


BMJ Open Plasma-glycated CD59 as an early biomarker for gestational diabetes mellitus: prospective cohort study protocol

Chloe Andrews ¹, Michelle Toth-Castillo,² Huseyin Aktas,^{2,3} Miguel-Angel Luque Fernandez,⁴ Steven Koon Wong,⁵ Sarbattama Sen,^{1,3} Jose Halperin^{2,3}

To cite: Andrews C, Toth-Castillo M, Aktas H, *et al.* Plasma-glycated CD59 as an early biomarker for gestational diabetes mellitus: prospective cohort study protocol. *BMJ Open* 2022;**12**:e054773. doi:10.1136/bmjopen-2021-054773

► Prepublication history for this paper is available online. To view these files, please visit the journal online (<http://dx.doi.org/10.1136/bmjopen-2021-054773>).

CA and MT-C contributed equally.

Received 23 June 2021
Accepted 28 March 2022



© Author(s) (or their employer(s)) 2022. Re-use permitted under CC BY-NC. No commercial re-use. See rights and permissions. Published by BMJ.

¹Pediatric Newborn Medicine, Brigham and Women's Hospital, Boston, Massachusetts, USA

²Department of Medicine, Brigham and Women's Hospital, Boston, Massachusetts, USA

³Harvard Medical School, Boston, Massachusetts, USA

⁴Department of Epidemiology and Population Health, London School of Hygiene and Tropical Medicine, London, UK

⁵Department of Digital Health eCare, Mass General Brigham Inc, Boston, Massachusetts, USA

Correspondence to

Professor Jose Halperin;
jhalperin@bwh.harvard.edu

ABSTRACT:

Introduction The significant maternal and neonatal outcomes of gestational diabetes mellitus (GDM) make it a major public health concern. Mothers with GDM are at greater risk of pregnancy complications and their offspring are at higher risk of diabetes and obesity. Currently, GDM is diagnosed with glucose load methods which are time-consuming and inconvenient to administer more than once during pregnancy; for this reason, there is a recognised need for a more accurate and simpler test for GDM. Previous studies indicate that plasma-glycated CD59 (pGCD59) is a novel biomarker for GDM. We present here the protocol of a prospective cohort study designed to (1) determine the accuracy of pGCD59 as an early, first trimester predictor of GDM and gestational impaired glucose tolerance and (2) assess the associations between pGCD59 levels and adverse maternal and neonatal outcomes.

Methods and analysis We will obtain discarded plasma samples from pregnant women at two time points: first prenatal visit (usually <14 weeks gestation) and gestational weeks 24–28. A study-specific medical record abstraction tool will be used to obtain relevant maternal and neonatal clinical data from the EPIC clinical database. The prevalence of GDM will be determined using standard of care glucose load test results. We will determine the sensitivity and specificity of pGCD59 to predict the diagnosis of GDM and gestational impaired glucose tolerance, as well as the associations between levels of pGCD59 and the prevalence of maternal and neonatal outcomes.

Ethics and dissemination This study has been approved by the Mass General Brigham Institutional Review Board (protocol 2011P002254). The results of this study will be presented at international meetings and disseminated in peer-reviewed journals.

INTRODUCTION

Gestational diabetes mellitus (GDM), defined as glucose intolerance that first occurs or is identified in pregnancy, is estimated to affect 7%–10% of pregnancies in the USA every year.^{1,2} This condition represents a major risk

Strengths and limitations of this study

- This study uses a biorepository of discarded clinical samples from high volume obstetrics clinics to achieve a large sample size and high proportion of gestational diabetes mellitus cases.
- This study leverages longitudinal electronic medical record data to obtain detailed assessments of maternal and neonatal outcomes.
- This study relies on accurate clinical documentation which may yield heterogeneity in our data.
- Longer-term infant follow-up after the postpartum hospitalisation is beyond the scope of this study and thus, we will not be able to determine associations between plasma-glycated CD59 levels and long-term offspring outcomes.

