

## Compartmental Pharmacokinetics of the Antifungal Echinocandin Caspofungin (MK-0991) in Rabbits

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**The pharmacokinetics of the antifungal echinocandin-lipopeptide caspofungin (MK-0991) in plasma were studied in groups of three healthy rabbits after single and multiple daily intravenous administration of doses of 1, 3, and 6 mg/kg of body weight. Concentrations were measured by a validated high-performance liquid chromatography method and fitted into a three-compartment open pharmacokinetic model. Across the investigated dosage range, caspofungin displayed dose-independent pharmacokinetics. Following administration over 7 days, the mean peak concentration in plasma ( $C_{max}$ )  $\pm$  standard error of the mean increased from  $16.01 \pm 0.61$   $\mu\text{g/ml}$  at the 1-mg/kg dose to  $105.52 \pm 8.92$   $\mu\text{g/ml}$  at the 6-mg/kg dose; the mean area under the curve from 0 h to infinity rose from  $13.15 \pm 2.37$  to  $158.43 \pm 15.58$   $\mu\text{g} \cdot \text{h/ml}$ , respectively. The mean apparent volume of distribution at steady state ( $V_{d,ss}$ ) was  $0.299 \pm 0.011$  liter/kg at the 1-mg/kg dose and  $0.351 \pm 0.016$  liter/kg at the 6-mg/kg dose (not significant [NS]). Clearance (CL) ranged from  $0.086 \pm 0.017$  liter/kg/h at the 1-mg/kg dose to  $0.043 \pm 0.004$  liter/kg/h at the 6-mg/kg dose (NS), and the mean terminal half-life was between 30 and 34 h (NS). Except for a trend towards an increased  $V_{d,ss}$ , there were no significant differences in pharmacokinetic parameters in comparison to those after single-dose administration. Caspofungin was well tolerated, displayed linear pharmacokinetics that fit into a three-compartment pharmacokinetic model, and achieved sustained concentrations in plasma that were multiple times in excess of reported MICs for susceptible opportunistic fungi.**

Caspofungin (MK-0991) is a novel, investigational parenteral antifungal agent that belongs to a new generation of semisynthetic cyclic lipopeptides of the echinocandin family. It acts by noncompetitive inhibition of the synthesis of 1, 3- $\beta$ -D-glucan, an essential homopolysaccharide in the cell wall of many pathogenic fungi (11, 13). Similar to other current investigational echinocandin derivatives, caspofungin has potent and fungicidal in vitro activity against most clinically relevant *Candida* species without cross-resistance to currently approved antifungal agents, and it has cell-wall damaging effects on several *Aspergillus* species (3, 6, 8, 9, 15, 16, 20). The drug has demonstrated very promising activity in infection models of oropharyngeal (A. M. Flattery, G. K. Abruzzo, J. G. Smith, C. J. Gill, H. Rosen, H. Kropp, and K. Bartizal, Abstr. 36th Intersci. Conf. Antimicrob. Agents Chemother., abstr. F-40, 1996) and disseminated (1, 10; K. Bartizal, J. G. Smith, C. J. Gill, A. M. Flattery, L. Kong, C. Leighton, J. Stone, D. Cylc, A. Yuan, and G. K. Abruzzo, Abstr. 36th Intersci. Conf. Antimicrob. Agents Chemother., abstr. F-80, 1997) candidiasis and significantly prolonged the survival in mouse models of disseminated (1; K. Bartizal et al., Abstr. 36th Intersci. Conf. Antimicrob. Agents Chemother.) and pulmonary (E. M. Bernard, T. Ishimaru, and D. Armstrong, Abstr. 36th Intersci. Conf. Antimicrob. Agents Chemother., abstr. F-39, 1996) as-

pergillosis, both in healthy and immunocompromised animals, respectively. In addition to its antifungal activity, caspofungin was also effective as a preventive and therapeutic modality against *Pneumocystis carinii* pneumonia in dexamethasone-immunocompromised rodents (19). Little is known, however, about the pharmacokinetics of caspofungin. The purpose of this study was to characterize the pharmacokinetics of caspofungin in plasma in the rabbit, to compare them with those of other echinocandins, and to provide the basis for the investigation of the concentration-response relationships of caspofungin in experimental rabbit models of invasive fungal infections.

**Study drug.** Caspofungin (MK-0991; Merck & Co., Rahway, N.J.) was provided as a lyophilized powder and dissolved according to the recommendations of the manufacturer in sterile water to produce a 50-mg/ml stock solution that was maintained at  $-70^\circ\text{C}$ . Prior to use, the drug was freshly diluted with sterile water to 10-, 5-, and 2-mg/ml solutions for the 6-, 3-, and 1 mg/kg dosage group, respectively. The reconstituted drug was administered at ambient temperature as a slow intravenous bolus over 1 min through the indwelling catheter.

