Menadione: a platform and a target to valuable compounds synthesis

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Review

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Abstract

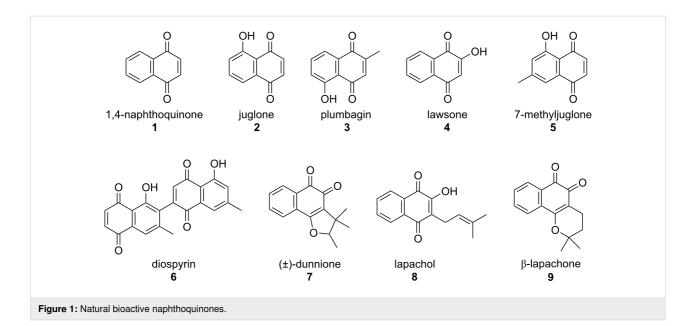
Naphthoquinones are important natural or synthetic compounds belonging to the general class of quinones. Many compounds in this class have become drugs that are on the pharmaceutical market for the treatment of various diseases. A special naphthoquinone derivative is menadione, a synthetic naphthoquinone belonging to the vitamin K group. This compound can be synthesized by different methods and it has a broad range of biological and synthetic applications, which will be highlighted in this review.

Introduction

Naphthoquinones belong to the chemical family of quinones and are widely present in synthetic and natural products (Figure 1). In nature, quinones are biosynthesized as secondary metabolites by various organisms, from simple single-celled microorganisms to more complex beings, such as higher plants and animals [1]. Actually, quinones play important roles in several physiological processes in these organisms, such as

photosynthesis [2] and oxidative phosphorylation [3,4], as well as many other metabolic processes [5-7]. Quinones also received considerable attention due to their importance in microbial systems, once several studies have shown that structural variations in microbial quinones have chemotaxonomic significance and can be used in the classification and identification of various microbial species [8]. With their particular and

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quite interesting chemical properties and bioactivities, naphthoquinones have aroused great interest, mainly in the pharmaceutical field, where they have been widely used in the development of new and more efficient drugs [1,9].

The naphthoquinone menadione has attracted a lot of attention. Menadione or 2-methyl-1,4-naphthoquinone (10), most known as vitamin K_3 , is a naphthoquinone derivative exclusively synthetic, not found in nature, used as an important precursor to synthesize vitamins K_1 and K_2 , being classified as a provitamin (Figure 2) [10]. Vitamins K, obtained through food, play an important role in maintaining animals' physiology, by acting on blood clotting and regulating bone calcification [10]. In animals, menadione can be converted in vitamin K_2 in the intestinal tract, by intestinal microbiota [10]. In humans, the menadione–vitamin K_2 conversion occurs after its alkylation in

the liver [11]. Moreover, in adult humans, vitamin K_1 can be converted into vitamin K_2 , a process that requires menadione as intermediate [12]. Menadione sodium bisulfite complex (MSB, 13) [13] and menadiol (vitamin K_4 , 14) [14], in turn, are two water-soluble derivatives converted in the body, to menadione. The MSB favors the formation of prothrombin and speeds up blood coagulation, improving its antihemorrhagic activity when compared to the natural vitamins K [13].

The action of menadione in live organisms is not restricted to its use as a biosynthetic precursor to vitamins K_1 and K_2 , a variety of studies has shown a wide range of biological activities of menadione, such as anticancer [15-22], antibacterial [23-26], antifungal [27,28], antimalarial [29-32], antichagasic [33], and anthelmintic [34] effects. In these cases, the redox cycle of menadione, followed by reactive oxygen species (ROS) genera-

tion, resulted from the interactions between nucleophilic biomolecules and 1,4-naphthoquinonic nucleus of menadione and its derivatives.

The presence of an α,β -unsaturated diketone in the quinone structures allows them to accept electrons through reduction processes, followed by oxidation, thus establishing a redox cycle. The main characteristics of the quinones (Q) redox cycle, comprises the one-electron reduction to generate a semiquinone intermediate (SQ) and the two-electron reduction leading to hydroquinone (HQ), in NAD(P)H oxidase-dependent processes [35-38]. In the presence of oxygen, the reduced species is oxidized back to the quinone, thus completing the cycle. In case of naphthoquinones such as menadione, the quinone-semiquinone or quinone-hydroquinone interconversion generates reactive oxygen species (ROS), such as superoxide anion (O2°-), hydrogen peroxide (H2O2), hydroxyl radical (°OH), and hydroperoxyl radical (*OOH) (Figure 3) [35]. Additionally, the menadione semiquinone radical can participate in another redox cycle, such as, the Fenton reaction, also resulting in the production of hydroxyl and hydroperoxyl radicals (Figure 3) [39-41].

The quinone-hydroquinone and quinone-semiquinone interconversions, with ROS generation, are responsible for the wide range of biological activities of menadione and its derivatives. An excess of ROS in the intracellular environment can cause harmful effects on structures such as nucleic acids (DNA and

RNA), enzymes, structural proteins, and cell membrane lipids. These effects can lead to dysfunctions that activate the apoptosis process resulting in cell death [42-46].

In addition to participating in redox cycles as biomolecules in various biochemical processes, the 1,4-naphthoquinone structure of the menadione core is also easily recognized and incorporated by different live organisms, as is evidenced by its presence in the structure of several natural naphthoquinones. The ability to participate in biochemical processes by interacting with biomolecules through redox cycles makes menadione a very interesting structural model for the development of new bioactive compounds. Considering the great potential of menadione, mainly in biological applications, in this review several applications involving its synthesis and its use as a versatile synthetic platform will be discussed.

Review

Preparation methods of menadione

Among the most common methods for the preparation of menadione (10), we can find, for example, the oxidation of 2-methylnaphthalene or 2-methylnaphthol. Other less frequent but equally efficient approaches to synthesize menadione include the demethylation of 2-methyl-1,4-dimethoxynaphthalene, the construction of the naphthoquinone ring, the methylation of 1,4-naphthoquinone, and the electrochemical synthesis from 2-methyl-1,4-dihydroxynaphthalene. The works discussed in

Fenton reaction

OH

HQ 10 14
$$Fe^2+ + H_2O_2 \longrightarrow Fe^3+ + HO + HO$$
 (step 1)

 $Fe^{3+} + H_2O_2 \longrightarrow Fe^2+ + HOO + H^+$ (step 2)

 $2 H_2O_2 \longrightarrow HO + HOO + H_2O$ (net reaction)

NAD(P)H, H⁺

1 e⁻

OH

Fenton reaction

OH

Fersh H2O2 \longrightarrow Fe²⁺ Fe³⁺ OH

NAD(P)H, H⁺

1 e⁻

OH

Fersh Fe²⁺

OH

Fenton reaction

OH

Fersh H2O2 \longrightarrow Fe²⁺ Fe³⁺

OH

reactions

OH

Fersh Fe²⁺

OH

OH

SQ 10 15

this section are grouped according to the synthetic approach that was employed to prepare menadione.

Oxidation of 2-methylnaphthalene

Menadione synthesis through the oxidation of 2-methylnaphthalene (16) includes the use of oxygen-rich oxidants using various reaction conditions and a broad range of well-succeeding methodologies has been reported. A summary is presented in Table 1 and will be discussed in this section.

In a pioneering study, Fieser reported the use of chromium(IV) oxide in glacial acetic acid for the oxidation of 2-methylnaphthalene (16) and obtained menadione (10) in 38-42% yield (Table 1, entry 1) [47]. A similar process was developed by Li and Elliot, who used sodium dichromate as oxidizing agent in the presence of sulfuric acid, instead of acetic acid, to obtain compound 10 in 62% yield within a shorter reaction time (Table 1, entry 2) [48]. The methodology using sodium dichromate and sulfuric acid was adapted to the industrial scale production of vitamin K_3 (10). However, this process is not ecofriendly, once, in this reaction, 18 kg of inorganic salts were obtained as a byproduct per kg of product and it was necessary

to treat the wastewater containing chromium. In this context, alternative approaches with a broad range of catalysts and oxidizing agents were studied [63]. Yamazaki reported the use of 10 mol % of chromium(VI) oxide and orthoperiodic acid, as a terminal oxidant, to obtain menadione (10) in 61% yield (Table 1, entry 3) [49].

Alternatives to chromium(VI) compounds to oxidize 2-methyl-naphthalene (16) to menadione (10) have also been evaluated and one of the most studied and used oxidants has been H₂O₂. Yamaguchi and co-workers described the oxidation of 16 with aqueous H₂O₂ in the presence of a palladium(II)-polystyrene sulfonic acid resin (Table 1, entry 4) [50]. According to the authors, in the absence of the catalysts, the oxidation took place slowly with 7.8% yield, meanwhile, Pd-catalysis improved the yield to 50–60% under otherwise identical conditions [50]. The approach described by the Adam's group used H₂O₂ (85%) and methyltrioxorhenium(VII) (MTO) as the catalyst (Table 1, entry 5) [51,64]. Without the catalyst, the reaction yield was only 10%, but after the addition of MTO and acetic anhydride, the yield increased to 46% (Table 1, entry 5) [51]. The authors suggested two simultaneous reaction pathways: a direct oxidation

 Table 1: Different approaches of 2-methylnaphthalene oxidation to menadione.

Entry	Conditions	Catalyst	Yield (%)	Ref.
1	CrO ₃ , H ₂ O, AcOH, 85–90 °C, 1 h	-	38–42	[47]
2	Na ₂ Cr ₂ O ₇ ·H ₂ O, H ₂ SO ₄ , CCl ₄ , 80 to 0 °C, 15 min	_	48–62	[48]
3	H ₅ IO ₆ , CH ₃ CN, 5 °C, 1 h	CrO ₃	61	[49]
4	H ₂ O ₂ (60%), AcOH, 50 °C, 8 h	palladium(II)-resin	50-60	[50]
5	H ₂ O ₂ (85%), Ac ₂ O, AcOH, 40 °C, N ₂ , 4 h	CH ₃ ReO ₃	46	[51]
6	H ₂ O ₂ (30%), AcOH, 100 °C, 3 h	_	86	[52]
7	H ₂ O ₂ (0.2 M), AcOH, rt, 24 h	iron(III) salts	13	[53]
8	H ₂ O ₂ (30%), AcOH, 60 °C, 3 h	MnPc	60	[54]
9	H ₂ O ₂ (30%), TAA, rt, 1.5 h	FeCl ₃ ·H ₂ O/H ₂ Pydic/benzylamine	44	[55]
10	H ₂ O ₂ , AcOH, 100 °C, 6 h	SeMCM-41	99 ^a	[56]
11	H ₂ O ₂ (30%), AcOH, H ₂ SO ₄ , 60–80 °C, 1 h	[(DIPAPTES)PdCl ₂] or [SiO ₂ (DIPAPES)PdCl ₂]	52 or 59	[57]
12	H ₂ O ₂ (35%), CH ₃ CN, AcOH, reflux, 8 h	GO@CHONHRN(CH2PPh2)2PdCl2	99 ^a	[58]
13	H ₂ O ₂ (35%), CH ₃ CN, AcOH, H ₂ SO ₄ , reflux, 12 h	L1-iron(III)	79 ^a	[59]
14	H ₂ O ₂ (35%), CH ₃ CN, H ₂ O, 60 °C, 20 min	$H[Cu^{II}(ttb)(H_2O)_3]_2[Cu^{II}(ttb)CI]_2$ [PW ₁₂ O ₄₀]·4H ₂ O	78 ^a	[60]
15	CH ₃ COOOH, AcOH, 80 °C, 2 h	Au/HPS	72	[61]
16	O ₃ , AcOH, 160 °C	chromium(III) and manganese(II) salts	70	[62]

^aConversion

by rhenium bisperoxo complex and an MTO-catalyzed in situ generation of peroxyacetic acid as oxidant from acetic anhydride [64]. Later in 2002, Narayanan and co-workers reported the oxidation of 16 with H₂O₂ (30%) in acetic acid at 100 °C without catalyst (Table 1, entry 6) [52]. The authors obtained the desired product 10 in 86% yield and 95% conversion. This approach could represent a cheap and more ecofriendly method for the synthesis of 10, because it avoids mineral acid and chromium salts [52]. Sobkowiak and co-workers reported the use of iron(III) as a catalyst to activate H₂O₂ for the oxidation of 16 in glacial acetic acid (Table 1, entry 7) [53]. However, the oxidation process was not selective, and traces of 6-methyl-1,4-naphthoquinone were also identified. The reaction yields are not dependent on the type of the iron(III) salt used (perchlorate or acetate), except for iron(III) chloride, which exclusively leads to 1-chloro-2-methylnaphthalene as product [53].

Xiao and co-workers reported a manganese(II) naphthenate (MnPc)-catalyzed oxidation of 16 to furnish 10 in 60% yield with 75.6% conversion and 80% selectivity (Table 1, entry 8) [54]. The MnPc catalyst improves the stability of H₂O₂ and thus promotes a selective oxidation [54]. The approach developed by Beller's group, which applied a three component catalyst system consisting of iron(III) chloride, pyridine-2,6-dicarboxylic acid (H₂Pydic), and benzylamine (1:1:2.2) for the oxidation of 16 with H₂O₂, in tert-amyl alcohol (TAA), allowed to obtain 10 in 44% yield (Table 1, entry 9) [55]. Subsequently, Kulkarni's group evaluated the catalytic activity of selenium mesoporous molecular sieves (SeMCM-41) in the oxidation of 16 (Table 1, entry 10). The approach was performed using H_2O_2 as oxidant in acetic acid over SeMCM-41 (Si/Se = 30) at 100 °C [56]. In this case, a conversion of 99% was achieved and menadione (10) was obtained with 68% selectivity. According to the authors, the reaction mechanism involves the formation of an active selenium peroxo species. Additionally, they mentioned that the catalyst was easily separated from the reaction mixture by simple filtration [56].

Serindağ and co-workers disclosed the bidentate tertiary aminomethylphosphine complexes of Ru(II), Pd(II), and Co(II) with N,N-bis(diphenylphosphinomethyl)aminopropyltrietoxysilane (DIPAPTES) and the best yields were obtained using [(DIPAPTES)PdCl₂] complex and silica supported [SiO₂(DIPAPES)PdCl₂] complex, in 52% and 59% yields, respectively (Table 1, entry 11). In both cases, the formation of product 10 was observed with conversions of up to 90% [57]. In another approach, Uruş and co-workers used graphene oxide (GO)-supported bis(diphenylphosphinomethyl)amino GO@CHONHRN(CH₂PPh₂)₂MX₂ (M:Pd(II) and Pt(II))-type complexes as heterogeneous nanocatalysts, with Pd(II) com-

plexes showing the best catalytic activities with high selectivity compared to the Pt(II) complex, leading to 95–99% conversion and 60–65% selectivity (Table 1, entry 12) [58].

Other less common oxidizing agents have also been used for the oxidation of 2-methylnaphthalene (16) to produce menadione (10). Sulman and co-workers, for instance, achieved the oxidation of 10 using peracetic acid as oxidant in the presence of gold nanoparticles deposited on hypercrosslinked polystyrene (Au/HPS) (Table 1, entry 15). The best result was obtained using 1% Au/HPS in glacial acetic acid, which led to 96% conversion and 75% selectivity [61]. Another interesting example of oxidation process was reported by Mamchur and Galstyan, who used ozone as oxidizing agent in the presence of a mixture of chromium(III) and manganese(II) salts to furnish product 10 in 70% yield (Table 1, entry 16) [62]. The authors proposed that the oxidation involves an initial ozonation of the transition metal salts, which then oxidized substrate 16 to the desired product 10.

Oxidation of 2-methylnaphthol

Menadione synthesis was also achieved by oxidation of 2-methylnaphthol (17). The main advantage of using substrate 17, compared to 2-methylnaphthalene, is to avoid the formation of byproducts such as 6-methyl-1,4-naphthoquinone [65]. The conditions for the oxidation of 2-methylnaphthol (17) to menadione (10) are quite similar to those employed for the oxidation of 2-methylnaphthalene (16) using H₂O₂, molecular oxygen, and *tert*-butyl hydroperoxide as oxidizing agents.

