REVIEW Open Access

Check for updates

Human placenta-derived amniotic epithelial cells as a new therapeutic hope for COVID-19-associated acute respiratory distress syndrome (ARDS) and systemic inflammation

(2022) 13:126

Amirhesam Babajani[†], Kasra Moeinabadi-Bidgoli[†], Farnaz Niknejad[†], Hamidreza Rismanchi[†], Sepehr Shafiee[†], Siavash Shariatzadeh[†], Elham Jamshidi, Mohammad Hadi Farjoo and Hassan Niknejad^{*}

Abstract

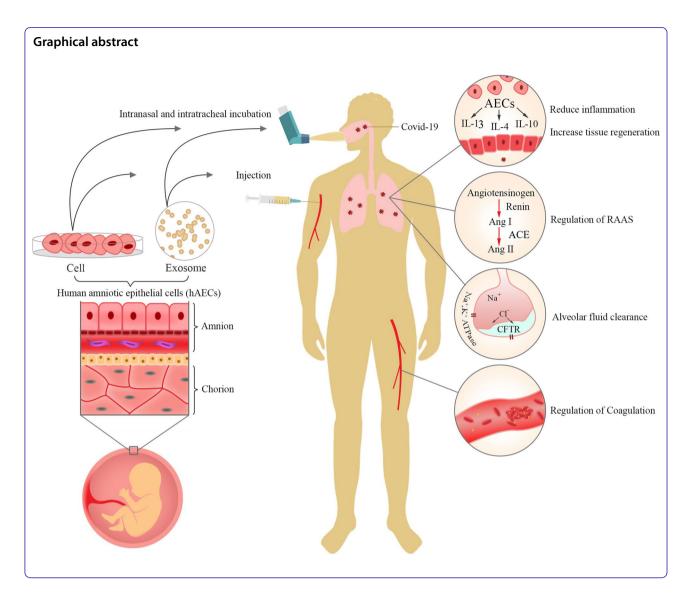
Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), which causes coronavirus disease 2019 (COVID-19), has become in the spotlight regarding the serious early and late complications, including acute respiratory distress syndrome (ARDS), systemic inflammation, multi-organ failure and death. Although many preventive and therapeutic approaches have been suggested for ameliorating complications of COVID-19, emerging new resistant viral variants has called the efficacy of current therapeutic approaches into question. Besides, recent reports on the late and chronic complications of COVID-19, including organ fibrosis, emphasize a need for a multi-aspect therapeutic method that could control various COVID-19 consequences. Human amniotic epithelial cells (hAECs), a group of placentaderived amniotic membrane resident stem cells, possess considerable therapeutic features that bring them up as a proposed therapeutic option for COVID-19. These cells display immunomodulatory effects in different organs that could reduce the adverse consequences of immune system hyper-reaction against SARS-CoV-2. Besides, hAECs would participate in alveolar fluid clearance, renin-angiotensin-aldosterone system regulation, and regeneration of damaged organs. hAECs could also prevent thrombotic events, which is a serious complication of COVID-19. This review focuses on the proposed early and late therapeutic mechanisms of hAECs and their exosomes to the injured organs. It also discusses the possible application of preconditioned and genetically modified hAECs as well as their promising role as a drug delivery system in COVID-19. Moreover, the recent advances in the pre-clinical and clinical application of hAECs and their exosomes as an optimistic therapeutic hope in COVID-19 have been reviewed.

Keywords: ARDS, COVID-19, Amniotic membrane, Epithelial stem cells, Immunomodulation, Regenerative medicine, SARS-CoV-2, Exosome

[†]Amirhesam Babajani, Kasra Moeinabadi-Bidgoli, Farnaz Niknejad, Hamidreza Rismanchi, Sepehr Shafiee and Siavash Shariatzadeh: These authors are alphabetically ordered co-first authors Department of Pharmacology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran



^{*}Correspondence: niknejad@sbmu.ac.ir



Introduction

Coronavirus disease 2019 (COVID-19), caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), has reached a new morbidity peak worldwide. New variants of SARS-CoV-2, including B.1.1.7 from the UK, 501Y.V2 and Omicron (B.1.1.529) from South Africa, and B.1.617 from India have made the spread and complication of COVID-19 more worrying [1-4]. So far, COVID-19 was symptom-free or showed flu-like syndrome in many patients and a minority of infected cases demonstrated severe pneumonia and acute respiratory distress syndrome (ARDS), the most severe complication of COVID-19 [5, 6]. However, studies have shown that new variants can enhance disease severity and increase the virulence of the virus [7, 8]. Despite the unclear pathophysiology of COVID-19, studies attribute the most complications to the uncontrolled immune system reaction. Previously, the term "cytokine storm" was used to describe maladapted induction of immune response in COVID-19. However, considering the lower amounts of some inflammatory mediators and dynamic alteration in the concentration of many cytokines, some researchers proposed alternative descriptions such as complex viral sepsis with immune deviation to describe the immune phenomenon in COVID-19 [9]. These suggestions mean that the exact pathophysiology and the possible future complications are still under question.

In order to reduce the severe complications and mortality of COVID-19, management of COVID-19 patients mainly consists of supportive care such as invasive mechanical ventilation and extracorporeal membrane oxygenation (ECMO), and there is still no proven curative drug, especially for susceptible patients [10]. Despite introducing various COVID-19 vaccines such as

Pfizer-BioNTech, Johnson & Johnson's Janssen, Sinopharm, Oxford–AstraZeneca, and Sputnik, the lower vaccine effectiveness has been reported for some variants, including β variant (B.1.351) and Omicron (B.1.1.529) [4, 11, 12]. Besides, many reports and hypotheses are released about the long-term consequences of COVID-19, including renal, cardiovascular, hepatic, and respiratory complications [13–16]. Since we are still at the early stages of the COVID-19 pandemic, vaccines' effect on these complications has not been evaluated yet.

Considering the current problems in COVID-19 management, there is still a need to develop practical therapeutic approaches that could reduce the early and late complications and mortality of COVID-19.

Human amniotic epithelial cells (hAECs) and their exosomes have been widely studied for various therapeutic effects on immune-related and degenerative diseases. These cells and exosomes display considerable healthgiving effects via immunomodulating properties on both immune cells and cytokine production. hAECs modulate immune cells such as lymphocytes, neutrophils, natural killer cells, and macrophages as primary sources of cytokine production during COVID-19 pathogenesis [17–19]. Additionally, hAECs release some types of angiotensin (Ang), such as Ang-(1-7), which confront destroying effects of SARS-CoV-2 on the renin-angiotensin-aldosterone system (RAAS), including lung fibrosis and pulmonary hypertension [20, 21]. As a severe complication of COVID-19, ARDS could be eliminated by hAECs-derived surfactants, which reduce air-liquid surface tension in damaged alveoli and improve patients' respiratory quality [22, 23]. Furthermore, studies have reported dysregulated coagulation such as deep vein thrombosis (DVT) and pulmonary thromboembolism (PTE) in COVID-19 [24, 25]. hAECs could control thrombotic events by releasing antithrombotic molecules such as perlecan, hyaluronic acid, and serpin F1 [26]. Besides, hAECs have shown promising regenerative potential in lung-related diseases, which could be used as a therapeutic approach in chronic complications of COVID-19 such as lung fibrosis [27, 28].

Within this context, comprehensive literature searches were performed using Scopus, PubMed, Medline, Embase, Clinicaltrials.gov, in order to identify publications written in English up to January 2022. Besides, the used keywords were stem cell, amnion, amniotic epithelial cell, exosome, regenerative medicine, cell therapy, immunomodulation, inflammation, lung fibrosis, reninangiotensin—aldosterone system, alveolar fluid, coagulation, coagulopathy, COVID-19, SARS-CoV-2, drug delivery, gene modification and immune system focusing on the biology of hAECs and the possible therapeutic role of hAECs in COVID-19. We merged founded records and removed the duplicates using EndNote X9 (Thomson

Reuters, New York, NY, USA). In order to increase the accuracy, two reviewers screened the publications by title, abstract and full texts to exclude unrelated records independently. Finally, three expert reviewers extracted data from all qualified studies.

Herein, we discuss the mechanisms involved in therapeutic features of human amniotic epithelial cells (hAECs) and their exosomes, the role of preconditioning, and gene modification of these cells to be used as a drug delivery system, and recent clinical trials in COVID-19.

Human amniotic epithelial cells

Human amniotic membrane contains two primary cell types, including human amniotic epithelial cells (hAECs), human amniotic mesenchymal stromal cells (hAMSCs), which can be isolated through appropriate methods [29]. Single-layer hAECs is located on a compact basement membrane adjacent to amniotic fluid. Beneath this structure is a stroma containing amniotic mesenchymal cells (AMCs) that secrete basement membrane contents such as collagen, laminin, and fibronectin. Amniotic epithelium originates prior to gastrulation, suggesting that some cell populations from this area could show pluripotency [30]. hAECs are differentiated from amnioblasts which originated directly from the epiblastic layer during the early gestational period. Unlike adult epithelial cell populations, it has been shown that they retain their stemness properties [31]. These multipotent cells can differentiate into mesodermal, ectodermal, and endodermal lineage cells [32]. It has been observed that hAECs express stem cell pluripotency markers such as octamer-binding-protein-4 (OCT-4), fibroblast growth factor (FGF)-4, tumor rejection antigens (TRA) 1-60, TRA-1-81, stage-specific embryonic antigen (SSEA)-3, and SSEA-4, as well as high rates of Homeobox protein NANOG that helps embryonic stem cells maintain their pluripotency [33]. However, it has been shown that they are a heterogeneous population consisting of mature, multipotent, pluripotent, and progenitor cells that express different grades of pluripotency markers [34]. One of the main reasons for this heterogeneity is that the hAECs lose some of their stemness during the gestational period [35]. For instance, it has been shown that during 16–17 weeks of gestation, a higher percentage of hAECs express pluripotency markers such as NANOG, OCT4, and SOX-2 compared to the epithelial cells isolated from the term placenta [36]. Despite their differentiation and proliferation potency, hAECs do not present the properties of teratogenicity that have been observed in the other type of stem cells [37, 38]. It seems that the lack of telomerase activity and expression of tumor rejection antigens (TRA) in hAECs can lead to tumor suppression [38].

There are many studies on the isolation of hAECs after elective Cesarean section. The isolation methods

are mainly based on peeling of chorion from amnion and separation of hAECs from the underlying basement membranes via enzymatic digestion by enzymatic solutions such as trypsin/EDTA and dispase II, and some detergents [39, 40]. Although some other cell types can be presented in the final isolated cells, studies have shown that the final isolated cell population mainly consists of hAECs under standard isolation [40, 41]. The identification of epithelial cells during the culture is based on morphologic characteristics and expression of epithelial and pluripotency markers. The epithelial cells can be flat, columnar, and cuboidal, while the mesenchymal cells are spindle-shaped [34]. Also, they express epithelial markers such as CK 1-8 and E-Cadherin [34]. After the isolation, density separation techniques can be used for identifying and proliferating hAECs with higher expression of stem cell markers such as SSEA-4 [42]. Other Stem cell markers such as TRA-1-60, TRA-1-81, and SSEA-3 have also been used for identifying hAECs with higher grades of stemness [43-45].

After the isolation, an average of 100 million cells of amniotic epithelium could be obtained from each amnion (about 100 million cells). After six passages, this cell number could even be increased to 10–60 billion by adding epidermal growth factor (EGF), which acts as a mitogen and enhances the proliferation of hAECs [46]. Besides, isolating the same cell numbers takes about four hours for hAECs, while it takes approximately 4–6 weeks for AMCs [47]. Based on the current studies, the appropriate cell number for intravenous (IV) administration of stem cells in treating COVID-19 patients is about one million cells per kilogram of weight; thus, the cells isolated from each placenta without any further passage could be enough for nearly 100 infected patients in one administration dose [48, 49].

They also affect host tissue cells by paracrine effect and consequently promote regeneration. hAECs have the immune privilege potency due to low expression of human leukocyte antigens such as HLA-A, HLA-B, HLA-C, HLA-DR while presenting immunomodulatory HLA-G [50, 51]. This feature reduces the chance of immune rejection in the human body. The other property which makes hAECs a candidate for cell therapy is their anti-inflammation feature. They increase the expression of interleukin (IL)10, a potent anti-inflammatory cytokine, and matrix metalloproteinase (MMP) 9, which has an anti-fibrotic effect. They also decrease PDGF secretion from macrophages M2, suppressing the immune response [52]. hAECs can reduce inflammation and the following fibrosis by secretion of TGF-β3 and suppression of TGF-β1. The TGF-β group includes three subtypes, TGF-β1 and TGF-β2, which have proinflammatory and fibrosis effects, and TGF-β3 with reverse potential. TGF- β 3 mainly has anti-inflammatory and anti-fibrotic roles by antagonizing TGF- β 1 [50]. Accumulating pieces of evidence have suggested that hAECs can provide a desirable microenvironment for tissue recovery by releasing bioactive cytokines and extracellular vesicles [53, 54]. It has been shown that hAECs can release antifibrotic, proangiogenic, and anti-inflammatory factors such as vascular endothelial growth factor (VEGF), angiogenin (AGN), and insulin-like growth factor (IGF) [55].

hAECs secreted extracellular vesicles (EVs) contain various bioactive molecules that can affect the microenvironment for inducing different therapeutic effects [56]. Utilizing exosomes instead of cells offers attractive benefits, such as calculating dosage and potency, storage capability, more stability, and an increased chance of crossing the blood-brain barrier (BBB) [57]. However, exosomes therapy has some blind spots such as lack of regenerative and proliferation capacity, as well as the inability of targeted migration to the injury site. Thus, whether using exosomes or cells are completely distinctive approaches with exclusive pros and cons which should be considered regarding pathology and injured organ situation [58]. Advantages of hAECs and their exosomes, including availability, low cost, and desirable biological features would bring them up as a proposed therapeutic approach in COVID-19. Herein, we discuss different therapeutic mechanisms of hAECs such as immunomodulation, renin-angiotensin-aldosterone system (RAAS) regulation, alveolar fluid clearance, coagulopathy elimination, and regenerative capacity in COVID-19.

COVID-19 pathophysiology and the potential therapeutic effects of amniotic epithelial cells Immunomodulatory effect of hAECs in COVID-19

Although earlier reports focused on respiratory failure in the COVID-19, it has been observed that besides the lungs, SARS-CoV-2 is capable of invading various organs, including kidneys, brain, and gastrointestinal tract [59-61]. Additionally, the response of the host immune system is also a determining factor regarding the course of the disease. In general, the course of the COVID-19 in severe cases can be divided into three stepwise phases. In the first phase, known as the viral replication phase, the SARS-CoV-2 invades host cells and replicates. This invasion initiates both innate and adaptive immune system responses. However, some patients' inadequate or misdirected immune response can lead to the second phase, known as "cytokine storm," resulting in a hyperinflammatory response and systemic inflammation [62]. This hyper-inflammation can cause ARDS, multi-organ failure, and death in the last phase. Understanding the bimolecular reactions in each phase can guide researchers into new preventive and curative actions.

