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Gastrointestinal involvement in Parkinson's disease: pathophysiology, diagnosis, and management

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Growing evidence suggests an increasing significance for the extent of gastrointestinal tract (GIT) dysfunction in Parkinson's disease (PD). Most patients suffer from GIT symptoms, including dysphagia, sialorrhea, bloating, nausea, vomiting, gastroparesis, and constipation during the disease course. The underlying pathomechanisms of this α -synucleinopathy play an important role in disease development and progression, i.e., early accumulation of Lewy pathology in the enteric and central nervous systems is implicated in pharyngeal discoordination, esophageal and gastric motility/peristalsis impairment, chronic pain, altered intestinal permeability and autonomic dysfunction of the colon, with subsequent constipation. Severe complications, including malnutrition, dehydration, insufficient drug effects, aspiration pneumonia, intestinal obstruction, and megacolon, frequently result in hospitalization. Sophisticated diagnostic tools are now available that permit more detailed examination of specific GIT impairment patterns. Furthermore, novel treatment approaches have been evaluated, although high-level evidence trials are often missing. Finally, the burgeoning literature devoted to the GIT microbiome reveals its importance for neurologists. We review current knowledge about GIT pathoanatomy, pathophysiology, diagnosis, and treatment in PD and provide recommendations for management in daily practice.

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INTRODUCTION

Thirty years ago, gastrointestinal tract (GIT) symptoms in Parkinson's disease (PD) played a subordinate role in clinical practice of most neurologists and movement disorders specialists, despite the existence of numerous very early clinical reports and neuropathological findings that clearly provided evidence of relevant GIT involvement^{1–3}. Although GIT dysfunction can precede somatomotor symptoms by up to 20 years^{4–6} and impact negatively on quality of life^{7,8}, clinicians mainly focused on the 'classical' motor symptomatology at that time. It was not until the late 1980s and early 1990s that the number of clinical studies and neuropathology publications about the complex interaction of PD and the GIT begin to increase, thereby, also stimulating the interest of clinical neurologists for this topic^{9–15}. After 2000, the quest for explanations regarding the potential role of the GIT and the peripheral nervous system in the pathogenesis of PD gained momentum owing to the work by the Braak group, among others^{16–19}. Thus, one finds in PubMed for the period 1960–1969 eleven (keywords 'Parkinson' and 'dysphagia') and four (keywords 'Parkinson' and 'constipation') publications, from 1970 to 1979 twenty ('Parkinson' and 'dysphagia') and seven ('Parkinson' and 'constipation') publications, as opposed to more than ten entries per year beginning in 1992 and 2003, and, alone for the year 2020, 109 articles ('Parkinson' and 'dysphagia') and 93 ('Parkinson' and 'constipation') (<[parkinson dysphagia - Search Results - PubMed \(nih.gov\)](#)> and <[parkinson constipation - Search Results - PubMed \(nih.gov\)](#)>).

Today, GIT research is a promising and still growing field of inquiry and continues to provide neurologists and movement disorders specialists with novel and valuable data that can help to better understand the complexity of PD. At this point, we still have

a multitude of jigsaw puzzle pieces that must be carefully pieced together. Periodic review articles are intended to serve as the basis for future work^{5,6,20,21}. Here, we provide an overview of the latest knowledge, hypotheses, and debates about the pathology, pathophysiology, diagnostic methods for oropharyngeal and esophageal affection as well as impairment of the lower GIT, including a summary of current treatment strategies from an interdisciplinary standpoint. These might be helpful for neurologists, speech- and language therapists, and other clinicians in their daily work with PD patients and PD-associated GIT dysfunction.

ANATOMY: GASTROINTESTINAL TRACT AND ASSOCIATED BRAIN AREAS: CENTRAL CONTROL OF GASTROINTESTINAL MOTILITY

The entire GIT is one of the major gateways for extrinsic influences upon the human body. It is autonomously innervated by the largest part of the peripheral nervous system, the so-called enteric nervous system (ENS), which contains several hundreds of neurons²² and even more glial cells. Both neurons and glial cell populations have a variety similar to that found in the brain. Neurons consist of motoneurons, secretomotor-, or interneurons that express acetylcholine, nitric oxide synthase, catecholamines, GABA, or a broad range of neuropeptides^{23–26}. Glial cells can be found in at least four different morphologies and chemical codings, expressing S100B, the reactive gliosis marker GFAP, PDGFR α , or proteolipid-protein-1^{27,28}. Both neurons and glial cells form complex networks that populate in ganglionic and aganglionic plexus the complete gut wall from esophagus to anus and from serosa to mucosal layer.

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The ENS varies significantly along the gut axis, analogue to the distinct functional differences between the individual gut segments. Different neuronal subtypes and glial cells form neuronal circuits that allow the autonomous regulation of gastrointestinal motility²⁹. Although the gut works independently, there is a varying influence of the central nervous system (CNS) via several additional extrinsic inputs, of which the vagus nerve is the largest. The vagus nerve is part of the so-called brain-gut-axis that connects the CNS with the GIT. The brain-gut-axis consists of two main routes between the two organs³⁰. One is based on humoral factors, such as cytokines, hormones, or even bacterial metabolites from the gut microbiome, whereas the other is a hard-wired connection: the vagus nerve. The vagus nerve contains up to 50,000 fibers that run in both directions, the afferent ones being the majority with ~90% of all fibers³¹. While the afferent fibers deliver information from the gut, the efferent fibers provide parasympathetic motor stimuli that originate in two brainstem nuclei, first the dorsal motor nucleus of the vagus and, second, the ambiguus nucleus, which both contribute to gastrointestinal motility. Additionally to this direct input from the brainstem, there are several routes of influence represented, i.e., by sympathetic fibers from prevertebral ganglia that connect the gut with thoracic segments of the spinal cord²⁹.

The CNS influence upon gastrointestinal motility is dependent on the location. While there is a considerable impact on both esophagus and stomach³², the gut becomes more independent in the small and large intestine, where autonomous reflex circuits control smooth muscle activity, local blood flow, or secretion and absorption along the mucosal barrier. Interestingly, recent studies provide evidence that there are neural connections between the vagal nuclei and various areas of the cortex that influence stomach motility³³ and, thus, might be affected in PD, as demonstrated in a rat model³⁴. Especially the latter might explain the top-down gastrointestinal symptoms in PD. Based on the dual-hit hypothesis³⁵, there will also be a bottom-up process, possibly initiated by local inflammation and a compromised mucosal barrier, that allows gut content, including lipopolysaccharides, short chain fatty acids (SCFA) or other bacterial metabolites to enter the gut wall. In PD patients, the mucosal barrier is compromised and corresponding markers, such as calprotectin, can be found in the feces³⁶. There is a vast amount of evidence that the microbiome in PD patients is disturbed³⁷, combined with an alteration of SCFAs³⁸. Recent studies demonstrate the existence of neuronal circuits that monitor the microbiome or its metabolites report to the CNS or lead to modification of the innervation³⁹. These findings open up perspectives for using the gut, its intrinsic nervous system, the mucosal barrier, or the microbiome as therapeutic targets.

PATHOLOGY: ALPHA-SYNUCLEINOPATHY IN THE GIT OF INCIDENTAL LEWY BODY DISEASE AND PARKINSON'S DISEASE

Lewy pathology (LP, Lewy bodies, Lewy neurites) in prodromal PD (at autopsy, incidental Lewy body disease, ILBD^{40–42}) and in sporadic PD occurs throughout the human GIT^{3,9,43–48}. Interpretation of ENS histological slides from intestinal biopsies requires caution because immunocytochemical protocols vary considerably, and α -synuclein immunoreactivity must be distinguished from α -synuclein aggregates (LP) and α -synuclein aggregating species⁴⁹. As staged cases show, LP exists in the olfactory bulb, spinal cord, peripheral autonomic ganglia, submandibular gland, cardiac nerves, and ENS before it appears in the substantia nigra, pars compacta, and before neuronal loss occurs there^{17,50–54}. The aggregated α -synuclein lesions are not transient.

One of the largest studies examined a wide range of organs from 92 autopsied individuals, including 17 PD, 7 ILBD, and 23 controls⁴⁶. In pure PD, LP was found in the GIT of 64.7% (11/17), peripheral vagal nerve (pN. X) of 73.3%, and sympathetic trunk of

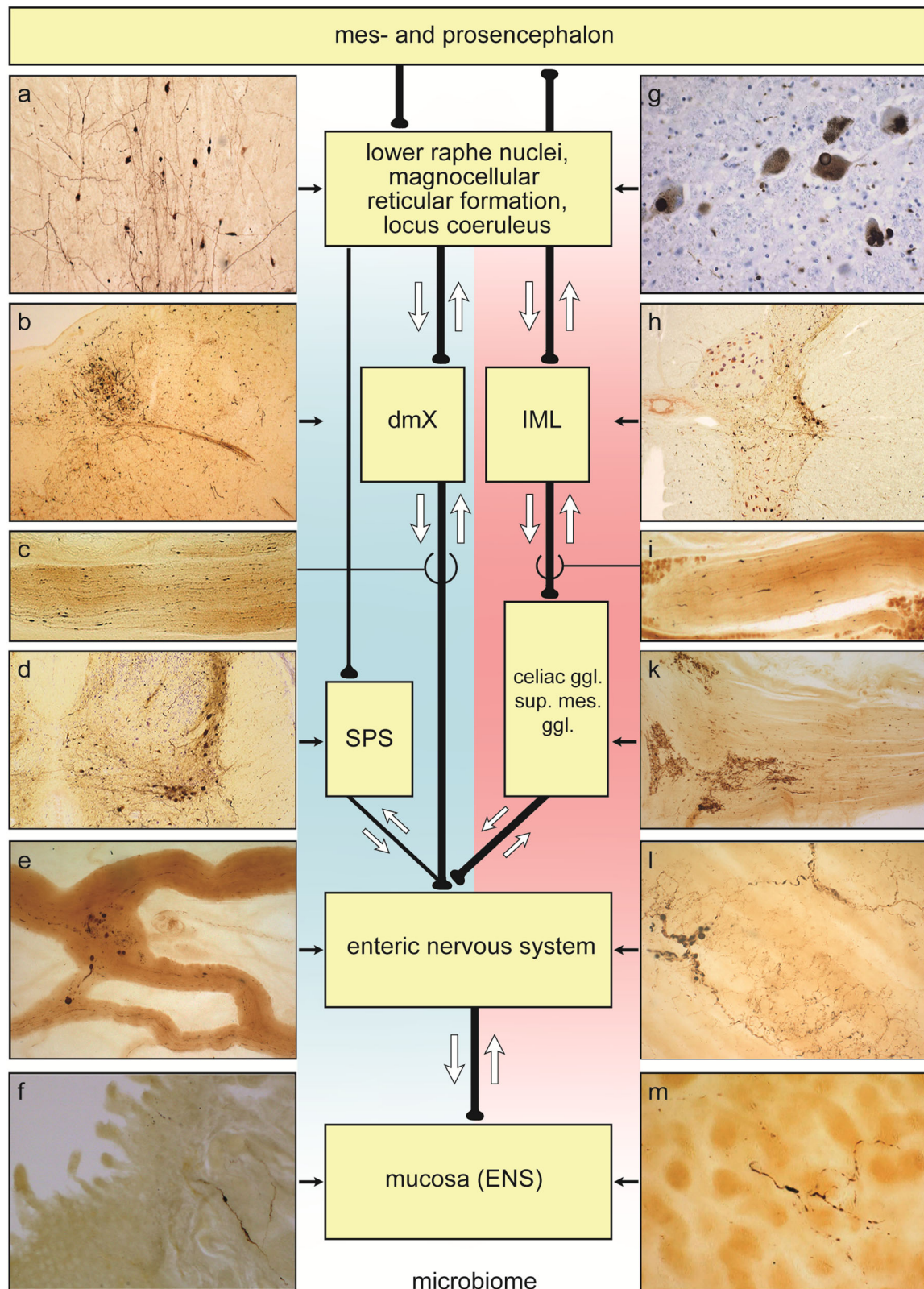
80% of cases. In ILBD, 14.2% (1/7) showed LP in the GIT and 28.57% in the pN. X. 50% of the ILBD group also displayed LP in the sympathetic trunk. A decreasing ENS rostral-caudal immunostaining gradient was seen, corroborating an earlier report⁹: The upper GIT, i.e., distal esophagus followed by the stomach, tended to display the highest pathological burdens⁴⁶; both sites are directly controlled by parasympathetic preganglionic fibers of the vagus nerve^{31,55,56} (see section "Anatomy: gastrointestinal tract and associated brain areas: central control of gastrointestinal motility" above). Although there were no 'ENS-only' (i.e., 'ENS-first') cases in the cohort, the authors surmised that "the findings of the present study are not incompatible with a GI entry for PD, ILBD and DLB"⁴⁶. In a subsequent investigation of pN. X tissue, they concluded that "the results [i.e., LP in 10/18 ILBD and 42/44 PD subjects; no LP in 49 controls] support initiation of Lewy-type alpha-synucleinopathy in the brain, with early, in some cases preclinical, subsequent progression to the peripheral nervous system"⁵⁷. However, a lack of α -synuclein immunopositivity in the true control (as opposed to ILBD) group is not surprising [see also⁵⁸] and, in any event, in 55.56% (ILBD) and 95.45% (manifest PD) of cases the pN. X was involved.

