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REVIEW



STAT3 pathway in cancers: Past, present, and future

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National Natural Science Foundation of China, Grant/Award Numbers: 81702703, 81872203; China Postdoctoral Science Foundation, Grant/Award Numbers: 2018M630883, 2019T120688; Hubei Province Natural Science Foundation of China, Grant/Award Number: Signal transducer and activator of transcription 3 (STAT3), a member of the STAT family, discovered in the cytoplasm of almost all types of mammalian cells, plays a significant role in biological functions. The duration of STAT3 activation in normal tissues is a transient event and is strictly regulated. However, in cancer tissues, STAT3 is activated in an aberrant manner and is induced by certain cytokines. The continuous activation of STAT3 regulates the expression of downstream proteins associated with the formation, progression, and metastasis of cancers. Thus, elucidating the mechanisms of STAT3 regulation and designing inhibitors targeting the STAT3 pathway are considered promising strategies for cancer treatment. This review aims to introduce the history, research advances, and prospects concerning the STAT3 pathway in cancer. We review the mechanisms of STAT3 pathway regulation and the consequent cancer hallmarks associated with tumor biology that are induced by the STAT3 pathway. Moreover, we summarize the emerging development of inhibitors that target the STAT3 pathway and novel drug delivery systems for delivering these inhibitors. The barriers against targeting the STAT3 pathway, the focus of future research on promising targets in the STAT3 pathway, and our perspective on the overall utility of STAT3 pathway inhibitors in cancer treatment are also discussed.

KEYWORDS

cancer hallmarks, cancer treatment, STAT3 pathway inhibitors, STAT3

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1 | INTRODUCTION

The incidence and mortality rates of cancer are rapidly increasing worldwide. An estimated 19.3 million new cancer cases and 10.0 million cancer-related deaths were recorded around the world in 2020. By 2040, the global cancer burden is expected to undergo a 47% rise from 2020.¹ According to World Health Statistics 2021 posted by the World Health Organization, there were 1.9 million new cases diagnosed in 2021. Even in high-income countries, cancer has been regarded as one of the leading causes of "premature death" defined as the death between the ages of 30 and 70.² Currently, most patients with cancer are treated with surgery combined with radiotherapy, chemotherapy, and/or immunotherapy, depending on the types of cancer.^{3–5} Although these conventional therapies may achieve the significant tumor elimination effect initially, various barriers such as surgical trauma, high cost, drug resistance, and cytotoxicity in normal tissues are associated with the risk of treatment discontinuation and cancer recurrence and metastasis.⁶⁻⁸ Therefore, searching for new therapeutic targets and treatment methods are crucial for improving the survival rate and quality of life of patients with cancer.

Signal transducer and activator of transcription (STAT) proteins are latent cytoplasmic transcription factors comprising seven members, including STAT1, STAT2, STAT3, STAT4, STAT5a, STAT5b, and STAT6.9 Among them, STAT3 is involved in various basic cell functions including cell growth, survival, differentiation, regeneration, immune response, and cellular respiration. STAT3 can be strictly regulated by upstream signaling molecules, such as Janus kinase (JAK) and epidermal growth factor receptor (EGFR). It then localizes to the nucleus of cells and binds to target DNA to regulate the expression of downstream proteins.^{10–14} However, in addition to its functions in the normal cells, STAT3 activation in the tumor microenvironment (TME) is regarded as an oncogenic event. High phospho-STAT3 expression is associated with poor prognosis in patients with various types of cancers such as non-small cell lung cancer, gastric cancer, and colorectal cancer.^{15–17} Constitutive activation of STAT3 plays an important role in tumor formation, development, metastasis, and recurrence. These are strongly associated with cancer hallmarks and lead to poor patient outcomes. Therefore, the STAT3 pathway is a promising target for cancer therapy.^{18–22}

In this review, we summarize the latest advances in STAT3 pathway in cancer. The mechanisms of STAT3 pathway regulation and the cancer hallmarks induced by the STAT3 pathway are also reviewed. We introduce the STAT3 pathway inhibitors and the novel drug delivery systems in preclinical studies and clinical trials for cancer treatment. Furthermore, the difficulties and prospects of targeting the STAT3 pathway in cancer are also discussed.

2 | HISTORY OF STAT3 PATHWAY INVESTIGATION

In 1990, interferon-stimulated gene factor 3 (ISGF3), the STAT protein complex activated by interferon α (IFN α), was purified and identified as a DNA-binding protein by Darnell et al. (Figure 1).²³ In 1992, Fu reported that the 113, 91, and 84 kDa (ISGF3 α) proteins of the IFN α induced primary transcription factor ISGF3 had conserved Src homology domains (SH2 and SH3). IFN α treatment stimulates the activating phosphorylation of ISGF3 α by protein tyrosine kinase (PTK), which facilitates direct signal transduction from the cell membrane to the nucleus.²⁴ The discovery of JAK/STAT pathway was considered as one of the top 10 breakthroughs in 1993 by Science. Subsequently, an acute-phase response factor (APRF) was observed to bind to interleukin-6 (IL-6) responsive elements and was activated in the cytoplasm to achieve the IL-6 signal transduction.²⁵ After the purification and cloning of APRF, Akira et al. demonstrated that APRF is highly homologous to the ISGF3 family and is involved in the glycoprotein 130 (gp130)-mediated signaling pathway. And this APRF was regarded as the novel transcription factor named STAT3.²⁶ The structure of STAT3 contains several conserved domains, including an N-terminal domain, coiled-coil domain, DNA-binding domain (DBD), linker domain, SH2 domain, and C-terminal domain, which are critical for its functions (Figure 2A,B).^{27,28} The Nterminal domain mediates tetramerization of two phosphorylated STAT3 dimers, dimerization of unphosphorylated STAT3, and binds other proteins to form functional complexes. These three main functions are associated with DNA binding, nuclear accumulation, and gene regulation



FIGURE 1 Brief history of signal transducer and activator of transcription 3 (STAT3) studies



FIGURE 2 (A, B) Schematic and functional architecture of STAT3 protein. STAT3 has the construction, including N-terminus domain, coiled-coil domain, DNA-binding domain, linker, SRC homology 2 and C-terminus domain. Each of them plays a role such as DNA binding, dimerization, and transactivation. Reprinted with permission from Mertens et al.²⁸ Copyright 2015 by National Academy of Sciences

of STAT3.²⁹ The coiled-coil domain is essential for early events in STAT3 signal transduction, including STAT3 recruitment for the receptor binding, tyrosine phosphorylation, dimerization, and DNA binding.³⁰ The C-terminal domain, also known as the trans-activation domain (TDA), contains a specific tyrosine residue (Tyr705) that can be phosphorylated through the interaction of ligands with their receptors. STAT3 has an alpha-helical linker domain spanning amino acid residues 500 to 575 that is followed by a classical SH2 domain. The SH2 domain is the highly conserved region that interacts with specific phosphotyrosine motifs of cytoplasmic signaling molecules and plays an important role in the dimerization of two STAT3.³¹

In 1999, Darnell et al. identified STAT3 as an oncogene that could be constitutively activated and cause growth dysregulation in human tumor samples.³² Thus, STAT3 inhibitors have been developed for a long time. In 2001, Turkson et al. designed a STAT3 inhibitor, the SH2 domain-binding phosphopeptide PY*LKTK, based on the key structural features of the STAT3 protein. PY*LKTK can bind to STAT3 in a manner similar to the Tyr(P)-SH2 interaction, block STAT3 dimerization, and selectively inhibits STAT3 activation.³³ Subsequently, the first non-peptidic small-molecule STAT3 inhibitor, Stattic, was developed. Stattic was reported to inhibit both the activation and dimerization of STAT3 by selectively inhibiting the binding of tyrosine-phosphorylated peptides to the STAT3 SH2 domain.34 Moreover, the orally available direct STAT3 inhibitor napabucasin (BBI608) was approved by the United States Food and Drug Administration (FDA) for the treatment of two cancers, including gastric/gastroesophageal junction (GEJ) cancer and pancreatic cancer in 2016. In 2017 and 2020, the FDA approved tocilizumab and WP1066, respectively, to reduce cytokine release syndrome (CRS) induced by chimeric antigen receptor T (CAR-T) cell therapy and to treat pediatric brain cancer. These results indicate that the STAT3 pathway is indispensable for treatment strategies against cancer in the past, present, and future.

3 | STAT3 AS AN ONCOGENE

Oncogenes are regarded as the viral or mutated cellular genes that play a decisive role in tumor formation. Activated oncogenes, such as K-*ras* and *src*, inducing malignant transformation in tumor cells were found.^{35,36} Generally, *STAT3* is considered an oncogene because of the following reasons.

Aberrant phosphorylation of STAT3 accumulates in nearly 70% of cancers, such as non-small cell lung cancer and breast tumor. In human breast tissues, the level of STAT3-binding activity was significantly higher in carcinomas than in normal and benign lesions.³⁷ And STAT3 expression is 10.6-fold higher in head and neck squamous cell carcinoma (HNSCC) tissues, compared to normal mucosa tissues derived from non-HNSCC patients, leading to that activated *STAT3* is regarded as an oncogene.^{38–42}

In addition to STAT3 overexpression in tumor tissues, cells bearing the activated STAT3 were transformed to form tumors in nude mice. Bromberg et al. replaced the C-terminal loop with two cysteine residues in the SH2 domain of STAT3 to develop a STAT3 molecule (STAT3-C) that can dimerize spontaneously without tyrosine phosphorylation. After transfection of immortalized fibroblasts with STAT3-C, colonies were observed in soft agar, and tumors were present in nude mice 2–4 weeks after the injection of STAT3-C clones.³²

Aberrantly, phosphorylated STAT3 is also involved in tumor formation, development, and metastasis, which would influence the clinical outcome of patients. Lin et al. obtained tumor samples from 90 patients with glioblastoma (GBM) to examine the association between p-STAT3 expression levels and patient outcomes. Patients with a large percentage of p-STAT3 positive tumor cells had shorter progression-free survival years and overall survival years. In this study, p-STAT3 expression was considered an independent prognostic indicator in the outcome of patients with GBM.⁴³ Fei et al. reported that high plasma STAT3 levels with the high programmed death ligand 1 (PD-L1) expression induced by STAT3 led to the worst overall survival in patients with diffuse large B-cell lymphoma.⁴⁴

Moreover, STAT3 pathway inhibitors are reported to be effective in cancer treatment. In 2000, Song et al. proposed that targeting STAT3 can inhibit tumor growth through downregulating Bcl-xL expression to increase apoptosis of HNSCC cancer cells, which supports the view of STAT3 as an oncogene.⁴⁵ Moreover, there is a significant correlation between downstream proteins regulated by STAT3 and lymph node metastasis, cancer stages, recurrence, and death in patients with lingual squamous cell carcinoma. Knocking down the upstream molecule of STAT3 or inhibiting STAT3 expression can decrease the growth and metastasis of HNSCC.^{18,46,47} The enhancer of zeste homolog 2/STAT3 signaling is demonstrated to play a promoting role in chemoresistance and chemo-related adverse events in prostate cancer treatment. Targeting this signaling can significantly block the neuroendocrine differentiation in prostate cancer and inhibit the growth of chemoresistant cancer in vivo.^{48,49} Collectively, the above studies provide evidence that the high expression level and constitutive activation of STAT3 in the TME are strongly associated with cancer formation and poor prognosis of patients.

However, some researchers hold the converse view that STAT3 is an oncosuppressor. Kenichi Shinagawa et al. showed that the clinicopathological factors are significantly associated with IL-6 expression rather than STAT3 expression levels. The expression of IL-6 and STAT3 in 116 patients with oral squamous cell carcinoma (OSCC) were independent, and only high IL-6 expression significantly promoted vascular invasion and decreased the 5year disease-free survival rate.⁵⁰ Moreover, STAT3-positive cells are associated with favorable outcomes in some cancers. High nuclear STAT3 levels were associated with a 72.4% overall survival rate in patients with HNSCC at 5 years, compared to 38.3% in the low nuclear STAT3level group. High STAT3 expression may contribute to the early stages of tumor formation and development by inhibiting apoptosis of tumor cells.⁵¹ Tissue microarraybased study of both node-negative and node-positive breast tumors showed that overexpression of phospho-STAT3 (Tyr705) improved the short-term survival and long-term survival of patients with breast tumor, which indicated a better prognosis of patients with STAT3 positive breast cancer.^{52,53} Pencik et al. reported that the ARF-Mdm2-p53 tumor suppressor axis is regulated by STAT3, and loss of STAT3 signal transduction increases the risk of prostate cancer metastasis and recurrence in mouse models, indicating a poor outcome after treatment with IL-6/STAT3 inhibitors.48,54

4 | REGULATORS OF THE STAT3 PATHWAY

4.1 | Activation of STAT3

STAT family proteins were first reported in the last three decades as the transcription factors that are activated in a ligand-dependent manner and promote the rapid induction of gene expression.^{55–57} It is reported that more than 40 different polypeptide ligands, including cytokines, JAK kinases, and growth factors, are associated with STAT phosphorylation. As one of the most important members of the STAT family, STAT3 is activated through tyrosine phosphorylation in response to epidermal growth factor (EGF) and IL-6 and others to play an important role in regulating cell proliferation, differentiation, and apoptosis after DNA binding.^{57,58} Once activated by JAK, STAT3 molecules start dimerizing mainly via the SH2 domain with the help of TDAs to form the reciprocal pY-SH2 interactions. Then, phosphorylated STAT3 can translocate into the nucleus and bind to specific DNA to activate the downstream proteins gene expression.^{56,59,60}

4.2 | Positive regulators

As a transcription factor, STAT3 is expressed in almost all types of cells and is tightly activated by upstream regulatory molecules under normal conditions. However, the continuous release of upstream molecules causes the constitutive activation of STAT3 in the TME (Figure 3). JAK/STAT pathway is an important oncogenic signaling cascade activated by multiple adaptor proteins such as IL-6, EGF, and IFN-y.⁶¹ Persistent activation of STAT3 induced by cytokines and growth factors is associated with cell proliferation, differentiation, and apoptosis in cancer.^{62,63} IL-6 is expressed at the high levels in the TME by immune cells, tumor cells, and stromal cells and acts as both pro- and anti-inflammatory cytokine. IL-6 binds to IL-6R on the cell membrane and subsequently forms a protein complex with IL-6R β (gp130 receptor), mediates the activation of STAT3, and promotes tumorigenesis.^{64–66} IL-6 signaling can also be mediated by a trans-signaling pathway, wherein the interaction of IL-6 with the secreted from IL-6R (sIL-6R) can form the IL-6-sIL-6R complex and subsequently bind to gp130.67 Moreover, activated STAT3 can induce IL-6 expression to generate a positivefeedback loop for STAT3 overexpression. The JAK family contains four non-receptor tyrosine kinases (RTKs) including JAK1, JAK2, JAK3, and TYK2. Except JAK3 expressed in hematopoietic cells, others are widely expressed in various cells. With the engagement of gp130, JAK is activated to phosphorylate STAT3 subsequently.⁶⁸

In addition to the IL-6/JAK/STAT3 pathway, the activation of EGFR, fibroblast growth factor receptor, C-X-C chemokine receptor (CXCR), G-protein coupled receptor, and B7-H3, as well as the stimulation with carcinogenic (areca nut extract) and others, can also cause STAT3 phosphorylation via the JAK/STAT3 pathway.^{69–73} It was reported that other classes of non-receptor PTKs could stimulate STAT3 activation. The Src family of kinases, including Src, Lck, Hck, Lyn, Fyn, and Fgr can mediate STAT3 activation. Among them, viral Src was reported to induce constitutive STAT3 activation in a JAKindependent manner. Cellular Src tyrosine kinase was later demonstrated to be a positive regulator of STAT3 activation via stimulation with platelet-derived growth factor and ligand of the human EGFR family.74-76 EGFR, a receptor PTK, is upregulated in most epithelial cancers, and EGFR signaling contributes to cancer cell proliferation and survival. Activated EGFR can directly interact with SH2 domains to phosphorylate STAT3. Moreover, treatment by targeting both EGFR and STAT3 was demonstrated to play an important role in EGFR-STAT3 feedback loop blockade and restriction of pancreatic cancer volume.⁷⁷



FIGURE 3 Schematic of pathways activating STAT3 signaling. Once cytokines and growth factors such as interleukin-6 (IL-6), epidermal growth factor (EGF) and G-proteins as positive regulators bind to their receptors, ligand-bound receptors undergo conformational changes and activate Janus kinase (JAK) family. STAT3 molecules are phosphorylated on Y705 by intracellular non-receptor tyrosine kinases (RTKs) such as JAK and RTKs such as EGF receptor, or STAT3 can also be activated directly by Src and Abl. While PTP, suppressor of cytokine signaling and protein inhibitor of activated STAT as the negative regulators can inhibit the activity of STAT3. The dimerization of two activated STAT3 molecules binding via SH2 domain enters the nucleus and then binds to target gene

Long non-coding RNAs (lncRNAs) with a length of more than 200 nucleotides can induce or reduce STAT3 expression.⁷⁸ MicroRNA (miRNA), as one of the noncoding endogenous RNA, can also induce or reduce STAT3 expression by regulating mRNA degradation or translational inhibition via binding to targeting mRNA molecules.^{79,80} Plenty of studies have found that lncRNAs such as HOX Transcript Antisense RNA (HOTAIR), Gastric Cancer Associated Transcript 3 (GACAT3), Nuclear Paraspeckle Assembly Transcript 1(NEAT1), Forkhead Box D2 Adjacent Opposite Strand RNA 1 (FOXD2-AS1), and Inter-Alpha-Trypsin Inhibitor Heavy Chain 4 Antisense RNA 1 (ITIH4-AS1), as well as miRNAs including miR-24, miR-629, miR-149, miR-495-3p, and miR-34a can act as the positive regulators of the STAT3 pathway.^{81–86}

4.3 | Negative regulators

STAT3 is temporarily activated and tightly regulated by negative regulators that can silence the STAT3 pathway in normal tissues. However, these negative regulators are inhibited in tumor cells. These negative regulators either block the STAT3 signaling pathway or directly act on the STAT3 protein and, thus, may inspire new ideas for cancer treatment (Figure 3).