After arriving at its destination, SARS-CoV-2 fuses with the host cells using Spike (S) proteins. Since previous reports suggested that angiotensin-converting enzyme (ACE)-2 was involved in the invasion of host cells by SARS-CoV and other coronaviruses, ACE-2 was one of the early culprits that have been investigated for its potential role in SARS-CoV-2 invasion [63]. Later findings indicated that ACE-2, with the help of other membrane proteins such as transmembrane serine protease 2 (TMPRSS-2), binding immunoglobulin protein (GRP78), and CD-147, an MMP inducer expressed in epithelial cells and WBC, acts as a target for the S2 domain of SARS-CoV-2 [64-67]. ACE2 is expressed on various cells, including the lung alveoli, gastrointestinal tract, brain, kidney, and liver. The co-expression pattern of ACE-2 with TMPRSS-2, GRP78, heat shock protein-70 (HSP-70), and CD-147 can decide the pattern of the symptoms and prognosis of patients [68].

After the host cells are invaded by SARS-CoV-2, the innate immune system and adaptive immune system are activated through paracrine signaling [69]. Since the complete activation of both the humoral immune system that produces neutralizing antibodies and the cellular immune system may need days to be completed, the early responses to the infections can be attributed to the innate immune system [70]. Interactions between infected cells and antigen-presenting cells (APCs) trigger innate immune system activation [71]. To activate the innate immune response, the single-strand RNA of the virus (ssRNA) interacts with pattern recognition receptors (PRRs) such as toll-like receptors (TLRs) and retinoic acid-inducible gene-I-like receptors (RLRs), which are expressed in pneumocytes, dendritic cells, monocytes, and macrophages [72, 73]. TLR-7 triggers the Janus kinase (JAK)-signal transducer and activator of transcription (STAT), activator protein 1 (AP-1), interferon regulatory factors (IRF)-3, IRF-7, and nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) [73, 74]. The NF-κB upregulates the production of IL-1, IL-6, tumor necrosis factor (TNF)- α , and interferon (IFN)- α , which are the central cytokines involved in the hyperinflammatory response in patients with COVID-19 [75]. The production of these cytokines attracts the cells of the innate immune system as well as lymphocytes. Additionally, the innate immune system tries to eliminate infected cells by secreting defensins and other antimicrobial peptides, reactive oxygen species (ROS), and neutrophil extracellular traps (NET), as well as producing more pro-inflammatory cytokines [9, 76].

The IFN- α has a crucial role in defending the host cells against the invading virus. However, the N protein of the SARS-CoV-2 redirects the signaling pathways from IFN- α producing cascades to IFN- β producing

cascades, resulting in a hyper-reactive but inadequate innate immune response [77, 78]. In COVID-19, it has been observed that the number of resident alveolar macrophages has been significantly decreased. In contrast, there has been an increase in the number cytokine-producing pro-inflammatory-monocytederived macrophages [79]. The overproduction of the pro-inflammatory cytokines can result in a cytokine storm and an exaggerated immune system response that can lead to acute respiratory distress syndrome (ARDS) [62]. The response from the adaptive immune system leads to an increase in the number of CD4+ cells, CD8+ cells, and Th-17 in the invasion sites [80]. The activation of the adaptive immune system will also further upregulate the NF-kB signaling cascade, which is responsible for the secretion of IL-1, IL-6, and TNF- α [81]. Furthermore, activation of the adaptive immune system will result in the production of IL-8, monocyte chemoattractant protein (MCP), granulocyte colony-stimulating factor (G-CSF)-α, TNF-α, prostaglandin E2 (PGE2), and IL-17 [82]. This overproduction of cytokines by both the innate and adaptive immune systems can lead to T cell exhaustion. The exhausted T cells (Tex), which can be identified by the presentation of specific markers such as CD279 and CD 366, have a reduced level of functionality and cannot provide an efficient immune response against SARS-CoV-2 [83]. In this regard, the lower number of CD3+ cells, CD4+ cells, CD8+ cells, CD19+ cells, and natural killer (NK) cells was observed in severe cases of COVID-19 [80]. It means that regulatory mechanisms that inhibit the overproduction of pro-inflammatory cytokines have also been disrupted. The uncontrolled overproduction of cytokines primarily by the innate immune system will result in a cytokine storm and a systemic hyper-inflammatory/immunodeficiency state. In this phase, the local immune system response turns into systemic inflammation and multi-organ failure [84].

Amniotic epithelial cells have been successfully used in treating several conditions such as bronchopulmonary dysplasia, lung injury, the aftermath of cerebrovascular accidents, and liver fibrosis [85, 86]. As mentioned earlier, the COVID-19 course includes a viral phase, a cytokine storm phase, and a multi-organ hyper-inflammation phase. The immunomodulatory features, anti-inflammatory and anti-fibrotic, and antimicrobial characteristics of hAECs can be beneficial in the viral phase, cytokine storm, hyper-inflammation phase, as well as repairing the degenerative damages in the lung tissue.

hAECs can regulate immune system response by altering the expression pattern of inflammatory cytokines through paracrine signaling or direct cell contact. hAEC can potentially control the overproduction of cytokines and prevent cytokine storms. TNF- α and IL-6 are two

of the most contributing factors in cytokine storm and occurrence of ARDS [87, 88]. AECs release some small molecules like migration inhibitory factor (MIF), IL-10, and PGE2, which can modulate cytokine-producing inflammatory cells and prevent the cytokine storm [89]. In a study conducted by Murphy et al., hAECs suppressed inflammation and promoted remodeling in mice with lung injury. This study demonstrated that levels of inflammatory cytokines such as TNF α, TGF-β, IF-γ, IL-6 have significantly decreased after hAECs injection [27]. Furthermore, it was observed that injection of hAECs prevented infiltration of inflammatory cells. In another study with similar aims, it was observed that hAECs decreased MCP-1, TNF- α , IL-1, IL-6, and TGF- β [90]. It is also worth noting that these immunomodulatory features of hAECs depend on regular host immune function and cannot be observed in patients with impaired immune systems. The innate immune system plays an essential role in hAECs immunomodulatory effect since observations indicate that the anti-inflammatory properties of hAECs are dependent on host normal macrophage function [91].

hAECs modulate the proliferation and differentiation of both the adaptive and innate immune system cells [17]. These cells present markers such as CD59 and FasL that modulate adaptive immune system response. Moreover, the HLA-G presented by hAECs regulates the differentiation of Treg, myeloid cells, and NK cells and prevents hyperinflammatory reactions [92]. As mentioned earlier, the cytokine storm is triggered by exaggerated innate immune system response and uncontrolled stimulation of the adaptive immune system. hAECs can prevent the differentiation of dendritic stem cells through contact inhibition and prevent unnecessary triggering of both the innate and adaptive immune systems at the beginning [93]. They also have shown promising results in reducing neutrophil infiltration and reducing oxidative bursts. Furthermore, it has been observed that in lung injuries, hAECs alter the polarization of macrophages and shift pro-inflammatory M1 macrophages into M2 macrophage, which can secrete anti-inflammatory cytokines like IL-10 through paracrine signaling [94]. Observations also indicated that hAECs could decrease the production of cytokines produced by CD4+ cells, Th1 cells, and Th17 cells while increasing cytokines produced by Th2 through IL-5 signaling cascade [95]. The shift toward the IL-5 pathway modulates hyper-inflammatory responses and improves the humoral immune system response. hAECs could help the maturation of FOXP3-expressing Treg cells, which can modulate inflammatory responses [96]. In another study, it was observed that hAECs could reduce the amount of pro-inflammatory cytokines such as IL-17 and caused an increase in the number of Treg and CD4+ T cells after an ischemic cerebrovascular accident (CVA) [97]. An increased proportion of Th2 cells in the peripheral lymphoid organs and within the central nervous system (CNS) were also observed in this study. It is also worth mentioning that hAECs do not suppress the immune system but prevent hyper-inflammation. An interesting observation in this study was prevention of pneumonia which is a common occurrence in the aftermath of CVA due to temporary immunocompromisation [98].

The hAECs secrete antimicrobial peptides (AMPs) such as human β -defensin (HBD) 1, HBD2, HBD3, secretory leukocyte protease inhibitor, and elafin [99]. We previously showed that inflammatory cytokines such as IL-1 β significantly upregulates the secretion of these antimicrobial peptides [100]. HBDs, which naturally are present in the mucosa, can trigger the activation of the innate and adaptive immune system and play an essential role in the early immune response to prevent the spread of the virus [101]. Furthermore, HBD3 has shown excellent antibacterial properties that can prevent nosocomial secondary bacterial infections in COVID-19 [102].

In recent years, exosomes have been the subject of many studies. It has been suggested that exosomes extracted from hAECs have anti-inflammatory properties. It has been shown that AEC-derived exosomes were enriched for factors that participate in immunomodulation, such as P38, PI3K-Akt, and proteins involved in Fc-gamma receptor-dependent phagocytosis pathways. The hAECs-derived exosomes increased macrophage's phagocytic activity and altered their polarization state mainly through the PI3K-Akt pathway. Exosomes could also reduce neutrophil myeloperoxidase and suppress CD3/CD28 activated T cell proliferation [103].

Effects of hAECs on renin–angiotensin–aldosterone system in COVID-19

The RAAS supervisory system is extended in different body organs that consist of several components, including angiotensinogen, various angiotensin (Ang) types, ACE, and AT1 receptors. ACE-1 converts Ang I to Ang II, which occurs extensively in the lung. Ang II can induce inflammation, profibrotic pathways, thrombosis, and vasoconstriction via the angiotensin II type 1 receptor (AT1R). Another arm of RAAS acts through the ACE-2-Ang-(1-7)-Mas receptor axis, in which the activity of ACE-2 leads to conversion of Ang II to the Ang-(1-7). This axis possesses counterregulatory actions on Ang II and results in vasodilation and reducing inflammation [63, 104]. The contributive roles of various components of RAAS in some viral infections are already established. Early studies investigated and confirmed the contribution of ACE-2 in SARS-CoV-2 invasion. SARS-CoV-2

infiltrates the host cells' cytosol using their envelopeanchored spike (S) glycoproteins binding to the ACE-2 cellular receptors [105]. Nearly all body organs express this receptor, especially type 2 alveolar epithelial cells [106]. The interaction between the viral spike protein and ACE-2 promotes the activation of metalloproteinase 17 (ADAM-17), which results in shedding and inhibition of ACE-2 [107]. Inhibition of ACE-2 redirects the RAAS axis toward ACE 1-Ang II-AT1 receptor, which is a lungdestructive factor by inducing fibrosis, inflammation, vasoconstriction, and alveolar epithelial cells apoptosis [108].

Amniotic epithelial cells can reduce the harmful effects of ACE 1-Ang II-AT1 receptor axis on pulmonary tissues. It has been demonstrated that Ang-(1-7) is presented in amniotic epithelial cells adjacent to the implanted embryo that indicates possible luminal secretion of these peptides. Besides, increased Ang-(1-7) secretion at late gestation was observed in the uterus and placenta. The presence of Ang-(1-7) induces inhibitory effects on Ang II that results in vasodilation and lower inflammation. It is hypothesized that the higher amount of Ang-(1-7) may be attributed to other enzymes such as neprilysin and prolyendopeptidase [20]. Some studies reveals that the amounts of ACE2 and TMPRSS2 are correlated to pregnancy stages. The expression of both ACE2 and TMPRSS2 are reduced gradually during first trimester and second trimester and it is undetectable in third trimester placenta [109]. Placental absence of ACE2 and TMPRSS2 as critical gateway of SARS-CoV-2 entry protect hAECs against possible infection during COVID-19 cell therapy. This protective ability prevents exclusion of hAECs from therapeutic process and provides safe cell therapy regardless of possible viral infection.

Lung-related alveolar fluid clearance effect of hAECs in COVID-19

Alveolar fluid clearance (AFC) impairment in COVID 19 can lead to related lung disease poor outcomes due to gas exchange disturbance and ARDS and/or coinfections. To achieve an efficient AFC, it is crucial to establish a tight alveolar epithelial barrier, as well as set a sufficient ions gradient across the alveolar epithelium in which sodium (Na⁺) plays a pivotal role. The sodium-potassium adenosine triphosphatase pump (Na/K-ATPase) actively provides the Na+ gradient. Subsequently, Na+ enters the epithelial cells through the epithelial sodium channel (ENaC), which reabsorbs water in the alveolar space. Besides, the Na/K-ATPase pump, localized at the lateral and basal sides of the epithelial cells, works as a tight junctional molecule and a regulator for the cytoskeletons network and the tight junctions' permeability. SARS-CoV-2 can alter the expression and function of the Na/K-ATPase pump by elevating levels of inflammatory cytokines, including IL-1 β , IL-6, IL-8, and TNF- α [110]. TNF- α can also alter sodium uptake and chloride secretion by reducing the expression of the ENaC mRNA and the stability of the cystic fibrosis transmembrane conductance regulator (CFTR) as a chloride channel, which can result in higher airway liquid film depth and pulmonary edema. Notably, type II alveolar cells use CFTR as the chloride channel for the active clearance of alveoli from water. In addition, TNF- α increases the permeability of the pulmonary capillary endothelial cells and alveolar epithelial cells through induction of cytoskeleton disruption [111].

Lung surfactant principally reduces the surface tension at the air-liquid interface in alveoli, prevents lung collapse, and facilitates breathing. Surfactant consists of phospholipids (80%), cholesterol (10%), and four surfactant proteins (SP), including the hydrophilic SP-A and SP-D (also named collectins), and hydrophobic SP-B and SP-C [112]. Type II alveolar cells are responsible for the secretion and recycling of surfactants. It has recently been suggested that administration of the surfactant can reduce pulmonary edema, improve blood oxygenation, regulate the uncontrolled inflammatory response, and facilitate respiratory effort in patients with critical COVID 19 pneumonia. Even though lung surfactant therapy is a standard and efficient treatment for ARDS, natural surfactants appear more beneficial than recombinant surfactants [113]. In addition, the intratracheal application of the SP-C-based surfactant has demonstrated immunoregulatory features and improved gas exchange [114]. Although the pulmonary anti-inflammatory properties of surfactants are well known, their non-pulmonary immune function is also noteworthy. For instance, the topical administration of surfactants on the skin has improved wound healing with less fibrotic composition through a significant reduction in the level of proinflammatory cytokines TNF-α and IL-6 [112]. Ujma et al. have reviewed the anti-inflammatory and antimicrobial properties of hydrophilic surfactant proteins (SP-A and SP-D) in tissues such as gastrointestinal and female reproductive tracts by reducing TNF- α and IL-1 in various sites like skin, eye, ear, nose, artery, CNS, and gastrointestinal, urinary, and female reproductive tracts, which can be possible targeted sites for SARS-CoV-2 and its co-infections [115].

The contribution of the hAECs in the secretion of pulmonary surfactant into amniotic fluid has been identified. It has been demonstrated that hAECs express all four surfactant proteins SP-A, SP-B, and SP-C, and SP-D. Moreover, the hAECs express ATP binding cassette subfamily A membrane 3 (ABCA3), a lamellar body protein requisite for pulmonary surfactant construction that also

expresses in alveolar type II cells. This study also showed that dipalmitoyl phosphatidyl choline (DPPC), one of the predominant components of the surfactant phospholipids and an essential contributor to surfactant biophysical functionality, could be detected in hAECs-derived supernatant [22]. These results confirmed that hAECs could express and secrete all four types of surfactant proteins with potential biophysical performance to reduce air—liquid surface tension.