**Animals.** Healthy female New Zealand White rabbits (Hazelton, Denver, Pa.) weighing 2.5 to 3.5 kg were used in all experiments. They were individually housed and maintained with water and standard rabbit feed ad libitum according to National Institutes of Health Guidelines for Laboratory Animal Care (4). Vascular access was established in each rabbit  $\geq 72$  h prior to experimentation by the surgical placement of a subcutaneous silastic central venous catheter as previously described (21).

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**Single-dose studies.** Three groups of three rabbits were studied by single-dose studies. Animals received caspofungin at either 1, 3, or 6 mg/kg of body weight as a single steady intravenous bolus over 1 min. Plasma samples were drawn immediately before administration, immediately after administration (maximum concentration of drug in plasma [ $C_{max}$ ]), and then at 10 and 30 min and 1, 2, 4, 8, 12, 18, 24, 48, 72, and 96 h postdosing.

**Multiple-dose studies.** A different set of three groups of three rabbits were studied by multiple-dose studies. Animals received caspofungin at either 1, 3, or 6 mg/kg of body weight daily as an intravenous bolus over 1 min for a total of seven doses. Plasma samples were drawn immediately before administration of the seventh dose, immediately after administration ( $C_{max}$ ), and then at 10 and 30 min and 1, 2, 4, 8, 12, 18, 24, 48, 72, and 96 h postdosing. Hepatic and renal toxicities were monitored in plasma 24 h after the last drug dose and compared to normal values. All animals were clinically evaluated each day and weighed before the first dose and at the end of the study.

**Processing of blood samples.** Blood samples were collected in heparinized syringes. Plasma was immediately separated by centrifugation and stored at  $-80^{\circ}\text{C}$  until shipment in dry ice to the laboratories of Merck, Sharp & Dohme-Chibret, Riom, France, for assay.

**Analytical method.** Drug levels in plasma were determined after solid-phase extraction and dilution in mobile phase by reversed-phase high-performance liquid chromatography. The mobile phase consisted of acetonitrile–0.01 M  $\text{KH}_2\text{PO}_4$  (60:40, vol/vol), pH 3. Separation was achieved using a Brownlee Cyano column (220 by 4.6 mm [inner diameter]; particle size, 5  $\mu\text{m}$ ; Perkin-Elmer, Norwalk, Conn.). Caspofungin was detected by fluorometric detection (excitation at 224 nm; emission at, 302 nm).

Quantitation was based on an internal standard method using the semisynthetic echinocandin L-733,560 as the internal standard. Eight-point standard curves (range of concentrations: 0.15 to 10  $\mu\text{g}/\text{ml}$ ) were linear with  $r^2$  values greater than 0.998. The lower limit of quantification (LLQ) was 0.15  $\mu\text{g}/\text{ml}$ . Accuracies were within  $\pm 15\%$  and intra- and interday variability (precision) was  $<5\%$ .

**Pharmacokinetic data analysis.** Pharmacokinetic parameters for caspofungin were determined using compartmental analysis. Experimental plasma concentration-time data were fitted to a three-compartment open model with intravenous bolus input and linear first-order elimination from the central compartment using iterative weighted nonlinear least-squares regression in the Adapt II (5) computer program. Model selection was guided by Akaike's information criterion (23). The model fit the data well, with  $r^2$  values for the individual fits ranging from 0.974 to 0.999 (mean, 0.991). The regression lines through the plot of observed versus estimated concentrations did not differ from the line of identity, and no bias was observed.  $C_{max}$  values were determined as model-estimated concentrations immediately after bolus administration, and  $\text{AUC}_{0-\infty}$  values were calculated from estimated plasma concentration profiles using the trapezoidal rule and extrapolation to infinity by standard techniques. Dose linearity after single and after multiple dosing was determined by comparison of the dose-normalized area under the curve from 0 h to infin-

ity ( $\text{AUC}_{0-\infty}$ ) across dosage levels by analysis of variance (ANOVA) and linear regression analysis. Accumulation was assessed for each dosage level by comparing the mean AUC between doses after multiple dosing as an approximation of AUC between doses at steady state with the mean  $\text{AUC}_{0-\infty}$  after single dosing.

**Statistical analysis.** Differences between the means of pharmacokinetic parameters across dosage levels were evaluated by ANOVA with Bonferroni's correction for multiple comparisons. Student's or Welch's  $t$  test was used in addition for comparison of pharmacokinetic parameters after single dosing with those after multiple dosing. A two-tailed  $P$  value of  $<0.05$  was considered statistically significant.