Similar to the oxidation of compound 16, it is possible to oxidize 2-methylnaphthol (17) with H_2O_2 to produce menadione (10), as was demonstrated by Minisci and co-workers [66]. In this work, the oxidation of 17 with 60% aqueous hydrogen peroxide, using bromine and sulfuric acid as catalysts, provided menadione in 90% yield (Table 2, entry 1) [66]. Accord-

Table 2: Approaches for the oxidation of 2-methylnaphthol to menadione (10). conditions 17 10 Entry Conditions Catalyst Yield (%) Ref. 1 H₂O₂ (60%), MeOH, reflux, 20 min Br₂, H₂SO₄ 90 [66] 2 H₂O₂ (30%), MeCN, 80 °C, 30 min Ti-MMM-2 78 [65] 3 H₂O₂ (30%), MeCN, reflux, 40 min TiSBA-15 93 [67] 4 H₂O₂, acetone, 70 °C, 2 h Nb₂O₂-SiO₂ 60 [68] 5 97 H₂O₂, MeCN, 75 °C, 45 min NbSBA-15(2.2pH) [69] 6 H₂O₂/CH₃COOH, CO₂ (150 bar), 50 °C, 2 h Au (5%)/HPS 89 [70] 7 H₂O₂ (30%), TAA, 0 °C, 1 h FeCl₃·H₂O/H₂Pydic/benzylamine 55 [55]

HPA-n

Au/TiO₂

Au/C-2

FePcS-SiO₂

^aConversion.

8

9

10

11

12

ing to the proposed mechanism, the first step involves the electrophilic bromination of the corresponding phenol, followed by hydrolysis promoted by H_2O_2 [66].

CO₂, benzene/H₂O, 50 °C, 25 min

O₂ (3 atm), toluene, 80 °C, 6 h

O₂ (3 atm), toluene, 80 °C, 6 h

O2 (3 atm), toluene, 80 °C, 8 h

t-BuOOH, DCM, 80 °C, 1 h

Variations in the methods of 2-methylnaphthol (17) oxidation to menadione (10) with H_2O_2 were made by changing the catalytic systems in order to increase the yield and selectivity. These include the catalysis by Ti-based [65,67], Nb-based [68,69], and Au-based [70] heterogeneous systems. In addition to being more efficient, given the obtained atom economy, the reactions that use these catalytic heterogeneous systems are also presented as environmentally friendly. They are cleaner, either because of the low generation of waste or the use of environmentally friendly conditions, and they also allow recycling catalysts without losing efficiency [67,68,70].

Kholdeeva and co-workers reported the use of 30% aqueous H_2O_2 as oxidant and hydrothermally stable mesoporous mesophase titanium silicates (Ti-MMM-2) as catalyst group, producing 10 in 78% yield at 100% substrate conversion (Table 2, entry 2) [65]. During the studies it was observed that crucial factors affected the product yield, such as substrate concentration, H_2O_2 /substrate molar ratio, solvent nature, reaction temperature, and mesoporous size [65]. Selvaraj and co-workers reported the liquid-phase oxidation of 17 using Ti-containing mesoporous silica catalysts, TiSBA-15 (Table 2, entry 3) [67]. According to the authors, the best result was achieved with

TiSBA-15 ($n_{\rm Si}/n_{\rm Ti}=6$) catalysis and H₂O₂, exhibiting 93% selectivity to menadione (**10**). In addition, catalyst recycling experiments showed the TiSBA-15(6) had higher catalytic stability in the liquid-phase oxidation as compared to other titanium-containing mesoporous catalysts [67].

81^a

57

49

മവ

55

[71]

[72]

[72]

[73]

[74]

Cavani and co-workers reported a heterogenous catalyst system for the oxidation of 2-methylnaphthol (17) using 35% aqueous $\rm H_2O_2$ and niobium oxide dispersed in silica (Nb₂O₂-SiO₂) as catalyst (Table 2, entry 4) to obtain menadione (10) in 60% yield [68]. Another approach using niobium was developed by the Selvaraj group, which used mesoporous NbSBA-15 catalysts in the liquid-phase oxidation of 17 (Table 2, entry 5) [69]. Different NbSBA-15 catalysts were evaluated and with the optimized reaction conditions, menadione (10) was synthetized in 100% conversion and 97.3% selectivity using NbSBA-15(2.2 pH) and $\rm H_2O_2$ [69].

Sulman and co-workers, for instance, reported the synthesis of menadione (10) using supercritical (SC) carbon dioxide as green solvent [70]. The authors studied the oxidation using three metal-supported hypercrosslinked polystyrene (HPS) catalysts, which were Au (5%)/HPS, Pd (5%)/HPS and Pt (5%)/HPS, in SC CO₂ medium. The best conversion (89%) was obtained with Au (5%)/HPS, using CO₂ (150 bar), and a mixture of H₂O₂ and acetic acid as oxidant. Additionally, the selectivity of this process was 99% (Table 2, entry 6).

Beller et al. developed another approach using H_2O_2 as oxidizing agent in combination with a three component catalyst system consisting of FeCl₃·6H₂O, pyridine-2,6-dicarboxylic acid (H₂Pydic), and different benzylamines (1:1:2.2) (Table 2, entry 7). The reaction was carried out in *tert*-amyl alcohol (TAA), which led to product **10** in 55% yield and 99% conversion of **17** [55]. In addition to hydrogen peroxide, other oxidizing agents can be used in the synthesis of menadione (**10**) from **17** and include heteropoly acids [71], molecular oxygen [72,73], and organic peroxides [74].

Matveev and co-workers studied phosphomolybdovanadium heteropoly acids of Keggin-type with the general structure $H_{3+n}PMo_{12-n}V_nO_{40}$ (HPA-n) and their acidic salts as reversibly acting oxidants to convert 17 to 10 (Table 2, entry 8) [71]. The reaction was carried out in a two-phase solvent system under CO_2 atmosphere and the best selectivity (89%) was achieved using $H_5PMo_{10}V_2O_{40}$. In the proposed mechanism, first the HPA-n was reduced by 17, followed by product isolation, and regeneration of HPA-n by dioxygen.

The Kholdeeva group also reported the oxidation of 2-methylnaphthol (17) using molecular oxygen in the presence of gold nanoparticles as catalyst and the best yield of menadione (10) was obtained using 1.5% Au/TiO₂ as catalyst (57%, Table 2, entry 9), while the best conversion of 17 was furnished using 1% Au/C-2 catalyst (94%, Table 2, entry 10) [72]. In 2011, the same group patented a 2-methylnaphthol (17) oxidation approach using molecular oxygen in absence of catalyst under mild reaction conditions (Table 2, entry 11) [73]. The authors reported an oxidation study for this approach involving three alternative reaction mechanisms: free radical autoxidation, cation radical autoxidation, and thermal intersystem crossing (ISC), using ¹⁸O₂ labeling, spin-trapping, spectroscopic, mass spectrometric, kinetic, and computational techniques. After several experiments, the obtained results have demonstrated that the 2-methylnaphthol (17) oxidation occurs via a thermal ISC (spin inversion) [73]. Additionally, a zwitterionic intermediate formed in the rate-limiting step contributes significantly to the O2-based selective oxidation. However, this oxidation mechanism could be modified by the addition of initiators or bases and the predominant path depends mainly on the dioxygen pressure and the solvent nature [73].

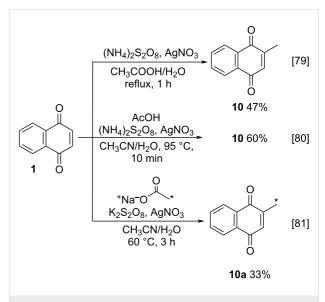
Another approach was reported by Zalomaeva and co-workers, which used an iron tetrasulfophthalocyanine (FePcS) supported catalyst (FePcS-SiO₂) in combination with the oxidizing agent *tert*-butyl hydroperoxide for the oxidation of 2-methylnaphthol (17) [74]. Interestingly, the oxidation was efficient with only 0.25 mol % of the catalyst providing 55% selectivity and approximately 50% yield. Indeed, the best product yield

(approximately 55%) was achieved at a 95% conversion and 55% selectivity, using dichloromethane and *tert*-butyl hydroperoxide at 80 °C (Table 2, entry 12).

Methylation of 1,4-naphthoguinone

Another route to prepare menadione (10) involves the methylation of 1,4-naphthoquinone. Because of their electron-deficient character, quinones are highly reactive with nucleophilic radicals [75]. The most useful alkylation approach is the Kochi–Anderson method [76] (or also known as Jacobsen–Torssell method [77,78]), via oxidative decarboxylation, where the quinone reacts with a carboxylic acid in the presence of silver(I) nitrate and ammonium or potassium peroxydisulfate. Nucleophilic free radicals are generated from the carboxylic acid through decarboxylation mediated by [Ag⁺]-peroxydisulfate, followed by their coupling with the quinone, providing the respective alkylated product.

One of the first practical applications of this methodology to produce menadione (10) was described by Ashnagar and co-workers [79]. 1,4-Naphthoquinone (1) was treated with acetic acid in the presence of ammonium persulfate, as oxidizing agent, and silver(I) catalysis for only 1 hour, furnishing menadione (10) in 47% yield (Scheme 1). After this pioneering work, some adaptations were reported. As an example, Liu and co-workers synthetized 10 using a much simpler way (Scheme 1) [80]. These authors reported the methylation and alkylation of 1,4-naphthoquinone (1) in the presence of (NH₄)₂S₂O₈ and AgNO₃ as catalyst to obtain 10 in 60% yield. Recently, Onuki and co-workers conducted dimerization reactions of 10, exploring an interesting artifice to track the dimer-



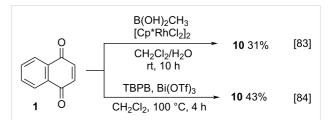
Scheme 1: Selected approaches for menadione synthesis using silver(I) as a catalyst.

ization reaction path: they synthesized 2-(methyl-¹³C)-1,4naphthoquinone (10) (Scheme 1) [81]. For that, sodium acetate-2-13C was used as the source of the methyl radical, generated by its treatment with K₂S₂O₈ and AgNO₃. After 3 hours at 60 °C, the ¹³C-labelled menadione was obtained in 33% yield.

In 1991, Coppa and co-workers reported the homolytic methylation of 1,4-naphthoquinone (1) using simple sources of methyl radicals [82]. In the methylation reaction using tert-butyl hydroperoxide and Fe(OAc)₂OH, menadione (10) and 2,3-dimethyl-1,4-naphthoquinone (18) were obtained in 80% yield with a 75:25 ratio, respectively (Scheme 2, method A). In the methylation reaction using methyl radicals generated by the redox decomposition of H₂O₂ in DMSO solution, compounds 10 and 18 were obtained in an overall yield of 80-90% with a 77:23 ratio, respectively (Scheme 2, method B). Finally, the use of H₂O₂ thermal decomposition in acetone with catalytic methanesulfonic acid, led to compounds 10 and 18 in 47% yield with 86:14 ratio (Scheme 2, method C). In all cases, the monomethylation was not selective and even at partial conversions of naphthoquinone, significant amounts of dimethyl derivatives were formed. The authors explained the unfavorable steric and polar effects of the methyl group in the quinone ring were probably very low or they were balanced by the favorable enthalpic effects.

Another interesting work was reported by Wang and co-workers, who studied rhodium complexes as catalysts for the arylation and alkylation of benzo- and naphthoquinones [83]. They synthesized menadione (10) (Scheme 3) by reacting 1,4naphthoquinone (1) with methylboronic acid in the presence of [Cp*RhCl₂]₂ as catalyst for 10 h, in 31% isolated yield. Later in 2019, Yang and co-workers performed a bismuth catalyst system study for the methylation and alkylation of quinone derivatives [84]. Furthermore, they also evaluated the methylation

without catalysts and with the use of lanthanum(II) and copper(II) salts as additive. However, the best results were achieved with bismuth(III) triflate. The use of tert-butyl hydroperoxide in the presence of bismuth(III) triflate for the methylation of 1,4-naphthoquinone (1) provided 10 in 43% yield, in already optimized conditions (Scheme 3).



Scheme 3: Methylation approach of 1,4-naphthoquinone using i) rhodium complexes/methylboronic acid and ii) bismuth(III) triflate and tert-butyl hydroperoxide.

Demethylation of 2-methyl-1,4dimethoxynaphthalene

The oxidative demethylation of 1,4-dimethoxyarenes is another valid synthetic approach to achieve 1,4-quinones, with the oxidative demethylation of 2-methyl-1,4-dimethoxynaphthalene (19) can be used to synthesize menadione (10). The oxidizing agents most commonly used in oxidative demethylation are cerium(IV) ammonium nitrate (CAN), and silver(II) oxide. However, the application of these oxidants is limited, as the synthesis of quinones requires milder reaction conditions. With the purpose of obtaining milder conditions applicable to the synthesis of more complex quinones, including menadione (10), some oxidative demethylation methods have been developed based on other oxidizing agents, such as cobalt(III) fluoride [85], phenyliodine(III) bis(trifluoroacetate) (PIFA) [86] and tert-butyl hydroperoxide [87] (Table 3).

In 1999, Tomatsu and co-workers performed the synthesis of menadione (10) through demethylation of 2-methyl-1,4-dimethoxynaphthalene (19), using cobalt(III) fluoride as oxidizing agent (Table 3, entry 1) [85]. The obtained results showed that the cobalt(III) fluoride catalyst was comparable with other oxidizing agents already well-established for this synthesis, like silver(II) oxide and ammonium cerium(IV) nitrate. Cobalt(III) fluoride proved to be a good oxidizing agent for the synthesis of menadione (10). This approach furnished 10 in 92% yield, although the reaction required a longer reaction time compared to the few minutes using AgO and CAN.

Another problem associated with the use of oxidizing metallic agents is the generation of metallic residues that can be toxic to the environment or act as contaminants of the desired products, such as pharmaceuticals. This problem has led to the search for clean and environmentally friendly methods that reduce or do not generate metallic residues. In 2001, Tohma and co-workers published an alternative and sustainable methodology, using phenyliodine(III) bis(trifluoroacetate) (PIFA) as an oxidizing agent of the demethylation reaction [86]. The hypervalent iodine(III) proved to be a good oxidizing agent in the formation of 10 (92% yield) (Table 3, entry 2). According to the authors, this is a good synthetic path, since PIFA has a low toxicity and it is easily accessible. Subsequently, Wójtowicz and co-workers studied a series of experiments in order to test the oxidative action of tert-butyl hydroperoxide and the role of organoselenes as catalysts in the demethylation reaction of hydroquinone ethers [87]. The combination of tert-butyl hydroperoxide and poly(bis-1,2-phenyl)diselenide was the one that showed the best result: 90% yield after 12 h reaction and recrystallization (Table 3, entry 3).

Construction of the naphthoquinone ring

In addition to the oxidation of naphthalene derivatives, the construction of the naphthoquinone ring is also a viable synthetic method to produce menadione. This is a very efficient strategy with great synthetic value, however, it is less used when compared to other methodologies, as it involves more steps and sometimes more complex reactions.

One of the pioneering methods for naphthoquinone ring construction was reported by Horii and co-workers who performed the synthesis of menadione (10) from itaconic acid, via 2-methyl-1-tetralone [88]. Two synthetic routes were performed (Scheme 4); route A proceeds via the oxidation of 2-methyl-1-tetralone (25) using chromium(IV) oxide and route B starts with C-2 bromination of 25, giving the intermediate 26, which was reduced to 27, and oxidized in the presence of chromium(IV) oxide leading to 10 with 37% yield. Although path B was the more complicated to perform, it was the higher yielding route.

In 2002, an interesting methodology for menadione synthesis was reported by Kacan and Karabulut (Scheme 5). The authors studied a Diels-Alder reaction, using LiClO₄-diethyl ether (LPDE) as a catalyst, 1-ketoxy-1,3-butadiene **28** as a diene and 2-methyl-1,4-benzoquinone (**29**) as dienophile. By this method, menadione was obtained in 90% yield in 5 hours [89].

Another route was developed by Murahashi and co-workers that used *p*-cresol as synthetic precursor [90]. First, *p*-cresol, in the presence of *tert*-butyl hydroperoxide, was oxidized to 4-methyl-4-*tert*-butyldioxycyclohexadienone using tris(triphenylphosphine)ruthenium(II) dichloride as catalyst. Then, a BF₃·OEt₂-catalzyed migration of the methyl group to the C-2 position and removal of the *tert*-butoxy group in a 1,1,1,3,3,3-hexafluoroisopropanol (HFIP)/toluene mixture afforded 2-methyl-1,4-benzoquinone (29). Finally, a Diels–Alder reaction was performed with 1,3-butadiene, followed by dehydrogenation gave menadione (10). This proved to be a good synthetic route, leading to menadione in approximately 80% overall yield (Scheme 6).

Scheme 5: Menadione synthesis via Diels-Alder reaction

Scheme 4: Synthesis of menadione (10) from itaconic acid.

Another interesting synthetic approach was reported by Mal and co-workers, who synthetized menadione (10) via a demethoxy-carbonylating annulation of methyl methacrylate (33) with 3-cyanophthalide (32), in the presence of lithium *tert*-butoxide as catalyst (64% yield) (Scheme 7) [91].