factor for both maternal and infant adverse pregnancy outcomes: for mothers with GDM, there is a greater risk of pre-eclampsia, caesarean section and preterm delivery; for their infants, there is a greater risk of macrosomia, large-for-gestational age (LGA), neonatal hypoglycaemia and future development of obesity, diabetes and cardiovascular disease.^{3–8} Treatment of GDM, as well as weight control following diagnosis,⁹ reduces the incidence of some adverse outcomes, strongly underscoring the importance of accurate and minimally burdensome universal screening, as well as early diagnosis.¹⁰

Current guidelines recommend GDM screening at 24–28 weeks gestation. However, markers of glycaemic load used in the non-pregnant population, such as fasting plasma glucose, haemoglobin A1c (HbA1c) and fructosamine, notably fail as screening or diagnostic tests for GDM. HbA1c, the standard of care test for management of individuals with diabetes, provides an integrated measure of glycaemic load over a period of 6–8 weeks

but has poor sensitivity for the screening/diagnosis of GDM due in part to the increased turnover rate of erythrocytes during pregnancy. Furthermore, the worsening insulin resistance in pregnancy is initially compensated for by an increase in insulin production, resulting in normal or close to normal fasting glucose levels.^{11 12} This also explains the low sensitivity of fasting plasma glucose, which provides a transient snapshot of glucose levels.¹² Regarding A1c and fructosamine, four studies that evaluated several HbA1c thresholds using different diagnostic criteria for GDM failed to identify a clear pattern between A1c levels and probability of GDM diagnosed by Carpenter-Coustan (C&C) criteria.¹³ Similarly, several human studies demonstrated that fructosamine, another marker that integrates glycaemic load over time, has very limited value as a screening test for GDM.^{14 15}

The low sensitivity of traditional glycaemic markers in pregnancy explains why glucose load tests (GLTs) are still universally used as the standard of care. A 'two-step' approach, consisting of a 1-hour 50-g GLT followed by a 3-hour 100-g oral glucose tolerance test (OGTT) for those who fail the GLT, is mostly used in the USA, whereas a 'one-step' approach, consisting of a 2-hour 75-g load, is used in many countries around the world.¹⁶ Both approaches are expensive, time consuming, unpleasant and have poor reproducibility on repeat testing of the same individual; moreover, their predictive value is low and differs according to ethnic origin.¹⁷ The failure rate of the OGTT to make a diagnosis of GDM is nearly 10%, with vomiting during the test being the major reason for this failure.¹⁷ In addition, the seminal Hyperglycemia and Adverse Pregnancy Outcome study has clearly documented a continuous positive association between increased values of maternal glycaemia and the risk of adverse maternal and fetal outcomes.⁴ As a consequence, there is currently debate as to the rationale of imposing an arbitrary dichotomous definition of 'normal' and 'abnormal' on gestational glucose tolerance. Indeed, empirical evidence accumulating over the last 20 years supports recognising an intermediate category of abnormal gestational glucose tolerance in women that falls in between the currently accepted 'normal' and 'abnormal' glucose thresholds. This intermediate category of glucose intolerance is also associated with considerable increases in risk of fetal macrosomia (27.5% vs 9.9% for intermediate vs normal, respectively), pre-eclampsia, caesarean-section delivery (40.0% vs 19.9% for intermediate vs normal, respectively), preterm delivery, neonatal jaundice and extended maternal and infant hospital length of stay.^{18–23}

Together, the discussion above explains the need for a simpler, easier-to-use, cost-effective, sensitive and specific biomarker for detection of GDM.²⁴ Prior evidence supports a strong link between the complement system, activity of the complement regulatory protein CD59 and the pathogenesis of vascular complications of diabetes.^{25 26} Complement activation ultimately leads to formation of the pore-forming membrane attack complex (MAC), the main effector of complement-mediated tissue damage.

