



Review

Small Vessel Disease: Ancient Description, Novel Biomarkers

Rita Moretti * and Paola Caruso

Neurology Clinic, Department of Medical, Surgical and Health Sciences, University of Trieste, 34127 Trieste, Italy; paolacaruso83@gmail.com

* Correspondence: moretti@units.it

Abstract: Small vessel disease (SVD) is one of the most frequent pathological conditions which lead to dementia. Biochemical and neuroimaging might help correctly identify the clinical diagnosis of this relevant brain disease. The microvascular alterations which underlie SVD have common origins, similar cognitive outcomes, and common vascular risk factors. Nevertheless, the arteriosclerosis process, which underlines SVD development, is based on different mechanisms, not all completely understood, which start from a chronic hypoperfusion state and pass through a chronic brain inflammatory condition, inducing a significant endothelium activation and a consequent tissue remodeling action. In a recent review, we focused on the pathophysiology of SVD, which is complex, involving genetic conditions and different co-morbidities (i.e., diabetes, chronic hypoxia condition, and obesity). Currently, many points still remain unclear and discordant. In this paper, we wanted to focus on new biomarkers, which can be the expression of the endothelial dysfunction, or of the oxidative damage, which could be employed as markers of disease progression or for future targets of therapies. Therefore, we described the altered response to the endothelium-derived nitric oxide-vasodilators (ENOV), prostacyclin, C-reactive proteins, and endothelium-derived hyperpolarizing factors (EDHF). At the same time, due to the concomitant endothelial activation and chronic neuroinflammatory status, we described hypoxia-endothelial-related markers, such as HIF 1 alpha, VEGFR2, and neuroglobin, and MMPs. We also described blood–brain barrier disruption biomarkers and imaging techniques, which can also describe perivascular spaces enlargement and dysfunction. More studies should be necessary, in order to implement these results and give them a clinical benefit.

Keywords: small vessel disease; vascular damage; blood–brain barrier damage; reactive oxygen species; endothelial dysfunction; metalloproteinases



Citation: Moretti, R.; Caruso, P. Small Vessel Disease: Ancient Description, Novel Biomarkers. *Int. J. Mol. Sci.* **2022**, *23*, 3508. <https://doi.org/10.3390/ijms23073508>

Academic Editor: Lorenzo Malatino

Received: 1 March 2022

Accepted: 21 March 2022

Published: 23 March 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Small vessel disease (SVD) (also called cerebral small vessel disease, cSVD) relies on the deep brain's small vessels alterations. *Small vessels* are univocally defined as small penetrating arteries, capillaries, and small veins. cSVD is strongly related to a chronic hypoperfusion condition, which predisposes the entire brain to hemorrhagic events, white (confluent or not) matter alterations, and lacunar events. SVD is the most important and common cause of all the vascular forms of dementia (up to 45%), but as previously underlined, it predisposes to a higher risk of vascular strokes (25–30% of cases) and 25–35% of all the lacunar events [1,2]. The other crucial common characteristic of SVD is that its pathological consequence could be represented by a silent lesion progression, which has its clinical confirmation in dramatic radiological imaging, without apparent, evident acute events. Thus, SVD is a clinical condition whose principal stigma is that the lesions may progress over time, for imprecise rules, and above all, with or without clinical consequences, in relationship with the extension and the confluency of the white matter alterations [3–5].

Generally, SVD clinical signs are concomitant psychological and behavioral sequelae, summarized by an essential executive function disruption and standard neuropsychological features (apathy and vascular depression) [1–4].

In sporadic cerebral SVD, aging, diabetes, chronic hypoxia, and hypertension are the most recognized clinical risk factors. Still, different hereditary forms of cerebral SVD have also been described [6]. Small arterioles show significant disruptions in both cases, easily described as arteriolosclerosis, lipohyalinosis, and severe endothelial disruption. Its principal consequence is a strong invalidation of the neurovascular coupling mechanisms and vessel tone dysregulation [7,8], and even venules are interested in the ongoing process [9]. With the arteriolosclerotic process, SVD is characterized by a substantial increment in cerebral amyloid angiopathy (CAA). This condition, which has been traditionally related to Alzheimer's disease, is, on the contrary, quite frequent in the normal aging process, and it is dramatically evident in the SVD process. It is related to a consistent deposition of amyloid β -peptide (Ab) in the walls of the small arterioles, and it increments the consequences of altered neurovascular coupling in small parenchymal and leptomeningeal arterioles [9–11].

The principal consequence of arteriolosclerosis is the chronic hypoperfusive state, which induces a perpetual neuro-inflammation state, and gives rise to an essential endothelial activation. These conditions induce an overwhelming alteration of the oxidative response, which potentiates the basal inflammation status of the deep brain structure, expanding through different neural networks, principally the basal-forebrain ones [12].

In a recent review [12], we focused on the contribution of the complex and multifaceted “vascular damage” in developing small vessel dementia, starting from small vessel disease condition. We have written that the SVD is “an ongoing process, which begins with altered microvessels and pial arteries and ends in subcortical dementia; CBF regional selective decrease seems to be one of the critical factors for the progression from small vessel disease to small vessel disease-related dementia, together with proved altered response to inflammation, and oxidative stress” [12].

Neuroimaging is the main helpful diagnostic instrument for managing brain SVD. Therefore, the main findings in SVD are subcortical infarcts, lacunes, white matter hyperintensities (WMHs), prominent perivascular spaces (PVS), and cerebral microbleeds (CMBs) [13]. T2 or FLAIR MRI reports indicate confluent and symmetrical white matter hyperintensities [14–23] into the frontal and prefrontal-thalamus-basal forebrain networks [24–29]. Many instruments have been implemented to relate the number of lacunes, the extension and the amount of surface of white matter hyperintensities, and their relation to the subsequent cognitive and behavioral impairment [30–32]. The confluence between clinical, neuropsychological, and neuroimaging findings helps to converge for a correct diagnosis of the Vascular cognitive impairment, as stated in NINDS-AIREN criteria [33–35] and the DSM-V R (Fifth Edition-revised) [36–39]. VCI refers to an ample spectrum of vascular brain pathologies that contribute to cognitive impairment, ranging from mild and subjective cognitive decline to overt dementia [40,41].

The state of the art in the VCI field is an ongoing definition [42], and many terms have been employed, such as the descriptive ones of vascular cognitive disorder, subcortical vascular dementia, and mild and major vascular neurocognitive disorders (Diagnostic and Statistical Manual of Mental Disorders, Fifth Edition (DSM-5) [36–39,43]. Others include vascular cognitive disorder (VCD), while subcortical VAD (sVAD) has been employed to define a circumscribed syndrome, related to small vessel disease [44–46]. However, it is well defined that the difference between VaD subtypes may depend on the anatomical distribution of the vascular insults [47]. Usually, small artery disease is more often associated with subcortical VaD than with cortical and cortical-subcortical VaD [47]. Executive dysfunctions and behavioral disorders (apathy and vascular depression, etc.) are the commonest findings [48].

Very recently, excellent studies emerged on the potential role of transcranial doppler findings in patients with white matter lesions, as possible markers for developing vascular dementia. A significant example is the demonstration that patients with white matter lesions, but without any other sign of cognitive impairment, showed a hemodynamic pattern of cerebral hypoperfusion and enhanced vascular resistance, as a distinctive marker of a possible predictor factor of developing dementia [49]. Another significant result is

the one obtained by a study using a transcranial Doppler, in which patients with $\geq 80\%$ unilateral internal carotid artery stenosis with no history of stroke were recruited [50]; this study demonstrated that cognitive impairment correlated linearly with lower flow in the hemisphere fed by the occluded internal carotid artery, but only below a threshold of MFV = 45 cm/s. [50]

Finally, transcranial doppler studies have shown that it could delineate a profile of low perfusion and high vascular resistance in patients with a defined diagnosis of vascular depression [49].

In this paper, we wanted to address the contribution of the chronic inflammatory brain condition due to SVD and, starting from this situation, verify the possibility of finding new biomarkers of endothelial dysfunction, inflammation, and oxidative damage, which could be possible future targets of focused therapies.

2. Possible and Proved New Markers of Blood–Brain Barrier Leakage, Perivascular Enlargements, and Mitochondrial Alterations

SVD has the small vessels (pial and the small penetrating) and white matter as a significant definite target. Nevertheless, growing attention has been dedicated to disrupting perivascular spaces, astrocytic end-feet, capillaries, and veins. As a final point, the blood–brain barrier (BBB) has been addressed as another potential target of the intriguing mechanisms that underlie the small vessel brain pathology complex. BBB is not only a solid defensive barrier but acts as an active and specific player of active selection crossover, possessing cell-cell signaling with the end-feet of astrocytes and disclosure a potential role of maintaining efflux pumps [51–55]. Thus, the disruption of the BBB is proportionately increased by normal aging but progresses as a hallmark in different pathologies, i.e., multiple sclerosis or in primary inflammatory disease. Nevertheless, it is an expression of white matter inflammation, even due to chronic hypoperfusion, such as the one which occurs in small vessel disease [SVD], accomplishing the progression and the extension of the white matter sufferance, named as white matter hyperintensities (WMH) [56–62], the confluency of which is synonymous with SVD progression, leading to subcortical vascular dementia (sVAD) [12,62]. In AD-prone patients, BBB disruption has been signaled even in hippocampal degeneration, which occurs after a major stroke [61,62].

A dynamic contrast-enhanced MRI (DCE-MRI) [63] has been employed for in-vivo quantification of the pathological passage of plasma through BBB [64,65]. Moreover, apart from the BBB leakage, the possibility of estimating the vascular permeability-surface area product (PS) and the plasma volume fraction (VP) in a given region of interest has also been described [66,67]. The model suggested that PS increased with WMH severity, aging, and other vascular risk factors, and at the same time, a lower blood vP [65]. The most promising in-vivo demonstration is that BBB integrity is compromised in more severe WMH, even beyond visible lesions [63] (Insert Figure 1).

Even if we know that BBB is disrupted in SVD, we do not know the reasons for BBB leakage in this condition. The most disputed involvement is one of the pericytes. Pericytes are capillary mural cells that stabilize newly formed vessels and induce repair. When a pericyte-deficient adult mouse model has been employed [68], different transcriptional changes in brain endothelial cells have been mapped due to a defective pericyte contact at a single-cell level. In that conformation, endothelial cells, deprived of pericyte contacts, seem to exhibit a “venous-shifted molecular pattern,” and therefore lack any capillary specialization, and upregulate proteins which are typically expressed during developmental stages, such as the Fibroblast Growth Factor Binding Protein (*Fgfbp1*), or those expressed during pathological angiogenesis, such as Angiopoietin 2 (*Angpt2*). These aspects permit a possible cell proliferation, with a very flawed arteriolar BBB regulation system, and reduction of the angiogenesis process [68]. *Fgfbp1* and *Angpt2* levels could probably be crucial markers of BBB leakage during SVD. More studies will be necessary to prove that.

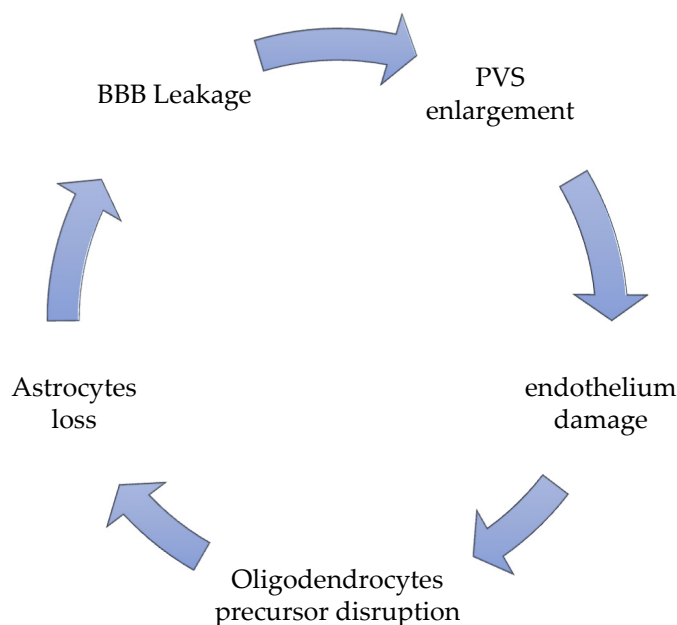


Figure 1. The vicious circle of SVD pathology.

Perivascular spaces (PVS) have gained an essential role in SVD pathogenesis; they are no longer considered as virtual empty spaces, but as the most efficacious catabolites clearance system [12]; they are resident sites of perivascular macrophages, pial cells, mast cells, nerve fibers, and collagen fibers [69]. PVS are virtual spaces intimately connected to deep arterioles [70]. Even in these conditions, they act as a lymphatic net, defined as a glymphatic-perivascular territory [71].