Recently, Dissanayake and co-workers tested the stability of furans to be used as a diene in Diels-Alder reactions for the

OH RuCl₂(PPh₃)₃
$$t$$
-BuOOH, benzene, rt t t -BuOOH, benzene, rt t t -BuOOH, benzene t -BuOOH, benzene

Scheme 7: Synthesis of menadione (10) via demethoxycarbonylating annulation of methyl methacrylate.

synthesis of p-benzoquinones and p-hydroquinones or the synthesis of menadione (10). The furan derivative 34 was used as a diene and 2-iodophenyltrifluoromethanesulfonate (35) as a dienophile, in the presence of n-butyllithium, forming 10 in 55% yield (Scheme 8) [92].

In the same year, Gogin and and co-workers developed a method for the synthesis of menadione (10) using dienophiles from o-cresol or o-toluidine [93]. Mo-V-P-HPA-X catalysts were tested, where X is the amount of V atoms present in the molecule. All reactions led to the product in good yields. The route from o-toluidine (36) to form 10, using Mo-VP-HPA-10 as catalyst, in the presence of 1,3-butadiene, presented the best yield (about 33%) (Scheme 9). This presented itself as a good synthetic route, considering that it used easily accessible reagents and there was no formation of polluting products.

Electrochemical synthesis

Although not common, menadione (10) can be readily produced through electrochemical synthesis. This methodology allows the reuse of the electrolyte and demonstrates a significant substrate conversion. Raju and co-workers [94], for instance, reported the electrochemical synthesis of 10 from menadiol (14) using galvanostatic biphasic electrolysis (Scheme 10). For this approach, two smooth platinum foil elec-

trodes placed 1 cm apart in the upper aqueous phase were electrolyzed galvanostatically (30 mA/cm²). Additionally, a NaBr solution acidified with $\rm H_2SO_4$ was used as electrolyte. The voltage during electrolysis was 2.1 V and when the reaction was completed (charge passed of 2 F/mol), menadione (10) was obtained in 99% yield.

Menadione as synthetic precursor

There are many strategies available for structural modifications of menadione (10) and in the following section we will discuss their evolution and present those methods generally used to access menadione derivatives, to highlight the importance of this substrate for organic synthesis. The most common reaction types and reaction sites of menadione derivatization are depicted in Figure 4. The vast majority of the methods involve the unsaturated α,β -system of the naphthoquinone nucleus, due to its greater reactivity when compared to the adjacent aromatic ring, which depends on previous modifications in the menadione intermediates [95].

Epoxidation reactions

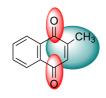
The use of menadione in the preparation of epoxides is widely reported in the scientific literature. In nature, menadione epoxides are formed through oxidation reactions in vivo, that occur in protein processes dependent on vitamin K [96,97]. Dwyer

Scheme 9: o-Toluidine as a dienophile in a Diels-Alder reaction for the synthesis of menadione (10)

Scheme 8: Furan 34 used as a diene in a Diels-Alder reaction for the synthesis of menadione (10).

Scheme 10: Representation of electrochemical synthesis of menadione.

reduction and acylation carbonyl condensation

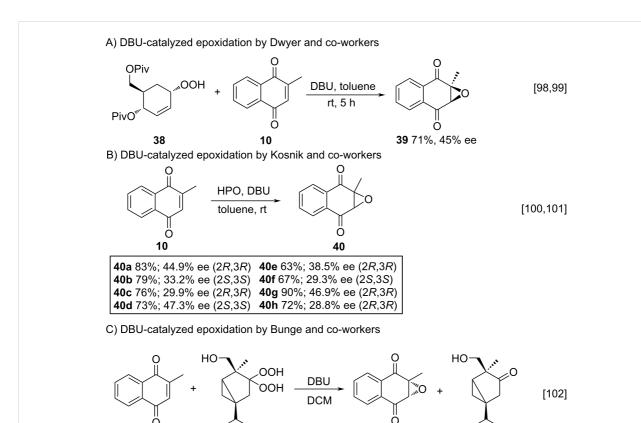


epoxidation
pericyclic reactions
menadione as nucleophile
alkylation and acylation via free radicals
1,4-addition-type reactions

Figure 4: Reaction sites and reaction types of menadione as substrate.

and co-workers described a procedure using sugar-derived hydroperoxides for the synthesis of epoxides in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) as base [98,99]. The authors studied a broad range of quinone derivatives among which compound 39 was obtained through reaction of menadione (10) and hydroperoxide 38 in 71% yield and 45% ee (Scheme 11A). Analogously, Kosnik and co-workers described a similar epoxidation methodology using a series of pyranosederived anomeric hydroperoxides (HPO) to obtain epoxides 40 with moderate ees (Scheme 11B) [100,101]. Bunge and co-workers used the enantiomerically pure dihydroperoxide 41 in the DBU-mediated epoxidation of menadione (10) for the enantioselective synthesis of epoxide 42 (92% yield and 45–66% ee) (Scheme 11C) [102].

Another interesting approach to menadione epoxidation is the use of a phase-transfer catalyst (PTC). Ooi and co-workers studied the epoxidation of menadione (10) and other carbony-lated substrates using tetrabutylammonium bromide (TBAB) as catalyst and the optimized reaction conditions involved ultrasound irradiation, obtaining quantitative yields when compared to the mechanical stirring procedure, thus demonstrating the best efficiency of the method [103]. In 2002, Arai and co-workers reported studies involving epoxidation reactions of menadione (10) using H_2O_2 as oxidant and a chiral salt derived from cinchonine PTC 44 as catalyst. Despite of good yields, the method did not demonstrate good enantioselectivity results [104]. Berkessel and co-workers, in turn, described the use of asymmetric Weitz–Scheffer-type epoxidation of menadione

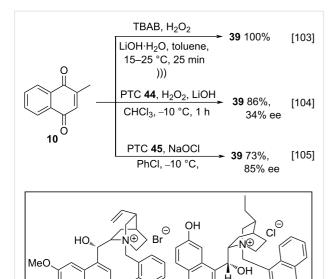


42 92%, 66% ee

Scheme 11: DBU-catalyzed epoxidation of menadione (10).

10

(10), mediated by cinchona alkaloid PTC 45, showing high enantioselectivity (85% ee) (Scheme 12) [105].



Scheme 12: Phase-transfer catalysis for the epoxidation of menadione

PTC 45

PTC 44

Exploring a different epoxidation reaction approach, Lattanzi and co-workers reported a methodology using a (+)-norcamphor hydroperoxide **46**, to generate the menadione-derived epoxide **40** in 51% ee, under optimized reaction conditions employing n-BuLi/THF [106]. The method proved to be effective

Scheme 13: Menadione epoxidation using a hydroperoxide derived from (+)-norcamphor.

at recovering approximately 95% of the enantiopure alcohol 47. This allowed the alcohol's effective reconversion to hydroperoxide 46 and proved to be a useful method for the enantioselective epoxidation of menadione (10) (Scheme 13).

Pericyclic reactions

The Diels–Alder reaction, among pericyclic reactions, is a very important synthetic approach to obtain several molecular scaffolds, including naturally occurring molecules, drugs, polymers, and heterocycles with promising biological activity. Especially, Diels–Alder reactions involving quinones and dienes as starting materials allow for the synthesis of more complex molecules such as natural products. Within this scope, the menadione (10) molecule has been explored as substrate for this versatile reaction.

Ryu and co-workers described an enantioselective and structurally selective Diels-Alder reaction for the synthesis of asymmetric compound **50** catalyzed by a chiral oxazaborolidinium cation (**49**) [107]. This type of catalyst has been used in several Diels-Alder reactions proving to be an excellent choice for highly enantioselective reactions [108]. For instance, the reaction of menadione (**10**) with 2-triisopropylsilyloxy-1,3-butadiene (**48**) gave compound **50** in 96% yield and 91% ee (Scheme 14).

In 2006, Nishimoto and co-workers described an interesting application of a Diels-Alder reaction conducted, among others, in water and fluorous solvents [109]. Especially, the employment of water as solvent has gained great attention in the last few

Scheme 14: Enantioselective Diels-Alder reaction for the synthesis of asymmetric quinone 50 catalyzed by a chiral oxazaborolidinium cation.

years in organic synthesis due to its physical and chemical properties. The authors studied the Diels-Alder reaction between menadione (10) and 2,3-dimethyl-1,3-butadiene (51) and it was possible to draw a comparison between the reported properties for this type of solvent (Table 4). The studies showed that the emulsion, prepared by sonication, of an equimolar mixture of lithium perfluorooctane sulfonate (LiFOS) and perfluorohexane (PFH) in aqueous medium resulted in a significant increase of the reaction rate, when compared to other reaction conditions [109]. As an example, using an equimolar ratio of 500 mM, the Diels-Alder reaction achieved 98% and 2% recovery of starting material after 72 h, at an estimated product generation rate of 142.5 µM/h. In this context, it was possible to apply a green methodology without the application of heating or the use of Lewis acids, contributing significantly to the progress of the synthesis studies in aqueous medium.

The Diels-Alder reaction using menadione (10) was also studied by Bendiabdellah and co-workers, who reported the intramolecular Diels-Alder domino reactions promoted by Lewis acids [110]. The reaction involving menadione (10) and

excess of triene 53 was carried out using boron trifluoride diethyl etherate or zirconium(IV) tetrachloride to furnish product 54 in good yield (Scheme 15).

The same group also explored the Diels-Alder reaction of menadione (10) with trienes 55, 57, and 59 (Scheme 16) [110]. The scandium(III) triflate-catalyzed reaction showed the best results in terms of performance, producing furanone 56 in 65% yield. Additionally, this study was also extended to the synthesis of hetero-tetracyclic derivatives containing endocyclic nitrogen atoms. The best result was obtained using triene 57, and 2.0 equivalents of scandium(III) triflate, affording compound 58 in 70% yield. Finally, the optimized conditions were employed to react menadione (10) with the triene 59, using 5.0 equivalents of scandium(III) triflate, forming compound 60 in 55% yield.

Additional studies were made, using protected trienes, and in Scheme 17 below the optimized reaction conditions are shown for the synthesis of compounds **62** and **64** (Scheme 17) [110].

Table 4: Media effect in the initial rates of Diels-Alder reaction between menadione and 2,3-dimethyl-1,3-butadiene.

Conditions (organic solvents)	Reaction rate $(\mu M/h)$	Conditions (aqueous)	Reaction rate (μM/h)
perfluorohexane (PFH)	0.25	aq SDS (100 mM)	3.87
<i>n</i> -hexane	0.38	aq TFE (100 mM)	3.80
diethyl ether	1.00	aq LiOTf (100 mM)	1.15
methanol	1.27	aq LiOS (100 mM)	2.35
acetonitrile	1.60	aq LiFOS (100 mM)	11.6
dichloromethane	3.16	neat	<0.2
toluene	0.73	LiFOS/PFH in water	140.5
water	1.40	(500 mM/500 mM)	142.5

Scheme 15: Optimized reaction conditions for the synthesis of anthra[9,1-bc]pyranone.

$$\begin{array}{c} H_2C \\ \hline \\ \textbf{61} \text{ (1.5 equiv)} \\ \hline \\ \textbf{CH}_2 \\ \hline \\ \textbf{BF}_3 \cdot \text{OEt}_2 \text{ (2 equiv)} \\ \textbf{DCM}, -45 \, ^{\circ}\text{C}, 5 \text{ h} \\ \hline \\ \textbf{62} \text{ (\pm)} 85\% \\ \hline \\ \textbf{RES}_3 \cdot \text{OEt}_2 \text{ (2 equiv)} \\ \textbf{DCM}, -45 \, ^{\circ}\text{C}, 5 \text{ h} \\ \hline \\ \textbf{62} \text{ (\pm)} 85\% \\ \hline \\ \textbf{NHBoc} \\ \textbf{CH}_2 \\ \textbf{Sc}(\text{OTf})_3 \text{ (2 equiv)} \\ \textbf{DCM}, 0 \, ^{\circ}\text{C}, 2.5 \text{ h} \\ \hline \\ \textbf{64} \text{ (\pm)} 70\% \\ \hline \\ \textbf{Scheme 17: Synthesis of derivatives employing protected trienes.} \end{array}$$

In 2019, Sultan and co-workers described a methodology for the synthesis of quinone derivatives using a combination of potassium persulfate, trifluoroacetic acid (TFA), and blue-LED light [111]. Under these conditions, menadione (10) and terminal alkynes 66 underwent a [2 + 2] cycloaddition reaction generating compounds containing cyclobutene rings (67a–c), that are important precursors in natural products syntheses. It is important to note that the choice of the blue-LED source was made after preliminary studies demonstrated the occurrence of reac-

tions with benzoquinones using the compact fluorescent lamp (Scheme 18).

Reduction and acylation reactions

Menadione reduction reactions are one of the most important types of reactions and are directly related to some of its characteristic properties, such as the biological redox cycle. The main and most common menadione reduction product is menadiol or vitamin K_4 , followed by its dialkyl ether and diacyl derivatives.

Menadione can be easily converted to menadiol (14) by reduction with sodium dithionite, as first described by Fieser (Scheme 19) [47]. The reaction was carried out in a separatory funnel to which was added menadione (10), sodium hydrosulfite, and water [47]. The mixture was shaken for a few minutes until the solution passed through a brown phase and became yellow. Despite of being an old method, it is very efficient and widely used with some adaptations. In 2003, Ito and co-workers also used sodium dithionite to obtain menadiol (14) from 10. However, in this case, the authors mixed menadione (10) and sodium hydrosulfite in acetic acid and water to obtain 14 in 97% yield [112]. More recently, still using sodium dithionite, Suhara and co-workers reported in their various works on the synthesis of vitamin K analogues, the use of menadione (10) to obtain 14 [113-117]. In these works, menadione (10) was reduced by using an aqueous 10% sodium dithionite solution in diethyl ether to furnish alcohol 14 in a quantitative yield.

Still about reduction reactions, Niemczyk and Van Arnum described a green methodology for reduction of menadione (10), during the pegylation of 14 to improving the solubility of the studied compounds [118]. The authors reduced 10 with sodium dithionite under ultrasound irradiation, generating the reduced

adduct in 79% yield (Scheme 20). Depending on the type of solvent used, the yield may vary due to oxidation of the alcohol **14** back to **10** because of its low stability in solution. The pegylation strategy involved monomethoxypoly(ethyleneglycol)succinimide carbonate (mPEG-SC, **68**), giving the pegylated product and *N*-hydroxysuccinimide (NHS) as the sole byproduct. The latter can be recycled again to the pegylation reagent. This study showed better results when compared to the methodology for a phosphorylation of **14** developed by Fieser [119], a procedure carried out in two steps whose main difficulties are the separation of pyridine byproducts and inorganic phosphate (Scheme 20).

Kulkarni and co-workers reported a method for menadione reduction mediated by 5,6-*O*-isopropylidene-L-ascorbic acid (70, R = H) under UV light irradiation [120]. Initial studies were carried out using lawsone (4) and after optimization of the reaction conditions, it was extended to other quinones, including menadione (10). The best conditions were 1,2-dimethoxyethane (DME) as solvent, temperature 25 °C, under ultraviolet light irradiation (125 W lamp) using a Pyrex filter in an immersion-well photoreactor. It was observed that the presence of free hydroxy groups in 70 was essential for the quinone reduction reaction to occur, when compared to the 2,3-di-*O*-methylated

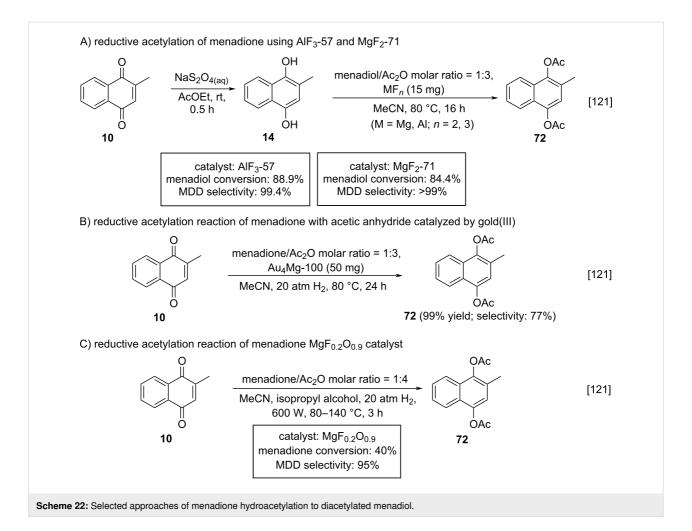
derivative (**70**, R = Me). Under the reported conditions, the reduction of menadione (**10**) gave compound **14** in 42% yield after 80 hours (Scheme 21).