CD59, a key complement regulator that inhibits MAC formation,²⁷ is universally expressed in mammalian cells. Glycation, the non-enzymatic attachment of glucose to protein amino groups, forms glycated CD59 (GCD59) and abrogates its activity as an inhibitor of MAC formation.^{28 29} Phospholipases shed both CD59 and GCD59 off cell membranes and release them into the circulation in soluble forms that can be measured in blood and other bodily fluids.

We have developed an assay to measure blood levels of GCD59 and demonstrated that these levels accurately distinguish individuals with and without diabetes (HbA1c >6.5% or <6%, respectively).³⁰ Furthermore, we previously reported that plasma GCD59 (pGCD59) levels at 24–28 weeks gestation identified GDM with high specificity and sensitivity.³¹ Also, higher maternal pGCD59 levels were associated with a higher prevalence of LGA even after adjustment for confounders such as body mass index (BMI), maternal and gestational age, race/ethnicity and history of diabetes.³¹ The prevalence of LGA newborns was 4.3% in the lowest quartile and 13.5% in the highest quartile of pGCD59. Recognising the importance of early detection of GDM, we also assessed the utility of pGCD59 as an early predictor (<20 weeks gestation) of GDM in 693 women with obesity from the Vitamin D and Lifestyle Intervention study.^{32 33} We found that pGCD59 successfully identified early GDM with an area under the receiver operating characteristic curve of 0.86. In this study, a one-unit increase in maternal pGCD59 concentration was associated with 36% increased odds of delivering an LGA infant.³³

The assay we have developed uses two specific monoclonal antibodies in a sandwich ELISA³¹; it is highly reproducible and measures serum and/or pGCD59 in a highly specific and sensitive manner. Indeed, assuming a basal level of CD59 glycation of ~5% (based on the basal glycation level of HbA1c in normoglycaemic individuals), and that the concentration of CD59 in human serum is ~100–150 ng/mL,³⁴ the detection of GCD59 in individuals without diabetes implies that the assay can detect GCD59 in the low picomolar range.

We report here the protocol of a human study designed to assess the accuracy of pGCD59 as an early, first trimester (<14 weeks gestation) predictor of GDM. Secondary objectives of this study are (1) to evaluate the potential association of maternal pGCD59 levels with adverse maternal and neonatal outcomes usually associated with GDM and (2) to assess the association of pGCD59 with the prevalence of gestational impaired glucose tolerance (GIGT), a form of glucose intolerance that does not meet American College of Obstetricians and Gynecologists criteria for GDM.

METHODS AND ANALYSIS

Human subjects, study design and sample acquisition

This is a prospective cohort study of pregnant women and their infants who receive prenatal care and deliver

Box 1 Eligibility criteria for plasma-glycated CD59 study
Eligibility criteria

- ▶ Consented to participate in Crimson Biobank.
- ▶ Discarded clinical plasma sample at <14 weeks gestation.
- ▶ ≥18 and ≤40 years old.

at Brigham and Women's Hospital (BWH) in Boston, Massachusetts. Patients who have consented to make their excess samples and associated clinical data available to researchers are included in the Partner's Crimson Biobank. We will obtain excess samples from the biobank based on study-specific criteria. The clinical data will be

extracted from the electronic medical records (EMRs) of patients who have consented to make their deidentified samples available to researchers.

Sample selection

Eligible plasma samples from donors meeting the criteria displayed in **box 1** will be separated by the Crimson Biobank from tubes routinely used to measure HbA1c at the first prenatal visit.

Sample size calculation

We estimate that including 500 women diagnosed with GDM at 24–28 weeks gestation, diagnosed based on current standard of care (C&C criteria), will provide

Table 1 Maternal and infant data obtained or derived from EPIC and the Clarity report