Their malfunction, the hallmarks of which are the combined enlargement and widening, is the principal responsibility for perivascular accumulation of catabolites and toxic substances, which is determinant for enhancing ongoing neural damage until starvation [72,73]. The perivascular debris accumulation, together with the BBB leakage, potentiates and accelerates the perivascular inflammation, strongly favored by the stagnation-induced process and by medical conditions which influence it, such as hypertension and diabetes [74–78]. PVS enlargement is responsible for an altered cerebrovascular reactivity (CVR) [12], due to the extension of the constant inflammatory response [41] present as a constant marker in SVD, due to the chronic hypoperfusion state. The PVS is never an isolated situation, but it is accompanied by an altered BBB disruption and a significant perivascular inflammation [75,79–82]. More recently, new actors contribute with BBB leakage and PVS enlargement to help the progress of SVD [83–86], such as the oligodendrocyte precursor cells (OPCs), which generally help BBB stabilization [86,87] and the astrocytes, which exert their fundamental role as regulating the signal of neuro-vascular coupling [12]. Oligodendrocytes are the first victims of chronic models of chronic cerebral hypoperfusion (CCH), together with the precocious sufferance of the perineural space [88–90], and with a hyperactivation of microglia, firstly in the hippocampus [91,92], then in the thalamus, up to in the cortical neuronal population [93]. Secondary to oligodendrocytes, astrocyte death occurs in proportion to the chronic ischemia condition's length and severity [94,95], due to the ongoing modifications of general and neuronal metabolic requests. Their death is a consequence of chronic hypoxia, but it worsens neuronal death due to a lack of functions, regulating the neurovascular coupling signal [96]. The process by which this occurs is that during the entire process of chronic ischemia, microglia retract its branches, with a consequent reduction of the length and strength of the microglial ramification, with a concomitant degeneration of the soma [97]. The frontal activation of microglia occurs in a two-step pattern: at the beginning, M1 activation upregulates TNF alpha, IL-23, IL-1beta, and IL12 production, which attack neurons, and directly contribute to their injury; only

after M2 activation occurs can the reparation process can begin [98]. In the SVD, due to the chronic hypoxia-hypoperfusion condition [12], the passage through M1 towards M2 activation does not occur [98]. In SVD, there is a substantial augmentation of M1 activation, together with a heavy reduction of M2 promotion [99,100]. The brisk oligodendrocyte degeneration, associated with M1 activation, increases calcium currents and induces a severe apoptosis process. The calcium increases, and the severe apoptosis is accompanied by an augmentation of caspase-3 RNA and matrix-metalloprotease 2 (MMP-2) [101]. At the beginning of the SVD process, these markers reflect the temptation reparation process induced by a standard M1/M2 passage, as described above. Nevertheless, until the chronic inflammatory condition occurs in SVD ongoing development, there is an alteration of the M1/M2 passage, with a predominant M1 event; therefore, in SVD patients' cerebrospinal fluid (CSF), there is a constant growth of oligodendrocyte-derived myelin sheath-like myelin lipid sulfatide (ODSMS) and myelin essential protein (MBP) due to the massive oligodendrocytes death [102–105]. For similar reasons, markers of axonal damage, i.e., neurofilament light chain (NFL), together with CSF α -1 antitrypsin, tissue inhibitor of metalloproteinase-1 (TIMP-1), plasminogen activator inhibitor-1 (PAI-1), and apolipoprotein H (ApoH) have been found to increase very early in the CSF in SVD [106–108]. Finally, due to the BBB leakage, ultrastructural studies find that in older animals as well as in those affected by SVD, there are severe alterations of the capillary basement membrane of the deeper arterioles, inside the white matter, filling plasma proteins into vascular bagging and collagen deposition inside PVS, in a phenomenon described as microvascular fibrosis [55,98,109]. Many studies have testified that microvascular fibrosis and BBB splitting have a higher CSF/serum albumin (SA) ratio in patients with SVD [109]. Matrix remodeling pathway (TIMP-1 and matrix metalloproteinases) as an expression of endothelium disruption in SVD has been described [109] (Insert Table 1 here).

Table 1. Possible hematic or CSF markers of SVD.

Functional Domain	Markers	Effectiveness on SVD
	DCE-MRI technique: Increase in permeability surface area Increase in white matter alterations Lower blood plasma volume in white matter altered regions	Demonstrate diagnostic confirmation Demonstrate the amount and the progression of SVD Determine the PS increasing together with a lowering of blood vP
BBB Leakage	Loss of pericytes: upregulation of FGFBP1 and ANGPT2	Altered angiogenesis and demonstration of a venous-shifted molecular pattern of BBB, due to the altered arterial regulatory properties
	Enlargement of PVS	Alteration of the glymphatic system
	M1 activation: increase in TNF-alpha, IL-23, IL-1 beta, and IL-12	Strong and chronic neuroinflammatory condition, shifted to a M1 vs. M2 activation
	General increment in caspase-3 RNA; of MMP-2	Promoting and overwhelming the active neuroinflammation condition

Table 1. *Cont.*

Functional Domain	Markers	Effectiveness on SVD
Endothelial dysfunction	Decrease in ENOV, prostacyclins, NO, eNOS, and VE-cadherins	Altered production of NO, due to decrease in its production and increment in its consumption, due to increment in ROS
	Increase in C-protein, EDHF, VEGF, ICAM-1, sTM, IL-6, PA-1, von Willebrand Factors, HIF-1 alpha; VEGFR, and Neuroglobin	Expression of endothelial altered activation, with important flawed permeability and activation of thrombotic pattern
	Increase in homocysteine	Endothelial toxicity, promotion of oxidative and inflammatory damages
	Increase in CSF/plasma albumin ratio	Proof of endothelial altered permeability
	albuminuria	Indirect proof of endothelial altered permeability
Oxidative damage	Increase in SOD, prostacyclin, and Hydrogen peroxide	Altered response to oxidative stress, with damages to mitochondria, altered oxygen delivery, and endothelial degeneration promotion
	Decrease in NOX2 NADPH oxidase	Further reduction of proper response to ROS accumulation; their decrease is proportional to endothelial inflammation and alteration
	APOE4	Promotion of endothelial reduced resistance to ROS

3. Markers of Endothelial Dysfunction

As previously described [12], there is a global endothelial altered function in SVD [110,111], which could be synthesized in an alteration of normal endothelial response to endothelium-derived nitric oxide-vasodilators (ENOV) [112], prostacyclin [113], C-reactive proteins [114], and endothelium-derived hyperpolarizing factors (EDHF) [115].

NO is rapidly removed in SVD for the mitochondrial alterations, with a consequent anti-oxidative response and consumed by peroxynitrite (O₂ anions plus NO) [116]. However, it can also be reduced in its production, as it occurs in normal aging [117], in an accelerated way, in SVD, with a consistent down-regulation of endothelial NO synthase (eNOS). Moreover, in SVD, there is an evident dysfunction of the Rho-associated protein kinase (ROCK) [118] and the related ERM proteins (ezrin, radixin, and moesin), fundamental for barrier properties' integrity [118–120] and their induction of the downregulation of the vascular endothelium cadherins (VE-cadherins) [121].

In diabetes, where SVD is a constant presentation form with a crucial endothelial hyper-permeability, a concomitant increase in arteriolar deposition of advanced glycation end products has been observed, which helps and maintains the increase in endothelial permeability through Rho activation and an upregulation of the vascular endothelial growth factor (VEGF) [122,123].

The superimposition of BBB disruption, endothelial dysfunction, and microvascular fibrosis causes a substantial permeability alteration, with albumin extravasation; the increased CSF/plasma albumin ration is a proven witness of a severe progression of con-

fluency of white matter lesions in SVD [124–127], together with albuminuria (even if not well-accepted) [128–131].

Other important markers of endothelial altered activation [12,132–134] in SVD are intercellular adhesion molecule-1 (ICAM-1), which has been considered as a generic expression of white matter progression [95], soluble thrombomodulin (sTM), interleukin-6 (IL-6), plasminogen activator inhibitor-1 (PAI-1), and von Willebrand factor [129–134]. Others, such as HIF 1 alpha, VEGFR2, and neuroglobin, are more evident when the confluency of different WMH becomes constant in different models [135,136].

4. Markers of Oxidative Damages in SVD

Reactive oxygen species (ROS) is an umbrella term for many ordinary derivatives of molecular oxygen, and their accumulation leads to a complex phenomenon called oxidative distress. There are two species, hydrogen peroxide (H_2O_2) and the superoxide anion radical (O_2^-), which are key redox signaling agents generated under the control of growth factors and cytokines by more than 40 enzymes, prominently including nicotinamide adenine dinucleotide phosphate (NADPH) oxidases [12] and the mitochondrial electron transport chain [126]. When mitochondrial cells usually function, the active process of oxidative phosphorylation converts oxygen to superoxide by oxidase enzymes, and superoxide can be transformed by superoxide dismutase (SOD) or to non-radical hydrogen peroxide [126,136,137], i.e., from glutathione peroxidase (Gpx), or when catalase enzymatically metabolizes hydrogen peroxide to water and oxygen [136].

Chronic cerebral conditions of constant hypoxia are the principal inductors of the uncontrolled production of ROS [138,139].

NADPH oxidase activity and mitochondrial are significantly higher in cerebral arteries when compared with systemic arteries in blood vessels from healthy animals (mouse, rat, pig, and rabbit) [140,141]. Thus, brain vessels are one of the most prominent productions of ROS, suggesting that there could be fundamental ROS-dependent signaling in cerebral arteries, which might be indispensable for vasoactive regulation properties.

Thus, the accumulation of ROS species, associated with mitochondrial dysfunction, BBB disruption, and chronic inflammatory status are three conditions in SVD and are proportionate to WMH extension. They lead to an altered endothelial further altered activation, which is reflected in a decoupling of the neurovascular coupling system, with significant sub-cortical and cortical signal alteration, with consequent reflex in oligodendrocytes astrocytes and finally to neurons [12,142]. An active role of flow-dependent responses in rat cerebral arteries has been recently demonstrated in vivo, directly exerted by the NADPH-oxidase reactions [143]. Specifically, Nox₂-NADPH oxidase dysfunction is related to the propagation of the ischemic brain injury, derived by the occlusion of larger pial arteries; Nox₂/NOx₂ knock-out mice, in the same condition, show the minor extension of brain injury after an ischemic infarct [144].

The induced alterations of mitochondrial DNA by ROS attacks and chronic ischemic conditions are some of the most critical contributors to neuronal aging and degeneration, either considering oxidative damage as a promoter or as a consequence of it [145–147].

The decline of mitochondrial functioning has been largely implicated in the aging process and is characterized by a reduced density of mitochondria and reduced mitogenesis [148–152]. Such changes, which originate as replication errors, accumulate in postmitotic tissues during aging, leading to increased proportions of impaired mitochondria [152]. In the aging brain, there has been a sufficient demonstration of impairment of synaptic mitochondria leading to impaired neurotransmission and cognitive failure [149–155]. Precocious forms of small vessel disease, leading to vascular dementia, have been described in specific mitochondrial point mutation [156]. Other mitochondrial mutation phenotypes have been described as pure brain involvement, including fluctuating encephalopathy, seizures, dementia, migraine, stroke-like episodes, ataxia, and spasticity [149,153–155]. Growing attention should be paid to mitochondrial DNA mutations for brain pathologies,

in order to gain more robust data on their possible relevance, and their correlation with postmortem neuropathologic features, to advance our understanding [156–161].

Oxidative stress potentiates the disorders of the endothelium-dependent NO signaling [162,163]. Uncoupling endothelial NO synthase (eNOS) (i.e., in relation with lower levels of tetrahydrobiopterin) switches the production of NO to that of superoxide, causing an overwhelming potentiation of ROS production, accelerating the oxidative stress, lowering the NO anti-inflammatory properties [164,165], and reducing NO modulation of Rho-kinase activity, inhibiting vascular tone control [166]. Rho-kinase, as a counterpart, influences mRNA-stability of eNOS [167].

The induction of oxidative stress is one of the most important promoters of pathological angiogenesis, by lipid oxygenation, thickening the blood vessel walls [168,169]. Moreover, the ApoE4 allele and the AD process seem to be involved in promoting vascular alterations independently of other recognized factors, i.e., age, diabetes, hypertension, and obesity, etc. However, it is supposed to worsen the confluency of WMH, probably somehow linked to ROS augmentation, without any other positive data [170–172].

5. Inflammation and SVD

As above written, neuroinflammation is a common finding in SVD models; it is tightly related to chronic hypoperfusion condition and defined as located hypoxia condition, the common finding of SVD. The pivotal role of neuroinflammation in SVD could accelerate the lipid peroxidation precipitation of the redox system and promote a more robust activation of M1 than M2 [173].

It has been demonstrated that NO-related metabolite, citrulline, and dimethylarginine (DMA) concentrations were significantly higher in patients with strategic infarcts [174]. Arginine depletion was an independent predictor of VaD [174]. S100B (calcium-binding protein B) is a protein that stimulates the expression of pro-inflammatory cytokines. SomIt has been described to have a significant correlation between S100B/asymmetric dimethylarginine levels and cognitive decline in patients with leukoaraiosis [175,176].

Homocysteine could be a potential marker of neuroinflammation inside SVD, promoting the increase in TNF-alpha and IL1-beta, upregulating the transcriptional fibroblast growth factor-2, IL-6, and IL-8, [177,178], and enhancing the VEGF/ERK1/2 signaling pathway [179,180], which can be seen frequently in the atherosclerosis process. Homocysteine is directly linked to the B-inflammatory pathway through a direct upregulation of pyruvate kinase muscle isoenzyme 2 (PKM-2), B-mediated, which mainly promotes the inflammatory basis of atherosclerosis cascade [181,182].

Homocysteine accumulation promotes an increase in the endoplasmic reticulum (ER) stress, upregulating metalloproteinases-9 (MMP-9), and inducing apoptosis [183]. Definitively, the accumulation of homocysteine in animal models enhanced the expression of the AGEs or vascular cell adhesion molecule [184] and MMP-9 [185]. The inflammation cascade could be mediated by the effects on smooth muscle cells rather than on the endothelium alterations [186,187].

Chronic inflammation and oxidative stress have been suggested as concurrent mechanisms of SVD. A possible link between accumulation products (i.e., homocysteine) and other markers could be the circulating metalloproteinases (MMPs) and the tissue inhibitors of metalloproteinases (TIMPs) [188].

MMPs, some of the Ca^{2+} -Zn endopeptidase, have been described as having six different properties: collagenase, gelatinases, stromelysins, matrilysin, membrane-specific metalloproteinases, and no other specified. Their specific role inside the brain is complex and multifaceted; it begins with the neuronal networks remodeling throughout the integrity of the BBB [189]. MMP remains inside the brain, probably in inactivate form, and is active only under special conditions, such as chronic hypoperfusion or chronic inflammatory status. The significant components are MMP2 and MMP14, which are present specifically inside astrocytes, whereas microglia present the MMP-3 and the MMP9, which, by definition, are called inflammatory metalloproteases. Their expression is more severe in acute

damage and gradually decreases in the reparation phases. They can be found near the damaged areas and in the propinquity vessel-related areas [190].

There are four possible mechanisms which have been related to MMP involvement in SVD and, in general, in the neuroinflammation process. The most obvious and well-studied MMP directly activates signaling cytokines, cell-receptors, and adhesion molecules. There are essential works that testify that, even directly, MT4-MMP upregulates a TNF-alpha convertase, and is able to activate TNF-alpha, in its soluble and active [191,192]. Secondly, there are many pieces of evidence in different clinical cases (neurological bacterial infection and PD, etc.), in which there is a direct activation, probably mediated by lipopolysaccharides, calcium currents and other apoptotic signals, and alpha-synuclein deposits, which activate MMP-3 into the interstitial brain fluid, and there, it triggers M1 activation, with a consequent (and above-described) M1 activation [193,194].

Thirdly, MMP seems to be tightly involved in the so-called Fas-FasL system. This system has been known as an inducer of extrinsic cell death responsible for cell-mediated cytotoxicity and peripheral immune regulation. MMP might improve the FAS system, probably through an intrinsic possibility of modulating chloride channel activity, inducing and promoting glutamate excitotoxicity currents, or altering the interactions between neuronal cells and extracellular matrix compounds [195].

Finally, the MMPs participate in many digestive processes at the BBB, particularly the tight junctions and the basement membrane. It has been proposed that MMPs digest tight junctions and basement membrane proteins, thus contributing to BBB leakage [196]. The increased activity of MMP, tightly associated with a higher permeability at the BBB, has been demonstrated in vivo during the reperfusion process through an increase in MMP-2 and MMP-9 mRNA activity [197]. The induction of BBB leakage has, as an indirect effect, an increment in the vasogenic edema, inside the WM, with a drastic increment in vascular demyelination process (Insert Figure 2).

