

HHS Public Access

Eur J Cancer Prev. Author manuscript; available in PMC 2023 September 01.

Published in final edited form as:

Author manuscript

Eur J Cancer Prev. 2022 September 01; 31(5): 401–407. doi:10.1097/CEJ.000000000000722.

TAS2R38 polymorphisms, *Helicobacter pylori* infection and susceptibility to gastric cancer and pre-malignant gastric lesions

Matteo Giaccherini^{1,2}, Cosmeri Rizzato³, Manuel Gentiluomo¹, Antonella Lupetti³, Lourdes Flores-Luna⁴, Jorge Vivas⁵, Maria Mercedes Bravo⁶, Elena Kasamatsu⁷, Nubia Muñoz⁸, Federico Canzian², Ikuko Kato⁹, Daniele Campa¹

¹Department of Biology, University of Pisa, Pisa, Italy

²Genomic Epidemiology Group, German Cancer Research Center (DKFZ), Heidelberg, Germany

³Department of Translational Research and New Technologies in Medicine and Surgery, University of Pisa, Pisa, Italy

⁴Center for Public Health Research, National Institute of Public Health, Cuernavaca, Morelos, Mexico

⁵Cancer Control Center of the Tachira State, San Cristobal, Venezuela

⁶Grupo de Investigación en Biología del Cáncer, Instituto Nacional de Cancerología, Bogotá, Colombia

⁷Instituto de Investigaciones en Ciencias de la Salud, National University of Asunción, Asunción, Paraguay

⁸Cancer Institute of Colombia, Bogotá, Colombia

⁹Department of Oncology and Pathology, Wayne State University School of Medicine, Detroit, Michigan, USA

Abstract

Background: Gastric cancer is worldwide the fourth more common cancer type by incidence, and the third by mortality. We analyzed three missense variants of *TAS2R38* gene: rs713598 (A49P), rs1726866 (V262A), and rs10246939 (I296V). These variants and their combination in haplotypes (PAV/tasters or AVI/non-tasters) and diplotypes are responsible for individual

Author contributions

Conflicts of interest

Corresponding author: Daniele Campa, Department of Biology, University of Pisa, Via Derna 1, 56126 Pisa, Italy, daniele.campa@unipi.it, phone +39-050-221510.

Daniele Campa conceived the study. Matteo Giaccherini performed the lab work.

Daniele Campa, Cosmeri Rizzato, Federico Canzian, Ikuko Kato drafted the manuscript.

Manuel Gentiluomo performed data quality control and reconstructed the haplotypes/phenotypes.

Ikuko Kato performed data analysis.

Antonella Lupetti, Lourdes Flores-Luna, Jorge Vivas, Maria Mercedes Bravo, Elena Kasamatsu, Nubia Muñoz, contributed in the data interpretation, analysis and in the drafting of the manuscript.

No author has potential conflicts of interest to disclose.

differences in bitter perception. The single nucleotide polymorphisms and the related phenotypes are known to be associated with susceptibility to Gram-negative bacterial infections, such as *Helicobacter pylori*, and with risk of various cancer types. An association between intermediate tasters (as defined by *TAS2R38* diplotypes) and increased risk of gastric cancer was reported in a Korean population.

Methods: we analyzed 2616 individuals of Latin American origin, representing the whole spectrum of lesions from gastritis to gastric cancer.

Results: Comparing cancer cases *vs* non-cancers we observed a decrease in risk associated with heterozygous carriers of rs10246939 (p=0.006) and rs1726866 (p=0.003) when compared with homozygotes of the more common allele. Also, the analysis of diplotypes/phenotypes reflected the same association, with super-tasters showing a borderline increased risk of developing gastric cancer compared to medium-tasters (OR=1.63, 95%CI 1.04-2.56, p=0.033). Also, non-tasters showed an increased risk when compared to medium-tasters although not reaching statistical significance (OR=1.58, 95%CI 0.80-2.87, p=0.203). We also tested the interactions between the *TAS2R38* genotypes and *H. pylori cagA* status in a subset of samples and found no interaction.

Conclusion: In conclusion, our results suggest only a modest contribution of *TAS2R38* gene genetic variability in gastric cancer etiology.

Keywords

TAS2R38; genetic polymorphisms; *Helicobacter pylori*; susceptibility; gastric cancer; premalignant gastric lesions

Introduction

Gastric cancer (GC) is worldwide the fourth more common cancer type by incidence, affecting over one million people annually, and the third by mortality with nearly 800,000 cases dying of the disease (data from 2018, https://gco.iarc.fr/today/, (Ferlay et al. 2019)).