Data category/time point	Data points collected	Data points derived
Demographics	Race, ethnicity, insurance, OB practice.	–
Obstetrics and medical history	Gravidity, parity, medical history (ICD10 codes, active/resolved), infertility diagnosis (ICD10 codes).	–
Prenatal visits (three time points: first (V1), second (V2), third (V3) trimester)	Date of visit, age (V1 only), height (V1 only), weight, GA, blood pressure, medications.	BMI
Prenatal laboratory studies (three time points: first, second, third trimester)	Date of laboratory studies, GA at laboratory studies, WBCs, haematocrit, platelets, neutrophils, lymphocytes, haemoglobin A1c.	–
Prenatal ultrasounds (three time points: first, second, third trimester)	Date of ultrasound, fetal measurements (EFW, femur length, abdominal circumference, biparietal diameter, head circumference, weight percentile).	IUGR
Glucose load test	Date of GLT, GLT result.	GDM (C&C criteria)
Glucose tolerance test	Date of GTT, GTT results (fasting and 1, 2 and 3 hours).	GDM, GIGT (C&C criteria)
Additional prenatal laboratory studies	Results and dates of 24-hour protein, random urine protein, random protein, random creatinine, TSH.	–
Pregnancy complications	Clinical diagnosis (ICD10 codes) of GDM, pregnancy-induced hypertension, pre-eclampsia, pre-eclampsia with severe features, HELLP syndrome, UTIs, chorioamnionitis, URIs, STDs, neural tube defects, congenital heart defects, abdominal wall defects.	–
Delivery	Date and time of delivery, maternal weight, MOD, GA, infant MRN, sex, GBS, antibiotics, Apgar scores (1 and 5 min), infant measurements at birth (birth weight, length and head circumference), infant weight at discharge.	Multiple gestation, gestational weight gain (V1 to delivery), ponderal index, macrosomia, LGA, SGA
Infant feeding	Maternal feeding intention, date and time of first feed, breast milk at first feed, feeding type at discharge.	–
Infant clinical data	Diagnoses (ICD10 codes) of LGA, SGA, fetal malformations, shoulder dystocia/birth injury, hyperbilirubinaemia, NH and NICU admission; NICU length of stay, BG values, dextrose containing IVs, O2 devices, bilirubin, phototherapy, haematocrit.	Lowest BG, BG <45 mg/dL
Postpartum maternal visit	Date of visit, weight, blood pressure, GTT results, infant feeding method.	Postpartum weight loss

BG, blood glucose; BMI, body mass index; C&C, Carpenter & Coustan; EFW, estimated fetal weight; GA, gestational age; GBS, group B streptococcus; GDM, gestational diabetes mellitus; GIGT, gestational impaired glucose tolerance; GLT, glucose load test; GTT, glucose tolerance test; ICD, International Classification of Diseases; IUGR, intrauterine growth restriction; LGA, large for gestational age; MOD, mode of delivery; MRN, medical record number; NH, neonatal hypoglycaemia; NICU, neonatal intensive care unit; OB, obstetrics; SGA, small for gestational age; STDs, sexually transmitted diseases; TSH, thyroid stimulating hormone; UTIs, urinary tract infections; WBCs, white blood cells.

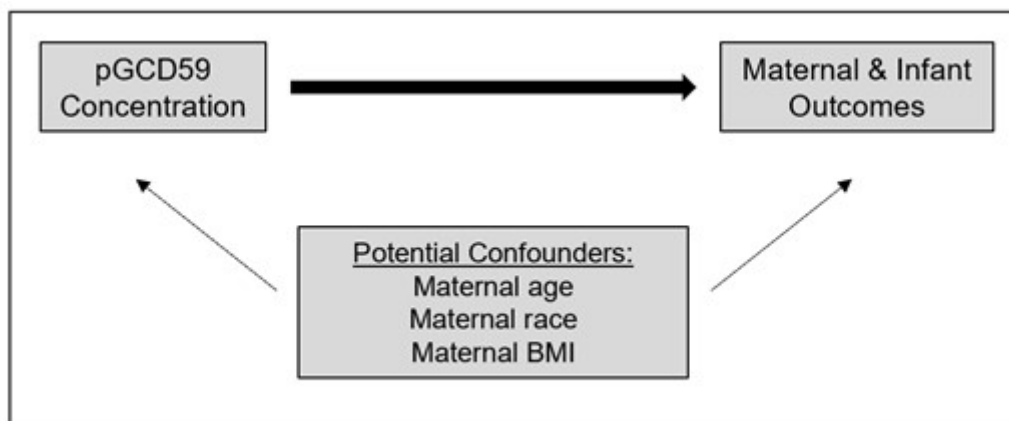


Figure 1 Directed acyclic graph of secondary objectives. BMI, body mass index; pGCD59, plasma-glycated CD59.