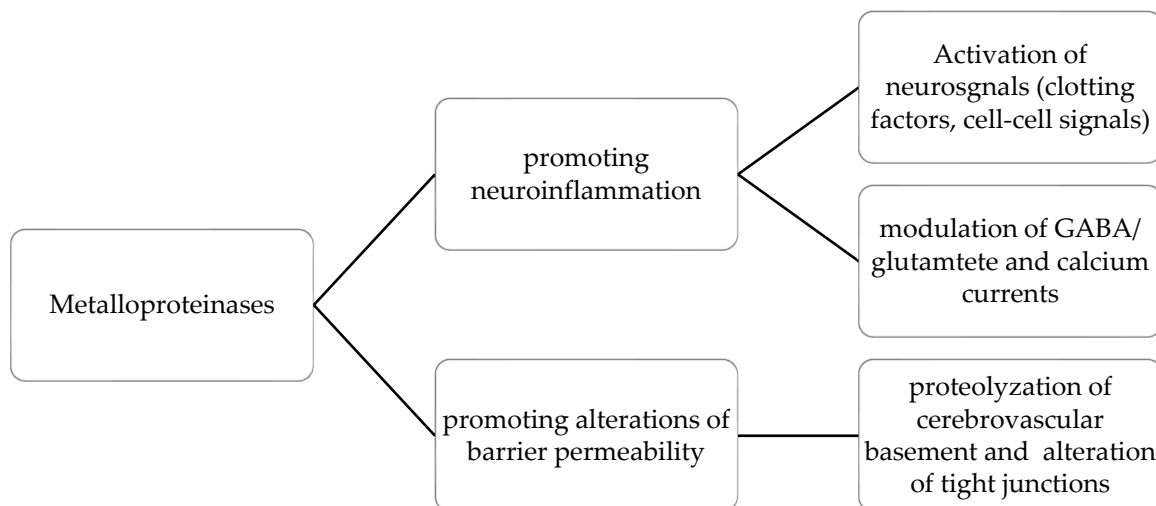


Figure 2. A synopsis of the metalloproteinases action inside the brain, redirecting in SVD pathology.

These data have been evoked in animal models and rare human models, and there is a substantial lack of information, i.e., on the possible relationship between MMP levels and extension and repairing of stroke lesions [198,199].

Nevertheless, some interesting points shed some light on the topic: increased confluency of WMH could be related to higher levels of TIMP-4, after three months of a primary stroke [199].

In a recent study, Arba et al. showed that increasing the grade of SVD sustains higher levels of TIMP-4 and supports the involvement of TIMP-4 in the pathologic process of SVD; they studied a population of an ischemic stroke patient, reporting that brain atrophy was associated with baseline TIMP-4 levels and leukoaraiosis was associated with 90-day TIMP-

4 levels. A global SVD score, expressed as a combined product of leukoaraiosis, lacunes, and brain atrophy, was associated with TIMP-4 levels at 90 days with a dose-response effect [199].

Increased levels of MMP have been associated with severe white matter alterations and a cognitive profile that resembles sVAD [200,201]. In particular, a positive relationship between MMP2 lower levels has been found, together with an increase in albumin index in CSF of SVD patients, as above written [202–204].

Due to lipohyalinosis substitution of smooth muscle cells in arterioles (as described in 12), there is an inverse correlation between TIMP-4 elevated levels (only in animal models) and reduction of lipohyalinosis and collagen bagging through an undescribed and uncertain mechanism [205–207].

All these aspects accounted for, MMPs and tissue inhibitors of metalloproteinases-1 (TIMP-1) could be promising SVD biomarkers [208,209].

6. Potential Future Therapies Approach

Different approaches can be employed to offer potential treatment for VCI, at the moment these are only symptomatic; many data have been obtained from cholinesterase inhibitors and memantine [210].

Potential treatment strategies for brain SVD might include those that target antioxidant effects for the endothelium of small cerebral vessels and the BBB. Due to the major decrease in NO bioavailability in SVD, NO donors could help release the functioning endothelium of small vessel disease, limited by their susceptibility to tolerance development. The apparent strategy, in the same manner as the administration of potent antioxidants such as Vitamins C and E, has shown to be beneficial for vascular function in several experimental and small clinical trials [211].

Disappointingly, the results of large clinical trials of antioxidant supplementation have largely failed to show any benefit. The ROS scavenger tempol is cell-permeable and has been used in experimental studies, as well as edaravone (O₂-scavenger). The ROS scavenger tempol is cell-permeable and has been used in experimental studies, as well as edaravone (O₂-scavenger). Problems derived from NADPH oxidase activity, particularly its primary contributor, Nox₂. It can be argued that prolonged selective therapies could help prevent brain SVD but invariably lead to an immunosuppression condition and many other side effects derived by other different Nox oxidases [137,145,212].

Notably, three of the most influential and frequently prescribed classes of drugs for the treatment of vascular risk factors, which have been shown to inhibit NADPH oxidases, reducing oxidative stress, are the Angiotensin-converting-enzyme inhibitors (ACE inhibitors), Angiotensin II receptor type 1 (AT 1) antagonists, and the statins [3,4]. There are no impressive studies on these drugs as primary NADPH inhibitors, rather than their well-known function per se.

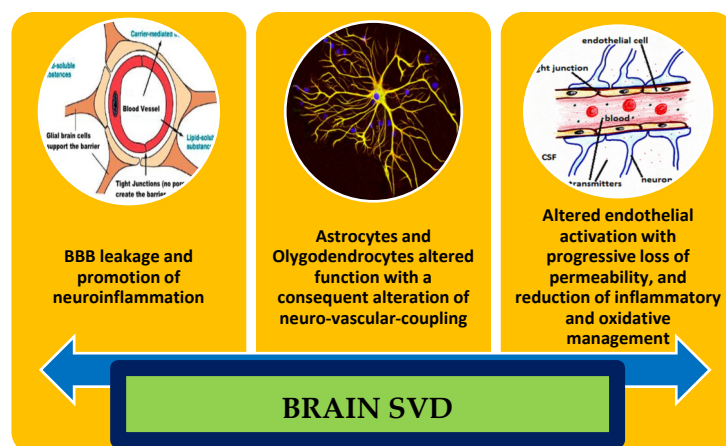
Many other trials have been conducted and are still ongoing [213].

Phenolic acids (or phenolcarboxylic acids) are aromatic acid compounds containing a phenolic ring and an organic carboxylic acid function [214]. Among the most studied molecules belonging to this group, caffeic, chlorogenic, o-coumaric, p-coumaric, m-coumaric, ferulic, and cinnamic acids are the most commonly consumed in the human diet, being contained in coffee [215], together with gallic, p-hydroxybenzoic, vanillic, syringic, and protocatechuic acids. They can be found in bran, grain brown rice, olive oil, tea, cherries, plums, gooseberries, and red wine [216]. These substances have been studied among middle-aged adults, showing a benefit of their intake in different cognitive domains [217–224].

In the same way, rosmarinic acid induced a promotion of oxidative stress response and a reduced lipid-peroxidation [225], but also reduced the gene expression of inducible nitric oxide synthase [225,226], and promoted neuroprotection, reducing matrix metalloproteinase 2 (MMP2), and IL-1 beta [225–227]. Myrtenal has been recently employed

as a multi-property substance (anti-inflammatory and anti-oxidant) [228] but results are only promising.

Apart from physical aerobic activity and avoiding vascular risk factors (smoking, high quantities of carbohydrates, and alcohol consumption, etc.), even external stimuli have been applied in studies; in order to implement cognitive abilities on vascular deterioration, transcranial magnetic stimulation has been studied, which it is still under debate, because its activity on the dorsal striatum with the consequential increase in dopamine release may contribute to the clinical and neurophysiological outcome in vascular depression and vascular cognitive impairment [229] (Insert Scheme 1).



Scheme 1. Synopsis of the pathophysiology of SVD.

7. Conclusions

In the last few decades, the concept of vascular contributions to cognitive impairment and dementia has been emphasized. Cerebral small vessel disease is a common neurocognitive disorder and source of disability. Pathophysiology of cSVD is complex, involving multiple pathways, as described before. Several risk factors, including genetic, co-morbid complications, and environmental factors, contribute to the pathogenesis or exacerbate the complications. Inflammation, chronic hypoperfusion, oxidative damage, glymphatic alterations, and BBB disruption might be potential contributors to the pathogenesis of this complex phenomenon. MMPs, ROS, and other reactive factors trigger inflammatory responses, leading to the abnormalities in small vessels and endothelium dysfunction associated with CSVD.

This study has several limits: Although comprehensive, the approach used in the examined investigations in the attempt to disentangle the complex pathomechanisms of VCI has a number of caveats and potential criticisms. So far in our study, we have tried to have the most homogenous definition, but otherwise, just examining different animal models could represent not a constant level of clinical reversibility. Therefore, the available results on a relatively small sample size might not be confirmed on larger populations, although most of them were obtained from homogeneous samples.

Another limitation is that the correlation between different techniques and the anatomical distribution and severity of vascular lesions has been rarely systematically investigated; therefore, without the contribution of advanced imaging, blood samples, cerebrospinal fluid, laboratory models, or the combination of techniques, the conclusions that can be reached cannot be sufficiently powerful.

Finally, results do not usually provide specific clinical information, although they are sensitive to the “global weight” of several biochemical pathways and neurotransmitter activities. Consequently, a panel of changes, rather than a single marker of disease, should be considered.

More detailed investigations are required to understand the pathophysiology of SVD. Several fluid biomarkers that might be used in diagnostic settings have been identified.

Thus, currently, there is little value in blood tests. CSF biomarkers may help physicians separate vascular and neurodegenerative causes based on BBB disruption and extracellular matrix breakdown. Alongside the need for a correct diagnosis of the disease, biomarkers could be valuable tools to monitor the progression of the disease itself and the possible response to treatment. In this work, we have tried to underline the importance of the inflammatory response in disease pathogenesis. Much further work needs to be conducted along with these positions. The search for an optimal panel of biomarkers with high sensitivity and specificity will provide the crucial tools to enhance success in identifying valid biomarkers in SVD. A combination of biochemical and imaging markers and psychometrics will be necessary to improve the diagnostic accuracy progression of the pathology and finally to monitor response to possible treatment. We believe that the contribution of inflammation on SVD is significant and should be further studied to identify new therapeutic possibilities.

Author Contributions: Conceptualization R.M. and P.C.; methodology, R.M.; software, R.M. and P.C.; formal analysis, R.M. and P.C.; investigation, R.M.; data curation R.M. and P.C.; writing—original draft preparation, R.M. and P.C.; writing—review and editing, R.M.; visualization, R.M.; supervision, R.M. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

Ab	Amyloid b-peptide
AD	Alzheimer's Disease
Angpt2	Angiopietin 2
ApoH	Apolipoprotein H
BBB	Blood—brain barrier
CAA	Cerebral amyloid angiopathy
CBF	Cerebral blood flow
CCH	Chronic cerebral hypoperfusion
CNS	Central Nervous System
COX	Cyclooxygenase
CSF	Cerebrospinal fluid
cSVD	Cerebral small vessel disease
CVR	Cerebrovascular reactivity
EC	Endothelial cells
EDHF	Endothelium-derived hyperpolarizing factors
eNOS	Endothelial NO synthase
ENOV	Endothelium-derived nitric oxide-vasodilators
Fgfbp1	Fibroblast growth factor binding protein
GABA	Gamma-aminobutyric acid
GPx	Glutathione Peroxidase
GSH	Glutathione
ICAM-1	Intercellular adhesion molecule-1
IL	Interleukin
ISF	Interstitial fluid
MBP	Myelin basic protein
MMPs	Matrix metalloproteinase
NADPH	Nicotinamide adenine dinucleotide phosphate
NfL	Neurofilament light chain
NO	Nitric oxide
ODMSMS	Oligodendrocyte-derived myelin sheath-like myelin lipid sulfatide

OPCs	Oligodendrocyte precursor cells
PAI-1	Plasminogen activator inhibitor-1
PS	Permeability-surface area product
PVS	Perivascular spaces
ROCK	Rho-associated protein kinase
ROS	Reactive oxygen species
SA	Serum albumin
SVD	Small vessel disease
SOD	Super Oxide Dismutase
sTM	Soluble thrombomodulin
TIMPs	Tissue inhibitors of metalloproteinases
TNF- α	Tumor necrosis factor- α
VCI	Vascular cognitive impairment
VE-cadherins	Vascular endothelium cadherins
VEGF	Vascular endothelial growth factor
vP	Plasma volume fraction
WMH	White matter hyperintensities