Due to the significance of tyrosine phosphorylation in STAT3 activation, tyrosine phosphatases can negatively regulate STAT3.⁸⁷ The protein tyrosine phosphatase (PTP) family, which includes PTP receptor-type D (PTPRD), PTP receptor-type T, PTP receptor-type K, SH2-domain-containing PTP1 (SHP1), SHP2, PTP-nonreceptor type 9, and T-cell PTP, is essential for the negative regulation of JAK-STAT3 signaling through STAT3 dephosphorylation.^{88–93} Pertinently, PTPRD is downregulated in nasopharyngeal carcinoma (NPC), and PTPRD overexpression can promote the sensitivity of NPC cells to radiotherapy owing to STAT3 dephosphorylation.⁹⁴ Gupta et al. reported that morin-induced SHP1 expression abrogated the effect of STAT3 and promoted chemosensitization of HNSCC cells.⁸⁸

The suppressor of cytokine signaling (SOCS) proteins family has eight members including SOCS 1–7 and cytokine-inducible SH2-containing protein. Each of them inhibits STAT3 phosphorylation via different mechanisms: directly binding its kinase inhibitory region and SH2 domains to JAK kinase domain to block STAT3 phosphorylation; binding its SH2 domain to JAK-phosphorylated cytokine receptors to prevent STAT3 recruitment.⁹⁵ Considering SOCS1, Yoshikawa et al. showed that silencing of SOCS1 induced the constitutive activation of the JAK2/STAT3 pathway in hepatocellular carcinoma (HCC). The growth of SOCS1 restoration cancer was suppressed with the same efficacy as treatment with the JAK2 inhibitor AG490, which indicated the negative regulation of SOCS in the JAK/STAT3 pathway.⁹⁶ Moreover, SOCS1 expression inhibits the growth and metastasis of prostate cancer by decreasing levels of cyclins D1 and E and cyclindependent kinases (CDK) 2 and 4.⁹⁷

Activated STAT3 is also regulated in the nucleus to regulate gene expression. It was reported that the protein inhibitor of activated STAT (PIAS) can bind to activated STAT dimers and prevent them from binding to DNA.⁹⁸ The most important negative regulator in the PIAS family is PIAS3, and many studies demonstrated that upregulation of PIAS3 expression can inhibit cell proliferation and increase drug chemosensitivity in various tumors.^{99–101} Jiang et al. identified that JAK/STAT hyperactivation induced by SOCS3 and PIAS3 deficiency is associated with the development of early-stage myeloid-derived suppressor cells (eMDSCs) and immunosuppression in breast cancer.¹⁰²

Moreover, various lncRNAs were demonstrated to induce the STAT3 gene expression and were downregulated in cancer tissues, which were associated with the tumor progression and poor prognosis. This negative correlation between STAT3 and specific lncRNAs indicated that lncRNAs could work as the negative regulators of STAT3 pathway.^{103–105} It was reported that lncRNAs such as lncRNA-p21, MEG3, and PTCSC3, as well as miRNAs such as miR-548d-3p and miR-17 cluster family members, can directly target STAT3 and thereby regulate E-cadherin expression.^{103,106–110}

5 | REGULATION OF CANCER HALLMARKS BY THE STAT3 PATHWAY

In 2000, Hanahan et al. proposed hallmarks of cancer, which contained six essential alterations inducing malignant cell growth.¹¹¹ Later in 2011, because of the advancement in cancer research, "energy metabolism" and "immune suppression" were also considered cancer hallmarks. A new generation of cancer hallmarks has been enumerated and includes "sustaining proliferative signaling," "evading growth suppressors," "avoiding immune destruction," "enabling replicative immortality," "tumorMedComm

promoting inflammation," "activating invasion and metastasis," "inducing angiogenesis," "genome instability and mutation," "resisting cell death," and "deregulating cellular energetics."¹¹² Recently, the number of cancer hallmarks increased from 10 to 14. "Unlocking phenotypic plasticity," "senescent cells," "non-mutational epigenetic reprogramming," and "polymorphic microbiomes" were considered as the emerging cancer hallmarks.¹¹³ STAT3, as a multifunctional regulator in cancer formation, development, and metastasis, can influence all cancer hallmarks through the following five ways (Figure 4).

5.1 | Cell proliferation and apoptosis

Cancer is considered a cell cycle disease, and many studies suggest that connections between oncogenes and an abnormal cell cycle are caused by mutations or abnormalities in the expression of cyclins and cyclin-CDKs in a variety of cancers.¹¹⁴⁻¹¹⁶ The activated cyclin-CDK complex formed by the binding of the cyclin box from cyclins with a well-conserved family of protein kinases regulates the progression of cells through the division cycle.¹¹⁴ Cyclin D1 (CCND1) protein, which is one of the major isoforms of D-type cyclins, can interact with CDK4/6 and accelerate the progression of the cell cycle through the G1 phase. CCND1 was reported to be overexpressed and accumulated in association with p-STAT3 in gastric and oral cancer cells. Moreover, after treatment with STAT3 pathway inhibitors, cyclin expression was significantly reduced.^{117,118} Luo et al. showed that upregulation of CCND1 by STAT3 enhanced the proliferation of gastric cancer cells. JAK/STAT3 blockade by AG490 could significantly decrease the CCND1 protein levels to inhibit cancer cells proliferation.¹¹⁸ Yang et al. reported that icaritin can be used to inhibit OSCC cell proliferation via the regulation of the the STAT3 pathway.¹¹⁹

Additionally, the STAT3 pathway is reported to promote cell survival and inhibit apoptosis by modulating the apoptotic regulatory proteins including Bcl-2 family.45,120 As regulator of programmed cell death, Bcl-2 family members are categorized into three subfamilies including prosurvival proteins, BH3-only proapoptotic proteins and multidomain proapoptotic proteins. Cell apoptosis can be evaluated through the expression ratio of proapoptotic to prosurvival factors.¹²¹ Pro-survival proteins such as Bcl-X_L, Bcl-2, and MCL-1 are overexpressed in many cancers and contribute to tumor initiation, progression, and therapeutic resistance.¹²²⁻¹²⁴ Catlett-Falcone et al. demonstrated that the growth and survival of multiple myeloma depended on IL-6 receptor signaling, which is associated with STAT3-induced Bcl-X_L expression. Consequently, blockade of gene regulation mediated by STAT3 could inhibit Bcl-X_L expression and induce apoptosis in cancer



FIGURE 4 Schematic of the relationship between STAT3 and cancer hallmarks. STAT3 improve the representation of tumor hallmarks through the following five aspects: inflammatory microenvironment and immunosuppression; cell proliferation and apoptosis; epithelial-mesenchymal transition and cancer stem-like cells; deregulation of the cellular energetics; angiogenesis and invasion and metastasis

cells.¹²⁵ Oh et al. used licochalconce (LC) H for OSCC treatment, and their results showed that the JAK2/STAT3 pathway and STAT3 target genes such as Bcl-2 were suppressed by LCH, which caused the cell apoptosis and inhibited cell proliferation and colony formation in OSCC cells.¹²⁶

In addition to cancer cells, STAT3 expression in senescent cancer-associated fibroblasts (CAFs) can improve the viability of cancer cells, which is associated with the senescence-associated secretory phenotype. Yasuda et al. demonstrated that the senescence of CAFs induced by proinflammatory cytokines was blocked after the treatment with JAK/STAT3 pathway inhibitors. The JAK/STAT3 inhibitor effectively blocks the peritoneal tumor formation in a mouse model.¹²⁷ Thus, constitutive activation of STAT3 can upregulate the expression of anti-apoptotic proteins and cyclins to regulate cell cycle, promote cell proliferation, and inhibit cell apoptosis.

5.2 | Chronic inflammatory microenvironment and immunosuppression

Tumors are also considered "wounds that do not heal," and many similarities can be found between the microenvironment of chronic inflammatory conditions and tumors. Chemokines and cytokines in TME interact with cancer cells.^{20,128} Dynamic participants in the TME, including tumor-associated macrophages (TAMs), neutrophils, dendritic cells, and regulatory T-cells (Tregs) induce an immunosuppressive TME and enhance the evasion of tumor from immune surveillance, which are associated with cancer progression.^{129–131} PD-L1, an immune checkpoint, plays an important role in delivering pro-survival signals to cancer cells to protect them from tumor-specific immunity.¹³² The over-expressions of PD-L1 in cancer cells and PD-1 in stromal cells were reported to be associated with the phosphorylation of STAT3, which demonstrated the immunosuppressive roles of the STAT3 pathway.¹³³ Xu et al. observed an effective cytotoxicity of natural killer (NK) cells after blockade of the IL-6/JAK/STAT3 pathway and PD-L1, which indicated that NK cell-mediated recognition and cytotoxicity against tumor cells were inhibited via STAT3 pathway-induced PD-L1 expression in tumor cells.¹³⁴ In addition to tumor cells, PD-L1 is also overexpressed via the STAT3 pathway on the surface of neutrophils with the engagement of PD-1 to block the activation of T-cells and NK cells. CAFs derived from HCC are reported to protect neutrophils from apoptosis and promote neutrophils activation via the IL-6/JAK/STAT3 pathway. Once activated, neutrophils inhibit T-cell immunity through the STAT3/PD-L1 axis.135 Tumor-derived granulocyte colony stimulating factor (G-CSF) can also activate the STAT3 pathway in neutrophils and show pro-tumor effects involving dysfunction of NK cells via PD-L1/PD-1 interactions.136

Macrophages, which are regarded as the tissue sentinels, can eliminate or repair damaged cells and matrices to maintain tissue integrity. After the stimulation with different activation programs, macrophages are polarized into two different modes with their own metabolic functions, including M1-like macrophages that produce pro-inflammatory cytokines and M2-like macrophages that have anti-inflammatory and wound healing characteristics.¹³⁷ IL-6-dependent STAT3 activation induces M2 polarization while inhibiting M1 activation, and this promotes cancer progression and reduces patient survival.¹³⁸⁻¹⁴⁰ Besides inducing M2 polarization, the STAT3 pathway also upregulates PD-L1 expression in cancer cells induced by TAMs. Zhang et al. conducted a study to demonstrate how TAMs suppress immune activation and promote the invasive capacity of cancer cells via the STAT3 pathway. The results showed that IFN- γ produced by TAMs enhances the constitutive activation of STAT3 in cancer cells, which is associated with PD-L1 expression.¹⁴¹ Simultaneously, MDSCs play an immunosuppressive role in many cancers, and IL-10/STAT3 signaling in MDSCs suppresses CD8⁺ T-cell proliferation and promotes the development of Tregs in tumors.142-144

Additionally, p-STAT3 in the TME can induce the production of IL-6, IL-10, and vascular endothelial growth factor (VEGF) and transform dendritic cells (DCs) into regulatory DCs that contribute to immune tolerance.^{20,130,145} Kortylewski et al. ablated *STAT3* in hematopoietic cells of adult mice with the *Mx1-Cre-loxP* system via injection of poly(I:C), and the anti-tumor immunity was evaluated. The results showed that *STAT3* deletion in hematopoiMedComm



FIGURE 5 Schematic of STAT3 signaling in tumor microenvironment (TME). STAT3 activation has the ability in affecting TME via up- or downregulating downstream molecules and promoting tumor cell proliferation and survival, angiogenesis, immune evasion as the result. The functions of natural killer cell and dendritic cell in antigen presentation and target cell recognition are inhibited. While macrophage polarization toward M2-like endotype and the immune checkpoint expression and proliferation of myeloid-derived suppressor cell, cancer-associated fibroblasts, and regulatory T-cell are promoted by phosphorylated STAT3

etic cells enhanced the ability of DCs to present antigens and activate T-cells, improved the ability of granulocytes and NKs to eliminate targeting tumor cells, increased T-cell responses to tumor antigens, and inhibited tumor growth in tumor-bearing mice.¹⁴⁶ Constitutive STAT3 activation decreases the expression of T-cell chemotactic factors (such as RANTES and IP-10) to inhibit Tcell chemotaxis. Further, S1PR1 promotes STAT3 activation in CD4⁺ T-cell associated with Tregs accumulation at tumor sites.^{147,148} However, some studies reported contradictory results. Hanna et al. showed that IL-10R/STAT3 pathway correlated with the accumulation of a PD-1^{int} CD8⁺ T-cell subset, which plays an important role in tumor elimination, and the loss of IL-10R/STAT3 signaling enhanced the accumulation of functionally impaired PD-1^{hi} CD8⁺ T-cells associated with the progression of tumors in a chronic lymphocytic leukemia model.¹⁴⁹ Taken together, constitutive activation of STAT3 regulates the adaptive immune response in the chronic inflammatory TME and mediates immunosuppression in cancers (Figure 5).

5.3 | Angiogenesis, invasion, and metastasis

Angiogenesis, which refers to the formation of new blood vessels, is activated by hypoxia and nutrient deprivation to meet the metabolic demands, to remove the waste products from solid tumors, and is involved in initiation, progression, and metastasis of cancer.^{150,151} Tumor angiogenesis is mediated by tumor-secreted angiogenic cytokines, growth factors, and integrins expressed both on angiogenic endothelium and tumor cells. Integrins can integrate signals between the extracellular matrix (ECM) and cellular and chemokines, such as CCL4, to coordinate the migration of circulating endothelial progenitor cells to the hypoxic area.¹⁵² VEGF is associated with vascular development and is regarded as one of the most important factors for the induction of angiogenesis. VEGF can be activated by hypoxic TME and directly upregulated by constitutive activation of STAT3.^{153,154} STAT3 directly regulates VEGF expression via its promoter, and mutations in the STAT3 binding site in the VEGF promoter abrogate this regulation.¹⁵³ VEGF expression can also be indirectly induced by the hypoxic TME through a STAT3/hypoxia-inducible factor 1 alpha (HIF- 1α)-dependent pathway. Hypoxia-induced pSTAT3 accelerates the accumulation of HIF-1 α protein and prolongs its half-life in solid tumor cells, which subsequently enhances VEGF expression.^{155,156} The study by Liu et al. employing sulforaphane to block angiogenesis in HCC demonstrated this indirect mechanism. After sulforaphane treatment, the STAT3/HIF-1 α /VEGF pathway was blocked and the angiogenesis and tumor growth were inhibited.¹⁵⁷ High expression levels of STAT3 and VEGF are also associated with lymph node involvement in esophageal squamous cell cancer, indicating that STAT3/VEGF pathway promotes cancer cell lymphatic metastasis and is correlated with pTNM stage.¹⁵⁸

Moreover, ECM remodeling induced by matrix metalloproteinases (MMPs), which can degrade ECM components is also involved in tumor angiogenesis.¹⁵⁹ In normal tissues, the function of MMPs is counteracted by tissue inhibitors of metalloproteinases (TIMPs) to maintain the balance between angiogenic and anti-angiogenic effects. However, the effect of MMPs is increased in tumor tissues, which break this balance and promote angiogenesis by remodeling the basement membrane and promoting pericyte recruitment.^{152,160} Overexpression of MMPs is also reported to be induced by pSTAT3. Considering MMP-9, Roy et al. identified the mechanisms by which A Disintegrin and Metalloproteinase domain-containing protein 12 (ADAM12) induces angiogenesis in breast cancer. The results showed that ADAM12-regulated angiogenesis was correlated with the upregulation of the proangiogenic factors including VEGF and MMP-9 and the anti-angiogenic factors including TIMP2. The silencing of ADAM12 inhibited STAT3 activation, which demonstrated that angiogenesis can be promoted by activating MMP-9 in a STAT3-dependent manner.¹⁶¹ The elevated gene transcription of MMP-9 was demonstrated to be associated with activation of the STAT3 in tumor-associated myeloid cells.¹⁶² Additionally, MMP overexpression contributes to cancer cell invasion and metastasis, and IL-8/STAT3 signaling in HNSCC can also upregulate the expression of MMP-2 and -9 and play a key role in cancer invasion and metastasis.^{163,164} Further, the migration of endothelial cells (ECs) during angiogenesis can be facilitated with the help of MMP-2/9 overexpression. In addition to important features of STAT3 pathway-related angiogenesis including cell migration, invasion, and tube formation, constitutive activation of STAT3 in CAFs and tumor cells can regulate collagen fibrogenesis and collagen disorganization and fibroblast contractility resulting in increased cancer invasion and metastasis.^{20,165,166} Thus, constitutive activation of STAT3 promotes angiogenesis, invasion, and metastasis of cancer cells.

5.4 | Epithelial-mesenchymal transition (EMT) and cancer stem-like cells (CSCs)

EMT is a general phenomenon observed in embryonic development, tissue remodeling, and wound repair. However, the loss of intracellular adhesion and the transition from an epithelial to the mesenchymal phenotype are correlated with tumor progression and can be observed in cancer lesion area by the loss of E-cadherin and gain of vimentin, N-cadherin, and fibronectin.^{167,168} In cancer tissues, the unlocking of the phenotypic plasticity of cancer cells concurs with EMT, and the EMT-related regulators including STAT3 are associated with cancer cell dedifferentiation.¹⁶⁹ Studies have demonstrated that downstream of STAT3, estrogen-regulated zinc transporter LIV-1, miR-21, IL-6, HIF, and VEGFR2 can repress Ecadherin and modulate EMT in many cancers.¹⁷⁰⁻¹⁷⁴ Dysregulation of the STAT3 pathway in CAFs in pancreatic ductal adenocarcinoma (PDAC) samples was reported to be associated with transforming growth factor beta 1 secretion and the significant single-cell population shifts toward EMT, leading to the aggressive behavior of PDAC.175

CSCs are a heterogeneous population of cancer cells that play an important role in cell self-renewal and differentiation. CSCs are associated with tumor regeneration and metastasis and are considered indicators for the prognosis of patients with cancer.^{176–178} Activation of the STAT3dependent pathways including the STAT3/FAK/ERK pathway, STAT3/Oct-4 pathway in various cancer cells, and the IL-8/STAT3 pathway and EGFR/STAT3/Sox-2 pathways in TAMs are shown to contribute to the cancer stemness.¹⁷⁹⁻¹⁸² Moreover, a link between EMT and CSCs is demonstrated.^{183,184} The CSCs are associated with EMT exhibit and maintain EMT after cancer treatment. EMT phenotypes promote the metastatic proliferation of CSCs among cancer cells. This is crucial for treatment resistance and occurrence, progression, and recurrence of various tumor types.^{151,185} STAT3 pathway is reported to regulate the EMT and CSC in cancer development. Oncostatin M (OSM), a crucial inducer of JAK/STAT3 pathway activation, promotes EMT and generates cells with CSC properties in a STAT3-dependent manner.^{186,187} To further explore the mechanism by which the OSM/STAT3 pathway induces EMT and CSCs, Junk et al. infected epithelial/non-CSCs with retroviruses encoding shS-MAD2, SMAD3, SMAD4 to inhibit SMAD proteins. SMAD3 knockdown significantly reduced mesenchymal/CSC expansion, compared to other treatment groups, which implicated the OSM/JAK/STAT3/SMAD3 pathway in EMT induction and CSCs generation.¹⁸⁸ Recent studies also demonstrated that STAT3 activation is involved in cisplatin chemotherapy resistance through the STAT3/Snail pathway, which is associated with the EMT-like phenotype and stem-like properties acquisition. Besides, resistance to 5-fluorouracil, temozolomide, and sorafenib treatment is also related to the enrichment of CSCs caused by p-STAT3 upregulation.^{189–193} Thus, activated STAT3 can promote EMT and maintain the stemness of cancer stem cells, thereby affecting the prognosis of patients with cancer.