sufficient power to assess the accuracy of first trimester pGCD59 to identify women with GDM. In 2016, the BWH clinical laboratory received approximately 6000 plasma samples from women undergoing a GLT in the course of their prenatal care. Of these women, 1156 (19.6%) had abnormal GLT results and 380 (6.4%) were subsequently diagnosed with GDM using C&C criteria.³⁵ Based on these data, we conservatively estimate that to include 500 women with GDM we will need to measure first trimester pGCD59 in ~8000 women, given a prevalence of 5% of GDM in the BWH population.

Sample and data collection

After enrollment and collection of the first trimester plasma sample, a follow-up plasma sample will be collected for each subject at pregnancy weeks 24–28, when the GLT is performed. Both of these plasma samples (approximately 0.5 mL) will be frozen (-80°) and stored by the Crimson Biobank until retrieval by the study investigators.

Two months post delivery, maternal and infant clinical data will be downloaded from the subject's medical record and both discarded plasma samples will be retrieved from the Crimson Biobank for measurement of pGCD59. After retrieving the samples, we will break all links to the subject's medical record and, thereafter, identify samples and data only through their code number without any access to the subjects' personal information.

Patient and public involvement statement

Patients and the public were not involved in the design or the recruitment and conduct of this study, and will not be involved in the plans to disseminate the study results.

CLARITY REPORT

To efficiently download specified clinical information on both mother and infant from the EPIC medical record database, we have created a study-specific abstraction tool (Clarity report). Clarity is an EPIC native relational database warehouse made available to provide a platform for data analysis and volume data reporting.^{32 36 37} Most data

from the EPIC real-time clinical database are downloaded to Clarity daily.

The Clarity system uses a variety of tools and algorithms to extract data attributes from the EMR. Examples of these methods and algorithms include:

- ▶ Extraction of problem lists and diagnoses using International Classification of Diseases categorisations.
- ▶ Extraction of problem lists using natural language processing.
- ▶ Extraction of data points from encounters across all Mass General Brigham health facilities (including both ambulatory and inpatient settings).
- ▶ Extraction of medications orders, imaging results and laboratory test results using dynamic date ranges based on prenatal/postpartum visits.
- ▶ Extraction of clinical data from different levels of the patient's medical record including flowsheets, health maintenance encounters and procedure descriptions.
- ▶ Extraction of neonatal intensive care unit (NICU)-related activities by examining health level seven events.

A total of 213 variables will be obtained from the EMRs of the mothers and their infants. [Table 1](#) presents a summary of the types of variables we will download from the EMR.

We have also created a Research Electronic Data Capture (REDCap) data import tool which we manually populate with the Clarity report. The data will be checked for accuracy, cleaned and imported into REDCap for storage or statistical analysis.

During the initial development and testing of the Clarity report we conducted data accuracy audits by reviewing pilot subjects' EMRs. Similarly, we will conduct regular data audits for each set of extracted data prior to breaking the link to the participant's personal identifiers.

The index test is levels of pGCD59 measured with the highly specific and sensitive ELISA assay described previously.³¹ The reference tests are the 50-g GLT and the 100-g 3-hour OGTT, respectively, administered as standard prenatal care to screen for and diagnose GDM.