The potential ability of the hAECs for differentiation to the alveolar epithelial type II cells can also bolster the hypothesis of utilizing hAECs for improving AFC in COVID 19 patients [88]. Besides, this study shows that hAECs administration represents lung fibrosis abrogation and repair of lung injury through anti-inflammatory features, such as decreasing TNF- α, IL-1, IL-6, TGF- β, and MMP-1 2. Formerly, it has been indicated that hAECs produce the tissue inhibitor of metalloproteinase (TIMP) type 1 and specifically type 2, which prevent inflammatory effects of MMPs [116]. In this regard, hAECs can potentially interfere with the destructive effect of COVID-19 on alveolar fluid clearance. These cells can preserve the transepithelial ion movement by regulating the inflammatory-induced down-regulation of ENaC and CFTR in lung injuries and COVID-19. A study on the combination of the hAECs and surfactant at clinically-used concentrations determined that this exposure has no damaging effect on the viability, functionality, and phenotype of the hAECs. Besides, the hAECs could differentiate to alveolar type II epithelial cells in small airway epithelial growth medium regardless of treating with surfactant [117].

To conclude, the distinctive properties of hAECs in clearing the alveolar space of fluid brought the convincing idea of utilizing these cells for patients with severe COVID 19. This can also be achieved by co-administrate of the hAECs and surfactants.

Effects of hAECs in COVID-19 hypercoagulopathies

COVID-19-associated dysregulated coagulation is one of the major complications that could be seen in COVID-19 infection. This condition could cause life-threatening exposures such as deep vein thrombosis (DVT), thromboembolism, platelet aggregation, and thrombosis. In a cohort study of 184 COVID-19 patients in the intensive care unit (ICU), the cumulative incidence of large-vessel thrombotic events was 49%, and pulmonary embolism was the main observed event in lung computed tomography (CT) [118]. The extensive expression of ACE2 receptors within endothelial cells simplifies the SARS-CoV-2 binding and entrance, causing infection, vascular injury, and endothelial dysfunction [119, 120]. Circulating cell-free (CF) nuclear materials (e.g., DNA, RNA,

and histone) that are released in the infectious situation due to cellular and tissue necrosis, apoptosis, autophagy, or mitotic catastrophe is known as NETosis [121]. Neutrophil extracellular traps (NETs) are DNA structures combined with histones and antimicrobial proteins that are released during cell injuries, especially by neutrophil cytotoxic enzymes [122]. Circulating cell-free nuclear materials could be a cofactor for coagulation activation factors VII, XI, and XII-activating proteases and provoke a thrombotic response [123, 124]. NET-associated histones could cause platelet aggregation by TLRs, which exist on platelets and other cells' surfaces [125]. Proinflammatory cytokines such as IL-6, IL-17A, and TNF-α are enhanced in moderate to severe condition patients. These pro-inflammatory cytokines significantly induce clot formation and platelet hyperactivation and disrupt physiological anticoagulant pathways [126]. Studies have shown that inhibition of IL-6 leads to the elimination of thrombin production. IL-1 and TNF- α show their effects by downregulation of thrombomodulin and inhibiting tissue plasminogen activator [127]. Vascular injury and disintegrated intercellular junction due to COVID-19 could expose the subendothelial matrix, including tissue factor (TF) and collagen [128]. Furthermore, in COVID-19, TF expression on macrophages and platelets is enhanced by inflammatory cytokines and tissue factor pathway inhibitor (TFPI), which inhibits the TF pathway, is impaired by inflammation. As a result, the coagulation cascade is activated, and an increase of fibrin level raises the risk of clot formation in the blood [129, 130].

Expression of IL-10 by hAECs inhibits coagulation by suppressing monocytes. Monocytes trigger coagulation cascades by activating tissue factor (TF) and stimulating thrombin production [26]. Furthermore, two types of glycosaminoglycans, including perlecan and hyaluronic acid are secreted from hAECs that inhibit thrombosis by inducing endothelial cell proliferation, which is necessary to suppress the initiation of thrombosis. Pigment-epithelium derived factor (PEDF) is another released factor from hAECs that inhibits platelet activation and aggregation through antioxidant capacity. MMP-9 secreted by amniotic cells can also prevent platelet activation by suppressing the Na+/K+ exchanger and adjusting intracellular calcium balance [26]. Platelet activation results in the adhesion of platelets to each other that leads to the release of their granule substances, such as P-selectin, which is mainly known for participating in platelet rolling during the clot formation process [131]. Research indicates that incubating platelets with hAECs can considerably reduce P-selectin levels and consequently expand prothrombin time (PT) and prevent thrombosis [132]. Furthermore, hAECs express anti-inflammatory factors like IL-4, IL-10, and IL-13 to suppress inflammation

and downregulate pro-inflammatory cytokines IL-1 and TNF- α [26]. The latter pro-inflammatory factors show procoagulant properties by inhibiting three critical substances of the anticoagulation procedure, including plasminogen activator, thrombomodulin, and protein C [127]. The proposed therapeutic effects of hAECs and common mediators are shown in Table 1.

Tissue regenerative ability of hAECs in COVID-19

COVID-19 can affect the tissues of the body, especially in the respiratory system. It has been demonstrated that 17% of the COVID-19 patients had ARDS, among whom 65% died of the disease worsening [133]. MMPs released in the inflammation phase of COVID-19 can damage the epithelium and endothelium; thus, the tissue could progress to fibrosis. Some substances secreted by epithelial cells, such as pro-fibrotic growth factors, chemokines, vascular inhibitors, and procoagulant mediators activate the fibroblasts and play a key role in lung fibrosis [134]. In the post-COVID-19 patients who have experienced ARDS during their disease, irreversible and progressive pulmonary fibrosis

is one of the critical long-term complications [135]. Besides, the cardiovascular system might be affected by COVID-19; however, the underlying pathophysiology is unclear. Several mechanisms such as ACE2-dependent myocardial infection, myocyte apoptosis due to excessive extracellular Ca²⁺ level caused by cytokine storm, myocardial injury due to increased myocardial demand in an acute infection, arrhythmia, and abnormal blood pressure are supposed for cardiac damage [136-138]. Kidneys are another organ that COVID-19 could damage. Sepsis caused by cytokine storm and direct attack of the virus through ACE-2 and dipeptidyl peptidase-4 (DPP-4), which are expressed on tubular cells, have been known as binding seats for SARS-CoV [139]. Liver injury with elevated hepatocyte-derived enzymes and serum bilirubin has been observed in COVID-19 patients. Studies showed that 14.8-53.1% of COVID-19 patients had abnormal levels of alanine aminotransferase (ALT) and aspartate aminotransferase (AST) during the disease, with a mostly mild rise in serum bilirubin which suggested hepatocyte damage [140]. The mechanism of liver injury is not precise, but it may

Table 1 Proposed therapeutic effects of hAECs and responsible mediators in COVID-19

Mediators	Mechanism	Effects	References	
MIF	Immunomodulation	Prevention of cytokine storm	[27, 89]	
IL-10				
PGE2				
HLA-G	Immunomodulation	Regulation of immune cells differentiation	[92]	
IL-5	Immunomodulation	Modulation of hyper-inflammatory responses	[95]	
		Improvement of humoral immune system		
AMPs	Immunomodulation	Triggering early immune response	[101, 102]	
		Prevention of nosocomial secondary bacterial infections		
Ang-(1-7)	Regulating RAAS	Reduction of inflammation, fibrosis, thrombosis, and vasoconstriction	[20, 63, 104]	
SP-A	Alveolar fluid clearance	Reduction of air-liquid surface tension	[22]	
SP-B				
SP-C				
SP-D				
TIMP-1	Alveolar fluid clearance	Prevention of inflammatory effects of MMPs	[116]	
TIMP-2				
IL-10	Eliminating hypercoagulopathies	Suppression of coagulation cascades by inhibiting monocytes-induced activation of TF	[26]	
Perlecan	Eliminating hypercoagulopathies	Inhibition of thrombosis by inducing endothelial cell proliferation	[26]	
Hyaluronic acid				
PEDF	Eliminating hypercoagulopathies	Inhibition of platelet activation and aggregation through antioxidant capacity	[26]	
MMP-9	Eliminating hypercoagulopathies	Prevention of platelet activation by suppressing the Na+/K+ exchanger and adjusting intracellular calcium balance	[26]	
IL-4	Eliminating hypercoagulopathies	Inhibition of plasminogen activator, thrombomodulin, and protein C	[26, 127–130]	
IL-10				
IL-13				

MIF, migration inhibitory factor; IL, interleukin; PGE2, prostaglandin E2; HLA, human leukocyte antigen; AMP, antimicrobial peptide; Ang, angiotensin; SP, surfactant; TIMP, tissue inhibitor of metalloproteinase; PEDF, pigment epithelium-derived factor; MMP, matrix metalloproteinase; RAAS, renin-angiotensin system

be due to direct virus-induced cytopathic effects and/ or immunopathology induced by overshooting inflammatory responses and drug-induced injury during the treatment of the disease [141]. COVID-19 can cause acute or chronic neurological deficits that could raise the risk of neurodegenerative diseases [142]. COVID-19 also attacks the endothelium of the vessel's wall. It was described that the cytokine storm and the inflammation afterwards could detach the endothelial cells. Whatever the severity of the disease is higher, the count of detached endothelial cells increases [143]. Considering the involvement of various body organs and possible tissue damage, proposing an effective regenerative approach for reducing tissue damage-related late complications is necessary.

Regenerative medicine is a promising treatment for chronic and disabling situations [144]. Transplanted stem cells participate in tissue regeneration via two main mechanisms: first, by direct differentiation of cells toward mature and functional cells, and second, via their paracrine impacts. It seems that paracrine influences are essential in tissue regeneration compared with direct differentiation. Paracrine effects consist of various cytokines, growth factors, and differentiation-promoting agents that play critical roles in tissue repair and regeneration [145]. hAECs have many capabilities, such as stem cell-like proliferation, differentiation, and immunomodulatory features [146]. Considering tissue damages due to SARS-CoV-2 infection and dysregulated immune response, hAECs could participate in the regeneration of COVID-19 affected organs.

The differentiation potential of hAECs makes them a source for cell replacement therapy [147]. The hAECs express some lung cells specific markers such as thyroid transcription factor or Nkx 2.1, which is among the earliest lineage specification markers of the developing lung. These markers give hAECs the potential to differentiate into type II alveolar pneumocytes and produce surfactants [88, 148]. In addition to the differentiation properties, hAECs-derived exosomes have a crucial role in regeneration medicine. Exosomes of hAECs contain P38 protein, which is critical factor for tissue regeneration. These exosomes contain miR-23a and miR-203a, which inhibit Smad 2 and 3, respectively, as well miR-34a that induces lung fibroblast senescence. The mentioned miRNAs cause the suppression of TGF-β and fibroblast proliferation [103]. Besides, it has been shown that the miRNAs content of hAEC-derived exosomes participated in their regeneration ability [149]. Tan et al. showed that hAECs could decrease tissue damages by inhibiting myeloperoxidase (MPO) produced by neutrophils [103]. hAECs application also can prevent lung injury by reducing the number of neutrophils and macrophages in the lung and inhibiting the overexpression of inflammatory cytokine genes such as TNF- α , IL-1, IL-6, and IFN-gamma [150].

In hypoxic cardiac damage, hAECs secrete pro-angiogenic cytokines, including angiogenin (ANG), epidermal growth factor (EGF), monocyte chemoattractant protein (MCP)-1, and IL-6 that have angiogenic effects through several pathways, including paracrine effects and chemoattractant-mediated effects [151]. When cultured hAECs are exposed to TGF- β 1, Epithelial-to-Mesenchymal Transition (EMT) can occur. EMT would enhance the cardiac regeneration capacity of hAECs by increasing their mobility and extracellular matrix-modulating capacity [152].

In kidney injury, intravenous injection of hAECs diminished the apoptosis and necrosis of peritubular capillaries and tubular cells and provoked cell proliferation. hAECs decrease the number of pro-inflammatory M1 macrophages and shift them to anti-inflammatory M2 macrophages. This shift is associated with the increased IL4 and IL13, and reduced levels of TNF α and IFN γ , which reduce the inflammation response [153]. hAECs motivate the expression of miR-101 in CD4+ T cells in acute kidney injury (AKI) patients that inhibit the activation of CD4+ T cells. As a result, IL-2, which plays a crucial role in kidney injury, is reduced in inflammation [154].

Native hAECs express hepatocyte-related genes such as alpha (1)-antitrypsin, glutamine synthetase, phosphate synthetase-I, phosphoenolpyruvate carboxykinase, cytochrome P (CYP)-450, CYP2D6, and CYP3A4. When hAECs are cultured in an in vitro environment, they express transthyretin, tyrosine aminotransferase, CYP2C9, and the key hepatocyte transcription factors, HNF-3g and C/EBP-a genes [155]. In vitro, hAECs differentiate into hepatocyte-like cells with the fetal hepatocytes features, whereas in vivo, they differentiate into adult hepatocytes [156]. It has been shown that transplanted hAECs reduced collagen deposition and diminished the pro-inflammatory and pro-fibrotic cytokines in an experimental animal model of liver fibrosis [157].

The therapeutic potential of hAECs is also seen in the treatment of neurological injuries and diseases. hAECs could differentiate into the neural cells. Moreover, they have paracrine effects by secreting cytokines, neurotrophic factors (NTFs), growth factors, and neurotransmitters that facilitate neural survival and regeneration, axonal outgrowth, and synapse reformation, thus leading to the reinnervation of lost neuronal connections and further recovery of neurological functions [158]. Considering the regenerative features of hAECs, including releasing many regenerative factors, differentiation to intended cells, and creating an appropriate reconstructive environment, they could emerge as a suitable

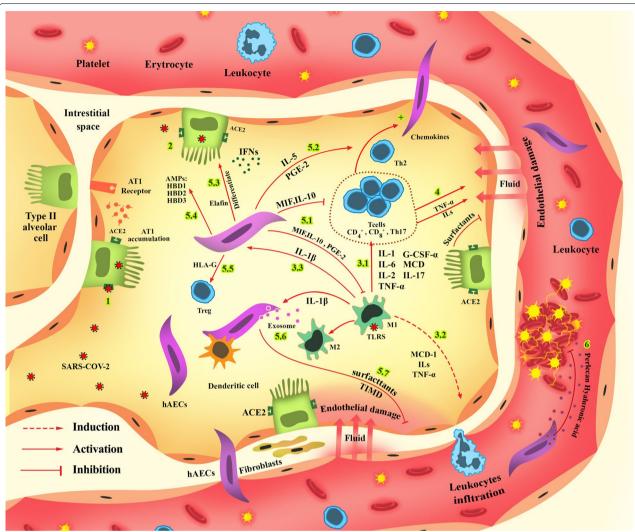


Fig. 1 Proposed therapeutic mechanisms of hAECs in COVID-19. (1) SARS-CoV-2 enters the respiratory system and attaches to ACE2 receptors located on the basolateral membrane of type II alveolar cells. ACE2 damage would result in AT1 accumulation in alveoli that could induce vasoconstriction, inflammation, fibrosis, and apoptosis of alveolar epithelial cells. (2) Interferons, especially type I, are released from infected type Il alveolar cells that defend against the invading virus. (3.1) M1 macrophages are activated in the immune response process that release IL-1, IL-6, IL-8, TNF-alpha, G-CSF-alpha, MCP, IL-17, and IL-5. These factors activate the CD4+, CD8+, and Th17 T cells. (3.2) MDC, IL-5, and TNF-alpha have chemotaxis effects that cause the migration of leukocytes to the alveolar space. (3.3) IL-1ß secreted by M1 macrophages could activate the hAECs. (4) Activated CD4+, CD8+, Th17 cause inflammation and cytokine storm that damage the endothelial and epithelial cells and alveolar fluid accumulation. T cells also have chemotaxis properties that can call the hAECs to the alveolar space. (5.1) hAECs release PGE-2, IL-10, and MIF that inhibit the activation of macrophages, CD4+, CD8+, and Th17 cells and modulate cytokine-producing inflammatory cells. (5.2) hAECs induce inhibitory cytokine production in T-helper 2 through IL-5 signaling cascade. (5.3) hAECs can be differentiated into alveolar cells that regenerate damaged lung tissue. (5.4) hAECs secrete AMPs such as HBD1, HBD2, HBD3, secretory leukocyte protease inhibitor, and elafin. AMPs play an essential role in the early immune response that reduces the spread of the virus. Furthermore, they have antimicrobial properties that prevent nosocomial infection. (5.5) HLA-G presented by hAECs can regulate the differentiation of Treg cells, preventing hyperinflammatory responses. This feature also reduces the chance of immune rejection in the human body. (5.6) Activated hAECs release some exosomes which contain regenerative agents and PI3K-Akt pathway activators that induce M1 to M2 macrophage polarization. (5.7) hAECs release surfactants and TIMP that prevent the accumulation of the fluid in the alveoli. (6) hAECs secrete two types of glycosaminoglycans, perlecan and hyaluronic acid, that inhibit thrombosis. Thus, hAECs could play an important role as an inhibitor of clot formation in coagulation dysregulation caused by COVID-19. Abbreviations: hAECs, human amnion epithelial cells; ACE, angiotensin-converting enzyme; AT1, angiotensin 1; IL, interleukin; G-CSF, granulocyte colony-stimulating factor; MCP, monocyte chemoattractant protein; MDC, macrophage-derived chemokine; PGE, prostaglandin; MIF, macrophage migration inhibitory factor; AMPs, antimicrobial peptides; HBD, human beta-defensin; HLA, human leukocyte antigen; TIMP, inhibitor of metalloproteinase

regenerative agent in COVID-19 degenerative complications. The possible therapeutic effects of hAECs are shown in Fig. 1.

