Antipneumococcal Activities of GAR-936 (a New Glycylcycline) Compared to Those of Nine Other Agents against Penicillin-Susceptible and -Resistant Pneumococci

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GAR-936, a new glycylcycline, had lower MICs (≤ 0.016 to 0.125 µg/ml) for 201 penicillin- and tetracyclinesusceptible and -resistant pneumococcal strains than tetracycline (≤ 0.06 to 128 µg/ml), minocycline (≤ 0.06 to 16.0 µg/ml), or doxycycline (≤ 0.06 to 32.0 µg/ml). GAR-936 was also bactericidal against 11 of 12 strains tested at the MIC after 24 h, with significant kill rates at earlier time points.

The incidence of pneumococcal resistance to penicillin G and other β -lactam and non- β -lactam compounds has increased worldwide at an alarming rate (1, 6, 7). In the United States, a recent survey of 1,476 strains showed that 49.6% were penicillin susceptible, 17.9% were of intermediate resistance (penicillin intermediate), and 32.5% were penicillin resistant. Macrolide MICs rose with those of penicillin G, with an overall macrolide resistance level of 33% (8).

Resistance to conventional oral tetracyclines (including minocycline and doxycycline) occurs with increased frequency among penicillin-intermediate and -resistant pneumococci, and genes coding for macrolide and tetracycline resistance are carried on the same transposon (6, 7). GAR-936 is a new broad-spectrum parenteral glycylcycline. Preliminary studies have shown that this drug is active against penicillin-susceptible and -resistant pneumococci (13, 15).

This study compares the activity of GAR-936 against 201 penicillin-susceptible, -intermediate, or -resistant pneumococcal strains with those of three other tetracyclines, four β -lactams, clarithromycin, and vancomycin. Additionally, the activities of these compounds against 12 penicillin-susceptible and -resistant strains of pneumococci were determined by the broth MIC and time-kill methods.

For determination of MICs by agar dilution, 51 penicillinsusceptible, 72 penicillin-intermediate, and 78 penicillin-resistant strains of pneumococci were utilized. For time-kill studies, four penicillin-susceptible, four penicillin-intermediate, and four penicillin-resistant strains were tested; of these, five were tetracycline susceptible (MIC, $\leq 2.0 \ \mu g/ml$), 1 was tetracycline intermediate (MIC, 4.0 $\mu g/ml$), and six were tetracycline resistant (MIC, $\geq 8.0 \ \mu g/ml$).

The agar dilution method used in this study to determine MICs of various antimicrobial agents for 201 pneumococcal strains has been used routinely for many years by our group (6, 7); Mueller-Hinton agar (BBL Microbiology Systems, Cockeysville, Md.) supplemented with 5% sheep blood was employed. Microbroth MIC determinations for 12 strains were

performed in accordance with the protocol recommended by the National Committee for Clinical Laboratory Standards (9). Standard quality control strains, including *Streptococcus pneumoniae* ATCC 49619 (9), were included in each run of agar and broth dilution MIC determinations. All MIC studies were performed in air. All strains, recently isolated from stocks stored at -70° C and subcultured twice, grew in air and did not require added CO₂ for growth. The susceptibility patterns of these strains are representative of those commonly encountered in the United States (8).

For time-kill studies, methods described previously were used (11, 12). The final inoculum was 5×10^5 to 5×10^6 CFU/ml. Growth controls with inoculum but no antibiotic were included with each experiment (11, 12). Viable-cell counts of antibiotic-containing suspensions were performed according to standard methods (11, 12). Colony counts were performed on plates yielding 30 to 300 colonies. The lower limit of sensitivity of colony counts was 300 CFU/ml. As mentioned above, all incubations were in ambient air without added CO₂.