Outcomes

The primary outcome is diagnosis of GDM at pregnancy weeks 24–28, extracted from the maternal medical record and adjudicated based on C&C criteria. A subject is classified as having GDM if she has at least two values at or above the following thresholds plasma glucose levels: fasting: ≥ 95 mg/dL, 1-hour 180 mg/dL, 2-hour 155 mg/dL and 3-hour 140 mg/dL.⁸ Subjects are also classified as having GDM if their 1-hour GLT result is ≥ 190 mg/dL.⁸

Secondary outcomes include glucose intolerance (defined as one abnormal OGTT result) and adverse maternal and neonatal outcomes associated with GDM. Secondary maternal outcomes include a diagnosis of pre-eclampsia and caesarean section. Secondary neonatal outcomes include low birth weight as determined by the Ponderal Index ($PI = \text{fetal wt (g)} \times (100/\text{fetal length (cm)})^3$), macrosomia (birth weight > 4000 g), LGA (based on the Fenton curve,³⁸ fetal malformations, including neural tube, congenital heart and abdominal wall defects, shoulder dystocia/birth injury, small for gestational age (SGA), neonatal hypoglycaemia, hyperbilirubinaemia and NICU admission.

Data analysis plan

The initial statistical analyses will involve generating a detailed description of the maternal and infant characteristics for the total cohort by GDM and GIGT status. Counts and percentages will be reported for categorical variables. For continuous variables, histograms will be generated to determine data distributions and identify potential outliers. Depending on data distribution we will report either means and SD or medians and IQRs.

Receiver operating characteristic curves will be used to evaluate the performance of pGCD59 in the first trimester to predict a diagnosis of GDM or GIGT. We will compute the sensitivity, specificity, percentage of correctly classified, and both positive and negative likelihood ratios for each of the percentile cut points of pGCD59. The area under the curve will be computed using the trapezoidal rule.

The associations between levels of pGCD59 and adverse maternal and infant outcomes will be modelled using linear or logistic regression, depending on the outcome, with adjustment for relevant confounders and covariates (figure 1). Linear regression will be used to determine the associations between pGCD59, birth weight and the PI. Additionally, logistic regression will be used to determine the associations between pGCD59 and pre-eclampsia, caesarean section, macrosomia, LGA/SGA, fetal malformations, intrauterine growth restriction, shoulder dystocia/birth injury, neonatal hypoglycaemia, hyperbilirubinaemia and NICU admission. Some potential confounders that will be considered are maternal age, race, BMI and gestational weight gain.

Missing data will be handled by assessing the pattern of missing and using multiple imputation if data are missing at random. The level for significance for all analyses will be set at $p < 0.05$.

ETHICS AND DISSEMINATION

Ethical and safety concerns

This study has been approved by the Mass General Brigham Institutional Review Board (protocol 2011P002254). This study poses minimal to no risks to subjects. All subjects have signed consent forms allowing research use of samples taken in the course of their care through the Crimson Biobank.

Dissemination plan

Recruitment for this study started in October 2019 and is expected to continue for four additional years. The results of this study will be presented at international meetings in the fields of obstetrics, paediatrics and endocrinology and disseminated in relevant peer-reviewed journals.

Contributors The authors' responsibilities were as follows JH, SS and HA were responsible for the overall design and planning of the study; CA and MT-C were responsible for writing the manuscript with feedback from M-ALF, SKW, SS and JH; SS and JH: had primary responsibility for the final content; and all authors: reviewed the manuscript for accuracy and read and approved the final manuscript.

Funding This work was supported by NIH/NIDDK grant number R01 DK118528.

Competing interests JH is a founder and has financial interest in Mellitus LLC. Mellitus had licensed related IP and has interests in developing diagnostic tools for diabetes. JH's interests were reviewed and are managed by Brigham and Women's Hospital and Brigham Health in accordance with their conflict of interest policies. All other authors declare no competing financial interests in relation to the work described.

Patient and public involvement Patients and/or the public were not involved in the design, or conduct, or reporting, or dissemination plans of this research.

Patient consent for publication Not applicable.

Provenance and peer review Not commissioned; externally peer reviewed.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>.