Helicobacter pylori (Hp), a Gram-negative bacterium, has been reconized as a class 1 carcinogen for GC by the International Agency for Research on Cancer.(Humans 1994) Hp is the most important established risk factor for GC (Helicobacter and Cancer Collaborative Group 2001) as well as a risk factor for pre-malignant gastric lesions, ranging from chronic gastritis to dysplasia, and gastric ulcer.(Uemura et al. 2001) Even though the involvement of the genetic variability to GC development is relatively understudied, its contribution has been investigated genome-wide association studies (GWAS) (Lott and Carvajal-Carmona 2018) and candidate gene studies.(Rizzato et al. 2013a; Durães et al. 2014; Duell et al. 2015)

In humans at least 25 *TAS2R* genes encode for receptors that are responsible for bitter perception. Bitter taste receptor expression has been reported at first in the oral cavity,(Adler et al. 2000) but in the last decades it has been observed in a large number of organs and tissues, including GI tract, testis and upper respiratory airways.(Behrens and Meyerhof 2011) These evidences clearly demonstrate that this group of genes do not have only a role in taste perception and are probably involved in several unrelated functions. Many poison and toxic compounds are bitter and therefore a plausible explanation for their evolution

has been, for many years, their ability to function as gatekeepers to avoid the ingestion of possibly harmful molecules.(Risso et al. 2016b) More recently, instead, mounting evidence suggest that the main function of taste receptors could be the defense of the organism from pathogens, especially Gram-negative bacteria.(Lee et al. 2012) Germline genetic variability in taste receptor genes has been associated with many human traits, such as body mass index (BMI),(Inoue et al. 2013) male infertility,(Gentiluomo et al. 2017) dietary and drinking habits,(Duffy et al. 2004; Diószegi, Llanaj and Ádány 2019) aging,(Campa et al. 2012) smoking behavior,(Risso et al. 2016a) human infections,(Gallo et al. 2016; Cantone et al. 2018) and risk of several cancer types.(Campa et al. 2010; Carrai et al. 2011; Barontini et al. 2017; Gentiluomo et al. 2019) Three single nucleotide polymorphisms (SNPs) in the *TAS2R38* gene, rs713598 (A49P), rs1726866 (V262A), and rs10246939 (I296V), are responsible for amino acid substitutions that modify the ability of the receptor to bind to ligands. In addition through linkage disequilibrium they capture the vast majority of the genetic variability of the gene and have extensively studied in relation to cancer development.(Carrai et al. 2011; Choi and Kim 2019; Lambert et al. 2019)

Considering these premises, the aim of the present study was to analyze the genetic variability of the *TAS2R38* gene in relation to GC and advanced gastric premalignant lesions, in several populations from Latin America where GC risk is high.

Materials and Methods

Study population

This study was designed as an ancillary project based on deidentified samples and data previously collected for three studies conducted in a wide range of Latin American populations from low to high risk of GC and representing the whole spectrum of lesions from gastritis to GC. Details concerning eligibility, recruitment, data and sample collection of each study have been published elsewhere. (Muñoz et al. 1996; Kato et al. 2004; Trejode la O et al. 2008; Rizzato et al. 2012; Flores-Luna et al. 2013, 2020) Briefly, the original studies include a randomized clinical trial (RCT) for pre-malignant lesions, a GC case-control study and cross-sectional studies for Hp and its associated gastric pathologies. Except for the participants in the RCT and neighborhood controls in the case-controls study, who were in general good health, the other study subjects were recruited from patients seen at gastroenterology or oncology clinics. Subjects were at least 30 years old, and recruited at hospitals from four countries in Latin America, i.e., Venezuela (Tachira province), Colombia (five centers, two from high and three from low risk areas based on gastric cancer incidence), Mexico (Mexico City) and Paraguay (Asunción City) in a varied period from early 1990s to early 2000s. In each site recruitment time spanned no more than three years and all disease groups were collected within the same time period. All study subjects signed an informed consent and ethical clearance was obtained from the committee of each recruitment as well as coordinating centers. While samples were archived regardless of Hp cagA status from Venezuelan RCT and case-control study (cases only), Mexican, Colombian, and Paraguayan samples were limited to cagA-positive only. cagA status was determined by PCR in all studies. For this study, we included 3021 samples.

Histological diagnosis

For non-cancer cases, five or six gastroscopic biopsies taken from predefined sites were used for histopathological assessment. For each biopsy, the following variables were recorded in a standardized form: type and depth of mucosa biopsied, degrees of neutrophil and monocyte infiltration, active regeneration, glandular atrophy, IM and dysplasia, depth of monocyte infiltration, type of intestinal metaplasia (IM), and quantity of Hp infection (none, difficult to find, easy to find, and abundant). Most of these lesions were graded as none, mild, moderate, or severe and scored as 0-3, respectively. The Hp, monocyte, and neutrophil infiltration scores were calculated as the mean of all evaluable biopsies rated 0-3. Global diagnoses of non-cancer patients were classified into superficial gastritis, chronic gastritis, atrophic gastritis, IM and dysplasia based on the most advanced lesion as previously described.(Muñoz et al. 1996; Kato et al. 2004; Rizzato et al. 2013b; Canzian et al. 2020; Flores-Luna et al. 2020) IMs were further subtyped into types I-III according to Filipe and Jass (Filipe and Jass 1986) and incomplete subtypes, IM II and III, and dysplasia were considered high-grade premalignant lesions based on our previous observation in relation to cagA-positive Hp infection.(Plummer et al. 2007) Duodenal ulcer diagnosis without any premalignant lesions were considered ineligible. GC cases were limited to epithelial origin, arising from non-cardiac gastric mucosa, and classified according to the Lauren classification.(Hu et al. 2012)