Time-kill assays were analyzed by determining the number of strains which yielded a $\Delta \log_{10}$ CFU/ml of -1, -2, and -3 at 0, 4, 6, 12, and 24 h compared to counts at time zero. Antimicrobial agents were considered bactericidal at the lowest concentration that reduced the original inoculum by $\geq 3 \log_{10}$ CFU/ml (99.9%) at each of the time periods, and they were considered bacteriostatic if the inoculum was reduced by 0 to 3 log₁₀ CFU/ml. With the sensitivity threshold and inocula used in these studies, no problems were encountered in delineating 99.9% killing when it occurred. The problem of bacterial carryover was addressed by dilution, as described previously (11, 12). For tetracycline and clarithromycin time-kill testing, only strains with MICs of <8.0 and <4.0 µg/ml, respectively, were tested. In all cases, growth controls grew satisfactorily, without spontaneous autolysis.

The results of agar dilution MIC testing are presented in Table 1. GAR-936 had lower MICs (≤ 0.016 to 0.125 µg/ml) than tetracycline (≤ 0.06 to 128 µg/ml), minocycline (≤ 0.06 to 16.0 µg/ml), and doxycycline (≤ 0.06 to 32.0 µg/ml) against all strains. MICs of all β-lactams rose with those of penicillin G, with imipenem being the most active, followed by meropenem, ceftriaxone, and amoxicillin. Macrolide and tetracycline resistance occurred mainly in penicillin-intermediate

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TABLE 1. Agar dilution MICs for 201 pneumococcal isolates

Dimen	Penicillin	MIC (µg/ml)						
Drug	susceptibility of strain ^a	Range	$\mathrm{MIC}_{50}^{\ b}$	MIC ₉₀ ^c				
Penicillin	S	≤0.008-0.06	≤0.016	0.03				
	Ι	0.125 - 1.0	0.25	1.0				
	R	2.0 - > 8.0	2.0	4.0				
GAR-936	S	$\leq 0.016 - 0.125$	0.03	0.125				
	Ι	$\leq 0.016 - 0.125$	0.03	0.06				
	R	$\leq 0.016 - 0.125$	0.06	0.125				
Tetracycline	S	≤0.06-64.0	0.25	32.0				
	Ι	≤0.06-64.0	1.0	64.0				
	R	$\leq 0.25 - 128.0$	32.0	64.0				
Minocycline	S	$\leq 0.06 - 16.0$	0.06	8.0				
	Ι	$\leq 0.06 - 16.0$	0.25	16.0				
	R	$\leq 0.06 - 16.0$	4.0	16.0				
Doxycycline	S	0.03 - 16.0	0.125	8.0				
	Ι	≤0.06-32.0	0.5	8.0				
	R	0.125 - 16.0	4.0	8.0				
Amoxicillin	S	≤0.008-0.25	0.016	0.03				
	Ι	≤0.016-2.0	0.125	1.0				
	R	0.5 - 4.0	2.0	2.0				
Ceftriaxone	S	$\leq 0.008 - 1.0$	0.016	0.03				
	Ι	≤0.016-2.0	0.25	1.0				
	R	0.25 - 8.0	1.0	2.0				
Clarithromycin	S	$\leq 0.016 -> 128.0$	0.03	8.0				
	Ι	$\leq 0.016 -> 128.0$	0.06	>128.0				
	R	$\leq 0.016 -> 128.0$	1.0	>128.0				
Imipenem	S	≤0.008-0.25	≤ 0.008	0.016				
	Ι	≤0.008-0.25	0.06	0.25				
	R	0.03-0.5	0.25	0.5				
Meropenem	S	$\leq 0.008 - 0.5$	0.016	0.03				
	Ι	≤0.016-0.5	0.125	0.5				
	R	0.125 - 2.0	0.5	1.0				
Vancomycin	S	$\leq 0.06 - 0.5$	0.25	0.25				
-	Ι	$\leq 0.06 - 0.5$	0.25	0.25				
	R	0.125-0.5	0.25	0.25				

^a S, penicillin susceptible; I, penicillin intermediate; R, penicillin resistant.

^{*b*} MIC₅₀, MIC at which 50% of the isolates tested were inhibited.

^c MIC₉₀, MIC at which 90% of the isolates tested were inhibited.

and -resistant strains, and all strains were susceptible to vancomycin.

