Holsteins are likely to produce more milk than dual-purpose or beef cows, although long-term selection of strains within a breed kept under different environmental conditions may result in a divergent lactational performance (e.g., North American vs. New Zealand Holstein strains; Lucy et al., 2009; Grala et al., 2011).

Moreover, a considerable variation of metabolic adaptation to identical feeding and management conditions exists among animals within a herd although selection targets were identical (Kessel et al., 2008; Gross and Bruckmaier, 2015). Therefore, it is not surprising that metabolic stress is not necessarily related to the overall lifetime performance (Gross et al., 2016). Obviously, there is no negative relationship between increased yearly milk production, disease occurrence, and culling, although culling risk increases in high-yielding dairy cows with a greater likelihood to develop health disorders (Beaudeau et al., 2000; Rilanto et al., 2020). Higher milk yields are undoubtedly more demanding in terms of dairy cow nutrition and management. Furthermore, a greater milk production is associated with a greater metabolic load particularly in early lactation, which increases the risk and susceptibility toward different production diseases (Ingvarstsen et al., 2003; McArt and Neves, 2020; Macmillan et al., 2021).

### Management Aspects and Epigenetic Regulation of Mammary Gland Capacity

In addition to all listed factors above, management of dairy cows exerts significant constraints on milk production. Epigenetic regulation of milk production in dairy cows was

comprehensively described by Singh et al. (2010). Early metabolic and fetal programming, respectively, may affect the production outcome of the offspring. Recently, studies showed that e.g., heat stress in pregnant dairy cows impairs lifetime milk production of daughters and granddaughters (Dado-Senn et al., 2020; Laporta et al., 2020). Also at an early stage of life in calves, feeding level with milk until weaning determines their future milk production (Soberon and van Amburgh, 2013; Korst et al., 2017). In comparison with calves receiving a restricted amount of milk, calves fed milk or milk replacer ad libitum produced more milk during their first lactation (Soberon et al., 2012; Korst et al., 2017). Furthermore, the rearing intensity of heifers during the pre-pubertal period impacts the future lifetime performance (Sejrsen and Purup, 1997). Being more specific, overfeeding during this phase promotes the development of greater fat pad instead of parenchymal tissue due to lower circulating bST concentrations (Sejrsen et al., 1982; Purup et al., 2000; Akers, 2017a). For further details on mammary gland development and function, the reader may consult the reviews of Akers (2017a, b).

Increasing milking frequency (e.g., three or more times vs. two times daily) was shown to increase daily milk production (Wall and McFadden, 2012; Ferneborg et al., 2017), whereas once-daily milking reduced daily milk yield in dairy cows (Stelwagen et al., 2013). Similarly, cows in automatic milking systems are milked on average more than two times daily and have a higher milk yield than cows milked only two times a day (Tse et al., 2018).

The impact of dry period length on dairy cow health and milk yield in the subsequent lactation was shown in a number of research and review papers (van Knegsel et al., 2013; Chen et al., 2016; Kok et al., 2017). This aspect gained more attention in dairy cow management as with the overall increasing milk yields during the last decades cows needed to be dried-off while still producing considerable amounts of milk. Not only economic interests are decisive for adjusting dry period length. Furthermore, the start into lactation is a challenging period for dairy cows, particularly for overconditioned dairy cows. Hence, a shortening or omission of the dry period goes along with a reduced milk production but concomitantly lowers the metabolic load after parturition in the subsequent lactation (Chen et al., 2016).

### Conclusions and Outlook

Currently, selection targets in dairy cows consider primarily traits related to longevity and fitness and are less oriented toward higher performance. However, milk production will further continue to increase as the physiological constraints addressed in this review give space for improvements. Of course, boundaries are set by the farming system, climate, etc. that cannot be overcome. Overall, dietary nutrient and energy availability seems the major limiting factor for milk production. Hence, glucose and AAs are the most limiting nutrients determining the amounts of milk volume and milk components, but also essential for the immune system. Efforts in improving welfare, husbandry, feeding, and management are likely to further enhance milk production, but will simultaneously improve other traits like reproductive performance and animal health. The existing variation in metabolic adaptation to different environmental stimuli provides further potential for appropriately selecting cows fitting best to the

respective conditions. However, increasing milk yields must not be dismissed as driving forces worsening animal health. Only healthy animals can perform well and produce high amounts of milk.

## Conflict of interest statement

The author declares no conflicts of interest.