SNP selection and genotyping

We analyzed the genetic variability of the *TAS2R38* gene, focusing on three common polymorphisms that determine an amino acid change: rs713598 (A49P), rs1726866 (V262A), and rs10246939 (I296V). The combination of these variants is responsible for phenotypic change in bitter tasting. Genotyping was conducted in 384-well plates using TaqMan technology (ThermoFisher Applied Biosystems, Waltham MA, USA) and KASP technology (LGC Genomics/KBioscience, Hoddesdon, UK), according to the manufacturer's instruction. Duplicated samples (8%) were used as positive control to check the quality of genotyping, and no-template controls were added to each plate. PCR plates were read on a QuantStudio 5 instrument and genotypes called with QuantStudio software (ThermoFisher Applied Biosystem, Waltham MA, USA).

Haplotype, diplotype and phenotype reconstruction

Haplotypes were reconstructed starting from individuals having 100% call rate (n=2177) using the Phase software.(Stephens and Scheet 2005) Diplotypes were assigned to each individual and a tasting phenotype, coded as a categorical variable, was also assigned to each diplotype combination. Considering the literature,(Choi and Kim 2019) we assigned to the PAV/PAV diplotype the highest degree of bitter sensitivity and to AVI/AVI the lowest one. All other diplotypes were considered as intermediate tasters.

Data filtering, statistical and bioinformatic analysis

Out of the 3021 samples, for which DNA was extracted, 326 were excluded from the statistical analysis because they had a call rate lower than 66% (less than two genotypes out of the three attempted), leaving 2695 subjects. Additional 79 samples were removed

Giaccherini et al.

due to missing covariates as well as ineligible histological types of gastric cancer. The association analyses were limited to the remaining 2616 subjects, that had an average call rate of 94%. The concordance rate between duplicates samples was higher than 98%. For haplotype/phenotype analysis the number was limited to those with a call rate of 100% (n = 2177). We use baseline (pre-treatment) information for the RCT participants.

Response variables in this study were global histological diagnosis, which was divided into 3 groups: low-grade, high-grade pre-malignant lesions and GC, and selected histological parameters, i.e., Hp, monocyte and neutrophil infiltration scores for non-cancer cases only. A conditional logistic regression model to accommodate stratified analyses on country was used to estimate odds ratios (OR) and 95% confidence intervals (CI) for GC vs. non-cancer and those for high-grade vs. low-grade diseases. The ORs for genotypes were calculated using major allele homozygotes as the reference. The ORs were also calculated using both dominant and recessive models. The linear trend with risk was also tested according to the number of minor alleles. Medium-taster phenotype was used as the reference in calculating the ORs for phenotypes. All ORs were adjusted for basic demographic variables (sex, age and educational level), and other risk factors reported previously in this population (cigarette smoking, and duration of refrigerator use), (Kato et al. 2004) most of which, except sex and age, were coded on the ordinal scale. In addition, among non-cancer cases, analysis of covariance was used to estimate covariate-adjusted mean Hp, monocyte, and neutrophil infiltration scores according to receptor phenotypes. These covariates included indicators variables for country (Venezuela as reference), histological group (low- vs high-grade) in addition to those described above for logistic regression. All analyses were performed with SAS 9.4. We also tested the interactions between *cagA* status and genotypes/haplotypes/ phenotypes in Venezuelan samples only, prior to combining their samples to the others, which were limited to cagA-positive specimens. We did not find any significant interactions (supplementary table 1) and thus the main results were based on all countries combined.

Results

We analyzed the associations between three polymorphic variants of the *TAS2R38* gene, (rs713598 rs1726866, rs10246939) and the risk of developing GC in a population recruited in Latin America. The relevant characteristics of the population are given in table 1, whereas the allelic frequencies are shown in supplementary table 2.

The outcomes of interest were high-grade *vs* low-grade lesions and cancer *vs* non-cancer (high-grade + low-grade pre-malignant lesions). None of the SNPs showed a statistically significant association comparing high-grade *vs* low-grade lesions. Comparing cancer *vs* non-cancer we observed a decrease in risk associated with heterozygous carriers of rs10246939 (p=0.006) and rs1726866 (p=0.003) when compared with homozygotes of the more common allele. Also, heterozygotes for rs713598 showed a lower risk of cancer, although the association did not reach statistical significance (p=0.098). The results are shown in table 2.

We also reconstructed the haplotypes and observed a remarkable difference in haplotype frequencies among the different countries in the study as shown in supplementary table 3.

Additionally, we inferred the tasting phenotypes from the diplotypes. We observed that super-tasters had an increased risk of developing GC compared to medium-tasters (OR=1.63, 95% CI 1.04-2.56, p=0.033). Also, non-tasters showed an increased risk when compared to medium-tasters although not reaching statistical significance (OR=1.58, 95% CI 0.80-2.87, p=0.203). We observed no statistically significant associations comparing high-grade vs low-grade lesions. The results are shown in table 3.