References

- Hakim, A.M. Small Vessel Disease. *Front. Neurol.* **2019**, *10*, 1020. [[CrossRef](#)] [[PubMed](#)]
- Uiterwijk, R.; Van Oostenbrugge, R.J.; Huijts, M.; De Leeuw, P.W.; Kroon, A.A.; Staals, J. Total Cerebral Small Vessel Disease MRI Score Is Associated with Cognitive Decline in Executive Function in Patients with Hypertension. *Front. Aging Neurosci.* **2016**, *8*, 301. [[CrossRef](#)] [[PubMed](#)]
- Taylor, W.D.; Aizenstein, H.J.; Alexopoulos, G.S. The vascular depression hypothesis: Mechanisms linking vascular disease with depression. *Mol. Psychiatry* **2013**, *18*, 963–974. [[CrossRef](#)] [[PubMed](#)]
- Pinter, D.; Ritchie, S.J.; Doubal, F.; Gattringer, T.; Morris, Z.; Bastin, M.; Hernández, M.D.C.V.; Royle, N.A.; Corley, J.; Maniega, S.M.; et al. Impact of small vessel disease in the brain on gait and balance. *Sci. Rep.* **2017**, *7*, 41637. [[CrossRef](#)] [[PubMed](#)]
- Wardlaw, J.M.; Smith, E.F.; Biessels, G.J.; Cordonnier, C.; Fazekas, F.; Frayne, R.; Lindley, R.I.; O'Brien, J.T.; Barkhof, F.; Benavente, O.R.; et al. Neuroimaging standards for research into small vessel disease and its contribution to ageing and neurodegeneration. *Lancet Neurol.* **2013**, *12*, 822–832. [[CrossRef](#)]
- Haffner, C.; Malik, R.; Dichgans, M. Genetic factors in cerebral small vessel disease and their impact on stroke and dementia. *J. Cereb. Blood Flow Metab.* **2016**, *36*, 158–171. [[CrossRef](#)]
- Moody, D.M.; Brown, W.R.; Challa, V.R.; Anderson, R.L. Periventricular venous collagenosis: Association with leukoaraiosis. *Radiology* **1995**, *194*, 469–476. [[CrossRef](#)] [[PubMed](#)]
- Smith, E.E.; Vijayappa, M.; Lima, F.; Delgado, P.; Wendell, L.; Rosand, J.; Greenberg, S.M. Impaired visual evoked flow velocity response in cerebral amyloid angiopathy. *Neurology* **2008**, *71*, 1424–1430. [[CrossRef](#)] [[PubMed](#)]
- Park, L.; Koizumi, K.; El Jamal, S.; Zhou, P.; Previti, M.L.; Van Nostrand, W.E.; Carlson, G.; Iadecola, C. Age-Dependent Neurovascular Dysfunction and Damage in a Mouse Model of Cerebral Amyloid Angiopathy. *Stroke* **2014**, *45*, 1815–1821. [[CrossRef](#)] [[PubMed](#)]
- Staals, J.; Booth, T.; Morris, Z.; Bastin, M.E.; Gow, A.J.; Corley, J.; Redmond, P.; Starr, J.M.; Deary, I.; Wardlaw, J.M. Total MRI load of cerebral small vessel disease and cognitive ability in older people. *Neurobiol. Aging* **2015**, *36*, 2806–2811. [[CrossRef](#)] [[PubMed](#)]
- Staals, J.; Makin, S.D.; Doubal, F.N.; Dennis, M.S.; Wardlaw, J.M. Stroke subtype, vascular risk factors, and total MRI brain small-vessel disease burden. *Neurology* **2014**, *83*, 1228–1234. [[CrossRef](#)] [[PubMed](#)]
- Moretti, R.; Caruso, P. Small Vessel Disease-Related Dementia: An Invalid Neurovascular Coupling? *Int. J. Mol. Sci.* **2020**, *21*, 1095. [[CrossRef](#)]
- Tonet, E.; Pompei, G.; Faragasso, E.; Cossu, A.; Pavasini, R.; Passarini, G.; Tebaldi, M.; Campo, G. Coronary Microvascular Dysfunction: PET, CMR and CT Assessment. *J. Clin. Med.* **2021**, *10*, 1848. [[CrossRef](#)]
- Rosenberg, G.A.; Wallin, A.; Wardlaw, J.M.; Markus, H.S.; Montaner, J.; Wolfson, L.; Iadecola, C.; Zlokovic, B.V.; Joutel, A.; Dichgans, M.; et al. Consensus statement for diagnosis of subcortical small vessel disease. *J. Cereb. Blood Flow Metab.* **2016**, *36*, 6–25. [[CrossRef](#)]
- Patel, B.; Markus, H.S. Magnetic Resonance Imaging in Cerebral Small Vessel Disease and its Use as a Surrogate Disease Marker. *Int. J. Stroke* **2011**, *6*, 47–59. [[CrossRef](#)]
- Erkinjuntti, T.; Inzitari, D.; Pantoni, L.; Wallin, A.; Scheltens, P.; Rockwood, K.; Roman, G.C.; Chui, H.; Desmond, D.W. Research criteria for subcortical vascular dementia in clinical trials. *J. Neur. Transm. Suppl.* **2000**, *59*, 23–30.
- O'Donnell, M.J.; Xavier, D.; Liu, L.; Zhang, H.; Chin, S.L.; Rao-Melacini, P.; Rangarajan, S.; Islam, S.; Pais, P.; McQueen, M.J.; et al. Risk factors for ischaemic and intracerebral haemorrhagic stroke in 22 countries (the INTERSTROKE study): A case-control study. *Lancet* **2010**, *376*, 112–123. [[CrossRef](#)]

18. Potter, G.M.; Marlborough, F.J.; Wardlaw, J.M. Wide variation in definition, detection and description of lacunar lesions on imaging. *Stroke* **2011**, *42*, 359–366. [[CrossRef](#)] [[PubMed](#)]
19. Shi, Y.; Wardlaw, J.M. Update on cerebral small vessel disease. A dynamic whole-brain disease. *Stroke Vasc. Neurol.* **2016**, *1*, e000035. [[CrossRef](#)] [[PubMed](#)]
20. Wardlaw, J.M.; Doubal, F.; Armitage, P.; Msc, F.C.; Carpenter, T.; Maniega, S.M.; Farrall, A.; Sudlow, C.; Dennis, M.; Dhillon, B. Lacunar stroke is associated with diffuse blood-brain barrier dysfunction. *Ann. Neurol.* **2009**, *65*, 194–202. [[CrossRef](#)]
21. Englund, E. White matter pathology of vascular dementia. In *Vascular Dementia*; Chui, E., Ed.; M. Dunitz: London, UK, 2004; pp. 117–130.
22. Englund, E.A.; Person, B. Correlations between histopathologic white matter changes and proton MR relaxation times in dementia. *Alzheimer Dis. Assoc. Disord.* **1987**, *1*, 156–170. [[CrossRef](#)] [[PubMed](#)]
23. Smallwood, A.; Oulhaj, A.; Joachim, C.; Christie, S.; Sloan, C.; Smith, A.D.; Esiri, M. Cerebral subcortical small vessel disease and its relation to cognition in elderly subjects: A pathological study in the Oxford Project to Investigate Memory and Ageing (OPTIMA) cohort. *Neuropathol. Appl. Neurobiol.* **2012**, *38*, 337–343. [[CrossRef](#)] [[PubMed](#)]
24. Gold, G.; Kovari, E.; Herrmann, F.R.; Canuto, A.; Hof, P.R.; Michel, J.P.; Bouras, C.; Giannakopoulos, P. Cognitive consequences of thalamic, basal ganglia, and deep white matter lacunes in brain aging and dementia. *Stroke* **2005**, *36*, 1184–1188. [[CrossRef](#)]
25. Klassen, A.C.; Sung, J.H.; Stadlan, E.M. Histological changes in cerebral arteries with increasing age. *J. Neuropathol. Exp. Neurol.* **1968**, *27*, 607–623. [[CrossRef](#)]
26. Cummings, J.L. Frontal-subcortical circuits and human behavior. *Arch. Neurol.* **1993**, *50*, 873–880. [[CrossRef](#)]
27. Mega, M.S.; Cummings, J.L. Frontal-subcortical circuits and neuropsychiatric disorders. *J. Neuropsychiatry Clin. Neurosci.* **1994**, *6*, 358–370.
28. Tak, S.; Yoon, S.J.; Jang, J.; Yoo, K.; Jeong, Y.; Ye, J.C. Quantitative analysis of hemodynamic and metabolic changes in subcortical vascular dementia using simultaneous near-infrared spectroscopy and fMRI measurements. *Neuroimage* **2011**, *55*, 176–184. [[CrossRef](#)] [[PubMed](#)]
29. McKhann, G.; Drachman, D.; Folstein, M.; Katzman, R.; Price, D.; Stadlan, E.M. Clinical diagnosis of Alzheimer's disease: Report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. *Neurology* **1984**, *34*, 939–944. [[CrossRef](#)] [[PubMed](#)]
30. Kramer, J.H.; Reed, B.R.; Mungas, D.; Weiner, M.W.; Chui, H. Executive dysfunction in subcortical ischaemic vascular disease. *J. Neurol. Neurosurg. Psychiatr.* **2002**, *72*, 217–220. [[CrossRef](#)] [[PubMed](#)]
31. Burton, E.; Ballard, C.; Stephens, S.; Kenny, R.A.; Kalaria, R.; Barber, R.; O'Brien, J. Hyperintensities and fronto-subcortical atrophy on MRI are substrates of mild cognitive deficits after stroke. *Dement. Geriatr. Cogn. Disord.* **2003**, *16*, 113–118. [[CrossRef](#)] [[PubMed](#)]
32. Tullberg, M.; Fletcher, E.; DeCarli, C.; Mungas, D.; Reed, B.R.; Harvey, D.J.; Weiner, M.W.; Chui, H.C.; Jagust, W.J. White matter lesions impair frontal lobe function regardless of their location. *Neurology* **2004**, *63*, 246–253. [[CrossRef](#)] [[PubMed](#)]
33. Román, G.C.; Tatemichi, T.K.; Erkinjuntti, T.; Cummings, J.L.; Masdeu, J.C.; Garcia, J.H.; Amaducci, L.; Orgogozo, J.M.; Brun, A.; Hofman, A. Vascular dementia: Diagnostic criteria for research studies. Report of the NINDS-AIREN International Workshop. *Neurology* **1993**, *43*, 250–260. [[CrossRef](#)] [[PubMed](#)]
34. Chui, H.C.; Victoroff, J.I.; MArgolin, D.; Jagust, W.; Shankle, R.; Katzman, R. Criteria for the diagnosis of ischemic vascular dementia proposed by the state of California Alzheimer's Disease Diagnostic and Treatment Centers. *Neurology* **1992**, *42*, 473–480. [[CrossRef](#)]
35. Kim, G.H.; Lee, J.H.; Seo, S.W.; Ye, B.S.; Cho, H.; Kim, H.J.; Noh, Y.; Yoon, C.W.; Chin, J.H.; Oh, S.J.; et al. Seoul criteria for PIB(-) subcortical vascular dementia based on clinical and MRI variables. *Neurology* **2014**, *82*, 1529–1535. [[CrossRef](#)]
36. Fazekas, F.; Chawluk, J.B.; Alavi, A.; Hurtig, H.I.; Zimmermann, R.A. MR signal abnormalities at 1.5 T in Alzheimer's dementia and normal aging. *Am. J. Roentgenol.* **1987**, *149*, 351–356. [[CrossRef](#)]
37. Cleutjens, F.A.H.M.; Ponds, R.W.H.M.; Spruit, M.A.; Burgmans, S.; Jacobs, H.I.L.; Gronenchild, H.B.M.; Stalls, J.; Franssen, F.M.E.; Dijkstra, J.B.; Vanfleteren, L.E.G.M.; et al. The relationship between cerebral small vessel disease, hippocampal volume and cognitive functioning in patients with COPD: An MRI study. *Front. Aging Neurosci.* **2017**, *9*, 88. [[CrossRef](#)]
38. Scheltens, P.; Barkhof, F.; Leys, D.; Pruvo, J.P.; Nauta, J.J.; Vermersch, P.; Steinling, M.; Valk, J. A semiquantitative rating scale for the assessment of signal hyperintensities on magnetic resonance imaging. *J. Neurol. Sci.* **1993**, *114*, 7–12. [[CrossRef](#)]
39. Kim, K.W.; MacFall, J.R.; Payne, M.E. Classification of white matter lesions on magnetic resonance imaging in elderly persons. *Biol. Psychiatry* **2008**, *64*, 273–280. [[CrossRef](#)]
40. O'Brien, J.T.; Erkinjuntti, T.; Reisberg, B.; Roman, G.; Sawada, T.; Pantoni, L.; Bowler, J.V.; Ballard, C.; DeCarli, C.; Gorelick, P.B.; et al. Vascular cognitive impairment. *Lancet Neurol.* **2003**, *2*, 89–98. [[CrossRef](#)]
41. Erkinjuntti, T.; Gauthier, S. Diagnosing vascular cognitive impairment and dementia. In *Concepts and Controversies in Vascular Cognitive Impairment in Clinical Practice*; Wahlund, L.O., Erkinjuntti, T., Gauthier, S., Eds.; Cambridge University Press: Cambridge, UK, 2009; pp. 3–9.
42. Van der Flier, W.M.; Skoog, I.; Schneider, J.A.; Pantoni, L.; Mok, V.; Chen, C.L.H.; Scheltens, P. Vascular cognitive impairment. *Nat. Rev. Dis. Primers* **2018**, *4*, 18003. [[CrossRef](#)]
43. Skrobot, O.A.; O'Brien, J.; Black, S.; Chen, C.; DeCarli, C.; Erkinjuntti, T.; Ford, G.A.; Kalaria, R.N.; Pantoni, L.; Pasquier, F.; et al. The Vascular Impairment of Cognition Classification Consensus Study. *Alzheimers Dement.* **2017**, *13*, 624–633. [[CrossRef](#)]