The 2010 study may have a selection bias because, to assess the relative LP frequency in the periphery, the ultimate choice of regions for further study was reduced to those with "a greater likelihood to have positive staining"⁴⁶ rather than shared innervation or neuronal circuitries. In addition, for the majority of their cases, the authors examined only a single slide for each ENS subdivision⁴⁶. Relevant ENS-related autonomic structures, e.g., the lumbar prevertebral celiac ganglion (sympathetic innervation of the esophagus, stomach, duodenum, pancreas, liver) and spinal cord SPS⁴⁴, but also the appendix vermiformis and superior mesenteric ganglion^{47,48,59}, were not included.

Results from investigations performed to date on large cohorts show, despite divergent findings from smaller studies^{60,61}, that 0% of ILBD cases displayed LP in the ENS in the absence of brain lesions^{44–46}. Of the prodromal PD cases analyzed by Stockholm et al., 44% (17/39) of subsequent PD patients displayed no ENS LP⁴⁷, and not even all manifest PD cases from autopsy-based studies had ENS involvement^{44,46,53,59} see also ref. ⁶². The heterogeneity of findings obtained from such studies is complicated not only by differences in immunocytochemical staining techniques and study design (cohort sampling size and stratification, retrospective vs. longitudinal), but also by dissection protocols. Borghammer & Van Den Berge point out that the human GIT "measures ~8–10 m at post-mortem and has a geometric surface area of at least 7000 cm². Thus, many hundreds of microscopy slides are required to rule out... gut pathology with any degree of confidence"⁶³. Similarly, pN. X specimens necessarily come from a very limited portion of the entire nerve^{44,52,57,58,64}, and multiple sections from both vagal trunks would be required to ascertain whether LP is present or absent in a given individual.

The argument that autopsy-based studies report only rare cases of LP in the ENS in the absence of brain LP does not eliminate the possibility of an ENS and/or peripheral nervous system origin for PD (ILBD), or of LP spread from the ENS to the brain^{16,17,35,62}. That a potential anatomical pathway from intrinsic ENS neurons to the pN. X exists, receives support from the fact that the myenteric plexus, epithelial enteroendocrine cells, and preganglionic portions of the N. X express normal α -synuclein^{65,66}. Converging lines of evidence also support the idea of propagation via cell-to-cell transsynaptic transmission of misfolded α -synuclein into recipient cells; there, misfolded α -synuclein can recruit native α -synuclein and become a template for development of pathological aggregates^{67–69}. Formalin-fixed tissue from the stomach is capable of limited to robust seeding in ILBD (5/8 cases), PD (10/12 cases), and controls (2/9)⁷⁰.

Overexpressed human α -synuclein and human α -synuclein lysates in animal models can mediate cellular dysfunction^{71,72} as



well as pathological spreading (seeding) along the pN. α bidirectionally^{73–75}, and anatomical connectivities^{73,76–82} make both directionalities conceivable in human α -synucleinopathy (Fig. 1). The latest findings in the appendix vermiformis of 46/48 ILBD cases^{47,48} indicate that additional ‘conduits’ for such pathological α -synuclein transport could exist (Fig. 1).

An awareness is gradually emerging that the presence of LP in the GIT and its patterns of progression may well differ between various PD subpopulations and in ILBD^{48,58,63,83,84}. Borghammer & Van Den Berge postulated the existence of a ‘PNS-first Lewy body disorder phenotype’, wherein early pathology in the peripheral autonomic nervous system might spread along retrograde

Fig. 1 Diagram showing possible bidirectional (white arrows) parasympathetic (blue background) and sympathetic (pink background) pathways along which pathological α -synuclein propagation in ILBD and PD could occur between the periphery, including the ENS, and the CNS. *Retrograde: parasympathetic* (distal esophagus/stomach \rightarrow pN. X \rightarrow dorsal motor nucleus of the vagus nerve, dmX); *parasympathetic* (appendix vermiformis \rightarrow RIM \rightarrow pN. X \rightarrow dmX); *parasympathetic* (descending colon and further distal \rightarrow ganglion pelvicum \rightarrow SPS preganglionic neurons \rightarrow lower brainstem level-setting nuclei); *sympathetic* (distal esophagus/stomach \rightarrow prevertebral celiac ganglion postganglionic neurons \rightarrow IML preganglionic neurons \rightarrow lower brainstem level-setting nuclei). Alternatively, *anterograde: parasympathetic* (dmX \rightarrow pN. X \rightarrow distal esophagus/stomach); *parasympathetic* (lower brainstem level-setting nuclei \rightarrow SPS preganglionic neurons \rightarrow prevertebral postganglionic ganglion pelvicum \rightarrow descending colon and portions further distal); *sympathetic* (appendix vermiformis \rightarrow RIM \rightarrow prevertebral SMG postganglionic neurons \rightarrow Nn. splanchnici \rightarrow IML preganglionic neurons \rightarrow lower brainstem level-setting nuclei). Abbreviations: pN. X peripheral vagus nerve, dmX dorsal motor nucleus of the vagus nerve, IML intermediate mediolateral nucleus, SPS sacral parasympathetic nucleus, RIM root of the small intestine mesentery, sup. mes. ggl. superior mesenteric ganglion. The level-setting nuclei consist of the lower raphe nuclei, magnocellular nucleus of the reticular formation, and locus coeruleus⁸¹. The RIM contains parasympathetic and sympathetic fibers innervating the upper GIT extending from the proximal jejunum to the distal ileum, thereby making it another potentially useful structure for neuropathological diagnosis of the existence of LP in the small intestine^{61,232}. Illustrations showing LP in **a–m** are not to scale: **a** great raphe nucleus. **b** dmX and intramedullary N. X. **c** pN. X at level of the carotid bifurcation. **d** SPS. **e** Gastric cardia, Auerbach plexus, tangential section. **f** Jejunum, Meissner (submucous) plexus, transversal section. **g** Locus coeruleus. **h** IML. **i** Splanchnic nerve at the level of the celiac ganglion. **k** Celiac ganglion. **l** Distal esophagus, Auerbach plexus, tangential section. **m** Gastric cardia, Meissner (submucous) plexus, tangential section. LP within the lamina propria reach the mucosa near gastric glands. Syn-1 immunohistochemistry (BD Biosciences, Eysins, Switzerland) in 100–150 μ m sections.

connectivities to nuclei of the lower brainstem that contribute to rapid-eye-sleep (REM) regulation⁶³ (Fig. 1). This intriguing hypothesis, drawing on insights gleaned from idiopathic REM sleep behavioral disorder (RBD) research^{85–92}, could also be tested neuropathologically, provided tissue from RBD patients were still available, in autopsy cohorts staged according to the 2003 PD staging protocol and including early ILBD^{93–95}.

NEUROLOGY: UPPER GIT

Oropharyngeal phase

Prevalence of dysphagia. Oropharyngeal dysphagia is a common and often disabling clinical manifestation in PD. Recent meta-analysis estimates the prevalence of oropharyngeal dysphagia up to 82% during the course of the disease⁹⁶. However, only 20–40% of patients are aware of their swallowing dysfunction, and only less than 10% report their complaint spontaneously^{97,98} which might result from early pharyngeal hyposensitivity^{21,99}. Recent research has led to the conclusion that oropharyngeal dysphagia is not only a late-stage PD symptom but can occur during any stage of the disease, including the preclinical or prodromal stages^{96,100,101}. Therefore, a comprehensive examination of oropharyngeal swallowing function should be performed regularly, even in early disease stages, when defined clinical predictors (see below) are present¹⁰².

Pathophysiology. In contrast to all other parts of the GIT, the oropharynx is not only innervated by the involuntary ENS but is also controlled from voluntarily triggered mechanisms of skeletal muscle movements. Thus, some additional pathophysiological mechanisms play a role that are also relevant for other somatomotor symptoms of PD, such as bradykinesia or tremor²¹:

1. Accumulation of Lewy pathology (see section “Pathology: alpha-synucleinopathy in the GIT of incidental Lewy body disease and Parkinson’s disease” above) takes place not only in the substantia nigra but also in various non-dopaminergic swallowing-relevant brainstem and cortical areas.
2. Putamen and globus pallidus are activated bilaterally during normal swallowing. Therefore, lack of dopamine in the striatum of PD patients may impair this part of the supramedullary swallowing network.
3. Peripheral mechanisms might be also involved as indicated by α -synuclein deposits in the peripheral sensory and motor nerves of the larynx as well as disease-induced neuromuscular alterations of pharyngeal muscles^{103–105}. Substance P plays an important role in these peripheral oropharyngeal mechanisms.

Substance P (SP). SP is an ubiquitous neuropeptide in the nervous system, with immunoreactive fibers having been detected in the laryngeal nerves, epithelium, and basal membrane of pharyngeal mucosa, especially on the surface of the epiglottis. SP mediates the response to local stimuli in the pharyngeal mucosa and thereby enhances the swallow and cough reflexes¹⁰⁶. A reduction of substance P, found in PD patients’ sputum, is postulated to lead to a disturbance of protective reflexes and, ultimately, silent aspiration¹⁰⁷. Reduced saliva concentrations of substance P may also be a predictor for the presence of early pharyngeal swallowing dysfunction¹⁰⁶. Table 1 provides an overview of PD-related oropharyngeal dysphagia clinical manifestations and postulated pathomechanisms¹⁰⁸.

Main pathological findings. The main phenotype characteristic of PD-related dysphagia is insufficient pharyngeal bolus clearing, with residues predominant located in the valleculae, in addition to pharyngolaryngeal movement disorders, above all pharyngeal bradykinesia¹⁰⁹. Silent aspiration is also a frequent clinical manifestation, even in early stages, but the risk increases with disease duration¹⁰⁰. In an analogy to freezing of gait with similar pathophysiologic mechanisms, a recent study described the presence of oropharyngeal freezing resulting in a temporally missing or delayed swallowing reflex¹¹⁰.

Dual task situations (i.e., cognitive or somatomotor tasks) are challenging to swallowing functional reserve capacities and therefore should be integrated into standard instrumental swallowing evaluations^{111,112} (Fig. 2). Furthermore, retention of medications in the hypopharynx for long periods of time may account for erratic absorption of levodopa with an insufficient or unpredictable clinical response to oral medication²¹ (Fig. 3). An association of delayed on-phenomena with pharyngeal residues could have been shown as well¹¹³. In a recent study, substantially impaired ability to swallow tablets or capsules was found in 28% ($n = 33/118$) of patients at all stages of the disease¹¹⁴. Capsules were the easiest to swallow, whereas oval tablets were the most difficult¹¹⁴.