## Literature Cited

- Akers, R. M. 2000. Selection for milk production from a lactation biology viewpoint. *J. Dairy Sci.* 83:1151–1158. doi:10.3168/jds.S0022-0302(00)74981-2
- Akers, R. M. 2017a. Triennial lactation Symposium/Bolfa: plasticity of mammary development in the prepubertal bovine mammary gland. *J. Anim. Sci.* 95:5653–5663. doi:10.2527/jas2017.1792
- Akers, R. M. 2017b. A 100-year review: mammary development and lactation. *J. Dairy Sci.* 100:10332–10352. doi:10.3168/jds.2017-12983
- Al-Trad, B., K. Reisberg, T. Wittek, G. B. Penner, A. Alkaassem, G. Gäbel, M. Füll, and J. R. Aschenbach. 2009. Increasing intravenous infusions of glucose improve body condition but not lactation performance in midlactation dairy cows. *J. Dairy Sci.* 92:5645–5658. doi:10.3168/jds.2009-2264
- Armentano, L. E., R. R. Grummer, S. J. Bertics, T. C. Skaar, and S. S. Donkin. 1991. Effects of energy balance on hepatic capacity for oleate and propionate metabolism and triglyceride secretion. *J. Dairy Sci.* 74:132–139. doi:10.3168/jds.S0022-0302(91)78153-8
- Aschenbach, J. R., N. B. Kristensen, S. S. Donkin, H. M. Hammon, and G. B. Penner. 2010. Gluconeogenesis in dairy cows: the secret of making sweet milk from sour dough. *IUBMB Life.* 62:869–877. doi:10.1002/iub.400
- Bargo, F., L. D. Muller, E. S. Kolver, and J. E. Delahoy. 2003. Invited review: production and digestion of supplemented dairy cows on pasture. *J. Dairy Sci.* 86:1–42. doi:10.3168/jds.S0022-0302(03)73581-4
- Bauman, D. E., and W. B. Currie. 1980. Partitioning of nutrients during pregnancy and lactation: a review of mechanisms involving homeostasis and homeorhesis. *J. Dairy Sci.* 63:1514–1529. doi:10.3168/jds.S0022-0302(80)83111-0
- Bauman, D. E., J. W. Perfield, 2nd, K. J. Harvatine, and L. H. Baumgard. 2008. Regulation of fat synthesis by conjugated linoleic acid: lactation and the ruminant model. *J. Nutr.* 138:403–409. doi:10.1093/jn/138.2.403
- Beaudeau, F., H. Seegers, V. Ducrocq, C. Fourichon, and N. Bareille. 2000. Effect of health disorders on culling in dairy cows: a review and a critical discussion. *Ann. Zootech.* 49:293–311. doi:10.1051/animres:2000102
- Bickerstaffe, R., E. F. Anison, and J. L. Linzell. 1974. The metabolism of glucose, acetate, lipids and amino acids in lactating dairy cows. *J. Agric. Sci.* 82:71–85. doi:10.1017/S0021859600050243
- Bines, J. A., and I. C. Hart. 1982. Metabolic limits to milk production, especially roles of growth hormone and insulin. *J. Dairy Sci.* 65:1375–1389. doi:10.3168/jds.S0022-0302(82)82358-8
- Bionaz, M., E. Vargas-Bello-Pérez, and S. Busato. 2020. Advances in fatty acids nutrition in dairy cows: from gut to cells and effects on performance. *J. Anim. Sci. Biotechnol.* 11:110. doi:10.1186/s40104-020-00512-8
- Bobe, G., J. W. Young, and D. C. Beitz. 2004. Invited review: pathology, etiology, prevention and treatment of fatty liver in dairy cows. *J. Dairy Sci.* 87:3105–3124. doi:10.3168/jds.S0022-0302(04)73446-3
- Boutinaud, M., J. Guinard-Flamenta, and H. Jammes. 2004. The number and activity of mammary epithelial cells, determining factors for milk production. *Reprod. Nutr. Dev.* 44:499–508. doi:10.1051/rnd:2004054
- Bradford, B. J., K. Yuan, J. K. Farney, L. K. Mamedova, and A. J. Carpenter. 2015. Invited review: inflammation during the transition to lactation: new adventures with an old flame. *J. Dairy Sci.* 98:6631–6650. doi:10.3168/jds.2015-9683
- Brito, L. F., N. Bedere, F. Douhard, H. R. Oliveira, M. Arnal, F. Peñaricano, A. P. Schinckel, C. F. Baes, and F. Miglior. 2021. Review: Genetic selection of high-yielding dairy cattle toward sustainable farming systems in a rapidly changing world. *Animal* 15(Suppl 1):100292. doi:10.1016/j.animal.2021.100292
- Brown, W. E., and M. S. Allen. 2013. Effects of intrajugular glucose infusion on feed intake, milk yield, and metabolic responses of early postpartum cows fed diets varying in protein and starch concentration. *J. Dairy Sci.* 96:7132–7142. doi:10.3168/jds.2013-6636
- Brunner, N., S. Groeger, J. Canelas Raposo, R. M. Bruckmaier, and J. J. Gross. 2019. Prevalence of subclinical ketosis and production diseases in dairy cows in central and South America, Africa, Asia, Australia, New Zealand, and eastern Europe. *Transl. Anim. Sci.* 3:84–92. doi:10.1093/tas/txy102
- Campbell, R. M., and C. G. Scanes. 1988. Inhibition of growth hormone-stimulated lipolysis by somatostatin, insulin, and insulin-like growth factors (somatomedins) in vitro. *Proc. Soc. Exp. Biol. Med.* 189:362–366. doi:10.3181/00379727-189-42819
- Cant, J. P., J. J. M. Kim, S. R. L. Cieslar, and J. Doelman. 2018. Symposium review: amino acid uptake by the mammary glands: where does the control lie? *J. Dairy Sci.* 101:5655–5666. doi:10.3168/jds.2017-13844
- Cant, J. P., D. R. Trout, F. Qiao, and N. G. Purdie. 2002. Milk synthetic response of the bovine mammary gland to an increase in the local concentration of arterial glucose. *J. Dairy Sci.* 85:494–503. doi:10.3168/jds.S0022-0302(02)74100-3
- Capuco, A. V., S. E. Ellis, S. A. Hale, E. Long, R. A. Erdman, X. Zhao, and M. J. Paape. 2003. Lactation persistency: insights from mammary cell proliferation studies. *J. Anim. Sci.* 81(Suppl 3):18–31. doi:10.2527/2003.81suppl\_318x
- Carlson, D. B., J. W. McFadden, A. D'Angelo, J. C. Woodworth, and J. K. Drackley. 2007. Dietary l-carnitine affects periparturient nutrient metabolism and lactation in multiparous cows. *J. Dairy Sci.* 90:3422–3441. doi:10.3168/jds.2006-811
- Chaiyabutr, N., A. Faulkner, and M. Peaker. 1980. The utilization of glucose for the synthesis of milk components in the fed and starved lactating goat in vivo. *Biochem. J.* 186:301–308. doi:10.1042/bj1860301
- Chen, J., G. J. Remmelink, J. J. Gross, R. M. Bruckmaier, B. Kemp, and A. T. M. van Knegsel. 2016. Effects of dry period length and dietary energy source on milk yield, energy balance, and metabolic status of dairy cows over two consecutive years: effects in the second year. *J. Dairy Sci.* 99:4826–4838. doi:10.3168/jds.2015-10742
- Chilliard, Y. 1992. Physiological constraints to milk production: factors which determine nutrient partitioning, lactation persistency, and mobilization of body reserves. *World Rev. Anim. Prod.* 27:19–26.
- Clark, J. H. 1975. Lactational responses to postruminal administration of proteins and amino acids. *J. Dairy Sci.* 58:1178–1197. doi:10.3168/jds.S0022-0302(75)84696-0
- Dado-Senn, B., J. Laporta, and G. E. Dahl. 2020. Carry over effects of late-gestational heat stress on dairy cattle progeny. *Theriogenology.* 154:17–23. doi:10.1016/j.theriogenology.2020.05.012
- Dahl, G. E. 2008. Effects of short day photoperiod on prolactin signaling in dry cows: a common mechanism among tissues and environments? *J. Anim. Sci.* 86(13 Suppl):10–14. doi:10.2527/jas.2007-0311
- Dahl, G. E., B. A. Buchanan, and H. A. Tucker. 2000. Photoperiodic effects on dairy cattle: a review. *J. Dairy Sci.* 83:885–893. doi:10.3168/jds.S0022-0302(00)74952-6
- Dahl, G. E., S. Tao, and I. M. Thompson. 2012. Lactation biology symposium: effects of photoperiod on mammary gland development and lactation. *J. Anim. Sci.* 90:755–760. doi:10.2527/jas.2011-4630
- Davis, S. R., and R. J. Collier. 1985. Mammary blood flow and regulation of substrate supply for milk synthesis. *J. Dairy Sci.* 68:1041–1058. doi:10.3168/jds.S0022-0302(85)80926-7