ORCID iD

Chloe Andrews <http://orcid.org/0000-0002-4282-8215>

REFERENCES

- American Diabetes Association. 2. classification and diagnosis of diabetes. *Diabetes Care* 2015;38:S8–16.
- McIntyre HD, Catalano P, Zhang C, *et al*. Gestational diabetes mellitus. *Nat Rev Dis Primers* 2019;5:47.
- McIntyre HD, Sacks DA, Barbour LA, *et al*. Issues with the diagnosis and classification of hyperglycemia in early pregnancy. *Diabetes Care* 2016;39:53–4.
- HAPO Study Cooperative Research Group, Metzger BE, Lowe LP, *et al*. Hyperglycemia and adverse pregnancy outcomes. *N Engl J Med* 2008;358:1991–2002.
- Catalano PM, Vargo KM, Bernstein IM, *et al*. Incidence and risk factors associated with abnormal postpartum glucose tolerance in women with gestational diabetes. *Am J Obstet Gynecol* 1991;165:914–9.
- Durnwald C, Huston-Presley L, Amini S, *et al*. Evaluation of body composition of large-for-gestational-age infants of women with gestational diabetes mellitus compared with women with normal glucose tolerance levels. *Am J Obstet Gynecol* 2004;191:804–8.
- Metzger BE. Long-term outcomes in mothers diagnosed with gestational diabetes mellitus and their offspring. *Clin Obstet Gynecol* 2007;50:972–9.
- Committee on Practice B-O. ACOG practice Bulletin No. 190: gestational diabetes mellitus. *Obstet Gynecol* 2018;131:e49–64.