5.5 | Deregulation of the cellular energetics

Cancer cells convert pyruvate into lactate irrespective of the presence of oxygen via aerobic glycolysis, also known as the "Warburg effect," and metabolism in cancer cells can accelerate adenosine triphosphate (ATP) synthesis and maintain rapid cell proliferation.¹⁹⁴ An increased glycolytic rate and concomitant limitation of the tricarboxylic acid cycle (TCA cycle), even in the presence of completely functional mitochondria and sufficient oxygen, typify the Warburg effect.^{195,196} The mechanism underlying the Warburg effect and its association with cancer cell proliferation is not fully understood, but the STAT3 pathway is demonstrated to contribute to Warburg's vicious circle. Studies show that STAT3 activation is associated with the M2 isozyme of pyruvate kinase (PKM2)/HIF-1 α positive feedback loop and promotes proliferation of many types of cancer cells, such as human HCC cells, breast canMedComm

cer cells, and colorectal cancer cells. PKM2 is typically present in tumor cells and can be activated by HIF-1 α in response to oxygen deprivation, cytokines, and oncogenes in the TME. Activated PKM2 can promote HIF gene transcription and STAT3 activation, and pSTAT3 can induce the expression of HIF-1 α and MEK5 and, thereby, enhance the Warburg effect.¹⁹⁷⁻²⁰¹ The expression of hexokinase 2 (HK2), which is accelerated by the STAT3 pathway, is also involved in the Warburg effect. Pu et al. showed that circCUL3 was upregulated in gastric cancer tissues and activated HK2 expression by targeting the miR-515-5p/STAT3 pathway. The results demonstrated molecular interactions between activated STAT3 and HK2.202 Thus, STAT3 signaling promotes the Warburg effect in cancer tissues by upregulating downstream glycolytic proteins; it also promotes cancer cell proliferation related to glycolysis through mechanisms that are not completely understood.

6 | STAT3 AS A THERAPEUTIC TARGET IN CANCER TREATMENT

Considering that aberrantly activated STAT3 is characteristic of cancer formation, progression, and metastasis, recent studies highlighted the effectiveness of blocking STAT3 signaling in cancer treatment (Table 1). In general, constitutive activation of STAT3 signaling can be disrupted by the following mechanisms: (1) indirect STAT3 inhibition by targeting upstream molecules, including IL-6, EGFR, and JAK; (2) direct STAT3 inhibition by blocking dimerization, preventing target gene transcription, or decreasing total STAT3 expression (Figure 6).

6.1 | Targeting the STAT3 pathway for cancer treatment

6.1.1 | Targeting upstream receptors

Targeting JAK

JAKs are the key activator of the STAT3 pathway. The JAK/STAT pathway is activated by various cytokines, and many JAK inhibitors focus on the potential treatment of chronic inflammatory disorders and CRS.^{262–264} The efficacy and safety of several JAK inhibitors are evaluated in clinical trials. Additionally, some orally available, ATP-competitive, small-molecule JAK inhibitors are considered for treating solid tumors.²⁶⁵

Lestaurtinib (CEP-701), the "first generation" FMS-like tyrosine kinase-3 (FLT3) inhibitor that also inhibits JAK2, is reported to inhibit the growth and migration of cancer cells and prevent colony formation of various cancer, 12 of 29

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TABLE 1 Inhibitors targeting signal transducer and activator of transcription 3 (STAT3) in cancers

Inhibitor	Target	Туре	Cancer type	Clinical trials		
Inhibitors targeting upstream receptors						
Dasatinib ^{203–208}	Src	Small molecule	Breast cancer, head and neck squamous cell carcinoma (HNSCC), leukemia, non-small-cell lung carcinoma (NSCLC), prostate cancer	NCT00924352 NCT00826449 NCT02744768 NCT00385580		
Cetuximab ^{209–213}	Epidermal growth factor receptor	Antibody	Colorectal cancer, HNSCC	NCT02928224 NCT02164916 NCT02358031		
AZD1480 ^{214,215}	Janus kinase (JAK)	Small molecule	HNSCC, colorectal cancer	-		
Ruxolitinib ^{216–218}	JAK	Small molecule	Breast cancer, pancreatic cancer, NSCLC	NCT01423604 NCT01594216 NCT02155465		
Tofacitinib ²¹⁹	JAK	Small molecule	Lymphocytic leukemia	-		
8αTGH	JAK	Natural compound	HNSCC	-		
Curcumin ^{221,222}	JAK	Natural compound	Lung cancer, HNSCC, breast cancer	NCT01160302 NCT01740323		
WP1066 ^{223–225}	JAK	Small molecule	HNSCC, gastric cancer, melanoma	-		
Tocilizumab ^{226–228}	IL-6R	Antibody	HNSCC, breast cancer, lymphocytic leukemia	NCT03135171 NCT02906371		
Siltuximab ^{229,230}	IL-6R	Antibody	Prostate cancer, NSCLC	NCT00433446 NCT00841191		
Bazedoxifene ²³¹	IL-6R	Small molecule	Pancreatic cancer	-		
Metformin ^{232,233}	IL-6R	Small molecule	HNSCC, ovarian cancer, breast cancer	NCT01579812 NCT01340300		
Inhibitors blocking STAT3 of	limerization					
S3I-M2001 ²³⁴	SH2	Peptidomimetic	Breast cancer	-		
S3I-201 ²³⁵⁻²³⁷	SH2	Small molecule	AdCC, HNSCC, ASCC	-		
STA-21 ²³⁸	SH2	Small molecule	Breast cancer	-		
Stattic ²³⁹	SH2	Small molecule	Nasopharyngeal carcinoma	-		
OPB-51602 ²⁴⁰⁻²⁴²	SH2	Small molecule	Hematological malignancie, refractory solid malignancies	NCT01344876 NCT01184807		
OPB-111077 ²⁴³	SH2	Small molecule	Hepatocellular carcinoma (HCC), acute myeloid leukemia	NCT01711034 NCT03197714		
TTI-101 ^{244,245}	SH2	Small molecule	HCC, HNSCC	-		
CJ-1383 ²⁴⁶	SH2	Small molecule	Breast cancer	-		
Inhibitors targeting STAT3	DNA-binding domain (DE	BD)				
STAT3 decoy ^{247,248}	DBD	Oligodeoxynucleotides (ODNs)	HNSCC, NSCLC	NCT00696176		
G-quartet ODN ^{249,250}	DBD	ODNs	HNSCC, NSCLC	-		
InS3-54A18 ²⁵¹	DBD	Small molecule	Lung cancer, breast cancer	-		
InS3-54 ²⁵²	DBD	Small molecule	NSCLC, breast cancer	-		
BBI608 ^{253–255}	DBD	Small molecule	Colorectal cancer, gastric cancer, glioblastoma	NCT01830621 NCT02315534 NCT02178956		
MMPP ^{256,257}	DBD	Small molecule	NSCLC, ovarian cancer	-		

(Continues)

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TABLE 1 (Continued)

Inhibitor	Target	Туре	Cancer type	Clinical trials			
Inhibitors decreasing STAT3 expression							
AZD9150 ^{258,259}	STAT3 mRNA	Antisense oligonucleotides	Lymphoma, lung cancer, HCC HNSCC	NCT01839604 NCT01563302 NCT03394144 NCT02549651			
MiR-124-3p ²⁶⁰	STAT3 mRNA	MicroRNA	Nasopharyngeal carcinoma	-			
SD-36 ²⁶¹	STAT3 protein	Small molecule	Lymphocytic leukemia	-			



FIGURE 6 Schematic of STAT3 signaling inhibitors. The principle of STAT3 inhibitors in tumor treatment is based on targeting upstream proteins of the STAT3 signaling or directly targeting STAT3; inhibition of upstream cytokine such as IL-6 and tyrosine kinases JAK with small-molecule inhibitors and natural compound such as tocilizumab, siltuximab, curcumin, ruxolitinib. As for inhibiting STAT3 in direct way, NH2-terminal, DNA-binding and SH2 domains can be targeted with OPB-51602, S3I-201, STAT3 decoy, G-quartet oligodeoxynucleotide, AZD9150. Negative regulators of STAT3 can also play a role as STAT3 inhibitors (Figure 3)

including anaplastic thyroid cancer, human neuroblastomas, and mutated acute myeloid leukemia (AML).^{266–268} However, Knapper et al. conducted a prospective randomized assessment to evaluate the efficacy of lestaurtinib administered after each cycle of chemotherapy in FLT3-AML treatment. The results showed that, compared to chemotherapy alone, the combination therapy of lestaurtinib and chemotherapy with concomitant anti-fungal prophylaxis promoted survival, while no other statistically significant clinical benefit was reported, indicating that lestaurtinib may not be the optimal drug for further AML clinical trials.²⁶⁹ Sen et al. showed that AZD1480 abrogated STAT3 phosphorylation induced by IL-6 and exhibited anti-tumor activity in HPV-HNSCC in vivo mouse model (in two patient-derived HNSCC xenograft models, the tumor volume derived from Patient 1 in the AZD1480 administration group was less than 1/2 compared with the vehicle treatment group).²¹⁴ Although AZD1480 is demonstrated to have a potential anti-cancer effect, its adverse events in clinical administration remain a problem. In a Phase 1 clinical trial, dose-limiting toxicities were observed after the administration of AZD1480 in patients with solid tumors.²⁷⁰

As a small-molecule inhibitor of JAK2 phosphorylation, WP1066 shows highly anti-tumor effectiveness in the treatment of AML, melanoma, and bladder cancer. Besides, WP1066 could improve the chemo-sensitivity in various tumor models.^{271–273} For instance, WP1066 sensitizes OSCC cells to cisplatin by targeting the STAT3/miR-21 axis. Zhang et al. reported that after the combination of WP1066 and cisplatin in the treatment of Tca8113/DDP xenograft tumor models, the growth rate was significantly less than the untreated group.²²³ Liu et al. demonstrated that AG490 plays a suppressing role in angiogenesis and reducing MDSCs in TME of HNSCC by inhibiting the JAK2/STAT3 pathway.²⁷⁴

Ruxolitinib (INC424) is an FDA-approved drug that targets JAK1/2 to treat inflammatory diseases. Although it is not approved for cancer treatment by FDA, clinical trials have been conducted. Dao et al. conducted a Phase II study of ruxolitinib to test its safety and efficacy in the treatment of chronic neutrophilic leukemia and atypical chronic myeloid leukemia. It was well-tolerated, with an overall hematologic response rate of 32% among treated patients.²⁷⁵

Natural compounds have also been reported to suppress JAK/STAT3 pathway.²⁷⁶ Chen et al. introduced a betulinic acid derivative, SH479, which can target the JAK/STAT3 pathway to inhibit arthritis.²⁷⁷ Lee et al. demonstrated the function of arctiin in abrogating JAK and Src activation as well as inhibiting the constitutive activation of pSTAT3.²⁷⁸ Pouyfung et al. showed that 8α-tiglovloxyhirsutinolide-13-O-acetate (8αTGH), a compound from Vernonia cinerea, has anti-tumor activity in an HNSCC mouse model through inhibiting the JAK/STAT3 pathway.²²⁰ JAK inhibitors were mainly designed for autoimmune dysfunction treatment. Although some of them demonstrated anti-tumor utility, the results of clinical trials for cancer treatment were still modest. Further studies using JAK inhibitors in combination therapies may provide more promising results.

Targeting IL-6 and IL-6R

As mentioned, IL-6 is an inflammatory cytokine that activates the JAK1 and JAK2 pathways, leading to STAT3 phosphorylation. IL-6 is elevated in the serum of cancer patients and has diagnostic and/or prognostic significance.^{64,65} The blockade of IL-6 activity by IL-6 and IL-6R neutralizing antibodies is already used to treat rheumatoid arthritis (RA).²⁷⁹ Moreover, it was reported that tocilizumab, an IL-6R blocking antibody that is FDA-approved for RA treatment, can sensitize tumor cells to radiation and overcome erlotinib resistance. Tocilizumab in combination with erlotinib is used to treat SQ20B xenograft tumor models. After 40 days of combination therapy, the survival rate of mice was 40%, compared with 0%, in the erlotinib only

group, which suggests that this combinatorial approach may improve the treatment response and survival rate of patients.^{226,280}

Siltuximab (CNTO-328) is a monoclonal antibody that can specifically target IL-6 and prevent its binding to IL-6R. IL-6 is upregulated in cholangiocarcinoma and non-small cell lung cancer, and treatment with siltuximab alters the IL-6/STAT3 pathway and suppresses the growth and invasion of tumor.^{281,282} The results of Phase I/II and II clinical trials conducted by Angevin et al. and Coward et al., respectively, indicated that siltuximab was safe in ovarian and Kirsten rat sarcoma-2-mutant cancers treatment.^{283,284} However, these several clinical trials of siltuximab in treating patients with cancer was still in their early stage such as Phases I or II due to the adverse events during the cancer treatment. For instance, in a Phase II trial that evaluated mitoxantrone/prednisone with or without siltuximab in prostate cancer treatment, the combination group presented higher percentages of patients with severe adverse events, grade P3 adverse events. Three patients in the combination-treated group were dead due to siltuximab side effects.²⁸⁵ Although siltuximab was regarded as a promising drug in cancer treatment, the way to reduce adverse events associated with siltuximab still need to be explored.

6.1.2 | Targeting STAT3 directly

Blocking dimerization

SH2 domains recognize and bind to select phosphotyrosine residues on receptors and other proteins to form the multiprotein complexes and mediate the intracellular protein–protein interactions.²⁸⁶ There are two ways to inhibit STAT3 directly by targeting SH2: blocking the recruitment of activated RTKs or non-receptor kinases that phosphorylate Tyr705 of STAT3 to the plasma membrane and blocking the dimerization of two activated STAT3 molecules. Two types of inhibitors, including peptides and small molecules targeting the SH2 domain of STAT3, have been reported to reduce the tumor cells proliferation.^{286,287}

Peptidomimetic inhibitors, which are designed via amino acid residues in a STAT3 structure-based manner, can directly interact with the SH2 domain. PY*LKTK can form inactive STAT3:PY*LKTK complexes and effectively reduces levels of active STAT3:STAT3 dimers in vivo.³³ ISS 610, which is based on PY*L, was reported to induce the apoptosis of STAT3-dependent transformed cancer cells without affecting normal cells. S3I-M2001, an oxazole-based peptidomimetic, can inhibit the growth of tumors in mice with breast cancer.^{234,288} Ac-pTyr-Leu-Pro-Gln-Thr-Val-NH2, BP-PM6, BP-PM7, and PMM-172 are all STAT3 inhibitors that target the SH2 domain to reduce

constitutive STAT3 phosphorylation in cancer cells such as HNSCC and breast cancer cells.^{289–291}

Non-peptidic small-molecule inhibitors also block STAT3 activation. Genini et al. investigated the mechanism by which OPB-51602 interferes with STAT3 function in cancer cells. Binding of OPB-51602 to the SH2 domain disrupts intradomain interactions and promotes STAT3 aggregation, and this causes mitochondrial dysfunction and ultimately leads to cancer cell death.²⁴⁰ Kim et al. assessed another inhibitor targeting the SH2 domain of STAT3 called ODZ10117. It can inhibit tyrosine phosphorylation and STAT3 dimerization and thereby reduce tumor growth and lung metastasis in patients with breast cancer.²⁹² Song et al. identified STA-21 as the best-match small-molecule inhibitor targeting the SH2 domain of STAT3 among nearly 429,000 compounds selected through serial functional evaluation based on their established cellbased assays. The breast cancer-bearing mouse was used to evaluate its anti-tumor effect, and STAT3 phosphorylation and dimerization were presented to be abrogated.²³⁸ Moreover, studies have shown that STA-21 treatment can inhibit cell growth, induce apoptosis in bladder cancer cell lines, and improve psoriasis lesions.^{293,294} Schust et al. introduced a small-molecule inhibitor of STAT3 activation and dimerization called Stattic, which also targets the SH2 domain. Stattic treatment with 10 μ M resulted in the apoptosis of approximately 18% of STAT3-dependent MDA-MB-435S breast cancer cells.³⁴ Chen et al. reported a novel small-molecule inhibitor, N4, which can bind to the SH2 of STAT3 and abolish p-STAT3 (Tyr705). N4 suppresses tumor growth and metastasis in animal models of pancreatic cancer. After 20 days of treatment with 20 μ M/kg N4 in pancreatic cancer-bearing mice, the tumor weights of the mice were significantly lower than those of the control group (p < 0.0001, n = 6).²⁹⁵ Wang et al. reported that HJC0152, designed as an O-alkylaminotethered derivative of niclosamide, can inhibit p-STAT3 (Tyr705) and decrease the invasion and migration ability of HNSCC cells.²⁹⁶ LY5 was designed as a small-molecule inhibitor that blocks the phosphotyrosine-binding site of the SH2 domain at nanomolar concentrations. However, Yu et al. showed that the anti-tumor effects of LY5 were not due to the inhibition of STAT3 phosphorylation.²⁹⁷ Bu et al. found that S3I-201 reduced MDSCs and improved CSC eradication, which enhanced the efficacy of conventional chemotherapy in an HNSCC mouse model. Six weeks after treatment with S3I-201 in this mouse model, tumor growth was significantly reduced, compared to that in the PBS control group (p < 0.001, n = 6). Further research has shown that S3I-201 can inhibit STAT3 activation and thereby impair cancer cell proliferation and immunosuppression without appreciable side effects in a mouse model of HPV-negative anal squamous cell carcinoma.^{236,237,298}

Targeting the STAT3 DBD

As mentioned previously, the function of activated STAT3 in cancer cell proliferation, migration, and invasion relies on the physical interaction between the DBD and DNA after translocation into the nucleus. Inhibitors targeting the STAT3 DBD may also decrease STAT3 activity by disrupting its binding to the target DNA.²⁹⁹ Previously, the DBD of STAT3 was considered undruggable because of its potentially limited selectivity.³⁰⁰ Turkson et al. reported that platinum-containing compounds can selectively block STAT3, and it has been shown that platinum (IV)-containing complexes, CPA-1 and CPA-7, disrupt the DNA-binding capacity of STAT3 and induce tumor regression.³⁰¹ Recently, a novel smallmolecule lead compound called (E)-2-methoxy-4-(3-(4methoxyphenyl)prop-1-en-1-yl)phenol (MMPP) was synthesized by Son et al., and it can suppress cancer growth by directly interacting with the DBD of STAT3.²⁵⁶ Compared to the bioactive compound (E)-2,4-bis(p-hydroxyphenyl)-2-butenal (BHPB), which was synthesized previously, MMPP circumvented the problem caused by the presence of α , β -unsaturated carbonyl groups in BHPB that form a non-selective covalent bond, which is conducive to side effects.^{256,302} Leong et al. showed that the using of a STAT3 decoy is an alternative approach for targeting the activated STAT3. Treatment of HNSCC cells with the STAT3 decoy suppresses cancer cells growth and STAT3mediated gene expression.²⁴⁷ Moreover, the decoy can also block STAT3 activation in vivo, induce apoptosis, and decrease the proliferation of HNSCC cells. After treatment with the STAT3 decoy combined with cisplatin for 10 days in an HNSCC xenograft model, there was a 7.4-fold increase in the number of apoptotic cells compared to that in the control group (p = 0.002).³⁰³ Jing et al. developed a series of G-quartet oligodeoxynucleotides (GQ-ODNs) as STAT3 inhibitors that can block its DNA-binding activity. Treatment with GQ-ODN plus paclitaxel in nude mice with HNSCC tumors over 21 days showed potent efficacy. The mean tumor size decreased by 35%, compared to the 9.4-fold increase in the untreated control group.²⁵⁰