- Davis, A. N., W. A. Myers, C. Chang, B. N. Tate, J. E. Rico, M. Moniruzzaman, N. J. Haughey, and J. W. McFadden. 2021. Somatotropin increases plasma ceramide in relation to enhanced milk yield in cows. *Domest Anim. Endocrinol.* 74:106480. doi:10.1016/j.domaniend.2020.106480
- De Koster, J. D., and G. Opsomer. 2013. Insulin resistance in dairy cows. *Vet. Clin. North Am. Food Anim. Pract.* 29:299–322. doi:10.1016/j.cvfa.2013.04.002
- Desvergne, B., L. Michalik, and W. Wahli. 2006. Transcriptional regulation of metabolism. *Physiol. Rev.* 86:465–514. doi:10.1152/physrev.00025.2005
- Djonov, V., A. C. Andres, and A. Ziemiecki. 2001. Vascular remodelling during the normal and malignant life cycle of the mammary gland. *Microsc. Res. Tech.* 52:182–189. doi:10.1002/1097-0029(20010115)52:2<182::AID-JEMT1004>3.0.CO;2-M
- Drackley, J. K. 1999. ADSA Foundation scholar award. Biology of dairy cows during the transition period: the final frontier? *J. Dairy Sci.* 82:2259–2273. doi:10.3168/jds.s0022-0302(99)75474-3
- FAOSTAT. 2021. Statistics Division. Food and Agriculture Organization of the United Nations. [Accessed December 14, 2021]. <http://www.fao.org/faostat/en/#data>.
- Ferneborg, S., L. Kovac, K. J. Shingfield, and S. Agenäs. 2017. Effect of increased milking frequency and residual milk removal on milk production and milk fatty acid composition in lactating cows. *J. Dairy Res.* 84:453–463. doi:10.1017/S0022029917000681
- Fleischer, P., M. Metzner, M. Beyerbach, M. Hoedemaker, and W. Kleer. 2001. The relationship between milk yield and the incidence of some diseases in dairy cows. *J. Dairy Sci.* 84:2025–2035. doi:10.3168/jds.S0022-0302(01)74646-2
- Freitas, P. H. F., Y. Wang, P. Yan, H. R. Oliveira, F. S. Schenkel, Y. Zhang, Q. Xu, and L. F. Brito. 2021. Genetic diversity and signatures of selection for thermal stress in cattle and other two *Bos* species adapted to divergent climatic conditions. *Front. Genet.* 12:604823. doi:10.3389/fgene.2021.604823
- Gaughan, J. B., T. L. Mader, S. M. Holt, M. L. Sullivan, and G. L. Hahn. 2010. Assessing the heat tolerance of 17 beef cattle genotypes. *Int. J. Biometeorol.* 54:617–627. doi:10.1007/s00484-009-0233-4
- Gauly, M., and S. Ammer. 2020. Review: Challenges for dairy cow production systems arising from climate changes. *Animal.* 14(Suppl 1):s196–s203. doi:10.1017/S1751731119003239
- Goff, J. P. 2014. Calcium and magnesium disorders. *Vet. Clin. North Am. Food Anim. Pract.* 30:359–381. doi:10.1016/j.cvfa.2014.04.003
- Götze, A., A. Honnens, G. Flachowsky, and H. Bollwein. 2010. Variability of mammary blood flow in lactating Holstein-Friesian cows during the first twelve weeks of lactation. *J. Dairy Sci.* 93:38–44. doi:10.3168/jds.2008-1781
- Grala, T. M., M. C. Lucy, C. V. C. Phyn, A. J. Sheahan, J. M. Lee, and J. R. Roche. 2011. Somatotropic axis and concentrate supplementation in grazing dairy cows of genetically diverse origin. *J. Dairy Sci.* 94:303–315. doi:10.3168/jds.2010-3773
- Greenberg, A. S., W. J. Shen, K. Muliro, S. Patel, S. C. Souza, R. A. Roth, and F. B. Kraemer. 2001. Stimulation of lipolysis and hormone-sensitive lipase via the extracellular signal-regulated kinase pathway. *J. Biol. Chem.* 276:45456–45461. doi:10.1074/jbc.M104436200
- Gröhn, Y. T., S. W. Eicker, and J. A. Hertl. 1995. The association between previous 305-day milk yield and disease in New York State dairy cows. *J. Dairy Sci.* 78:1693–1702. doi:10.3168/jds.S0022-0302(95)76794-7
- Gross, J. J., and R. M. Bruckmaier. 2015. Repeatability of metabolic responses to a nutrient deficiency in early and mid-lactation and implications for robustness of dairy cows. *J. Dairy Sci.* 98:8634–8643. doi:10.3168/jds.2014-9246
- Gross, J. J., and R. M. Bruckmaier. 2019. Invited review: metabolic challenges and adaptation during different functional stages of the mammary gland in dairy cows: perspectives for sustainable milk production. *J. Dairy Sci.* 102:2828–2843. doi:10.3168/jds.2018-15713
- Gross, J., H. A. van Dorland, R. M. Bruckmaier, and F. J. Schwarz. 2011. Performance and metabolic profile of dairy cows during a lactational and deliberately induced negative energy balance by feed restriction with subsequent realimentation. *J. Dairy Sci.* 94:1820–1830. doi:10.3168/jds.2010-3707