- 9 Aiken CEM, Hone L, Murphy HR, *et al.* Improving outcomes in gestational diabetes: does gestational weight gain matter? *Diabet Med* 2019;36:167–76.
- 10 Zera CA, Seely EW. Diabetes: treatment of gestational diabetes reduces obstetric morbidity. *Nat Rev Endocrinol* 2010;6:69–70.
- 11 Cousins L, Dattel BJ, Hollingsworth DR, *et al.* Glycosylated hemoglobin as a screening test for carbohydrate intolerance in pregnancy. *Am J Obstet Gynecol* 1984;150:455–60.
- 12 Hanna FWF, Peters JR. Screening for gestational diabetes; past, present and future. *Diabet Med* 2002;19:351–8.
- 13 Coustan DR. Diabetes mellitus in pregnancy: screening and diagnosis. UpToDate. Waltham, MA 2016.
- 14 Cefalu WT, Prather KL, Chester DL, *et al.* Total serum glycosylated proteins in detection and monitoring of gestational diabetes. *Diabetes Care* 1990;13:872–5.
- 15 Nasrat HA, Ajabnoor MA, Ardawi MS. Fructosamine as a screening-test for gestational diabetes mellitus: a reappraisal. *Int J Gynaecol Obstet* 1991;34:27–33.
- 16 ACOG practice Bulletin No. 190 summary: gestational diabetes mellitus. *Obstet Gynecol* 2018;131:406–8.
- 17 Agarwal MM, Punnose J, Dhatt GS. Gestational diabetes: problems associated with the oral glucose tolerance test. *Diabetes Res Clin Pract* 2004;63:73–4.
- 18 Tallarigo L, Giampietro O, Penno G, *et al.* Relation of glucose tolerance to complications of pregnancy in nondiabetic women. *N Engl J Med* 1986;315:989–92.
- 19 Langer O, Brustman L, Anyaegbunam A, *et al.* The significance of one abnormal glucose tolerance test value on adverse outcome in pregnancy. *Am J Obstet Gynecol* 1987;157:758–63.
- 20 Sermer M, Naylor CD, Gare DJ, *et al.* Impact of increasing carbohydrate intolerance on maternal-fetal outcomes in 3637 women without gestational diabetes. The Toronto Tri-Hospital gestational diabetes project. *Am J Obstet Gynecol* 1995;173:146–56.
- 21 Forsbach G, Cantú-Díaz C, Vázquez-Lara J, *et al.* Gestational diabetes mellitus and glucose intolerance in a Mexican population. *Int J Gynaecol Obstet* 1997;59:229–32.
- 22 Aberg A, Rydhstroem H, Frid A. Impaired glucose tolerance associated with adverse pregnancy outcome: a population-based study in southern Sweden. *Am J Obstet Gynecol* 2001;184:77–83.
- 23 Jensen DM, Damm P, Sørensen B, *et al.* Clinical impact of mild carbohydrate intolerance in pregnancy: a study of 2904 nondiabetic Danish women with risk factors for gestational diabetes mellitus. *Am J Obstet Gynecol* 2001;185:413–9.
- 24 Bogdanet D, Reddin C, Murphy D, *et al.* Emerging protein biomarkers for the diagnosis or prediction of gestational Diabetes—A scoping review. *J Clin Med* 2021;10:jcm10071533 doi:10.3390/jcm10071533
- 25 Qin X, Goldfine A, Krumrei N, *et al.* Glycation inactivation of the complement regulatory protein CD59: a possible role in the pathogenesis of the vascular complications of human diabetes. *Diabetes* 2004;53:2653–61.
- 26 Ghosh P, Sahoo R, Vaidya A, *et al.* Role of complement and complement regulatory proteins in the complications of diabetes. *Endocr Rev* 2015;36:272–88.
- 27 Meri S, Morgan BP, Davies A, *et al.* Human protectin (CD59), an 18,000–20,000 MW complement lysis restricting factor, inhibits C5b-8 catalysed insertion of C9 into lipid bilayers. *Immunology* 1990;71:1–9.
- 28 Acosta J, Hettinga J, Flückiger R, *et al.* Molecular basis for a link between complement and the vascular complications of diabetes. *Proc Natl Acad Sci U S A* 2000;97:5450–5.
- 29 Acosta J, Qin X, Halperin J. Complement and complement regulatory proteins as potential molecular targets for vascular diseases. *Curr Pharm Des* 2004;10:203–11.
- 30 Ghosh P, Sahoo R, Vaidya A, *et al.* A specific and sensitive assay for blood levels of glycosylated CD59: a novel biomarker for diabetes. *Am J Hematol* 2013;88:670–6.
- 31 Ghosh P, Luque-Fernandez MA, Vaidya A, *et al.* Plasma glycosylated CD59, a novel biomarker for detection of pregnancy-induced glucose intolerance. *Diabetes Care* 2017;40:981–4.
- 32 Jelsma JGM, van Poppel MNM, Galjaard S, *et al.* DALI: Vitamin D and lifestyle intervention for gestational diabetes mellitus (GDM) prevention: an European multicentre, randomised trial - study protocol. *BMC Pregnancy Childbirth* 2013;13:142.
- 33 Ma D, Luque-Fernandez MA, Bogdanet D, *et al.* Plasma glycosylated CD59 predicts early gestational diabetes and large for gestational age newborns. *J Clin Endocrinol Metab* 2020;105:dga087:e1033–40. doi:10.1210/clinem/dgaa087
- 34 Landi APG, Wilson AB, Davies A, *et al.* Determination of CD59 protein in normal human serum by enzyme immunoassay, using octyl-glucoside detergent to release glycosyl-phosphatidylinositol-CD59 from lipid complex. *Immunol Lett* 2003;90:209–13.
- 35 Carpenter MW, Coustan DR. Criteria for screening tests for gestational diabetes. *Am J Obstet Gynecol* 1982;144:768–73.
- 36 Turner D, Monthé-Drèze C, Cherkerzian S, *et al.* Maternal obesity and cesarean section delivery: additional risk factors for neonatal hypoglycemia? *J Perinatol* 2019;39:1057–64.
- 37 Sen S, Andrews C, Anderson E, *et al.* Type of feeding provided with dextrose gel impacts hypoglycemia outcomes: comparing donor milk, formula, and breastfeeding. *J Perinatol* 2020;40:1705–11.
- 38 Fenton TR, Nasser R, Eliasziw M, *et al.* Validating the weight gain of preterm infants between the reference growth curve of the fetus and the term infant. *BMC Pediatr* 2013;13:92.