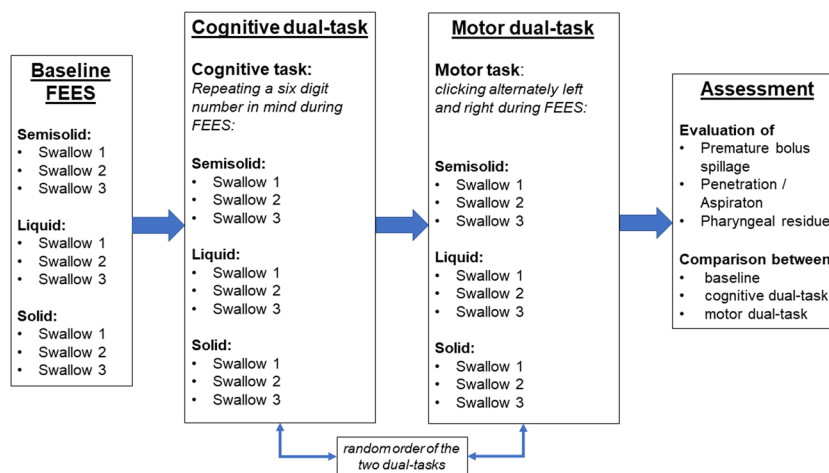
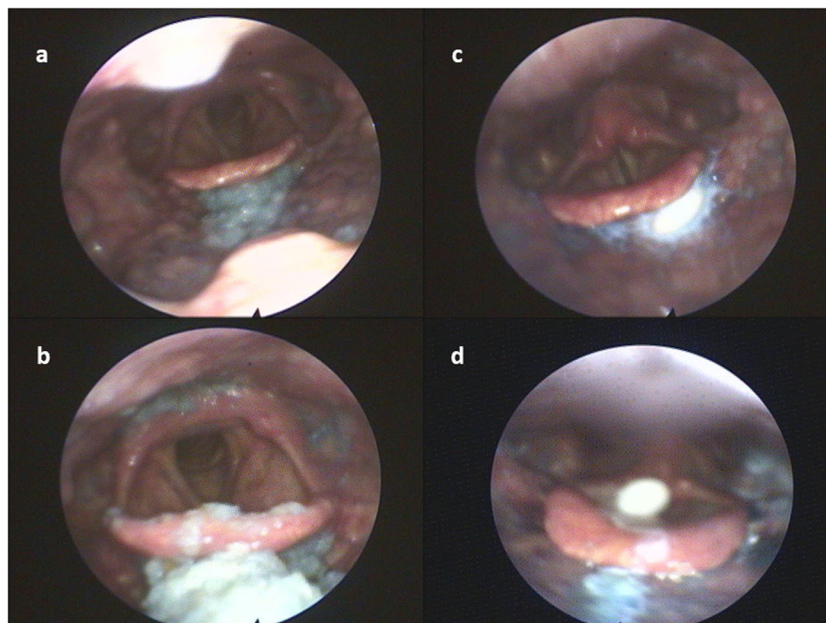
Clinical predictors. The following clinical conditions have been linked to oropharyngeal dysphagia in PD and can be considered predictors^{21,115,116}:

- Hoehn and Yahr stage \geq III
- Relevant weight loss
- Body Mass Index (BMI) \leq 20 kg/m²
- Severe drooling or sialorrhea
- Dementia

Another major problem for PD patients is drooling^{117,118}. Sialorrhea in PD patients usually does not result from an increased

Table 1. Overview of PD-related oropharyngeal dysphagia clinical manifestations and postulated pathomechanisms.

Clinical manifestation	Pathomechanisms
Prolonged oral transit time:	Dopaminergic + non-dopaminergic (especially Lewy pathology in swallowing cortex?)
Premature spillage:	Dopaminergic + non-dopaminergic (Lewy pathology in swallowing cortex?)
Delayed swallow reflex:	Dopaminergic + decreased Substance P concentration
Prolonged pharyngeal transit time:	Dopaminergic + non-dopaminergic (Lewy pathology in brainstem?)
Penetration:	Dopaminergic + non-dopaminergic
Aspiration:	Dopaminergic + non-dopaminergic
Residue in valleculae:	Primarily dopaminergic
Residue in piriform sinus:	Dopaminergic + non-dopaminergic
Dysfunction of upper esophageal sphincter:	Primarily non-dopaminergic (Lewy pathology in swallowing centers of medulla oblongata?)
Insufficient cough reflex:	Decreased Substance P concentration

Source ¹⁰⁸.**Fig. 2** Dual Task examination algorithm via Flexible Endoscopic Evaluation of Swallowing (FEES) (adapted from ref. ¹¹²). FEES examination protocol including cognitive and motor dual-task for evaluation of swallowing function in PD patients.**Fig. 3** Examples for pharyngeal residue via FEES. **a** Mild residue for solid food located in the valleculae. **b** Moderate to severe residue for solid food located in the valleculae and piriform sinus with penetration into the laryngeal vestibule. **c** Tablet residue located in the valleculae. **d** Tablet penetration.

production of saliva but from a reduced spontaneous swallowing rate (48/h vs. 71/h) and/or from oropharyngeal dysphagia with a reduced ability to swallow saliva⁵². The extent of sialorrhea correlates with the severity of PD-related dysphagia¹¹⁹.

Diagnostic management

Screening-tools. Questionnaires and a specific water swallow test may be used as screening tools:

1. The Swallowing Disturbance Questionnaire (SDQ) with a sensitivity of 80.5% and a specificity of 81.3% is simple to apply using a score of 15 dysphagia-associated questions to detect PD-related dysphagia¹²⁰. A score >10 recommends further dysphagia diagnostic (max. score 44.5). Additionally, the patient-rated Radboud Oral Motor Inventory for Parkinson's disease (ROMP) questionnaire is used for assessment of speech, swallowing, and saliva control¹²¹.
2. The Munich Dysphagia Test—Parkinson's disease (MDT-PD) with a sensitivity of 82% and a specificity of 71% was designed to detect milder forms of dysphagia without aspiration risk¹²², although its usefulness as a screening tool for aspiration events is controversial¹²³. A French version is also available¹²⁴.
3. The Non-Motor Symptoms Questionnaire (NMS-Quest; Question 3: "difficulty swallowing food or drink or problems with choking") and the Movement Disorder Society—Unified Parkinson's disease rating scale (MDS-UPDRS; Question 2.3 of the UPDRS II: "problems swallowing pills or eating meals") each also include one question about swallowing difficulties^{125,126}. However, in a recent study, NMS and MDS-UPDRS were identified as unreliable tools for detecting previous aspiration¹²⁷.
4. Normal water tests that are useful for diagnosing severe dysphagia, e.g., in stroke patients, are not reliable screening tools for PD-related dysphagia when compared with instrumental diagnostic tools¹¹⁵. A detection of a 'wet voice' after different bolus consistencies showed a too low sensitivity to be a solid marker of penetration/aspiration in PD¹²⁸. Therefore, a modified water test was developed to evaluate the stimulability of drinking by using a maximum performance test (maximum swallowing volume <20 mL, maximum swallowing speed <10 mL/s)¹²⁹. Nonetheless, in a more recent study, swallowing speed was found to be prone to methodological errors and not suitable as a screening instrument to predict aspiration in PD patients¹³⁰.

Instrumental diagnostic tools. Flexible Endoscopic Evaluation of Swallowing (FEES) and Videofluoroscopic Swallowing Study are both considered to be the gold standard for evaluating oropharyngeal dysphagia^{131,132}. These instrumental tools should be applied in cases of unclear and severe PD-associated dysphagia, especially to detect silent aspiration as well as specific dysphagia phenotypes^{129,133}.

Therapeutic management. Over the years, a number of studies have provided evidence-based recommendations for treatment of oropharyngeal dysphagia^{21,101,134,135}. Swallowing therapy, especially as performed by speech and language therapists, and other specific therapeutic options might help to improve oropharyngeal swallowing impairment:

Pharmacotherapy. The effects of dopaminergic medication and levodopa on swallowing function and its role in dysphagia treatment are controversially discussed^{136–138}. Oropharyngeal swallowing parameters with good levodopa-responsiveness could be pharyngeal residue (especially in the valleculae), penetration,

and oral as well as pharyngeal transit times^{139,140}. Furthermore, some studies indicate positive effects of the dopamine agonists apomorphine and transdermal rotigotine^{141–143}. As such, an examination whether an improvement in swallowing function could be achieved by increasing or optimizing dopaminergic medication should be performed on a case-by-case basis, i.e., by using the FEES-Levodopa-Test. In this test, three salient parameters (premature spillage, penetration/aspiration events, and residues, each tested with liquid, semisolid, and solid food consistencies) are assessed in off- and on-stage conditions performing a specific score¹⁴⁰. A score improvement of >30% indicates levodopa responsiveness of dysphagia¹⁴⁰. Subsequently, in such cases, optimizing dopaminergic medication should be considered. Furthermore, levodopa-carbidopa intestinal gel (LCIG) infusion therapy might be capable of alleviating pharyngeal bradykinesia and premature bolus spillage¹⁴⁴.

Deep brain stimulation (DBS). To date, detailed information about the effects of DBS on swallowing function in PD patients remain limited^{145,146}. Using stimulation of the subthalamic nucleus (STN), low-frequency stimulations (i.e., 60 Hz) may have a beneficial effect on swallowing dysfunction in patients with freezing of gait^{145,147}, whereas high-frequency stimulation might result in beneficial, no, or detrimental effects¹⁴⁵. A short-term improvement (lower aspiration rate) was indicated as well but without confirmation in the long-term observation¹⁴⁸. Simultaneous STN and substantia nigra (SNr) stimulation seem to have no additional beneficial effect on dysphagia compared with conventional STN stimulation, but swallowing function does not deteriorate as a result¹⁴⁹.

Neuromuscular electrical stimulation (NMES). The use of NMES in PD patients seems to have no measurable benefit^{150,151}. A recent study using new electrode placement methods indicates increased hyoid bone movement and reduced aspiration risks¹⁵², but NMES cannot be recommended for PD dysphagia treatment at the present time.

Behavioral swallowing therapy. Because of heterogenous study populations and therapeutic methods as well as different outcome measures, general recommendations for non-pharmacological treatment are difficult to provide¹³⁵. However, some therapeutic strategies are promising for individual treatment of specific patterns of PD-related dysphagia: Thickened liquids and the chin-tuck maneuver might help to prevent liquid aspiration^{153,154}. The Lee Silverman Voice Treatment (LVST®), originally developed for treatment of PD-associated dysarthria, can also improve swallowing function, although controlled clinical trials are not yet available and the effects are unspecific¹⁵⁵. With regard to therapeutic strategies, dual task situations should be avoided in real-life circumstances to focus attention on swallowing performance¹¹². Two larger, randomized placebo-controlled studies showed a positive effect on swallowing safety and efficiency in PD patients who had performed a 4-week- expiratory muscle strength training regimen^{156,157}. Video-assisted swallowing therapy and specific swallowing skill training using surface electromyography might also be helpful for providing biofeedback to patients^{158,159}. In general, every affected PD patient should receive a detailed examination of swallowing disturbance patterns resulting in an individual training program based on available therapeutic methods. The efficacy of the method(s) selected should be confirmed via instrumental testing¹³³.

Treatment of sialorrhea. Parkinson-related sialorrhea can be managed effectively with injections of botulinum toxin A or B into the parotid and submandibular glands¹⁶⁰. Another pharmacological treatment option might be the application of the anticholinergic drug glycopyrrolate because it crosses the

blood-brain barrier and therefore does not have central anticholinergic side-effects¹⁶¹. In addition, gum chewing also helps to improve PD-related sialorrhoea in the short term but without maintaining a long-term effect¹⁶².

Esophago-gastral phase

Prevalence and main clinical findings. The prevalence of impaired gastric emptying in PD ranges from 70 to 100% and may be present in both early and advanced stages^{6,163}. Major clinical manifestations include nausea, vomiting, early satiety, and postprandial fullness, and these can lead to weight loss, malnutrition, and dehydration¹⁶⁴. Furthermore, there is growing evidence for a significant relationship between delayed gastric emptying and levodopa pharmacokinetics leading to drug-response fluctuations with delayed or missed on-phases after medication intake¹⁶⁵.

Esophageal motility disorders appear to occur very early and even in premotor stages of PD^{166,167}. A hypotensive peristalsis of the tubular esophagus occurs most frequently and early in the disease course, whereas in later stages diffuse esophageal spasms and multiple contractions may develop¹⁶⁷. However, primary opening disorders of the upper esophageal sphincter are rare¹⁶⁷.