44. Thal, D.R.; Grinberg, L.T.; Attems, J. Vascular dementia: Different forms of vessel disorders contribute to the development of dementia in the elderly brain. *Exp. Gerontol.* **2012**, *47*, 816–824. [[CrossRef](#)] [[PubMed](#)]
45. Pantoni, L. Cerebral small vessel disease: From pathogenesis and clinical characteristics to therapeutic challenges. *Lancet Neurol.* **2010**, *9*, 689–701. [[CrossRef](#)]
46. Bowler, J.V. Vascular cognitive impairment. *J. Neurol. Neurosurg. Psychiatr.* **2005**, *76*, 35–44. [[CrossRef](#)] [[PubMed](#)]
47. Caruso, P.; Signori, R.; Moretti, R. Small vessel disease to subcortical dementia: A dynamic model, which interfaces aging, cholinergic dysregulation and the neurovascular unit. *Vasc. Health Risk Manag.* **2019**, *15*, 259–281. [[CrossRef](#)]
48. Jellinger, K.A. Pathomechanisms of Vascular Depression in Older Adults. *Int. J. Mol. Sci.* **2021**, *23*, 308. [[CrossRef](#)] [[PubMed](#)]
49. Puglisi, V.; Bramanti, A.; Lanza, G.; Cantone, M.; Vinciguerra, L.; Pennisi, M.; Bonanno, L.; Pennisi, G.; Bella, R. Impaired Cerebral Haemodynamics in Vascular Depression: Insights from Transcranial Doppler Ultrasonography. *Front. Psychiatry* **2018**, *9*, 316. [[CrossRef](#)]
50. Vinciguerra, L.; Lanza, G.; Puglisi, V.; Pennisi, M.; Cantone, M.; Bramanti, A.; Pennisi, G.; Bella, R. Transcranial Doppler ultrasound in vascular cognitive impairment-no dementia. *PLoS ONE* **2019**, *14*, e0216162. [[CrossRef](#)]
51. Abbott, N.J.; Patabendige, A.A.K.; Dolman, D.E.M.; Yusof, S.R.; Begley, D.J. Structure and function of the blood-brain barrier. *Neurobiol. Dis.* **2010**, *37*, 13–25. [[CrossRef](#)]
52. Zlokovic, B.V. The Blood-Brain Barrier in Health and Chronic Neurodegenerative Disorders. *Neuron* **2008**, *57*, 178–201. [[CrossRef](#)]
53. Cserr, H.F.; DePasquale, M.; Patlak, C.S. Regulation of brain water and electrolytes during acute hyperosmolality in rats. *Am. J. Physiol.* **1987**, *253*, F522–F529. [[CrossRef](#)] [[PubMed](#)]
54. Fraser, P.A.; Dallas, A.D. Permeability of disrupted cerebral microvessels in the frog. *J. Physiol.* **1993**, *461*, 619–663. [[CrossRef](#)]
55. Bridges, L.R.; Andoh, J.; Lawrence, A.; Khoong, C.H.; Poon, W.W.; Esiri, M.M.; Markus, H.S.; Hainsworth, A.H. Blood-Brain Barrier Dysfunction and Cerebral Small Vessel Disease (Arteriolosclerosis) in Brains of Older People. *J. Neuropathol. Exp. Neurol.* **2014**, *73*, 1026–1033. [[CrossRef](#)]
56. Seo, J.H.; Miyamoto, N.; Hayakawa, K.; Pham, L.-D.D.; Maki, T.; Ayata, C.; Kim, K.-W.; Lo, E.H.; Arai, K. Oligodendrocyte precursors induce early blood-brain barrier opening after white matter injury. *J. Clin. Investig.* **2013**, *123*, 782–786. [[CrossRef](#)]
57. Wardlaw, J.M.; Sandercock, P.A.G.; Dennis, M.S.; Starr, J. Ithe s breakdown of the blood-brain barrier responsible for lacunar stroke, leukoaraiosis, and dementia? *Stroke* **2003**, *34*, 806–812. [[CrossRef](#)]
58. Wardlaw, J.M.; Doubal, F.N.; Valdes-Hernandez, M.; Wang, X.; Chappell, F.M.; Shuler, K.; Armitage, P.A.; Carpenter, T.C.; Dennis, M.S. Blood-Brain Barrier Permeability and Long-Term Clinical and Imaging Outcomes in Cerebral Small Vessel Disease. *Stroke* **2013**, *44*, 525–527. [[CrossRef](#)] [[PubMed](#)]
59. Erdo, F.; Denes, L.; de Lange, E. Age-associated physiological and pathological changes at the blood-brain barrier: A review. *J. Cereb. Blood Flow Metab.* **2017**, *37*, 4–24. [[CrossRef](#)]
60. Farrall, A.J.; Wardlaw, J.M. Blood-brain barrier: Aging and microvascular disease—systematic review and meta-analysis. *Neurobiol. Aging* **2009**, *30*, 337–352. [[CrossRef](#)] [[PubMed](#)]
61. Li, Y.; Li, M.; Zhang, X.; Shi, Q.; Yang, S.; Fan, H.; Qin, W.; Yang, L.; Yuan, J.; Jiang, T.; et al. Higher blood-brain barrier permeability is associated with higher white matter hyperintensities burden. *J. Neurol.* **2017**, *264*, 1474–1481. [[CrossRef](#)] [[PubMed](#)]
62. Li, Y.; Li, M.; Zuo, L.; Shi, Q.; Qin, W.; Yang, L.; Jiang, T.; Hu, W. Compromised blood-brain barrier integrity is associated with the total magnetic resonance imaging burden of cerebral small vessel disease. *Front. Neurol.* **2018**, *9*, 221. [[CrossRef](#)] [[PubMed](#)]
63. Stringer, M.S.; Heye, A.K.; Armitage, P.A.; Chappell, F.; Hernández, M.D.C.V.; Makin, S.D.J.; Sakka, E.; Thrippleton, M.J.; Wardlaw, J.M. Tracer kinetic assessment of blood-brain barrier leakage and blood volume in cerebral small vessel disease: Associations with disease burden and vascular risk factors. *NeuroImage* **2021**, *32*, 102883. [[CrossRef](#)]
64. Thrippleton, M.J.; Backes, W.H.; Sourbron, S.; Ingrid, M.; Osch, M.J.P.; Dichgans, M.; Fazekas, F.; Ropele, S.; Frayne, R.; Oostenbrugge, R.J.; et al. Quantifying blood-brain barrier leakage in small vessel disease: Review and consensus recommendations. *Alzheimer's Dement.* **2019**, *15*, 840–858. [[CrossRef](#)] [[PubMed](#)]
65. Manning, C.; Stringer, M.; Dickie, B.; Clancy, U.; Hernandez, M.C.V.; Wiseman, S.J.; Garcia, D.J.; Sakka, E.; Backes, W.H.; Ingrid, M.; et al. Sources of systematic error in DCE-MRI estimation of low-level blood-brain barrier leakage. *Magn. Reson. Med.* **2021**, *86*, 1888–1903. [[CrossRef](#)]
66. Stewart, C.R.; Stringer, M.S.; Shi, Y.; Thrippleton, M.J.; Wardlaw, J.M. Associations Between White Matter Hyperintensity Burden, Cerebral Blood Flow and Transit Time in Small Vessel Disease: An Updated Meta-Analysis. *Front. Neurol.* **2021**, *12*, 647848. [[CrossRef](#)] [[PubMed](#)]
67. Heye, A.K.; Thrippleton, M.J.; Armitage, P.A.; Hernandez, M.D.C.V.; Makin, S.D.; Glatz, A.; Sakka, E.; Wardlaw, J.M. Tracer kinetic modelling for DCE-MRI quantification of subtle blood-brain barrier permeability. *Neuroimage* **2016**, *125*, 446–455. [[CrossRef](#)]
68. Mäe, M.A.; He, L.; Nordling, S.; Vazquez-Liebanas, E.; Nahar, K.; Jung, B.; Li, X.; Tan, B.C.; Chin Foo, J.; Cazenave-Gassiot, A.; et al. Single-Cell Analysis of Blood-Brain Barrier Response to Pericyte Loss. *Circ. Res.* **2021**, *128*, e46–e62. [[CrossRef](#)]
69. Zhang, E.T.; Inman, C.B.; Weller, R.O. Interrelationships of the pia mater and the perivascular (Virchow-Robin) spaces in the human cerebrum. *J. Anat.* **1990**, *170*, 111–123.
70. Iadecola, C. The neurovascular Unit coming of age: A journey through neurovascular coupling in health and disease. *Neuron* **2017**, *96*, 17–42. [[CrossRef](#)]
71. Hendriks, D.; Smits, A.; Lavanga, M.; De Wel, O.; Thewissen, L.; Jansen, K.; Caicedo, A.; Van Huffe, S.; Naulaers, G. Measurement of Neurovascular Coupling in Neonates. *Front. Physiol.* **2019**, *10*, 65. [[CrossRef](#)] [[PubMed](#)]

72. Sweeney, M.D.; Sagare, A.P.; Zlokovic, B.V. Blood-brain barrier breakdown in Alzheimer disease and other neurodegenerative disorders. *Nat. Rev. Neurol.* **2018**, *14*, 133–150. [[CrossRef](#)]
73. Abbott, N.J.; Pizzo, M.E.; Preston, J.E.; Janigro, D.; Thorne, R.G. The role of brain barriers in fluid movement in the CNS: Is there a 'glymphatic' system? *Acta Neuropathol.* **2018**, *135*, 387–407. [[CrossRef](#)]
74. Huijts, M.; Duits, A.; Staals, J.; Kroon, A.A.; De Leeuw, P.W.; Van Oostenbrugge, R.J. Basal ganglia enlarged perivascular spaces are linked to cognitive function in patients with cerebral small vessel disease. *Curr. Neurovasc. Res.* **2014**, *11*, 136–141. [[CrossRef](#)] [[PubMed](#)]
75. Dalkara, T.; Alarcon-Martinez, L. Cerebral micro-vascular signaling in health and disease. *Brain Res.* **2015**, *1623*, 3–17. [[CrossRef](#)] [[PubMed](#)]
76. Wardlaw, J.M.; Benveniste, H.; Nedergaard, M.; Zlokovic, B.V.; Mestre, H.; Lee, H.; Doubal, F.N.; Brown, R.; Ramirez, J.; MacIntosh, B.J.; et al. Perivascular spaces in the brain: Anatomy, physiology and pathology. *Nat. Rev. Neurol.* **2020**, *16*, 137–153. [[CrossRef](#)]
77. Zhang, W.; Zhou, Y.; Wang, J.; Gong, X.; Chen, Z.; Zhang, X.; Cai, J.; Chen, S.; Fang, L.; Sun, J.; et al. Glymphatic clearance function in patients with cerebral small vessel disease. *Neuroimage* **2021**, *238*, 118257. [[CrossRef](#)]
78. Benveniste, H.; Nedergaard, M. Cerebral small vessel disease: A glymphopathy? *Curr. Opin. Neurobiol.* **2022**, *72*, 15–21. [[CrossRef](#)]
79. Jiménez-Balado, J.; Riba-Llena, I.; Garde, E.; Valor, M.; Gutiérrez, B.; Pujadas, F.; Delgado, P. Prevalence of hippocampal enlarged perivascular spaces in a sample of patients with hypertension and their relation with vascular risk factors and cognitive function. *J. Neurol. Neurosurg. Psychiatry* **2018**, *89*, 651–656. [[CrossRef](#)]
80. Giannakopoulos, P.; Gold, G.; Kowaru, E.; von Gunten, A.; Imhof, A.; Bouras, C. Assessing the cognitive impact of Alzheimer disease pathology and vascular burden in the aging brain: The Geneva experience. *Acta Neuropathol.* **2007**, *113*, 1–12. [[CrossRef](#)]
81. Van der Veen, P.H.; Muller, M.; Vinken, K.L.; Hendrikse, J.; Mali, W.P.; van der Graaf, Y.; Geerlings, M.I.; SMART Study Group. Longitudinal relationship between cerebral small vessel disease and cerebral blood flow. The second manifestations of arterial disease-magnetic resonance study. *Stroke* **2015**, *46*, 1233–1238. [[CrossRef](#)]
82. Cuadrado-Godia, E.; Dwivedi, P.; Sharma, S.; Santiago, A.O.; Roquer Gonzalez, J.; Balcells, M.; Laird, J.; Turk, M.; Suri, H.S.; Nicolaidis, A.; et al. Cerebral Small Vessel Disease: A Review Focusing on Pathophysiology, Biomarkers, and Machine Learning Strategies. *J. Stroke* **2018**, *20*, 302–320. [[CrossRef](#)]
83. Zhang, C.E.; Wong, S.M.; van de Haar, H.J.; Staals, J.; Jansen, J.F.; Jeukens, C.R.; Hofman, P.A.; van Oostenbrugge, R.J.; Backes, W.H. Blood-brain barrier leakage is more widespread in patients with cerebral small vessel disease. *Neurology* **2017**, *88*, 426–432. [[CrossRef](#)] [[PubMed](#)]
84. Huisa, B.N.; Caprihan, A.; Thompson, J.; Prestopnik, J.; Qualls, C.R.; Rosenberg, G.A. Long-term blood-brain barrier permeability changes in Binswanger disease. *Stroke* **2015**, *46*, 2413–2418. [[CrossRef](#)] [[PubMed](#)]
85. Wardlaw, J.M.; Makin, S.J.; Hernández, M.C.V.; Armitage, P.A.; Heye, A.K.; Chappell, F.M.; Muñoz-Maniega, S.; Sakka, E.; Shuler, K.; Dennis, M.S.; et al. Blood-brain barrier failure as a core mechanism in cerebral small vessel disease and dementia: Evidence from a cohort study. *Alzheimer's Dement.* **2017**, *13*, 634–643. [[CrossRef](#)]
86. Ihara, M.; Yamamoto, Y. Emerging evidence for pathogenesis of sporadic cerebral small vessel disease. *Stroke* **2016**, *47*, 554–560. [[CrossRef](#)]
87. Rajani, R.M.; Williams, A. Endothelial cell-oligodendrocyte interactions in small vessel disease and aging. *Clin. Sci.* **2017**, *131*, 369–379. [[CrossRef](#)]
88. Furukawa, S.; Sameshima, H.; Yang, L.; Hariskuma, M.; Ikenoue, T. Regional differences of microglial accumulation within 72 hours of hypoxia-ischemia and the effect of acetylcholine receptor agonist on brain damage and microglial activation in newborn rats. *Brain Res.* **2014**, *1562*, 52–58. [[CrossRef](#)]
89. Petito, C.K. Transformation of postischemic perineuronal glial cells. *J. Cereb. Blood Flow Metabol.* **1986**, *6*, 616–624. [[CrossRef](#)]
90. Petito, C.K.; Olarte, J.P.; Roberts, B.; Nowak, T.S.; Pulsinelli, W.A. Selective glial vulnerability following transient global ischemia in rat brain. *J. Neuropathol. Exp. Neurol.* **1998**, *57*, 231–238. [[CrossRef](#)]
91. Masuda, T.; Croom, D.; Hida, H.; Kirov, S.A. Capillary blood flow around microglial somata determines dynamics of microglial processes in ischemic conditions. *Glia* **2011**, *59*, 1744–1753. [[CrossRef](#)]
92. Ju, F.; Ran, Y.; Zhu, L.; Cheng, X.; Gao, H.; Xi, X.; Yang, Z.; Zhang, S. Increased BBB Permeability Enhances Activation of Microglia and Exacerbates Loss of Dendritic Spines after Transient Global Cerebral Ischemia. *Front. Cell Neurosci.* **2018**, *12*, 236. [[CrossRef](#)]
93. Zhang, S. Microglial activation after ischaemic stroke. *Stroke Vasc. Neurol.* **2019**, *4*, 71–74. [[CrossRef](#)] [[PubMed](#)]
94. Iadecola, C. The pathobiology of vascular dementia. *Neuron* **2013**, *80*, 844–866. [[CrossRef](#)] [[PubMed](#)]
95. Filous, A.S.; Silver, J. Targeting astrocytes in CNS injury and disease: A translational research approach. *Prog. Neurobiol.* **2016**, *144*, 173–187. [[CrossRef](#)]
96. Forsberg, K.M.E.; Zhang, Y.; Reiners, J.; Ander, M.; Niedermayer, A.; Fang, L.; Neugebauer, H.; Kassubek, J.; Katona, I.; Weis, J.; et al. Endothelial damage, vascular bagging and remodeling of the microvascular bed in human microangiopathy with deep white matter lesions. *Acta Neuropathol. Commun.* **2018**, *6*, 128. [[CrossRef](#)] [[PubMed](#)]
97. Szalay, G.; Martinecz, B.; Lénárt, N.; Környei, Z.; Orsolits, B.; Judák, L.; Császár, E.; Fekete, R.; West, B.L.; Katona, G.; et al. Microglia protect against brain injury and their selective elimination dysregulates neuronal network activity after stroke. *Nat. Commun.* **2016**, *7*, 11499. [[CrossRef](#)] [[PubMed](#)]