Decreasing STAT3 expression

Antisense oligonucleotides (ASOs) can selectively inhibit the translation of STAT3 mRNA by binding to single-stranded RNA sequences in a complementary fashion.^{304,305} Li et al. showed that treatment with a specific 2-O-methoxyethyl-modified ASOs, used to knock down STAT3 expression in HCC cells can decrease circulating VEGF, reduce neovascularization, and inhibit metastasis and growth of cancer cells.³⁰⁶ Oweida et al. demonstrated that treatment with STAT3 ASOs in combination with radiotherapy can increase the anti-tumor effect and decrease radioresistance. After treatment for LY2 and MOC2 tumor-bearing mice with STAT3 ASOs and radiation, the average tumor volumes were 53.0 (5.6) mm^3 and 254.8(81.6) mm,³ respectively, compared to 277.4(53.8)mm³ and 1042.9 (326.8) mm³ in mice treated with STAT3 ASOs alone.³⁰⁷ AZD9150, a second-generation ASO, inhibited STAT3 expression and downstream signaling targets by reducing STAT3 mRNA in lymphoma and lung cancer models.²⁵⁸ Treatment with AZD9150 combined with cisplatin in mice can significantly sensitize tumors to cisplatin and improve the survival rate, compared to treatment with either agent alone, which demonstrates a potential clinical benefit to overcome chemoresistance in HNSCC.²⁵⁹ Moreover, in addition to ASOs, miR-124-3p was reported to downregulate the transcription of STAT3 by interfering with its 3'untranslated region, and this was associated with apoptosis of NPC cells as well as inhibition of proliferation, migration, and invasion.²⁶⁰

6.2 | STAT3 pathway inhibitors in clinical trials against cancer

To date, several STAT3 pathway inhibitors are approved by the FDA for clinical cancer treatment. However, various other STAT3 pathway inhibitors have been used in clinical trials. As mentioned above, lestaurtinib (NCT00557193), AZD1480, ruxolitinib (NCT02092324), and siltuximab have been evaluated for their safety and anti-tumor effects in cancer treatment in clinical trials.^{269,270,275,283,308,309} EGFR, an activator of the STAT3 pathway, is targeted in clinical trials. Cetuximab is a monoclonal antibody against EGFR that can inhibit STAT3 activation. Carter et al. conducted a Phase 1 trial (NCT01445405) to explore the effect of bortezomib and cetuximab in combination with radiation therapy for advanced HNSCC. The median progression-free survival of patients treated with combination therapies was just 4.8 months, which indicated limited anti-tumor efficacy.³¹⁰ However, a Phase 2 trial (NCT00084318) showed that delivery of postoperative chemoradiotherapy (using cisplatin or docetaxel) with cetuximab for HNSCC treatment achieved 69% and 79% 2year overall survival and 57% and 66% 2-year disease-free survival.311

Besides the inhibitors of STAT3 pathway activators, including JAK, IL-6, and EGFR, the results of drugs directly targeting STAT3 in early-phase clinical trials demonstrated their potential in clinical use. Grandis et al. conducted an early Phase 0 trial (NCT00696176) of a STAT3 decoy for HNSCC treatment, and results demon-

strated that none of the patients suffered Grade 3/4 or dose-limiting toxicities. Moreover, patients with HNSCC received intratumoral injections of the STAT3 decoy before surgery, and levels of STAT3 target genes, including cyclin D1 and Bcl-XL, were reduced, compared to those in the control group (saline).³¹² Owing to the promising antitumor effect of AZD9150 in preclinical studies, Hong et al. conducted a Phase I dose-escalation clinical trial. A total of 25 patients with lymphoma and lung cancer was included in this study; 5% of them experienced drug-related adverse events, and 11 of the 25 evaluated patients had stable disease or partial response. Moreover, three of six patients had refractory diffuse large B-cell lymphoma (DLBCL), and two presented with a durable partial response.²⁵⁸ Subsequently, AZD9150 was demonstrated to be well-tolerated, and four of 30 patients with DLBCL achieved a response for at least 4 months associated with a 13% of clinical benefit rate in a Phase 1b clinical trial (NCT01563302).³¹³ In 2016, BBI608, a small molecule that directly targets the STAT3 DBD to inhibit STAT3 activation, was approved by the FDA for GEJ and pancreatic cancer treatment. Later, Jonker et al. conducted a Phase III clinical trial (NCT01830621) to test the efficacy of BBI608 for the treatment of refractory advanced colorectal cancer, and the results showed that BBI608 can prolong overall survival to 5.1 months in patients with pSTAT3-positive tumors, compared to 3 months in the placebo group.²⁵³

6.3 | Drug delivery systems for STAT3 inhibitors

In cancer therapy, STAT3 inhibitors play a significant role in improving anti-tumor efficacy and can augment conventional chemotherapeutic effects.^{314,315} However, conventional STAT3 inhibitors have some limitations, including the lack of anti-tumor effects, low selectivity, and high toxicity. To overcome these barriers, many studies have introduced novel drug delivery systems for STAT3 inhibitors to overcome these limitations. In nanomaterial-based drug delivery systems, STAT3 inhibitors described above are encapsulated into nanomaterials including biomimetic materials, liposomes, polymers, and inorganic materials (Table 2). Compared to the conventional STAT3 inhibitors, nanomaterial-based drug delivery systems have the following advantages: (1) biocompatibility and tumor targeting that reduce cytotoxicity to normal tissues; (2) efficient drug-loading capacity that can be used as a carrier for more than one drug; and (3) protection of loaded drugs from clearance during blood circulation.^{316,317}

Liposomes, which are cell membrane-like structures, have an efficient drug-loading ability as well as biocompatibility; hydrophobic drugs can be readily loaded inside the

Material	Drug	Delivery routes	Cells/Tissues Specificity	Ref
Liposomes	Curcumin-loaded liposomes-STAT3 siRNA	Intratumoral administration	Skin cancer	318
	Hyaluronic acid/TN-CCLP	Intravenous administration	Breast cancer	319
	Rop-DPRL and calorie restriction	Intraperitoneal injection	Melanoma	320
	Stattic	_	Melanoma cells	321
Polymers	Gel- NSC74859-ICG	Intravenous administration	HNSCC	322
	Ritonavir derivative	Intravenous administration	HNSCC	323
	Cucurbitacin-D; doxorubicin	Intravenous administration	Breast cancer	324
	$HA/^{siSTAT3}PPL_{PTX}$	Intravenous administration	Breast cancer	325
Biomimetic material	Exo-JSI124	Intranasal delivery	Glioblastoma tumor	326
	Tumor-derived exosomes- miR-34a	_	Colorectal cancer cells	327
	Corosolic acid-long-circulating liposomes-αCD163	-	Tumor-associated macrophages	328
	NPs-αIL6R Ab-CD44	Intravenous administration	Breast cancer	329
	CaP-cored low-density lipoprotein nanovehicle-STAT3 decoy ODNs	Intravenous administration	HCC cells	330
Inorganic material	AuNP-NUAP-STAT3d	-	HNSCC cells	331
	AIRISE-02(STAT3 siRNA-CpG- mesoporous silica nanoparticle)	Intratumoral administration	Breast cancer	332
	Layer-by-layer assembled gold nanoparticles-STAT3 siRNA -imatinib	Intratumoral administration	Melanoma	333
	SPION-TMC-ChT-TAT-H NPs	Intravenous administration	Colorectal cancer	334

lipid bilayer membranes.³³⁵ Shi et al. designed a calcium phosphate-cored low-density lipoprotein nanovehicle as a Trojan horse to load STAT3 decoy ODNs to overcome tumor necrosis factor-related apoptosis-inducing ligand resistance.³³⁰ Wu et al. introduced PEGylated liposomal FLLL32, a specific STAT3 inhibitor that binds to the SH2 domain. It enhanced anti-tumor efficacy and reduced toxicity through intravenous administration in treating pancreatic cancer.³³⁶ Exosomes that are liposomes with antigens, mRNAs, and miRNAs derived from various cells are promising vehicles for cancer treatment to overcome the tumor barrier to drug delivery. Zhuang et al. introduced a non-invasive method to deliver a STAT3 inhibitor loaded into exosomes via an intranasal route. This inhibitor can be taken up by microglial cells, and it can prolong the survival time of GL26 tumor-bearing mice to 44.5 days, compared to the control group.³²⁶

Zheng et al. introduced poly lactic-co-glycolic acid (PLGA) nanoparticles to co-deliver the cancer drug doxorubicin (Dox) and the STAT3 inhibitor nifurate (DNNPs) and achieve long-term drug release. After 2 h of treatment with DNNPs, the fluorescence intensity of Dox, which reflected DNNP uptake by BGC-823 gastric tumors in mice, was approximately three-fold that of the

Dox control group (p < 0.001).³³⁷ Tavares et al. introduced HPMA-based copolymers containing cucurbitacin-D (CuD), a STAT3 inhibitor, and Dox. In this drug delivery system, CuD can be released in a sustained, controlled manner and can effectively target breast cancer. After treatment with linear-CuD in combination with micellar-Dox in 4T1 tumor-bearing mice, the tumor volume decreased significantly, compared to that in the linear-CuD control group.³²⁴ Additionally, drug delivery systems introduced by Lavasanifar and colleagues including PEO-b-P(CL-JSI-124) conjugates, the PLGA-JSI-124 conjugate, PEO-b-PBCL micelles are demonstrated to be efficacy in cancer treatment.^{338–340}

In addition to enhancing the anti-tumor efficacy of STAT3 inhibitors through different drug delivery systems. STAT3 inhibitors can also be combined with other therapies to enhance the efficacy and safety of tumor treatment strategies. The immune checkpoint protein blockade is a promising immunotherapy for cancer treatment. However, resistance to durvalumab (the anti-PD-L1 antibody) and tremelimumab (the anti-CTLA4 antibody) associated with STK11 gene mutations is reported in non-small-cell lung carcinoma (NSCLC) treatment, and this has impacted the efficacy of immune blockade. It was demonstrated that STAT3 ASOs reversed this resistance, and STAT3 ASOs administration in combination with immune checkpoint blockade significantly enhanced the anti-tumor efficacy, compared to immunotherapy alone.³⁴¹

CAR-T cell therapy is a novel immunotherapy that demonstrated excellent efficacy in the treatment of hematological malignancies. However, overexpression of cytokines, such as IL-6 and IL-10, caused by STAT3 hyperactivation is associated with the occurrence of CRS. Therefore, treatment with the STAT3 signaling pathway inhibitor tocilizumab in combination with CAR-T cell therapy can reduce adverse events without impeding CAR-T cell expansion.³⁴² Guha et al. demonstrated that STAT3 inhibitors of liver-associated MDSCs (STATTIC or BBI608) can enhance the anti-tumor efficacy of CAR-T cells. In the murine liver metastasis model, STAT3 inhibition did not decrease CAR-T cell cytotoxicity nor cause the death of CAR-T cells, and apoptosis of tumor cells dosedependently increased, compared to the control group.³⁴³ Because STAT3 is associated with upregulation of PD-L1 expression, STAT3 inhibitors play a role in immunotherapy, and STAT3 inhibition combined with photothermal therapy based on gelatinase-sensitive gelatin nanoparticles was reported to enhance anti-tumor efficacy in HNSCC cancer treatment.¹³³ On the 15th day after combination therapy, levels of immunosuppressive MDSCs and PD-1 in the blood, tumor, and spleen were significantly reduced, and the growth of tumor volume was limited in the Tgfbr1/Pten double conditional knockout (2cKO) mice model, compared to the untreated group.³²²

As mentioned, STAT3 inhibitors loaded into organic materials, inorganic materials, and unique biomimetic materials have demonstrated great tumor-targeting, biocompatibility, and anti-tumor efficacy.

7 | CONCLUSION AND PERSPECTIVES

The STAT3 pathway plays a physiological role in normal cell growth, differentiation, and survival. Meanwhile, as the "Achilles' heel" of many cancers, owing to the association of constitutive STAT3 activation with cancer hallmarks as well as the relationship between STAT3 pathway and poor patient outcomes, many researchers believe that STAT3 acts as an oncogene and that blockade of STAT3 activity is a viable strategy for cancer treatment. Additionally, based on clinical trial results, various STAT3 inhibitors are well-tolerated and potentially improve the overall survival of patients with cancer.

Despite improvements in STAT3 targeting therapeutics, the conventional view is that STAT3 is "undruggable" owing to its ubiquitous expression. Only a few clinically available STAT3 inhibitors for cancer treatment are approved by the FDA, and there are several concerns that remain worthy of attention and consideration. First, STAT3 inhibitors should not affect the functions of structurally similar proteins, such as STAT1, and the side effects of STAT3 inhibitors, including hematologic toxicity, are still major problems to be addressed. Further efforts should be made to clarify the mechanisms of selectivity for STAT3 inhibition and to explore the novel delivery of STAT3 inhibitors to reduce toxicity, increase local concentrations, enhance cellular internalization, and achieve long-term release. Second, the development of STAT3 inhibitors is focused on STAT3:STAT3 dimerization, which is a dilemma because of the lack of a reliable off-target biomarker. Thus, in addition to the SH2 domain, further research should explore the targeting of other STAT3 domains. Recently, with the development of purchasable chemical databases and computer-aided drug discovery, the undruggable STAT3-DNA interface is targeted via artificial intelligence, which can virtually screen (VS) billions of molecular structures.³⁴⁴ For example, inS3-54, an inhibitor of STAT3 that targets its DBD, was designed via a VS of 200,000 compounds that excluded molecules predicted to bind STAT1. Moreover, to prevent off-target side effects, inS3-54 analogs were searched in the virtual Chemdiv database, and analog A18 was demonstrated to specifically bind to the DBD of STAT3 and inhibit its activity.^{251,252} Third, the drug delivery system and the combination of STAT3 inhibitors with other anti-tumor therapies can greatly enhance cancer treatment. For instance, STAT3 inhibitors in combination with CAR-T cell therapy can reduce adverse events, STAT3 inhibitors loaded into nanocarriers have enhanced serum stability to promote anti-cancer efficacy, and STAT3 inhibitors in combination with radiation can overcome radiotherapy resistance. In summary, with the progress of research on STAT3 pathway and advances in clinical trials of STAT3 inhibitors, the significance of STAT3 pathway in anti-tumor activity is evident. In the future, multimodal delivery of drugs targeting STAT3 pathway and other conventional therapies could be a promising strategy for cancer treatment.

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CONFLICT OF INTEREST

The authors have no conflict of interest to declare.

ETHICS STATEMENT

No ethical approval was required for this study.

AUTHOR CONTRIBUTION

L.L.B., L.L.C., and B.L. conceived the idea; H.Q.W., Q.W.M., and F.Y.H. performed the literature search and drafted the manuscript; X.G., H.L., S.R.L., J.W., F.C.S., Y.S. revised and edited the manuscript; L.L.B., L.L.C., and B.L. supervised and revised the manuscript. The authors read and approved the final manuscript.

DATA AVAILABILITY STATEMENT

Not applicable.

REFERENCES

- 1. Sung H, Ferlay J, Siegel RL, et al. Global cancer statistics 2020: gLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* 2021;71(3):209-249.
- 2. World Health Organization. World Health Statistics 2021: Monitoring Health for the SDGs, Sustainable Development Goals. World Health Organization; 2021.
- 3. Ciombor KK, Bekaii-Saab T. A comprehensive review of sequencing and combination strategies of targeted agents in metastatic colorectal cancer. *Oncologist.* 2018;23(1):25.
- 4. Addeo A, Banna GL, Metro G, Di Maio M. Chemotherapy in combination with immune checkpoint inhibitors for the firstline treatment of patients with advanced non-small cell lung cancer: a systematic review and literature-based meta-analysis. *Front Oncol.* 2019;9:264.
- Chen J, Ni Y, Sun G, et al. Comparison of current systemic combination therapies for metastatic hormone-sensitive prostate cancer and selection of candidates for optimal treatment: a systematic review and Bayesian network meta-analysis. *Front Oncol.* 2020;10:1806.
- Dobson GP. Addressing the global burden of trauma in major surgery. *Front Surg*. 2015;2:43-43.
- Nikolaou M, Pavlopoulou A, Georgakilas AG, Kyrodimos E. The challenge of drug resistance in cancer treatment: a current overview. *Clin Exp Metastasis*. 2018;35(4):309-318.
- Nurgali K, Jagoe RT, Abalo R. Adverse effects of cancer chemotherapy: anything new to improve tolerance and reduce sequelae? *Front Pharmacol.* 2018;9:245.
- 9. Loh C-Y, Arya A, Naema AF, Wong WF, Sethi G, Looi CY. Signal transducer and activator of transcription (STATs) proteins in cancer and inflammation: functions and therapeutic implication. *Front Oncol.* 2019;9:48.
- Wegrzyn J, Potla R, Chwae YJ, et al. Function of mitochondrial Stat3 in cellular respiration. *Science*. 2009;323(5915):793-797.
- Kano A, Wolfgang MJ, Gao Q, et al. Endothelial cells require STAT3 for protection against endotoxin-induced inflammation. *J Exp Med.* 2003;198(10):1517-1525.