Finally, our results did not show any evidence of interaction between *TAS2R38* polymorphisms and *cagA* status (supplementary table 1). We did not find statistically significant differences in the geno-haplo-phenotype frequencies between *CagA* positive Venezuelan samples and all samples from all other countries (compare supplementary table 1 and supplementary table 4).

Discussion

TAS2R38 polymorphisms have been associated with a very long list of human traits such as dietary preferences,(Diószegi, Llanaj and Ádány 2019) smoking behavior,(Baker et al. 2018) alcohol drinking, choice and abuse,(Duffy et al. 2004; Ramos-Lopez et al. 2015) glucose homeostasis,(Dotson et al. 2008) BMI,(Inoue et al. 2013) risk of various cancer types and microbial infection, especially from Gram-negative microorganisms.(Gallo et al. 2016; Cantone et al. 2018)

The haplotypes (PAV and AVI) and diplotypes resulting from the allelic and genotypic combinations explain the individual differences in the perception of bitter molecules. AVI/AVI individuals are considered non-tasters and PAV/PAV have been defined as super-tasters with all the intermediate combinations associated with intermediate tasting phenotypes. The T2R38 receptor has a very strong affinity for a synthetic compound called 6-n-propylthiouracil (PROP), and also binds to several bitter compounds present in vegetables, drugs, smoke particulate and alcoholic (e.g. beer, wine) and non-alcoholic beverages (e.g. coffee and tea).(Mikołajczyk-Stecyna, Malinowska and Chmurzynska 2017) TAS2R38 polymorphisms, alone or in combination, and PROP sensitivity phenotypes have been studied in relation to colorectal cancer, (Carrai et al. 2011) pancreatic cancer, (Gentiluomo et al. 2019) and GC.(Choi et al. 2016) Considering that the AVI haplotype is supposedly non-functional and therefore gives an obvious disadvantage to AVI/AVI individuals both in respect of defense from infections and in rejecting the ingestion of toxic compounds, a long standing question has been the reason of the relatively high frequency of this population across the world. (Risso et al. 2016b) A possible explanation could be balancing selection, that favors heterozygous individuals who have an intermediate tasting phenotype.(Wooding et al. 2004)

Our results show that heterozygotes for the *TAS2R38*-rs713598 (A49P), *TAS2R38*-rs1726866 (V262A), and/or *TAS2R38*-rs10246939 (I296V) have a lower risk of developing GC. According to tasting phenotype determination (through haplotype and diplotype reconstruction) we observed that medium-tasters had a decreased risk of developing the disease compared to both non-tasters and super-tasters. Cancer is not generally considered to exert selective pressure, due to the fact that arises at older stages in life, however one of the

main etiological factor of GC is the inflammation due to Hp (a Gram-negative bacterium) that colonizes the host at younger age. Thus, it may be possible to explain this association from an evolutionary perspective. The AVI haplotype (conferring the non-taster phenotype) has been associated with increased risk of developing infection even though in very small studies conducted in individuals of Caucasian descent.(Gallo et al. 2016; Cantone et al. 2018) On the other hand, the PAV haplotype and the PAV/PAV combination (super-tasters) have been reported to increase the risk of several cancer types (Mikhina, Motorina and Glekov 1985; Choi et al. 2017) possibly through an interaction with food intake. These observations could lead to hypothesize that a balanced mix of the two haplotypes AVI and PAV could decrease the risk to develop Hp infection and cancer. However, it needs to be noted that several other reports suggest that the AVI haplotype increases the risk of developing neoplastic pathologies (Carrai et al. 2011; Yamaki et al. 2017) or no association at all.(Choi and Kim 2019; Gentiluomo et al. 2019) In addition, the evidences supporting an association between vegetable preferences and intake are heterogenous and conflicting. (Lambert et al. 2019; Smith et al. 2020)

In a recent manuscript Choi and colleagues investigated the role of *TAS2R38*-rs713598, *TAS2R38*-rs1726866, and *TAS2R38*-rs10246939 alone and in combination, in relation to GC in 449 cases and 1,131 controls from Korea. The authors observed that medium-tasters had an increase in risk in developing the disease compared to super-tasters while non-tasters did not show any association.(Choi et al. 2016) These results go in the opposite direction of what we observed in our data. The frequencies of the SNPs are very different in the Korean study and in ours. The two populations are therefore genetically different, which makes a direct comparison of the different findings difficult. It is worth mentioning though that the minor allele frequencies are similar to what expected in both populations when compared to other published work (Risso et al. 2016b) or to the 1000 Genomes project (https://www.internationalgenome.org/1000-genomes-browsers). A limitation of this study is, indeed, that all subjects enrolled in the analysis are from Latin America, which makes hard to generalize our findings.