Dobrinescu and co-workers studied, besides acetylation reactions, the hydroacetylation of menadione for the synthesis of diacetylated menadiol derivatives through heterogeneous catalysis [121]. The first methodology involved the reduction of menadione (10) to menadiol (14), with sodium dithionite, followed by hydroacetylation of 14 with acetic anhydride, using nanoscopic acidic hydroxylated metal fluorides $MF_{n-x}(OH)_x$ (M = Mg, Al; n = 2, 3; x < 0.1) as catalysts. This type of catalysts has a huge acidic versatility, once they can behave as Brønsted or Lewis acids. The reductive acetylation of 10 occurred in two steps and at high selectivity conversion rates when using AlF₃-57 and MgF₂-71 (Scheme 22A). The second proposal explored the reductive acetylation reaction of menadione (10) with acetic anhydride catalyzed by gold(III) deposited on the qualified metallic fluorides. The deposition of gold on metallic fluorides allowed the one-pot hydroacetylation of menadione (10) to diacetylated menadiol 72, while the deposition of gold on silica allowed only the hydrogenation of 10 to 14 (Scheme 22B). The authors also observed that catalysis by hydroxylated fluorides led to a higher reaction speed when compared to the use of gold-impregnated catalysts, indicating that gold(III) impregnation blocks are part of the active sites for

menadiol acetylation. The third approach was based on the Meerwein–Ponndorf–Verley (MPV) reduction coupled with an acetylation reaction, in combination with acid–base magnesium oxide fluoride (MgF $_x$ O $_y$) as catalyst. The MPV reduction is known as being highly selective for the reduction of C=O bonds [122]. The best methodology for this third catalytic approach was based on the MgF $_{0.2}$ O $_{0.9}$ catalyst, with the reaction time being optimized from 48 h to 3 h, which resulted in an increase of menadione (10) conversion (40%) and also in the reaction selectivity (95%) (Scheme 22C).

Another approach involving menadione acetylation was reported by Yadav and co-workers, who described a methodology for the synthesis of acetylated quinone derivatives catalyzed by bismuth(III) triflate [123], through an adapted Thiele–Winter acetoxylation reaction. The standard procedure involved the use of acetic anhydride and sulfuric acid catalysis. However, the use of sulfuric acid, a strong acid and oxidizing agent, can produce tar in some cases. In order to get around this problem a strategy applying Lewis acids can be used. Among the options, bismuth triflate is a catalyst, with the additional advantages of low-cost and easy preparation from commercially available bismuth oxide and triflic acid. Quinone acylation reactions took place under mild conditions using acetic anhydride and 2 mol % bismuth(III) triflate catalyst and compound 73 was obtained in 75% yield (Scheme 23).

Scheme 21: Menadione reduction by 5,6-O-isopropylidene-L-ascorbic acid under UV light irradiation.



$$\begin{array}{c} O \\ Ac_2O \\ 2 \text{ mol } \% \text{ Bi(OTf)}_3 \\ \hline CH_3CN, 50 \text{ min, rt} \\ OAc \\ \hline 10 \\ Scheme 23: Thiele-Winter reaction catalyzed by Bi(OTf)}_3. \end{array} \end{subarray}$$

Carbonyl condensation

Another possible derivatization involves transformations at the carbonyl groups in menadione that can be achieved through the addition of nucleophiles to the carbonyl group at position C-4. In this context, Nagaraja and co-workers reported the condensation of menadione (10) under different mild conditions, during the development of analytical methods for determining 10 in pharmaceutical preparations [124]. In method A, menadione (10) was treated with concentrated sulfuric acid and resorcinol (74), generating the intermediate 75 which underwent intramolecular condensation, to furnish 76. In method B, menadione (10) was treated with 3-methyl-2-benzothiazolinone hydrazone

(77) in alkaline medium forming diazocompounds as addition products, which spontaneously lose N_2 to form product 78 (Scheme 24).

Tang and co-workers described the synthesis of thiosemicarbazone 80 from menadione (10) through a condensation reaction with thiosemicarbazide (79), which was used as a ligand in the synthesis of metal complexes using different transition metals, in refluxing ethanol (Scheme 25) [125].

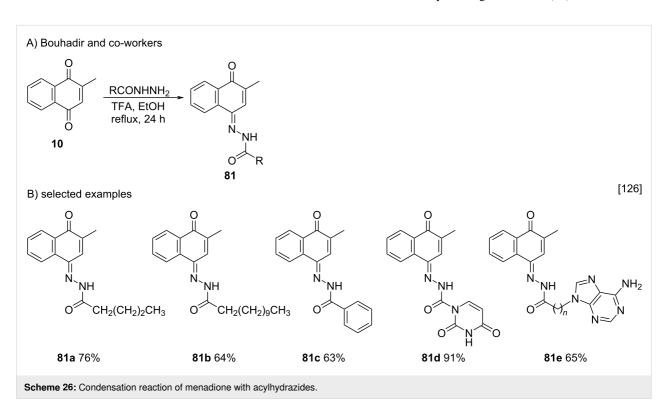
Examining a broad range of hydrazides in the condensation reaction with menadione, Bouhadir and co-workers reported the synthesis of various menadione acylhydrazone derivatives [126]. In this work, various acylhydrazides prepared by reaction of hydrazine hydrate with different esters were reacted with menadione (10) in trifluoroacetic acid under ethanol reflux conditions (Scheme 26) to synthesize acylhydrazones 81 in 63–91% yield. In view of the different structures that compounds 81a–e could adopt, after analysis by 2D NMR-NOESY spectra, it was found that all products were obtained as *E*-geometrical isomers and *trans*-conformers.

Scheme 24: Carbonyl condensation of menadione using resorcinol and a hydrazone derivative.

Scheme 25: Condensation reaction of menadione with thiosemicar-

Menadione as a nucleophile

Electron-rich 1,4-naphthoquinones, such as 2-hydroxy-, 2-amino-, and 2-alkylnaphtho-1,4-quinones may react as nucleophiles. Hence, menadione (10) can act as a nucleophile in, for example, bromination reactions [127] and aldol-type reactions with aldehydes and ketones [128]. In this context, Fry and co-workers explored the electrophilic substitution reaction to synthesize 2-methyl-3-bromonaphthalene-1,4-dione (82), an important intermediate used for the synthesis of naphthoquinones functionalized with organochalcogens [127]. Compound 82 was obtained by treating menadione (10) with molecular



bromine in the presence of sodium acetate and acetic acid in 76% yield. With brominated compound 82 at hands, the authors obtained four menadione derivatives 83a–d functionalized with organochalcogenic moieties after treatment of compound 82 with the respective ditellurides, a disulfide and a diselenide (Scheme 27).

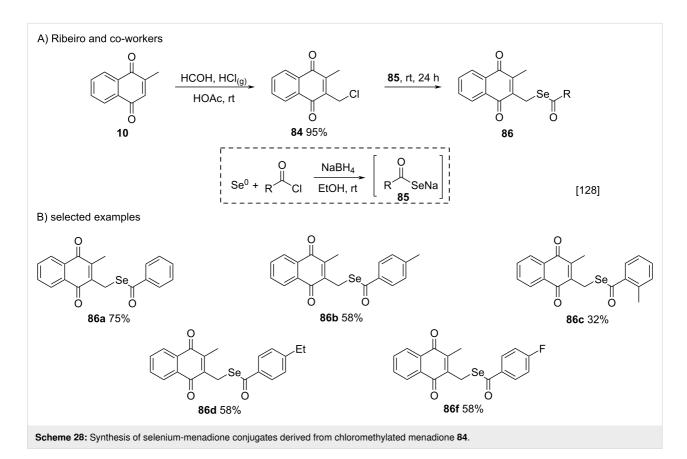
Recently, Ribeiro and co-workers used menadione as a nucleophile for the synthesis of 3-chloromethylated menadione **84**, a key intermediate used to prepare selenium-menadione conjugates **86** [128]. In this work, compound **84** was prepared through the reaction of menadione (**10**) with formaldehyde in the presence of gaseous HCl bubbled into the reaction medium

[129,130]. Then, the chloromethyl derivative **84** was treated with diselenides, generated in situ from the reaction between Se⁰, NaBH₄ and different acid chlorides, to form conjugates **86** in 24–75% yields (Scheme 28).

Alkylation and acylation by free radicals

One of the largest groups of reactions that use menadione (10) as substrate comprises free radical alkylation and acylation reactions. The most useful alkylation approach is the Kochi–Anderson method [76] or (Jacobsen–Torssell method [77]) where 10 reacts with a carboxylic acid in the presence of silver(I) nitrate and ammonium or potassium peroxydisulfate (Scheme 29).

$$\begin{array}{c} 1) \text{ NaOAc/HOAc, } 0 \text{ °C} \\ \hline 2) \text{ Br}_2 \\ \hline \\ \textbf{82 76}\% \\ \hline \\ \textbf{83a X} = \text{Te, R} = \text{H 9}\% \\ \textbf{83b X} = \text{Te, R} = \text{OCH}_3 \text{ 75}\% \\ \textbf{83c X} = \text{S, R} = \text{OCH}_3 \text{ 2}\% \\ \textbf{83d X} = \text{Se, R} = \text{OCH}_3 \text{ yield not reported} \\ \hline \\ \textbf{Scheme 27: Menadione derivatives functionalized with organochalcogens.} \\ \end{array}$$



In the past twenty years, several menadione alkylation studies have been carried out based on the Kochi–Anderson method [131-141]. In 2001, Salmon-Chemin and co-workers described the preparation of alkylated compounds **89** and **90** via oxidative decarboxylation of diacids and *N*-Boc-protected amino acids β-alanine, γ-aminobutyric acid, 5-aminovaleric acid, and 6-aminocaproic acid [131]. The compounds were obtained in moderate yields (37–63%), and a decreased yield was observed with an increase of the aliphatic chain length, which was more accentuated for the diacid derivatives, and more subtle for the amino acid derivatives (Scheme 30). The same methodology was applied during the synthesis of oligopeptides linked to **10** (Scheme 30) [132-134].

In detailed studies, initiated by Commandeur and co-workers [135] and expanded by Naturale and co-workers [139], the alkylation capabilities of menadione (10) were evaluated, exclu-

sively, with several amino acid types by Kochi–Anderson radical decarboxylation [76]. In the studies developed by Naturale's group, α -, β -, and γ -amino acids of linear and branched chains were used, as well as different amine protection groups (Table 5). The results revealed that the functionalization of naphthoquinones by a radical addition of decarboxylated α -, β - and γ -N-protected amino acids was possible. However, the high conversion rates of the reagents to the desired products were not reflected in the isolated product yields, which was attributed to the workup and purification processes. It was also possible to demonstrate a moderate influence of the N-protecting group on the reaction outcome, although electronic effects can be considered to play a role, especially with substituted α -amino acids.

Lanfranchi and co-workers also studied the menadione (10) alkylation by oxidative decarboxylation using carboxylic acids containing nitrogenous heterocycles as substituents, and achieved very interesting results [137]. The authors observed that the desired product 95 was obtained with very low yield, due to the competition between the Kochi–Anderson [76] reaction and the Minisc reaction (Scheme 31) generating a mixture of polymeric pyridine derivatives [142]. The Minisci reaction [142] is a nucleophilic radical substitution to an electron-deficient aromatic compound, in the presence of silver(I) nitrate, ammonium persulfate, and heat, reaction conditions that are very similar to those of the Kochi–Anderson procedure. Under

10
$$\frac{AgNO_3, (NH_4)_2S_2O_8}{MeCN/H_2O, 60-65 °C}$$
 $\frac{AgNO_3, (NH_4)_2S_2O_8}{2.5 \text{ h}}$ $\frac{AgNO_3, (NH_4)_2S_2O_8}{NeCN/H_2O, 60-65 °C}$ $\frac{AgNO_3, (NH_4)_2S_2O_8}{MeCN/H_2O, 60-65 °C}$ $\frac{AgNO_3, (NH_4)_2S_2O_8}{2.5 \text{ h}}$ $\frac{AgNO_3, (NH_4)_2S_2O_8}{NeCN/H_2O, 65-70 °C}$ $\frac{A$

Entry	Amino acid	Product	Protective group	Conversion (%)	Yield (%)
		92a	Вос	74	40
4	HO ₂ C NHPHG	92b	Troc	89	58
1	H	92c	Ac	98	49
		92d	TFA	100	42
		89a	Вос	96	58
2	HO ₂ C 1NHPHG	92e	Troc	100	42
2	H	92f	Ac	57	13
		92g	TFA	100	51
		89b	Boc	88	61
0	HO ₂ C NHPHG	92h	Troc	86	60
3	HO ₂ C NHPHG	92i	Ac	75	54
		92j	TFA	98	53
		92k	Boc	0	0
4	HO ₂ C NHPHG	921	Troc	35	18
4	"""	92m	Ac	29	16
		92n	TFA	92	27
	110.0 ()	92o	Boc	0	0
_	HO ₂ C NHPHG	92p	Troc	42	34
5		92q	Ac	24	24
	Ph	92r	TFA	29	29
		92s	Boc	100	50
0	HO ₂ C NHPHG	92t	Troc	100	55
6	INTERIG	92u	Ac	93	57
		92v	TFA	100	39

$$\begin{array}{c} \text{CO}_2\\ \text{HO}_2\text{C} \\ \text{93a X} = \text{Cl}\\ \text{93b X} = \text{NO}_3 \end{array} \begin{array}{c} \text{AgNO}_3\\ \text{H}_2\text{O}, \text{ rt, 1 h} \end{array} \begin{array}{c} \text{10}\\ \text{AgNO}_3, (\text{NH}_4)_2\text{S}_2\text{O}_8\\ \text{MeCN/H}_2\text{O}, 85 \, ^{\circ}\text{C}\\ \text{3 h} \end{array} \begin{array}{c} \text{[137]}\\ \text{95 10\%} \end{array}$$

these conditions, the γ -picoline radical preferentially reacts with itself (or the starting pyridylacetic acid) rather than with menadione (10) [137].

The Kochi-Anderson reaction can also be used for the alkylation of menadione with bromoalkyl-substituted carboxylic acids as described by Terasaki and co-workers [136] and Liu

and co-workers [140]. The alkylation products **96a,b** were obtained in good yields, demonstrating a greater resistance of this type of compounds to the workup and purification processes, when compared to derivatives with other types of long chain acids and substituents in the methyl terminal (Scheme 32).

An example for the alkylation of menadione by the Kochi–Anderson method with a complex carboxylic acid was described by Goebel and Barany in the synthesis of **98**, a human metabolite formed from vitamin K with biological activity [141]. In this work, a carboxylic acid derived from diethyl methylmalonate was used, and the product **98** was obtained in 56% yield, which is comparable to yields found with short chain diacids, as previously mentioned (Scheme 33).

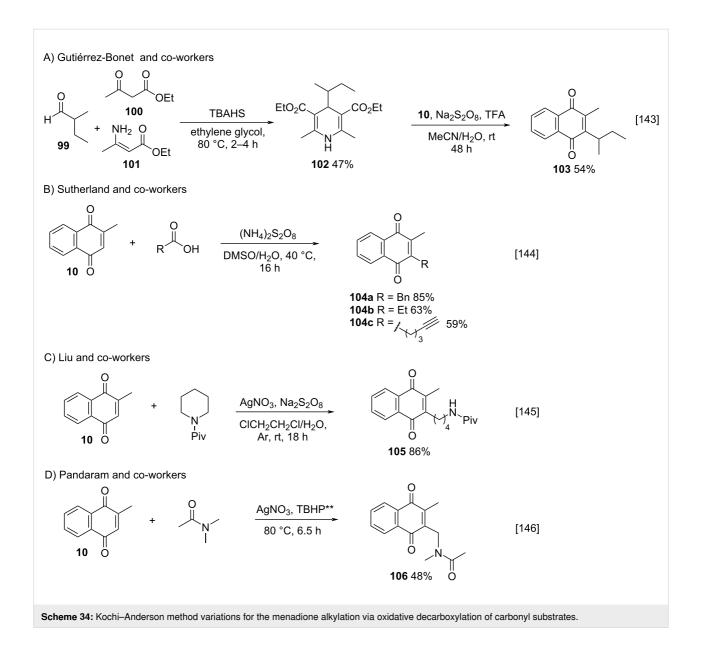
Some variations of the Kochi–Anderson method have also been described in the literature, such as the methods reported by Gutiérrez–Bonet and co-workers [143] and Sutherland and co-workers [144], both with no use of silver as radical generator, the method reported by Liu and co-workers [145], using cyclic amines as alkylating agents, and the method described by Pandaram [146], using silver salt and TBHP as oxidizing agent.