Modification of hAECs for administration in COVID-19

Preconditioning and gene modification of hAECs

Although cell-based therapy (CBT) has shown promising potential in treating various disorders, its efficacy needs to be promoted to reach desirable outcomes. One of the major causes of poor therapeutic performance of transplanted cells is the host's harsh environment. Hypoxia, acidosis, and excessive inflammation in COVID-19 prevent administered cells from establishing sufficient function needed for the favorable therapeutic function [159]. Preconditioning is a promising strategy to overcome the negative impacts of the harsh environment on administered stem cells. Preconditioning is applied on the cells before transplantation in order to adjust them to the host's milieu and improve their function [160]. Preconditioning has various methods such as hypoxic preconditioning, acidic preconditioning, and pharmacologic preconditioning to enhance cell survival, anti-oxidative capacity, angiogenesis induction, migration, and immunomodulatory impacts [161–163].

In COVID-19, the body's destructive environment causes cell death and dysfunction due to various harassments such as hypoxia and acidosis [164, 165]. As a promising stem cell modification strategy, preconditioning could be utilized. Hypoxic preconditioning is shown to improve the survival and performance of administered cells in hypoxic conditions. The transcription factor hypoxia-inducible factor- 1α (HIF- 1α) is upregulated during hypoxic stress. HIF-1 α induces transcription of survival-related genes such as PI3K/Akt and ERK1/2, as well as proliferation-related genes such as Nrf-2 [166-168]. Acidic preconditioning could also improve hAECs therapeutic potential. It has been demonstrated that acidic preconditioning of hAECs improves their survival and hampers apoptosis through upregulation of PI3K/ Akt and ERK1/2 pathways [169].

Oxidative stress occurs in various conditions, such as ROS accumulation during hypoxia. The excessive ROS production by neutrophils causes tissue damage, thrombosis, and red blood cell dysfunction in critically ill COVID-19 patients [170]. Preconditioning with $\rm H_2O_2$ and hypoxia is shown to improve stem cells' antioxidative capacity through upregulation of genes such as Nrf-2, heme oxygenase-1, autocrine motility factor, and hexokinase-2 [171, 172]. Enhanced antioxidative activity promotes cell survival after injection and improves their therapeutic impacts against COVID-19 [170]. It has been reported that glial cell line-derived neurotrophic factor

(GDNF) preconditioning of amniotic cells improves stem cells' resistance against hydrogen peroxide-induced cytotoxicity. Moreover, preconditioning of amniotic cells with GDNF improves their migration and engraftment to the injured tissue by upregulating mobility and engraftment-related factors, including CD44, C-X-C chemokine receptor type 4 (CXCR4), and CX3CR1 [173].

Since the immunomodulatory features of hAECs are pivotal in COVID-19 treatment, it is therapeutically beneficial to improve cells' immunomodulatory capacities. Depending on the stem cells' microenvironment, they can shift toward both inflammatory and anti-inflammatory phenotypes. Preconditioning with high amounts of inflammatory cytokines such as TNF- α and IL-6 increases the shift towards anti-inflammatory phenotypes [174, 175]. It has been elucidated that preconditioning of hAECs with remifentanil (an opioid analgesic) alleviated the production of IL-1 β and TNF- α proinflammatory factors when exposed to an inflammation inducer lipopolysaccharides (LPS). Remifentanil preconditioning also decreases the expression of COX-2, a cytokine with proinflammatory impacts [176].

Gene modification is a cell manipulation strategy in which specific genes are upregulated or downregulated through various mechanisms such as utilizing viral vectors. It has been shown that gene modification of hAECs improves their viability, differentiation, and metabolic characteristics.

It has been reported that enhancing GDNF gene expression by gene modification improved amniotic cells' survival under oxidative stress and inflammatory conditions and promoted their migration and engraftment into the damaged site. Besides, GDNF gene-modified amniotic cells had augmented levels of HIF-1α, transforming growth factor beta 1 (TGF- β 1), and VEGF production. Reduced oxygen supply and tissue ischemia result in mitochondrial injury, mitochondrial ROS accumulation, and mitochondrial bioenergy dysfunction, leading to cell apoptosis. It has been shown that GDNF gene modification improved the anti-oxidative capacity of amniotic cells and preserved mitochondrial integrity and bioenergy. Upregulation of GDNF through gene modification also promoted cells' anti-inflammatory capacity. Transplanted gene-modified amniotic cells into mice models of obstructive nephropathy decreased MCP-1 and TNF-α inflammatory cytokine levels robustly.

hAECs as a drug delivery tool

In recent years, many efforts have been made to use hAECs as drug reservoirs due to their low immunogenicity, prolonged circulation time, and ability to controlled release. Drugs loaded on stem cells have higher concentrations at the target site on account of stem cell homing

ability to the damaged tissues. Moreover, these drugs possess reduced side effects and higher bioavailability due to lower kidney or hepatic clearance [177–179].

In this regard, some studies tried to load antiviral agents and antibiotics on AECs. It is demonstrated that AECs treated with acyclovir and trifluridine antiviral drugs inhibited replication of viruses in cell culture, showing their potential for usage as a drug reservoir for antiviral purposes [180]. Besides, amniotic cells could be an appropriate choice for ofloxacin delivery, a quinolone antibiotic, in a sustained drug release manner [181]. Moxifloxacin is a fourth-generation fluoroquinolone used for the treatment of bacterial infections. It has been reported that amniotic cells could be a proper choice for moxifloxacin drug delivery since they can release the drug even 7 weeks after loading of drug [182]. Amniotic cells treated 30 min with netilmicin, an aminoglycoside antibiotic, inhibited Staphylococcus epidermidis infection for at least 3 days. These cells could uptake netilmicin in a dose-dependent manner in which the higher drug dose results in higher uptake by amniotic cells [183].

It seems that the idea of applying drug-loaded and modified hAECs is a hopeful and encouraging strategy as a therapeutic approach to encounter complications and progression of COVID-19.

Routes of delivery

For successful cell-based therapy, the administered stem cells must migrate to the target tissue. Migration is the arrival of stem cells from the delivery site to the target tissue, which occurs through the circulation system in systemic administration strategies [184]. Various molecules are involved in the migration and homing process of stem cells, such as chemotactic agents, adhesion molecules, and MMPs. Chemotactic agents such as CXCR4 promote stem cell tendency to the injured area and aid the migration process, leading to a successful homing. Adhesion molecules such as ICAM-1, ICAM-3, and integrins are responsible for stem cell adhesion and incorporation into the target tissue. MMPs such as MMP-2 and MMP-9 digest the target tissue's extracellular matrix (ECM) to facilitate stem cell embedment into the target tissue [185–187].

Different routes are available for hAECs delivery into the body, including intravenous injection, intratracheal, and intranasal routes. In addition, hAECs could be administered intraperitoneally in in vivo experimental studies [86, 88, 150, 188]. As discussed, administered stem cells migrate to the injury site with the assistance of systemic circulation, chemotactic agents, released inflammatory cytokines from the damaged tissue, and cell–cell adhesion molecules [87]. The most desired delivery route

has the fewest side effects, appropriate accessibility, and the highest cell number in the injury site.

Intravenous injection (IV), as the most common and feasible systemic administration route, has an advantage for treating pulmonary disorders, which is called the "pulmonary first-pass effect". It has been demonstrated that most IV injected stem cells are trapped into the lung, which augments the administered cells concentration in the damaged pulmonary site, which is beneficial for treating COVID-19 [189]. It has been shown that hAECs could migrate to the injured pulmonary tissue, differentiate into type I and II alveolar cells, and mitigate ventilation-induced lung injury [190]. Nevertheless, IV administered stem cells must reach the target site via systemic circulation, which diminishes their population in the target tissue compared to the local routes such as intranasal and intratracheal [191].

Intranasal and intratracheal administrations have been shown to improve stem cells' viability and population in the target tissue. In addition, intranasal delivery is a non-invasive route which makes it an attractive choice. However, some disadvantages are associated with these routes, including the need for intubation for intratracheal delivery and a limited number of cells that could be administered through these routes [87].

It has been shown that hAECs-derived exosomes possess many therapeutic effects. Exosomes could be administered through different routes such as intravenous, intranasal, subcutaneous, and oral. The intravenous route is the most frequent route for exosome administration due to its favorable feasibility. It is shown that exosomes can escape the immune system-mediated removal and liver clearance when administered intravenously. Moreover, exosomes are trapped into the lung during intravenous administration, similar to transplanted hAECs, which could be beneficial for the treatment of pulmonary disorders. Intranasal administration is a proper and noninvasive administration route for exosome delivery into the systemic circulation, and in some studies, it has been demonstrated that intranasal exosome delivery is even more efficient compared to IV injection [190, 192-194].

Pre-clinical and clinical application of hAECs

Considering various therapeutic potentials of hAECs, evaluating the possibility and safety of administration of these cells in animal models and humans is an essential step in developing AEC-based therapeutic approaches. Similar to drugs, Cellular and Tissue-Based Products (HCT/Ps) safety is a prior issue in clinical application. It has been shown that the most tolerated dose and the no observed adverse effect level (NOAEL) of hAECs used in a mouse model were both higher than MSCs. Besides, administering hAECs even at the most tolerated

dose had no adverse effect on growth, behavior, and general histology of animal models [195]. Tumorigenicity, another concerning issue in using HCT/Ps, is the main reason for failure in clinical trials on cell-based therapy [196, 197]. As mentioned above, some features of hAECs, including restricted proliferation due to the limited telomerase function, prevent tumorigenicity of hAECs [33]. Accordingly, the use of hAECs in animal models has not shown tumor formation that ensures another pivotal aspect of safety for clinical application of hAECs [195]. In another preclinical study, Hodges et al. utilized in utero ovine model of ventilation that caused ventilationinduced lung injury (VILI) to investigate whether hAECs would relieve lung injury. It has been shown that hAECs could reduce profibrotic mediators such as TGF-\(\beta\)1 and pro-inflammatory cytokines like IL-8 [190]. Vosdoganes et al. administered hAECs in bleomycin-induced pulmonary inflammation and fibrosis. They observed that intraperitoneally administration of hAECs returned lung tissue density, collagen content, and α -smooth muscle actin (α-SMA) production to a normal condition. It also reduced pulmonary leukocyte infiltration and expression of TGF- β , PDGF- α , and PDGF- β in the lungs [198]. The same study revealed that intraperitoneal administration of hAECs could downregulate proinflammatory cytokines such as TNF-α, TGF-β, IFN-γ, and IL-6 and also diminish subsequent pulmonary fibrosis by lowering pulmonary collagen deposition, levels of α -SMA, and inflammatory cells infiltration [27]. In another preclinical study, intranasal administration of AEC-derived exosomes in bleomycin-induced lung fibrosis reduced lung inflammation after 1 day. Besides, the treatment on the seventh day enhanced the tissue-to-airspace ratio and considerably ameliorated lung fibrosis [103]. The preclinical uses of hAECs in lung injuries are summarized in Table 2.

When the efficacy of using hAECs in pathologic conditions is approved in pre-clinical studies, it is critical to guarantee the production and preservation of HCT/ Ps under clinical-grade standards. In order to achieve acceptable standards for manufacturing procedures and controlling production quality of HCT/Ps, Food and Drug Administration (FDA), Health Canada, and European Medicines Agency (EMA) have considered particular guidelines called "Good Manufacturing Practice (GMP)" [202]. Pandemic situations and limitations such as time restriction, unavailability of appropriate equipment, and financial issues may influence fulfilling GMP standards. Viral contamination is a fundamental problem in preparing HCT/Ps during the COVID-19 pandemic. Contamination can originate from different sources, including operation room, placenta, biological raw materials, manufacturing environment,

and administration clinics. However, testing clinicians, related personnel, and the final products by utilizing validated tests for SARS-CoV-2 along with strictly adopting guidelines of GMP would reduce the chance of contamination [203].

As the first clinical use, hAECs have been administered in premature infants with bronchopulmonary dysplasia (BPD), a chronic lung disease with bronchi damage and alveolar dysplasia. Allogeneic hAECs were administered by intravenous infusion to six premature babies with BPD, and they were followed for 2 years. By utilizing an inline filter and lowering the concentration and the rate of infused hAECs, no adverse effects, including local site reaction, anaphylaxis, infection, features of rejection, or tumor formation was observed [204].

Some clinical studies have been launched to assess the safety and efficacy of hAECs in pathologic conditions. Up to January 2022, our search by the keywords "Amniotic epithelial cell" revealed 24 registered clinical trials of hAECs application in clinicaltrial.gov, of which seven are active and recruiting patients, and one has completed its trial. These clinical trials evaluate the efficacy of hAECs in various pathologic diseases such as bronchial fistula (NCT02959333), spastic cerebral palsy (NCT03107975), primary ovarian insufficiency (NCT03207412), infertility (NCT02912104), nonunion fracture (NCT03031509), corneal epithelial dystrophy (NCT00344708), acutegraft-versus-host disease (NCT03764228), Asherman's syndrome (NCT03223454), and Parkinson's disease (NCT04414813). To the best of our knowledge, there is only one clinical trial on utilizing hAECs for treating complications of COVID-19 in which Sandora et al. tried to regenerate cardiomyocytes damages due to myocardial infarction (MI) as an essential complication of COVID-19. They utilized a heart patch consisting of an amnion bilayer seeded with hAECs and autologous cardiomyocytes of patients who underwent coronary artery bypass graft (CABG) surgery. Autologous cardiomyocytes were isolated from waste tissues of the patient's heart after CABG surgery. They also used HLA-DR negative hAECs to reduce the risk of rejection. After the intervention, the condition of patients was evaluated by electrocardiography (ECG), echocardiography, blood test, and radiology (technetium-99m). However, no result of this study was published up to January 2022 (NCT04728906). Considering the proposed benefits of hAECs in COVID-19, including immunomodulation, alveolar fluid clearance, controlling RAAS, and regulating coagulation, along with the reported advantages of hAECs transplantation in lung injury models, these cells can be a remarkable resource for ameliorating the early and late complications of COVID-19.