Diagnostic management. Impaired gastric emptying is defined as >60% retention at 2 h postprandially and/or >10% retention at 4 h after ingestion of a radioactive technetium Tc 99m-labeled solid food¹⁶⁸. Other quantitative methods are the use of breath tests with nonradioactive ¹³C-sodium octanoate bound into a solid meal, or real time visualization by magnetic resonance imaging and electrogastrography¹⁶⁴. Because clinical evaluation is difficult, diagnostic examination of esophageal motility disorders nowadays is normally performed by using High Resolution Manometry to detect esophageal alterations¹⁶⁹.

Therapeutic management. Therapeutic options for managing PD-associated esophageal motility disorders are rare to date. A pilot study indicates a possible usefulness of botulinum toxin injections for treatment of esophageal spasms, but more evidence is needed¹⁷⁰. STN stimulation might also improve esophageal motility¹⁷¹. The use of capsaicin seems to be capable of improving esophageal motility as well as upper esophageal sphincter contraction, and it might be a promising tool for further

treatment^{172,173}. In gastroparesis, the increase of levodopa dosage may impair delayed gastric emptying¹⁷⁴. Pharmacotherapy options using domperidone might be useful but are said to increase the risk of a long QT syndrome. Recent studies have indicated possible positive effects by using nizatidine or ghrelin agonists but these require further evaluation⁶. Benefits from botulinum toxin injection in the pyloric sphincter and possible use of STN-DBS have been reported as well^{175,176}. However, LCIG (with or without entacapone application), subcutaneous apomorphin, and the rotigotine patch are helpful solutions for bypassing the GIT and therefore could be administered in cases of clinically relevant effects of esophageal spasms as well as gastroparesis on somatomotor symptoms.

Summary/Practical algorithm for management

Disturbances of the upper GIT in PD, especially oropharyngeal dysphagia, are complex syndromes that occur early in disease duration but often remain unnoticed until severe complications, such as aspiration pneumonia, develop. Accordingly, standardized and early diagnostic approaches as well as focused treatment of specific dysphagia patterns are required to help affected individuals. Table 2 provides a summary of the most relevant clinical manifestations of upper GIT impairment and feasible treatment approaches.

NEUROLOGY: LOWER GIT

Colon

Prevalence of constipation. Since initially being mentioned by James Parkinson, constipation has been considered a very frequent symptom that occurs in up to 80% of PD patients^{6,14,177–181}. As is often the case, constipation is described as the most frequent autonomic symptom^{14,182,183}. Notably, healthy people with constipation complaints, including delayed passage of stools, hard stools, or a sensation of incomplete evacuation, have shown a greater risk for subsequently developing PD^{184–186}. This fits well the neuropathological studies published by Braak and coworkers^{16,17} (see chapter 3). Constipation is currently considered one of the most relevant early signs of PD, and its frequency seems to be higher than the subjective complaints^{19,184,185,187–189}. The GIT may even play an important role in PD pathogenesis^{16,80,185} and as a prognostic factor,

Table 2. Summary of most relevant clinical manifestations of upper GIT impairment and feasible treatment approaches.

Symptom	Pharmacotherapy	Swallowing therapy by Speech language therapists
Oropharyngeal freezing:	Increase dose of L-dopa before meal times Amantadine?	Triggering of swallowing reflex External triggers?
Premature spillage:		Oral bolus control Avoid dual tasks
Penetration/Aspiration:	Non-oral delivery: patch or pump?	Protective reflexes Sensory stimulation Supraglottic swallow maneuver Safe food consistencies? PEG?
Pharyngeal residues without motor fluctuations:	Individual assessment of L-dopa responsiveness, if positive: Increase dose of L-dopa before meals	Effortful swallow exercise
Pharyngeal residues without motor fluctuations:	Individual assessment of L-dopa responsiveness, if positive: Optimize oral treatment Non-oral delivery: patch or pump?	Meal times during on state condition Effortful swallow exercise in off state condition
Esophageal spasms:	Non-oral delivery: patch or pump? Botulinum toxin injections into upper esophageal sphincter?	Protective reflexes Mendelsohn swallow exercise Safe food consistencies? PEG?

inasmuch as a significant relationship between constipation severity and progression to dementia has recently been demonstrated^{190,191}. Additional studies are needed to determine whether a similar relationship exists in patients who develop dementia with Lewy bodies (DLB).

Pathophysiology. Medications, reduced physical movement, a reduced muscle tone in the diaphragm and abdominal musculature, and reduced intake of fibers and liquids have been advanced as causes for constipation¹⁴. Beginning with the earliest studies and onwards, anticholinergic agents have been particularly related to severe constipating effects, including even the development of a megacolon. Constipation in PD is definitely disease-related¹⁸⁹. It was described long before any specific therapy had been found^{14,177}, and many studies of yet untreated patients were able to demonstrate delayed transit^{6,14}. It is much more probable that, in PD patients, a delayed transit plays an intrinsic and prominent role, and that constipation even can be exacerbated by the medical treatment itself^{14,192}.

The causes underlying the delayed transit are most probably degenerative changes involving Lewy pathology located centrally, including the spinal cord intermediolateral nucleus, and peripherally extending from the upper esophagus to the rectum in the Auerbach plexus (myenteric plexus) and Meissner plexus (submucous plexus)^{3,43,47,53}. Additionally, anismus, a failure of relaxation, or involuntary contractions of the anal sphincters during defecation (extremely rare!), can lead to so-called “outlet” constipation¹⁴.

Diagnostic management. Because constipation can develop into a megacolon, pseudo-obstruction, or volvulus, adequate diagnosis is essential¹⁴. Unfortunately, a megacolon usually remains asymptomatic, with the exception of the singular symptom of constipation, although an ileus followed by surgery¹⁴ and colon perforation have been described as megacolon consequences. Surprisingly, despite the existence of severe constipation at the time of presentation, patients seldom report this problem spontaneously, often because of embarrassment, which indicates that it is most probably underdiagnosed¹⁸⁹.

A simple method during the diagnostic work-up involves administering radiopaque markers (ROM) (Fig. 4) as the gold standard diagnostic test¹⁴: Abdominal X-rays are taken at defined time intervals to identify the retained numbers of ingested ROMs to calculate colonic transit time. Alternatively, a gamma camera can be used to track the movements of radioisotope test meals or capsules at specified time points for quantitative evaluation of scintigraphic colonic transit times¹⁹³. For orientation, it is also useful to ask patients to eat poppy seed cake and then note when the poppy seeds are excreted.

Although there is no gold standard method for the assessment of outlet constipation, defecography is widely preferred. It involves the instillation of barium in the rectum, and subjects are then asked to empty it during recording of a cinematic film¹⁹⁴. Anorectal dysfunction can also be assessed by external anal sphincter electromyography, a balloon distension and expulsion test, and anorectal manometry¹⁹⁵.

Therapeutic management. At present, there are no specific guidelines for the management of PD-associated constipation available. A fiber-rich diet, psyllium as a bulk laxative, stool softener, and sufficient liquid-intake have high therapeutic value in treating constipation, but regular physical exercise and physical therapy are also advisable¹⁷⁸. Exclusion of aggravating factors, such as anticholinergics, should be considered¹⁷⁸. Unfortunately, these measures are only useful in mild or moderate cases. In many instances, a colonic transit of more than 7 days is reported, and no improvement of colonic transit can be achieved by the various therapeutic options owing to the upper threshold. In this case,



Fig. 4 Colonic transit time in a 72-year-old male PD patient. White spots in the entire colon are radiopaque markers (erect, anterior-posterior).

additional medication must be prescribed. An effect of domperidone in the upper GIT has not been shown for constipation¹⁷⁸.

Stimulants, such as bisacodyl, sodium picosulfate, and senna are safe and helpful¹⁹⁶. In addition, stimulant laxatives and osmotic laxatives are recommended. The best results to date are achieved with macrogol^{197,198}. A disadvantage of lactulose is flatulence¹⁴. Positive data are also available for therapies with probiotics and probiotic fibers^{196,199}.

There are still no studies on the effects of modern prokinetic agents, such as serotonin (5-HT₄) agonists, e.g., mosapride²⁰⁰. In the meantime, prucaloprid²⁰¹ has been approved for severe constipation and may be administered to PD patients, although specific studies in this population are still lacking. Several new drugs, including relamorelin (ghrelin agonist)¹⁸¹, and chlorid channel activators, such as linaclotide, lubiprostone, and plencatanide^{188,201–203} are in discussion. In rare cases of anismus, we recommend botulinum toxin injections¹⁴.

EXCURSUS: THE ROLE OF THE GIT MICROBIOME

The GIT microbiome in PD has been intensively researched in recent years^{6,30,37,38,204–210}. By applying metagenomic and next-generation sequencing procedures, it is now possible to distinguish PD patients from healthy individuals^{204,210} at a very early disease stage by means of individually altered microbiota²⁰⁴. There may even be a ‘prodromal’ GIT microbiome because a microbial shift has been found, for instance, in patients with RBD²¹¹. In one large cohort, reduced GIT microbial diversity in PD patients correlated significantly with greater GIT symptom severity in comparison to controls²¹², and evidence exists for an ‘enteric pro-inflammatory profile’ in PD^{213,214}. Intestinal dysbiosis and small intestinal bacterial overgrowth in PD patients²¹⁵ might increase intestinal barrier permeability, thereby triggering excessive stimulation of the innate immune system and systemic inflammation, mechanisms possibly involved in the initiation of a-

synuclein deposition^{216–218}. According to this scenario, α -synuclein expression in the GIT would reflect an immune defense mechanism²¹⁹, which is further supported by the finding that the protein is capable of triggering T cell responses that may also potentiate neurodegeneration²²⁰.

At present, interpretation of the available findings is difficult because a great variety of factors can influence the microbial configuration of the GIT. For example, evaluation of the GIT microbiome in patients undergoing treatment for PD is still of limited use, inasmuch as levodopa and other antiparkinson medications act upon the intestinal flora^{192,206,221}, and, at least in a subset of patients, the opposite is also the true²²². In addition, particularly for PD, it cannot be clarified retrospectively whether the altered GIT microbiome is the cause or the effect of motility disturbances, such as severe constipation^{30,207,223}, and the association between the microbiome and neuroinflammation in PD still remains unclear^{63,204,206,224}, in part because the cohorts studied to date, with few notable exceptions^{222,225}, have been small^{204,207}. Finally, if the microbiome and its metabolites were to play a key pathogenetic role in PD, then considerable differences should be observable between populations on different continents owing simply to dietary variability, but this has not proved to be the case. Nonetheless, it is imperative, going forward, to examine not only the precise role of the GIT microbiome and the effects a targeted diet and probiotics might have on PD patients^{196,226,227} but also the potential advantages and adverse side effects associated with fecal microbiota transplantation^{228–231}.

FINAL CONCLUSIONS/PRACTICAL ALGORITHM FOR MANAGEMENT

Dysfunction of the upper GIT in PD, especially oropharyngeal dysphagia, are complex syndromes occurring early in disease that often remain unnoticed until severe complications, such as aspiration pneumonia, become manifest. In the lower GIT, constipation is a widespread and debilitating symptom with the potential of leading to severe bowel complications and even cognitive dysfunction.

In closing, standardized and early diagnostic approaches together with continuous and long-term treatment are necessary to help patients (Table 2).

Reporting summary

Further information on research design is available in the Nature Research Reporting Summary linked to this article.

DATA AVAILABILITY

All data generated or analyzed during this study are included in this published review (see references).