Hp infection and *cagA* status did not show any interaction with the tasting phenotype and therefore cannot explain the differences observed in the two studies. However we were not well powered to detect interaction between the microorganism and the host genetic variability and it is known that Hp strains are dramatically different in different populations across the globe,(Thorell et al. 2017; Muñoz-Ramirez et al. 2021) it is therefore possible that the difference in the directions of the association depends on different strains of Hp.

Finally, given multiple comparisons made on the two different histological endpoints as well all modest level of the statistical associations found, we cannot rule out the possibility that both associations reflect random statistical fluctuation and chance findings.

In conclusion, our results suggest at best a limited influence of the *TAS2R38* genetic variability in gastric cancer susceptibility.

Refer to Web version on PubMed Central for supplementary material.

Funding

This work was supported by intramural funding of the University of Pisa. Additional funding was provided by the National Cancer Institute, National Institutes of Health, United States (R01CA98309 and R21CA182822, P.I. Ikuko Kato).

Data availability

The primary data for this work will be made available to researchers who submit a reasonable request to the PI of the external funding (I. Kato), conditional to approval by all the collaborators. Data will be stripped from all information allowing identification of study participants. The data are not publicly available due to privacy or ethical restrictions.

References

- Adler E, Hoon MA, Mueller KL, Chandrashekar J, Ryba NJP, Zuker CS. A novel family of mammalian taste receptors. Cell 2000;100:693–702. [PubMed: 10761934]
- Baker AN, Miranda AM, Garneau NL, Hayes JE. Self-reported Smoking Status, TAS2R38 Variants, and Propylthiouracil Phenotype: An Exploratory Crowdsourced Cohort Study. Chem Senses 2018;43:617–25. [PubMed: 30137252]
- Barontini J, Antinucci M, Tofanelli S, Cammalleri M, Dal Monte M, Gemignani F et al. Association between polymorphisms of TAS2R16 and susceptibility to colorectal cancer. BMC Gastroenterol 2017;17:104. [PubMed: 28915899]
- Behrens M, Meyerhof W. Gustatory and extragustatory functions of mammalian taste receptors. Physiol Behav 2011;105:4–13. [PubMed: 21324331]
- Campa D, De Rango F, Carrai M, Crocco P, Montesanto A, Canzian F et al. Bitter taste receptor polymorphisms and human aging. Glendinning JI (ed.). PLoS One 2012;7:e45232. [PubMed: 23133589]
- Campa D, Vodicka P, Pardini B, Naccarati A, Carrai M, Vodickova L et al. A gene-wide investigation on polymorphisms in the taste receptor 2R14 (TAS2R14) and susceptibility to colorectal cancer. BMC Med Genet 2010;11:88. [PubMed: 20534144]
- Cantone E, Negri R, Roscetto E, Grassia R, Catania MR, Capasso P et al. In Vivo Biofilm Formation, Gram-Negative Infections and TAS2R38 Polymorphisms in CRSw NP Patients. Laryngoscope 2018;128:E339–45. [PubMed: 29570813]
- Canzian F, Rizzato C, Obazee O, Stein A, Flores-Luna L, Camorlinga-Ponce M et al. Genetic polymorphisms in the cag pathogenicity island of Helicobacter pylori and risk of stomach cancer and high-grade premalignant gastric lesions. Int J Cancer 2020;147:2437–45. [PubMed: 32363734]
- Carrai M, Steinke V, Vodicka P, Pardini B, Rahner N, Holinski-Feder E et al. Association between TAS2R38 gene polymorphisms and colorectal cancer risk: a case-control study in two independent populations of Caucasian origin. Song Y (ed.). PLoS One 2011;6:e20464. [PubMed: 21674048]
- Choi J-H, Kim J. TAS2R38 Bitterness Receptor Genetic Variation and Risk of Gastrointestinal Neoplasm: A Meta-Analysis. Nutr Cancer 2019;71:585–93. [PubMed: 30663393]
- Choi J-H, Lee J, Choi IJ, Kim Y-W, Ryu KW, Kim J. Genetic Variation in the TAS2R38 Bitter Taste Receptor and Gastric Cancer Risk in Koreans. Sci Rep 2016;6:26904. [PubMed: 27245112]
- Choi JH, Lee J, Oh JH, Chang HJ, Sohn DK, Shin A et al. Variations in the bitterness perceptionrelated genes TAS2R38 and CA6 modify the risk for colorectal cancer in Koreans. Oncotarget 2017;8:21253–65. [PubMed: 28423504]