- Gross, J. J., H. A. van Dorland, O. Wellnitz, and R. M. Bruckmaier. 2015a. Glucose transport and milk secretion during manipulated plasma insulin and glucose concentrations and during LPS-induced mastitis in dairy cows. *J. Anim. Physiol. Anim. Nutr. (Berl)* 99:747–756. doi:10.1111/jpn.12259
- Gross, J. J., E. C. Kessler, C. Albrecht, and R. M. Bruckmaier. 2015b. Response of the cholesterol metabolism to a negative energy balance in dairy cows depends on the lactational stage. *PLoS One* 10:e0121956. doi:10.1371/journal.pone.0121956
- Gross, J. J., L. Grossen-Rösti, F. Schmitz-Hsu, and R. M. Bruckmaier. 2016. Metabolic adaptation recorded during one lactation does not allow predicting longevity in dairy cows. *Schweiz. Arch. Tierheilkd.* 158:565–571. doi:10.17236/sat00078
- Gross, J. J., F. J. Schwarz, K. Eder, H. A. van Dorland, and R. M. Bruckmaier. 2013. Liver fat content and lipid metabolism in dairy cows during early lactation and during a mid-lactation feed restriction. *J. Dairy Sci.* 96:5008–5017. doi:10.3168/jds.2012-6245
- Grossen-Rösti, L., E. C. Kessler, A. Tröschler, R. M. Bruckmaier, and J. J. Gross. 2018. Hyperglycaemia in transition dairy cows: effects of lactational stage and conjugated linoleic acid supplementation on glucose metabolism and turnover. *J. Anim. Physiol. Anim. Nutr. (Berl)* 102:483–494. doi:10.1111/jpn.12771
- Grummer, R. R. 1993. Etiology of lipid-related metabolic disorders in periparturient dairy cows. *J. Dairy Sci.* 76:3882–3896. doi:10.3168/jds.S0022-0302(93)77729-2
- Grummer, R. R., D. G. Mashek, and A. Hayirli. 2004. Dry matter intake and energy balance in the transition period. *Vet. Clin. North Am. Food Anim. Pract.* 20:447–470. doi:10.1016/j.cvfa.2004.06.013
- Gutgesell, A., R. Ringseis, and K. Eder. 2009. Short communication: dietary conjugated linoleic acid down-regulates fatty acid transporters in the mammary glands of lactating rats. *J. Dairy Sci.* 92:1169–1173. doi:10.3168/jds.2008-1640
- Harmon, D. L., R. M. Yamka, and N. A. Elam. 2004. Factors affecting intestinal starch digestion in ruminants: a review. *Can. J. Anim. Sci.* 84:309–318. doi:10.4141/A03-077
- Hernández-Castellano, L. E., L. L. Hernandez, S. Weaver, and R. M. Bruckmaier. 2017a. Increased serum serotonin improves parturient calcium homeostasis in dairy cows. *J. Dairy Sci.* 100:1580–1587. doi:10.3168/jds.2016-11638
- Hernández-Castellano, L. E., L. L. Hernandez, H. Sauerwein, and R. M. Bruckmaier. 2017b. Endocrine and metabolic changes in transition dairy cows are affected by prepartum infusions of a serotonin precursor. *J. Dairy Sci.* 100:5050–5057. doi:10.3168/jds.2016-12441
- Hoard's Dairyman. 2020. The world record milk cow: she's just one of the herd. Accessed 14 December 2021. <https://hoards.com/article-27383-the-world-record-milk-cow-shes-just-one-of-the-herd.html>
- Hötger, K., H. M. Hammon, C. Weber, S. Görs, A. Tröschler, R. M. Bruckmaier, and C. C. Metges. 2013. Supplementation of conjugated linoleic acid in dairy cows reduces endogenous glucose production during early lactation. *J. Dairy Sci.* 96:2258–2270. doi:10.3168/jds.2012-6127
- Huhtanen, P., A. Vanhatalo, and T. Varvikko. 2002. Effects of abomasal infusions of histidine, glucose, and leucine on milk production and plasma metabolites of dairy cows fed grass silage diets. *J. Dairy Sci.* 85:204–216. doi:10.3168/jds.S0022-0302(02)74069-1
- Humer, E., J. R. Aschenbach, V. Neubauer, I. Kröger, R. Khiaosa-Ard, W. Baumgartner, and Q. Zebeli. 2018. Signals for identifying cows at risk of subacute ruminal acidosis in dairy veterinary practice. *J. Anim. Physiol. Anim. Nutr. (Berl)* 102:380–392. doi:10.1111/jpn.12850
- Huntington, G. B., D. L. Harmon, and C. J. Richards. 2006. Sites, rates, and limits of starch digestion and glucose metabolism in growing cattle. *J. Anim. Sci.* 84(Suppl):E14–E24. doi:10.2527/2006.8413\_supple14x
- Ingvarsen, K. L. 2006. Feeding- and management-related diseases in the transition cow: physiological adaptations around calving