Table 2 Pre-clinical use of hAEC in lung disease

No	Pathologic condition	Type of HCT/Ps	Mechanism of action	Administered dose	Route of administration	References
1	Lipopolysaccharide- induced lung injury	hAECs	Reducing lung inflam- matory mediators	1.8×10^8 (intratracheal)	Intratracheal	[86]
				0.9×10^8 (intravenous)	Intravenous	
2	Lung fibrosis	hAECs	Surfactant production	1×10^6	Intravenous	[88]
			Differentiation to pneu- mocytes			
			Immunomodulation			
3	Bleomycin-induced lung injury	hAECs	Reducing lung inflam- matory mediators	4×10^6	Intraperitoneal	[27]
			Reducing leukocyte infiltration			
			Preventing collagen deposition			
	Bleomycin-induced lung injury	hAECs	Reducing macrophage infiltration	4×10^6	Intraperitoneal	[94]
			Switching M1 to M2 macrophages			
5	Idiopathic pulmonary Fibrosis	hAEC-derived exosomes	Promoting proliferation of bronchioalveolar stem cell Increasing macrophage phagocytosis Reducing neutrophil myeloperoxidase Suppressing T cell proliferation	10 μg	Intranasal	[103]
6	Ventilation-induced lung injury	hAECs	Differentiated into type I and II alveolar cells	120×10^6 (total	Intratracheal	[190]
			Reducing fibrosis	I dose)	Intravenous	
	Bleomycin-induced lung injury	hAECs	Reducing lung inflam- mation	4×10^{6}	Intraperitoneal	[198]
			Reducing fibroblast activation			
8	Ventilation-induced lung injury	hAECs	Reducing T cell infiltration	0.9×10^8 (both routes)	Intratracheal	[199]
					Intravenous	
9	Bronchopulmonary dysplasia	hAECs	Reducing lung inflam- matory mediators	0.1×10^6	Intratracheal	[200]
			Reducing leukocyte infiltration		Intravenous	
10	Ventilation-induced lung injury	hAECs	Regenerating lung tissue	0.3×10^8	Intravenous	[201]

Discussion and future directions

Human amniotic epithelial cells and their exosomes can prevent or reduce hyper-inflammatory responses, tissue fibrosis, accumulation of alveolar fluid in ARDS, and severe thrombotic events. Also, hAECs can regulate transepithelial ion transportation by reversing hyper-inflammation on ENaC and CFTR in the lungs. The most prominent feature that makes hAECs a promising cell source for treating severe forms of COVID-19 is their immunomodulatory effects, which can be attributed to

the regulation of NF- κ B cascade and secretion of immunomodulatory cytokines such as IL-10. Furthermore, the secretion of all four natural surfactant proteins already used for treating ARDS prioritize the hAECS from other cell sources proposed to treat COVID-19. Moreover, the regeneration ability of hAECs makes them an appropriate choice for eliminating organ fibrosis as a late complication of COVID-19. Unfortunately, there are only a few reports on using hAECs in COVID-19; thus, further investigations must be conducted on the possible

mechanisms, safety, and therapeutic outcomes of hAEC therapy in COVID-19. Besides, hAECs produce antiviral and antimicrobial peptides which are parts of innate immune system [99]. We believe that these hAECs-produced peptides could participate in defending the human body against SARS-CoV-2. However, further experiments need to be done to clarify the AMPs' effects on this virus. Additionally, modification strategies such as preconditioning, gene modification, and drug loading have been shown to extensively improve hAECs capacity for tissue regeneration, immunomodulation, and antiviral combatting. These strategies ameliorate hAECs survival, migratory ability, paracrine impacts, AMP production, and anti-oxidative capacity. Modification strategies are promising tools to enhance the success chance of cell therapy [159]. Nevertheless, there are only a few reports on using hAECs in COVID-19; thus, further analysis and reconsideration in the expectations of surmounting their hurdles should be performed in future studies.

Conclusion

Human amniotic epithelial cells and their secretome would induce beneficial therapeutic effects in COVID-19 through modulating immune system reactions and controlling coagulopathies. They can also prevent lung tissue fibrosis, clear alveolar fluid, and participate in tissues regeneration in ARDS clinical and preclinical models. Modification methods such as preconditioning and gene modification can enhance the therapeutic effects of hAECs. However, there are only a few reports of administrating hAECs and their exosomes in treating COVID-19; thus, further studies are required regarding the possible therapeutic mechanisms and efficacy of hAECs in severe COVID-19 cases.

Acknowledgements

The authors would also like to express their most sincere words of appreciation to Dr. Tahere Tayebi for her valuable contribution.

Authors' contributions

A.B. and H.N. contributed to the conception. A.B., K.M., F.N., H.R., S.S., S.S., and E.J. reviewed the articles and wrote the original draft. E.J. designed the figures. H.N. and A.B. contributed to the manuscript revision and editing. All the authors read and approved the submitted version.

Funding

This work was supported by the Pharmacology Department of School of Medicine, Shahid Beheshti University of Medical Sciences. In addition, it was supported by the National Institutes for Medical Research Development (NIMAD), Tehran, Iran [963951].

Availability of data and materials

Not applicable.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

The authors grant the Publisher the sole and exclusive license of the full copyright in the Contribution, which licenses the Publisher hereby accepts. Consequently, the Publisher shall have the exclusive right throughout the world to publish and sell the Contribution in all languages.

Competing interests

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest

Received: 20 September 2021 Accepted: 25 February 2022 Published online: 25 March 2022

References

- Gupta RK. Will SARS-CoV-2 variants of concern affect the promise of vaccines? Nat Rev Immunol. 2021. https://doi.org/10.1038/s41577-021-00556-5.
- Harrington D, Kele B, Pereira S, Couto-Parada X, Riddell A, Forbes S, et al. Confirmed reinfection with SARS-CoV-2 variant VOC-202012/01. Clin Infect Dis. 2021. https://doi.org/10.1093/cid/ciab014.
- 3. Wibmer CK, Ayres F, Hermanus T, Madzivhandila M, Kgagudi P, Oosthuysen B, et al. SARS-CoV-2 501Y.V2 escapes neutralization by South African COVID-19 donor plasma. Nat Med. 2021;27:622–5. https://doi.org/10.1038/s41591-021-01285-x.
- Chen J, Wang R, Gilby NB, Wei G-W. Omicron variant (B.1.1.529): infectivity, vaccine breakthrough, and antibody resistance. J Chem Inf Model. 2022;62:412–22. https://doi.org/10.1021/acs.jcim.1c01451.
- Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet. 2020:395:497–506.
- Jamshidi E, Asgary A, Tavakoli N, Zali A, Dastan F, Daaee A, et al. Symptom prediction and mortality risk calculation for COVID-19 using machine learning. Front Artif Intell. 2021;4: 673527. https://doi.org/10. 3389/frai.2021.673527.
- Yadav PD, Mohandas S, Shete AM, Nyayanit DA, Gupta N, Patil DY, et al. SARS CoV-2 variant B.1.617.1 is highly pathogenic in hamsters than B.1 variant. bioRxiv. 2021. https://doi.org/10.1101/2021.05.05.442760.
- Davies NG, Jarvis CI, CMMID COVID-19 Working Group, Edmunds WJ, Jewell NP, Diaz-Ordaz K, et al. Increased mortality in community-tested cases of SARS-CoV-2 lineage B.1.1.7. Nature. 2021;593:270–4. https://doi. org/10.1038/s41586-021-03426-1.
- Schultze JL, Aschenbrenner AC. COVID-19 and the human innate immune system. Cell. 2021;184:1671–92. https://doi.org/10.1016/j.cell. 2021.02.029.
- Barbaro RP, MacLaren G, Boonstra PS, Iwashyna TJ, Slutsky AS, Fan E, et al. Extracorporeal membrane oxygenation support in COVID-19: an international cohort study of the Extracorporeal Life Support Organization registry. Lancet. 2020;396:1071–8.
- Liu Y, Liu J, Xia H, Zhang X, Fontes-Garfias CR, Swanson KA, et al. Neutralizing activity of BNT162b2-elicited serum. N Engl J Med. 2021;384:1466–8. https://doi.org/10.1056/NEJMc2102017.
- Abu-Raddad LJ, Chemaitelly H, Butt AA, National Study Group for COVID-19 Vaccination. Effectiveness of the BNT162b2 Covid-19 vaccine against the B.1.1.7 and B.1.351 variants. N Engl J Med. 2021;385:187–9. https://doi.org/10.1056/NEJMc2104974.
- Kunutsor SK, Laukkanen JA. Renal complications in COVID-19: a systematic review and meta-analysis. Ann Med. 2020;52:345–53. https://doi. org/10.1080/07853890.2020.1790643.
- Petrovic V, Radenkovic D, Radenkovic G, Djordjevic V, Banach M. Pathophysiology of cardiovascular complications in COVID-19. Front Physiol. 2020;11: 575600. https://doi.org/10.3389/fphys.2020.575600.
- Kunutsor SK, Laukkanen JA. Hepatic manifestations and complications of COVID-19: a systematic review and meta-analysis. J Infect. 2020. https://doi.org/10.1016/j.jinf.2020.06.043.
- Fraser E. Long term respiratory complications of covid-19. BMJ. 2020;370: m3001. https://doi.org/10.1136/bmj.m3001.
- Li H, Niederkorn JY, Neelam S, Mayhew E, Word RA, McCulley JP, et al. Immunosuppressive factors secreted by human amniotic epithelial

- cells. Investig Ophthalmol Vis Sci. 2005;46:900–7. https://doi.org/10. 1167/iovs.04-0495.
- Li J, Koike-Soko C, Sugimoto J, Yoshida T, Okabe M, Nikaido T. Human amnion-derived stem cells have immunosuppressive properties on NK cells and monocytes. Cell Transplant. 2015;24:2065–76. https://doi.org/ 10.3727/096368914X685230.
- Magatti M, Vertua E, De Munari S, Caro M, Caruso M, Silini A, et al. Human amnion favours tissue repair by inducing the M1-to-M2 switch and enhancing M2 macrophage features. J Tissue Eng Regen Med. 2017;11:2895–911. https://doi.org/10.1002/term.2193.
- Neves LAA, Stovall K, Joyner J, Valdés G, Gallagher PE, Ferrario CM, et al. ACE2 and ANG-(1–7) in the rat uterus during early and late gestation. Am J Physiol Regul Integr Comp Physiol. 2008;294:R151–61. https://doi.org/10.1152/ajpregu.00514.2007.
- Maron BA, Leopold JA. The role of the renin-angiotensin-aldosterone system in the pathobiology of pulmonary arterial hypertension (2013 Grover Conference series). Pulmon Circ. 2014;4:200–10. https://doi.org/ 10.1086/675984.
- Lemke A, Castillo-Sánchez JC, Prodinger F, Ceranic A, Hennerbichler-Lugscheider S, Pérez-Gil J, et al. Human amniotic membrane as newly identified source of amniotic fluid pulmonary surfactant. Sci Rep. 2017;7:6406. https://doi.org/10.1038/s41598-017-06402-w.
- Piva S, DiBlasi RM, Slee AE, Jobe AH, Roccaro AM, Filippini M, et al. Surfactant therapy for COVID-19 related ARDS: a retrospective casecontrol pilot study. Respir Res. 2021;22:20. https://doi.org/10.1186/ s12931-020-01603-w
- 24. Ng JJ, Liang ZC, Choong AMTL. The incidence of pulmonary thromboembolism in COVID-19 patients admitted to the intensive care unit: a meta-analysis and meta-regression of observational studies. J Intensive Care. 2021;9:20. https://doi.org/10.1186/s40560-021-00535-x.
- Longchamp G, Manzocchi-Besson S, Longchamp A, Righini M, Robert-Ebadi H, Blondon M. Proximal deep vein thrombosis and pulmonary embolism in COVID-19 patients: a systematic review and meta-analysis. Thromb J. 2021;19:15. https://doi.org/10.1186/s12959-021-00266-x.
- Niknejad H, Yazdanpanah G, Kakavand M. Extract of fetal membrane would inhibit thrombosis and hemolysis. Med Hypotheses. 2015. https://doi.org/10.1016/j.mehy.2015.04.030.
- Murphy S, Lim R, Dickinson H, Acharya R, Rosli S, Jenkin G, et al. Human amnion epithelial cells prevent bleomycin-induced lung injury and preserve lung function. Cell Transplant. 2011;20:909–23. https://doi.org/ 10.3727/096368910X543385.
- 28. Udwadia ZF, Koul PA, Richeldi L. Post-COVID lung fibrosis: the tsunami that will follow the earthquake. Lung India Medknow. 2021;38:S41–7. https://doi.org/10.4103/lungindia.lungindia_818_20.
- Evangelista M, Soncini M, Parolini O. Placenta-derived stem cells: new hope for cell therapy? Cytotechnology. 2008;58:33–42. https://doi.org/ 10.1007/s10616-008-9162-z.
- Ilancheran S, Michalska A, Peh G, Wallace EM, Pera M, Manuelpillai U. Stem cells derived from human fetal membranes display multilineage differentiation potential. Biol Reprod. 2007;77:577–88. https://doi.org/ 10.1095/biolreprod.106.055244.
- Miki T, Strom SC. Amnion-derived pluripotent/multipotent stem cells. Stem Cell Rev. 2006;2:133–42. https://doi.org/10.1007/ s12015-006-0020-0.
- Murphy S, Rosli S, Acharya R, Mathias L, Lim R, Wallace E, et al. Amnion epithelial cell isolation and characterization for clinical use. Curr Protoc Stem Cell Biol. 2010. https://doi.org/10.1002/9780470151808.sc01e 06s13
- Miki T. Stem cell characteristics and the therapeutic potential of amniotic epithelial cells. Am J Reprod Immunol. 2018. https://doi.org/10. 1111/aji.13003.
- Ghamari S-H, Abbasi-Kangevari M, Tayebi T, Bahrami S, Niknejad H. The bottlenecks in translating placenta-derived amniotic epithelial and mesenchymal stromal cells into the clinic: current discrepancies in marker reports. Front Bioeng Biotechnol. 2020;8:180. https://doi.org/10. 3389/fbioe.2020.00180.
- Barboni B, Russo V, Curini V, Martelli A, Berardinelli P, Mauro A, et al. Gestational stage affects amniotic epithelial cells phenotype, methylation status, immunomodulatory and stemness properties. Stem Cell Rev. 2014;10:725–41. https://doi.org/10.1007/s12015-014-9519-y.