- Laouar Y, Welte T, Fu XY, Flavell RA. STAT3 is required for Flt3L-dependent dendritic cell differentiation. *Immunity*. 2003;19(6):903-912.
- Moh A, Iwamoto Y, Chai GX, et al. Role of STAT3 in liver regeneration: survival, DNA synthesis, inflammatory reaction and liver mass recovery. *Lab Invest*. 2007;87(10):1018-1028.
- Hirano T, Ishihara K, Hibi M. Roles of STAT3 in mediating the cell growth, differentiation and survival signals relayed through the IL-6 family of cytokine receptors. *Oncogene*. 2000;19(21):2548-2556.
- Kim DY, Cha ST, Ahn DH, et al. STAT3 expression in gastric cancer indicates a poor prognosis. *J Gastroenterol Hepatol*. 2009;24(4):646-651.
- 16. Xu Y, Lu S. A meta-analysis of STAT3 and phospho-STAT3 expression and survival of patients with non-small-cell lung cancer. *Eur J Surg Oncol.* 2014;40(3):311-317.
- 17. Kusaba T, Nakayama T, Yamazumi K, et al. Activation of STAT3 is a marker of poor prognosis in human colorectal cancer. *Oncol Rep.* 2006;15(6):1445-1451.
- Zhao Y, Zhang J, Xia H, et al. Stat3 is involved in the motility, metastasis and prognosis in lingual squamous cell carcinoma. *Cell Biochem Funct*. 2012;30(4):340-346.
- Johnson DE, Burtness B, Leemans CR, Lui VWY, Bauman JE, Grandis JR. Head and neck squamous cell carcinoma. *Nat Rev Dis Primers*. 2020;6(1):92.
- Huynh J, Chand A, Gough D, Ernst M. Therapeutically exploiting STAT3 activity in cancer-using tissue repair as a road map. *Nat Rev Cancer*. 2019;19(2):82-96.
- Yu H, Pardoll D, Jove R. STATs in cancer inflammation and immunity: a leading role for STAT3. *Nat Rev Cancer*. 2009;9(11):798-809.
- 22. Lee H, Jeong AJ, Ye SK. Highlighted STAT3 as a potential drug target for cancer therapy. *BMB Rep.* 2019;52(7):415-423.
- Fu X-Y, Kessler DS, Veals SA, Levy DE, Darnell J. ISGF3, the transcriptional activator induced by interferon alpha, consists of multiple interacting polypeptide chains. *Proc Natl Acad Sci.* 1990;87(21):8555-8559.
- 24. Fu XY. A transcription factor with SH2 and SH3 domains is directly activated by an interferon alpha-induced cytoplasmic protein tyrosine kinase(s). *Cell*. 1992;70(2):323-335.
- Wegenka UM, Buschmann J, Lütticken C, Heinrich PC, Horn F. Acute-phase response factor, a nuclear factor binding to acute-phase response elements, is rapidly activated by interleukin-6 at the posttranslational level. *Mol Cell Biol.* 1993;13(1):276-288.
- Akira S, Nishio Y, Inoue M, et al. Molecular cloning of APRF, a novel IFN-stimulated gene factor 3 p91-related transcription factor involved in the gp130-mediated signaling pathway. *Cell*. 1994;77(1):63-71.
- Sgrignani J, Garofalo M, Matkovic M, Merulla J, Catapano CV, Cavalli A. Structural biology of STAT3 and its implications for anticancer therapies development. *Int J Mol Sci.* 2018;19(6):1591.
- Mertens C, Haripal B, Klinge S, Darnell JE. Mutations in the linker domain affect phospho-STAT3 function and suggest targets for interrupting STAT3 activity. *Proc Natl Acad Sci.* 2015;112(48):14811-14816.

- Hu T, Yeh JE, Pinello L, et al. Impact of the N-terminal domain of STAT3 in STAT3-dependent transcriptional activity. *Mol Cell Biol.* 2015;35(19):3284-3300.
- Zhang T, Kee WH, Seow KT, Fung W, Cao X. The coiledcoil domain of Stat3 is essential for its SH2 domainmediated receptor binding and subsequent activation induced by epidermal growth factor and interleukin-6. *Mol Cell Biol.* 2000;20(19):7132-7139.
- Haan S, Hemmann U, Hassiepen U, et al. Characterization and binding specificity of the monomeric STAT3-SH2 domain. J Biol Chem. 1999;274(3):1342-1348.
- 32. Bromberg JF, Wrzeszczynska MH, Devgan G, et al. Stat3 as an oncogene. *Cell*. 1999;98(3):295-303.
- Turkson J, Ryan D, Kim JS, et al. Phosphotyrosyl peptides block Stat3-mediated DNA binding activity, gene regulation, and cell transformation. *J Biol Chem.* 2001;276(48):45443-45455.
- Schust J, Sperl B, Hollis A, Mayer TU, Berg T. Stattic: a small-molecule inhibitor of STAT3 activation and dimerization. *Chem Biol.* 2006;13(11):1235-1242.
- Slebos RJ, Kibbelaar RE, Dalesio O, et al. K-ras oncogene activation as a prognostic marker in adenocarcinoma of the lung. N Engl J Med. 1990;323(9):561-565.
- Jove R, Hanafusa H. Cell transformation by the viral src oncogene. Annu Rev Cell Biol. 1987;3(1):31-56.
- 37. Watson CJ, Miller WR. Elevated levels of members of the STAT family of transcription factors in breast carcinoma nuclear extracts. *Br J Cancer*. 1995;71(4):840-844.
- Levy DE, Inghirami G. STAT3: a multifaceted oncogene. Proc Natl Acad Sci U S A. 2006;103(27):10151-10152.
- Attili I, Karachaliou N, Bonanno L, et al. STAT3 as a potential immunotherapy biomarker in oncogeneaddicted non-small cell lung cancer. *Ther Adv Med Oncol.* 2018;10:1758835918763744.
- Resemann HK, Watson CJ, Lloyd-Lewis B. The Stat3 paradox: a killer and an oncogene. *Mol Cell Endocrinol*. 2014;382(1):603-611.
- Tolomeo M, Cascio A. The multifaced role of STAT3 in cancer and its implication for anticancer therapy. *Int J Mol Sci.* 2021;22(2):603.
- Grandis JR, Drenning SD, Zeng Q, et al. Constitutive activation of Stat3 signaling abrogates apoptosis in squamous cell carcinogenesis in vivo. *Proc Natl Acad Sci U S A*. 2000;97(8):4227-4232.
- Lin GS, Yang LJ, Wang XF, et al. STAT3 Tyr705 phosphorylation affects clinical outcome in patients with newly diagnosed supratentorial glioblastoma. *Med Oncol.* 2014;31(4):924.
- 44. Fei Y, Yu J, Li Y, et al. Plasma soluble PD-L1 and STAT3 predict the prognosis in diffuse large B cell lymphoma patients. *J Cancer*. 2020;11(23):7001.
- 45. Song JI, Grandis JR. STAT signaling in head and neck cancer. *Oncogene*. 2000;19(21):2489-2495.
- Trivedi S, Concha-Benavente F, Srivastava RM, et al. Immune biomarkers of anti-EGFR monoclonal antibody therapy. *Ann Oncol.* 2015;26(1):40-47.
- Lai SY, Johnson FM. Defining the role of the JAK-STAT pathway in head and neck and thoracic malignancies: implications for future therapeutic approaches. *Drug Resist Updat*. 2010;13(3):67-78.
- 48. Luo J, Wang K, Yeh S, et al. LncRNA-p21 alters the antiandrogen enzalutamide-induced prostate cancer neuroendocrine

differentiation via modulating the EZH2/STAT3 signaling. *Nat Commun.* 2019;10(1):2571.

- Li X, Gera L, Zhang S, et al. Pharmacological inhibition of noncanonical EED-EZH2 signaling overcomes chemoresistance in prostate cancer. *Theranostics*. 2021;11(14):6873-6890.
- Shinagawa K, Yanamoto S, Naruse T, et al. Clinical roles of interleukin-6 and STAT3 in oral squamous cell carcinoma. *Pathol Oncol Res.* 2017;23(2):425-431.
- Pectasides E, Egloff AM, Sasaki C, et al. Nuclear localization of signal transducer and activator of transcription 3 in head and neck squamous cell carcinoma is associated with a better prognosis. *Clin Cancer Res.* 2010;16(8):2427-2434.
- 52. Dolled-Filhart M, Camp RL, Kowalski DP, Smith BL, Rimm DL. Tissue microarray analysis of signal transducers and activators of transcription 3 (Stat3) and phospho-Stat3 (Tyr705) in node-negative breast cancer shows nuclear localization is associated with a better prognosis. *Clin Cancer Res.* 2003;9(2):594-600.
- 53. Sonnenblick A, Shriki A, Galun E, et al. Tissue microarraybased study of patients with lymph node-positive breast cancer shows tyrosine phosphorylation of signal transducer and activator of transcription 3 (tyrosine705-STAT3) is a marker of good prognosis. *Clin Transl Oncol.* 2012;14(3):232-236.
- Pencik J, Schlederer M, Gruber W, et al. STAT3 regulated ARF expression suppresses prostate cancer metastasis. *Nat Commun.* 2015;6:7736.
- 55. Schindler C. STATs as activators of apoptosis. *Trends Cell Biol.* 1998;8(3):97-98.
- Bromberg J. The role of STATs in transcriptional control and their impact on cellular function. *Oncogene*. 2000;19(21):2468-2473.
- 57. Mui AL. The role of STATs in proliferation, differentiation, and apoptosis. *Cell Mol Life Sci.* 1999;55(12):1547-1558.
- Zhong Z, Wen Z, Darnell JE. Stat3: a STAT family member activated by tyrosine phosphorylation in response to epidermal growth factor and interleukin-6. *Science*. 1994;264(5155): 95-98.
- Fan Y, Mao R, Yang J. NF-kappaB and STAT3 signaling pathways collaboratively link inflammation to cancer. *Protein Cell*. 2013;4(3):176-185.
- 60. Horvath CM, Wen Z. A STAT protein domain that determines DNA sequence recognition suggests a novel DNA-binding domain. *Genes Dev.* 1995;9(8):984-994.
- Hu X, li J, Fu M, Zhao X, Wang W. The JAK/STAT signaling pathway: from bench to clinic. *Signal Transduc Target Ther*. 2021;6(1):402.
- Mali SB. Review of STAT3 (Signal Transducers and Activators of Transcription) in head and neck cancer. Oral Oncol. 2015;51(6):565-569.
- 63. Bournazou E, Bromberg J. Targeting the tumor microenvironment: JAK-STAT3 signaling. *JAKSTAT*. 2013;2(2):e23828.
- 64. Sansone P, Bromberg J. Targeting the interleukin-6/Jak/stat pathway in human malignancies. *J Clin Oncol*. 2012;30(9):1005-1014.
- 65. Dineshkumar T, Ashwini BK, Rameshkumar A, Rajashree P, Ramya R, Rajkumar K. Salivary and serum interleukin-6 levels in oral premalignant disorders and squamous cell carcinoma: diagnostic value and clinicopathologic correlations. *Asian Pac J Cancer Prev.* 2016;17(11):4899-4906.

- 66. Sriuranpong V, Park JI, Amornphimoltham P, Patel V, Nelkin BD, Gutkind JS. Epidermal growth factor receptorindependent constitutive activation of STAT3 in head and neck squamous cell carcinoma is mediated by the autocrine/ paracrine stimulation of the interleukin 6/gp130 cytokine system. *Cancer Res.* 2003;63(11):2948-2956.
- Johnson DE, O'Keefe RA, Grandis JR. Targeting the IL-6/JAK/STAT3 signalling axis in cancer. *Nat Rev Clin Oncol.* 2018;15(4):234-248.
- Jin W. Role of JAK/STAT3 signaling in the regulation of metastasis, the transition of cancer stem cells, and chemoresistance of cancer by epithelial-mesenchymal transition. *Cells*. 2020;9(1):217.
- Shi T, Papay RS, Perez DM. α(1A)-adrenergic receptor differentially regulates STAT3 phosphorylation through PKCε and PKCδ in myocytes. *J Recept Signal Transduct Res.* 2012;32(2):76-86.
- Chang YF, Liu TY, Liu ST, Tseng CN. Arecoline inhibits myogenic differentiation of C2C12 myoblasts by reducing STAT3 phosphorylation. *Food Chem Toxicol.* 2012;50(10):3433-3439.
- Fan TF, Deng WW, Bu LL, Wu TF, Zhang WF, Sun ZJ. B7-H3 regulates migration and invasion in salivary gland adenoid cystic carcinoma via the JAK2/STAT3 signaling pathway. *Am J Transl Res.* 2017;9(3):1369-1380.
- Li P, Huang T, Zou Q, et al. FGFR2 promotes expression of PD-L1 in colorectal cancer via the JAK/STAT3 signaling pathway. *J Immunol.* 2019;202(10):3065-3075.
- Tan Y, Shu L, Xu P, Bai S. Mesenchymal stem cells attract endothelial progenitor cells via a positive feedback loop between CXCR2 and CXCR4. *Stem Cells Int.* 2019;2019:4197164.
- Yu CL, Meyer DJ, Campbell GS, et al. Enhanced DNA-binding activity of a Stat3-related protein in cells transformed by the Src oncoprotein. *Science*. 1995;269(5220):81-83.
- Silva CM. Role of STATs as downstream signal transducers in Src family kinase-mediated tumorigenesis. *Oncogene*. 2004;23(48):8017-8023.
- Cirri P, Chiarugi P, Marra F, et al. c-Src activates both STAT1 and STAT3 in PDGF-stimulated NIH3T3 cells. *Biochem Biophys Res Commun.* 1997;239(2):493-497.
- Zhao C, Yang L, Zhou F, et al. Feedback activation of EGFR is the main cause for STAT3 inhibition-irresponsiveness in pancreatic cancer cells. *Oncogene*. 2020;39(20):3997-4013.
- Ashrafizadeh M, Gholami MH, Mirzaei S, et al. Dual relationship between long non-coding RNAs and STAT3 signaling in different cancers: new insight to proliferation and metastasis. *Life Sci.* 2021;270:119006.
- Dong H, Lei J, Ding L, Wen Y, Ju H, Zhang X. MicroRNA: function, detection, and bioanalysis. *Chem Rev.* 2013;113(8):6207-6233.
- Kohanbash G, Okada H. MicroRNAs and STAT interplay. Semin Cancer Biol. 2012;22(1):70-75.
- Zhou W, Wang L, Miao Y, Xing R. Novel long noncoding RNA GACAT3 promotes colorectal cancer cell proliferation, invasion, and migration through miR-149. *Onco Targets Ther*. 2018;11:1543-1552.
- Liang C, Zhao T, Li H, et al. Long non-coding RNA ITIH4-AS1 accelerates the proliferation and metastasis of colorectal cancer by activating JAK/STAT3 signaling. *Mol Ther Nucleic Acids*. 2019;18:183-193.

- Deng S, Wang J, Zhang L, Li J, Jin Y. LncRNA HOTAIR promotes cancer stem-like cells properties by sponging miR-34a to activate the JAK2/STAT3 pathway in pancreatic ductal adenocarcinoma. *Onco Targets Ther.* 2021;14:1883-1893.
- Xia D, Yao R, Zhou P, Wang C, Xia Y, Xu S. LncRNA NEAT1 reversed the hindering effects of miR-495-3p/STAT3 axis and miR-211/PI3K/AKT axis on sepsis-relevant inflammation. *Mol Immunol.* 2020;117:168-179.
- Li R, Chen S, Zhan J, et al. Long noncoding RNA FOXD2-AS1 enhances chemotherapeutic resistance of laryngeal squamous cell carcinoma via STAT3 activation. *Cell Death Dis.* 2020;11(1):41.
- Gougelet A, Colnot S. MicroRNA-feedback loop as a key modulator of liver tumorigenesis and inflammation. World J Gastroenterol. 2013;19(4):440-444.
- Chen W, Daines MO, Khurana Hershey GK. Turning off signal transducer and activator of transcription (STAT): the negative regulation of STAT signaling. *J Allergy Clin Immunol*. 2004;114(3):476-489. quiz 490.
- Gupta SC, Phromnoi K, Aggarwal BB. Morin inhibits STAT3 tyrosine 705 phosphorylation in tumor cells through activation of protein tyrosine phosphatase SHP1. *Biochem Pharmacol.* 2013;85(7):898-912.
- Yang Y, Jiang B, Huo Y, et al. Shp2 suppresses PyMT-induced transformation in mouse fibroblasts by inhibiting Stat3 activity. *Virology*. 2011;409(2):204-210.
- Kumar V, Cheng P, Condamine T, et al. CD45 phosphatase inhibits STAT3 transcription factor activity in myeloid cells and promotes tumor-associated macrophage differentiation. *Immunity*. 2016;44(2):303-315.
- 91. Lee H, Kim M, Baek M, et al. Targeted disruption of TC-PTP in the proliferative compartment augments STAT3 and AKT signaling and skin tumor development. *Sci Rep.* 2017;7:45077.
- Lee JH, Mohan CD, Shanmugam MK, et al. Vitexin abrogates invasion and survival of hepatocellular carcinoma cells through targeting STAT3 signaling pathway. *Biochimie*. 2020;175:58-68.
- Kim M, Morales LD, Jang I-S, Cho Y-Y, Kim DJ. Protein tyrosine phosphatases as potential regulators of STAT3 signaling. *Int J Mol Sci.* 2018;19(9):2708.
- 94. Lin Y, Zhou X, Yang K, et al. Protein tyrosine phosphatase receptor type D gene promotes radiosensitivity via STAT3 dephosphorylation in nasopharyngeal carcinoma. *Oncogene*. 2021;40(17):3101-3117.
- Durham GA, Williams JJ, Nasim MT, Palmer TM. Targeting SOCS proteins to control JAK-STAT signalling in disease. *Trends Pharmacol Sci.* 2019;40(5):298-308.
- Yoshikawa H, Matsubara K, Qian GS, et al. SOCS-1, a negative regulator of the JAK/STAT pathway, is silenced by methylation in human hepatocellular carcinoma and shows growthsuppression activity. *Nat Genet*. 2001;28(1):29-35.
- Neuwirt H, Puhr M, Santer FR, et al. Suppressor of cytokine signaling (SOCS)-1 is expressed in human prostate cancer and exerts growth-inhibitory function through downregulation of cyclins and cyclin-dependent kinases. *Am J Pathol.* 2009;174(5):1921-1930.
- Wu M, Song D, Li H, et al. Negative regulators of STAT3 signaling pathway in cancers. *Cancer Manag Res.* 2019;11:4957-4969.