Microbroth MICs of the 12 strains tested by the time-kill method were either identical to or differed by one dilution from those obtained by agar dilution (Table 2). Time-kill results are presented in Table 3. GAR-936 was bactericidal at the MIC after 24 h for 11 of the 12 strains tested; the remaining strain (for which the GAR-936 MIC was $0.06 \mu g/ml$) was killed

at eight times the MIC (0.5 μ g/ml). GAR-936 showed significant kill rates at earlier time periods: at 12 h, 90% killing of all 12 strains occurred at twice the MIC, while after 6 h, 90% killing of all strains occurred at four times the MIC. The killing rates of other tetracyclines for the six susceptible strains were slightly lower than those of GAR-936. The bactericidal activities of β -lactams were similar, with uniform bactericidal activity being exhibited at twice the MIC for amoxicillin, ceftriaxone, and imipenem and at eight times the MIC for meropenem after 24 h. Clarithromycin MICs of <4.0 μ g/ml at twice the MIC after 24 h; vancomycin was bactericidal against 11 of 12 strains at twice the MIC at the same time point. The kill kinetics of GAR-936 against a tetracycline-resistant strain is depicted in Fig. 1.

The glycylcyclines are semisynthetic tetracyclines exhibiting a broad spectrum of activity compared to conventional tetracyclines, with activity against gram-positive and -negative aerobes, anaerobes, chlamydiae, and mycoplasmas (2–5, 10, 14, 16, 17). GAR-936 is a newly described 9-*t*-butylglycylamido derivative of minocycline. This compound possesses activity against isolates possessing the two major determinants of tetracycline resistance, ribosomal protection and efflux (13, 15).

In the present study, MICs of GAR-936 against all pneumococci, irrespective of their susceptibility to ß-lactams, macrolides, or other tetracyclines, ranged between ≤ 0.016 and 0.125 μ g/ml. These results are similar to those obtained by Petersen et al (13). Bactericidal activity was also found after 24 h for all 12 strains tested at $\leq 0.5 \mu \text{g/ml}$. At the MIC, after 24 h, GAR-936 showed better kill kinetics, relative to the MIC, than other tetracyclines and was bactericidal for more strains than were the ß-lactams tested. It should be emphasized, however, that he relatively small number of strains tested makes interpretation of relatively small differences in kill kinetics problematic. More strains must be tested before such conclusions can be considered valid. Although tetracyclines are traditionally thought to be bacteriostatic agents, the present study shows that GAR-936 (and other tetracyclines, relative to their MICs) may be bactericidal after 24 h.

Of the other tetracyclines tested, minocycline and doxycycline had lower MICs than tetracycline; the significance of this phenomenon is unknown at this time. The MICs of and timekills results for the other compounds tested are similar to those reported by others (7, 11, 12).

Although the results of toxicologic and pharmacokinetic studies are awaited, GAR-936 appears to be a promising

TABLE 2. Microbroth MICs for 12 strains tested by the time-kill method

Drug	MIC (µg/ml) for strain ^a :											
	1(S)	2(S)	3(S)	4(S)	5(I)	6(I)	7(I)	8(I)	9(R)	10(R)	11(R)	12(R)
Penicillin	0.06	0.016	0.03	0.03	0.5	0.5	0.25	0.25	4.0	2.0	4.0	4.0
GAR-936	0.06	0.03	0.03	0.06	0.125	0.06	0.03	0.03	0.03	0.03	0.06	0.06
Tetracycline	64.0	0.5	64.0	0.5	4.0	64.0	0.125	0.25	32.0	32.0	1.0	32.0
Minocycline	16.0	0.125	8.0	0.125	0.5	64.0	0.06	0.06	16.0	8.0	0.25	4.0
Doxycycline	8.0	0.25	16.0	0.25	1.0	16.0	0.06	0.125	8.0	8.0	1.0	8.0
Amoxicillin	0.03	0.03	0.016	0.016	0.06	0.125	0.125	0.06	1.0	1.0	2.0	2.0
Ceftriaxone	0.06	0.016	0.016	0.016	0.125	1.0	0.06	0.06	2.0	2.0	1.0	2.0
Clarithromycin	>64.0	0.03	0.016	0.03	>64.0	0.008	0.016	0.03	>64.0	>64.0	0.03	>64.0
Imipenem	0.008	0.008	0.008	0.008	0.03	0.06	0.03	0.03	0.25	0.25	0.5	0.25
Meropenem	0.016	0.016	0.016	0.008	0.06	0.125	0.125	0.06	1.0	0.5	1.0	1.0
Vancomycin	0.25	0.25	0.5	0.5	0.25	0.5	0.5	0.125	0.5	0.25	0.25	0.25