In the reaction described by Gutiérrez-Bonet [143], the authors used persulfate as oxidizing agent, trifluoroacetic acid as menadione-activator and 1,4-dihydropyridine **102**, which was readily prepared from aldehyde **99** in one step, to achieve homolysis [143]. The advantage of the method is the absence of a noble metal salt and milder reaction conditions; however, it presents also some disadvantages such as a longer reaction time and the

need of pre-functionalized aldehydes. In turn, Sutherland and co-workers [144] described the menadione (10) alkylation with persulfate, alkylcarboxylic acids, and dimethyl sulfoxide (DMSO) at 40 °C, in a process without metals, photocatalysts, light or pre-functionalized alkyl substrates (Scheme 34B). This study demonstrated that the silver salt is not essential for the alkylation to occur. Compounds 104a-c were obtained in good yields, similar to the yields obtained by the original process (Scheme 34B). The success of this procedure is related to the easier decomposition of persulfate to form sulfate radicals (SO₄•-) in DMSO. In their work, Liu and co-workers described the access to distal aminoalkyl-substituted menadione 105 by silver-catalyzed site-selective ring-opening and C-C-bond functionalization of the cyclic amine in good yields (Scheme 34C) [145]. This approach overcame other methods' problems, such as multi-stage transformations or the use of short-chain amino acids. Recently, Pandaram and co-workers demonstrated that menadione (10) also could be amidoalkylated using silver(I) nitrate – *tert*-butyl hydroperoxide in *N*,*N*-dimethylacetamide as alkylating agent and solvent (Scheme 34D) [146]. This was the first reported synthesis of several amidoalkylated quinones that were obtained in moderate and good yields. No pre-functionalization of starting materials was required, and only nonhazardous reagents were used.

Despite its widespread use, the Kochi–Anderson method for the alkylation of menadione has as its main limitation the exclusive use of carboxylic acids as alkyl chain source, thus restricting the substrate scope. In this context, several approaches have been developed to replace carboxylic acids with a different radical

HO₂C
$$\underset{n}{\longleftrightarrow}$$
 Br $\underset{n}{\longleftrightarrow}$ AgNO₃, (NH₄)₂S₂O₈ MeCN/H₂O, reflux 2 h 96a $n = 15$ 79% [136] 96b $n = 10$ 56% [140]



source, such as alkyl halides, alkanes or activated alkenes, in conjunction with transition-metal catalysis or metal-free processes.

In a work published by Baral and co-workers, the unprecedented Csp²–Csp³ alkylation of menadione (**10**) with medium and large-size cyclic alkanes was achieved by the combination of copper(II) triflate and *tert*-butyl hydroperoxide (Scheme 35) [147]. The products **107a**–**d** were obtained in 58–64% yield range. This is a one-step process with no need of activated alkylating substrates. In turn, Li and Yang also reported the use of copper as alkylation promoter without the use of an oxidizing agent. However, in their method functionalized alkyl halides and high temperatures were used, to obtain compound **108b** in 84% yield (Scheme 35) [148].

Gu and co-workers explored the reactivity of cyclobutanone oximes as alkylation substrates for menadione (10), in a reaction catalyzed by nickel and oxidizing agents free [149]. With this method, it was possible to obtain cyanoalkylated compounds 110 in excellent yields, from a wide range of cyclobutanone oximes with aryl, benzyl or alkyl groups (Scheme 36).

Recently, the use of iron as catalyst in the alkylation reaction of menadione via free radicals has also been reported [150,151]. Iron has many advantages, such as its high abundance, low-cost, and low toxicity to humans and the environment, that make iron quite attractive to be used in synthetic processes. Liu and co-workers described a radical alkylation of menadione (10) with an olefin as the radical precursor, during the iron(III)-mediated C–H conversion of quinones with non-activated

$$\begin{array}{c} & & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & &$$

alkenes (Scheme 37A) [150]. In their work, Li and Shen used a general iron-catalyzed protocol for the synthesis of alkylated quinones, including menadione, with alkyl bromides as alkylating reagents, with a broad substrate scope, densely functional group tolerance, and good yields (Scheme 37B) [151]. A

common advantage to both methods is the absence of oxidizing agents to generate the radical species. However, the method described by Li requires harsher conditions, such as temperatures above 100 $^{\circ}$ C and reaction times longer than 24 hours, when compared to the method developed by Liu.

Other menadione alkylation methods, by free radicals, which do not involve the Kochi–Anderson procedure [76], its adaptations, or catalysis mediated by transition metals have also been reported. These methods include alkylation by thermal decomposition of diacid peroxides [152], the use of organotellurium compounds [153,154], and perfluoroalkylation from perfluoroalkyl radicals [155].

Boudalis and co-workers reported a selective alkylation method for menadione with radicals generated from the thermal decomposition of diacyl peroxides **113a,b** (Scheme 38) [152], as an adaptation of the route developed by Fieser [156]. Yamago and co-workers also described the alkylation of menadione, during the synthesis mediated by radicals of quinones substituted with organotellurium compounds (Scheme 38) [153,154]. A very specific type of radical alkylation of menadione was described by Antonietti [75] for the synthesis of perfluoronaphtho-quinones from perfluoroalkyl radicals, with or without alkenes presence (Scheme 38). In these studies, it was observed that in the absence of alkenes, perfluoroalkylation occurs directly on the menadione C-3 carbon, generating product **115**. On the other hand, in the presence of a terminal alkene, perfluoroalkylation occurs first on the alkene and then the obtained free

radical reacts with menadione, leading to products **116a,b**. The same results were obtained for Sansotera and co-workers who used perfluorodiacyl peroxides as alkylating agents [155].

In contrast to alkylation, the radical acylation of menadione is not very common. In one of the few reports that exclusively is dedicated to the study of the radical acylation of menadione, Waske and co-workers described a versatile method for the preparation of photoacylated menadione products from aldehydes using free radical conditions [157], also called photo-Friedel–Crafts acylation, a term introduced by Oelgemöller [158-161]. According to this protocol, a mixture of menadione (10) and an aliphatic or aromatic aldehyde in excess, in benzene, is irradiated under direct excitation conditions (λ_{max} = 419 nm). The acylated menadione derivatives 117a–f were obtained in moderate yields (Scheme 39).

Westwood and co-workers developed a decarboxylative acylation for the direct C–H acylation and carbamoylation of heterocycles, including menadione, under metal-, photocatalyst, and light-free conditions [162] based on the method developed by Minisci [142]. The reaction occurs between menadione (10) and acyl radicals derived from α-keto acids and

alkyl-substituted oxamic acid in the presence of persulfate in DMSO [142], providing acylated products **117d**,**e** and **118**, respectively, in moderate yields (Scheme 40).

Borah and co-workers described a methodology for the free radical benzoylation of 2-substituted-1,4-naphthoquinones, such as menadione, as an alternative approach to the use of organometallic reagents [163]. Considering some limitations of the

methods commonly used in acylation reactions via free radicals, such as the use of metallic catalysts, long reaction times, and acyl/benzoyl source, in Borah's work the acylation of menadione via benzoyl radicals was performed using the metal-free tetra-*n*-butylammonium iodide/*tert*-butyl hydroperoxide (TBAI/TBHP) system [163]. Under optimized conditions the three benzoylated compounds **119a–c** were obtained with 37–43% yield (Scheme 41). The modest yields of the menadione deriva-

tives, when compared to halogenated derivatives, can be explained by the interaction of the methyl group with the TBAI/TBHP system [163].

1.4-Addition reactions

Menadione behaves like a typical Michael acceptor in the presence of nucleophiles, such as amines and thiols, and the addition of the nucleophile occurs at the C-3 carbon, which is less steric impeded and more electrophilic [164]. The formed adduct is a naphthohydroquinone which is then oxidized, regenerating the quinone structure, in a process that can be spontaneous or induced by oxidizing agents depending on the reaction conditions [131].

The best-known method for the addition of nucleophiles to menadione was developed by Kallmayer [165]. In this method, initially proposed for the Michael-type addition of ethanolamine, menadione and an amine were solubilized in benzene and the reaction was maintained at room temperature (rt), leading to the amino-substituted menadione 120 in moderate yield (Scheme 42) [165]. Afterwards, ethanol/dichloromethane mixtures were used, as they increased the solubility of both menadione and the synthesized products.

In recent years, the Kallmayer method [165] has been the most common approach to promote the addition of amines to menadione, proving to be quite efficient and robust [131,166-174]. Salmon-Chemin and co-workers described the synthesis of amino-substituted menadione derivatives using polyalkylamines to form the adducts 3-polyaminomenadione and 3,3'-polyamino-bis(menadione) [131]. Several reaction conditions were employed to obtain products 121a-d or 122a-d, such as the amount of polyamine and reaction time, requiring 5.0 equivalents of polyamine and 1 hour of reaction to form 121a-d and 0.5 equivalents of polyamine and 3 days of reaction to obtain 122a-d. The yields of each product type were also different, with 121a-d being obtained in moderate to good yields and 122a-d in low to moderate yields (Scheme 43).

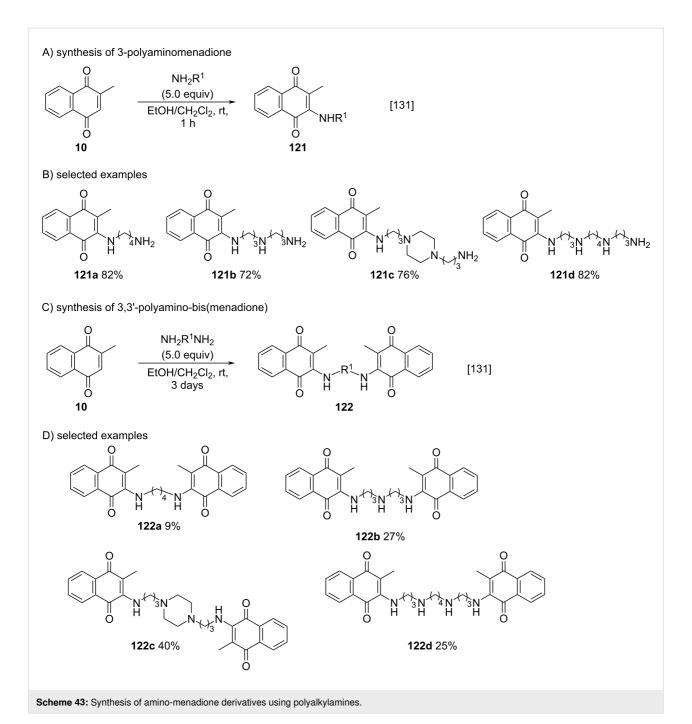
Karunan's group [166], Wang's group [167] and Li's group [168] used the Kallmayer method to prepare aminomenadiones **123** and **124** through the addition of amines containing linear,

cyclic, and branched aliphatic chains (Scheme 44). Jing and co-workers, in turn, applied this methodology for the addition of propargylamine to 10, to form the propargylamino-substituted product 125 (Scheme 44) [169]. Bowen and co-workers also carried out the reaction between amino alcohols and 10, in ethanol at rt, demonstrating that this method remains the most interesting option, even after 30 years, for the Michael-type addition of amino alcohols to 10, yielding products of type 126 (Scheme 44) [170]. In all these cases, the yields of the adducts varied according to the nature of the respective precursor amine.

Other authors have also succeeded with the Michael-type addition of complex amines to menadione (10), containing arenes and heteroarenes as substituents in the aliphatic chain. In the work developed by Namsa-aid and Ruchirawat, homoveratrylamine (127) was used as nucleophile [171], while Zacconi and co-workers applied benzyl- or phenethylamines 128 [172]. However, the protocols required different solvents and reaction temperatures (Scheme 45). In the works developed by Wu and co-workers [173] and Patil and co-workers [174], amines containing heterocycles such as pyridine 129 or thiophene 130 were used as nucleophiles, to provide the corresponding compounds 132a,b and 133a,b, respectively, in good yields (Scheme 45). A possible reason for the higher yields obtained by Patil compared to those reported by Wu could have been an additional sonication step after the partial menadione dissolution in methanol, increasing the solubility of this reac-

The method developed by Kallmayer [165] also supports the use of amino acids, as described by Ge and co-workers [175]. They reported a Michael-type addition of different natural α -amino acids **134a–e** to menadione (**10**). However, the only product that formed with a measurable yield was the L-glycine derivative **135a** (23% yield) (Scheme 46).

A variation of the Kallmayer method [165] was described by Mital and co-workers, which involved the addition of several amines to menadione in the presence of an inorganic base (K₂CO₃) to afford products **136** [176]. The compounds were obtained in moderate yields, in a very short reaction time, when compared to the original method (Table 6).



Besides to the Kallmayer method [165], many other protocols for the Michael-type addition of amines to menadione have been developed and described. These protocols differ, basically, in the used solvent and the obtained adduct yields, when compared to the original method. With regard to solvents, the use of hot water [177], diethyl ether [178], acetonitrile at 45 °C [179], the amino reagent [180], and pure dichloromethane [181-183] were reported as solvents. Conditions that resulted in reduced menadione solubility, may explain the drop of the reaction yield.

A particular method that deserves to be highlighted was described by Sharma and co-workers. It consisted in the addition of amines to menadione (10), via a solvent-free Michael-type addition, using silica-supported perchloric acid (HClO₄-SiO₂) and ultrasound irradiation (Scheme 47) [184] and provided the adducts 137a,b in good yields in up to 20 minutes. This method is highly efficient and can be considered environmentally friendly when compared to the previously described protocols, which used solvents such as dichloromethane or required longer reaction times.

$$\begin{array}{c} \text{H}_2\text{N} \\ \text{OCH}_3 \\ \text{EtOH, rt, 24 h} \\ \text{O} \\ \text{H}_2\text{N} \\ \text{O} \\ \text{128} \\ \text{50 °C, 4 h} \\ \text{H}_2\text{N} \\ \text{129 N} \\ \text{129 N} \\ \text{MeOH, rt, 12 h} \\ \text{MeOH, rt, 12 h} \\ \text{Scheme 45: Selected examples of Michael-type addition of complex amines to menadione (10).} \\ \end{array} \begin{tabular}{c} \text{OMe} \\ \text{OMe$$

The Michael-type addition reaction of menadione could also be carried out using the appropriate indole to obtain indolylnaph-thoquinones 139 or indolylnaphthalene-1,4-diols 140 in excel-

lent yields. In this regard, Yadav and co-workers [185] reported the indium(III) bromide catalyzed conjugate addition of 2-methylindole (138a) to 10 to obtain the product 139a

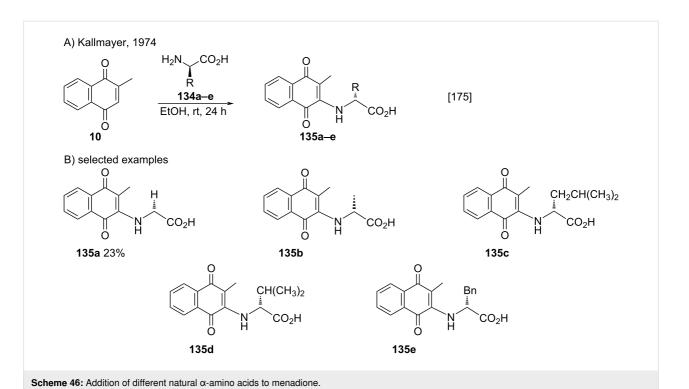


Table 6: Reaction conditions for the addition of amines to menadione by Mital and co-workers.

Entry	Compounds	R^1 , R^2	Yield (%)	
1	136a	-H, -CH ₂ Ph	45	
2	136b	-H, -CH ₂ CO ₂ H	52	
3	136c	-H, -CH ₃	45	
4	136d	-H, -C(CH ₂) ₅	45	
5	136e	-H, -Ph	44	
6	136f	-CH ₃ , -CH ₂ Ph	39	

Scheme 47: Michael-type addition of amines to menadione using silica-supported perchloric acid.

(Scheme 48). The same group also published a microwave-accelerated solvent- and catalyst-free synthesis of 3-indolylhydroquinones **140a**,**b** (Scheme 48) [186].

Tanoue and co-workers [187] described the synthesis of indolylnaphthoquinone **142** using a Michael-type addition reaction of **10** with 3-iodoindole (**141**) (Scheme 49). The reaction was carried out in acetic acid at rt for 4 days resulting in 2-methyl-3-(3-indolyl)-1,4-naphthoquinone (**142**) in 62% yield. When the reaction was carried out in the presence of cesium carbonate in acetonitrile at rt for 1 day, the product **142** was obtained in 23% yield.

An ecofriendly approach to this methodology was reported by Escobeto-González and co-workers [188], who investigated three different non-conventional reaction activation modes: microwave (MW) and near-infrared irradiation (NIR) as well as high-speed ball milling (HSBM) (Scheme 50). The alternative approaches were compared with typical mantle heating conditions (MH) and all methods were carried out under solvent-free conditions in the presence of Tonsil Actisil FF (TAFF) as a green catalyst. The best results were obtained using NIR at 121 °C for 10 min furnishing product 142 in 51% yield. According to the authors, the reaction mechanism proceeds via a classical Michael-type addition of indole (138b) to 10, assisted

by an oxygen interaction of a carbonyl group with the Lewis acidic sites of TAFF, followed by in situ oxidation to obtain the product **142** [188].