**Pharmacokinetics in plasma.** The estimated plasma concentration-versus-time profiles of caspofungin are shown in Fig. 1, and the corresponding mean compartmental pharmacokinetic parameters are listed in Table 1. Administration of caspofungin at single doses of 1, 3, and 6 mg/kg resulted in escalating peak levels in plasma that ranged from  $20.02 \pm 1.18$  to  $123.4 \pm 5.17$   $\mu\text{g}/\text{ml}$  (means  $\pm$  standard errors of the means). The drug exhibited a rapid initial distribution phase, followed by a second, somewhat slower distribution-elimination phase, and a prolonged elimination phase with a mean terminal half-life ranging from 26 to 31 h. Mean plasma levels fell below LLQ in a dose-dependent manner after 8, 12, and 18 h postdosing. Caspofungin demonstrated linear pharmacokinetics in plasma with no changes in dose-normalized  $\text{AUC}_{0-\infty}$  or total clearance (CL) across the investigated dosage range. The apparent volume of distribution at steady state ( $V_{ss}$ ) was comparatively small and independent of the dosage.

After multiple dosing over 7 days, peak concentrations in plasma were not significantly different from those observed after administration of a single dose. Mean levels in plasma fell below the LLQ in a dose-dependent manner after 8, 24, and 48 h. Levels in plasma at the end of the dosing interval were below the LLQ in all rabbits receiving the 1-mg/kg dose and in two of three rabbits receiving the 3-mg/kg dose, respectively. There were no significant differences in AUC, CL, and half-life compared to those after single dosing. The  $V_{ss}$ , however, showed a trend towards larger values after multiple dosing ( $P < 0.05$ ,  $P = 0.16$ , and  $P < 0.05$  for the 1-, 3-, and 6-mg/kg dosages, respectively, by  $t$  test only). No differences in dose-normalized  $\text{AUC}_{0-\infty}$  across the investigated dosage range were noted by ANOVA and linear regression, indicating dose-independent, linear pharmacokinetics of the compound also after multiple dosing.

Blood urea nitrogen, serum creatinine, bilirubin, and alanine aminotransferase levels determined after 7 days of treatment with caspofungin were within the range of normal values determined in 24 healthy, drug-naive animals. Infusion-related toxicity or other clinical abnormalities, including abnormal weight changes, were not observed.

The results of this study demonstrate linear pharmacokinetics of caspofungin over the investigated dosage range of 1 to 6 mg/kg/day, with dose-proportional increases in the  $\text{AUC}_{0-\infty}$  with increasing dosage. Plasma concentration data fitted well into a three-compartment open pharmacokinetic model that revealed a prolonged terminal elimination half-life in the range of 30 to 35 h. There were no significant differences in pharmacokinetic parameters between single-dose and multiple-dose