- and strategies to reduce feeding-related diseases. *Anim. Feed Sci. Technol.* 126:175–213. doi:10.1016/j.anifeeds.2005.08.003
- Ingvarstsen, K. L., R. J. Dewhurst, and N. C. Friggens. 2003. On the relationship between lactational performance and health: is it yield or metabolic imbalance that cause production diseases in dairy cattle? a position paper. *Livest. Prod. Sci.* 83:277–308. doi:10.1016/S0301-6226(03)00110-6
- Ingvarstsen, K. L., and K. Moyes. 2013. Nutrition, immune function and health of dairy cattle. *Animal* 7(Suppl 1):112–122. doi:10.1017/S175173111200170X.
- Institut National de la Recherche Agronomique (INRA) 2018. *INRA feeding system for ruminants*. Wageningen (The Netherlands): Wageningen Academic Publishers.
- Jaakson, H., P. Karis, K. Ling, A. Ilves-Luht, J. Samarütel, M. Henno, I. Jõudu, A. Waldmann, E. Reimann, P. Pärn, et al. 2018. Adipose tissue insulin receptor and glucose transporter 4 expression, and blood glucose and insulin responses during glucose tolerance tests in transition holstein cows with different body condition. *J. Dairy Sci.* 101:752–766. doi:10.3168/jds.2017-12877.
- Kadzere, C. T., M. R. Murphy, N. Silanikove, and E. Maltz. 2002. Heat stress in lactating dairy cows: a review. *Livest. Prod. Sci.* 77:59–91. doi:10.1016/S0301-6226(01)00330-X.
- Karis, P., H. Jaakson, K. Ling, R. M. Bruckmaier, J. J. Gross, P. Pärn, T. Kaart, and M. Ots. 2020. Body condition and insulin resistance interactions with periparturient gene expression in adipose tissue and lipid metabolism in dairy cows. *J. Dairy Sci.* 103:3708–3718. doi:10.3168/jds.2019-17373.
- Kennedy, B. W. 1984. Selection limits: have they been reached with the dairy cow? *Can. J. Anim. Sci.* 64:207–215. doi:10.4141/cjas84-028
- Kessel, S., M. Stroehl, H. H. D. Meyer, S. Hiss, H. Sauerwein, F. J. Schwarz, and R. M. Bruckmaier. 2008. Individual variability in physiological adaptation to metabolic stress during early lactation in dairy cows kept under equal conditions. *J. Anim. Sci.* 86:2903–2912. doi:10.2527/jas.2008-1016
- Kessler, E. C., J. J. Gross, R. M. Bruckmaier, and C. Albrecht. 2014. Cholesterol metabolism, transport, and hepatic regulation in dairy cows during transition and early lactation. *J. Dairy Sci.* 97:5481–5490. doi:10.3168/jds.2014-7926
- Kessler, E. C., S. K. Wall, L. L. Hernandez, R. M. Bruckmaier, and J. J. Gross. 2018. Short communication: circulating serotonin is related to the metabolic status and lactational performance at the onset of lactation in dairy cows. *J. Dairy Sci.* 101:11455–11460. doi:10.3168/jds.2018-14626
- Kok, A., A. T. M. van Knegsel, C. E. van Middelaar, B. Engel, H. Hogeveen, B. Kemp, and I. J. M. de Boer. 2017. Effect of dry period length on milk yield over multiple lactations. *J. Dairy Sci.* 100:739–749. doi:10.3168/jds.2016-10963
- Kolver, E. S. 2003. Nutritional limitations to increased production on pasture-based systems. *Proc. Nutr. Soc.* 62:291–300. doi:10.1079/pns2002200
- Komatsu, T., F. Itoh, S. Kushibiki, and K. Hodate. 2005. Changes in gene expression of glucose transporters in lactating and nonlactating cows. *J. Anim. Sci.* 83:557–564. doi:10.2527/2005.833557x
- Korst, M., C. Koch, J. Kesser, U. Müller, F. J. Romberg, J. Rehage, K. Eder, and H. Sauerwein. 2017. Different milk feeding intensities during the first 4 weeks of rearing in dairy calves: part 1: effects on performance and production from birth over the first lactation. *J. Dairy Sci.* 100:3096–3108. doi:10.3168/jds.2016-11594
- Lacasse, P., and C. G. Prosser. 2003. Mammary blood flow does not limit milk yield in lactating goats. *J. Dairy Sci.* 86:2094–2097. doi:10.3168/jds.S0022-0302(03)73798-9
- Lacasse, P., and D. Petitclerc. 2021. Effect of photoperiod before and during first gestation on milk production and prolactin concentration in dairy heifers. *J. Dairy Sci.* 104:4991–4998. doi:10.3168/jds.2020-19514
- Lacasse, P., C. M. Vinet, and D. Petitclerc. 2014. Effect of prepartum photoperiod and melatonin feeding on milk production and prolactin concentration in dairy heifers and cows. *J. Dairy Sci.* 97:3589–3598. doi:10.3168/jds.2013-7615
- Laporta, J., F. C. Ferreira, V. Ouellet, B. Dado-Senn, A. K. Almeida, A. D. Vries, and G. E. Dahl. 2020. Late-gestation heat stress impairs daughter and granddaughter lifetime performance. *J. Dairy Sci.* 103:7555–7568. doi:10.3168/jds.2020-18154
- Larsen, M., and N. B. Kristensen. 2013. Precursors for liver gluconeogenesis in periparturient dairy cows. *Animal* 7:1640–1650. doi:10.1017/S1751731113001171
- Levit, H., S. Pinto, T. Amon, E. Gershon, A. Kleinjan-Elazary, V. Bloch, Y. A. Ben Meir, Y. Portnik, S. Jacoby, A. Arnin, et al. 2021. Dynamic cooling strategy based on individual animal response mitigated heat stress in dairy cows. *Animal* 15:100093. doi:10.1016/j.animal.2020.100093
- Linzell, J. L. 1974. Mammary blood flow and methods of identifying and measuring precursors of milk. In: Larson, B. L., and V. R. Smith, ed. *Lactation*. Vol. 1. New York (NY): Academic Press; p. 143.
- Locher, L., S. Häussler, L. Laubenthal, S. P. Singh, J. Winkler, A. Kinoshita, A. Kenéz, J. Rehage, K. Huber, H. Sauerwein, et al. 2015. Effect of increasing body condition on key regulators of fat metabolism in subcutaneous adipose tissue depot and circulation of nonlactating dairy cows. *J. Dairy Sci.* 98:1057–1068. doi:10.3168/jds.2014-8710
- Locher, L. F., N. Meyer, E. M. Weber, J. Rehage, U. Meyer, S. Dänicke, and K. Huber. 2011. Hormone-sensitive lipase protein expression and extent of phosphorylation in subcutaneous and retroperitoneal adipose tissues in the periparturient dairy cow. *J. Dairy Sci.* 94:4514–4523. doi:10.3168/jds.2011-4145
- Longo, N., M. Frigeni, and M. Pasquali. 2016. Carnitine transport and fatty acid oxidation. *Biochim. Biophys. Acta* 1863:2422–2435. doi:10.1016/j.bbamcr.2016.01.023
- Lucy, M. C., G. A. Verkerk, B. E. Whyte, K. A. Macdonald, L. Burton, R. T. Cursons, J. R. Roche, and C. W. Holmes. 2009. Somatotrophic axis components and nutrient partitioning in genetically diverse dairy cows managed under different feed allowances in a pasture system. *J. Dairy Sci.* 92:526–539. doi:10.3168/jds.2008-1421
- Ma, L., K. L. Cook, D. E. Bauman, and K. J. Harvatine. 2015. Short communication: milk fat depression induced by conjugated linoleic acid and a high-oil and low-fiber diet occurs equally across the day in Holstein cows. *J. Dairy Sci.* 98:1851–1855. doi:10.3168/jds.2014-8614
- Macmillan, K., M. Gobikrushanth, A. Behrouzi, B. Hoff, and M. G. Colazo. 2021. Prevalence of early postpartum health disorders in holstein cows and associations with production, reproduction, and survival outcomes on Alberta dairy farms. *Can. Vet. J.* 62:273–280.
- Malacco, V. M. R., M. Erickson, F. F. Cardoso, B. P. Biese, J. G. Laguna, and S. S. Donkin. 2020. Short communication: effect of glucose infusion dose and stage of lactation on glucose tolerance test kinetics in lactating dairy cows. *J. Dairy Sci.* 103:7547–7554. doi:10.3168/jds.2019-17139
- Mann, S., D. V. Nydam, A. Abuelo, F. A. Leal Yepes, T. R. Overton, and J. J. Wakshlag. 2016. Insulin signalling, inflammation, and lipolysis in subcutaneous adipose tissue of transition dairy cows either overfed energy during the prepartum period or fed a controlled-energy diet. *J. Dairy Sci.* 99:6737–6752. doi:10.3168/jds.2016-10969.
- Matsumoto, M., H. Nishinakagawa, M. Kurohmaru, Y. Hayashi, and J. Otsuka. 1992. Pregnancy and lactation affect the microvasculature of the mammary gland in mice. *J. Vet. Med. Sci.* 54:937–943. doi:10.1292/jvms.54.937
- Mattmiller, S. A., C. M. Corl, J. C. Gandy, J. J. Looor, and L. M. Sordillo. 2011. Glucose transporter and hypoxia-associated gene expression in the mammary gland of transition dairy cattle. *J. Dairy Sci.* 94:2912–2922. doi:10.3168/jds.2010-3936
- McArt, J. A. A., R. C. Neves. 2020. Association of transient, persistent, or delayed subclinical hypocalcemia with early lactation disease, removal, and milk yield in Holstein cows. *J. Dairy Sci.* 103:690–701. doi:10.3168/jds.2019-17191
- McArt, J. A., D. V. Nydam, and G. R. Oetzel. 2012. Epidemiology of subclinical ketosis in early lactation dairy cattle. *J. Dairy Sci.* 95:5056–5066. doi:10.3168/jds.2012-5443