- Izumi M, Pazin BJ, Minervini CF, Gerlach J, Ross MA, Stolz DB, et al. Quantitative comparison of stem cell marker-positive cells in fetal and term human amnion. J Reprod Immunol. 2009;81:39–43. https://doi.org/10.1016/j.jri.2009.02.007.
- Lee AS, Tang C, Rao MS, Weissman IL, Wu JC. Tumorigenicity as a clinical hurdle for pluripotent stem cell therapies. Nat Med. 2013. https://doi. org/10.1038/nm.3267.
- Tian Y-B, Wang N-X, Xu Y, Yu C-Y, Liu R-M, Luo Y, et al. Hyaluronic acid ameliorates the proliferative ability of human amniotic epithelial cells through activation of TGF-β/BMP signaling. PeerJ. 2020. https://doi.org/ 10.7717/peerj.10104.
- Niknejad H, Deihim T, Peirovi H, Abolghasemi H. Serum-free cryopreservation of human amniotic epithelial cells before and after isolation from their natural scaffold. Cryobiology. 2013;67:56–63. https://doi.org/10.1016/j.cryobiol.2013.05.001.
- Motedayyen H, Esmaeil N, Tajik N, Khadem F, Ghotloo S, Khani B, et al. Method and key points for isolation of human amniotic epithelial cells with high yield, viability and purity. BMC Res Notes. 2017;10:552. https://doi.org/10.1186/s13104-017-2880-6.
- Biniazan F, Manzari-Tavakoli A, Safaeinejad F, Moghimi A, Rajaei F, Niknejad H. The differentiation effect of bone morphogenetic protein (BMP) on human amniotic epithelial stem cells to express ectodermal lineage markers. Cell Tissue Res. 2021;383:751–63. https://doi.org/10. 1007/s00441-020-03280-z.
- Miki T, Marongiu F, Dorko K, Ellis ECS, Strom SC. Isolation of amniotic epithelial stem cells. Curr Protoc Stem Cell Biol. 2010; Chapter 1: Unit 1E.3. https://doi.org/10.1002/9780470151808.sc01e03s12.
- 43. Centurione L, Passaretta F, Centurione MA, Munari SD, Vertua E, Silini A, et al. Mapping of the human placenta: experimental evidence of amniotic epithelial cell heterogeneity. Cell Transplant. 2018;27:12–22. https://doi.org/10.1177/0963689717725078.
- García-Castro IL, García-López G, Ávila-González D, Flores-Herrera H, Molina-Hernández A, Portillo W, et al. Markers of pluripotency in human amniotic epithelial cells and their differentiation to progenitor of cortical neurons. PLoS ONE. 2015;10:e0146082. https://doi.org/10.1371/ journal.pone.0146082.
- Spitzhorn L-S, Rahman MS, Schwindt L, Ho H-T, Wruck W, Bohndorf M, et al. Isolation and molecular characterization of amniotic fluid-derived mesenchymal stem cells obtained from Caesarean sections. Stem Cells Int. 2017;2017:5932706. https://doi.org/10.1155/2017/5932706.
- Miki T, Lehmann T, Cai H, Stolz DB, Strom SC. Stem cell characteristics of amniotic epithelial cells. Stem Cells. 2005. https://doi.org/10.1634/ stemcells.2004-0357.
- Lim R, Hodge A, Moore G, Wallace EM, Sievert W. A pilot study evaluating the safety of intravenously administered human amnion epithelial cells for the treatment of hepatic fibrosis. Front Pharmacol. 2017;8:549. https://doi.org/10.3389/fphar.2017.00549.
- Leng Z, Zhu R, Hou W, Feng Y, Yang Y, Han Q, et al. Transplantation of ACE2-mesenchymal stem cells improves the outcome of patients with COVID-19 pneumonia. Aging Dis. 2020. https://doi.org/10.14336/ad. 2020.0228.
- Guo Z, Chen Y, Luo X, He X, Zhang Y, Wang J. Administration of umbilical cord mesenchymal stem cells in patients with severe COVID-19 pneumonia. Crit Care. 2020;24:420. https://doi.org/10.1186/ s13054-020-03142-8.
- Farhadihosseinabadi B, Farahani M, Tayebi T, Jafari A, Biniazan F, Modaresifar K, et al. Amniotic membrane and its epithelial and mesenchymal stem cells as an appropriate source for skin tissue engineering and regenerative medicine. Artif Cells Nanomed Biotechnol. 2018;46:431–40. https://doi.org/10.1080/21691401.2018.1458730.
- Niknejad H, Peirovi H, et al. Properties of the amniotic membrane for potential use in tissue engineering. Eur Cells Mater. 2008. https://doi. org/10.22203/ecm.v015a07.
- Andrewartha N, Yeoh G. Human Amnion Epithelial Cell Therapy for Chronic Liver Disease. Stem Cells Int. 2019;2019:8106482. https://doi. org/10.1155/2019/8106482.
- Uchida S, Inanaga Y, Kobayashi M, Hurukawa S, Araie M, Sakuragawa N. Neurotrophic function of conditioned medium from human amniotic epithelial cells. J Neurosci Res. 2000;62:585–90. https://doi.org/10.1002/ 1097-4547(20001115)62:4%3c585::AID-JNR13%3e3.0.CO;2-U.

- Yang S, Sun H-M, Yan J-H, Xue H, Wu B, Dong F, et al. Conditioned medium from human amniotic epithelial cells may induce the differentiation of human umbilical cord blood mesenchymal stem cells into dopaminergic neuron-like cells. J Neurosci Res. 2013;91:978–86. https:// doi.org/10.1002/jnr.23225.
- Wu Q, Fang T, Lang H, Chen M, Shi P, Pang X, et al. Comparison of the proliferation, migration and angiogenic properties of human amniotic epithelial and mesenchymal stem cells and their effects on endothelial cells. Int J Mol Med. 2017;39:918–26. https://doi.org/10.3892/ijmm.2017. 2897
- Jafari A, Babajani A, Abdollahpour-Alitappeh M, Ahmadi N, Rezaei-Tavirani M. Exosomes and cancer: from molecular mechanisms to clinical applications. Med Oncol. 2021. https://doi.org/10.1007/ s12032-021-01491-0.
- Yin K, Wang S, Zhao RC. Exosomes from mesenchymal stem/stromal cells: a new therapeutic paradigm. Biomark Res. 2019;7:8. https://doi. org/10.1186/s40364-019-0159-x.
- Wang X, Chen Y, Zhao Z, Meng Q, Yu Y, Sun J, et al. Engineered exosomes with ischemic myocardium-targeting peptide for targeted therapy in myocardial infarction. J Am Heart Assoc. 2018;7:e008737. https://doi.org/10.1161/JAHA.118.008737.
- Gu J, Han B, Wang J. COVID-19: Gastrointestinal manifestations and potential fecal-oral transmission. Gastroenterology. 2020;158:1518–9.
- Werion A, Belkhir L, Perrot M, Schmit G, Aydin S, Chen Z, et al. SARS-CoV-2 causes a specific dysfunction of the kidney proximal tubule. Kidney Int. 2020;98:1296–307. https://doi.org/10.1016/j.kint.2020.07. 019
- Solomon IH, Normandin E, Bhattacharyya S, Mukerji SS, Keller K, Ali AS, et al. Neuropathological features of Covid-19. N Engl J Med. 2020;383:989–92. https://doi.org/10.1056/NEJMc2019373.
- Ragab D, Salah Eldin H, Taeimah M, Khattab R, Salem R. The COVID-19 cytokine storm; what we know so far. Front Immunol. 2020;11:1446. https://doi.org/10.3389/fimmu.2020.01446.
- Dalan R, Bornstein SR, El-Armouche A, Rodionov RN, Markov A, Wielockx B, et al. The ACE-2 in COVID-19: foe or friend? Horm Metab Res. 2020;52:257–63. https://doi.org/10.1055/a-1155-0501.
- Pourani MR, Abdollahimajd F. CD147 as a novel receptor in the pathogenesis of SARS-CoV-2: Is there any correlation with the risk of COVID-19 in dermatological diseases? Dermatol Ther. 2020. https://doi.org/10. 1111/dth.14443.
- 65. Acosta Saltos F, Acosta Saltos AD. Entry of SARS-CoV2 through the basal surface of alveolar endothelial cells—a proposed mechanism mediated by CD147 in Covid-19. MDPI AG; 2020. https://www.researchgate.net/profile/Alejandro-Acosta-Saltos/publication/341618106_Entry_of_SARS-CoV2_through_the_Basal_Surface_of_Alveolar_Endothelial_Cells__A_Proposed_Mechanism_Mediated_by_CD147_in_COVID-19/links/5f52091d299bf13a319e9d99/Entry-of-SARS-CoV2-through-the-Basal-Surface_of-Alveolar-Endothelial-Cells-A-Proposed-Mechanism-Mediated-by-CD147-in-COVID-19.pdf.
- Ibrahim IM, Abdelmalek DH, Elshahat ME, Elfiky AA. COVID-19 spikehost cell receptor GRP78 binding site prediction. J Infect. 2020;80:554– 62. https://doi.org/10.1016/j.jinf.2020.02.026.
- Bittmann S. The role of TMPRSS2 and TMPRSS2-inhibitors in cell entry mechanism of COVID-19. J Regen Biol Med. 2020. https://doi.org/10. 37191/mapsci-2582-385x-2(3)-026.
- Piva F, Sabanovic B, Cecati M, Giulietti M. Expression and co-expression analyses of TMPRSS2, a key element in COVID-19. Eur J Clin Microbiol Infect Dis. 2021;40:451–5. https://doi.org/10.1007/s10096-020-04089-y.
- Tan Y, Tang F. SARS-CoV-2-mediated immune system activation and potential application in immunotherapy. Med Res Rev. 2021;41:1167– 94. https://doi.org/10.1002/med.21756.
- Lowery SA, Sariol A, Perlman S. Innate immune and inflammatory responses to SARS-CoV-2: implications for COVID-19. Cell Host Microbe. 2021;29:1052–62. https://doi.org/10.1016/j.chom.2021.05.004.
- Zheng J, Wang Y, Li K, Meyerholz DK, Allamargot C, Perlman S. Severe acute respiratory syndrome Coronavirus 2-induced immune activation and death of monocyte-derived human macrophages and dendritic cells. J Infect Dis. 2021;223:785–95. https://doi.org/10.1093/infdis/jiaa7 53.

- Salvi V, Nguyen HO, Sozio F, Schioppa T, Gaudenzi C, Laffranchi M, et al. SARS-CoV-2-associated ssRNAs activate inflammation and immunity via TLR7/8. JCI Insight. 2021. https://doi.org/10.1172/jci.insight.150542.
- Manik M, Singh RK. Role of toll-like receptors in modulation of cytokine storm signaling in SARS-CoV-2-induced COVID-19. J Med Virol. 2021;94:869–77. https://doi.org/10.1002/jmv.27405.
- Gadanec LK, McSweeney KR, Qaradakhi T, Ali B, Zulli A, Apostolopoulos V. Can SARS-CoV-2 virus use multiple receptors to enter host cells? Int J Mol Sci. 2021;22:992. https://doi.org/10.3390/ijms22030992.
- Hariharan A, Hakeem AR, Radhakrishnan S, Reddy MS, Rela M. The role and therapeutic potential of NF-kappa-B pathway in severe COVID-19 patients. Inflammopharmacology. 2021;29:91–100. https://doi.org/10. 1007/s10787-020-00773-9.
- Babajani A, Hosseini-Monfared P, Abbaspour S, Jamshidi E, Niknejad H. Targeted mitochondrial therapy with over-expressed MAVS protein from mesenchymal stem cells: a new therapeutic approach for COVID-19. Front Cell Dev Biol. 2021;9: 695362. https://doi.org/10.3389/fcell. 2021.695362.
- Acharya D, Liu G, Gack MU. Dysregulation of type I interferon responses in COVID-19. Nat Rev Immunol. 2020;20:397–8. https://doi.org/10.1038/ s41577-020-0346-x
- Prompetchara E, Ketloy C, Palaga T. Immune responses in COVID-19 and potential vaccines: Lessons learned from SARS and MERS epidemic. Asian Pac J Allergy Immunol. 2020;38:1–9. https://doi.org/10.12932/ AP-200220-0772.
- Zhang D, Guo R, Lei L, Liu H, Wang Y, Wang Y, et al. Frontline Science: COVID-19 infection induces readily detectable morphologic and inflammation-related phenotypic changes in peripheral blood monocytes. J Leukoc Biol. 2021;109:13–22. https://doi.org/10.1002/JLB.4HI07 20-470R.
- 80. He R, Lu Z, Zhang L, Fan T, Xiong R, Shen X, et al. The clinical course and its correlated immune status in COVID-19 pneumonia. J Clin Virol. 2020;127: 104361. https://doi.org/10.1016/j.jcv.2020.104361.
- 81. Carcaterra M, Caruso C. Alveolar epithelial cell type II as main target of SARS-CoV-2 virus and COVID-19 development via NF-Kb pathway deregulation: a physio-pathological theory. Med Hypotheses. 2021;146: 110412. https://doi.org/10.1016/j.mehy.2020.110412.
- Bouayad A. Innate immune evasion by SARS-CoV-2: comparison with SARS-CoV. Rev Med Virol. 2020;30:1–9. https://doi.org/10.1002/rmv. 2135.
- 83. De Biasi S, Meschiari M, Gibellini L, Bellinazzi C, Borella R, Fidanza L, et al. Marked T cell activation, senescence, exhaustion and skewing towards TH17 in patients with COVID-19 pneumonia. Nat Commun. 2020;11:3434. https://doi.org/10.1038/s41467-020-17292-4.
- Webb BJ, Peltan ID, Jensen P, Hoda D, Hunter B, Silver A, et al. Clinical criteria for COVID-19-associated hyperinflammatory syndrome: a cohort study. Lancet Rheumatol. 2020;2:e754–63. https://doi.org/10.1016/ S2665-9913(20)30343-X.
- Baker EK, Malhotra A, Lim R, Jacobs SE, Hooper SB, Davis PG, et al. Human amnion cells for the prevention of bronchopulmonary dysplasia: a protocol for a phase I dose escalation study. BMJ Open. 2019;9:e026265. https://doi.org/10.1136/bmjopen-2018-026265.
- 86. Vosdoganes P, Hodges RJ, Lim R, Westover AJ, Acharya RY, Wallace EM, et al. Human amnion epithelial cells as a treatment for inflammation-induced fetal lung injury in sheep. Am J Obstet Gynecol. 2011;205(156):e26-33.
- Jamshidi E, Babajani A, Soltani P, Niknejad H. Proposed mechanisms of targeting COVID-19 by delivering mesenchymal stem cells and their exosomes to damaged organs. Stem Cell Rev Rep. 2021;17:176–92. https://doi.org/10.1007/s12015-020-10109-3.
- Moodley Y, Ilancheran S, Samuel C, Vaghjiani V, Atienza D, Williams ED, et al. Human amnion epithelial cell transplantation abrogates lung fibrosis and augments repair. Am J Respir Crit Care Med. 2010;182:643– 51. https://doi.org/10.1164/rccm.201001-0014OC.
- 89. Koizumi NJ, Inatomi TJ, Sotozono CJ, Fullwood NJ, Quantock AJ, Kinoshita S. Growth factor mRNA and protein in preserved human amniotic membrane. Curr Eye Res. 2000;20:173.
- McDonald CA, Payne NL, Sun G, Moussa L, Siatskas C, Lim R, et al. Immunosuppressive potential of human amnion epithelial cells in the treatment of experimental autoimmune encephalomyelitis. J Neuroinflammation. 2015;12:112. https://doi.org/10.1186/s12974-015-0322-8.