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REFERENCES

- Eadie, M. J. & Tyrer, J. H. Alimentary disorder in parkinsonism. *Australas. Ann. Med* **14**, 13–22 (1965).
- Eadie, M. J. & Tyrer, J. H. Radiological abnormalities of the upper part of the alimentary tract in Parkinsonism. *Australas. Ann. Med* **14**, 23–27 (1965).
- den Hartog Jager, W. A. & Bethlem, J. The distribution of Lewy bodies in the central and autonomic nervous systems in idiopathic paralysis agitans. *J. Neurol. Neurosurg. Psychiatry* **23**, 283–290 (1960).
- Coelho, M. et al. Late-stage Parkinson's disease: the Barcelona and Lisbon cohort. *J. Neurol.* **257**, 1524–1532 (2010).
- Jost, W. H. Gastrointestinal dysfunction in Parkinson's disease. *J. Neurol. Sci.* **289**, 69–73 (2010).
- Fasano, A., Visanji, N. P., Liu, L. W., Lang, A. E. & Pfeiffer, R. F. Gastrointestinal dysfunction in Parkinson's disease. *Lancet Neurol.* **14**, 625–639 (2015).
- Lubomski, M., Rushworth, R. L. & Tisch, S. Hospitalisation and comorbidities in Parkinson's disease: a large Australian retrospective study. *J. Neurol. Neurosurg. Psychiatry* **86**, 324–330 (2015).
- Lubomski, M., Davis, R. L. & Sue, C. M. Gastrointestinal dysfunction in Parkinson's disease. *J. Neurol.* **267**, 1377–1388 (2020).
- Wakabayashi, K., Takahashi, H., Takeda, S., Ohama, E. & Ikuta, F. Parkinson's disease: the presence of Lewy bodies in Auerbach's and Meissner's plexuses. *Acta Neuropathol.* **76**, 217–221 (1988).
- Wakabayashi, K., Takahashi, H., Takeda, S., Ohama, E. & Ikuta, F. Parkinson's disease: an immunohistochemical study of Lewy body-containing neurons in the enteric nervous system. *Acta Neuropathol.* **79**, 581–583 (1990).
- Wakabayashi, K., Takahashi, H., Ohama, E., Takeda, S. & Ikuta, F. Lewy bodies in the visceral autonomic nervous system in Parkinson's disease. *Adv. Neurol.* **60**, 609–612 (1993).
- Bushmann, M., Dobmeyer, S. M., Leeker, L. & Perlmutter, J. S. Swallowing abnormalities and their response to treatment in Parkinson's disease. *Neurology* **39**, 1309–1314 (1989).
- Jost, W. H. & Schmirgk, K. Constipation in Parkinson's disease. *Klin. Wochenschr.* **69**, 906–909 (1991).
- Jost, W. H. Gastrointestinal motility problems in patients with Parkinson's disease: effects of antiparkinsonian treatment and guidelines for management. *Drugs Aging* **10**, 249–258 (1997).
- Edwards, L., Quigley, E. M. M., Hofman, R. & Pfeiffer, R. F. Gastrointestinal symptoms in Parkinson disease: 18-month follow-up study. *Mov. Disord.* **8**, 83–86 (1993).
- Braak, H., Rüb, U., Gai, W. P. & Del Tredici, K. Idiopathic Parkinson's disease: possible routes by which vulnerable neuronal types may be subject to neuroinvasion by an unknown pathogen. *J. Neural Transm.* **110**, 517–536 (2003).
- Braak, H., de Vos, R. A. I., Bohl, J. & Del Tredici, K. Gastric α -synuclein immunoreactive inclusions in Meissner's and Auerbach's plexuses in cases staged for Parkinson's disease related brain pathology. *Neurosci. Lett.* **396**, 67–72 (2006).
- Langston, J. W. The Parkinson's complex: parkinsonism is just the tip of the iceberg. *Ann. Neurol.* **59**, 591–596 (2006).
- Cersósimo, M. G. et al. Gastrointestinal manifestations in Parkinson's disease: prevalence and occurrence before motor symptoms. *J. Neurol.* **260**, 1332–1338 (2013).
- Klingelhoefer, L. & Reichmann, H. Parkinson's disease and gastrointestinal non motor symptoms: Diagnostic and therapeutic options – a practice guide. *J. Parkinsons Dis.* **5**, 647–658 (2015).
- Suttrup, I. & Warnecke, T. Dysphagia in Parkinson's disease. *Dysphagia* **31**, 24–32 (2016).
- Furness, J. B., Callaghan, B. P., Rivera, L. & Cho, H. J. The enteric nervous system and gastrointestinal innervation: integrated local and central control. *Adv. Exp. Med. Biol.* **817**, 39–71 (2014).
- Jessen, K. R. GABA and the enteric nervous system. A neurotransmitter function? *Mol. Cell Biochem.* **38 Spec No**, 69–76 (1981).
- Hens, J., Vanderwinden, J. M., De Laet, M. H., Scheuermann, D. W. & Timmermans, J. P. Morphological and neurochemical identification of enteric neurones with mucosal projections in the human small intestine. *J. Neurochem.* **76**, 464–471 (2001).
- Timmermans, J. P., Hens, J. & Adriaensens, D. Outer submucosal plexus: an intrinsic nerve network involved in both secretory and motility processes in the intestine of large mammals and humans. *Anat. Rec.* **262**, 71–78 (2001).
- Qu, Z. D. et al. Immunohistochemical analysis of neuron types in the mouse small intestine. *Cell Tissue Res.* **334**, 147–161 (2008).
- Gulbransen, B. D. & Sharkey, K. A. Novel functional roles for enteric glia in the gastrointestinal tract. *Nat. Rev. Gastroenterol. Hepatol.* **9**, 625–632 (2012).
- Grundmann, D. et al. Enteric glia: S100, GFAP, and beyond. *Anat. Rec.* **302**, 1333–1344 (2019).
- Spencer, N. J. & Hu, H. Enteric nervous system: sensory transduction, neural circuits and gastrointestinal motility. *Nat. Rev. Gastroenterol. Hepatol.* **17**, 338–351 (2020).
- Endres, K. & Schäfer, K. H. Influence of commensal microbiota on the enteric nervous system and its role in neurodegenerative diseases. *J. Innate Immun.* **10**, 172–180 (2018).
- Beyak, M. J., Bulmer, D., Jiang, W., Keating, W., Grundy, D. Extrinsic Sensory Afferent Nerves Innervating the Gastrointestinal Tract, Physiology of the Gastrointestinal Tract Ch. 25 In *Physiology of the Gastrointestinal Tract, 4th ed* (eds Leonard R. Johnson). (Academic Press, San Diego, 2006), pages 685–725.
- Travagli, R. A. & Anselmi, L. Vagal neurocircuitry and its influence on gastric motility. *Nat. Rev. Gastroenterol. Hepatol.* **13**, 389–401 (2016).
- Levinthal, D. J. & Strick, P. L. Multiple areas of the cerebral cortex influence the stomach. *Proc. Natl Acad. Sci. USA* **117**, 13078–13083 (2020).

34. Anselmi, L., Toti, L., Bove, C., Hampton, J. & Travaglini, R. A. A nigro-vagal pathway controls gastric motility and is affected in a rat model of parkinsonism. *Gastroenterology* **153**, 1581–1593 (2017).
35. Hawkes, C. H., Del Tredici, K. & Braak, H. Parkinson's disease: a dual hit hypothesis. *Neuropathol. Appl. Neurobiol.* **33**, 599–614 (2007).
36. Schwierz, A. et al. Fecal markers of intestinal inflammation and intestinal permeability are elevated in Parkinson's disease. *Parkinsonism Relat. Disord.* **50**, 104–107 (2018).
37. Boertjen, M., Pereira, P. A. B., Aho, V. T. E. & Scheperjans, F. Increasing comparability and utility of gut microbiome studies in Parkinson's disease: A systematic review. *J. Parkinsons Dis.* **9**, S297–S312 (2019).
38. Unger, M. M. et al. Short chain fatty acids and gut microbiota differ between patients with Parkinson's disease and age-matched controls. *Parkinsonism Relat. Disord.* **32**, 66–72 (2016).
39. Muller, P. A. et al. Microbiota modulate sympathetic neurons via a gut-brain circuit. *Nature* **583**, 441–446 (2020).
40. Dickson, D. W. et al. Evidence that incidental Lewy body disease is presymptomatic Parkinson's disease. *Acta Neuropathol.* **115**, 437–444 (2008).
41. Markesbery, W. R., Jicha, G. A., Liu, H. & Schmitt, F. A. Lewy body pathology in normal elderly subjects. *J. Neuropathol. Exp. Neurol.* **68**, 816–822 (2009).
42. Hyman, B. T. et al. National Institute on Aging-Alzheimer's association guidelines for the neuropathologic assessment of Alzheimer's disease. *Alzheimers Dement* **8**, 1–13 (2012).
43. Kupsky, W. J., Grimes, M. M., Sweeting, J., Bertsch, R. & Cote, L. J. Parkinson's disease and megacolon: concentric hyaline inclusions (Lewy bodies) in enteric ganglion cells. *Neurology* **37**, 1253–1255 (1987).
44. Bloch, A., Probst, A., Bissig, H., Adams, H. & Tolnay, M. Alpha-synuclein pathology of the spinal and peripheral autonomic nervous system in neurologically unimpaired elderly subjects. *Neurobiol. Appl. Neurol.* **32**, 284–295 (2006).
45. Probst, A., Bloch, A. & Tolnay, M. New insights into the pathology of Parkinson's disease: does the peripheral autonomic nervous system become central? *Eur. J. Neurol.* **15**, 1–4 (2008).
46. Beach, T. G. et al. Multi-organ distribution of phosphorylated α -synuclein histopathology in subjects with Lewy body disorders. *Acta Neuropathol.* **119**, 689–702 (2010).
47. Stokholm, M. G., Danielsen, H. K., Hamilton-Dutoit, S. J. & Borghammer, P. Pathological α -synuclein in gastrointestinal tissues from prodromal Parkinson disease patients. *Ann. Neurol.* **79**, 940–949 (2016).
48. Killinger, B. A. et al. The vermiform appendix impacts the risk of developing Parkinson's disease. *Sci. Transl. Med.* **10**, eaar5280 (2018).
49. Shannon, K. & Vanden Berghe, P. The enteric nervous system in PD: gateway, bystander victim, or source of solutions. *Cell Tiss. Res.* **373**, 313–326 (2018).
50. Braak, H. et al. Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiol. Aging* **24**, 197–211 (2003).
51. Del Tredici, K., Rüb, U., de Vos, R. A. I., Bohl, J. R. E. & Braak, H. Where does Parkinson disease pathology begin in the brain? *J. Neuropathol. Exp. Neurol.* **61**, 413–426 (2002).
52. Del Tredici, K., Hawkes, C. H., Ghebremedhin, E. & Braak, H. Lewy pathology in the submandibular gland of individuals with incidental Lewy body disease and sporadic Parkinson's disease. *Acta Neuropathol.* **119**, 703–713 (2010).
53. Del Tredici, K. & Braak, H. Spinal cord lesions in sporadic Parkinson's disease. *Acta Neuropathol.* **124**, 643–664 (2012).
54. Ghebremedhin, E., Del Tredici, K., Langston, J. W. & Braak, H. Diminished tyrosine hydroxylase immunoreactivity in the cardiac conduction system and myocardium in Parkinson's disease: an anatomical study. *Acta Neuropathol.* **118**, 777–784 (2009).
55. Hopkins, D. A., Bieger, D., de Vente, J. & Steinbusch, H. W. M. Vagal efferent projections: viscerotropy, neurochemistry and effects of vagotomy. *Prog. Brain Res.* **107**, 79–96 (1996).
56. Goyal, R. K. & Hirano, I. The enteric nervous system. *N. Engl. J. Med.* **334**, 1106–1115 (1996).
57. Lack and relative lack of vagus nerve alpha-synuclein pathology in an autopsy series of 49 normal elderly and 18 with incidental Lewy body disease. American Association of Neuropathologists, Inc. Abstracts of the 93rd Annual Meeting June 8–11, 2017 Garden Grove, CA. *J. Neuropathol. Exp. Neurol.* **76**, Abstract 152 (2017). <https://academic.oup.com/jnen/article/76/6/491/3832872>.
58. Beach, T. G. et al. Vagus nerve and stomach synucleinopathy in Parkinson's disease, incidental Lewy body disease, and normal elderly subjects: Evidence against the "body-first" hypothesis. *J. Parkinsons Dis.* **11**, 1833–1843 (2021).
59. Gelpi, E. et al. Multiple organ involvement by alpha-synuclein pathology in Lewy body disorders. *Mov. Disord.* **29**, 1010–1018 (2014).
60. Shannon, K. M., Keshavarzian, A., Dodiya, H. B., Jakate, S. & Kordower, J. H. Is alpha-synuclein in the colon a biomarker for premotor 19. Parkinson's disease? Evidence from 3 cases. *Mov. Disord.* **27**, 716–719 (2012).
61. Ito, S. et al. Alpha-synuclein immunohistochemistry of gastrointestinal and biliary surgical specimens for diagnosis of Lewy body disease. *Int. J. Clin. Exp. Pathol.* **15**, 1714–1723 (2014).
62. Borghammer, P. How does Parkinson's disease begin? Perspectives on neuroanatomical pathways, prions, and histology. *Mov. Disord.* **33**, 48–57 (2018).
63. Borghammer, P. & Van Den Berge, N. Brain-first versus gut-first Parkinson's disease: a hypothesis. *J. Parkinsons Dis.* **9**, S281–S295 (2019).
64. Del Tredici, K. & Braak, H. A not entirely benign procedure: progression of Parkinson's disease. *Acta Neuropathol.* **115**, 379–384 (2008).
65. Phillips, R. J., Walter, G. C., Wilder, S. L., Baronowsky, E. A. & Powley, T. L. Alpha-synuclein-immunopositive myenteric neurons and vagal preganglionic terminals: autonomic pathway implicated in Parkinson's disease? *Neuroscience* **153**, 733–750 (2008).
66. Chandra, R., Hiniker, A., Kuo, Y. M., Nussbaum, R. L. & Liddle, R. A. α -Synuclein in gut endocrine cells and its implications for Parkinson's disease. *JCI Insight* **2**, e92295 (2017).
67. Desplats, P. et al. Inclusion formation and neuronal cell death through neuron-to-neuron transmission of alpha-synuclein. *Proc. Natl. Acad. Sci. USA* **106**, 13010–13015 (2009).
68. George, S., Rey, N. L., Reichenbach, N., Steiner, J. A. & Brundin, P. α -Synuclein: the long distance runner. *Brain Pathol.* **23**, 350–357 (2013).
69. Goedert, M., Masuda-Suzukake, M. & Falcon, B. Like prions: the propagation of aggregated tau and α -synuclein in neurodegeneration. *Brain* **140**, 266–278 (2017).
70. Fenyi, A. et al. Seeding propensity and characteristics of pathogenic α Syn assemblies in formalin-fixed human tissue from the enteric nervous system, olfactory bulb, and brainstem in cases staged for Parkinson's disease. *Cells* **10**, 139 (2021).
71. Recasens, A. et al. Lewy body extracts from Parkinson disease brains trigger α -synuclein pathology in neurodegeneration in mice and monkeys. *Ann. Neurol.* **75**, 351–362 (2014).
72. Chen, F. et al. α -Synuclein aggregation in the olfactory bulb induces olfactory deficits by perturbing granule cells and granular-mitral synaptic transmission. *NPJ Parkinsons Dis.* **7**, 144 (2021).
73. Ulusoy, A. et al. Caudo-rostral brain spreading of α -synuclein through vagal connections. *EMBO Mol. Med.* **5**, 1119–1127 (2013).
74. Ulusoy, A. et al. Brain-to-stomach transfer of α -synuclein via vagal preganglionic projections. *Acta Neuropathol.* **133**, 381–393 (2017).
75. Van Den Berge, N. et al. Evidence for bidirectional and trans-synaptic parasympathetic and sympathetic propagation of alpha-synuclein in rats. *Acta Neuropathol.* **138**, 535–550 (2019).
76. Holmqvist, S. et al. Melki R. Direct evidence of Parkinson pathology spread from the gastrointestinal tract to the brain in rats. *Acta Neuropathol.* **128**, 805–820 (2014).
77. Jackson, R. G. Anatomy of the vagus nerves in the region of the lower esophagus and the stomach. *Anat. Rec.* **103**, 1–18 (1949).
78. Travaglini, R. A., Hermann, G. E., Browning, K. N. & Rogers, R. C. Brainstem circuits regulating gastric function. *Ann. Rev. Physiol.* **68**, 279–305 (2006).
79. Braak, H. & Del Tredici, K. Invited article: Nervous system pathology in sporadic Parkinson's disease. *Neurology* **70**, 1916–1925 (2008).
80. Braak, H. & Del Tredici, K. Potential pathways of abnormal tau and α -synuclein dissemination in sporadic Alzheimer's and Parkinson's diseases. *Cold Spring Harb. Perspect. Biol.* **8**, pii: a023630 (2016).
81. Del Tredici, K. & Braak, H. Sporadic Parkinson's disease: development and distribution of α -synuclein pathology. *Neuropathol. Appl. Neurobiol.* **42**, 33–50 (2016).
82. Orimo, S., Ghebremedhin, E. & Gelpi, E. Peripheral and central autonomic nervous system: does the sympathetic or parasympathetic nervous system bear the brunt of the pathology during the course of sporadic PD? *Cell Tiss. Res.* **373**, 267–286 (2018).
83. Breen, D. P., Halliday, G. M. & Lang, A. E. Gut-brain axis and the spread of α -synuclein pathology: vagal highway or dead end? *Mov. Disord.* **34**, 307–316 (2019).
84. Horsager, J. et al. Brain-first versus body-first Parkinson's disease: a multimodal imaging case-control study. *Brain* **143**, 3077–3088 (2020).
85. Boeve, B. F. et al. Pathophysiology of REM sleep behavior disorder and relevance to neurodegenerative disease. *Brain* **130**, 2770–2788 (2007).
86. Boeve, B. F. et al. Clinicopathologic correlations in 172 cases of rapid eye movement sleep behavior disorder with or without a coexisting neurodegenerative disorder. *Sleep. Med.* **14**, 754–762 (2013).
87. Iranzo, A., Toloso, E. & Gelpi, E. Neurodegenerative disease status and post-mortem pathology in idiopathic rapid-eye-movement sleep behavior disorder: an observational cohort study. *Lancet Neurol.* **12**, 443–453 (2013).
88. Sprenger, F. S. et al. Enteric nervous system α -synuclein immunoreactivity in idiopathic REM sleep behavior disorder. *Neurology* **85**, 1761–1768 (2015).