98. Zhao, S.-C.; Ma, L.-S.; Chu, Z.-H.; Xu, H.; Wu, W.-Q.; Liu, F. Regulation of microglial activation in stroke. *Acta Pharmacol. Sin.* **2017**, *38*, 445–458. [[CrossRef](#)] [[PubMed](#)]
99. Morrison, H.W.; Filosa, J.A. A quantitative spatiotemporal analysis of microglia morphology during ischemic stroke and reperfusion. *J. Neuroinflamm.* **2013**, *10*, 4. [[CrossRef](#)]
100. Perego, C.; Fumagalli, S.; De Simoni, M.G. Temporal pattern of expression and colocalization of microglia/macrophage phenotype markers following brain ischemic injury in mice. *J. Neuroinflamm.* **2011**, *8*, 174. [[CrossRef](#)]
101. Farkas, E.; Donka, G.; de Vries, R.A.L.; Mihaly, A.; Bari, F.; Luiten, P.G.M. Experimental cerebral hypoperfusion induces white matter injury and microglial activation in the rat brain. *Acta Neuropathol.* **2004**, *108*, 57–64. [[CrossRef](#)]
102. Paolini Paoletti, F.; Simoni, S.; Parnetti, L.; Gaetani, L. The Contribution of Small Vessel Disease to Neurodegeneration: Focus on Alzheimer's Disease, Parkinson's Disease and Multiple Sclerosis. *Int. J. Mol. Sci.* **2021**, *22*, 4958. [[CrossRef](#)]
103. Fredman, P.; Wallin, A.; Blennow, K.; Davidsson, P.; Gottfries, C.; Svennerholm, L. Sulfatide as a biochemical marker in cerebrospinal fluid of patients with vascular dementia. *Acta Neurol. Scand.* **1992**, *85*, 103–106. [[CrossRef](#)] [[PubMed](#)]
104. Tullberg, M.; Månsson, J.E.; Fredman, P.; Lekman, A.; Blennow, K.; Ekman, R.; Rosengren, L.E.; Tisell, M.; Wikkelso, C. CSF sulfatide distinguishes between normal pressure hydrocephalus and subcortical arteriosclerotic encephalopathy. *J. Neurol. Neurosurg. Psychiatry* **2000**, *69*, 74–81. [[CrossRef](#)] [[PubMed](#)]
105. Thibert, K.A.; Raymond, G.V.; Nascene, D.R.; Miller, W.P.; Tolar, J.; Orchard, P.J.; Lund, T.C. Cerebrospinal fluid matrix metalloproteinases are elevated in cerebral adrenoleukodystrophy and correlate with MRI severity and neurologic dysfunction. *PLoS ONE* **2012**, *7*, e50430. [[CrossRef](#)] [[PubMed](#)]
106. Jonsson, M.; Zetterberg, H.; Van Straaten, E.; Lind, K.; Syversen, S.; Edman, Å.; Blennow, K.; Rosengren, L.; Pantoni, L.; Inzitari, D.; et al. Cerebrospinal fluid biomarkers of white matter lesions—Cross-sectional results from the LADIS study. *Eur. J. Neurol.* **2010**, *17*, 377–382. [[CrossRef](#)] [[PubMed](#)]
107. Bjerke, M.; Andreasson, U.; Rolstad, S.; Nordlund, A.; Lind, K.; Zetterberg, H.; Edman, Å.; Blennow, K.; Wallin, A. Subcortical vascular dementia biomarker pattern in mild cognitive impairment. *Dement. Geriatr. Cogn. Disord.* **2009**, *28*, 348–356. [[CrossRef](#)]
108. Öhrfelt, A.; Andreasson, U.; Simon, A.; Zetterberg, H.; Edman, Å.; Potter, W.; Holder, D.; Devanarayan, V.; Seeburger, J.; Smith, A.D.; et al. Screening for New Biomarkers for Subcortical Vascular Dementia and Alzheimer's Disease. *Dement. Geriatr. Cogn. Dis. Extra* **2011**, *1*, 31–42. [[CrossRef](#)]
109. Peters, A.; Sethares, C. Age-related changes in the morphology of cerebral capillaries do not correlate with cognitive decline. *J. Comp. Neurol.* **2012**, *520*, 1339–1347. [[CrossRef](#)]
110. Iejima, D.; Itabashi, T.; Kawamura, Y.; Noda, T.; Yuasa, S.; Fukuda, K.; Oka, C.; Iwata, T. HTRA1 (high temperature requirement A serine peptidase 1) gene is transcriptionally regulated by insertion/deletion nucleotides located at the 3' end of the ARMS2 (age related maculopathy susceptibility 2) gene in patients with age-related macular degeneration. *J. Biol. Chem.* **2015**, *290*, 2784–2797.
111. Zlokovic, B.V. Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat. Rev. Neurosci.* **2011**, *12*, 723–728. [[CrossRef](#)]
112. Cai, W.; Zhang, K.; Li, P.; Zhu, L.; Xu, J.; Yang, B.; Hu, X.; Lu, Z.; Chen, J. Dysfunction of the neurovascular unit in ischemic stroke and neurodegenerative diseases: An aging effect. *Ageing Res. Rev.* **2017**, *34*, 77–87. [[CrossRef](#)]
113. Prisby, R.D.; Ramsey, M.W.; Behnke, B.J.; Dominguez, J.M.; Donato, A.J.; Allen, M.R.; Delp, M.D. Aging reduces skeletal blood flow endothelium dependent vasodilation, and NO bioavailability in Rats. *J. Bone Miner. Res.* **2007**, *22*, 1280–1288. [[CrossRef](#)] [[PubMed](#)]
114. Nicholson, W.T.; Vaa, B.; Hesse, C.; Eisenach, J.H.; Joyner, M.J. Aging is associated with reduced prostacyclin-mediated dilation in the human forearm. *Hypertension* **2009**, *53*, 973–978. [[CrossRef](#)] [[PubMed](#)]
115. Van Dijk, E.J.; Prins, N.D.; Vermeer, S.E.; Vrooman, H.A.; Hofman, A.; Koudstaal, P.J.; Breteler, M.M.B. C-reactive protein and cerebral small-vessel disease: The Rotterdam Scan Study. *Circulation* **2005**, *112*, 900–905. [[CrossRef](#)]
116. Long, D.A.; Newaz, M.A.; Prabahakar, S.S.; Price, K.L.; Truong, L.; Feng, L.; Mu Oyekan, A.O.; Johnson, R.J. Loss of nitric oxide and endothelial-derived hyperpolarizing factor-mediated responses in ageing. *Kidney Int.* **2005**, *68*, 2154–2163. [[CrossRef](#)]
117. Van der Loo, B.; Labugger, R.; Skepper, J.N.B.; Achschmid, M.; Kilo, J.; Powell, J.M.; Palacios-Callendere, M.; Erusalimsky, J.D.; Quaschnig, T.; Malinski, T. Enhanced peroxynitrite formation is associated with vascular ageing. *J. Exp. Med.* **2000**, *18*, 1731–1744. [[CrossRef](#)]
118. Puca, A.A.; Carrizzo, A.; Ferrario, A.; Villa, F.; Vecchione, C. Endothelial nitric oxide synthase, vascular integrity and human exceptional longevity. *Immun. Ageing* **2012**, *9*, 26. [[CrossRef](#)]
119. Flentje, A.; Kalsi, R.; Monahan, T.S. Small GTPases and Their Role in Vascular Disease. *Int. J. Mol. Sci.* **2019**, *20*, 917. [[CrossRef](#)]
120. Hartmann, S.; Ridley, A.J.; Lutz, S. The Function of Rho-Associated Kinases ROCK1 and ROCK2 in the Pathogenesis of Cardiovascular Disease. *Front. Pharmacol.* **2015**, *6*, 276. [[CrossRef](#)]
121. Szulcek, R.; Beckers, C.M.; Hodzic, J.; de Wit, J.; Chen, Z.; Grob, T.; Musters, R.J.; Minshall, R.D.; van Hinsbergh, V.W.; van Nieuw Amerongen, G.P. Localized RhoA GTPase activity regulates dynamics of endothelial monolayer integrity. *Cardiovasc. Res.* **2013**, *99*, 471–482. [[CrossRef](#)]
122. Van Nieuw Amerongen, G.P.; Beckers, C.M.; Achekar, I.D.; Zeeman, S.; Musters, R.J.; van Hinsbergh, V.W. Involvement of Rho kinase in endothelial barrier maintenance. *Arterioscler. Thromb. Vasc. Biol.* **2007**, *27*, 2332–2339. [[CrossRef](#)]
123. Wang, J.; Liu, H.; Chen, B.; Li, Q.; Huang, X.; Wang, L.; Guo, X.; Huang, Q. RhoA/ROCK-dependent moesin phosphorylation regulates AGE-induced endothelial cellular response. *Cardiovasc. Diabetol.* **2012**, *11*, 7. [[CrossRef](#)] [[PubMed](#)]

124. Sun, H.; Breslin, J.W.; Zhu, J.; Yuan, S.Y.; Wu, M.H. Rho and ROCK signaling in VEGF-induced microvascular endothelial hyperpermeability. *Microcirculation* **2006**, *13*, 237–247. [[CrossRef](#)] [[PubMed](#)]
125. Simpson, J.E.; Fernando, M.S.; Clark, L.; Ince, P.G.; Matthews, F.; Forster, G.; O'Brien, J.T.; Barber, R.; Kalaria, R.N.; Brayne, C.; et al. White matter lesions in an unselected cohort of the elderly: Astrocytic, microglial and oligodendrocyte precursor cell responses. *Neuropathol. Appl. Neurobiol.* **2007**, *33*, 410–419. [[CrossRef](#)] [[PubMed](#)]
126. Sies, H.; Jones, D.P. Reactive oxygen species (ROS) as pleiotropic physiological signalling agents. *Nat. Rev. Mol. Cell Biol.* **2020**, *21*, 363–383. [[CrossRef](#)]
127. Pantoni, L.; Inzitari, D.; Pracucci, G.; Lolli, F.; Giordano, G.; Bracco, L.; Amaducci, L. Cerebrospinal fluid proteins in patients with leucoaraiosis: Possible abnormalities in blood-brain barrier function. *J. Neurol. Sci.* **1993**, *115*, 125–131. [[CrossRef](#)]
128. Musaeus, C.S.; Gleerup, H.S.; Høgh, P.; Waldemar, G.; Hasselbalch, S.G.; Simonsen, A.H. Cerebrospinal Fluid/Plasma Albumin Ratio as a Biomarker for Blood-Brain Barrier Impairment Across Neurodegenerative Dementias. *J. Alzheimers Dis.* **2020**, *75*, 429–436. [[CrossRef](#)]
129. Georgakis, M.K.; Chatzopoulou, D.; Tsvigoulis, G.; Petridou, E.T. Albuminuria and cerebral small vessel disease: A systematic review and meta-analysis. *J. Am. Geriatr. Soc.* **2018**, *66*, 509–517. [[CrossRef](#)]
130. Wada, M.; Takahashi, Y.; Iseki, C.; Kawanami, T.; Daimon, M.; Kato, T. Plasma fibrinogen, global cognitive function, and cerebral small vessel disease: Results of a cross-sectional study in community-dwelling Japanese elderly. *Intern. Med.* **2011**, *50*, 999–1007. [[CrossRef](#)]
131. Kulikauskas, M.R.; Shaka, X.; Bautch, V.L. The versatility and paradox of BMP signaling in endothelial cell behaviors and blood vessel function. *Cell. Mol. Life Sci.* **2022**, *79*, 77. [[CrossRef](#)]
132. Knottnerus, I.L.; Govers-Riemslog, J.W.; Hamulyak, K.; Rouhl, R.P.; Staals, J.; Spronk, H.M. Endothelial activation in lacunar stroke subtypes. *Stroke* **2010**, *41*, 1617–1622. [[CrossRef](#)]
133. Knottnerus, I.L.; Cate, H.; Lodder, J.; Kessels, F.; van Oostenbrugge, R.J. Endothelial dysfunction in lacunar stroke: A systematic review. *Cerebrovasc. Dis.* **2009**, *27*, 519–526. [[CrossRef](#)] [[PubMed](#)]
134. Stevenson, S.F.; Doubal, F.N.; Shuler, K.; Wardlaw, J.M. A systematic review of dynamic cerebral and peripheral endothelial function in lacunar stroke versus controls. *Stroke* **2010**, *41*, e434–e442. [[CrossRef](#)] [[PubMed](#)]
135. Markus, H.S.; Hunt, B.; Palmer, K.; Enzinger, C.; Schmidt, H.; Schmidt, R. Markers of endothelial and hemostatic activation and progression of cerebral white matter hyperintensities: Longitudinal results of the Austrian Stroke Prevention Study. *Stroke* **2005**, *36*, 1410–1414. [[CrossRef](#)] [[PubMed](#)]
136. Fernando, M.S.; Simpson, J.E.; Matthews, F.; Brayne, C.; Lewis, C.E.; Barber, R.; Kalaria, R.N.; Forster, G.; Esteves, F.; Wharton, S.B.; et al. White matter lesions in an unselected cohort of the elderly: Molecular pathology suggests origin from chronic hypoperfusion injury. *Stroke* **2006**, *37*, 1391–1398. [[CrossRef](#)]
137. Egea, J.; Fabregat, I.; Frapart, Y.M.; Ghezzi, P.; Görlach, A.; Kietzmann, T.; Kubaichuk, K.; Knaus, U.G.; Lopez, M.G.; Olaso-Gonzalez, G.; et al. European contribution to the study of ROS: A summary of the findings and prospects for the future from the COST action BM1203 (EU-ROS). *Redox Biol.* **2017**, *13*, 94–162, Erratum in *Redox Biol.* **2018**, *14*, 694–696. [[CrossRef](#)]
138. Dikalov, S.I.; Dikalova, A.E.; Bikineyeva, A.T.; Schmidt, H.H.; Harrison, D.G.; Griendling, K.K. Distinct roles of Nox1 and Nox4 in basal and angiotensin II-stimulated superoxide and hydrogen peroxide production. *Free Radic. Biol. Med.* **2008**, *45*, 1340–1351. [[CrossRef](#)]
139. Zhang, X.; Wu, B.; Nie, K.; Jia, Y.; Yu, J. Effects of acupuncture on declined cerebral blood flow, impaired mitochondrial respiratory function and oxidative stress in multi-infarct dementia rats. *Neurochem. Int.* **2014**, *65*, 23–29. [[CrossRef](#)]
140. Huang, J.L.; Fu, S.T.; Jiang, Y.Y.; Cao, Y.B.; Guo, M.L.; Wang, Y.; Xu, Z. Protective effects of Nicotiflorin on reducing memory dysfunction, energy metabolism failure, and oxidative stress in multi-infarct dementia model rats. *Pharmacol. Biochem. Behav.* **2007**, *86*, 741–748. [[CrossRef](#)]
141. Takac, I.; Schröder, K.; Brandes, R.P. The Nox family of NADPH oxidases: Friend or foe of the vascular system? *Curr. Hypertens. Rep.* **2012**, *14*, 70–78. [[CrossRef](#)]
142. Miller, A.A.; Drummond, G.R.; DeSilva, T.M.; Mast, A.E.; Hickey, H.; Williams, J.P.; Broughton, B.R.; Sobey, C.G. NADPH oxidase activity is higher in cerebral versus systemic arteries of four animal species: Role of Nox2. *Am. J. Physiol. Heart Circ. Physiol.* **2009**, *296*, H220–H225. [[CrossRef](#)]
143. Liu, H.; Zhang, J. Cerebral hypoperfusion and cognitive impairment: The pathogenic role of vascular oxidative stress. *Int. J. Neurosci.* **2012**, *122*, 494–499. [[CrossRef](#)] [[PubMed](#)]
144. Paravicini, T.M.; Miller, A.A.; Drummond, G.R.; Sobey, C.G. Flow-induced cerebral vasodilatation in vivo involves activation of phosphatidylinositol-3kinase, NADPH-oxidase, and nitric oxide synthase. *J. Cereb. Blood Flow Metab.* **2006**, *26*, 836–845. [[CrossRef](#)] [[PubMed](#)]
145. De Silva, T.M.; Brait, V.H.; Drummond, G.R.; Sobey, C.G.; Miller, A.A. Nox2 oxidase activity accounts for the oxidative stress and vasomotor dysfunction in mouse cerebral arteries following ischemic stroke. *PLoS ONE* **2011**, *6*, e28393. [[CrossRef](#)] [[PubMed](#)]
146. Lin, M.T.; Beal, M.F. Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature* **2006**, *443*, 787–795. [[CrossRef](#)]
147. Raz, N. The aging brain: Structural changes and their implications for cognitive aging. In *New Frontiers in Cognitive Aging*; Dixon, R., Bäckman, L., Nilsson, L., Eds.; Oxford University Press: Telangana, India, 2004; pp. 115–134.
148. Sun, N.; Youle, R.J.; Finkel, T. The mitochondrial basis of aging. *Mol. Cell* **2016**, *61*, 654–666. [[CrossRef](#)]