^a S, penicillin susceptible; I, penicillin intermediate; R, penicillin resistant.

Drug and concn	No. of strains killed by time point, with $\Delta \log_{10}$ CFU/ml (from time zero value) of:												
	3 h				6 h			12 h			24 h		
	-1	-2	-3	-1	-2	-3	-1	-2	-3	-1	-2	-3	
GAR-936	7	2	1	10	6	4	10	11	7	10	10	10	
$8 \times MIC$ $4 \times MIC$	7 6	3 2	1 1	12 12	6 6	4 4	12 12	$\begin{array}{c} 11 \\ 10 \end{array}$	7 6	12 12	12 12	12 11	
$2 \times MIC$	5	2	1	11	5	3	12	9	6	12	11	11	
MIC	4	1	1	7	5	2	10	8	4	12	11	11	
$0.5 \times MIC$	0	0	0	3	0	0	3	2	1	5	3	2	
Tetracycline ^a													
$8 \times MIC$	3	1	0	5	3	3	5	4	3	6	6	5	
$4 \times MIC$ $2 \times MIC$	2 2	0 0	$\begin{array}{c} 0\\ 0\end{array}$	4 3	3 2	0 0	5 5	4	2 2	6 6	6 5	5 5	
MIC	1	0	0	2	$\tilde{0}$	0	3	2 2	$\tilde{0}$	3	3	1	
$0.5 \times \text{MIC}$	0	0	0	0	0	0	1	0	0	0	0	0	
Minocycline ^a													
$8 \times MIC$	4	2	1	4	4	2	5	4	4	6	5	5	
$4 \times MIC$	3	1	1	4	3	2	5	4	3	6	5	5	
2× MIC MIC	2 2	$\begin{array}{c} 1\\ 0\end{array}$	$\begin{array}{c} 0\\ 0\end{array}$	4 3	$\frac{2}{2}$	$^{2}_{0}$	5 5	4 3	3 2	6 6	5 4	4 3	
$0.5 \times MIC$	1	0	0	1	$\tilde{0}$	0	0	0	$\tilde{0}$	0	0	0	
Doxycycline ^a													
8× MIC	4	2	0	5	3	3	5	5	3	6	5	5	
$4 \times MIC$	3	1	0	5	3	0	5	5	3	6	5	5	
$2 \times MIC$	1	0	0	4	1	0	5	4	1	6	5	4	
MIC $0.5 \times MIC$		0 0	0 0	2 1	$1 \\ 0$	0 0	2 1	$1 \\ 0$		2 0	$2 \\ 0$	2 0	
	0	0	0	1	0	0	1	0	0	0	0	0	
Amoxicillin 8× MIC	10	4	0	12	10	6	12	12	10	12	12	12	
$4 \times MIC$	9	4 2	0	12	9	6 3	12	12	8	12	12	12	
$2 \times MIC$	8	3	Ő	11	8	2	12	12	7	12	12	12	
MIC	5	2	0	11	4	2	10	5	3	10	8	8	
$0.5 \times MIC$	1	0	0	2	1	0	1	1	0	2	1	1	
Ceftriaxone	_	_	_			_							
$8 \times MIC$	8 8	$\begin{array}{c} 0 \\ 1 \end{array}$	$\begin{array}{c} 0\\ 0\end{array}$	12 12	10 9	$\begin{array}{c} 0 \\ 1 \end{array}$	12 12	12 11	10 8	12 12	12 12	12 12	
$4 \times MIC$ $2 \times MIC$	8 8		0	12	9 7		12	11 10	8 8	12	12	12	
MIC	5	1	ŏ	9	6	Ő	10	7	1	10	10	12	
$0.5 \times \text{MIC}$	2	0	0	2	0	0	1	1	0	0	0	0	
Clarithromycin ^b													
$8 \times MIC$	4	2	0	7	4	0	7	4	4	7	7	6	
$4 \times MIC$	3	1	0	6	3	0	7	4	4	7	6	6	
2× MIC MIC	3 2	1 1	0 0	4 4	3 2	$\begin{array}{c} 1\\ 0\end{array}$	7 6	4 4	4 3	7 7	6 6	6 4	
$0.5 \times MIC$	1	0	0	2	$\tilde{0}$	0	2	1	0	2	1	1	
Imipenem													
8× MIC	12	4	0	12	10	2	12	11	10	12	12	12	
$4 \times MIC$	11	5	0	12	9	2	12	11	10	12	12	12	
$2 \times MIC$	11	3	0	12	9	4	12	11	10	12	12	12	
MIC $0.5 \times MIC$		$ \begin{array}{c} 2\\ 0 \end{array} $	$\begin{array}{c} 0\\ 0\end{array}$	12 6	8 3	$ \begin{array}{c} 2\\ 0 \end{array} $	12 4	10 2	8 0	11 3	11 3	9 2	
$\frac{\text{Meropenem}}{8 \times \text{MIC}}$	11	3	1	12	5	2	12	12	9	12	12	12	
$4 \times MIC$	9	2	0	12	4	2	12	12	7	12	12	11	
$2 \times MIC$	6	4	0	12	7	3	12	11	8	12	12	11	
MIC $0.5 \times MIC$	6 1	2 0	$\begin{array}{c} 1\\ 0\end{array}$	11 2	5 0	$\frac{2}{0}$	11 1	9 0	5 0	$10 \\ 2$	9 1	8 1	
	1	U	0	-	0	0	Ŧ	0	0	-	1	T	
Vancomycin 8× MIC	7	1	0	12	6	1	12	11	9	12	12	12	
$4 \times MIC$	7	1	0	12	6	2	12	10	9	12	12	12	
$2 \times MIC$	7	1	0	12	6	2	12	10	8	12	12	11	
MIC	6	1	0	12	4	0	11	9	5	10	9	7	
$0.5 \times MIC$	0	0	0	2	0	0	2	0	0	2	2	0	