- McFadden, J. W. 2020. Review: lipid biology in the periparturient dairy cow: contemporary perspectives. *Animal* 14(Suppl 1):s165–s175. doi:10.1017/S1751731119003185
- McFadden, J. W., and J. E. Rico. 2019. Invited review: sphingolipid biology in the dairy cow: the emerging role of ceramide. *J. Dairy Sci.* 102:7619–7639. doi:10.3168/jds.2018-16095
- McNamara, J. P., K. Huber, and A. Kenéz. 2016. A dynamic, mechanistic model of metabolism in adipose tissue of lactating dairy cattle. *J. Dairy Sci.* 99:5649–5661. doi:10.3168/jds.2015-9585
- Miller, P. S., B. L. Reis, C. C. Calvert, E. J. DePeters, and R. L. Baldwin. 1991. Patterns of nutrient uptake by the mammary glands of lactating dairy cows. *J. Dairy Sci.* 74:3791–3799. doi:10.3168/jds.S0022-0302(91)78571-8
- Neubauer, V., R. M. Petri, E. Humer, I. Kröger, N. Reisinger, W. Baumgartner, M. Wagner, and Q. Zebeli. 2020. Starch-rich diet induced rumen acidosis and hindgut dysbiosis in dairy cows of different lactations. *Animals (Basel)*. 10:1727. doi:10.3390/ani10101727
- NorFor. 2011. *The Nordic feed evaluation system*. In: H. Volden, editor. EAAP Scientific Series, Volume 130. Wageningen (The Netherlands): Wageningen Academic Publishers.
- NRC. 2001. *Nutrient requirements of dairy cattle*. 7th ed. Washington (DC): National Academies Press.
- Özçelik, R., R. M. Bruckmaier, and L. E. Hernández-Castellano. 2017. Parturition daylight exposure increases serum calcium concentrations in dairy cows at the onset of lactation. *J. Anim. Sci.* 95:4440–4447. doi:10.2527/jas2017.1834
- Owens, F. N., R. A. Zinn, and Y. K. Kim. 1986. Limits to starch digestion in the ruminant small intestine. *J. Anim. Sci.* 63:1634–1648. doi:10.2527/jas1986.6351634x
- Peel, C. J., and D. E. Bauman. 1987. Somatotropin and lactation. *J. Dairy Sci.* 70:474–486. doi:10.3168/jds.S0022-0302(87)80030-9
- Ponchon, B., P. Lacasse, S. Ollier, and X. Zhao. 2017. Effects of photoperiod modulation and melatonin feeding around drying-off on bovine mammary gland involution. *J. Dairy Sci.* 100:8496–8506. doi:10.3168/jds.2016-12272
- Prosser, C. G., S. R. Davis, V. C. Farr, and P. Lacasse. 1996. Regulation of blood flow in the mammary microvasculature. *J. Dairy Sci.* 79:1184–1197. doi:10.3168/jds.S0022-0302(96)76472-X
- Pulina, G., A. Tondo, P. P. Danieli, R. Primi, G. M. Crovetto, A. Fantini, N. P. P. Macciotta, and A. S. Atzori. 2020. How to manage cows yielding 20,000 kg of milk: technical challenges and environmental implications. *Ital. J. Anim. Sci.* 19:865–879. doi:10.1080/1828051X.2020.1805370
- Purup, S., M. Vestergaard, M. S. Weber, K. Plaut, R. M. Akers, and K. Sejrsen. 2000. Local regulation of pubertal mammary growth in heifers. *J. Anim. Sci.* 78(Suppl. 3):36–47. doi:10.2527/2000.78suppl\_336x
- Rabiee, A. R., K. Breinhild, W. Scott, H. M. Golder, E. Block, and I. J. Lean. 2012. Effect of fat additions to diets of dairy cattle on milk production and components: a meta-analysis and meta-regression. *J. Dairy Sci.* 95:3225–3247. doi:10.3168/jds.2011-4895
- Raboisson, D., C. Caubet, C. Tasca, L. D. Marchi, J. M. Ferraton, S. Gannac, A. Millet, F. Enjalbert, F. Schelcher, and G. Foucras. 2014a. Effect of acute and chronic excesses of dietary nitrogen on blood neutrophil functions in cattle. *J. Dairy Sci.* 97:7575–7585. doi:10.3168/jds.2014-8303
- Raboisson, D., M. Mounié, and E. Maigné. 2014b. Diseases, reproductive performance, and changes in milk production associated with subclinical ketosis in dairy cows: a meta-analysis and review. *J. Dairy Sci.* 97:7547–7563. doi:10.3168/jds.2014-8237
- Raschka, C., L. Ruda, P. Wenning, C.-I. von Stemm, C. Pfarrer, K. Huber, U. Meyer, S. Dänicke, and J. Rehage. 2016. In vivo determination of subcutaneous and abdominal adipose tissue depots in German Holstein dairy cattle. *J. Anim. Sci.* 94:2821–2834. doi:10.2527/jas.2015-0103
- Reinhardt, T. A., J. D. Lippolis, B. J. McCluskey, J. P. Goff, and R. L. Horst. 2011. Prevalence of subclinical hypocalcemia in dairy herds. *Vet. J.* 188:122–124. doi:10.1016/j.tvjl.2010.03.025