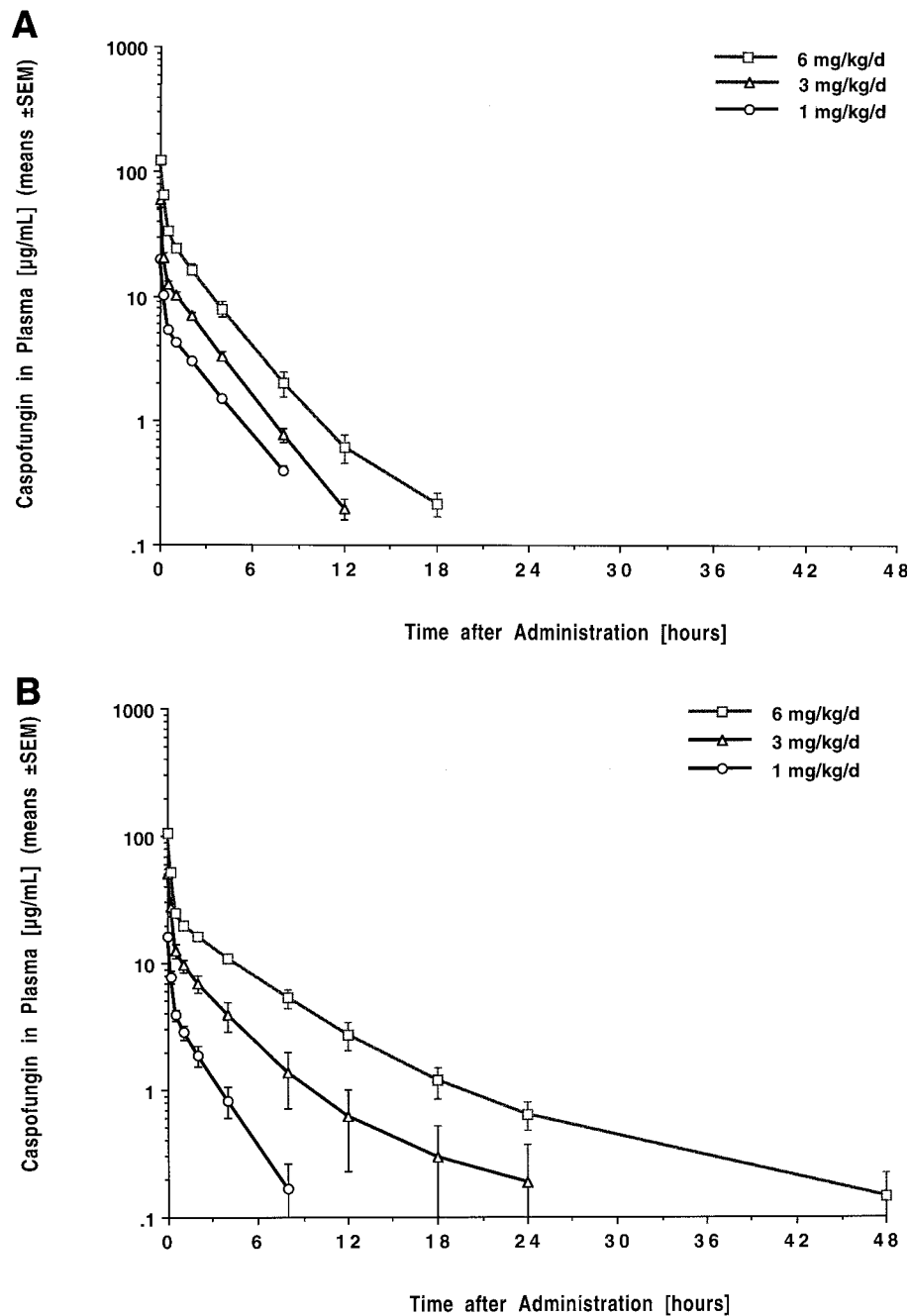


FIG. 1. Concentration-versus-time plots after single dosing (A) and after multiple daily dosing over seven days (B) with 1, 3, and 6 mg of caspofungin per kg, respectively. Each point plots the mean concentration  $\pm$  standard error of the mean (error bars) for three rabbits each at that time.

administration except for a trend towards an increased  $V$  after multiple dosing. The investigated dosages achieved sustained concentrations in plasma that were multiple times in excess of MICs reported for susceptible opportunistic fungi (8, 15). Caspofungin was well tolerated without evidence of renal or hepatic toxicities.

The favorable pharmacokinetic profile of caspofungin stands in marked contrast to that of cilofungin, the first echinocandin derivative that had entered clinical development. The pharmacokinetics of this drug in the rabbit were characterized by a very rapid elimination from the bloodstream via first-order

kinetics, and its antifungal efficacy *in vivo* was limited. Increased antifungal efficacy, particularly in the brain, could only be achieved through intermittent and continuous infusion of daily dosages of as much as 180 mg/kg that elicited nonlinear saturation kinetics (14, 22).

The pharmacokinetics of caspofungin in plasma in the rabbit appear somewhat different from those of the investigational echinocandin VER-002 (*v*-echinocandin; formerly LY303366) in the same species. In comparison to caspofungin, at similar dosages, VER-002 exhibited an approximately twofold faster clearance and twofold lower  $C_{\text{max}}$  and AUC values but a three-

TABLE 1. Estimated compartmental pharmacokinetic parameters of caspofungin in plasma<sup>a</sup>