Thiols can also be employed as nucleophiles in the Michael-type addition to menadione. This approach is quite similar to the biological processes that naturally run between menadione and cysteine derivatives. Chen and co-workers described the synthesis of menadione analogues functionalized with thiols [189], using an adaptation of the already described method by Borovkov [190], where menadione was reacted with thioalcohols, thioethers, and ethers. The reaction occurs between menadione (10) and the respective thiols using copper sulfate pentahydrate as catalyst, in ethanol, at rt for 24 hours, to furnish products 143 in 30–50% yield (Scheme 51).

Scheme 51: Synthesis of menadione analogues functionalized with thiols.

Singh and co-workers described the synthesis of bis-menadione derivatives through Michael-type addition of different dithiols and menadione used in excess [191]. The reaction proceeded in dichloromethane at rt for 5 h, furnishing products **144** in

65–84% yield. These results make this method very effective for the synthesis of menadione-derived symmetrical molecules (Scheme 52).

Mercaptoacetic (145a) and mercaptopropanoic (145b) acids were also used as nucleophiles in Michael-type addition reactions to menadione, as described by Garbay's group [178,192] and Singh's group [193]. In the method developed by Garbay and co-workers, that was based on Tamure and co-workers' methodology for the addition of 2-mercaptoethanol to menadione (10) [194], the addition reaction occurred in the presence of DBU and ethyl ether, providing products 146a and 146b in 67% and 20% yield, respectively (Scheme 53). In turn, the method applied by Singh required milder conditions and was more effective, using only menadione (10), the nucleophile 145a and methanol as a solvent to synthesize the addition product 146a in 71% yield (Scheme 53).

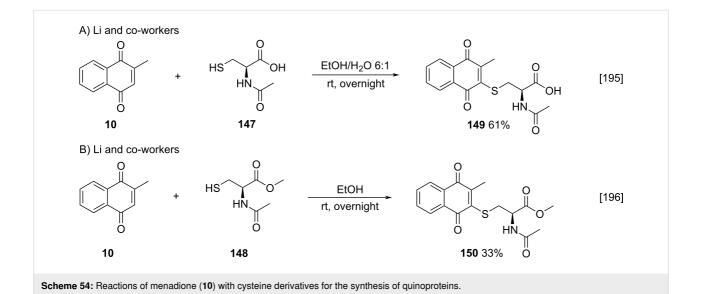
In a study on the introduction of quinones into proteins to obtain quinoproteins, Li and co-workers reported a methodology for the reaction of menadione (10) with L-cysteine [195]. The reaction of 10 and N-acetyl-L-cysteine (147) occurs in an ethanol/water mixture, at room temperature overnight, to furnish the product 149 in 61% yield (Scheme 54A). Li and co-workers, in a similar study, reacted menadione (10) with N-acetylcysteine methyl ester (148) to obtain compound 150 in 33% yield [196] (Scheme 54B).

Kumar and co-workers focused their work on the synthesis of menadione-glutathione conjugates by Michael-type addition

DBU, ether, rt, 23 h (from 145a,b) or MeOH, rt, 2 h (from 145a) 145b
$$n = 2$$

DBU, ether, rt, 23 h (from 145a,b) or MeOH, rt, 2 h (from 145a) 146a $n = 1$ 67% (from 145a) [178,192] 146a $n = 1$ 71% (from 145a) [193] 146b $n = 2$ 20% (from 145b) [178,192]

Scheme 53: Mercaptoalkyl acids as nucleophiles in Michael-type addition reaction to menadione.



reaction [197], based on the method of Nickerson and co-workers [198]. In this method an aqueous solution of L-glutathione (151) was treated with a menadione (10) solution in DMSO/ethanol at 0 °C for 1 h, then diluted with ethyl

acetate, and stirred at room temperature overnight (Scheme 55). The menadione-glutathione conjugate 152 was separated by filtration, being obtained in 33% yield without further purification.

Conclusion

Organic synthesis is the most active subarea of chemistry that uses structural models to plan and develop new products and new reactions. The use of abundant natural products, even if produced by synthetic means, is one of the central strategies in research for the development of new bioactive compounds. Since the first reports on the biological activities of menadione and the development of methods for its preparation on an indus-

Scheme 55: Synthesis of menadione-glutathione conjugate 152 by Michael-type addition.

trial scale, various compounds have been synthesized using this important 1,4-naphthoquinone. As a structural platform, this commercially available organic compound offers multiple possibilities for chemical modification in a search for new hit compounds that can become a new drug. This review represents an update and overview of aspects of menadione chemistry, synthetic opportunities, and its derivatives. The number of applications of menadione highlighted in this review clearly demonstrates the central role this compound plays in synthetic organic and medicinal chemistry.

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References

- de Carvalho da Silva, F.; Francisco Ferreira, V. Curr. Org. Synth. 2016, 13, 334–371. doi:10.2174/1570179412666150817220343
- Goodwin, T. W.; Mercer, E. I. Introduction to plant Biochemistry; Pergamon Press, 1972.
- Kashket, E. R.; Brodie, A. F. J. Biol. Chem. 1963, 238, 2564–2570. doi:10.1016/s0021-9258(19)68006-3
- Ziegler, D. M. Am. J. Clin. Nutr. 1961, 9, 43–49. doi:10.1093/ajcn/9.4.43
- Castro, F. A. V.; Mariani, D.; Panek, A. D.; Eleutherio, E. C. A.; Pereira, M. D. *PLoS One* **2008**, *3*, e3999. doi:10.1371/journal.pone.0003999
- da Silva Júnior, E. N.; de Souza, M. C. B. V.; Fernandes, M. C.; Menna-Barreto, R. F. S.; Pinto, M. d. C. F. R.; de Assis Lopes, F.; de Simone, C. A.; Andrade, C. K. Z.; Pinto, A. V.; Ferreira, V. F.; de Castro, S. L. Bioorg. Med. Chem. 2008, 16, 5030–5038. doi:10.1016/j.bmc.2008.03.032
- Coates, C. S.; Ziegler, J.; Manz, K.; Good, J.; Kang, B.; Milikisiyants, S.; Chatterjee, R.; Hao, S.; Golbeck, J. H.; Lakshmi, K. V. *J. Phys. Chem. B* 2013, *117*, 7210–7220. doi:10.1021/jp401024p
- Hiraishi, A. J. Biosci. Bioeng. 1999, 88, 449–460. doi:10.1016/s1389-1723(00)87658-6
- Hook, I.; Mills, C.; Sheridan, H. Bioactive Naphthoquinones from Higher Plants. Studies in Natural Products Chemistry, 1st ed.; Elsevier, 2014; Vol. 41, pp 119–160. doi:10.1016/b978-0-444-63294-4.00005-x
- Combs, G. F., Jr.; McClung, J. P. Vitamin K. The Vitamins, Fundamental Aspects in Nutrition and Health; Academic Press: London, UK, 2017; pp 243–265. doi:10.1016/b978-0-12-802965-7.00009-5

- Ivanova, D.; Zhelev, Z.; Getsov, P.; Nikolova, B.; Aoki, I.; Higashi, T.; Bakalova, R. *Redox Biol.* 2018, 16, 352–358. doi:10.1016/j.redox.2018.03.013
- Thijssen, H. H. W.; Vervoort, L. M. T.; Schurgers, L. J.; Shearer, M. J. Br. J. Nutr. 2006, 95, 260–266. doi:10.1079/bjn20051630
- Hassan, G. S. Menadione. In *Profiles of Drug Substances, Excipients and Related Methodology;* Brittain, H. G., Ed.; Academic Press: San Diego, CA, USA, 2013; pp 227–313. doi:10.1016/b978-0-12-407691-4.00006-x
- Fiore, L. D.; Scola, M. A.; Cantillon, C. E.; Brophy, M. T. *J. Thromb. Thrombolysis* 2001, *11*, 175–183. doi:10.1023/a:1011237019082
- Nutter, L. M.; Ann-Lii, C.; Hsiao-Ling, H.; Ruey-Kun, H.; Ngo, E. O.; Tsang-Wu, L. *Biochem. Pharmacol.* 1991, 41, 1283–1292. doi:10.1016/0006-2952(91)90099-q
- Kishore, C.; Sundaram, S.; Karunagaran, D. Chem.-Biol. Interact. 2019, 309, 108725. doi:10.1016/j.cbi.2019.108725
- Majiene, D.; Kuseliauskyte, J.; Stimbirys, A.; Jekabsone, A. *Nutrients* 2019, 11, 1294. doi:10.3390/nu11061294
- Zhang, W.; Negoro, T.; Satoh, K.; Jiang, Y.; Hashimoto, K.; Kikuchi, H.; Nishikawa, H.; Miyata, T.; Yamamoto, Y.; Nakano, K.; Yasumoto, E.; Nakayachi, T.; Mineno, K.; Satoh, T.; Sakagami, H. Anticancer Res. 2001, 21, 3439–3444.
- Gilloteaux, J.; Jamison, J. M.; Lorimer, H. E.; Jarjoura, D.;
 Taper, H. S.; Calderon, P. B.; Neal, D. R.; Summers, J. L. *Tissue Cell* 2004, 36, 197–209. doi:10.1016/j.tice.2004.01.006
- Baran, I.; Ionescu, D.; Filippi, A.; Mocanu, M. M.; Iftime, A.; Babes, R.; Tofolean, I. T.; Irimia, R.; Goicea, A.; Popescu, V.; Dimancea, A.; Neagu, A.; Ganea, C. Leuk. Res. 2014, 38, 836–849. doi:10.1016/j.leukres.2014.04.010
- Gilloteaux, J.; Jamison, J. M.; Neal, D. R.; Loukas, M.; Doberzstyn, T.; Summers, J. L. *Ultrastruct. Pathol.* 2010, 34, 140–160. doi:10.3109/01913121003662304
- Gilloteaux, J.; Jamison, J. M.; Neal, D.; Summers, J. L. *Ultrastruct. Pathol.* 2014, 38, 116–140. doi:10.3109/01913123.2013.852645
- Sreelatha, T.; Kandhasamy, S.; Dinesh, R.; Shruthy, S.; Shweta, S.; Mukesh, D.; Karunagaran, D.; Balaji, R.; Mathivanan, N.; Perumal, P. T. *Bioorg. Med. Chem. Lett.* 2014, 24, 3647–3651. doi:10.1016/j.bmcl.2014.04.080
- Tintino, S. R.; Oliveira-Tintino, C. D. M.; Campina, F. F.; Weslley Limaverde, P.; Pereira, P. S.; Siqueira-Junior, J. P.; Coutinho, H. D. M.; Quintans-Júnior, L. J.; da Silva, T. G.; Leal-Balbino, T. C.; Balbino, V. Q. Med. Chem. Res. 2018, 27, 261–267. doi:10.1007/s00044-017-2063-y
- Andrade, J. C.; Morais Braga, M. F. B.; Guedes, G. M. M.; Tintino, S. R.; Freitas, M. A.; Quintans, L. J., Jr.; Menezes, I. R. A.; Coutinho, H. D. M. Saudi J. Biol. Sci. 2017, 24, 59–64. doi:10.1016/j.sjbs.2015.09.004
- Lee, M. H.; Yang, J. Y.; Cho, Y.; Woo, H. J.; Kwon, H. J.; Kim, D. H.;
 Park, M.; Moon, C.; Yeon, M. J.; Kim, H. W.; Seo, W.-D.; Kim, S.-H.;
 Kim, J.-B. *Int. J. Mol. Sci.* 2019, 20, 1169. doi:10.3390/ijms20051169
- Gyetvai, Ä.; Emri, T.; Fekete, A.; Varga, Z.; Gazdag, Z.; Pesti, M.;
 Belágyi, J.; Emõdy, L.; Pócsi, I.; Lenkey, B. FEMS Yeast Res.
 2007, 7, 265–275. doi:10.1111/j.1567-1364.2006.00179.x
- Jakab, Á.; Emri, T.; Sipos, L.; Kiss, Á.; Kovács, R.; Dombrádi, V.; Kemény-Beke, Á.; Balla, J.; Majoros, L.; Pócsi, I. *J. Basic Microbiol.* 2015, 55, 973–981. doi:10.1002/jobm.201400903

- Cyrklaff, M.; Srismith, S.; Nyboer, B.; Burda, K.; Hoffmann, A.; Lasitschka, F.; Adjalley, S.; Bisseye, C.; Simpore, J.; Mueller, A.-K.; Sanchez, C. P.; Frischknecht, F.; Lanzer, M. Nat. Commun. 2016, 7, 13401. doi:10.1038/ncomms13401
- Afianda, F.; Wiraswati, H. L.; Alisjahbana, B.; Fauziah, N. IOSR J. Pharm. Biol. Sci. 2020, 15, 56–60.
- Müller, T.; Johann, L.; Jannack, B.; Brückner, M.; Lanfranchi, D. A.;
 Bauer, H.; Sanchez, C.; Yardley, V.; Deregnaucourt, C.; Schrével, J.;
 Lanzer, M.; Schirmer, R. H.; Davioud-Charvet, E. J. Am. Chem. Soc.
 2011, 133, 11557–11571. doi:10.1021/ja201729z
- Ehrhardt, K.; Deregnaucourt, C.; Goetz, A.-A.; Tzanova, T.; Gallo, V.;
 Arese, P.; Pradines, B.; Adjalley, S. H.; Bagrel, D.; Blandin, S.;
 Lanzer, M.; Davioud-Charvet, E. *Antimicrob. Agents Chemother.* 2016, 60, 5146–5158. doi:10.1128/aac.02975-15
- Cristina Desoti, V.; Lazarin-Bidóia, D.; Martins Ribeiro, F.;
 Cardoso Martins, S.; da Silva Rodrigues, J. H.; Ueda-Nakamura, T.;
 Vataru Nakamura, C.; Farias Ximenes, V.; de Oliveira Silva, S.
 PLoS One 2015, 10, e0144033. doi:10.1371/journal.pone.0144033
- Kapadia, G. J.; Soares, I. A. O.; Rao, G. S.; Badoco, F. R.; Furtado, R. A.; Correa, M. B.; Tavares, D. C.; Cunha, W. R.; Magalhães, L. G. Acta Trop. 2017, 167, 163–173. doi:10.1016/j.actatropica.2016.12.001
- Gant, T. W.; Ramakrishna Rao, D. N.; Mason, R. P.; Cohen, G. M. *Chem.-Biol. Interact.* 1988, 65, 157–173. doi:10.1016/0009-2797(88)90052-x
- Klotz, L.-O.; Hou, X.; Jacob, C. Molecules 2014, 19, 14902–14918. doi:10.3390/molecules190914902
- Jordão, A. K.; Vargas, M. D.; Pinto, A. C.; da Silva, F. d. C.; Ferreira, V. F. RSC Adv. 2015, 5, 67909–67943. doi:10.1039/c5ra12785h
- Schieber, A. J. Agric. Food Chem. 2018, 66, 13051–13055. doi:10.1021/acs.jafc.8b05215
- D'Odorico, A.; Sturniolo, G. C.; Bilton, R. F.; Morris, A. I.;
 Gilmore, I. T.; Naccarato, R. Carcinogenesis 1997, 18, 43–46.
 doi:10.1093/carcin/18.1.43
- Sun, J.-S.; Tsuang, Y.-H.; Huang, W.-C.; Chen, L.-T.; Hang, Y.-S.;
 Lu, F.-J. Cell. Mol. Life Sci. 1997, 53, 967–976.
 doi:10.1007/s000180050118
- Chen, R.; Pignatello, J. J. Environ. Sci. Technol. 1997, 31, 2399–2406. doi:10.1021/es9610646
- Keyer, K.; Imlay, J. A. Proc. Natl. Acad. Sci. U. S. A. 1996, 93, 13635–13640. doi:10.1073/pnas.93.24.13635
- Brynildsen, M. P.; Winkler, J. A.; Spina, C. S.; MacDonald, I. C.; Collins, J. J. Nat. Biotechnol. 2013, 31, 160–165. doi:10.1038/nbt.2458
- Imlay, J. A. Curr. Opin. Microbiol. 2015, 24, 124–131. doi:10.1016/j.mib.2015.01.004
- Lushchak, O. V.; Bayliak, M. M.; Korobova, O. V.; Levine, R. L.; Lushchak, V. I. *Redox Rep.* **2009**, *14*, 214–220. doi:10.1179/135100009x12525712409454
- Zaccaria, M.; Ludovici, M.; Sanzani, S. M.; Ippolito, A.; Cigliano, R. A.; Sanseverino, W.; Scarpari, M.; Scala, V.; Fanelli, C.; Reverberi, M. Toxins 2015, 7, 4315–4329. doi:10.3390/toxins7104315
- 47. Fieser, L. F. *J. Biol. Chem.* **1940**, *133*, 391–396. doi:10.1016/s0021-9258(18)73319-x
- Li, L.; Elliott, W. H. J. Am. Chem. Soc. 1952, 74, 4089–4090. doi:10.1021/ja01136a036
- Yamazaki, S. Tetrahedron Lett. 2001, 42, 3355–3357.
 doi:10.1016/s0040-4039(01)00432-4