- Rilanto, T., K. Reimus, T. Orro, U. Emanuelson, A. Viltrop, and K. Mõtus. 2020. Culling reasons and risk factors in Estonian dairy cows. *BMC Vet. Res.* 16:173. doi:10.1186/s12917-020-02384-6
- Ringseis, R., and K. Eder. 2011. Regulation of genes involved in lipid metabolism by dietary oxidized fat. *Mol. Nutr. Food Res.* 55:109–121. doi:10.1002/mnfr.201000424
- Ringseis, R., J. Keller, and K. Eder. 2018. Regulation of carnitine status in ruminants and efficacy of carnitine supplementation on performance and health aspects of ruminant livestock: a review. *Arch. Anim. Nutr.* 72:1–30. doi:10.1080/1745039X.2017.1421340
- Roth, Z. 2020. Reproductive physiology and endocrinology responses of cows exposed to environmental heat stress—experiences from the past and lessons for the present. *Theriogenology* 155:150–156. doi:10.1016/j.theriogenology.2020.05.040
- Rukkamsuk, T., T. A. Kruij, and T. Wensing. 1999. Relationship between overfeeding and overconditioning in the dry period and the problems of high producing dairy cows during the postparturient period. *Vet. Q.* 21:71–77. doi:10.1080/01652176.1999.9694997
- Sadri, H., F. Giallongo, A. N. Hristov, J. Werner, C. H. Lang, C. Parys, B. Saremi, and H. Sauerwein. 2016. Effects of slow-release urea and rumen-protected methionine and histidine on mammalian target of rapamycin (mTOR) signaling and ubiquitin proteasome-related gene expression in skeletal muscle of dairy cows. *J. Dairy Sci.* 99:6702–6713. doi:10.3168/jds.2015-10673
- Sammad, A., Y. J. Wang, S. Umer, H. Lirong, I. Khan, A. Khan, B. Ahmad, and Y. Wang. 2020. Nutritional physiology and biochemistry of dairy cattle under the influence of heat stress: consequences and opportunities. *Animals (Basel)* 10:793. doi:10.3390/ani10050793
- Schäff, C., S. Börner, S. Hacke, U. Kautzsch, H. Sauerwein, S. K. Spachmann, M. Schweigel-Röntgen, H. M. Hammon, and B. Kuhla. 2013. Increased muscle fatty acid oxidation in dairy cows with intensive body fat mobilization during early lactation. *J. Dairy Sci.* 96:6449–6460. doi:10.3168/jds.2013-6812
- Schlegel, G., R. Ringseis, J. Keller, F. J. Schwarz, and K. Eder. 2012. Changes in the expression of hepatic genes involved in cholesterol homeostasis in dairy cows in the transition period and at different stages of lactation. *J. Dairy Sci.* 95:3826–3836. doi:10.3168/jds.2011-5221
- Schuh, K., H. Sadri, S. Häussler, L. A. Webb, C. Urh, M. Wagner, C. Koch, J. Frahm, S. Dänicke, G. Dusel, et al. 2019. Comparison of performance and metabolism from late pregnancy to early lactation in dairy cows with elevated v. normal body condition at dry-off. *Animal* 13:1478–1488. doi:10.1017/S1751731118003385
- Schwab, C. G., and G. A. Broderick. 2017. A 100-year review: protein and amino acid nutrition in dairy cows. *J. Dairy Sci.* 100:10094–10112. doi:10.3168/jds.2017-13320
- Seck, M., J. A. V. Linton, M. S. Allen, D. S. Castagnino, P. Y. Chouinard, and C. L. Girard. 2017. Apparent ruminal synthesis of B vitamins in lactating dairy cows fed diets with different forage-to-concentrate ratios. *J. Dairy Sci.* 100:1914–1922. doi:10.3168/jds.2016-12111
- Sejrsen, K., J. T. Huber, H. A. Tucker, and R. M. Akers. 1982. Influence of nutrition of mammary development in pre- and postpubertal heifers. *J. Dairy Sci.* 65:793–800. doi:10.3168/jds.s0022-0302(82)82268-6
- Sejrsen, K., and S. Purup. 1997. Influence of prepubertal feeding level on milk yield potential of dairy heifers: a review. *J. Anim. Sci.* 75:828–835. doi:10.2527/1997.753828x
- Singh, K., R. A. Erdman, K. M. Swanson, A. J. Molenaar, N. J. Maqbool, T. T. Wheeler, J. A. Arias, E. C. Quinn-Walsh, and K. Stelwagen. 2010. Epigenetic regulation of milk production in dairy cows. *J. Mamm. Gland Biol. Neoplasia* 15:101–112. doi:10.1007/s10911-010-9164-2
- Soberon, F., E. Raffrenato, R. W. Everett, and M. E. Van Amburgh. 2012. Prewaning milk replacer intake and effects on long-term productivity of dairy calves. *J. Dairy Sci.* 95:783–793. doi:10.3168/jds.2011-4391
- Soberon, F., and M. E. Van Amburgh. 2013. Lactation biology symposium: the effect of nutrient intake from milk or milk replacer of preweaned dairy calves on lactation milk yield as adults: a meta-