- Ma J, Yang Y, Fu Y, et al. PIAS3-mediated feedback loops promote chronic colitis-associated malignant transformation. *Theranostics*. 2018;8(11):3022-3037.
- 100. Lee JH, Kim C, Sethi G, Ahn KS. Brassinin inhibits STAT3 signaling pathway through modulation of PIAS-3 and SOCS-3 expression and sensitizes human lung cancer xenograft in nude mice to paclitaxel. *Oncotarget.* 2015;6(8):6386-6405.
- 101. Saydmohammed M, Joseph D, Syed V. Curcumin suppresses constitutive activation of STAT-3 by up-regulating protein inhibitor of activated STAT-3 (PIAS-3) in ovarian and endometrial cancer cells. *J Cell Biochem.* 2010;110(2):447-456.
- 102. Jiang M, Zhang W, Zhang R, et al. Cancer exosome-derived miR-9 and miR-181a promote the development of early-stage MDSCs via interfering with SOCS3 and PIAS3 respectively in breast cancer. *Oncogene*. 2020;39(24):4681-4694.
- 103. Tan J, Xiang L, Xu G. LncRNA MEG3 suppresses migration and promotes apoptosis by sponging miR-548d-3p to modulate JAK-STAT pathway in oral squamous cell carcinoma. *IUBMB Life*. 2019;71(7):882-890.
- 104. Sharma U, Barwal TS, Khandelwal A, et al. LncRNA ZFAS1 inhibits triple-negative breast cancer by targeting STAT3. *Biochimie*. 2021;182:99-107.
- 105. Wang Y, Wu S, Zhu X, et al. LncRNA-encoded polypeptide ASRPS inhibits triple-negative breast cancer angiogenesis. J Exp Med. 2020;217(3):jem.20190950.
- 106. Najm A, Masson FM, Preuss P, et al. MicroRNA-17-5p reduces inflammation and bone erosions in mice with collageninduced arthritis and directly targets the JAK/STAT pathway in rheumatoid arthritis fibroblast-like synoviocytes. *Arthritis Rheumatol*. 2020;72(12):2030-2039.
- 107. Carraro G, El-Hashash A, Guidolin D, et al. miR-17 family of microRNAs controls FGF10-mediated embryonic lung epithelial branching morphogenesis through MAPK14 and STAT3 regulation of E-Cadherin distribution. *Dev Biol.* 2009;333(2):238-250.
- 108. Zhang H, Cai K, Wang J, et al. MiR-7, inhibited indirectly by lincRNA HOTAIR, directly inhibits SETDB1 and reverses the EMT of breast cancer stem cells by downregulating the STAT3 pathway. *Stem Cells*. 2014;32(11):2858-2868.
- 109. Xiao D, Cui X, Wang X. LncRNA PTCSC3 inhibits cell proliferation in laryngeal squamous cell carcinoma by down-regulating lncRNA HOTAIR. *Biosci Rep.* 2019;39(6): BSR20182362.
- 110. Jin S, Yang X, Li J, Yang W, Ma H, Zhang Z. p53targeted lincRNA-p21 acts as a tumor suppressor by inhibiting JAK2/STAT3 signaling pathways in head and neck squamous cell carcinoma. *Mol Cancer*. 2019;18(1):38.
- Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell*. 2000;100(1):57-70.
- 112. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646-674.
- Hanahan D. Hallmarks of cancer: new dimensions. *Cancer Discov*. 2022;12(1):31-46.
- 114. Pines J. Cyclins, CDKs and cancer. Semin Cancer Biol. 1995;6(2):63-72.
- Mittnacht S, Boshoff C. Viral cyclins. *Rev Med Virol*. 2000;10(3):175-184.
- Deshpande A, Sicinski P, Hinds PW. Cyclins and cdks in development and cancer: a perspective. *Oncogene*. 2005;24(17):2909-2915.

- 117. Deepak Roshan VG, Sinto MS, Thomas S, Kannan S. Cyclin D1 overexpression associated with activation of STAT3 in oral carcinoma patients from South India. *J Cancer Res Ther.* 2018;14(2):403-408.
- 118. Luo J, Yan R, He X, He J. Constitutive activation of STAT3 and cyclin D1 overexpression contribute to proliferation, migration and invasion in gastric cancer cells. *Am J Transl Res.* 2017;9(12):5671.
- 119. Yang JG, Lu R, Ye XJ, Zhang J, Tan YQ, Zhou G. Icaritin reduces oral squamous cell carcinoma progression via the inhibition of STAT3 signaling. *Int J Mol Sci.* 2017;18(1):132.
- Adams JM, Cory S. The Bcl-2 apoptotic switch in cancer development and therapy. Oncogene. 2007;26(9):1324-1337.
- Warren CFA, Wong-Brown MW, Bowden NA. BCL-2 family isoforms in apoptosis and cancer. *Cell Death Dis.* 2019;10(3): 177.
- 122. Bessou M, Lopez J, Gadet R, et al. The apoptosis inhibitor BclxL controls breast cancer cell migration through mitochondriadependent reactive oxygen species production. *Oncogene*. 2020;39(15):3056-3074.
- 123. Lee EF, Harris TJ, Tran S, et al. BCL-XL and MCL-1 are the key BCL-2 family proteins in melanoma cell survival. *Cell Death Dis.* 2019;10(5):1-14.
- 124. Jiao Y, Ding H, Huang S, et al. Bcl-XL and Mcl-1 upregulation by calreticulin promotes apoptosis resistance of fibroblastlike synoviocytes via activation of PI3K/Akt and STAT3 pathways in rheumatoid arthritis. *Clin Exp Rheumatol.* 2018;36(5): 841-849.
- 125. Catlett-Falcone R, Landowski TH, Oshiro MM, et al. Constitutive activation of Stat3 signaling confers resistance to apoptosis in human U266 myeloma cells. *Immunity*. 1999;10(1):105-115.
- 126. Oh HN, Oh KB, Lee MH, et al. JAK2 regulation by licochalcone H inhibits the cell growth and induces apoptosis in oral squamous cell carcinoma. *Phytomedicine*. 2019;52:60-69.
- 127. Yasuda T, Koiwa M, Yonemura A, et al. Inflammationdriven senescence-associated secretory phenotype in cancerassociated fibroblasts enhances peritoneal dissemination. *Cell Rep.* 2021;34(8):108779.
- 128. Zhao H, Wu L, Yan G, et al. Inflammation and tumor progression: signaling pathways and targeted intervention. *Signal Transduc Target Ther*. 2021;6(1):263.
- 129. Powell DR, Huttenlocher A. Neutrophils in the tumor microenvironment. *Trends Immunol.* 2016;37(1):41-52.
- Raposo TP, Beirão BC, Pang LY, Queiroga FL, Argyle DJ. Inflammation and cancer: till death tears them apart. *Vet J*. 2015;205(2):161-174.
- 131. Huang L, Guo Y, Liu S, et al. Targeting regulatory T cells for immunotherapy in melanoma. *Mol Biomed*. 2021;2(1):11.
- 132. Sun C, Mezzadra R, Schumacher TN. Regulation and function of the PD-L1 checkpoint. *Immunity*. 2018;48(3):434-452.
- 133. Bu L, Yu G, Wu L, et al. STAT3 induces immunosuppression by upregulating PD-1/PD-L1 in HNSCC. J Dent Res. 2017;96(9):1027-1034.
- 134. Xu L, Chen X, Shen M, et al. Inhibition of IL-6-JAK/Stat3 signaling in castration-resistant prostate cancer cells enhances the NK cell-mediated cytotoxicity via alteration of PD-L1/NKG2D ligand levels. *Mol Oncol.* 2018;12(3):269-286.
- 135. Cheng Y, Li H, Deng Y, et al. Cancer-associated fibroblasts induce PDL1+ neutrophils through the IL6-STAT3 pathway

that foster immune suppression in hepatocellular carcinoma. *Cell Death Dis.* 2018;9(4):1-11.

- 136. Sun R, Xiong Y, Liu H, et al. Tumor-associated neutrophils suppress antitumor immunity of NK cells through the PD-L1/PD-1 axis. *Transl Oncol.* 2020;13(10):100825.
- 137. Italiani P, Boraschi D. From monocytes to M1/M2 macrophages: phenotypical vs. functional differentiation. *Front Immunol.* 2014;5:514.
- 138. Fu X-L, Duan W, Su C-Y, et al. Interleukin 6 induces M2 macrophage differentiation by STAT3 activation that correlates with gastric cancer progression. *Cancer Immunol Immunother*. 2017;66(12):1597-1608.
- 139. Yin Z, Ma T, Lin Y, et al. IL-6/STAT3 pathway intermediates M1/M2 macrophage polarization during the development of hepatocellular carcinoma. *J Cell Biochem.* 2018;119(11):9419-9432.
- 140. Di Conza G, Tsai CH, Gallart-Ayala H, et al. Tumor-induced reshuffling of lipid composition on the endoplasmic reticulum membrane sustains macrophage survival and pro-tumorigenic activity. *Nat Immunol.* 2021;22(11):1403-1415.
- 141. Zhang X, Zeng Y, Qu Q, et al. PD-L1 induced by IFN-γ from tumor-associated macrophages via the JAK/STAT3 and PI3K/AKT signaling pathways promoted progression of lung cancer. *Int J Clin Oncol.* 2017;22(6):1026-1033.
- 142. Vasquez-Dunddel D, Pan F, Zeng Q, et al. STAT3 regulates arginase-I in myeloid-derived suppressor cells from cancer patients. *J Clin Invest*. 2013;123(4):1580-1589.
- 143. Wu AA, Drake V, Huang HS, Chiu S, Zheng L. Reprogramming the tumor microenvironment: tumor-induced immunosuppressive factors paralyze T cells. *Oncoimmunol*ogy. 2015;4(7):e1016700.
- 144. Li K, Shi H, Zhang B, et al. Myeloid-derived suppressor cells as immunosuppressive regulators and therapeutic targets in cancer. *Signal Transduc Target Ther*. 2021;6(1):362.
- 145. Waldner MJ, Neurath MF. Master regulator of intestinal disease: IL-6 in chronic inflammation and cancer development. *Semin Immunol.* 2014;26(1):75-79.
- 146. Kortylewski M, Kujawski M, Wang T, et al. Inhibiting Stat3 signaling in the hematopoietic system elicits multicomponent antitumor immunity. *Nat Med.* 2005;11(12):1314-1321.
- 147. Jewett A, Head C, Cacalano NA. Emerging mechanisms of immunosuppression in oral cancers. *J Dent Res.* 2006;85(12):1061-1073.
- 148. Priceman Saul J, Shen S, Wang L, et al. S1PR1 is crucial for accumulation of regulatory T cells in tumors via STAT3. *Cell Rep.* 2014;6(6):992-999.
- 149. Hanna BS, Llaó-Cid L, Iskar M, et al. Interleukin-10 receptor signaling promotes the maintenance of a PD-1int TCF-1+ CD8+ T cell population that sustains anti-tumor immunity. *Immunity*. 2021;54(12):2825-2841.e10.
- 150. Gupta MK, Qin RY. Mechanism and its regulation of tumorinduced angiogenesis. *World J Gastroenterol*. 2003;9(6):1144-1155.
- Mathonnet M, Perraud A, Christou N, et al. Hallmarks in colorectal cancer: angiogenesis and cancer stem-like cells. *World J Gastroenterol.* 2014;20(15):4189-4196.
- 152. Weis SM, Cheresh DA. Tumor angiogenesis: molecular pathways and therapeutic targets. *Nat Med.* 2011;17(11):1359-1370.

- Niu G, Wright KL, Huang M, et al. Constitutive Stat3 activity up-regulates VEGF expression and tumor angiogenesis. *Onco*gene. 2002;21(13):2000-2008.
- 154. Tamura R, Tanaka T, Akasaki Y, Murayama Y, Yoshida K, Sasaki H. The role of vascular endothelial growth factor in the hypoxic and immunosuppressive tumor microenvironment: perspectives for therapeutic implications. *Med Oncol.* 2019;37(1):2.
- 155. Tong F, Xiong CJ, Wei CH, et al. Hypo-fractionation radiotherapy normalizes tumor vasculature in non-small cell lung cancer xenografts through the p-STAT3/HIF-1 alpha signaling pathway. *Ther Adv Med Oncol.* 2020;12:1758835920965853.
- 156. Zhang P-C, Liu X, Li M-M, et al. AT-533, a novel Hsp90 inhibitor, inhibits breast cancer growth and HIF-1α/VEGF/VEGFR-2-mediated angiogenesis in vitro and in vivo. *Biochem Pharmacol.* 2020;172:113771.
- 157. Liu P, Atkinson SJ, Akbareian SE, et al. Sulforaphane exerts anti-angiogenesis effects against hepatocellular carcinoma through inhibition of STAT3/HIF-1α/VEGF signalling. *Sci Rep.* 2017;7(1):12651.
- 158. Zhang N, Zhang M, Wang Z, Gao W, Sun Z-G. Activated STAT3 could reduce survival in patients with esophageal squamous cell carcinoma by up-regulating VEGF and cyclin D1 expression. *J Cancer*. 2020;11(7):1859-1868.
- 159. Quintero-Fabián S, Arreola R, Becerril-Villanueva E, et al. Role of matrix metalloproteinases in angiogenesis and cancer. *Front Oncol.* 2019;9:1370.
- 160. Jodele S, Chantrain CF, Blavier L, et al. The contribution of bone marrow-derived cells to the tumor vasculature in neuroblastoma is matrix metalloproteinase-9 dependent. *Cancer Res.* 2005;65(8):3200-3208.
- Roy R, Dagher A, Butterfield C, Moses MA. ADAM12 Is a novel regulator of tumor angiogenesis via STAT3 signaling. *Mol Cancer Res.* 2017;15(11):1608-1622.
- 162. Kujawski M, Kortylewski M, Lee H, Herrmann A, Kay H, Yu H. Stat3 mediates myeloid cell-dependent tumor angiogenesis in mice. *J Clin Invest.* 2008;118(10):3367-3377.
- 163. Xu Q, Ma H, Chang H, Feng Z, Zhang C, Yang X. The interaction of interleukin-8 and PTEN inactivation promotes the malignant progression of head and neck squamous cell carcinoma via the STAT3 pathway. *Cell Death Dis.* 2020;11(5): 405.
- 164. Ko HS, Park BJ, Choi SK, et al. STAT3 and ERK signaling pathways are implicated in the invasion activity by oncostatin M through induction of matrix metalloproteinases 2 and 9. *Yonsei Med J.* 2016;57(3):761-768.
- 165. Park SY, Jang WJ, Yi EY, et al. Melatonin suppresses tumor angiogenesis by inhibiting HIF-1alpha stabilization under hypoxia. *J Pineal Res.* 2010;48(2):178-184.
- 166. Wu F, Yang J, Liu J, et al. Signaling pathways in cancerassociated fibroblasts and targeted therapy for cancer. *Signal Transduc Target Ther*. 2021;6(1):218.
- 167. Smith A, Teknos TN, Pan Q. Epithelial to mesenchymal transition in head and neck squamous cell carcinoma. Oral Oncol. 2013;49(4):287-292.
- Pastushenko I, Blanpain C. EMT transition states during tumor progression and metastasis. *Trends Cell Biol.* 2019;29(3):212-226.

- Chao J, Zhao S, Sun H. Dedifferentiation of hepatocellular carcinoma: molecular mechanisms and therapeutic implications. *Am J Transl Res.* 2020;12(5):2099-2109.
- 170. Taylor KM, Hiscox S, Nicholson RI. Zinc transporter LIV-1: a link between cellular development and cancer progression. *Trends Endocrinol Metab.* 2004;15(10):461-463.
- 171. Zhang W, Zhang J, Zhang Z, et al. Overexpression of indoleamine 2,3-dioxygenase 1 promotes epithelialmesenchymal transition by activation of the IL-6/STAT3/PD-L1 pathway in bladder cancer. *Transl Oncol.* 2019;12(3):485-492.
- 172. Sun SS, Zhou X, Huang YY, et al. Targeting STAT3/miR-21 axis inhibits epithelial-mesenchymal transition via regulating CDK5 in head and neck squamous cell carcinoma. *Mol Cancer*. 2015;14:213.
- 173. Zhao M, Hu X, Xu Y, et al. Targeting of EZH2 inhibits epithelial-mesenchymal transition in head and neck squamous cell carcinoma via regulating the STAT3/VEGFR2 axis. *Int J Oncol.* 2019;55(5):1165-1175.
- 174. Yadav A, Kumar B, Datta J, Teknos TN, Kumar P. IL-6 promotes head and neck tumor metastasis by inducing epithelialmesenchymal transition via the JAK-STAT3-SNAIL signaling pathway. *Mol Cancer Res.* 2011;9(12):1658-1667.
- 175. Ligorio M, Sil S, Malagon-Lopez J, et al. Stromal microenvironment shapes the intratumoral architecture of pancreatic cancer. *Cell*. 2019;178(1):160-175.e27.
- 176. Chen YW, Chen KH, Huang PI, et al. Cucurbitacin I suppressed stem-like property and enhanced radiation-induced apoptosis in head and neck squamous carcinoma-derived CD44(+)ALDH1(+) cells. *Mol Cancer Ther.* 2010;9(11):2879-2892.
- 177. Wu TF, Chen L, Bu LL, Gao J, Zhang WF, Jia J. CD44+ cancer cell-induced metastasis: a feasible neck metastasis model. *Eur J Pharm Sci.* 2017;101:243-250.
- 178. Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. *Nature*. 2001;414(6859):105-111.
- 179. Huang WC, Jang TH, Tung SL, Yen TC, Chan SH, Wang LH. A novel miR-365-3p/EHF/keratin 16 axis promotes oral squamous cell carcinoma metastasis, cancer stemness and drug resistance via enhancing beta5-integrin/c-met signaling pathway. J Exp Clin Cancer Res. 2019;38(1):89.
- 180. Kim S-Y, Kang JW, Song X, et al. Role of the IL-6-JAK1-STAT3-Oct-4 pathway in the conversion of non-stem cancer cells into cancer stem-like cells. *Cell Signalling*. 2013;25(4):961-969.
- 181. Yang J, Liao D, Chen C, et al. Tumor-associated macrophages regulate murine breast cancer stem cells through a novel paracrine EGFR/Stat3/Sox-2 signaling pathway. *Stem Cells*. 2013;31(2):248-258.
- 182. Ning Y, Cui Y, Li X, et al. Co-culture of ovarian cancer stemlike cells with macrophages induced SKOV3 cells stemness via IL-8/STAT3 signaling. *Biomed Pharmacother*. 2018;103:262-271.
- 183. Tanabe S, Quader S, Cabral H, Ono R. Interplay of EMT and CSC in cancer and the potential therapeutic strategies. *Front Pharmacol.* 2020;11:904.
- Dave B, Mittal V, Tan NM, Chang JC. Epithelial-mesenchymal transition, cancer stem cells and treatment resistance. *Breast Cancer Res.* 2012;14(1):1-5.
- 185. Zhou P, Li B, Liu F, et al. The epithelial to mesenchymal transition (EMT) and cancer stem cells: implication for treatment resistance in pancreatic cancer. *Mol Cancer*. 2017;16(1):1-11.