89. Ehrminger, M. et al. The coeruleus/subcoeruleus complex in idiopathic rapid eye movement sleep behaviour disorder. *Brain* **139**, 1180–1188 (2016).
90. Vilas, D. et al. Assessment of α -synuclein in submandibular glands of patients with idiopathic rapid-eye-movement sleep behaviour disorder: a case-control study. *Lancet Neurol.* **15**, 708–718 (2016).
91. Knudsen, K. et al. In-vivo staging of pathology in REM sleep behaviour disorder: a multimodality imaging case-control study. *Lancet Neurol.* **17**, 618–628 (2018).
92. Rees, R. N., Noyce, A. J. & Schrag, A. The prodromes of Parkinson's disease. *Eur. J. Neurosci.* **49**, 320–327 (2019).
93. van de Berg, W. D. et al. Patterns of alpha-synuclein pathology in incidental cases and clinical subtypes of Parkinson's disease. *Parkinsonism Relat. Disord.* **18**, S28–S30 (2012).
94. Coughlin, D. G. et al. Most cases with Lewy pathology in a population-based cohort adhere to the Braak progression pattern but 'failure to fit' is highly dependent on staging system applied. *Parkinsonism Relat. Disord.* **64**, 124–131 (2019).
95. Jellinger, K. Is Braak staging valid for all types of Parkinson's disease. *J. Neural Transm.* **126**, 423–431 (2019).
96. Kalf, J., de Swart, B. J., Bloem, B. R. & Munneke, M. Prevalence of oropharyngeal dysphagia in Parkinson's disease: A meta-analysis. *Parkinsonism Relat. Disord.* **18**, 311–315 (2012).
97. Bushmann, M., Dobmeyer, S. M., Leeker, L. & Perlmutter, J. S. Swallowing abnormalities and their responses to treatment in Parkinson's disease. *Neurology* **39**, 1309–1314 (1989).
98. Bird, M., Woodward, M. C., Gibson, E. M., Phyland, D. J. & Fonda, D. Asymptomatic swallowing disorders in elderly patients with Parkinson's disease: A description of findings on clinical examination and videofluoroscopy in sixteen patients. *Age Ageing* **23**, 251–254 (1994).
99. Hammer, M. J., Murphy, C. A. & Abrams, T. M. Airway somatosensory deficits and dysphagia in Parkinson's disease. *J. Parkinson Dis.* **3**, 39–44 (2013).
100. Pflug, C. Critical dysphagia is common in Parkinson disease and occurs even in early stages: A prospective cohort study. *Dysphagia* **33**, 41–50 (2018).
101. Patel, B. et al. A comprehensive review of the diagnosis and treatment of Parkinson's disease dysphagia and aspiration. *Exp. Rev. Gastroenterol. Hepatol.* **14**, 411–424 (2020).
102. Burgos, R. et al. ESPEN guidelines clinical nutrition in neurology. *Clin. Nutr.* **37**, 354–396 (2018).
103. Mu, L. et al. Altered pharyngeal muscles in Parkinson disease. *J. Neuropathol. Exp. Neurol.* **71**, 520–530 (2012).
104. Mu, L. et al. Alpha-synuclein pathology and axonal degeneration of the peripheral motor nerves innervating pharyngeal muscles in Parkinson's disease. *J. Neuropathol. Exp. Neurol.* **72**, 119–129 (2013).
105. Mu, L. et al. Parkinson's disease affects peripheral sensory nerves in the pharynx. *J. Neuropathol. Exp. Neurol.* **72**, 614–623–38 (2013).
106. Schröder, J. B. et al. Substance P saliva reduction predicts pharyngeal dysphagia in Parkinson's disease. *Front. Neurol.* **10**, 386 (2019).
107. Troche, M. S., Brandimore, A. E., Okun, M. S., Davenport, P. W. & Hegland, K. W. Decreased cough sensitivity and aspiration in Parkinson disease. *Chest* **146**, 1294–1299 (2014).
108. Warnecke, T., Dziewas, R. & Langmore, S. Neurogenic dysphagia. Originally published in German: Neurogene Dysphagien: Diagnostik und Therapie (W. Kohlhammer Verlag, Stuttgart, 2013; 2nd extended and rev. ed. 2018) (Springer, Cham, 2021).
109. Warnecke, T. et al. Neurogenic dysphagia: A systematic review and proposal of a classification system. *Neurology* **96**, e876–e889 (2021).
110. Labeit, B. et al. Oropharyngeal freezing and its relation to dysphagia—An analogy to freezing of gait. *Parkinsonism Relat. Disord.* **75**, 1–6 (2020).
111. Troche, M. S., Okun, M. S., Rosenbek, J. C., Altmann, L. J. & Sapienza, C. M. Attentional resource allocation and swallowing safety in Parkinson's disease: a dual task study. *Parkinsonism Relat. Disord.* **20**, 439–443 (2014).
112. Labeit, B. et al. Effect of cognitive and motor dual-task on oropharyngeal swallowing in Parkinson's disease. *Eur. J. Neurol.* **28**, 754–762 (2021).
113. Fukae, J. et al. Impact of residual drug in the pharynx on the delayed-on phenomenon in Parkinson's disease patients. *Mov. Disord. Clin. Pr.* **7**, 273–278 (2020).
114. Buhmann, C. et al. Pill swallowing in Parkinson's disease: a prospective study based on flexible endoscopic evaluation of swallowing. *Parkinsonism Relat. Disord.* **62**, 51–56 (2019).
115. Lam, K. et al. Simple clinical tests may predict severe oropharyngeal dysphagia in Parkinson's disease. *Mov. Disord.* **22**, 640–644 (2007).
116. Cereda, E. et al. Swallowing disturbances in Parkinson's disease: a multivariate analysis of contributing factors. *Parkinsonism Relat. Disord.* **20**, 1382–1387 (2014).
117. Cersósimo, M. G. et al. Hyposialorrhea as an early manifestation of Parkinson disease. *Auton. Neurosci.* **150**, 150–151 (2009).
118. Nienstedt, J. C. et al. Drooling is no early sign of dysphagia in Parkinson's disease. *Neurogastroenterol. Motil.* **30**, e13259 (2018).
119. Nóbrega, A. C., Rodrigues, B. & Melo, A. Silent aspiration in Parkinson's disease patients with diurnal sialorrhoea. *Clin. Neurol. Neurosurg.* **110**, 117–119 (2008).
120. Manor, Y., Giladi, N., Cohen, A., Fliss, D. M. & Cohen, J. T. Validation of a swallowing disturbance questionnaire for detecting dysphagia in patients with Parkinson's disease. *Mov. Disord.* **22**, 1917–1921 (2007).
121. Kalf, J. G. et al. Reproducibility and validity of patient-related assessment of speech, swallowing, and saliva control in Parkinson's disease. *Arch. Phys. Med Rehabil.* **92**, 1152–1158 (2011).
122. Simons, J. A. et al. Development and validation of a new screening questionnaire for dysphagia in early stages of Parkinson's disease. *Parkinsonism Relat. Disord.* **20**, 992–998 (2014).
123. Buhmann, C. et al. Is the Munich dysphagia Test—Parkinson's disease (MDT-PD) a valid screening tool for patients at risk of aspiration? *Parkinsonism Relat. Disord.* **61**, 138–143 (2019).
124. Simons, J. A. et al. Multilingual validation of the first French version of Munich Dysphagia Test—Parkinson's disease (MDT-PD) in the Luxembourg Parkinson's study. *Front Neurol.* **10**, 1180 (2019).
125. Chaudhuri, K. R., Healy, D. G. & Schapira, A. H., National Institute for Clinical Excellence. Non-motor symptoms of Parkinson's disease: diagnosis and management. *Lancet Neurol.* **5**, 235–245 (2006).
126. Goetz, C. G. et al. Movement Disorder Society-sponsored revision of the Unified Parkinson's disease Rating Scale (MDS-UPDRS): scale presentation and clinimetric testing results. *Mov. Disord.* **23**, 2129–2170 (2008).
127. Nienstedt, J. et al. Predictive clinical factors for penetration and aspiration in Parkinson's disease. *Neurogastroenterol. Motil.* **31**, e13524 (2019).
128. Sampaio, M., Argolo, N., Melo, A. & Nóbrega, A. C. Wet voice as a sign of penetration/aspiration in Parkinson's disease: does testing material matter? *Dysphagia* **29**, 610–615 (2014).
129. Kalf, H. et al. Guidelines for speech-language therapy in Parkinson's disease. *Nijmegen, the Netherlands/Miami (FL)*. (ParkinsonNet/NPF, U.S.A., 2011). <https://www.parkinsonnet.com/discipline/speech-and-language>.
130. Pflug, C., Niessen, A., Buhmann, C. & Bihler, M. Swallowing speed is no adequate predictor of aspiration in Parkinson's disease. *Neurogastroenterol. Motil.* **31**, e13713 (2019).
131. Logemann, J. A. *Evaluation and treatment of swallowing disorders*. 2nd ed. (Pro-Ed, Austin, 1998).
132. Langmore, S. E. Evaluation of oropharyngeal dysphagia: which diagnostic tool is superior? *Curr. Opin. Otolaryngol. Head. Neck Surg.* **11**, 485–489 (2003).
133. Dziewas, R. et al. German guideline for neurogenic dysphagia (2020) https://dgn.org/wp-content/uploads/2013/01/030111_LL_Neurogene_Dysphagie_2020.pdf.
134. Ciucci, M. R. et al. Early identification and treatment of communication and swallowing deficits in Parkinson disease. *Semin Speech Lang.* **34**, 185–202 (2013).
135. Van Hooren, M. R. A. et al. Treatment effects for dysphagia in Parkinson's disease: a systematic review. *Parkinsonism Relat. Disord.* **20**, 800–807 (2014).
136. Melo, A. & Monteiro, L. Swallowing improvement after levodopa treatment in idiopathic Parkinson's disease: lack of evidence. *Parkinsonism Relat. Disord.* **19**, 279–281 (2013).
137. Sutton, J. Dysphagia in Parkinson's disease is responsive to levodopa. *Parkinsonism Relat. Disord.* **19**, 282–284 (2013).
138. Chang, M. C., Park, J. S., Lee, B. J. & Park, D. Effectiveness of pharmacologic treatment for dysphagia in Parkinson's disease: a narrative review. *Neurological Sci.* **42**, 513–519 (2021).
139. Monte, F., da Silva-Júnior, F. P., Braga-Neto, P., Nobre e Souza, M. A. & de Bruin, V. M. Swallowing abnormalities and dyskinesia in Parkinson's disease. *Mov. Disord.* **20**, 457–462 (2005).
140. Warnecke, T. et al. Levodopa responsiveness of dysphagia in advanced Parkinson's disease and reliability testing of the FEES-Levodopa-test. *Parkinsonism Relat. Disord.* **28**, 100–106 (2016).
141. Hirano, M. et al. Rotigotine transdermal patch improves swallowing in dysphagic patients with Parkinson's disease. *Dysphagia* **30**, 452–456 (2015).
142. Hirano, M. et al. Effects of the rotigotine transdermal patch versus oral levodopa on swallowing in patients with Parkinson's disease. *J. Neurol. Sci.* **15**, 404–405 (2019).
143. Torti, M., Bravi, D., Vacca, L. & Stocchi, F. Are all dopamine agonists essentially the same? *Drugs* **79**, 693–703 (2019).
144. Labeit, B. et al. Effect of intestinal levodopa-carbidopa infusion on pharyngeal dysphagia: results from a retrospective levodopa-carbidopa infusion on pharyngeal dysphagia: results from a retrospective levodopa-carbidopa infusion on pharyngeal dysphagia: results from a retrospective pilot study in patients with Parkinson's disease. *Parkinsons Dis.* **11**, 4260501 (2020).
145. Yu, H., Takahashi, K., Bloom, L., Quaynor, S. D. & Xie, T. Effect on Deep Brain Stimulation on swallowing function: A systematic review. *Front Neurol.* **11**, 547 (2020).