- analysis of current data. *J. Anim. Sci.* 91:706–712. doi:[10.2527/jas.2012-5834](https://doi.org/10.2527/jas.2012-5834)
- Sordillo, L. M., G. A. Contreras, and S. L. Aitken. 2009. Metabolic factors affecting the inflammatory response of periparturient dairy cows. *Anim. Health Res. Rev.* 10:53–63. doi:[10.1017/S1466252309990016](https://doi.org/10.1017/S1466252309990016)
- Spek, J. W., A. Bannink, G. Gort, W. H. Hendriks, and J. Dijkstra. 2013. Interaction between dietary content of protein and sodium chloride on milk urea concentration, urinary urea excretion, renal recycling of urea, and urea transfer to the gastrointestinal tract in dairy cows. *J. Dairy Sci.* 96:5734–5745. doi:[10.3168/jds.2013-6842](https://doi.org/10.3168/jds.2013-6842)
- Stelwagen, K., C. V. Phyn, S. R. Davis, J. Guinard-Flament, D. Pomiès, J. R. Roche, and J. K. Kay. 2013. Invited review: reduced milking frequency: milk production and management implications. *J. Dairy Sci.* 96:3401–3413. doi:[10.3168/jds.2012-6074](https://doi.org/10.3168/jds.2012-6074)
- Suriyasathaporn, W., A. J. Daemen, E. N. Noordhuizen-Stassen, S. J. Dieleman, M. Nielen, and Y. H. Schukken. 1999. Beta-hydroxybutyrate levels in peripheral blood and ketone bodies supplemented in culture media affect the in vitro chemotaxis of bovine leukocytes. *Vet. Immunol. Immunopathol.* 68:177–186. doi:[10.1016/s0165-2427\(99\)00017-3](https://doi.org/10.1016/s0165-2427(99)00017-3)
- Thompson-Crispi, K., H. Atalla, F. Miglior, and B. A. Mallard. 2014. Bovine mastitis: frontiers in immunogenetics. *Front. Immunol.* 5:493. doi:[10.3389/fimmu.2014.00493](https://doi.org/10.3389/fimmu.2014.00493)
- Tse, C., H. W. Barkema, T. J. DeVries, J. Rushen, and E. A. Pajor. 2018. Impact of automatic milking systems on dairy cattle producers' reports of milking labour management, milk production and milk quality. *Animal* 12:2649–2656. doi:[10.1017/S1751731118000654](https://doi.org/10.1017/S1751731118000654)
- Trevisi, E., M. Amadori, S. Cogrossi, E. Razzuoli, and G. Bertoni. 2012. Metabolic stress and inflammatory response in high-yielding, periparturient dairy cows. *Res. Vet. Sci.* 93:695–704. doi:[10.1016/j.rvsc.2011.11.008](https://doi.org/10.1016/j.rvsc.2011.11.008)
- Van der Kolk, J. H., J. J. Gross, V. Gerber, and R. M. Bruckmaier. 2017. Disturbed bovine mitochondrial lipid metabolism: a review. *Vet. Q.* 37:262–273. doi:[10.1080/01652176.2017.1354561](https://doi.org/10.1080/01652176.2017.1354561)
- Van Knegsel, A. T., S. G. van der Drift, J. Cermáková, and B. Kemp. 2013. Effects of shortening the dry period of dairy cows on milk production, energy balance, health, and fertility: a systematic review. *Vet. J.* 198:707–713. doi:[10.1016/j.tvjl.2013.10.005](https://doi.org/10.1016/j.tvjl.2013.10.005)
- Varga, G. A., H. M. Dann, and V. A. Ishler. 1998. The use of fiber concentrations for ration formulation. *J. Dairy Sci.* 81:3063–3074. doi:[10.3168/jds.S0022-0302\(98\)75871-0](https://doi.org/10.3168/jds.S0022-0302(98)75871-0)
- Venjakob, P. L., S. Borchardt, and W. Heuwieser. 2017. Hypocalcemia-cow-level prevalence and preventive strategies in German dairy herds. *J. Dairy Sci.* 100:9258–9266. doi:[10.3168/jds.2016-12494](https://doi.org/10.3168/jds.2016-12494)
- Vergara, C. F., D. Döpfer, N. B. Cook, K. V. Nordlund, J. A. McArt, D. V. Nydam, and G. R. Oetzel. 2014. Risk factors for postpartum problems in dairy cows: explanatory and predictive modeling. *J. Dairy Sci.* 97:4127–4140. doi:[10.3168/jds.2012-6440](https://doi.org/10.3168/jds.2012-6440)
- Wall, E. H., and T. B. McFadden. 2012. Triennial lactation symposium: a local affair: how the mammary gland adapts to changes in milking frequency. *J. Anim. Sci.* 90:1695–1707. doi:[10.2527/jas.2011-4790](https://doi.org/10.2527/jas.2011-4790)
- Weld, K. A., and L. E. Armentano. 2017. The effects of adding fat to diets of lactating dairy cows on total-tract neutral detergent fiber digestibility: a meta-analysis. *J. Dairy Sci.* 100:1766–1779. doi:[10.3168/jds.2016-11500](https://doi.org/10.3168/jds.2016-11500)
- White, H. M. 2015. The role of TCA cycle anaplerosis in ketosis and fatty liver in periparturient dairy cows. *Animals (Basel)*. 5:793–802. doi:[10.3390/ani5030384](https://doi.org/10.3390/ani5030384)
- Xiao, C. T., and J. P. Cant. 2005. Relationship between glucose transport and metabolism in isolated bovine mammary epithelial cells. *J. Dairy Sci.* 88:2794–2805. doi:[10.3168/jds.S0022-0302\(05\)72959-3](https://doi.org/10.3168/jds.S0022-0302(05)72959-3)
- Yang, Y., H. Sadri, C. Prehn, J. Adamski, J. Rehage, S. Dänicke, D. von Soosten, C. C. Metges, M. H. Ghaffari, and H. Sauerwein. 2020. Proteasome activity and expression of mammalian target of rapamycin signaling factors in skeletal muscle of dairy cows supplemented with conjugated linoleic acids during early lactation. *J. Dairy Sci.* 103:2829–2846. doi:[10.3168/jds.2019-17244](https://doi.org/10.3168/jds.2019-17244)
- Yasugi, T., T. Kaido, and Y. Uehara. 1989. Changes in density and architecture of microvessels of the rat mammary gland during pregnancy and lactation. *Arch. Histol. Cytol.* 52:115–122. doi:[10.1679/aohc.52.115](https://doi.org/10.1679/aohc.52.115)
- Zarrin, M., L. D. Matteis, M. C. Vernay, O. Wellnitz, H. A. van Dorland, and R. M. Bruckmaier. 2013. Long-term elevation of  $\beta$ -hydroxybutyrate in dairy cows through infusion: effects on feed intake, milk production, and metabolism. *J. Dairy Sci.* 96:2960–2972. doi:[10.3168/jds.2012-6224](https://doi.org/10.3168/jds.2012-6224)
- Zarrin, M., L. Grosse-Rösti, R. M. Bruckmaier, and J. J. Gross. 2017. Elevation of blood  $\beta$ -hydroxybutyrate concentration affects glucose metabolism in dairy cows before and after parturition. *J. Dairy Sci.* 100:2323–2333. doi:[10.3168/jds.2016-11714](https://doi.org/10.3168/jds.2016-11714)
- Zbinden, R. S., M. Falk, A. Münger, F. Dohme-Meier, H. A. van Dorland, R. M. Bruckmaier, and J. J. Gross. 2017. Metabolic load in dairy cows kept in herbage based feeding systems and suitability of potential markers for compromised well-being. *J. Anim. Physiol. Anim. Nutr. (Berl)* 101:767–778. doi:[10.1111/jpn.12498](https://doi.org/10.1111/jpn.12498)
- Zebeli, Q., J. R. Aschenbach, M. Tafaj, J. Boguhn, B. N. Ametaj, and W. Drochner. 2012. Invited review: role of physically effective fiber and estimation of dietary fiber adequacy in high-producing dairy cattle. *J. Dairy Sci.* 95:1041–1056. doi:[10.3168/jds.2011-442](https://doi.org/10.3168/jds.2011-442)
- Zhao, F.-Q., and A. F. Keating. 2007a. Functional properties and genomics of glucose transporters. *Curr. Genomics* 8:113–128. doi:[10.2174/138920207780368187](https://doi.org/10.2174/138920207780368187)
- Zhao, F.-Q., and A. F. Keating. 2007b. Expression and regulation of glucose transporters in the bovine mammary gland. *J. Dairy Sci.* 90(E. Suppl.):E76–E86. doi:[10.3168/jds.2006-470](https://doi.org/10.3168/jds.2006-470)