- Diószegi J, Llanaj E, Ádány R. Genetic Background of Taste Perception, Taste Preferences, and Its Nutritional Implications: A Systematic Review. Front Genet 2019;10, DOI: 10.3389/ fgene.2019.01272.
- Dotson CD, Zhang L, Xu H, Shin Y-K, Vigues S, Ott SH et al. Bitter taste receptors influence glucose homeostasis. PLoS One 2008;3:e3974. [PubMed: 19092995]
- Duell EJ, Bonet C, Muñoz X, Lujan-Barroso L, Weiderpass E, Boutron-Ruault M-C et al. Variation at ABO histo-blood group and FUT loci and diffuse and intestinal gastric cancer risk in a European population. Int J Cancer 2015;136:880–93. [PubMed: 24947433]
- Duffy VB, Davidson AC, Kidd JR, Kidd KK, Speed WC, Pakstis AJ et al. Bitter Receptor Gene (TAS2R38), 6-n-Propylthiouracil (PROP) Bitterness and Alcohol Intake. Alcohol Clin Exp Res 2004;28:1629–37. [PubMed: 15547448]
- Durães C, Muñoz X, Bonet C, García N, Venceslá A, Carneiro F et al. Genetic variants in the IL1A gene region contribute to intestinal-type gastric carcinoma susceptibility in European populations. Int J Cancer 2014;135:1343–55. [PubMed: 24615437]
- Ferlay J, Colombet M, Soerjomataram I, Mathers C, Parkin DM, Piñeros M et al. Estimating the global cancer incidence and mortality in 2018: GLOBOCAN sources and methods. Int J Cancer 2019;144:1941–53. [PubMed: 30350310]
- Filipe M, Jass J. Intestinal Metaplasia Subtypes and Cancer Risk. Gastric ca. In Filipe M and Jass J (eds) (ed.). Edimburgh: Churchill Livingstone, 1986.
- Flores-Luna L, Bravo MM, Kasamatsu E, Lazcano Ponce EC, Martinez T, Torres J et al. Risk factors for gastric precancerous and cancers lesions in Latin American counties with difference gastric cancer risk. Cancer Epidemiol 2020;64:101630. [PubMed: 31756677]
- Flores-Luna L, Camorlinga-Ponce M, Hernandez-Suarez G, Kasamatsu E, Martínez ME, Murillo R et al. The utility of serologic tests as biomarkers for Helicobacter pylori-associated precancerous lesions and gastric cancer varies between Latin American countries. Cancer Causes Control 2013;24:241–8. [PubMed: 23184121]
- Gallo S, Grossi S, Montrasio G, Binelli G, Cinquetti R, Simmen D et al. TAS2R38 taste receptor gene and chronic rhinosinusitis: new data from an Italian population. BMC Med Genet 2016;17:54. [PubMed: 27515546]
- Gentiluomo M, Crifasi L, Luddi A, Locci D, Barale R, Piomboni P et al. Taste receptor polymorphisms and male infertility. Hum Reprod 2017;32:2324–31. [PubMed: 29040583]
- Gentiluomo M, Lu Y, Canzian F, Campa D. Genetic variants in taste-related genes and risk of pancreatic cancer. Mutagenesis 2019;34:391–4. [PubMed: 31606007]
- Helicobacter and Cancer Collaborative Group. Gastric cancer and Helicobacter pylori: a combined analysis of 12 case control studies nested within prospective cohorts. Gut 2001;49:347–53. [PubMed: 11511555]
- Hu B, El Hajj N, Sittler S, Lammert N, Barnes R, Meloni-Ehrig A. Gastric cancer: Classification, histology and application of molecular pathology. J Gastrointest Oncol 2012;3:251–61. [PubMed: 22943016]
- Humans IWG on the E of CR to. Infection with Helicobacter pylori. IARC Monogr Eval Carcinog risks to humans 1994;61:177–240.
- Inoue H, Yamakawa-Kobayashi K, Suzuki Y, Nakano T, Hayashi H, Kuwano T. A case study on the association of variation of bitter-taste receptor gene TAS2R38 with the height, weight and energy intake in Japanese female college students. J Nutr Sci Vitaminol (Tokyo) 2013;59:16–21. [PubMed: 23535535]
- Kato I, Vivas J, Plummer M, Lopez G, Peraza S, Castro D et al. Environmental factors in Helicobacter pylori-related gastric precancerous lesions in Venezuela. Cancer Epidemiol Biomarkers Prev 2004;13:468–76. [PubMed: 15006925]
- Lambert JD, VanDusen SR, Cockroft JE, Smith EC, Greenwood DC, Cade JE. Bitter taste sensitivity, food intake, and risk of malignant cancer in the UK Women's Cohort Study. Eur J Nutr 2019;58:2111–21. [PubMed: 29980925]
- Lee RJ, Xiong G, Kofonow JM, Chen B, Lysenko A, Jiang P et al. T2R38 taste receptor polymorphisms underlie susceptibility to upper respiratory infection. J Clin Invest 2012;122:4145– 59. [PubMed: 23041624]