TABLE 3. Results of time-kill assays of 12 pneumococcal strains

 a Only strains with tetracycline MICs of <8.0 µg/ml were tested. b Only strains with clarithromycin MICs of <4.0 µg/ml were tested.

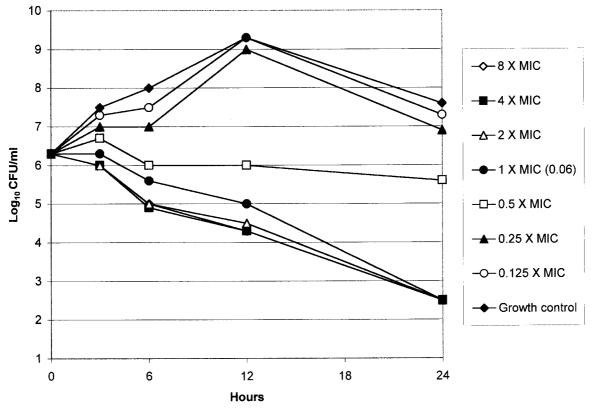


FIG. 1. Graphical depiction of GAR-936 kill kinetics versus a tetracycline-resistant (MIC, 64.0 µg/ml) strain of S. pneumoniae.

new agent for treatment of infections caused by drug-resistant pneumococci.

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