- Yamaguchi, S.; Inoue, M.; Enomoto, S. Chem. Lett. 1985, 14, 827–828. doi:10.1246/cl.1985.827
- Adam, W.; Herrmann, W. A.; Lin, J.; Saha-Möller, C. R.; Fischer, R. W.; Correia, J. D. G. *Angew. Chem.* 1994, 106, 2545–2546. doi:10.1002/ange.19941062322
- Narayanan, S.; Murthy, K. V. V. S. B. S. R.; Reddy, K. M.; Premchander, N. Appl. Catal., A 2002, 228, 161–165. doi:10.1016/s0926-860x(01)00962-0
- Kowalski, J.; Płoszyńska, J.; Sobkowiak, A. Catal. Commun. 2003, 4, 603–608. doi:10.1016/j.catcom.2003.09.003
- Yan, T.; Hong, M.; Niu, L.; Jiang, F.; Xiao, G. Res. Chem. Intermed.
 2012. 38. 1839–1846. doi:10.1007/s11164-012-0507-z
- Möller, K.; Wienhöfer, G.; Schröder, K.; Join, B.; Junge, K.; Beller, M. Chem. – Eur. J. 2010, 16, 10300–10303. doi:10.1002/chem.201001429
- Narender, N.; Suresh Kumar Reddy, K.; Krishna Mohan, K. V. V.;
 Kulkarni, S. J.; Koeckritrz, A.; Raghavan, K. V. J. Porous Mater. 2011, 18, 337–343. doi:10.1007/s10934-010-9383-3
- Uruş, S.; Keleş, M.; Serindağ, O.
 Phosphorus, Sulfur Silicon Relat. Elem. 2010, 185, 1416–1424.
 doi:10.1080/10426500903061533
- Uruş, S.; Çaylar, M.; Karteri, İ. Chem. Eng. J. 2016, 306, 961–972. doi:10.1016/i.cei.2016.08.009
- Çapan, A.; Uruş, S.; Sönmez, M. J. Saudi Chem. Soc. 2018, 22, 757–766. doi:10.1016/j.jscs.2017.12.007
- Chang, S.; An, H.; Chen, Y.; Hou, Y.; Zhang, J.; Zhu, Q. ACS Appl. Mater. Interfaces 2019, 11, 37908–37919. doi:10.1021/acsami.9b14928
- Shimanskaya, E. I.; Sulman, E. M.; Doluda, V. Y. *Catal. Sustainable Energy* 2015, 2, 28–32. doi:10.1515/cse-2015-0008
- Mamchur, A. V.; Galstyan, G. A. Russ. J. Org. Chem. 2004, 40, 1779–1781. doi:10.1007/s11178-005-0098-x
- 63. Sheldon, R. A. Top. Curr. Chem. 1993, 164, 29-30.
- Herrmann, W. A.; Haider, J. J.; Fischer, R. W. J. Mol. Catal. A: Chem. 1999, 138, 115–121. doi:10.1016/s1381-1169(98)00133-2
- Kholdeeva, A. O.; Zalomaeva, O. V.; Shmakov, A. N.;
 Melgunov, M. S.; Sorokin, A. B. J. Catal. 2005, 236, 62–68.
 doi:10.1016/j.jcat.2005.09.022
- Minisci, F.; Citterio, A.; Vismara, E.; Fontana, F.; De Bernardinis, S.;
 Correale, M. J. Org. Chem. 1989, 54, 728–731.
 doi:10.1021/jo00264a047
- Selvaraj, M.; Kandaswamy, M.; Park, D. W.; Ha, C. S. Catal. Today 2010, 158, 377–384. doi:10.1016/j.cattod.2010.04.041
- Strukul, G.; Somma, F.; Ballarini, N.; Cavani, F.; Frattini, A.;
 Guidetti, S.; Morselli, D. *Appl. Catal.*, A 2009, 356, 162–166.
 doi:10.1016/j.apcata.2008.12.041
- Selvaraj, M.; Park, D.-W.; Kim, I.; Kawi, S.; Ha, C. S. Dalton Trans.
 2012, 41, 9633–9638. doi:10.1039/c2dt31096a
- Shimanskaya, E.; Doluda, V.; Sulman, M.; Matveeva, V.; Sulman, E. Chem. Eng. J. 2014, 238, 206–209. doi:10.1016/j.cej.2013.07.087
- Matveev, K. I.; Odyakov, V. F.; Zhizhina, E. G. J. Mol. Catal. A: Chem. 1996, 114, 151–160. doi:10.1016/s1381-1169(96)00313-5
- Kholdeeva, O. A.; Zalomaeva, O. V.; Sorokin, A. B.; Ivanchikova, I. D.; Della Pina, C.; Rossi, M. Catal. Today 2007, 121, 58–64. doi:10.1016/j.cattod.2006.11.011
- Kholdeeva, O. A.; Ivanchikova, I. D.; Zalomaeva, O. V.; Sorokin, A. B.; Skobelev, I. Y.; Talsi, E. P. J. Phys. Chem. B 2011, 115, 11971–11983. doi:10.1021/jp2055975

- Zalomaeva, O. V.; Kholdeeva, O. A.; Sorokin, A. B. C. R. Chim. 2007, 10, 598–603. doi:10.1016/j.crci.2007.02.001
- Antonietti, F.; Gambarotti, C.; Mele, A.; Minisci, F.; Paganelli, R.;
 Punta, C.; Recupero, F. Eur. J. Org. Chem. 2005, 4434–4440.
 doi:10.1002/eioc.200500314
- Anderson, J. M.; Kochi, J. K. J. Am. Chem. Soc. 1970, 92, 1651–1659. doi:10.1021/ja00709a039
- Jacobsen, N.; Torssell, K. Justus Liebigs Ann. Chem. 1972, 763, 135–147. doi:10.1002/jlac.19727630115
- Jacobsen, N.; Torssell, K. Acta Chem. Scand. 1973, 27, 3211–3216. doi:10.3891/acta.chem.scand.27-3211
- Ashnagar, A.; Bruce, J. M.; Lloyd-Williams, P.
 J. Chem. Soc., Perkin Trans. 1 1988, 559–561. doi:10.1039/p19880000559
- Liu, B.; Gu, L.; Zhang, J. Recl. Trav. Chim. Pays-Bas 1991, 110, 99–103. doi:10.1002/recl.19911100402
- Onuki, M.; Ota, M.; Otokozawa, S.; Kamo, S.; Tomoshige, S.; Tsubaki, K.; Kuramochi, K. *Tetrahedron* **2020**, *76*, 130899. doi:10.1016/j.tet.2019.130899
- Coppa, F.; Fontana, F.; Minisci, F.; Barbosa, M. C. N.; Vismara, E. *Tetrahedron* 1991, *47*, 7343–7352. doi:10.1016/s0040-4020(01)89736-2
- 83. Wang, D.; Ge, B.; Du, L.; Miao, H.; Ding, Y. Synlett **2014**, *25*, 2895–2898. doi:10.1055/s-0034-1379472
- 84. Yang, J.; Dong, Y.; He, S.; Shi, Z.-C.; Wang, Y.; Wang, J.-Y. Tetrahedron 2019, 75, 130729. doi:10.1016/j.tet.2019.130729
- Tomatsu, A.; Takemura, S.; Hashimoto, K.; Nakata, M. Synlett 1999, 1474–1476. doi:10.1055/s-1999-2875
- Tohma, H.; Morioka, H.; Harayama, Y.; Hashizume, M.; Kita, Y. Tetrahedron Lett. 2001, 42, 6899–6902. doi:10.1016/s0040-4039(01)01407-1
- Wójtowicz, H.; Młochowski, J.; Syper, L.; Yadav, H. S. Synth. Commun. 2006, 36, 1991–2000. doi:10.1080/00397910600634175
- Horii, Z.-i.; Tanaka, T.; Murakami, Y. Pharm. Bull. 1957, 5, 82–84. doi:10.1248/cpb1953.5.82
- 89. Kacan, M.; Karabulut, H. R. F. Turk. J. Chem. 2002, 26, 251-254.
- Murahashi, S.-I.; Fujii, A.; Inubushi, Y.; Komiya, N. Tetrahedron Lett. 2010, 51, 2339–2341. doi:10.1016/j.tetlet.2010.02.134
- Mal, D.; Ghosh, K.; Jana, S. Org. Lett. 2015, 17, 5800–5803. doi:10.1021/acs.orglett.5b02920
- Dissanayake, I.; Hart, J. D.; Becroft, E. C.; Sumby, C. J.;
 Newton, C. G. J. Am. Chem. Soc. 2020, 142, 13328–13333. doi:10.1021/jacs.0c06306
- Gogin, L. L.; Zhizhina, E. G. Kinet. Catal. 2020, 61, 276–282. doi:10.1134/s0023158420010012
- Shanmugam, V. M.; Kulangiappar, K.; Ramaprakash, M.; Vasudevan, D.; Senthil Kumar, R.; Velayutham, D.; Raju, T. *Tetrahedron Lett.* 2017, 58, 2294–2297. doi:10.1016/j.tetlet.2017.04.099
- Couladouros, E. A.; Strongilos, A. T. Sci. Synth. 2006, 28, 217. doi:10.1055/sos-sd-028-00226
- 96. Siguret, V. Hematologie 2006, 12, 389-399.
- Mueller, N. J.; Stueckler, C.; Hall, M.; Macheroux, P.; Faber, K. Org. Biomol. Chem. 2009, 7, 1115–1119. doi:10.1039/b819057g
- Dwyer, C. L.; Gill, C. D.; Ichihara, O.; Taylor, R. J. K. Synlett 2000, 704–706. doi:10.1055/s-2000-6619
- Bundu, A.; Berry, N. G.; Gill, C. D.; Dwyer, C. L.; Stachulski, A. V.; Taylor, R. J. K.; Whittall, J. *Tetrahedron: Asymmetry* 2005, *16*, 283–293. doi:10.1016/j.tetasy.2004.11.015

- 100. Kośnik, W.; Stachulski, A. V.; Chmielewski, M. Tetrahedron: Asymmetry 2005, 16, 1975–1981. doi:10.1016/j.tetasy.2005.04.009
- 101. Kosnik, W.; Bocian, W.; Kozerski, L.; Tvaroska, I.; Chmielewski, M. Chem. – Eur. J. 2008, 14, 6087–6097. doi:10.1002/chem.200800051
- 102. Bunge, A.; Hamann, H.-J.; McCalmont, E.; Liebscher, J. *Tetrahedron Lett.* **2009**, *50*, 4629–4632. doi:10.1016/j.tetlet.2009.05.096
- 103. Ooi, T.; Tayama, E.; Doda, K.; Takeuchi, M.; Maruoka, K. Synlett 2000, 1500–1502. doi:10.1055/s-2000-7652
- 104. Arai, S.; Tsuge, H.; Oku, M.; Miura, M.; Shioiri, T. *Tetrahedron* **2002**, *58*. 1623–1630. doi:10.1016/s0040-4020(02)00028-5
- 105.Berkessel, A.; Guixà, M.; Schmidt, F.; Neudörfl, J. M.; Lex, J. Chem. – Eur. J. **2007**, *13*, 4483–4498. doj:10.1002/chem.200600993
- 106. Lattanzi, A.; Cocilova, M.; Iannece, P.; Scettri, A. Tetrahedron: Asymmetry 2004, 15, 3751–3755. doi:10.1016/j.tetasy.2004.10.009
- 107. Ryu, D. H.; Zhou, G.; Corey, E. J. J. Am. Chem. Soc. 2004, 126, 4800–4802. doi:10.1021/ja049323b
- 108. Corey, E. J. Angew. Chem., Int. Ed. 2002, 41, 1650–1667. doi:10.1002/1521-3773(20020517)41:10<1650::aid-anie1650>3.0.co; 2-b
- 109. Nishimoto, K.; Kim, S.; Kitano, Y.; Tada, M.; Chiba, K. Org. Lett. 2006, 8, 5545–5547. doi:10.1021/ol0623116
- 110. Bendiabdellah, Y.; Villanueva-Margalef, I.; Misale, A.; Nahar, K.; Haque, M.; Thurston, D.; Zinzalla, G. Synthesis 2011, 2321–2333. doi:10.1055/s-0030-1260073
- 111. Sultan, S.; Bhat, M.-u.-S.; Rizvi, M. A.; Shah, B. A. J. Org. Chem. 2019, 84, 8948–8958. doi:10.1021/acs.joc.9b00855
- 112.lto, T.; Ikemoto, T.; Yamano, T.; Mizuno, Y.; Tomimatsu, K. Tetrahedron: Asymmetry 2003, 14, 3525–3531. doi:10.1016/s0957-4166(03)00580-9
- 113. Suhara, Y.; Watanabe, M.; Nakagawa, K.; Wada, A.; Ito, Y.; Takeda, K.; Takahashi, K.; Okano, T. J. Med. Chem. 2011, 54, 4269–4273. doi:10.1021/jm200025f
- 114. Suhara, Y.; Hanada, N.; Okitsu, T.; Sakai, M.; Watanabe, M.; Nakagawa, K.; Wada, A.; Takeda, K.; Takahashi, K.; Tokiwa, H.; Okano, T. *J. Med. Chem.* **2012**, *55*, 1553–1558. doi:10.1021/jm2013166
- 115. Sakane, R.; Kimura, K.; Hirota, Y.; Ishizawa, M.; Takagi, Y.; Wada, A.; Kuwahara, S.; Makishima, M.; Suhara, Y. *Bioorg. Med. Chem. Lett.* 2017, *27*, 4881–4884. doi:10.1016/j.bmcl.2017.09.038
- 116. Suhara, Y.; Hirota, Y.; Hanada, N.; Nishina, S.; Eguchi, S.; Sakane, R.; Nakagawa, K.; Wada, A.; Takahashi, K.; Tokiwa, H.; Okano, T. J. Med. Chem. 2015, 58, 7088–7092. doi:10.1021/acs.jmedchem.5b00999
- 117. Kimura, K.; Hirota, Y.; Kuwahara, S.; Takeuchi, A.; Tode, C.; Wada, A.; Osakabe, N.; Suhara, Y. J. Med. Chem. 2017, 60, 2591–2596. doi:10.1021/acs.jmedchem.6b01717
- 118.Niemczyk, H. J.; Van Arnum, S. D. *Green Chem. Lett. Rev.* **2008**, *1*, 165–171. doi:10.1080/17518250802401273
- 119. Fieser, L. F.; Fry, E. M. J. Am. Chem. Soc. 1940, 62, 228–229. doi:10.1021/ja01858a504
- 120. Kulkarni, M. G.; Kate, S. D. J. Chem. Soc., Perkin Trans. 1 2000, 4242–4244. doi:10.1039/b005120i
- 121. Dobrinescu, C.; Iorgulescu, E. E.; Mihailciuc, C.; Macovei, D.; Wuttke, S.; Kemnitz, E.; Parvulescu, V. I.; Coman, S. M. Adv. Synth. Catal. 2012, 354, 1301–1306. doi:10.1002/adsc.201100718