- 91. Murphy SV, Shiyun SC, Tan JL, Chan S, Jenkin G, Wallace EM, et al. Human amnion epithelial cells do not abrogate pulmonary fibrosis in mice with impaired macrophage function. Cell Transplant. 2012;21:1477–92. https://doi.org/10.3727/096368911X601028.
- Strom SC, Gramignoli R. Human amnion epithelial cells expressing HLA-G as novel cell-based treatment for liver disease. Hum Immunol. 2016;77:734–9. https://doi.org/10.1016/j.humimm.2016.07.002.
- Magatti M, Caruso M, De Munari S, Vertua E, De D, Manuelpillai U, et al. Human amniotic membrane-derived mesenchymal and epithelial cells exert different effects on monocyte-derived dendritic cell differentiation and function. Cell Transplant. 2015;24:1733–52. https://doi.org/10. 3727/096368914X684033.
- Tan JL, Chan ST, Wallace EM, Lim R. Human amnion epithelial cells mediate lung repair by directly modulating macrophage recruitment and polarization. Cell Transplant. 2014;23:319–28. https://doi.org/10. 3727/096368912X661409.
- Liu YH, Vaghjiani V, Tee JY, To K, Cui P, Oh DY, et al. Amniotic epithelial cells from the human placenta potently suppress a mouse model of multiple sclerosis. PLoS ONE. 2012;7: e35758. https://doi.org/10.1371/journal.pone.0035758.
- Tan JL, Chan ST, Lo CY, Deane JA, McDonald CA, Bernard CC, et al. Amnion cell-mediated immune modulation following bleomycin challenge: controlling the regulatory T cell response. Stem Cell Res Ther. 2015;6:8. https://doi.org/10.1186/scrt542.
- Evans MA, Broughton BRS, Drummond GR, Ma H, Phan TG, Wallace EM, et al. Amnion epithelial cells—a novel therapy for ischemic stroke? Neural Regen Res. 2018;13:1346–9. https://doi.org/10.4103/1673-5374. 235223.
- Evans MA, Lim R, Kim HA, Chu HX, Gardiner-Mann CV, Taylor KWE, et al. Acute or delayed systemic administration of human amnion epithelial cells improves outcomes in experimental stroke. Stroke. 2018. https://doi.org/10.1161/strokeaha.117.019136.
- Stock SJ, Kelly RW, Riley SC, Calder AA. Natural antimicrobial production by the amnion. Am J Obstet Gynecol. 2007;196(255):e1-6. https://doi. org/10.1016/j.ajog.2006.10.908.
- Tehrani FA, Modaresifar K, Azizian S, Niknejad H. Induction of antimicrobial peptides secretion by IL-1β enhances human amniotic membrane for regenerative medicine. Sci Rep. 2017;7:17022. https://doi.org/10.1038/s41598-017-17210-7.
- 101. Marin-Luevano SP, Rodriguez-Carlos A, Jacobo-Delgado Y, Valdez-Miramontes C, Enciso-Moreno JA, Rivas-Santiago B. Steroid hormone modulates the production of cathelicidin and human β-defensins in lung epithelial cells and macrophages promoting Mycobacterium tuberculosis killing. Kekkaku. 2021;128:102080.
- 102. Suff N, Karda R, Diaz JA, Ng J, Baruteau J, Perocheau D, et al. Cervical gene delivery of the antimicrobial peptide, human β-defensin (HBD)-3, in a mouse model of ascending infection-related preterm birth. Front Immunol. 2020;11:106. https://doi.org/10.3389/fimmu.2020.00106.
- Tan JL, Lau SN, Leaw B, Nguyen HPT, Salamonsen LA, Saad MI, et al. Amnion epithelial cell-derived exosomes restrict lung injury and enhance endogenous lung repair. Stem Cells Transl Med. 2018;7:180– 96. https://doi.org/10.1002/sctm.17-0185.
- Lambert DW, Hooper NM, Turner AJ. Angiotensin-converting enzyme 2 and new insights into the renin-angiotensin system. Biochem Pharmacol. 2008;75:781–6. https://doi.org/10.1016/j.bcp.2007.08.012.
- Shereen MA, Khan S, Kazmi A, Bashir N, Siddique R. COVID-19 infection: origin, transmission, and characteristics of human coronaviruses. J Advert Res. 2020;24:91–8. https://doi.org/10.1016/j.jare.2020.03.005.
- Kuba K, Imai Y, Rao S, Gao H, Guo F, Guan B, et al. A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus-induced lung injury. Nat Med. 2005;11:875–9. https://doi.org/10.1038/nm1267.
- Samavati L, Uhal BD. ACE2, much more than just a receptor for SARS-COV-2. Front Cell Infect Microbiol. 2020;10:317. https://doi.org/10.3389/ fcimb.2020.00317.
- Kuba K, Imai Y, Rao S, Jiang C, Penninger JM. Lessons from SARS: control of acute lung failure by the SARS receptor ACE2. J Mol Med. 2006;84:814–20. https://doi.org/10.1007/s00109-006-0094-9.
- 109. Wong YP, Khong TY, Tan GC. The effects of COVID-19 on placenta and pregnancy: what do we know so far? Diagnostics. 2021;11:94.
- Kryvenko V, Vadász I. Molecular mechanisms of Na, K-ATPase dysregulation driving alveolar epithelial barrier failure in severe COVID-19. Am J

- Physiol Lung Cell Mol Physiol. 2021;320:L1186–93. https://doi.org/10. 1152/ajplung.00056.2021.
- 111. Eisenhut M, Shin Jl. Pathways in the pathophysiology of coronavirus 19 lung disease accessible to prevention and treatment. Front Physiol. 2020;11:872. https://doi.org/10.3389/fphys.2020.00872.
- Mirastschijski U, Schwab I, Coger V, Zier U, Rianna C, He W, et al. Lung surfactant accelerates skin wound healing: a translational study with a randomized clinical phase I study. Sci Rep. 2020;10:2581. https://doi. org/10.1038/s41598-020-59394-5.
- Mirastschijski U, Dembinski R, Maedler K. Lung surfactant for pulmonary barrier restoration in patients with COVID-19 pneumonia. Front Med. 2020;7:254. https://doi.org/10.3389/fmed.2020.00254.
- Spragg RG, Lewis JF, Walmrath H-D, Johannigman J, Bellingan G, Laterre P-F, et al. Effect of recombinant surfactant protein C-based surfactant on the acute respiratory distress syndrome. N Engl J Med. 2004;351:884–92. https://doi.org/10.1056/NEJMoa033181.
- Ujma S, Horsnell WGC, Katz AA, Clark HW, Schäfer G. Non-pulmonary immune functions of surfactant proteins A and D. J Innate Immun. 2017;9:3–11. https://doi.org/10.1159/000451026.
- Rowe TF, King LA, MacDonald PC, Casey ML. Tissue inhibitor of metalloproteinase-1 and tissue inhibitor of metalloproteinase-2 expression in human amnion mesenchymal and epithelial cells. Am J Obstet Gynecol. 1997;176:915–21.
- McDonald CA, Melville JM, Polglase GR, Jenkin G, Moss TJM. Maintenance of human amnion epithelial cell phenotype in pulmonary surfactant. Stem Cell Res Ther. 2014;5:107. https://doi.org/10.1186/scrt4 95
- Klok FA, Kruip MJHA, van der Meer NJM, Arbous MS, Gommers D, Kant KM, et al. Confirmation of the high cumulative incidence of thrombotic complications in critically ill ICU patients with COVID-19: an updated analysis. Thromb Res. 2020;191:148–50. https://doi.org/10.1016/j.throm res.2020.04.041.
- Monteil V, Kwon H, Prado P, Hagelkrüys A, Wimmer RA, Stahl M, et al. Inhibition of SARS-CoV-2 infections in engineered human tissues using clinical-grade soluble human ACE2. Cell. 2020;181:905-913.e7. https://doi.org/10.1016/j.cell.2020.04.004.
- García-Ortega A, de la Rosa D, Oscullo G, Castillo-Villegas D, López-Reyes R, Martínez-García MÁ. Coagulation disorders and thromboembolic disease in COVID-19: review of current evidence in search of a better approach. J Thorac Dis. 2021;13:1239–55.
- Jahr S, Hentze H, Englisch S, Hardt D, Fackelmayer FO, Hesch RD, et al. DNA fragments in the blood plasma of cancer patients: quantitations and evidence for their origin from apoptotic and necrotic cells. Cancer Res. 2001;61:1659–65.
- 122. Thiam HR, Wong SL, Wagner DD, Waterman CM. Cellular Mechanisms of NETosis. Annu Rev Cell Dev Biol. 2020;36:191–218. https://doi.org/10.1146/annurev-cellbio-020520-111016.
- Nakazawa F, Kannemeier C, Shibamiya A, Song Y, Tzima E, Schubert U, et al. Extracellular RNA is a natural cofactor for the (auto-)activation of Factor VII-activating protease (FSAP). Biochem J. 2005. https://doi.org/ 10.1042/bj20041021.
- Kannemeier C, Shibamiya A, Nakazawa F, Trusheim H, Ruppert C, Markart P, et al. Extracellular RNA constitutes a natural procoagulant cofactor in blood coagulation. Proc Natl Acad Sci. 2007. https://doi.org/ 10.1073/pnas.0608647104.
- Becker RC. COVID-19 update: Covid-19-associated coagulopathy. J Thromb Thrombolysis. 2020;50:54–67. https://doi.org/10.1007/ s11239-020-02134-3.
- Miesbach W, Makris M. COVID-19: coagulopathy, risk of thrombosis, and the rationale for anticoagulation. Clin Appl Thromb/Hemost. 2020. https://doi.org/10.1177/1076029620938149.
- Levi M, van der Poll T. Inflammation and coagulation. Crit Care Med. 2010;38:S26-34.
- Shovlin CL, Vizcaychipi MP. Vascular inflammation and endothelial injury in SARS-CoV-2 infection: the overlooked regulatory cascades implicated by the ACE2 gene cluster. QJM. 2020. https://doi.org/10.1093/qjmed/ bcaa241
- Bautista-Vargas M, Bonilla-Abadía F, Cañas CA. Potential role for tissue factor in the pathogenesis of hypercoagulability associated with in COVID-19. J Thromb Thrombolysis. 2020;50:479–83. https://doi.org/10. 1007/s11239-020-02172-x.

- Jose RJ, Manuel A. COVID-19 cytokine storm: the interplay between inflammation and coagulation. Lancet Respir Med. 2020;8:e46–7. https://doi.org/10.1016/S2213-2600(20)30216-2.
- Polgar J, Matuskova J, Wagner DD. The P-selectin, tissue factor, coagulation triad. J Thromb Haemost. 2005. https://doi.org/10.1111/j.1538-7836.2005.01373.x
- Kakavand M, Yazdanpanah G, Ahmadiani A, Niknejad H. Blood compatibility of human amniotic membrane compared with heparin-coated ePTFE for vascular tissue engineering. J Tissue Eng Regen Med. 2017. https://doi.org/10.1002/term.2064.
- Chen J, Qi T, Liu L, Ling Y, Qian Z, Li T, et al. Clinical progression of patients with COVID-19 in Shanghai, China. J Infect. 2020;80:e1-6. https://doi.org/10.1016/j.jinf.2020.03.004.
- Li Z, Niu S, Guo B, Gao T, Wang L, Wang Y, et al. Stem cell therapy for COVID-19, ARDS and pulmonary fibrosis. Cell Prolif. 2020;53:e12939. https://doi.org/10.1111/cpr.12939.
- Spagnolo P, Balestro E, Aliberti S, Cocconcelli E, Biondini D, Della Casa G, et al. Pulmonary fibrosis secondary to COVID-19: a call to arms? Lancet Respir Med. 2020. https://doi.org/10.1016/s2213-2600(20)30222-8.
- Aggarwal G, Cheruiyot I, Aggarwal S, Wong J, Lippi G, Lavie CJ, et al. Association of cardiovascular disease with coronavirus disease 2019 (COVID-19) severity: a meta-analysis. Curr Probl Cardiol. 2020;45: 100617. https://doi.org/10.1016/j.cpcardiol.2020.100617.
- 137. Khashkhusha TR, Chan JSK, Harky A. ACE inhibitors and COVID-19: we don't know yet. J Card Surg. 2020;35:1172–3. https://doi.org/10.1111/jocs.14582.
- Zaim S, Chong JH, Sankaranarayanan V, Harky A. COVID-19 and multiorgan response. Curr Probl Cardiol. 2020;45:100618.
- Naicker S, Yang C-W, Hwang S-J, Liu B-C, Chen J-H, Jha V. The novel coronavirus 2019 epidemic and kidneys. Kidney Int. 2020;97:824–8. https://doi.org/10.1016/j.kint.2020.03.001.
- Wong SH, Lui RNS, Sung JJY. Covid-19 and the digestive system. J Gastroenterol Hepatol. 2020. https://doi.org/10.1111/jgh.15047.
- Xu L, Liu J, Lu M, Yang D, Zheng X. Liver injury during highly pathogenic human coronavirus infections. Liver Int. 2020;40:998–1004. https://doi. org/10.1111/liv.14435.
- 142. DeKosky ST, Kochanek PM, Valadka AB, Clark RSB, Chou SH-Y, Au AK, et al. Blood biomarkers for detection of brain injury in COVID-19 patients. J Neurotrauma. 2021;38:1–43. https://doi.org/10.1089/neu.
- 143. Guervilly C, Burtey S, Sabatier F, Cauchois R, Lano G, Abdili E, et al. Circulating endothelial cells as a marker of endothelial injury in severe COVID-19. J Infect Dis. 2020. https://doi.org/10.1093/infdis/jiaa528.
- Tsukamoto A, Abbot SE, Kadyk LC, DeWitt ND, Schaffer DV, Wertheim JA, et al. Challenging regeneration to transform medicine. Stem Cells Transl Med. 2016;5:1–7. https://doi.org/10.5966/sctm.2015-0180.
- Maddaluno L, Urwyler C, Werner S. Fibroblast growth factors: key players in regeneration and tissue repair. Development. 2017;144:4047–60. https://doi.org/10.1242/dev.152587.
- Zhang Q, Lai D. Application of human amniotic epithelial cells in regenerative medicine: a systematic review. Stem Cell Res Ther. 2020;11:439. https://doi.org/10.1186/s13287-020-01951-w.
- Liu Q-W, Huang Q-M, Wu H-Y, Zuo G-S-L, Gu H-C, Deng K-Y, et al. Characteristics and therapeutic potential of human amnion-derived stem cells. Int J Mol Sci. 2021;22:970. https://doi.org/10.3390/ijms22020970.
- 148. Ahmad S. Amnion epithelial cells: cell therapy going green! Sci Transl Med. American Association for the Advancement of Science; 2010. https://stm.sciencemag.org/content/2/48/48ec138.short. Cited 11 Sept 2021.
- 149. Zhao B, Li X, Shi X, Shi X, Zhang W, Wu G, et al. Exosomal microRNAs derived from human amniotic epithelial cells accelerate wound healing by promoting the proliferation and migration of fibroblasts. Stem Cells Int. 2018;2018:5420463. https://doi.org/10.1155/2018/5420463.
- Vosdoganes P, Lim R, Koulaeva E, Chan ST, Acharya R, Moss TJM, et al. Human amnion epithelial cells modulate hyperoxia-induced neonatal lung injury in mice. Cytotherapy. 2013;1021:9. https://doi.org/10.1016/j. jcyt.2013.03.004.