149. Valiente-Pallejà, A.; Tortajada, J.; Bulduk, B.K.; Vilella, E.; Garrabou, G.; Muntané, G.; Martorell, L. Comprehensive summary of mitochondrial DNA alterations in the postmortem human brain: A systematic review. *EBioMedicine* **2022**, *76*, 103815. [[CrossRef](#)]
150. Corral-Debrinski, M.; Horton, T.; Lott, M.T.; Shoffner, J.M.; Beal, M.F.; Wallace, D.C. Mitochondrial DNA deletions in human brain: Regional variability and increase with advanced age. *Nat. Genet.* **1992**, *2*, 324–329. [[CrossRef](#)]
151. Taylor, S.D.; Ericson, N.G.; Burton, J.N.; Prolla, T.A.; Silber, J.R.; Shendure, J.; Bielas, J.H. Targeted enrichment and high-resolution digital profiling of mitochondrial DNA deletions in human brain. *Aging Cell* **2014**, *13*, 29–38. [[CrossRef](#)]
152. Kennedy, S.R.; Salk, J.J.; Schmitt, M.W.; Loeb, L.A. Ultra-sensitive sequencing reveals an age-related increase in somatic mitochondrial mutations that are inconsistent with oxidative damage. *PLoS Genet.* **2013**, *9*, 1003794. [[CrossRef](#)]
153. Diaz, F.; Bayona-Bafaluy, M.P.; Rana, M.; Mora, M.; Hao, H.; Moraes, C.T. Human mitochondrial DNA with large deletions repopulates organelles faster than full-length genomes under relaxed copy number control. *Nucleic Acids Res.* **2002**, *30*, 4626–4633. [[CrossRef](#)]
154. Guo, L.; Tian, J.; Du, H. Mitochondrial dysfunction and synaptic transmission failure in Alzheimer's disease. *J. Alzheimers Dis.* **2017**, *57*, 1071–1086. [[CrossRef](#)] [[PubMed](#)]
155. Ballif, B.C.; Theisen, A.; Coppinger, J.; Gowans, G.C.; Hersh, J.H.; Madan-Khetarpal, S.; Schmidt, K.R.; Tervo, R.; Escobar, L.F.; Friedrich, C.A.; et al. Expanding the clinical phenotype of the 3q29 microdeletion syndrome and characterization of the reciprocal microduplication. *Mol. Cytogenet.* **2008**, *1*, 8. [[CrossRef](#)] [[PubMed](#)]
156. Lanza, G.; Cantone, M.; Musso, S.; Borgione, E.; Scuderi, C.; Ferri, R. Early-onset subcortical ischemic vascular dementia in an adult with mtDNA mutation 3316G>A. *J. Neurol.* **2018**, *265*, 968–969. [[CrossRef](#)] [[PubMed](#)]
157. Campbell, G.R.; Ziabreva, I.; Reeve, A.K.; Krishnan, K.J.; Reynolds, R.; Howell, O.; Lassmann, H.; Turnbull, D.M.; Mahad, D.J. Mitochondrial DNA deletions and neurodegeneration in multiple sclerosis. *Ann. Neurol.* **2011**, *69*, 481–492. [[CrossRef](#)]
158. Chinnery, P.F. Mitochondrial disorders overview. In *GeneReviews [Internet]*; Margaret, P.A., Ardinger, H.H., Pagon, R.A., Wallace, S.E., Bean, L.J.H., Karen, S.A.A., Eds.; University of Washington, Seattle: Seattle, WA, USA, 2020; p. 1993.
159. Basel, D. Mitochondrial DNA Depletion Syndromes. *Clin. Perinatol.* **2020**, *47*, 123–141. [[CrossRef](#)]
160. Coskun, P.E.; Wyrembak, J.; Derbereva, O.; Melkonian, G.; Doran, E.; Lott, I.T.; Head, E.; Cotman, C.W.; Wallace, D.C. Systemic mitochondrial dysfunction and the etiology of Alzheimer's disease and down syndrome dementia. *J. Alzheimers Dis.* **2010**, *20*, 293–310. [[CrossRef](#)]
161. Roca-Bayerri, C.; Robertson, F.; Pyle, A.; Hudson, G.; Payne, B.A.I. Mitochondrial DNA damage and brain aging in human immunodeficiency virus. *Clin. Infect. Dis.* **2021**, *73*, e466–e473. [[CrossRef](#)]
162. Touyz, R.M.; Briones, A.M. Reactive oxygen species and vascular biology: Implications in human hypertension. *Hypertens. Res.* **2011**, *34*, 5–14. [[CrossRef](#)]
163. Mayhan, W.G.; Arrick, D.M.; Sharpe, G.M.; Sun, H. Age-related alterations in reactivity of cerebral arterioles: Role of oxidative stress. *Microcirculation* **2008**, *15*, 225–236. [[CrossRef](#)]
164. Dong, Y.F.; Kataoka, K.; Toyama, K.; Sueta, D.; Koibuchi, N.; Yamamoto, E.; Yata, K.; Tomimoto, H.; Ogawa, H.; Kim-Mitsuyama, S. Attenuation of brain damage and cognitive impairment by direct renin inhibition in mice with chronic cerebral hypoperfusion. *Hypertension* **2011**, *58*, 635–642. [[CrossRef](#)]
165. Santhanam, A.V.; d'Uscio, L.V.; Katusic, Z.S. Erythropoietin increases bioavailability of tetrahydrobiopterin and protects cerebral microvasculature against oxidative stress induced by eNOS uncoupling. *J. Neurochem.* **2014**, *131*, 521–529. [[CrossRef](#)] [[PubMed](#)]
166. Xie, H.; Ray, P.E.; Short, B.L. NF-kappa B activation plays a role in superoxide-mediated cerebral dysfunction after hypoxia/reoxygenation. *Stroke* **2005**, *36*, 1047–1052. [[CrossRef](#)] [[PubMed](#)]
167. Aghajanian, A.; Witthen, E.S.; Campbell, S.L.; BurrIDGE, K. Direct activation of RhoA by reactive oxygen species requires a redox-sensitive motif. *PLoS ONE* **2009**, *4*, e8045. [[CrossRef](#)] [[PubMed](#)]
168. Faraco, G.; Moraga, A.; Moore, J.; Anrather, J.; Pickel, V.M.; Iadecola, C. Circulating endothelin-1 alters critical mechanisms regulating cerebral microcirculation. *Hypertension* **2013**, *62*, 759–766. [[CrossRef](#)]
169. Bochkov, V.N.; Philippova, M.; Oskolkova, O.; Kadl, A.; Furnkranz, A.; Karabeg, E.; Afonyushkin, T.; Gruber, F.; Breuss, J.; Minchenko, A.; et al. Oxidized phospholipids stimulate angiogenesis via autocrine mechanisms, implicating a novel role for lipid oxidation in the evolution of atherosclerotic lesions. *Circ. Res.* **2006**, *99*, 900–908. [[CrossRef](#)]
170. Tai, L.M.; Thomas, R.; Marottoli, F.M.; Koster, K.P.; Kanekiyo, T.; Morris, A.W.; Bu, G. The role of APOE in cerebrovascular dysfunction. *Acta Neuropathol.* **2016**, *131*, 709–723. [[CrossRef](#)]
171. Han, B.H.; Zhou, M.L.; Johnson, A.W.; Singh, I.; Liao, F.; Vellimana, A.K.; Nelson, J.W.; Milner, E.; Cirrito, J.R.; Basak, J.; et al. Contribution of reactive oxygen species to cerebral amyloid angiopathy, vasomotor dysfunction, and micro-hemorrhage in aged Tg2576 mice. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, E881–E890. [[CrossRef](#)]
172. Grochowski, C.; Litak, J.; Kamieniak, P.; Maciejewski, R. Oxidative stress in cerebral small vessel disease. Role of reactive species. *Free Radic. Res.* **2018**, *52*, 1–13. [[CrossRef](#)]
173. Han, S.; Wu, H.; Li, W.; Gao, P. Protective effects of genistein in homocysteine-induced endothelial cell inflammatory injury. *Mol. Cell. Biochem.* **2015**, *403*, 43–49. [[CrossRef](#)]
174. Fleszar, M.G.; Wiśniewski, J.; Zboch, M.; Diakowska, D.; Gamian, A.; Krzystek-Korpacka, M. Targeted metabolomic analysis of nitric oxide/L-arginine pathway metabolites in dementia: Association with pathology, severity, and structural brain changes. *Sci. Rep.* **2019**, *9*, 1376. [[CrossRef](#)]

175. Gao, Q.; Fan, Y.; Mu, L.-Y.; Ma, L.; Song, Z.-Q.; Zhang, Y.-N. S100B and ADMA in cerebral small vessel disease and cognitive dysfunction. *J. Neurol. Sci.* **2015**, *354*, 27–32. [[CrossRef](#)] [[PubMed](#)]
176. Vinciguerra, L.; Lanza, G.; Puglisi, V.; Fiscaro, F.; Pennisi, M.; Bella, R.; Cantone, M. Update on the Neurobiology of Vascular Cognitive Impairment: From Lab to Clinic. *Int. J. Mol. Sci.* **2020**, *21*, 2977. [[CrossRef](#)] [[PubMed](#)]
177. Li, J.-J.; Li, Q.; Du, H.-P. Homocysteine Triggers inflammatory responses in macrophages through inhibiting CSE-H2S signaling via DNA hypermethylation of CSE promoter. *Int. J. Mol. Sci.* **2015**, *16*, 12560–12577. [[CrossRef](#)] [[PubMed](#)]
178. Moretti, R.; Giuffr , M.; Caruso, P.; Gazzin, S.; Tiribelli, C. Homocysteine in Neurology: A Possible Contributing Factor to Small Vessel Disease. *Int. J. Mol. Sci.* **2021**, *22*, 2051. [[CrossRef](#)]
179. Ahmad, S.; Siddiqi, M.I. Insights from molecular modeling into the selective inhibition of cathepsin S by its inhibitor. *J. Mol. Model.* **2017**, *23*, 92. [[CrossRef](#)]
180. Leng, Y.P.; Ma, Y.S.; Li, X.G.; Chen, R.F.; Zeng, P.Y.; Li, X.H.; Qiu, C.F.; Li, Y.P.; Zhang, Z.; Chen, A.F. I-Homocysteine-induced cathepsin V mediates the vascular endothelial inflammation in hyperhomocysteinemia. *Br. J. Pharmacol.* **2018**, *175*, 1157–1172. [[CrossRef](#)]
181. Moretti, R.; Dal Ben, M.; Gazzin, S.; Tiribelli, C. Homocysteine in neurology: From endothelium to neurodegeneration. *Curr. Nutr. Food Sci.* **2017**, *13*, 163–175. [[CrossRef](#)]
182. Deng, J.; Lu, S.; Li, H. Homocysteine activates B cells via regulating PKM-2 dependent metabolic reprogramming. *J. Immunol.* **2017**, *198*, 170–183. [[CrossRef](#)]
183. Kumar, A.; Palfrey, H.A.; Pathak, R.; Kadowitz, P.J.; Gettys, T.W.; Murthy, S.N. The metabolism and significance of homocysteine in nutrition and health. *Nutr. Metab.* **2017**, *14*, 78. [[CrossRef](#)]
184. Dayal, S.; Wilson, K.M.; Leo, L.; Arning, E.; Bottiglieri, T.; Lentz, S.R. Enhanced susceptibility to arterial thrombosis in a murine model of hyperhomocysteinemia. *Blood* **2006**, *108*, 2237–2243. [[CrossRef](#)]
185. Perla-Kajan, J.; Twardowski, T.; Jakubowski, H. Mechanisms of homocysteine toxicity in humans. *Amino Acids* **2007**, *32*, 561–572. [[CrossRef](#)] [[PubMed](#)]
186. Li, T.; Chen, Y.; Li, J.; Yang, X.; Zhang, H.; Qin, X.; Hu, Y.; Mo, Z. Serum Homocysteine Concentration Is Significantly Associated with Inflammatory/Immune Factors. *PLoS ONE* **2015**, *10*, e0138099. [[CrossRef](#)] [[PubMed](#)]
187. Reddy, V.S.; Trinath, J.; Reddy, G.B. Implication of homocysteine in protein quality control processes. *Biochimie* **2019**, *165*, 19–31. [[CrossRef](#)] [[PubMed](#)]
188. Heo, J.H.; Lucero, J.; Abumiya, T.; Koziol, J.A.; Copeland, B.R.; del Zoppo, G.J. Matrix metalloproteinases increase very early during experimental focal cerebral ischemia. *J. Cereb. Blood Flow Metab.* **1999**, *19*, 624–633. [[CrossRef](#)]
189. Zhang, M.; Zhu, W.; Yun, W.; Wang, Q.; Cheng, M.; Zhang, Z.; Liu, X.; Zhou, X.; Xu, G. Correlation of matrix metalloproteinase-2 single nucleotide polymorphisms with the risk of small vessel disease (SVD). *J. Neurol. Sci.* **2015**, *356*, 61–64. [[CrossRef](#)]
190. Yang, Y.; Estrada, E.Y.; Thompson, J.F.; Liu, W.; Rosenberg, G.A. Matrix metalloproteinase-mediated disruption of tight junction proteins in cerebral vessels is reversed by synthetic matrix metalloproteinase inhibitor in focal ischemia in rat. *J. Cereb. Blood Flow Metab.* **2007**, *27*, 697–709. [[CrossRef](#)]
191. Stamenkovic, I. Extracellular matrix remodeling: The role of matrix metalloproteinases. *J. Pathol.* **2003**, *200*, 448–464. [[CrossRef](#)]
192. English, W.R.; Suarez-Puente, X.S.; Freije, J.M.; Knauper, V.; Amour, A.; Merryweather, A.; L pez-Ot n, C.; Murphy, G. Membrane type 4 matrix metalloproteinase (MMP17) has tumor necrosis factor- α convertase activity but does not activate pro-MMP2. *J. Biol. Chem.* **2000**, *275*, 14046–14055. [[CrossRef](#)]
193. Kim, Y.S.; Choi, D.H.; Block, M.L.; Lorenzl, S.; Yang, L.; Kim, Y.J.; Sugama, S.; Cho, B.P.; Hwang, O.; Browne, S.E.; et al. A pivotal role of matrix metalloproteinase-3 activity in dopaminergic neuronal degeneration via microglial activation. *FASEB J.* **2007**, *21*, 179–187. [[CrossRef](#)]
194. Woo, M.S.; Park, J.S.; Choi, I.Y.; Kim, W.K.; Kim, H.S. Inhibition of MMP-3 or -9 suppresses lipopolysaccharide-induced expression of proinflammatory cytokines and iNOS in microglia. *J. Neurochem.* **2008**, *106*, 770–780. [[CrossRef](#)]
195. Powell, W.C.; Fingleton, B.; Wilson, C.L.; Boothby, M.; Matrisian, L.M. The metalloproteinase matrilysin proteolytically generates active soluble Fas ligand and potentiates epithelial cell apoptosis. *Curr. Biol.* **1999**, *9*, 1441–1447. [[CrossRef](#)]
196. Gu, Y.; Zheng, G.; Xu, M.; Li, Y.; Chen, X.; Zhu, W.; Tong, Y.; Chung, S.K.; Liu, K.J.; Shen, J. Caveolin-1 regulates nitric oxide-mediated matrix metalloproteinases activity and blood–brain barrier permeability in focal cerebral ischemia and reperfusion injury. *J. Neurochem.* **2012**, *120*, 147–156. [[CrossRef](#)] [[PubMed](#)]
197. Chandler, S.; Miller, K.M.; Clements, J.M.; Lury, J.; Corkill, D.; Anthony, D.C.C.; Adams, S.E.; Gearing, A.J.H. Matrix metalloproteinases, tumor necrosis factor and multiple sclerosis: An overview. *J. Neuroimmunol.* **1997**, *72*, 155–161. [[CrossRef](#)]
198. Inzitari, D.; Giusti, B.; Nencini, P.; Gori, A.M.; Nesi, M.; Palumbo, V.; Piccardi, B.; Armillis, A.; Pracucci, G.; Bono, G.; et al. MMP9 Variation After Thrombolysis Is Associated with Hemorrhagic Transformation of Lesion and Death. *Stroke* **2013**, *44*, 2901–2903. [[CrossRef](#)] [[PubMed](#)]
199. Montaner, J.; Molina, C.A.; Monasterio, J.; Abilleira, S.; Arenillas, J.F.; Ribo, M.; Quintana, M.; Alvarez-Sabin, J. Matrix Metalloproteinase-9 Pretreatment Level Predicts Intracranial Hemorrhagic Complications After Thrombolysis in Human Stroke. *Circulation* **2003**, *107*, 598–603. [[CrossRef](#)]
200. Arba, F.; Piccardi, B.; Palumbo, V.; Giusti, B.; Nencini, P.; Gori, A.M.; Sereni, A.; Nesi, M.; Pracucci, G.; Bono, G.; et al. Small Vessel Disease Is Associated with Tissue Inhibitor of Matrix Metalloproteinase-4 After Ischaemic Stroke. *Transl. Stroke Res.* **2018**, *10*, 44–51. [[CrossRef](#)]