- 122.Ooi, T.; Ichikawa, H.; Maruoka, K. *Angew. Chem., Int. Ed.* **2001**, *40*, 3610–3612. doi:10.1002/1521-3773(20011001)40:19<3610::aid-anie3610>3.0.co;
- 123. Yadav, J. S.; Reddy, B. V. S.; Swamy, T.; Rao, K. R. *Tetrahedron Lett.* **2004**, *45*, 6037–6039. doi:10.1016/j.tetlet.2004.06.032
- 124. Nagaraja, P.; Vasantha, R. A.; Yathirajan, H. S. J. Pharm. Biomed. Anal. 2002, 28, 161–168. doi:10.1016/s0731-7085(01)00599-4
- 125. Tang, H. A.; Wang, L. F.; Yang, R. D. *Transition Met. Chem.* **2003**, *28*, 395–398. doi:10.1023/a:1023609724972
- 126. Bouhadir, K.; Atallah, H.; Mezher, R.; Fatfat, M.; Gali-Muhtasib, H.; Elaridi, J. Org. Commun. 2017, 10, 259–272. doi:10.25135/acq.oc.26.17.07.040
- 127.Fry, F. H.; Holme, A. L.; Giles, N. M.; Giles, G. I.; Collins, C.; Holt, K.; Pariagh, S.; Gelbrich, T.; Hursthouse, M. B.; Gutowski, N. J.; Jacob, C. Org. Biomol. Chem. 2005, 3, 2579–2587. doi:10.1039/b502197a
- 128. Ribeiro, R. C. B.; de Marins, D. B.; Di Leo, I.; da Silva Gomes, L.; de Moraes, M. G.; Abbadi, B. L.; Villela, A. D.; da Silva, W. F.; da Silva, L. C. R. P.; Machado, P.; Bizarro, C. V.; Basso, L. A.; Cristina de Moraes, M.; Ferreira, V. F.; da Silva, F. d. C.; Nascimento, V. Eur. J. Med. Chem. 2021, 209, 112859. doi:10.1016/j.ejmech.2020.112859
- 129. Thomson, R. H. J. Chem. Soc. 1953, 1196–1199. doi:10.1039/jr9530001196
- 130. Ferreira, V. F.; Pinto, A. V.; Pinto, M. d. C. F. R.; Santos, S. C. J. Braz. Chem. Soc. 1996, 7, 169–172. doi:10.5935/0103-5053.19960026
- 131. Salmon-Chemin, L.; Buisine, E.; Yardley, V.; Kohler, S.; Debreu, M.-A.; Landry, V.; Sergheraert, C.; Croft, S. L.; Krauth-Siegel, R. L.; Davioud-Charvet, E. J. Med. Chem. 2001, 44, 548–565. doi:10.1021/jm001079l
- 132. Yamada, H.; Tanabe, K.; Nishimoto, S.-i. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 665–668. doi:10.1016/j.bmcl.2004.11.038
- 133. Bauer, H.; Fritz-Wolf, K.; Winzer, A.; Kühner, S.; Little, S.; Yardley, V.; Vezin, H.; Palfey, B.; Schirmer, R. H.; Davioud-Charvet, E. *J. Am. Chem. Soc.* **2006**, *128*, 10784–10794. doi:10.1021/ja061155v
- 134.Tanabe, K.; Yamada, H.; Nishimoto, S.-i. *J. Am. Chem. Soc.* **2007**, *129*, 8034–8040. doi:10.1021/ja071369s
- 135.Commandeur, C.; Chalumeau, C.; Dessolin, J.; Laguerre, M. *Eur. J. Org. Chem.* **2007**, 3045–3052. doi:10.1002/ejoc.200700135
- 136. Terasaki, N.; Yamamoto, N.; Tamada, K.; Hattori, M.; Hiraga, T.; Tohri, A.; Sato, I.; Iwai, M.; Iwai, M.; Taguchi, S.; Enami, I.; Inoue, Y.; Yamanoi, Y.; Yonezawa, T.; Mizuno, K.; Murata, M.; Nishihara, H.; Yoneyama, S.; Minakata, M.; Ohmori, T.; Sakai, M.; Fujii, M. Biochim. Biophys. Acta, Bioenerg. 2007, 1767, 653–659. doi:10.1016/j.bbabio.2006.11.008
- 137. Lanfranchi, D. A.; Cesar-Rodo, E.; Bertrand, B.; Huang, H.-H.; Day, L.; Johann, L.; Elhabiri, M.; Becker, K.; Williams, D. L.; Davioud-Charvet, E. Org. Biomol. Chem. 2012, 10, 6375–6387. doi:10.1039/c2ob25812a
- 138.Long, T. E.; Lu, X.; Galizzi, M.; Docampo, R.; Gut, J.; Rosenthal, P. J. Bioorg. Med. Chem. Lett. 2012, 22, 2976–2979. doi:10.1016/j.bmcl.2012.02.045
- 139. Naturale, G.; Lamblin, M.; Commandeur, C.; Felpin, F.-X.; Dessolin, J. *Eur. J. Org. Chem.* **2012**, 5774–5788. doi:10.1002/ejoc.201200722

- 140. Liu, Y.; Liu, Y.; Li, Z.; Zhou, G.-B.; Yao, Z.-J.; Jiang, S. *Bioorg. Med. Chem. Lett.* **2014**, *24*, 1650–1653. doi:10.1016/j.bmcl.2014.02.072
- 141. Goebel, E. S.; Barany, G. *Tetrahedron Lett.* **2016**, *57*, 5850–5851. doi:10.1016/j.tetlet.2016.11.043
- 142. Minisci, F.; Bernardi, R.; Bertini, F.; Galli, R.; Perchinummo, M. *Tetrahedron* **1971**, *27*, 3575–3579. doi:10.1016/s0040-4020(01)97768-3
- 143. Gutiérrez-Bonet, Á.; Remeur, C.; Matsui, J. K.; Molander, G. A. J. Am. Chem. Soc. 2017, 139, 12251–12258. doi:10.1021/jacs.7b05899
- 144.Sutherland, D. R.; Veguillas, M.; Oates, C. L.; Lee, A.-L. *Org. Lett.* **2018**, *20*, 6863–6867. doi:10.1021/acs.orglett.8b02988
- 145.Liu, R.-H.; He, Y.-H.; Yu, W.; Zhou, B.; Han, B. *Org. Lett.* **2019**, *21*, 4590–4594. doi:10.1021/acs.orglett.9b01496
- 146. Pandaram, S.; T. P., A. K.; Ilangovan, A. *Org. Biomol. Chem.* **2020**, *18*, 3027–3031. doi:10.1039/d0ob00323a
- 147.Baral, E. R.; Kim, S. H.; Lee, Y. R. *Asian J. Org. Chem.* **2016**, *5*, 1134–1141. doi:10.1002/ajoc.201600219
- 148.Li, D.; Yang, W.-C. Tetrahedron Lett. 2019, 60, 1792–1795. doi:10.1016/j.tetlet.2019.06.005
- 149.Gu, Y.-R.; Duan, X.-H.; Yang, L.; Guo, L.-N. Org. Lett. 2017, 19, 5908–5911. doi:10.1021/acs.orglett.7b02902
- 150.Liu, S.; Shen, T.; Luo, Z.; Liu, Z.-Q. Chem. Commun. 2019, 55, 4027–4030. doi:10.1039/c9cc01704f
- 151.Li, D.; Shen, X. *Org. Biomol. Chem.* **2020**, *18*, 750–754. doi:10.1039/c9ob02289a
- 152. Boudalis, A. K.; Policand, X.; Sournia-Saquet, A.; Donnadieu, B.; Tuchagues, J.-P. *Inorg. Chim. Acta* **2008**, *361*, 1681–1688. doi:10.1016/j.ica.2007.02.027
- 153. Yamago, S.; Hashidume, M.; Yoshida, J.-i. *Chem. Lett.* **2000**, *29*, 1234–1235. doi:10.1246/cl.2000.1234
- 154. Yamago, S.; Hashidume, M.; Yoshida, J.-i. Tetrahedron 2002, 58, 6805–6813. doi:10.1016/s0040-4020(02)00784-6
- 155. Sansotera, M.; Gambarotti, C.; Famulari, A.; Baggioli, A.; Soave, R.; Venturini, F.; Meille, S. V.; Wlassics, I.; Navarrini, W. *Tetrahedron* 2014, 70, 5298–5309. doi:10.1016/j.tet.2014.05.024
- 156. Fieser, L. F.; Oxford, A. E. J. Am. Chem. Soc. 1942, 64, 2060–2065. doi:10.1021/ja01261a015
- 157. Waske, P. A.; Mattay, J.; Oelgemöller, M. *Tetrahedron Lett.* **2006**, *47*, 1329–1332. doi:10.1016/j.tetlet.2005.12.060
- 158.Oelgemöller, M.; Schiel, C.; Fröhlich, R.; Mattay, J. Eur. J. Org. Chem. 2002, 2465–2474. doi:10.1002/1099-0690(200208)2002:15<2465::aid-ejoc2465>3.0.co;2
- 159. Schiel, C.; Oelgemöller, M.; Mattay, J. Synthesis 2001, 1275–1279. doi:10.1055/s-2001-15072
- 160. Schiel, C.; Oelgemöller, M.; Ortner, J.; Mattay, J. *Green Chem.* **2001**, *3*. 224–228. doi:10.1039/b106425h
- 161.Oelgemöller, M.; Schiel, C.; Ortner, J.; Mattay, J. The solarchemical photo-Friedel-Crafts acylation of quinones. AG-Solar Nordrhein-Westfalen - Solare Chemie und Solare Materialforschung; AG-Solar NRW: Jülich, Germany, 2002.
- 162. Westwood, M. T.; Lamb, C. J. C.; Sutherland, D. R.; Lee, A.-L. Org. Lett. 2019, 21, 7119–7123. doi:10.1021/acs.orglett.9b02679
- 163. Borah, A.; Sharma, A.; Hazarika, H.; Gogoi, P. ChemistrySelect 2017, 2, 9999–10003. doi:10.1002/slct.201702404
- 164. Scott, G. K.; Atsriku, C.; Kaminker, P.; Held, J.; Gibson, B.; Baldwin, M. A.; Benz, C. C. Mol. Pharmacol. 2005, 68, 606–615. doi:10.1124/mol.105.013474

- 165. Kallmayer, H. J. Arch. Pharm. (Weinheim, Ger.) 1974, 307, 806–814. doi:10.1002/ardp.19743071014
- 166. Karunan, T.; Mathew, N.; Srinivasan, L.; Muthuswamy, K. Drug Dev. Res. 2013, 74, 216–226. doi:10.1002/ddr.21065
- 167. Wang, M.; Zhang, C.; Sun, L.-P.; Ding, C.; Zhang, A. J. Org. Chem. **2014**, *79*, 4553–4560. doi:10.1021/jo500572u
- 168.Li, X.; Himes, R. A.; Prosser, L. C.; Christie, C. F.; Watt, E.; Edwards, S. F.; Metcalf, C. S.; West, P. J.; Wilcox, K. S.; Chan, S. S. L.; Chou, C. J. *J. Med. Chem.* **2020**, *63*, 5865–5878. doi:10.1021/acs.imedchem.0c00168
- 169. Jing, L.; Wu, G.; Hao, X.; Olotu, F. A.; Kang, D.; Chen, C. H.; Lee, K.-H.; Soliman, M. E. S.; Liu, X.; Song, Y.; Zhan, P. Eur. J. Med. Chem. 2019, 183, 111696. doi:10.1016/j.eimech.2019.111696
- 170. Bowen, R. D.; Mahmood, T.; Maitland, D. J.; Mercer, K. E.; Gallagher, R. T. Eur. J. Mass Spectrom. 2009, 15, 617–626. doi:10.1255/ejms.1023
- 171.Namsa-aid, A.; Ruchirawat, S. *Org. Lett.* **2002**, *4*, 2633–2635. doi:10.1021/ol026074s
- 172.Zacconi, F. C.; Nuñez, O. N.; Cabrera, A. L.; Valenzuela, L. M.; del Valle, J. M.; de la Fuente, J. C. *J. Chem. Thermodyn.* **2016**, *103*, 325–332. doi:10.1016/j.jct.2016.08.016
- 173.Wu, S.-P.; Huang, R.-Y.; Du, K.-J. *Dalton Trans.* **2009**, 4735–4740. doi:10.1039/b822613j
- 174. Patil, A.; Lande, D. N.; Nalkar, A.; Gejji, S. P.; Chakrovorty, D.; Gonnade, R.; Moniz, T.; Rangel, M.; Pereira, E.; Salunke-Gawali, S. *J. Mol. Struct.* 2017, 1143, 495–514. doi:10.1016/j.molstruc.2017.04.094
- 175.Ge, Y.; Li, A.; Wu, J.; Feng, H.; Wang, L.; Liu, H.; Xu, Y.; Xu, Q.; Zhao, L.; Li, Y. Eur. J. Med. Chem. 2017, 128, 180–191. doi:10.1016/j.ejmech.2017.01.034
- 176.Mital, A.; Mahlavat, S.; Bindal, S.; Sonawane, M.; Negi, V. *Der Pharm. Chem.* **2010**, *2* (3), 63–73.
- 177.Bittner, S.; Gorohovsky, S.; Paz-Tal (Levi), O.; Becker, J. Y. Amino Acids **2002**, *22*, 71–93. doi:10.1007/s726-002-8202-3
- 178. Brun, M.-P.; Braud, E.; Angotti, D.; Mondésert, O.; Quaranta, M.; Montes, M.; Miteva, M.; Gresh, N.; Ducommun, B.; Garbay, C. *Bioorg. Med. Chem.* **2005**, *13*, 4871–4879. doi:10.1016/j.bmc.2005.05.005
- 179. Cao, S.; Murphy, B. T.; Foster, C.; Lazo, J. S.; Kingston, D. G. I. Bioorg. Med. Chem. 2009, 17, 2276–2281. doi:10.1016/j.bmc.2008.10.090
- 180. Kayashima, T.; Mori, M.; Mizutani, R.; Nishio, K.; Kuramochi, K.; Tsubaki, K.; Yoshida, H.; Mizushina, Y.; Matsubara, K. Bioorg. Med. Chem. 2010, 18, 6305–6309. doi:10.1016/j.bmc.2010.07.022
- 181. Chadar, D.; Camilles, M.; Patil, R.; Khan, A.; Weyhermüller, T.; Salunke-Gawali, S. J. Mol. Struct. 2015, 1086, 179–189. doi:10.1016/j.molstruc.2015.01.029
- 182.Chadar, D.; Banerjee, P.; Saha, S. K.; Bhand, S.; Patil, R.; Salunke-Gawali, S. J. Mol. Struct. 2019, 1179, 443–452. doi:10.1016/j.molstruc.2018.11.024
- 183. Prasad, C. V.; Nayak, V. L.; Ramakrishna, S.; Mallavadhani, U. V. Chem. Biol. Drug Des. 2018, 91, 220–233. doj:10.1111/cbdd.13073
- 184. Sharma, U.; Katoch, D.; Sood, S.; Kumar, N.; Singh, B.; Thakur, A.; Gulati, A. *Indian J. Chem.* **2013**, *52B*, 1431–1440.
- 185. Yadav, J. S.; Reddy, B. V. S.; Swamy, T. *Synthesis* **2004**, 106–110. doi:10.1055/s-2003-44364

- 186. Yadav, J. S.; Subba Reddy, B. V.; Shiva Shankar, K.; Swamy, T.; Premalatha, K. Bull. Korean Chem. Soc. 2008, 29, 1418–1420. doi:10.5012/bkcs.2008.29.7.1418
- 187.Tanoue, Y.; Teraoka, T.; Kai, N.; Nagai, T.; Ushio, K. *J. Heterocycl. Chem.* **2010**, *47*, 1447–1449. doi:10.1002/jhet.492
- 188. Escobedo-González, R. G.; Pérez Martínez, H.; Nicolás-Vázquez, M. I.; Martínez, J.; Gómez, G.; Serrano, J. N.; Carranza Téllez, V.; Vargas-Requena, C. L.; Miranda Ruvalcaba, R. J. Chem. 2016, 3870529. doi:10.1155/2016/3870529
- 189.Chen, C.; Liu, Y.-Z.; Shia, K.-S.; Tseng, H.-Y. Bioorg. Med. Chem. Lett. **2002**, *12*, 2729–2732. doi:10.1016/s0960-894x(02)00532-2
- 190. Borovkov, V. V.; Filippovich, E. I.; Evstigneeva, R. P. Chem. Heterocycl. Compd. 1988, 24, 494–501. doi:10.1007/bf00755687
- 191. Singh, W. M.; Baruah, J. B. J. Mol. Struct. 2009, 931, 82–86. doi:10.1016/j.molstruc.2009.05.031
- 192. Braud, E.; Goddard, M.-L.; Kolb, S.; Brun, M.-P.; Mondésert, O.; Quaranta, M.; Gresh, N.; Ducommun, B.; Garbay, C. *Bioorg. Med. Chem.* **2008**, *16*, 9040–9049. doi:10.1016/j.bmc.2008.08.009
- 193.Singh, W. M.; Baruah, J. B. J. Chem. Crystallogr. 2011, 41, 952–958. doi:10.1007/s10870-011-0024-8
- 194. Tamura, K.; Southwick, E. C.; Kerns, J.; Rosi, K.; Carr, B. I.; Wilcox, C.; Lazo, J. S. Cancer Res. 2000, 60, 1317–1325.
- 195.Li, W.-W.; Heinze, J.; Haehnel, W. *J. Am. Chem. Soc.* **2005**, *127*, 6140–6141. doi:10.1021/ja050974x
- 196.Li, W.-W.; Hellwig, P.; Ritter, M.; Haehnel, W. Chem. Eur. J. 2006, 12, 7236–7245. doi:10.1002/chem.200501212
- 197. Kumar, S.; Malachowski, W. P.; DuHadaway, J. B.; LaLonde, J. M.; Carroll, P. J.; Jaller, D.; Metz, R.; Prendergast, G. C.; Muller, A. J. J. Med. Chem. 2008, 51, 1706–1718. doi:10.1021/jm7014155
- 198. Nickerson, W. J.; Falcone, G.; Strauss, G. *Biochemistry* **1963**, *2*, 537–543. doi:10.1021/bi00903a025

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