Drug dose (mg/kg)	C <sub>max</sub> (μg/ml)	C <sub>min</sub> (24 h) (μg/ml)	AUC <sub>0-∞</sub> (μg/ml · h)	V <sub>ss</sub> (liter/kg)	CL (liter/h/kg)	t <sub>1/2α</sub> <sup>b</sup> (h)	t <sub>1/2β</sub> (h)	t <sub>1/2γ</sub> <sup>*</sup> (h)
Single dose								
1	20.02 ± 1.18	0.00 ± 0.00	20.57 ± 0.89	0.239 ± 0.011	0.050 ± 0.002	0.09 ± 0.00	2.00 ± 0.06	31.61 ± 0.12
3	60.47 ± 8.49	0.00 ± 0.00	47.14 ± 3.60	0.277 ± 0.033	0.068 ± 0.005	0.05 ± 0.00	1.84 ± 0.07	30.23 ± 1.19
6	123.40 ± 5.17	0.10 ± 0.00	119.30 ± 10.91	0.275 ± 0.006	0.053 ± 0.006	0.11 ± 0.01	1.85 ± 0.14	26.19 ± 1.29
Multiple dose								
1	16.01 ± 0.61	0.00 ± 0.00	13.15 ± 2.37	0.299 ± 0.011	0.086 ± 0.017	0.09 ± 0.01	1.62 ± 0.21	31.87 ± 0.18
3	51.05 ± 4.59	0.18 ± 0.18	63.70 ± 21.34	0.378 ± 0.050	0.065 ± 0.015	0.09 ± 0.01	2.20 ± 0.37	34.77 ± 3.09
6	105.52 ± 8.92	0.63 ± 0.26	158.43 ± 15.58	0.351 ± 0.016	0.043 ± 0.004	0.10 ± 0.00	3.53 ± 0.48	30.86 ± 0.96

<sup>a</sup> All values represent the means ± standard errors of the means of values three rabbits each. Abbreviations: C<sub>min</sub>(24 h), plasma concentration at the end of the dosing interval (24 h); t<sub>1/2α</sub>, distributional half-life; t<sub>1/2β</sub>, apparent elimination half-life; t<sub>1/2γ</sub>, terminal elimination half-life.

<sup>b</sup> P ≤ 0.05 for the comparison among dosage groups by ANOVA.

to fourfold larger V (18; A. H. Groll, D. Mickiene, V. Petraitis, R. Petraitiene, A. Field-Ridley, M. Candelario, J. Bacher, C. McMillian, S. C. Piscitelli, and T. J. Walsh, Abstr. 38th Intersci. Conf. Antimicrob. Agents Chemother., abstr. J-59, 1998.). It is yet unknown whether these subtle differences in the pharmacokinetics of both compounds also result in different pharmacodynamics.

Similar to cilofungin (22), VER-002 (17), and the original diamino analog L-733,560 (2), the in vitro fungicidal activity of caspofungin against *Candida* spp. appears to be concentration dependent (7). The implications of these observations remain to be investigated in vivo.

The low apparent V<sub>ss</sub> in the rabbit is reflective of the extensive protein binding of caspofungin across all species (12; J. A. Stone, S. D. Holland, W. D. Ju, Z. Zhang, M. Schwartz, V. L. Hoagland, K. E. Mazina, T. L. Hunt, and S. Waldman, Abstr. 38th Intersci. Conf. Antimicrob. Agents Chemother., abstr. C-51, 1998). Tissue distribution studies with radiolabeled drug in mice following intraperitoneal administration revealed preferential distribution to liver, kidney, and small intestine and a slower CL from all tissues than from plasma (12). The slow equilibration rates of most tissues is consistent with the existence of a terminal elimination phase beyond the dosing interval of 24 h and the increasing V<sub>ss</sub> after multiple dosing in our study and may be important for the dynamics of the drug against infections located in tissues. Indeed, infection models investigating cilofungin and VER-002 have demonstrated a correlation of antifungal efficacy with concentrations in tissue, and time of exposure was important for achieving effective concentrations in tissue and antifungal efficacy (22; A. H. Groll, D. Mickiene, V. Petraitis, R. Petraitiene, C. McMillian, S. Piscitelli, and T. J. Walsh, Abstr. 39th Intersci. Conf. Antimicrob. Agents Chemother., abstr. 2001, 1999].

While caspofungin does not interact in a significant manner with the cytochrome P450 enzyme system, it undergoes substantial hepatic metabolism and biliary excretion (11). The experimental work with cilofungin indicates saturable elimination pathways for the class of echinocandins, either by saturable biliary excretion of the parent or by metabolite inhibition (14, 22). These circumstances may also be relevant to caspofungin at higher dosages or in more narrow dosing regimens and should specifically be considered when the drug is projected to be given concurrently with other drugs that may compete with its biliary elimination mechanisms. Thus, the potential of drug-drug interactions with drugs such as cyclo-

sporine, amphotericin B, certain antineoplastic agents (anthracyclines, vinca alkaloids, cis-platinum, etoposide) and antibacterial agents (macrolides, metronidazole) needs to be carefully studied during preclinical development.

In conclusion, caspofungin displayed linear pharmacokinetics in plasma that fit into a three-compartment open pharmacokinetic model. No accumulation in plasma was observed after multiple dosing, and the compound was well tolerated without hepatic or renal laboratory toxicity. The characterization of the pharmacokinetics in the rabbit will aid in selecting appropriate dosing regimens in infection models and may be useful for the translation of the findings of these models into clinical studies.

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