- 186. Smigiel JM, Parameswaran N, Jackson MW. Potent EMT and CSC phenotypes are induced by oncostatin-M in pancreatic cancer. *Mol Cancer Res.* 2017;15(4):478-488.
- 187. Jin W. Role of JAK/STAT3 signaling in the regulation of metastasis, the transition of cancer stem cells, and chemoresistance of cancer by epithelial-mesenchymal transition. *Cells*. 2020;9(1):217.
- 188. Junk DJ, Bryson BL, Smigiel JM, Parameswaran N, Bartel CA, Jackson MW. Oncostatin M promotes cancer cell plasticity through cooperative STAT3-SMAD3 signaling. *Oncogene*. 2017;36(28):4001-4013.
- 189. Zhao C, Li H, Lin HJ, Yang S, Lin J, Liang G. Feedback activation of STAT3 as a cancer drug-resistance mechanism. *Trends Pharmacol Sci.* 2016;37(1):47-61.
- 190. Liu WH, Chen MT, Wang ML, et al. Cisplatin-selected resistance is associated with increased motility and stemlike properties via activation of STAT3/Snail axis in atypical teratoid/rhabdoid tumor cells. *Oncotarget.* 2015;6(3):1750-1768.
- 191. Zhang JL, Liu XZ, Wang PY, et al. Targeting HCCR expression resensitizes gastric cancer cells to chemotherapy via downregulating the activation of STAT3. *Sci Rep.* 2016;6:24196.
- 192. Huang W, Zhong Z, Luo C, et al. The miR-26a/AP-2α/Nanog signaling axis mediates stem cell self-renewal and temozolomide resistance in glioma. *Theranostics*. 2019;9(19):5497-5516.
- 193. Su JC, Tseng PH, Wu SH, et al. SC-2001 overcomes STAT3mediated sorafenib resistance through RFX-1/SHP-1 activation in hepatocellular carcinoma. *Neoplasia*. 2014;16(7):595-605.
- 194. Liberti MV, Locasale JW. The Warburg effect: how does it benefit cancer cells? *Trends Biochem Sci.* 2016;41(3):211-218.
- 195. DeBerardinis RJ, Chandel NS. We need to talk about the Warburg effect. *Nat Metab.* 2020;2(2):127-129.
- 196. Zhu S, Dong Z, Ke X et-al. The roles of sirtuins family in cell metabolism during tumor development. *Semin Cancer Biol.*. 2019;57:59-71.
- 197. Bi YH, Han WQ, Li RF, et al. Signal transducer and activator of transcription 3 promotes the Warburg effect possibly by inducing pyruvate kinase M2 phosphorylation in liver precancerous lesions. *World J Gastroenterol*. 2019;25(16):1936-1949.
- 198. Li Q, Zhang D, Chen X, et al. Nuclear PKM2 contributes to gefitinib resistance via upregulation of STAT3 activation in colorectal cancer. *Sci Rep.* 2015;5:16082.
- 199. Guan M, Tong Y, Guan M, et al. Lapatinib inhibits breast cancer cell proliferation by influencing PKM2 expression. *Technol Cancer Res Treat*. 2018;17:1533034617749418.
- Demaria M, Poli V. PKM2, STAT3 and HIF-1α: the Warburg's vicious circle. Jak-stat. 2012;1(3):194-196.
- 201. Fan M, Sun W, Gu X, et al. The critical role of STAT3 in biogenesis of tumor-derived exosomes with potency of inducing cancer cachexia in vitro and in vivo. *Oncogene*. 2022;41:1050-1062.
- 202. Pu Z, Xu M, Yuan X, Xie H, Zhao J. Circular RNA circ-CUL3 accelerates the Warburg effect progression of gastric cancer through regulating the STAT3/HK2 axis. *Mol Ther Nucleic Acids*. 2020;22:310-318.
- 203. Yu GT, Mao L, Wu L, et al. Inhibition of SRC family kinases facilitates anti-CTLA4 immunotherapy in head and neck squamous cell carcinoma. *Cell Mol Life Sci.* 2018;75(22):4223-4234.
- 204. Bauman JE, Duvvuri U, Gooding WE, et al. Randomized, placebo-controlled window trial of EGFR, Src, or

combined blockade in head and neck cancer. JCI Insight. 2017;2(6):e90449.

- 205. Kim DW, Saussele S, Williams LA, et al. Outcomes of switching to dasatinib after imatinib-related low-grade adverse events in patients with chronic myeloid leukemia in chronic phase: the DASPERSE study. *Ann Hematol.* 2018;97(8):1357-1367.
- 206. Finn RS, Bengala C, Ibrahim N, et al. Dasatinib as a single agent in triple-negative breast cancer: results of an open-label phase 2 study. *Clin Cancer Res.* 2011;17(21):6905-6913.
- 207. Foà R, Bassan R, Vitale A, et al. Dasatinib-Blinatumomab for Ph-positive acute lymphoblastic leukemia in adults. N Engl J Med. 2020;383(17):1613-1623.
- 208. Yu EY, Wilding G, Posadas E, et al. Phase II study of dasatinib in patients with metastatic castration-resistant prostate cancer. *Clin Cancer Res.* 2009;15(23):7421-7428.
- 209. Bonner JA, Yang ES, Trummell HQ, Nowsheen S, Willey CD, Raisch KP. Inhibition of STAT-3 results in greater cetuximab sensitivity in head and neck squamous cell carcinoma. *Radiother Oncol.* 2011;99(3):339-343.
- 210. Park SH, Jo MJ, Kim BR, et al. Sonic hedgehog pathway activation is associated with cetuximab resistance and EPHB3 receptor induction in colorectal cancer. *Theranostics*. 2019;9(8):2235-2251.
- 211. Kopetz S, Grothey A, Yaeger R, et al. Encorafenib, binimetinib, and cetuximab in BRAF V600E-mutated colorectal cancer. N Engl J Med. 2019;381(17):1632-1643.
- 212. Kopetz S, Guthrie KA, Morris VK, et al. Randomized trial of irinotecan and cetuximab with or without vemurafenib in BRAF-mutant metastatic colorectal cancer (SWOG S1406). J Clin Oncol. 2021;39(4):285-294.
- 213. Burtness B, Harrington KJ, Greil R, et al. Pembrolizumab alone or with chemotherapy versus cetuximab with chemotherapy for recurrent or metastatic squamous cell carcinoma of the head and neck (KEYNOTE-048): a randomised, open-label, phase 3 study. *Lancet*. 2019;394(10212):1915-1928.
- 214. Sen M, Pollock NI, Black J, et al. JAK kinase inhibition abrogates STAT3 activation and head and neck squamous cell carcinoma tumor growth. *Neoplasia*. 2015;17(3):256-264.
- 215. Wang SW, Hu J, Guo QH, et al. AZD1480, a JAK inhibitor, inhibits cell growth and survival of colorectal cancer via modulating the JAK2/STAT3 signaling pathway. *Oncol Rep.* 2014;32(5):1991-1998.
- 216. Stover DG, Gil Del Alcazar CR, Brock J, et al. Phase II study of ruxolitinib, a selective JAK1/2 inhibitor, in patients with metastatic triple-negative breast cancer. *NPJ Breast Cancer*. 2018;4:10.
- 217. Park JS, Hong MH, Chun YJ, Kim HR, Cho BC. A phase Ib study of the combination of afatinib and ruxolitinib in EGFR mutant NSCLC with progression on EGFR-TKIs. *Lung Cancer*. 2019;134:46-51.
- 218. Hurwitz HI, Uppal N, Wagner SA, et al. Randomized, doubleblind, phase II study of ruxolitinib or placebo in combination with capecitabine in patients with metastatic pancreatic cancer for whom therapy with gemcitabine has failed. *J Clin Oncol.* 2015;33(34):4039-4047.
- 219. Bilori B, Thota S, Clemente MJ, et al. Tofacitinib as a novel salvage therapy for refractory T-cell large granular lymphocytic leukemia. *Leukemia*. 2015;29(12):2427-2429.

- 220. Pouyfung P, Choonate S, Wongnoppavich A, Rongnoparut P, Chairatvit K. Anti-proliferative effect of 8α-tigloyloxyhirsutinolide-13-O-acetate (8αTGH) isolated from Vernonia cinerea on oral squamous cell carcinoma through inhibition of STAT3 and STAT2 phosphorylation. *Phytomedicine*. 2019;52:238-246.
- 221. Kumar B, Yadav A, Hideg K, Kuppusamy P, Teknos TN, Kumar P. A novel curcumin analog (H-4073) enhances the therapeutic efficacy of cisplatin treatment in head and neck cancer. *PLoS One.* 2014;9(3):e93208.
- 222. Mehta HJ, Patel V, Sadikot RT. Curcumin and lung cancer–a review. *Target Oncol.* 2014;9(4):295-310.
- 223. Zhou X, Ren Y, Liu A, et al. WP1066 sensitizes oral squamous cell carcinoma cells to cisplatin by targeting STAT3/miR-21 axis. *Sci Rep.* 2014;4:7461.
- 224. Judd LM, Menheniott TR, Ling H, et al. Inhibition of the JAK2/STAT3 pathway reduces gastric cancer growth in vitro and in vivo. *PLoS One*. 2014;9(5):e95993.
- 225. Sau S, Mondal SK, Kashaw SK, Iyer AK, Banerjee R. Combination of cationic dexamethasone derivative and STAT3 inhibitor (WP1066) for aggressive melanoma: a strategy for repurposing a phase I clinical trial drug. *Mol Cell Biochem*. 2017;436(1-2):119-136.
- 226. Matsuoka Y, Nakayama H, Yoshida R, et al. IL-6 controls resistance to radiation by suppressing oxidative stress via the Nrf2antioxidant pathway in oral squamous cell carcinoma. *Br J Cancer*. 2016;115(10):1234-1244.
- 227. Alraouji NN, Al-Mohanna FH, Ghebeh H, et al. Tocilizumab potentiates cisplatin cytotoxicity and targets cancer stem cells in triple-negative breast cancer. *Mol Carcinog.* 2020;59(9):1041-1051.
- Liu FT, Jia L, Wang P, et al. CD126 and targeted therapy with tocilizumab in chronic lymphocytic leukemia. *Clin Cancer Res.* 2016;22(10):2462-2469.
- 229. Karkera J, Steiner H, Li W, et al. The anti-interleukin-6 antibody siltuximab down-regulates genes implicated in tumorigenesis in prostate cancer patients from a phase I study. *Prostate*. 2011;71(13):1455-1465.
- 230. Song L, Rawal B, Nemeth JA, Haura EB. JAK1 activates STAT3 activity in non-small-cell lung cancer cells and IL-6 neutralizing antibodies can suppress JAK1-STAT3 signaling. *Mol Cancer Ther.* 2011;10(3):481-494.
- 231. Wu X, Cao Y, Xiao H, Li C, Lin J. Bazedoxifene as a novel GP130 inhibitor for pancreatic cancer therapy. *Mol Cancer Ther.* 2016;15(11):2609-2619.
- 232. Curry J, Johnson J, Tassone P, et al. Metformin effects on head and neck squamous carcinoma microenvironment: window of opportunity trial. *Laryngoscope*. 2017;127(8):1808-1815.
- 233. Brown JR, Chan DK, Shank JJ, et al. Phase II clinical trial of metformin as a cancer stem cell-targeting agent in ovarian cancer. *JCI Insight*. 2020;5(11):e133247.
- 234. Siddiquee KA, Gunning PT, Glenn M, et al. An oxazole-based small-molecule Stat3 inhibitor modulates Stat3 stability and processing and induces antitumor cell effects. ACS Chem Biol. 2007;2(12):787-798.
- 235. Bu LL, Deng WW, Huang CF, Liu B, Zhang WF, Sun ZJ. Inhibition of STAT3 reduces proliferation and invasion in salivary gland adenoid cystic carcinoma. *Am J Cancer Res.* 2015;5(5):1751-1761.

- 236. Bu LL, Yu GT, Deng WW, et al. Targeting STAT3 signaling reduces immunosuppressive myeloid cells in head and neck squamous cell carcinoma. *Oncoimmunology*. 2016;5(5):e1130206.
- 237. Bu LL, Li YC, Yu GT, et al. Targeting phosphorylation of STAT3 delays tumor growth in HPV-negative anal squamous cell carcinoma mouse model. *Sci Rep.* 2017;7(1):6629.
- 238. Song H, Wang R, Wang S, Lin J. A low-molecular-weight compound discovered through virtual database screening inhibits Stat3 function in breast cancer cells. *Proc Natl Acad Sci U S A*. 2005;102(13):4700-4705.
- 239. Pan Y, Zhou F, Zhang R, Claret FX. Correction: stat3 inhibitor stattic exhibits potent antitumor activity and induces chemoand radio-sensitivity in nasopharyngeal carcinoma. *PLoS One*. 2020;15(8):e0237943.
- 240. Genini D, Brambilla L, Laurini E, et al. Mitochondrial dysfunction induced by a SH2 domain-targeting STAT3 inhibitor leads to metabolic synthetic lethality in cancer cells. *Proc Natl Acad Sci U S A*. 2017;114(25):E4924-E4933.
- 241. Ogura M, Uchida T, Terui Y, et al. Phase I study of OPB-51602, an oral inhibitor of signal transducer and activator of transcription 3, in patients with relapsed/refractory hematological malignancies. *Cancer Sci.* 2015;106(7):896-901.
- 242. Wong AL, Soo RA, Tan DS, et al. Phase I and biomarker study of OPB-51602, a novel signal transducer and activator of transcription (STAT) 3 inhibitor, in patients with refractory solid malignancies. *Ann Oncol.* 2015;26(5):998-1005.
- 243. Yoo C, Kang J, Lim HY, et al. Phase I dose-finding study of OPB-111077, a novel STAT3 inhibitor, in patients with advanced hepatocellular carcinoma. *Cancer Res Treat.* 2019;51(2): 510-518.
- 244. Jung KH, Yoo W, Stevenson HL, et al. Multifunctional effects of a small-molecule STAT3 inhibitor on NASH and hepatocellular carcinoma in mice. *Clin Cancer Res.* 2017;23(18):5537-5546.
- 245. Di JX, Zhang HY. C188-9, a small-molecule STAT3 inhibitor, exerts an antitumor effect on head and neck squamous cell carcinoma. *Anticancer Drugs*. 2019;30(8):846-853.
- 246. Chen J, Bai L, Bernard D, et al. Structure-based design of conformationally constrained, cell-permeable STAT3 inhibitors. ACS Med Chem Lett. 2010;1(2):85-89.
- 247. Leong PL, Andrews GA, Johnson DE, et al. Targeted inhibition of Stat3 with a decoy oligonucleotide abrogates head and neck cancer cell growth. *Proc Natl Acad Sci U S A*. 2003;100(7):4138-4143.
- 248. Carmicheal J, Kaur S, Batra SK, Ganti AK. Hunting for transcription factors: STAT3 decoy in non-small cell lung cancer. *Transl Lung Cancer Res*. 2018;7(3):S254-s257.
- 249. Weerasinghe P, Garcia GE, Zhu Q, et al. Inhibition of Stat3 activation and tumor growth suppression of non-small cell lung cancer by G-quartet oligonucleotides. *Int J Oncol.* 2007;31(1):129-136.
- 250. Jing N, Zhu Q, Yuan P, Li Y, Mao L, Tweardy DJ. Targeting signal transducer and activator of transcription 3 with G-quartet oligonucleotides: a potential novel therapy for head and neck cancer. *Mol Cancer Ther.* 2006;5(2):279-286.
- 251. Huang W, Dong Z, Chen Y, et al. Small-molecule inhibitors targeting the DNA-binding domain of STAT3 suppress tumor growth, metastasis and STAT3 target gene expression in vivo. *Oncogene*. 2016;35(6):783-792.

- 252. Huang W, Dong Z, Wang F, Peng H, Liu J-Y, Zhang J-T. A small molecule compound targeting STAT3 DNA-binding domain inhibits cancer cell proliferation, migration, and invasion. ACS Chem Biol. 2014;9(5):1188-1196.
- 253. Jonker DJ, Nott L, Yoshino T, et al. Napabucasin versus placebo in refractory advanced colorectal cancer: a randomised phase 3 trial. *Lancet Gastroenterol Hepatol.* 2018;3(4):263-270.
- 254. Shitara K, Yodo Y, Iino S. A phase I study of napabucasin plus paclitaxel for Japanese patients with advanced/recurrent gastric cancer. *In Vivo*. 2019;33(3):933-937.
- 255. Han D, Yu T, Dong N, Wang B, Sun F, Jiang D. Napabucasin, a novel STAT3 inhibitor suppresses proliferation, invasion and stemness of glioblastoma cells. *J Exp Clin Cancer Res.* 2019;38(1):289.
- 256. Son DJ, Zheng J, Jung YY, et al. MMPP attenuates non-small cell lung cancer growth by inhibiting the STAT3 DNA-Binding activity via direct binding to the STAT3 DNA-binding domain. *Theranostics*. 2017;7(18):4632-4642.
- 257. Zheng J, Son DJ, Lee HL, et al. E)-2-methoxy-4-(3-(4-methoxyphenyl)prop-1-en-1-yl)phenol suppresses ovarian cancer cell growth via inhibition of ERK and STAT3. *Mol Carcinog.* 2017;56(9):2003-2013.
- 258. Hong D, Kurzrock R, Kim Y, et al. AZD9150, a next-generation antisense oligonucleotide inhibitor of STAT3 with early evidence of clinical activity in lymphoma and lung cancer. *Sci Transl Med.* 2015;7(314):314ra185.
- 259. Sun CY, Nie J, Huang JP, Zheng GJ, Feng B. Targeting STAT3 inhibition to reverse cisplatin resistance. *Biomed Pharmacother*. 2019;117:109135.
- 260. Xu S, Zhao N, Hui L, Song M, Miao ZW, Jiang XJ. MicroRNA-124-3p inhibits the growth and metastasis of nasopharyngeal carcinoma cells by targeting STAT3. *Oncol Rep.* 2016;35(3):1385-1394.
- 261. Bai L, Zhou H, Xu R, et al. A potent and selective smallmolecule degrader of STAT3 achieves complete tumor regression in vivo. *Cancer Cell*. 2019;36(5):498-511 e17.
- 262. Gaykalova DA, Manola JB, Ozawa H, et al. NF-kappaB and stat3 transcription factor signatures differentiate HPV-positive and HPV-negative head and neck squamous cell carcinoma. *Int J Cancer*. 2015;137(8):1879-1889.
- 263. Cordes F, Foell D, Ding JN, Varga G, Bettenworth D. Differential regulation of JAK/STAT-signaling in patients with ulcerative colitis and Crohn's disease. *World J Gastroenterol.* 2020;26(28):4055-4075.
- 264. Luo W, Li YX, Jiang LJ, Chen Q, Wang T, Ye DW. Targeting JAK-STAT signaling to control cytokine release syndrome in COVID-19. *Trends Pharmacol Sci.* 2020;41(8):531-543.
- 265. Huynh J, Etemadi N, Hollande F, Ernst M, Buchert M. The JAK/STAT3 axis: a comprehensive drug target for solid malignancies. *Semin Cancer Biol.* 2017;45:13-22.
- 266. Pinto N, Prokopec SD, Vizeacoumar F, et al. Lestaurtinib is a potent inhibitor of anaplastic thyroid cancer cell line models. *PLoS One.* 2018;13(11):e0207152.
- 267. Minturn JE, Evans AE, Villablanca JG, et al. Phase I trial of lestaurtinib for children with refractory neuroblastoma: a new approaches to neuroblastoma therapy consortium study. *Cancer Chemother Pharmacol.* 2011;68(4):1057-1065.
- 268. Knapper S, Burnett AK, Littlewood T, et al. A phase 2 trial of the FLT3 inhibitor lestaurtinib (CEP701) as first-line treat-

ment for older patients with acute myeloid leukemia not considered fit for intensive chemotherapy. *Blood.* 2006;108(10): 3262-3270.