201. Candelario-Jalil, E.; Thompson, J.; Taheri, S.; Grossetete, M.; Adair, J.C.; Edmonds, E.; Prestopnik, J.; Wills, J.; Rosenberg, G.A. Matrix Metalloproteinases Are Associated with Increased Blood–Brain Barrier Opening in Vascular Cognitive Impairment. *Stroke* **2011**, *42*, 1345–1350. [[CrossRef](#)]
202. Rosenberg, G.A.; Sullivan, N.; Esiri, M.M. White matter damage is associated with matrix metalloproteinases in vascular dementia. *Stroke* **2001**, *32*, 1162–1168. [[CrossRef](#)]
203. Rosenberg, G.A. Inflammation and white matter damage in vascular cognitive impairment. *Stroke* **2009**, *40*, S20–S23. [[CrossRef](#)]
204. Ketsawatsomkron, P.; Keen, H.L.; Davis, D.R.; Lu, K.T.; Stump, M.; De Silva, T.M.; Hilzendeger, A.M.; Grobe, J.L.; Faraci, F.M.; Sigmund, C.D. Protective role for tissue inhibitor of Metalloproteinase-4, a novel peroxisome proliferator-activated receptor- γ target gene, in smooth muscle in Deoxycorticosterone acetate-salt hypertension. *Hypertension* **2016**, *67*, 214–222. [[CrossRef](#)]
205. Radomski, A.; Jurasz, P.; Sanders, E.J.; Overall, C.M.; Bigg, H.F.; Edwards, D.R.; Radomski, M.W. Identification, regulation and role of tissue inhibitor of metalloproteinases-4 (TIMP-4) in human platelets. *Br. J. Pharmacol.* **2002**, *137*, 1330–1338. [[CrossRef](#)] [[PubMed](#)]
206. Tomimoto, H.; Akiguchi, I.; Wakita, H.; Osaki, A.; Hayashi, M.; Yamamoto, Y. Coagulation activation in patients with Binswanger disease. *Arch. Neurol.* **1999**, *56*, 1104–1108. [[CrossRef](#)]
207. Iwamoto, T.; Kubo, H.; Takasaki, M. Platelet activation in the cerebral circulation in different subtypes of ischaemic stroke and Binswanger’s disease. *Stroke* **1995**, *26*, 52–56. [[CrossRef](#)] [[PubMed](#)]
208. Bjerke, M.; Zetterberg, H.; Edman, A.; Blennow, K.; Wallin, A.; Andreasson, U. Cerebrospinal fluid matrix metalloproteinases and tissue inhibitor of metalloproteinases in combination with subcortical and cortical biomarkers in vascular dementia and Alzheimer’s disease. *J. Alzheimers Dis.* **2011**, *27*, 665–676. [[CrossRef](#)] [[PubMed](#)]
209. Zhang, J.; Liu, N.; Yang, C. Effects of rosuvastatin in combination with nimodipine in patients with mild cognitive impairment caused by cerebral small vessel disease. *Panminerva Med.* **2019**, *61*, 439–443. [[CrossRef](#)]
210. Moretti, R.; Torre, P.; Antonello, R.M.; Cazzato, G.; Pizzolato, G. Different responses to rivastigmine in subcortical vascular dementia and multi-infarct dementia. *Am. J. Alzheimers Dis. Other Dementias* **2008**, *23*, 167–176. [[CrossRef](#)]
211. Laleu, B.; Gaggini, F.; Orchard, M.; Fioraso-Cartier, L.; Cagnon, L.; Houngninou-Molango, S.; Gradia, A.; Duboux, G.; Merlot, C.; Heitz, F.; et al. First in class, potent, and orally bioavailable NADPH oxidase isoform 4 (Nox4) inhibitors for the treatment of idiopathic pulmonary fibrosis. *J. Med. Chem.* **2010**, *53*, 7715–7730. [[CrossRef](#)]
212. De Silva, T.M.; Miller, A.A. Cerebral Small Vessel Disease: Targeting Oxidative Stress as a Novel Therapeutic Strategy? *Front. Pharmacol.* **2016**, *7*, 61. [[CrossRef](#)]
213. Pretnar-Oblak, J.; Sebestjen, M.; Sabovic, M. Statin treatment improves cerebral more than systemic endothelial dysfunction in patients with arterial hypertension. *Am. J. Hypertens.* **2008**, *21*, 674–678. [[CrossRef](#)]
214. Amarenco, P.; Benavente, O.; Goldstein, L.B.; Callahan, A.; Silleisen, H.; Hennerici, M.G.; Gilbert, S.; Rudolph, A.E.; Simunovic, L.; Zivin, J.A.; et al. Stroke Prevention by Aggressive Reduction in Cholesterol Levels Investigators. Results of the stroke prevention by aggressive reduction in cholesterol levels (SPARCL) trial by stroke subtypes. *Stroke* **2009**, *40*, 1405–1409. [[CrossRef](#)]
215. Caruso, G.; Godos, J.; Privitera, A.; Lanza, G.; Castellano, S.; Chillemi, A.; Bruni, O.; Ferri, R.; Caraci, F.; Grosso, G. Phenolic Acids and Prevention of Cognitive Decline: Polyphenols with a Neuroprotective Role in Cognitive Disorders and Alzheimer’s Disease. *Nutrients* **2022**, *14*, 819. [[CrossRef](#)] [[PubMed](#)]
216. Del Rio, D.; Rodriguez-Mateos, A.; Spencer, J.P.E.; Tognolini, M.; Borges, G.; Crozier, A. Dietary (poly)phenolics in human health: Structures, bioavailability, and evidence of protective effects against chronic diseases. *Antioxid. Redox Signal.* **2013**, *18*, 1818–1892. [[CrossRef](#)] [[PubMed](#)]
217. Manach, C.; Scalbert, A.; Morand, C.; Rémésy, C.; Jiménez, L. Polyphenols: Food sources and bioavailability. *Am. J. Clin. Nutr.* **2004**, *79*, 727–747. [[CrossRef](#)] [[PubMed](#)]
218. Cullen, A.E.; Centner, A.M.; Deitado, R.; Salazar, J.F.A. The Impact of Dietary Supplementation of Whole Foods and Polyphenols on Atherosclerosis. *Nutrients* **2020**, *12*, 2069. [[CrossRef](#)] [[PubMed](#)]
219. Kesse-Guyot, E.; Fezeu, L.; Andreeva, V.A.; Touvier, M.; Scalbert, A.; Hercberg, S.; Galan, P. Total and specific polyphenol intakes in midlife are associated with cognitive function measured 13 years later. *J. Nutr.* **2012**, *142*, 76–83. [[CrossRef](#)]
220. Goni, L.; Fernández-Matarrubia, M.; Romanos-Nanclares, A.; Razquin, C.; Ruiz-Canela, M.; Martínez-González, M.Á.; Toledo, E. Polyphenol intake and cognitive decline in the Seguimiento Universidad de Navarra (SUN) Project. *Br. J. Nutr.* **2021**, *126*, 43–52. [[CrossRef](#)]
221. Shakoor, H.; Feehan, J.; Apostolopoulos, V.; Platat, C.; Al Dhaheri, A.S.; Ali, H.I.; Ismail, L.C.; Bosevski, M.; Stojanovska, L. Immunomodulatory Effects of Dietary Polyphenols. *Nutrients* **2021**, *13*, 728. [[CrossRef](#)]
222. Godos, J.; Caraci, F.; Micek, A.; Castellano, S.; D’Amico, E.; Paladino, N.; Ferri, R.; Galvano, F.; Grosso, G. Dietary Phenolic Acids and Their Major Food Sources Are Associated with Cognitive Status in Older Italian Adults. *Antioxidants* **2021**, *10*, 700. [[CrossRef](#)]
223. Ran, L.S.; Liu, W.H.; Fang, Y.Y.; Xu, S.B.; Li, J.; Luo, X.; Pan, D.J.; Wang, M.H.; Wang, W. Alcohol, coffee and tea intake and the risk of cognitive deficits: A dose-response meta-analysis. *Epidemiol. Psychiatr. Sci.* **2021**, *30*, e13. [[CrossRef](#)]
224. Mallik, S.B.; Mudgal, J.; Nampoothiri, M.; Hall, S.; Dukie, S.A.; Grant, G.; Rao, C.M.; Arora, D. Caffeic acid attenuates lipopolysaccharide-induced sickness behaviour and neuroinflammation in mice. *Neurosci. Lett.* **2016**, *632*, 218–223. [[CrossRef](#)]
225. Lee, A.Y.; Wu, T.T.; Hwang, B.R.; Lee, J.; Lee, M.-H.; Lee, S.; Cho, E.J. The Neuro-Protective Effect of the Methanolic Extract of *Perilla frutescens* var. *japonica* and Rosmarinic Acid against H₂O₂-Induced Oxidative Stress in C6 Glial Cells. *Biomol. Ther.* **2016**, *24*, 338–345. [[CrossRef](#)] [[PubMed](#)]

226. De Mello Andrade, J.M.; Dos Santos Passos, C.; Kieling Rubio, M.A.; Mendonça, J.N.; Lopes, N.P.; Henriques, A.T. Combining in vitro and in silico approaches to evaluate the multifunctional profile of rosmarinic acid from *Blechnum brasiliense* on targets related to neurodegeneration. *Chem. Biol. Interact.* **2016**, *254*, 135–145. [[CrossRef](#)] [[PubMed](#)]
227. Rahbardar, M.G.; Amin, B.; Mehri, S.; Mirnajafi-Zadeh, S.J.; Hosseinzadeh, H. Anti-inflammatory effects of ethanolic extract of *Rosmarinus officinalis* L. and rosmarinic acid in a rat model of neuropathic pain. *Biomed. Pharmacother.* **2017**, *86*, 441–449. [[CrossRef](#)] [[PubMed](#)]
228. Dragomanova, S.; Pavlov, S.; Marinova, D.; Hodzev, Y.; Petralia, M.C.; Fagone, P.; Nicoletti, F.; Lazarova, M.; Tzvetanova, E.; Alexandrova, A.; et al. Neuroprotective Effects of Myrtenal in an Experimental Model of Dementia Induced in Rats. *Antioxidants* **2022**, *11*, 374. [[CrossRef](#)] [[PubMed](#)]
229. Lanza, G.; Bramanti, P.; Cantone, M.; Pennisi, M.; Pennisi, G.; Bella, R. Vascular Cognitive Impairment through the Looking Glass of Transcranial Magnetic Stimulation. *Behav. Neurol.* **2017**, *2017*, 1421326. [[CrossRef](#)]