- Lott PC, Carvajal-Carmona LG. Resolving gastric cancer aetiology: an update in genetic predisposition. lancet Gastroenterol Hepatol 2018;3:874–83. [PubMed: 30507471]
- Mikhina ZP, Motorina LI, Glekov I V. [Prophylactic brain irradiation and radiotherapy of brain metastases of small cell lung cancer]. Vopr Onkol 1985;31:61–70.
- Mikołajczyk-Stecyna J, Malinowska AM, Chmurzynska A. TAS2R38 and CA6 genetic polymorphisms, frequency of bitter food intake, and blood biomarkers among elderly woman. Appetite 2017;116:57–64. [PubMed: 28455260]
- Muñoz-Ramirez ZY, Pascoe B, Mendez-Tenorio A, Mourkas E, Sandoval-Motta S, Perez-Perez G et al. A 500-year tale of co-evolution, adaptation, and virulence: Helicobacter pylori in the Americas. ISME J 2021;15:78–92. [PubMed: 32879462]
- Muñoz N, Kato I, Peraza S, Lopez G, Carrillo E, Ramirez H et al. Prevalence of precancerous lesions of the stomach in Venezuela. Cancer Epidemiol Biomarkers Prev 1996;5:41–6. [PubMed: 8770465]
- Plummer M, van Doorn L-J, Franceschi S, Kleter B, Canzian F, Vivas J et al. Helicobacter pylori Cytotoxin-Associated Genotype and Gastric Precancerous Lesions. JNCI J Natl Cancer Inst 2007;99:1328–34. [PubMed: 17728213]
- Ramos-Lopez O, Roman S, Martinez-Lopez E, Gonzalez-Aldaco K, Ojeda-Granados C, Sepulveda-Villegas M et al. Association of a novel TAS2R38 haplotype with alcohol intake among Mexican-Mestizo population. Ann Hepatol 2015;14:729–34. [PubMed: 26256902]
- Risso DS, Kozlitina J, Sainz E, Gutierrez J, Wooding S, Getachew B et al. Genetic Variation in the TAS2R38 Bitter Taste Receptor and Smoking Behaviors. PLoS One 2016a;11:e0164157. [PubMed: 27711175]
- Risso DS, Mezzavilla M, Pagani L, Robino A, Morini G, Tofanelli S et al. Global diversity in the TAS2R38 bitter taste receptor: revisiting a classic evolutionary PROPosal. Sci Rep 2016b;6:25506. [PubMed: 27138342]
- Rizzato C, Kato I, Plummer M, Muñoz N, Stein A, Jan van Doorn L et al. Risk of advanced gastric precancerous lesions in Helicobacter pylori infected subjects is influenced by ABO blood group and cagA status. Int J cancer 2013a;133:315–22. [PubMed: 23319424]
- Rizzato C, Kato I, Plummer M, Muñoz N, Canzian F. Genetic variation in PSCA and risk of gastric advanced preneoplastic lesions and cancer in relation to Helicobacter pylori infection. PLoS One 2013b;8:e73100. [PubMed: 24023815]
- Rizzato C, Torres J, Plummer M, Muñoz N, Franceschi S, Camorlinga-Ponce M et al. Variations in Helicobacter pylori cytotoxin-associated genes and their influence in progression to gastric cancer: implications for prevention. PLoS One 2012;7:e29605. [PubMed: 22235308]
- Smith JL, Estus S, Lennie TA, Moser DK, Chung ML, Mudd-Martin G. TAS2R38 PAV Haplotype Predicts Vegetable Consumption in Community-Dwelling Caucasian Adults at Risk for Cardiovascular Disease. Biol Res Nurs 2020;22:326–33. [PubMed: 32207317]
- Stephens M, Scheet P. Accounting for decay of linkage disequilibrium in haplotype inference and missing-data imputation. Am J Hum Genet 2005;76:449–62. [PubMed: 15700229]
- Thorell K, Yahara K, Berthenet E, Lawson DJ, Mikhail J, Kato I et al. Rapid evolution of distinct Helicobacter pylori subpopulations in the Americas. PLoS Genet 2017;13:e1006546. [PubMed: 28231283]
- Trejo-de la O A, Torres J, Pérez-Rodríguez M, Camorlinga-Ponce M, Luna LF, Abdo-Francis JM et al. TLR4 single-nucleotide polymorphisms alter mucosal cytokine and chemokine patterns in Mexican patients with Helicobacter pylori-associated gastroduodenal diseases. Clin Immunol 2008;129:333–40. [PubMed: 18755634]
- Uemura N, Okamoto S, Yamamoto S, Matsumura N, Yamaguchi S, Yamakido M et al. Helicobacter pylori Infection and the Development of Gastric Cancer. N Engl J Med 2001;345:784–9. [PubMed: 11556297]
- Wooding S, Kim U-K, Bamshad MJ, Larsen J, Jorde LB, Drayna D. Natural selection and molecular evolution in PTC, a bitter-taste receptor gene. Am J Hum Genet 2004;74:637–46. [PubMed: 14997422]

Giaccherini et al.

Yamaki M, Saito H, Isono K, Goto T, Shirakawa H, Shoji N et al. Genotyping Analysis of Bitter-Taste Receptor Genes TAS2R38 and TAS2R46 in Japanese Patients with Gastrointestinal Cancers. J Nutr Sci Vitaminol (Tokyo) 2017;63:148–54. [PubMed: 28552880]

Table 1.

Distribution of demographic factors of study subjects by histological diagnosis.