- 269. Knapper S, Russell N, Gilkes A, et al. A randomized assessment of adding the kinase inhibitor lestaurtinib to first-line chemotherapy for FLT3-mutated AML. *Blood.* 2017;129(9):1143-1154.
- Plimack ER, Lorusso PM, McCoon P, et al. AZD1480: a phase I study of a novel JAK2 inhibitor in solid tumors. *Oncologist*. 2013;18(7):819-820.
- 271. Tsujita Y, Horiguchi A, Tasaki S, et al. STAT3 inhibition by WP1066 suppresses the growth and invasiveness of bladder cancer cells. *Oncol Rep.* 2017;38(4):2197-2204.
- 272. Ferrajoli A, Faderl S, Van Q, et al. WP1066 disrupts Janus kinase-2 and induces caspase-dependent apoptosis in acute myelogenous leukemia cells. *Cancer Res.* 2007;67(23):11291-11299.
- 273. Sau S, Mondal SK, Kashaw SK, Iyer AK, Banerjee R. Combination of cationic dexamethasone derivative and STAT3 inhibitor (WP1066) for aggressive melanoma: a strategy for repurposing a phase I clinical trial drug. *Mol Cell Biochem*. 2017;436(1):119-136.
- 274. Liu JF, Deng WW, Chen L, et al. Inhibition of JAK2/STAT3 reduces tumor-induced angiogenesis and myeloid-derived suppressor cells in head and neck cancer. *Mol Carcinog.* 2018;57(3):429-439.
- 275. Dao KT, Gotlib J, Deininger MMN, et al. Efficacy of ruxolitinib in patients with chronic neutrophilic leukemia and atypical chronic myeloid leukemia. *J Clin Oncol.* 2020;38(10):1006-1018.
- 276. Mohan CD, Rangappa S, Preetham HD, et al. Targeting STAT3 signaling pathway in cancer by agents derived from Mother Nature. *Semin Cancer Biol*. Published online April 20, 2020. doi: 10.1016/j.semcancer.2020.03.016
- 277. Chen S, Bai Y, Li Z, et al. A betulinic acid derivative SH479 inhibits collagen-induced arthritis by modulating T cell differentiation and cytokine balance. *Biochem Pharmacol.* 2017;126:69-78.
- 278. Lee JH, Kim C, Lee J, Um JY, Sethi G, Ahn KS. Arctiin is a pharmacological inhibitor of STAT3 phosphorylation at tyrosine 705 residue and potentiates bortezomib-induced apoptotic and anti-angiogenic effects in human multiple myeloma cells. *Phytomedicine*. 2019;55:282-292.
- 279. Rincon M. Interleukin-6: from an inflammatory marker to a target for inflammatory diseases. *Trends Immunol*. 2012;33(11):571-577.
- Stanam A, Love-Homan L, Joseph TS, Espinosa-Cotton M, Simons AL. Upregulated interleukin-6 expression contributes to erlotinib resistance in head and neck squamous cell carcinoma. *Mol Oncol.* 2015;9(7):1371-1383.
- 281. Nguyen MLT, Bui KC, Scholta T, et al. Targeting interleukin 6 signaling by monoclonal antibody siltuximab on cholangiocarcinoma. *J Gastroenterol Hepatol*. 2021;36(5):1334-1345.
- 282. Song L, Smith MA, Doshi P, et al. Antitumor efficacy of the anti-interleukin-6 (IL-6) antibody siltuximab in mouse xenograft models of lung cancer. *J Thorac Oncol.* 2014;9(7):974-982.
- 283. Angevin E, Tabernero J, Elez E, et al. A phase I/II, multipledose, dose-escalation study of siltuximab, an anti-interleukin-6

monoclonal antibody, in patients with advanced solid tumors. *Clin Cancer Res.* 2014;20(8):2192-2204.

- 284. Coward J, Kulbe H, Chakravarty P, et al. Interleukin-6 as a therapeutic target in human ovarian cancer. *Clin Cancer Res.* 2011;17(18):6083-6096.
- 285. Fizazi K, De Bono JS, Flechon A, et al. Randomised phase II study of siltuximab (CNTO 328), an anti-IL-6 monoclonal antibody, in combination with mitoxantrone/prednisone versus mitoxantrone/prednisone alone in metastatic castrationresistant prostate cancer. *Eur J Cancer*. 2012;48(1):85-93.
- Kraskouskaya D, Duodu E, Arpin CC, Gunning PT. Progress towards the development of SH2 domain inhibitors. *Chem Soc Rev.* 2013;42(8):3337-3370.
- 287. Morlacchi P, Robertson FM, Klostergaard J, McMurray JS. Targeting SH2 domains in breast cancer. *Future Med Chem*. 2014;6(17):1909-1926.
- 288. Turkson J, Kim JS, Zhang S, et al. Novel peptidomimetic inhibitors of signal transducer and activator of transcription 3 dimerization and biological activity. *Mol Cancer Ther*. 2004;3(3):261-269.
- 289. Mandal PK, Limbrick D, Coleman DR, et al. Conformationally constrained peptidomimetic inhibitors of signal transducer and activator of transcription. 3: evaluation and molecular modeling. J Med Chem. 2009;52(8):2429-2442.
- 290. McMurray JS, Mandal PK, Liao WS, Ren Z, Chen X. Inhibition of Stat3 by cell-permeable peptidomimetic prodrugs targeted to its SH2 domain. *Adv Exp Med Biol*. 2009;611:545-546.
- 291. Qiu HY, Zhu X, Luo YL, et al. Identification of new shikonin derivatives as antitumor agents targeting STAT3 SH2 domain. *Sci Rep.* 2017;7(1):2863.
- 292. Kim BH, Lee H, Song Y, et al. Development of oxadiazole-based ODZ10117 as a small-molecule inhibitor of STAT3 for targeted cancer therapy. *J Clin Med.* 2019;8(11):1847.
- 293. Chen CL, Cen L, Kohout J, et al. Signal transducer and activator of transcription 3 activation is associated with bladder cancer cell growth and survival. *Mol Cancer*. 2008;7:78.
- 294. Miyoshi K, Takaishi M, Nakajima K, et al. Stat3 as a therapeutic target for the treatment of psoriasis: a clinical feasibility study with STA-21, a Stat3 inhibitor. *J Invest Dermatol*. 2011;131(1):108-117.
- 295. Chen H, Bian A, Yang LF, et al. Targeting STAT3 by a small molecule suppresses pancreatic cancer progression. *Oncogene*. 2021;40(8):1440-1457.
- 296. Wang Y, Wang S, Wu Y, et al. Suppression of the growth and invasion of human head and neck squamous cell carcinomas via regulating STAT3 signaling and the miR-21/beta-catenin axis with HJC0152. *Mol Cancer Ther.* 2017;16(4):578-590.
- 297. Yu PY, Gardner HL, Roberts R, et al. Target specificity, in vivo pharmacokinetics, and efficacy of the putative STAT3 inhibitor LY5 in osteosarcoma, Ewing's sarcoma, and rhabdomyosarcoma. *PLoS One.* 2017;12(7):e0181885.
- 298. Bu LL, Zhao ZL, Liu JF, et al. STAT3 blockade enhances the efficacy of conventional chemotherapeutic agents by eradicating head neck stemloid cancer cell. *Oncotarget*. 2015;6(39):41944-41958.
- 299. Furtek SL, Backos DS, Matheson CJ, Reigan P. Strategies and approaches of targeting STAT3 for cancer treatment. ACS Chem Biol. 2016;11(2):308-318.

- 300. Leung CH, Chan DS, Ma VP, Ma DL. DNA-binding small molecules as inhibitors of transcription factors. *Med Res Rev.* 2013;33(4):823-846.
- 301. Turkson J, Zhang S, Palmer J, et al. Inhibition of constitutive signal transducer and activator of transcription 3 activation by novel platinum complexes with potent antitumor activity. *Mol Cancer Ther.* 2004;3(12):1533-1542.
- 302. Cho SH, Park MH, Lee HP, et al. E)-2,4-Bis(p-hydroxyphenyl)-2-butenal enhanced TRAIL-induced apoptosis in ovarian cancer cells through downregulation of NF-κB/STAT3 pathway. *Arch Pharm Res.* 2014;37(5):652-661.
- 303. Xi S, Gooding WE, Grandis JR. In vivo antitumor efficacy of STAT3 blockade using a transcription factor decoy approach: implications for cancer therapy. Oncogene. 2005;24(6):970-979.
- 304. Lau YK, Ramaiyer M, Johnson DE, Grandis JR. Targeting STAT3 in cancer with nucleotide therapeutics. *Cancers (Basel)*. 2019;11(11).
- 305. Shiah JV, Grandis JR, Johnson DE. Targeting STAT3 with proteolysis targeting chimeras and next-generation antisense oligonucleotides. *Mol Cancer Ther*. 2021;20(2):219-228.
- 306. Li WC, Ye SL, Sun RX, et al. Inhibition of growth and metastasis of human hepatocellular carcinoma by antisense oligonucleotide targeting signal transducer and activator of transcription 3. *Clin Cancer Res.* 2006;12(23):7140-7148.
- 307. Oweida AJ, Darragh L, Phan A, et al. STAT3 modulation of regulatory T cells in response to radiation therapy in head and neck cancer. *J Natl Cancer Inst.* 2019;111(12):1339-1349.
- 308. Brown PA, Kairalla JA, Hilden JM, et al. FLT3 inhibitor lestaurtinib plus chemotherapy for newly diagnosed KMT2Arearranged infant acute lymphoblastic leukemia: Children's Oncology Group trial AALL0631. *Leukemia*. 2021;35(5):1279-1290.
- 309. Chan E, Mulkerin D, Rothenberg M, et al. A phase I trial of CEP-701 + gemcitabine in patients with advanced adenocarcinoma of the pancreas. *Invest New Drugs*. 2008;26(3):241-247.
- 310. Argiris A, Duffy AG, Kummar S, et al. Early tumor progression associated with enhanced EGFR signaling with bortezomib, cetuximab, and radiotherapy for head and neck cancer. *Clin Cancer Res.* 2011;17(17):5755-5764.
- 311. Harari PM, Harris J, Kies MS, et al. Postoperative chemoradiotherapy and cetuximab for high-risk squamous cell carcinoma of the head and neck: Radiation Therapy Oncology Group RTOG-0234. J Clin Oncol. 2014;32(23):2486-2495.
- 312. Sen M, Thomas SM, Kim S, et al. First-in-human trial of a STAT3 decoy oligonucleotide in head and neck tumors: implications for cancer therapy. *Cancer Discov*. 2012;2(8):694-705.
- 313. Reilley MJ, McCoon P, Cook C, et al. STAT3 antisense oligonucleotide AZD9150 in a subset of patients with heavily pretreated lymphoma: results of a phase 1b trial. *J Immunother Cancer*. 2018;6(1):119.
- 314. Bharadwaj U, Kasembeli MM, Robinson P, Tweardy DJ. Targeting Janus kinases and signal transducer and activator of transcription 3 to treat inflammation, fibrosis, and cancer: rationale, progress, and caution. *Pharmacol Rev.* 2020;72(2): 486-526.
- Bosch-Barrera J, Queralt B, Menendez JA. Targeting STAT3 with silibinin to improve cancer therapeutics. *Cancer Treat Rev.* 2017;58:61-69.

- Hui Y, Yi X, Hou F, et al. Role of nanoparticle mechanical properties in cancer drug delivery. ACS Nano. 2019;13(7):7410-7424.
- 317. Liu J, Zhang R, Xu ZP. Nanoparticle-based nanomedicines to promote cancer immunotherapy: recent advances and future directions. *Small.* 2019;15(32):1900262.
- 318. Jose A, Labala S, Ninave KM, Gade SK, Venuganti VVK. Effective skin cancer treatment by topical co-delivery of curcumin and STAT3 siRNA using cationic liposomes. *AAPS Pharm-SciTech*. 2018;19(1):166-175.
- 319. Sun Y, Li X, Zhang L, et al. Cell permeable NBD peptidemodified liposomes by hyaluronic acid coating for the synergistic targeted therapy of metastatic inflammatory breast cancer. *Mol Pharmaceutics*. 2019;16(3):1140-1155.
- 320. Zhang J, Zhu S, Tan Q, et al. Combination therapy with ropivacaine-loaded liposomes and nutrient deprivation for simultaneous cancer therapy and cancer pain relief. *Theranostics*. 2020;10(11):4885-4899.
- 321. Mohammadian J, Mahmoudi S, Pourmohammad P, et al. Formulation of Stattic as STAT3 inhibitor in nanostructured lipid carriers (NLCs) enhances efficacy of doxorubicin in melanoma cancer cells. *Naunyn-Schmiedeberg's Arch Pharma*col. 2020;393(12):2315-2323.
- 322. Bu L-L, Wang H-Q, Pan Y, et al. Gelatinase-sensitive nanoparticles loaded with photosensitizer and STAT3 inhibitor for cancer photothermal therapy and immunotherapy. *J Nanobiotechnology*. 2021;19(1):1-13.
- 323. Sivák L, Šubr V, Kovářová J, et al. Polymer-ritonavir derivate nanomedicine with pH-sensitive activation possesses potent anti-tumor activity in vivo via inhibition of proteasome and STAT3 signaling. *J Controlled Release*. 2021;332:563-580.
- 324. Tavares MR, Hrabánková K, Konefał R, et al. HPMAbased copolymers carrying STAT3 inhibitor cucurbitacin-D as stimulus-sensitive nanomedicines for oncotherapy. *Pharmaceutics*. 2021;13(2):179.
- 325. Luo K, Gao Y, Yin S, et al. Co-delivery of paclitaxel and STAT3 siRNA by a multifunctional nanocomplex for targeted treatment of metastatic breast cancer. *Acta Biomater*. 2021;134:649-663.
- 326. Zhuang X, Xiang X, Grizzle W, et al. Treatment of brain inflammatory diseases by delivering exosome encapsulated anti-inflammatory drugs from the nasal region to the brain. *Mol Ther*. 2011;19(10):1769-1779.
- 327. Hosseini M, Baghaei K, Amani D, Ebtekar M. Tumorderived exosomes encapsulating miR-34a promote apoptosis and inhibit migration and tumor progression of colorectal cancer cells under in vitro condition. *DARU J Pharm Sci.* 2021;29(2):267-278.
- 328. Andersen MN, Etzerodt A, Graversen JH, et al. STAT3 inhibition specifically in human monocytes and macrophages by CD163-targeted corosolic acid-containing liposomes. *Cancer Immunol Immunother*. 2019;68(3):489-502.
- 329. Guo C, Chen Y, Gao W, et al. Liposomal nanoparticles carrying anti-IL6R antibody to the tumour microenvironment inhibit metastasis in two molecular subtypes of breast cancer mouse models. *Theranostics*. 2017;7(3):775.
- 330. Shi K, Xue J, Fang Y, et al. Inorganic kernel-reconstituted lipoprotein biomimetic nanovehicles enable efficient targeting "Trojan horse" delivery of STAT3-Decoy oligonucleotide

for overcoming TRAIL resistance. *Theranostics*. 2017;7(18): 4480.

- 331. Zhang S, Gupta S, Fitzgerald TJ. Dual radiosensitization and anti-STAT3 anti-proliferative strategy based on delivery of gold nanoparticle oligonucleotide nanoconstructs to head and neck cancer cells. *Nanotheranostics*. 2018;2(1):1-11.
- 332. Ngamcherdtrakul W, Reda M, Nelson MA, et al. In situ tumor vaccination with nanoparticle co-delivering CpG and STAT3 siRNA to effectively induce whole-body antitumor immune response. *Adv Mater*. 2021:2100628.
- 333. Labala S, Jose A, Chawla SR, et al. Effective melanoma cancer suppression by iontophoretic co-delivery of STAT3 siRNA and imatinib using gold nanoparticles. *Int J Pharm*. 2017;525(2):407-417.
- 334. Budi HS, Izadi S, Timoshin A, et al. Blockade of HIF-1α and STAT3 by hyaluronate-conjugated TAT-chitosan-SPION nanoparticles loaded with siRNA molecules prevents tumor growth. Nanomed Nanotechnol Biol Med. 2021;34:102373.
- 335. Malam Y, Loizidou M, Seifalian AM. Liposomes and nanoparticles: nanosized vehicles for drug delivery in cancer. *Trends Pharmacol Sci.* 2009;30(11):592-599.
- Wu X, Tang W, Marquez RT, et al. Overcoming chemo/radioresistance of pancreatic cancer by inhibiting STAT3 signaling. *Oncotarget.* 2016;7(10):11708-11723.
- 337. Zheng H, Chen Z, Cai A, et al. Nanoparticle mediated codelivery of nifuratel and doxorubicin for synergistic anticancer therapy through STAT3 inhibition. *Colloids Surf B Biointerfaces*. 2020;193:111109.
- 338. Garg SM, Vakili MR, Molavi O, Lavasanifar A. Self-associating poly(ethylene oxide)-block-poly(alpha-carboxyl-epsiloncaprolactone) drug conjugates for the delivery of STAT3

inhibitor JSI-124: potential application in cancer immunotherapy. *Mol Pharm*. 2017;14(8):2570-2584.

- 339. Molavi O, Mahmud A, Hamdy S, et al. Development of a poly(d,l-lactic-co-glycolic acid) nanoparticle formulation of STAT3 inhibitor JSI-124: implication for cancer immunotherapy. *Mol Pharm*. 2010;7(2):364-374.
- 340. Molavi O, Ma Z, Mahmud A, et al. Polymeric micelles for the solubilization and delivery of STAT3 inhibitor cucurbitacins in solid tumors. *Int J Pharm*. 2008;347(1-2):118-127.
- 341. Pore N, Wu S, Standifer N, et al. Resistance to durvalumab and durvalumab plus tremelimumab is associated with functional STK11 mutations in patients with non-small cell lung cancer and is reversed by STAT3 knockdown. *Cancer Discov.* 2021;11(11):2828-2845.
- 342. Lee DW, Gardner R, Porter DL, et al. Current concepts in the diagnosis and management of cytokine release syndrome. *Blood*. 2014;124(2):188-195.
- 343. Guha P, Gardell J, Darpolor J, et al. STAT3 inhibition induces Bax-dependent apoptosis in liver tumor myeloid-derived suppressor cells. Oncogene. 2019;38(4):533-548.
- Chan HCS, Shan H, Dahoun T, Vogel H, Yuan S. Advancing drug discovery via artificial intelligence. *Trends Pharmacol Sci.* 2019;40(8):592-604.

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