- Song Y-S, Joo H-W, Park I-H, Shen G-Y, Lee Y, Shin JH, et al. Transplanted human amniotic epithelial cells secrete paracrine proangiogenic cytokines in rat model of myocardial infarctio. Cell Transplant. 2015;24:2055–64. https://doi.org/10.3727/096368914X685609.
- Roy R, Kukucka M, Messroghli D, Kunkel D, Brodarac A, Klose K, et al. Epithelial-to-mesenchymal transition enhances the cardioprotective capacity of human amniotic epithelial cells. Cell Transplant. 2015;24:985–1002. https://doi.org/10.3727/096368913X675151.
- 153. Ren Y, Chen Y, Zheng X, Wang H, Kang X, Tang J, et al. Human amniotic epithelial cells ameliorate kidney damage in ischemia-reperfusion mouse model of acute kidney injury. Stem Cell Res Ther. 2020;11:410. https://doi.org/10.1186/s13287-020-01917-y.
- 154. Liu J, Hua R, Gong Z, Shang B, Huang Y, Guo L, et al. Human amniotic epithelial cells inhibit CD4T cell activation in acute kidney injury patients by influencing the miR-101-c-Rel-IL-2 pathway. Mol Immunol. 2017. https://doi.org/10.1016/j.molimm.2016.11.019.
- Ilancheran S, Moodley Y, Manuelpillai U. Human fetal membranes: a source of stem cells for tissue regeneration and repair? Placenta. 2009;30:2–10. https://doi.org/10.1016/j.placenta.2008.09.009.
- Marongiu F, Gramignoli R, Dorko K, Miki T, Ranade AR, Paola Serra M, et al. Hepatic differentiation of amniotic epithelial cells. Hepatology. 2011;53:1719–29. https://doi.org/10.1002/hep.24255.
- Manuelpillai U, Tchongue J, Lourensz D, Vaghjiani V, Samuel CS, Liu A, et al. Transplantation of human amnion epithelial cells reduces hepatic fibrosis in immunocompetent CCl4-treated mice. Cell Transplant. 2010. https://doi.org/10.3727/096368910x504496.
- Xu H, Zhang J, Tsang KS, Yang H, Gao W-Q. Therapeutic potential of human amniotic epithelial cells on injuries and disorders in the central nervous system. Stem Cells Int. 2019;2019:5432301. https://doi.org/10. 1155/2019/5432301.
- 159. Moeinabadi-Bidgoli K, Babajani A, Yazdanpanah G, Farhadihosseinabadi B, Jamshidi E, Bahrami S, et al. Translational insights into stem cell preconditioning: from molecular mechanisms to preclinical applications. Biomed Pharmacother. 2021;142:112026.
- Wei L, Fraser JL, Lu Z-Y, Hu X, Yu SP. Transplantation of hypoxia preconditioned bone marrow mesenchymal stem cells enhances angiogenesis and neurogenesis after cerebral ischemia in rats. Neurobiol Dis. 2012;46:635–45. https://doi.org/10.1016/j.nbd.2012.03.002.
- 161. Wang X, Shen K, Wang J, Liu K, Wu G, Li Y, et al. Hypoxic preconditioning combined with curcumin promotes cell survival and mitochondrial quality of bone marrow mesenchymal stem cells, and accelerates cutaneous wound healing via PGC-1α/SIRT3/HIF-1α signaling. Free Radic Biol Med. 2020;159:164–76. https://doi.org/10.1016/j.freeradbiomed.2020.7.023
- 162. Malik YS, Sheikh MA, Zhu X. Doxycycline can stimulate cytoprotection in neural stem cells with oxygen-glucose deprivation-reoxygenation injury: a potential approach to enhance effectiveness of cell transplantation therapy. Biochem Biophys Res Commun. 2013;432:355–8. https:// doi.org/10.1016/j.bbrc.2013.01.097.
- 163. Mena HA, Zubiry PR, Dizier B, Schattner M, Boisson-Vidal C, Negrotto S. Acidic preconditioning of endothelial colony-forming cells (ECFC) promote vasculogenesis under proinflammatory and high glucose conditions in vitro and in vivo. Stem Cell Res Ther. 2018;9:120. https:// doi.org/10.1186/s13287-018-0872-7.
- Somers VK, Kara T, Xie J. Progressive hypoxia: a pivotal pathophysiologic mechanism of COVID-19 pneumonia. Mayo Clin Proc. 2020;95:2339–42. https://doi.org/10.1016/j.mayocp.2020.09.015.
- Chhetri S, Khamis F, Pandak N, Al Khalili H, Said E, Petersen E. A fatal case of COVID-19 due to metabolic acidosis following dysregulate inflammatory response (cytokine storm). IDCases. 2020;21:e00829. https://doi. org/10.1016/j.idcr.2020.e00829.
- Francis KR, Wei L. Human embryonic stem cell neural differentiation and enhanced cell survival promoted by hypoxic preconditioning. Cell Death Dis. 2010;1: e22. https://doi.org/10.1038/cddis.2009.22.
- Kakudo N, Morimoto N, Ogawa T, Taketani S, Kusumoto K. Hypoxia enhances proliferation of human adipose-derived stem cells via HIF-1a

- activation. PLoS ONE. 2015;10: e0139890. https://doi.org/10.1371/journal.pone.0139890.
- Sakata H, Niizuma K, Yoshioka H, Kim GS, Jung JE, Katsu M, et al. Minocycline-preconditioned neural stem cells enhance neuroprotection after ischemic stroke in rats. J Neurosci. 2012;32:3462–73. https://doi.org/10.1523/JNEUROSCI.5686-11.2012.
- Okajima F. Regulation of inflammation by extracellular acidification and proton-sensing GPCRs. Cell Signal. 2013. https://doi.org/10.1016/j.cells ig.2013.07.022.
- Laforge M, Elbim C, Frère C, Hémadi M, Massaad C, Nuss P, et al. Tissue damage from neutrophil-induced oxidative stress in COVID-19. Nat Rev Immunol. 2020;20:515–6. https://doi.org/10.1038/s41577-020-0407-1.
- Kubo M, Li T-S, Suzuki R, Shirasawa B, Morikage N, Ohshima M, et al. Hypoxic preconditioning increases survival and angiogenic potency of peripheral blood mononuclear cells via oxidative stress resistance. Am J Physiol Heart Circ Physiol. 2008;294:H590–5. https://doi.org/10.1152/ aipheart.00856.2007.
- 172. Garrido-Pascual P, Alonso-Varona A, Castro B, Burón M, Palomares T. HO-preconditioned human adipose-derived stem cells (HC016) increase their resistance to oxidative stress by overexpressing Nrf2 and bioenergetic adaptation. Stem Cell Res Ther. 2020;11:335. https://doi.org/10.1186/s13287-020-01851-z.
- Rota C, Imberti B, Pozzobon M, Piccoli M, De Coppi P, Atala A, et al. Human amniotic fluid stem cell preconditioning improves their regenerative potential. Stem Cells Dev. 2012;21:1911–23. https://doi.org/10. 1089/scd.2011.0333.
- 174. Philipp D, Suhr L, Wahlers T, Choi Y-H, Paunel-Görgülü A. Preconditioning of bone marrow-derived mesenchymal stem cells highly strengthens their potential to promote IL-6-dependent M2b polarization. Stem Cell Res Ther. 2018;9:286. https://doi.org/10.1186/s13287-018-1039-2.
- Lin T, Pajarinen J, Nabeshima A, Lu L, Nathan K, Jämsen E, et al. Preconditioning of murine mesenchymal stem cells synergistically enhanced immunomodulation and osteogenesis. Stem Cell Res Ther. 2017;8:277. https://doi.org/10.1186/s13287-017-0730-z.
- Kim C-H, Lee S-H, Kim E-J, Ahn J-H, Choi E-J, Yoon J-U, et al. Effects of remifentanil preconditioning on factors related to uterine contraction in WISH cells. J Dent Anesth pain Med. 2019;19:343–51. https://doi.org/ 10.17245/jdapm.2019.19.6.343.
- Wu H-H, Zhou Y, Tabata Y, Gao J-Q. Mesenchymal stem cell-based drug delivery strategy: from cells to biomimetic. J Control Release. 2019;294:102–13. https://doi.org/10.1016/j.jconrel.2018.12.019.
- Thanuja MY, Anupama C, Ranganath SH. Bioengineered cellular and cell membrane-derived vehicles for actively targeted drug delivery: so near and yet so far. Adv Drug Deliv Rev. 2018;132:57–80. https://doi.org/10. 1016/j.addr.2018.06.012.
- 179. Babajani A, Soltani P, Jamshidi E, Farjoo MH, Niknejad H. Recent advances on drug-loaded mesenchymal stem cells with anti-neoplastic agents for targeted treatment of cancer. Front Bioeng Biotechnol. 2020;8:748. https://doi.org/10.3389/fbioe.2020.00748.
- Mencucci R, Paladini I, Menchini U, Gicquel JJ, Dei R. Inhibition of viral replication in vitro by antiviral-treated amniotic membrane. Possible use of amniotic membrane as drug-delivering tool. Br J Ophthalmol. 2011. https://doi.org/10.1136/bjo.2010.179556.
- Resch MD, Resch BE, Csizmazia E, Imre L, Németh J, Szabó-Révész P, et al. Drug reservoir function of human amniotic membrane. J Ocul Pharmacol Ther. 2011;27:323–6. https://doi.org/10.1089/jop.2011.0007.
- 182. Ho M-CD, Ring N, Amaral K, Doshi U, Li AP. Human enterocytes as an in vitro model for the evaluation of intestinal drug metabolism: characterization of drug-metabolizing enzyme activities of cryopreserved human enterocytes from twenty-four donors. Drug Metab Dispos. 2017. https://doi.org/10.1124/dmd.116.074377.
- Mencucci R, Menchini U, Dei R. Antimicrobial activity of antibiotictreated amniotic membrane. Cornea. 2006. https://doi.org/10.1097/01. ico.0000214207.06952.23.
- Liesveld JL, Sharma N, Aljitawi OS. Stem cell homing: from physiology to therapeutics. Stem Cells. 2020. https://doi.org/10.1002/stem.3242.

- Kollar K, Cook MM, Atkinson K, Brooke G. Molecular mechanisms involved in mesenchymal stem cell migration to the site of acute myocardial infarction. Int J Cell Biol. 2009;2009: 904682. https://doi.org/ 10.1155/2009/904682.
- 186. Meng S-S, Xu X-P, Chang W, Lu Z-H, Huang L-L, Xu J-Y, et al. LincRNA-p21 promotes mesenchymal stem cell migration capacity and survival through hypoxic preconditioning. Stem Cell Res Ther. 2018;9:280. https://doi.org/10.1186/s13287-018-1031-x.
- 187. Chang C-H, Lin Y-L, Tyan Y-S, Chiu Y-H, Liang Y-H, Chen C-P, et al. Interleukin-1β-induced matrix metalloproteinase-3 via ERK1/2 pathway to promote mesenchymal stem cell migration. PLoS ONE. 2021;16: e0252163. https://doi.org/10.1371/journal.pone.0252163.
- 188. van den Heuij LG, Fraser M, Miller SL, Jenkin G, Wallace EM, Davidson JO, et al. Delayed intranasal infusion of human amnion epithelial cells improves white matter maturation after asphyxia in preterm fetal sheep. J Cereb Blood Flow Metab. 2019;39:223–39. https://doi.org/10.1177/0271678X17729954.
- 189. Fischer UM, Harting MT, Jimenez F, Monzon-Posadas WO, Xue H, Savitz SI, et al. Pulmonary passage is a major obstacle for intravenous stem cell delivery: the pulmonary first-pass effect. Stem Cells Dev. 2009;18:683–92. https://doi.org/10.1089/scd.2008.0253.
- 190. Hodges RJ, Jenkin G, Hooper SB, Allison B, Lim R, Dickinson H, et al. Human amnion epithelial cells reduce ventilation-induced preterm lung injury in fetal sheep. Am J Obstet Gynecol. 2012;206(448):e8-15. https://doi.org/10.1016/j.ajog.2012.02.038.
- Kalimuthu S, Oh JM, Gangadaran P, Zhu L, Lee HW, Rajendran RL, et al. In vivo tracking of chemokine receptor CXCR4-engineered mesenchymal stem cell migration by optical molecular imaging. Stem Cells Int. 2017;2017;8085637. https://doi.org/10.1155/2017/8085637.
- Geffen Y, Perets N, Horev R, Yudin D, Oron O, Elliott E, et al. Exosomes derived from adipose mesenchymal stem cells: a potential non-invasive intranasal treatment for autism. Cytotherapy. 2020. https://doi.org/10. 1016/j.jcyt.2020.03.059.
- 193. Johnsen KB, Gudbergsson JM, Skov MN, Pilgaard L, Moos T, Duroux M. A comprehensive overview of exosomes as drug delivery vehicles—endogenous nanocarriers for targeted cancer therapy. Biochim Biophys Acta (BBA) Rev Cancer. 2014. https://doi.org/10.1016/j.bbcan.2014.04. 005
- 194. Arntz OJ, Pieters BCH, Oliveira MC, Broeren MGA, Bennink MB, de Vries M, et al. Oral administration of bovine milk derived extracellular vesicles attenuates arthritis in two mouse models. Mol Nutr Food Res. 2015;59:1701–12. https://doi.org/10.1002/mnfr.201500222.
- Yang P-J, Yuan W-X, Liu J, Li J-Y, Tan B, Qiu C, et al. Biological characterization of human amniotic epithelial cells in a serum-free system and their safety evaluation. Acta Pharmacol Sin. 2018;39:1305–16. https://doi.org/10.1038/aps.2018.22.
- 196. Fujikawa T, Oh S-H, Pi L, Hatch HM, Shupe T, Petersen BE. Teratoma formation leads to failure of treatment for type I diabetes using embryonic stem cell-derived insulin-producing cells. Am J Pathol. 2005;166:1781–91. https://doi.org/10.1016/S0002-9440(10)62488-1.
- Bongso A, Fong C-Y, Gauthaman K. Taking stem cells to the clinic: major challenges. J Cell Biochem. 2008;105:1352–60. https://doi.org/10.1002/ icb.21957.
- Vosdoganes P, Wallace EM, Chan ST, Acharya R, Moss TJM, Lim R. Human amnion epithelial cells repair established lung injury. Cell Transplant. 2013;22:1337–49. https://doi.org/10.3727/096368912X657657.
- Melville JM, McDonald CA, Bischof RJ, Polglase GR, Lim R, Wallace EM, et al. Human amnion epithelial cells modulate the inflammatory response to ventilation in preterm lambs. PLoS ONE. 2017;12: e0173572. https://doi.org/10.1371/journal.pone.0173572.
- Zhu D, Tan J, Maleken AS, Muljadi R, Chan ST, Lau SN, et al. Human amnion cells reverse acute and chronic pulmonary damage in experimental neonatal lung injury. Stem Cell Res Ther. 2017;8:257. https://doi. org/10.1186/s13287-017-0689-9.
- Papagianis PC, Ahmadi-Noorbakhsh S, Lim R, Wallace E, Polglase G, Jane Pillow J, et al. The effect of human amnion epithelial cells on lung development and inflammation in preterm lambs exposed to antenatal inflammation. PLoS ONE. 2021;16:e0253456. https://doi.org/10.1371/ journal.pone.0253456&type=printable.

- Bedford P, Jy J, Collins L, Keizer S. Considering cell therapy product "Good Manufacturing Practice" status. Front Med. 2018;5:118. https://doi.org/10.3389/fmed.2018.00118.
- Gramignoli R, Srinivasan RC, Kannisto K, Strom SC. Isolation of human amnion epithelial cells according to current good manufacturing procedures. Curr Protoc Stem Cell Biol. 2016;37:1E.10.1-1E.10.13. https://doi.org/10.1002/cpsc.2.
- 204. Lim R, Malhotra A, Tan J, Chan ST, Lau S, Zhu D, et al. First-in-human administration of allogeneic amnion cells in premature infants with bronchopulmonary dysplasia: a safety study. Stem Cells Transl Med. 2018;7:628–35. https://doi.org/10.1002/sctm.18-0079.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