		Low-grade ^a	(%)	High-grade ^b	(%)	Cancer ^c	(%)	Total	(%)
Sex	Female	1119	53.2%	179	52.0%	58	34.1%	1356	51.8%
	Male	983	46.8%	165	48.0%	112	65.9%	1260	48.2%
Age	30-39	537	25.5%	38	11.0%	10	5.9%	585	22.4%
	40-49	751	35.7%	90	26.2%	30	17.6%	871	33.3%
	50-59	519	24.7%	109	31.7%	35	20.6%	663	25.3%
	60+	295	14.0%	107	31.1%	95	55.9%	497	19.0%
Country ^d	Mexico	101	4.8%	19	5.5%	4	2.4%	124	4.7%
	Paraguay	33	1.6%	7	2.0%	9	5.3%	49	1.9%
	Colombia ^e	188	8.9%	41	11.9%	23	13.5%	252	9.6%
	Venezuela	1780	84.7%	277	80.5%	134	78.8%	2191	83.8%
	(Hp negative)	155	8.7%	10	3.6%	2	1.5%	167	7.6%
	(cagA negative)	555	31.2%	37	13.4%	26	19.4%	618	28.2%
	(cagA positive)	1070	60.1%	230	83.0%	106	79.1%	1406	64.2%

% represents column percentage.

 a Low-grade includes superficial gastritis to type I intestinal metaplasia.

b. High-grade includes types II and III intestinal metaplasia and dysplasia.

^cCancer includes non-cardia cancer, divided into 95 intestinal, 62 diffuse and 13 unknown/other cases.

 $d_{\text{All samples from countries except Venezuela were$ *cagA* $positive.}$

 $e_{\text{includes 179 from high-risk}}$ area and 73 from low-risk area

Table 2.

Associations between TAS2R38 SNPs and risk of high-grade pre-malignant gastric lesions or gastric cancer.

SNP	Genotype	Low- grade	High- grade	Cancer	OR ^a (95% CI)	P-value	OR ^b (95% CI)	P-value
rs10246939 (I296V)	CC	869	141	81	1.00		1.00	
	CT	860	149	51	1.04 (0.80-1.34)	0.783	0.58 (0.39-0.85)	0.006
	TT	255	45	26	1.08 (0.74-1.57)	0.697	1.01 (0.61-1.68)	0.963
	Trend					0.678		0.301
	Dominant	1115	194	77	1.05 (0.82-1.33)	0.717	0.67 (0.47-0.96)	0.028
	Recessive	255	45	26	1.06 (0.74-1.50)	0.755	1.30 (0.81-2.11)	0.281
rs1726866 (V262A)	GG	901	141	77	1.00		1.00	
	AG	835	136	41	1.01 (0.78-1.31)	0.952	0.52 (0.34-0.80)	0.003
	AA	240	42	19	1.07 (0.72-1.57)	0.866	0.87 (0.50-1.54)	0.641
	Trend					0.785		0.099
	Dominant	1075	178	60	1.02 (0.80-1.31)	0.872	0.60 (0.41-0.88)	0.008
	Recessive	240	42	19	1.06 (0.74-1.53)	0.748	1.04 (0.61-1.78)	0.605
rs713598 (A49P)	GG	813	128	73	1.00		1.00	
	GC	893	145	62	1.00 (0.77-1.31)	0.997	0.72 (0.49-1.06)	0.098
	CC	314	49	23	0.96 (0.66-1.38)	0.811	0.72 (0.42-1.23)	0.231
	Trend					0.845		0.119
	Dominant	1207	194	85	0.99 (0.77-1.27)	0.931	0.72 (0.51-1.03)	0.074
	Recessive	314	49	23	0.96 (0.68-1.34)	0.794	0.85 (0.52-1.40)	0.399

^aodds ratio for high-grade lesions *vs* low-grade, adjusted for age, sex, educational level, length of refrigerator use, smoking status and high starch diet tertile, and stratified by country.

b odds ratio for cancer vs non cancer (low+ high-grade), adjusted for age, sex, educational level, length of refrigerator use, smoking status and high starch diet tertile, and stratified by country.

ORs, 95%CIs and p-values in bold are statistically significant (p<0.05).

Table 3.

Odds ratios for risk of gastric cancer and high-grade premalignant lesions by taster status.

Taster status	Low-grade	High-grade	Cancer	OR ^{<i>a</i>} (95% CI)	P-value	OR ^{<i>b</i>} (95% CI)	P-value
Non-taster	212	37	16	1.01 (0.68-1.51)	0.783	1.58 (0.80-2.87)	0.203
Medium-taster	841	141	44	1.00		1.00	
Super-taster	723	110	53	0.95 (0.72-1.25)	0.697	1.63 (1.04-2.56)	0.033

 a^{a} odds ratio for high-grade lesions vs low-grade, adjusted for age, sex, educational level, length of refrigerator use, smoking status and high starch diet tertile, and stratified by country.

b odds ratio for cancer vs non-cancer (low+ high-grade), adjusted for age, sex, educational level, length of refrigerator use, smoking status and high starch diet tertile, and stratified by country.

ORs, 95%CIs and p-values in bold are statistically significant (p<0.05).