146. Chang, M. C., Park, J. S., Lee, B. J., Park, D. The effect of deep brain stimulation on swallowing function in Parkinson's disease: A narrative review. *Dysphagia*. <https://doi.org/10.1007/s00455-020-10214-y> (2021).
147. Xie, T. et al. Low-frequency stimulation of STN-DBS reduces aspiration and freezing of gait in patients with PD. *Neurology* **84**, 415–420 (2015).
148. Xie, T. et al. Long-term effect of low frequency stimulation of STN on dysphagia, freezing of gait and other motor symptoms in PD. *J. Neurol. Neurosurg. Psychiatry* **89**, 989–994 (2018).
149. Pflug, C. et al. Impact of simultaneous subthalamic and nigral stimulation on dysphagia in Parkinson's disease. *Ann. Clin. Transl. Neurol.* **7**, 628–638 (2020).
150. Heijnen, B. J., Speyer, R., Baijens, L. W. & Bogaardt, H. C. Neuromuscular electrical stimulation versus traditional therapy in patients with Parkinson's disease and oropharyngeal dysphagia: effects on quality of life. *Dysphagia* **27**, 336–345 (2012).
151. Baijens, L. W. et al. Surface electrical stimulation in dysphagic Parkinson patients: a randomized clinical trial. *Laryngoscope* **123**, E38–E44 (2013).
152. Park, J. S., Oh, D. H., Hwang, N. K. & Lee, J. H. Effects of neuromuscular electrical stimulation in patients with Parkinson's disease and dysphagia: A randomized, single-blind, placebo-controlled trial. *NeuroRehab* **42**, 457–463 (2018).
153. Logemann, J. A. et al. A randomized study of three interventions for aspiration of thin liquids in patients with dementia or Parkinson's disease. *J. Speech Lang. Hear Res.* **51**, 173–183 (2008).
154. Ayres, A. L., Jotz, G. P., Rieder, C. R. & Olchik, M. R. Benefit from the chin-down maneuver in the swallowing performance and self-perception of Parkinson's disease patients. *Parkinsons Dis.* **2017**, e7460343 (2017).
155. Sharkawi, A. E. et al. Swallowing and voice effects on Lee Silverman Voice. *J. Neurol. Neurosurg. Psychiatry* **72**, 31–37 (2002).
156. Troche, M. S. et al. Aspiration and swallowing in Parkinson's disease and rehabilitation with EMST: a randomized trial. *Neurology* **75**, 1912–1919 (2010).
157. Claus, I. et al. Expiratory muscle strength training for therapy of pharyngeal dysphagia in Parkinson's disease. *Mov. Disord.* **36**, 1815–1824 (2021).
158. Manor, Y., Mootanah, R., Freud, D., Giladi, N. & Cohen, J. T. Video-assisted swallowing therapy for patients with Parkinson's disease. *Parkinsonism Relat. Disord.* **19**, 207–211 (2013).
159. Athukorala, R. P., Jones, R. D., Sella, O. & Huckabee, M. L. Skill training for swallowing rehabilitation in patients with Parkinson's disease. *Arch. Phys. Med. Rehabil.* **95**, 1374–1382 (2014).
160. Jost, W. H. et al. SIAxI: Placebo-controlled, randomized, double-blind study of incobotulinumtoxin A for sialorrhea. *Neurology* **92**, e1982–e1991 (2019).
161. Arbouw, M. E. L. et al. Glycopyrrolate for sialorrhea in Parkinson's disease: a randomized, double-blind, crossover trial. *Neurology* **74**, 1203–1207 (2010).
162. South, A. R., Somers, S. M. & Jog, M. S. Gum chewing improves swallow frequency and latency in Parkinson patients: a preliminary study. *Neurology* **74**, 1198–1202 (2010).
163. Marrinan, S., Emmanuel, A. V. & Burn, D. J. Delayed gastric emptying in Parkinson's disease. *Mov. Disord.* **29**, 23–32 (2014).
164. Mukherjee, A., Biswas, A. & Das, S. K. Gut dysfunction in Parkinson's disease. *World J. Gastroenterol.* **22**, 5742–5752 (2016).
165. Doi, H. et al. Plasma levodopa peak delay and impaired gastric emptying in Parkinson's disease. *J. Neurol. Sci.* **319**, 86–88 (2012).
166. Sung, H. Y. et al. The prevalence and patterns of pharyngo-esophageal dysmotility in patients with early stage Parkinson's disease. *Mov. Disord.* **25**, 2361–2368 (2010).
167. Suttrup, I. et al. Esophageal dysfunction in different stages of Parkinson's disease. *Neurogastroenterol. Motil.* **29**, e12915 (2017).
168. Pasricha, P. J. & Parkman, H. P. Gastroparesis: definitions and diagnosis. *Gastroenterol. Clin. North Am.* **44**, 1–7 (2015).
169. Kahrilas, P. J. et al. Smout AJ, The Chicago classification of esophageal motility disorders, v3.0. *Neurogastroenterol. Motil.* **27**, 160–174 (2015).
170. Triadafilopoulos, G., Gandhi, R. & Barlow, C. Pilot cohort study of endoscopic botulinum neurotoxin injection in Parkinson's disease. *Parkinsonism Relat. Disord.* **44**, 33–37 (2017).
171. Derrey, S. et al. Impact of deep brain stimulation on pharyngo-esophageal motility: a randomized cross-over study. *Neurogastroenterol. Motil.* **27**, 1214–1222 (2015).
172. Alvarez-Berdugo, D. et al. A comparative study on the therapeutic effect of TRPV1, TRPA1, and TRPM8 agonists on swallowing dysfunction associated with aging and neurological diseases. *Neurogastroenterol. Motil.* **30**, e13185 (2018).
173. Suntrup-Krueger, S. et al. Effect of capsaicinoids on neurophysiological, biochemical and mechanical parameters of swallowing function. *Neurotherapeutics* **18**, 1360–1370 (2021).
174. Heetun, Z. S. & Quigley, E. M. Gastroparesis and Parkinson's disease: a systematic review. *Parkinsonism Relat. Disord.* **18**, 433–440 (2012).
175. Gil, R. A., Hwynn, N., Fabian, T., Joseph, S. & Fernandez, H. H. Botulinum toxin type A for the treatment of gastroparesis in Parkinson's disease patients. *Parkinsonism Relat. Disord.* **17**, 285–287 (2011).
176. Arai, F. et al. Subthalamic deep brain stimulation can improve gastric emptying in Parkinson's disease. *Brain* **135**, 1478–1485 (2012).
177. Parkinson, J. *An essay on the shaking palsy*. (Sherwood, Neely, and Jones, London, 1817).
178. Jost, W. H. An update on the recognition and treatment of autonomic symptoms in Parkinson's disease. *Exp. Rev. Neurother.* **17**, 791–799 (2017).
179. Adler, C. H. & Beach, T. G. Neuropathological basis of nonmotor manifestations of Parkinson's disease. *Mov. Disord.* **31**, 1114–1119 (2016).
180. Knudsen, K., Krogh, K., Østergaard, K. & Borghammer, P. Constipation in Parkinson's disease: subjective symptoms, objective markers, and new perspectives. *Mov. Disord.* **32**, 94–105 (2017).
181. Parkinson Study Group. A randomized trial of relamorelin for constipation in Parkinson's disease (MOVE-PD): Trial results and lessons learned. *Parkinsonism Relat. Disord.* **37**, 101–105 (2017).
182. Jost, W. H. Autonome Regulationsstörungen beim Parkinson Syndrom [Disorders of autonomic regulation in Parkinson syndrome]. *Fortschr. Neurol. Psychiatr.* **63**, 194–205 (1995).
183. Barone, P. et al. The PRIAMO study: A multicenter assessment of nonmotor symptoms and their impact on quality of life in Parkinson's disease. *Mov. Disord.* **24**, 1641–1649 (2009).
184. Abbott, R. D. et al. Frequency of bowel movements and the future risk of Parkinson's disease. *Neurology* **57**, 456–464 (2001).
185. Stirpe, P., Hoffman, M., Badiali, D. & Colosimo, C. Constipation: an emerging risk factor for Parkinson's disease? *Eur. J. Neurol.* **23**, 1606–1613 (2016).
186. Svensson, E., Henderson, V. W., Borghammer, P., Horváth-Puhó, E. & Sørensen, H. T. Constipation and risk of Parkinson's disease: A Danish population-based cohort study. *Parkinsonism Relat. Disord.* **28**, 18–22 (2016).
187. Pierantozzi, M. et al. Helicobacter pylori eradication and l-dopa absorption in patients with PD and motor fluctuations. *Neurology* **66**, 1824–1829 (2006).
188. Savica, R. et al. Medical records documentation of constipation preceding Parkinson disease: A case-control study. *Neurology* **73**, 1752–1758 (2009).
189. Gage, H. et al. Correlates of constipation in people with Parkinson's. *Parkinsonism Relat. Disord.* **17**, 106–111 (2011).
190. Camacho, M. et al. Early constipation predicts faster dementia onset in Parkinson's disease. *NPJ Parkinsons Dis.* **7**, 45 (2021).
191. García Roca, L. et al. COPPADIS Study Group. Constipation predicts cognitive decline in Parkinson's disease: results from the COPPADIS Cohort at 2-year follow-up and comparison with a control group. *J. Parkinsons Dis.* <https://doi.org/10.3233/JPD-212868> (2021).
192. Kenna, J. E. et al. Characterization of gastrointestinal symptom type and severity in Parkinson's disease: A case-control study in an Australian cohort. *Mov. Disord. Clin. Pr.* **8**, 245–253 (2021).
193. Lundin, E. et al. Segmental colonic transit studies: Comparison of a radiological and a scintigraphic method. *Colorectal Dis.* **9**, 344–351 (2007).
194. Agachan, F., Pfeifer, J. & Wexner, S. D. Defecography and proctography. Results of 744 patients. *Dis. Colon Rectum* **39**, 899–905 (1996).
195. Jost, W. H., Schrank, B., Herold, A. & Leib, O. Functional outlet obstruction: Anismus, spastic pelvic floor syndrome, and dyscoordination of the voluntary sphincter muscles. *Scand. J. Gastroenterol.* **34**, 449–453 (1999).
196. Barichella, M. et al. Probiotics and prebiotic fiber for constipation associated with PD. *Neurology* **87**, 1274–1280 (2016).
197. Zangaglia, R. et al. Macrogol for the treatment of constipation in Parkinson's disease. A randomized placebo-controlled study. *Mov. Disord.* **22**, 1239–1244 (2007).
198. Zesiewicz, T. A. et al. Practice parameter: treatment of nonmotor symptoms of Parkinson disease: report of the quality standards subcommittee of the American Academy of Neurology. *Neurology* **74**, 924–931 (2010).
199. Tan, A. H. et al. Probiotics for constipation in Parkinson disease: A randomized placebo-controlled study. *Neurology* **96**, e772–e782 (2021).
200. Liu, Z. et al. Mosapride citrate, a novel 5-HT₄ agonist and partial 5-HT₃ antagonist, ameliorates constipation in parkinsonian patients. *Mov. Disord.* **20**, 680–686 (2005).
201. Freitas, M. E., Alqaraawi, A., Lang, A. E. & Liu, L. W. C. Linaclotide and prucalopride for management of constipation in patients with parkinsonism. *Mov. Disord. Clin. Pr.* **5**, 218–220 (2018).
202. Bassotti, G., Usai Satta, P. & Bellini, M. Plecanatide for the treatment of chronic idiopathic constipation in adult patients. *Expert Rev. Clin. Pharm.* **12**, 1019–1026 (2019).
203. Ondo, W. G. et al. Placebo-controlled trial of lubiprostone for constipation associated with Parkinson disease. *Neurology* **78**, 1650–1654 (2012).

204. Bedarf, J. R. et al. Functional implications of microbial and viral gut metagenome changes in early stage L-DOPA-naïve Parkinson's disease patients. *Genome Med* **9**, 39 (2017).
205. Pietrucci, D. et al. Dysbiosis of gut microbiota in a selected population of Parkinson's patients. *Parkinsonism Relat. Disord.* **65**, 124–130 (2019).
206. Weis, S. et al. Effect of Parkinson's disease and related medications on the composition of the fecal bacterial microbiota. *NPJ Parkinsons Dis.* **5**, 28 (2019).
207. Chapelet, G., Leclair-Visonneau, L., Clairembault, T., Neunlist, M. & Derkinderen, P. Can the gut be the missing piece in uncovering PD pathogenesis? *Parkinsonism Relat. Disord.* **59**, 26–31 (2020).
208. Cryan, J. F., O'Riordan, K. J., Sandhu, K., Peterson, V. & Dinan, T. G. The gut microbiome in neurological disorders. *Lancet Neurol.* **19**, 179–194 (2020).
209. Metta, V. et al. Gastrointestinal dysfunction in Parkinson's disease: molecular pathology and implications of gut microbiome, probiotics, and fecal microbiota transplantation. *J. Neurol.* <https://doi.org/10.1007/s00415-021-10567-w> (2021).
210. Weis, S. et al. Association between Parkinson's disease and the faecal eukaryotic microbiota. *NPJ Parkinsons Dis.* **7**, 101 (2021).
211. Heintz-Buschart, A. et al. The nasal and gut microbiome in Parkinson's disease and idiopathic rapid eye movement sleep behavior disorder. *Mov. Disord.* **33**, 88–98 (2018).
212. Does the truth lie within the gut? Investigating the gut microbiome in an Australian cohort of Parkinson's disease patients. American Academy of Neurology, Abstracts of the 72nd Annual Meeting April 25–May 1, 2020 Toronto, Ontario. *Neurology* **94**, Abstract 1438, https://n.neurology.org/content/94/15_Supplement/1438 (2020).
213. Devos, D. et al. Colonic inflammation in Parkinson's disease. *Neurobiol. Dis.* **50**, 42–48 (2013).
214. Nielsen, S. D., Pearson, N. M. & Seidler, K. The link between the gut microbiota and Parkinson's disease: A systematic mechanism review with focus on α -synuclein transport. *Brain Res.* **1769**, 147609 (2021).
215. Hill-Burns, E. M. et al. Parkinson's disease and Parkinson's disease medications have distinct signatures of the gut microbiome. *Mov. Disord.* **32**, 739–749 (2017).
216. Forsyth, C. B. et al. Increased intestinal permeability correlates with sigmoid mucosa α -synuclein staining and endotoxin exposure markers in early Parkinson's disease. *PLoS One* **6**, e28032 (2011).
217. de Vos, W. M. & de Vos, E. A. Role of the intestinal microbiome in health and disease: from correlation to causation. *Nutr. Rev.* **70**, S45–S56 (2012).
218. Visanji, N. P., Brooks, P. L., Hazrati, L. N. & Lang, A. E. The prion hypothesis in Parkinson's disease: Braak to the future. *Acta Neuropathol. Commun.* **1**, 2 (2013).
219. Stolzenberg, E. et al. A role for neuronal α -synuclein in gastrointestinal immunity. *J. Innate Immun.* **9**, 456–463 (2017).
220. Sulzer, D. et al. T cells from patients with Parkinson's disease recognize α -synuclein peptides. *Nature* **546**, 656–661 (2017). Erratum: *Nature*. 549, 292 (2017).
221. van Kessel, S. P. et al. Gut bacterial tyrosine decarboxylases restrict levels of levodopa in the treatment of Parkinson's disease. *Nat. Commun.* **10**, 310 (2019).
222. van Kessel, S. P., Auvinen, P., Scheperjans, F. & El Aidi, S. Gut bacterial tyrosine decarboxylase associates with clinical variables in a longitudinal cohort study of Parkinson's disease. *NPJ Parkinsons Dis.* **7**, 115 (2021).
223. Keshavarzian, A., Engen, P., Bonvegna, S. & Cilia, R. The gut microbiome in Parkinson's disease: A culprit or a bystander? *Prog. Brain Res.* **252**, 357–450 (2020).
224. Seguela, L., Sarnelli, G. & Esposito, G. Leaky gut, dysbiosis, and enteric glia activation: the trilogy behind the intestinal origin of Parkinson's disease. *Neural Regen. Res.* **15**, 1037–1038 (2020).
225. Gorecki, A. M. et al. Single nucleotide polymorphisms associated with gut homeostasis influence risk and age-at-onset of Parkinson's disease. *Front. Aging Neurosci.* **12**, 603849 (2020).
226. Bedarf, J. R. et al. Das Darm-Mikrobiom bei der Parkinson-Krankheit [The gut microbiome in Parkinson's disease]. *Nervenarzt* **90**, 160–166 (2019).
227. Jackson, A. et al. Diet in Parkinson's disease: critical role for the microbiome. *Front. Neurol.* **10**, 1245 (2019).
228. Sampson, T. R. et al. Gut microbiota regulate motor deficits and neuroinflammation in a model of Parkinson's disease. *Cell* **167**, 1469–1480 (2017).
229. Vendrik, K. E. W. et al. Fecal microbiota transplantation in neurological disorders. *Front. Cell Infect. Microbiol.* **10**, 98 (2020).
230. Segal, A., Zlotnik, Y., Moyal-Atias, K., Abuhasira, R. & Ifergane, G. Fecal microbiota transplant as a potential treatment for Parkinson's disease—A case series. *Clin. Neurol. Neurosurg.* **207**, 106791 (2021).
231. Fecal Microbiota Transplantation for Parkinson's Disease. <https://clinicaltrials.gov/ct2/show/NCT03808389>. Accessed 31 Dec 2021.
232. Okino, Y. et al. Root of the small-bowel mesentery: correlative anatomy and CT features of pathologic conditions. *Radiographics* **21**, 1475–1490 (2001).

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COMPETING INTERESTS

K.-H.S., K.D.T., I.C., T.W., and W.J. declare no current or potential conflicts of interest. I.C. has previously received honoraria from Abbvie, BIAL, STADAPHARM, Georg Thieme Verlag KG, and consultancies from STADAPHARM. T.W. is an advisory board member of AbbVie, UCB, Archimedes, Phogenesis, Zambon, Bial, and Kyowa; he has received honoraria for lectures from Bial, AbbVie, STADA, UCB, Biogen, Licher, Desitin, Pfizer, Zambon, Teva, and Bayer; he also has received grants (investigator-initiated) from UCB, Licher, Abbvie, as well as academic grants from the G-BA Innovation Fund, Deutsche Parkinson-Vereinigung, (dPV), IZKF, and Neuro NRW (Germany). W.J. is a speaker and advisor for Abbie, Bial, Desitin, Licher, Stada, UCB, and Zambon.

ADDITIONAL INFORMATION

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