ROYAL SOCIETY OF CHEMISTRY

## PAPER

Check for updates

# Zinc(II) triazole *meso*-arylsubstituted porphyrins for UV-visible chloride and bromide detection. Adsorption and catalytic degradation of malachite green dye†

Mouhieddinne Guergueb,[a] Jihed Brahmi,[a] Soumaya Nasri,*[ab] Frédérique Loiseau,[c] Kaïss Aouadi,[d] Vincent Guerineau,[e] Shabir Najmudin [iD][f] and Habib Nasri [iD][a]

Three new triazole *meso*-arylporphyrins (**4a–c**) were synthesized by the copper(I)-catalyzed azide alkyne cycloaddition (CuAAC) "click" reaction in high yield. The corresponding zinc(II) coordination compounds (**5a–c**) have also been prepared. All **4a–c** and **5a–c** porphyrin species were fully characterized by elemental analysis, electrospray ionization and MALDI-TOF mass spectrometry, infrared spectroscopy, proton nuclear magnetic resonance, UV-visible, fluorescence and cyclic voltammetry. The zinc(II) **5a–c** complexes have been tested as detectors for Cl⁻ and Br⁻ anions. UV-visible titrations reveal that these host systems exhibit strong anion binding affinities. The efficiency of the adsorption of the malachite green dye (MG) dye on the **4a–c** free base porphyrins and the corresponding zinc(II) complexes **5a–c** was investigated by a kinetic study using these synthetic porphyrin derivatives as adsorbents. The use of our triazole Zn(II) complexes in the catalytic degradation of the MG dye is the first example where a metalloporphyrin is involved in the MG dye decolorization reaction. The degradation reactions were carried out using an ecological oxidant (H₂O₂), where the efficiency of the decolorization has been characterized by UV-visible spectroscopic analysis. Several factors affecting the degradation phenomenon have been studied. The energetic parameters concerning the degradation process have also been determined.

## 1. Introduction

In the fields of chemistry, biochemistry and material sciences, porphyrins and metalloporphyrins have been extensively studied due to their vast range of potential applications such as: photodynamic therapy, catalysis,[1,2] as building blocks,[3] in artificial photosynthetic systems[4] and as sensors.[5,6] The chemical modifications at the *meso* positions of porphyrins give rise to a large number of porphyrin derivatives with unique electronic and optical properties. Phosphines, pyridines, imidazoles and crown ethers, to name a few, have been used to functionalize *meso*-arylporphyrins.[7–10] Since its introduction in 2001 by Sharpless *et al.*,[11] the copper catalyzed azide–alkyne cycloaddition (CuAAC) known as "click chemistry" has been widely used to link moieties together in a very efficient and reliable way to prepare new functionalized *meso*-arylporphyrins. Beer *et al.*,[12,13] reported the synthesis and the characterization of several functionalized *meso*-porphyrins such as the "porphyrin-cage" and the tetra-triazole-appended "picket-fence" zinc(II) coordination compounds. These species were prepared to be tested as receptors for halide and other inorganic salts. Thus, the aim is to investigate the role of various substituents at the porphyrin *meso*-positions for receptor efficiency.

Interestingly, porphyrins and metalloporphyrins have been tested effectively in the degradation of chemical dyes. Indeed, these species constitute a large part of water pollutants due to the sewage discharge containing carcinogenic and chronically toxic organic dyes,[14] which are extensively used in textile, printing and photographic industries. During the late decade, several free bases porphyrins have been used as catalysts in presence of TiO₂, in the photocatalytic degradation of dyes, *e.g.* the *meso*-arylporphyrins substituted by phenyl carboxylic acid

*ᵃUniversity of Monastir, Laboratoire de Physico-chimie des Matériaux, Faculté des Sciences de Monastir, Avenue de l'environnement, 5019 Monastir, Tunisia. E-mail: hnasri1@gmail.com; Habib.Nasri@fsm.rnu.tn; Fax: +216 73500278*

*ᵇDepartment of Chemistry, College of Science Al-Zulfi, Majmaah University, Saudi Arabia*

*ᶜDépartement de Chimie Moléculaire, Université Grenoble Alpes, 301 rue de la Chimie, CS 40700, 38058 Grenoble Cedex 9, France*

*ᵈDepartment of Chemistry, College of Science, Qassim University, Buraidah 51452, Saudi Arabia*

*ᵉInstitut de Chimie des Substances Naturelles CNRS, Avenue de la Terrasse, F-91198 Gif-sur-Yvette, France*

*ᶠRandall Centre for Cell and Molecular Biophysics, Faculty of Life Sciences and Medicine, King's College, 3rd Floor New Hunt's House, London SE1 1UL, UK*

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0ra03070h
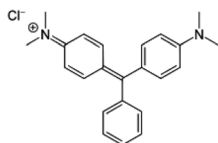
groups and di-*tert*-butyl-substituted phenyl rings have been used in the degradation of the methylene blue (MB) dye.[15] Metal porphyrins such as the manganese(III) metalloporphyrin based polymers have also been used in the methylene blue dye (MB) degradation in an aqueous solution of hydrogen peroxide.[16] We also reported the use of two 4-cyanopyridine–cobaltous-porphyrin complexes type $[Co^{II}(Porph)(4-CNpy)]$ (Porph = *meso*-tetrakis(*para*-methoxyphenyl)porphyrinato and *meso*-tetra(*para*-chlorophenyl)porphyrin) in the same MB dye degradation in presence of $H_2O_2$.[17] In 2018, we reported the catalytic degradation of the Calmagite dye using an aqueous solution of $H_2O_2$ with the (4,4′-bipyridine)[(tetrakis(ethyl-4(4-butyryl)oxy-phenyl)porphyrinato)]zinc(II) complex.[18]

Among dyes, the malachite green (MG) dye, with the formula $[C_6H_5C(C_6H_4N(CH_3)_2)_2]Cl$ (Scheme 1) is widely used in the textile and paper industry. The $[C_6H_5C(C_6H_4N(CH_3)_2)_2]^+$ cation which gives the very intense green color to this dye exhibits a very strong absorption band at 621 nm.
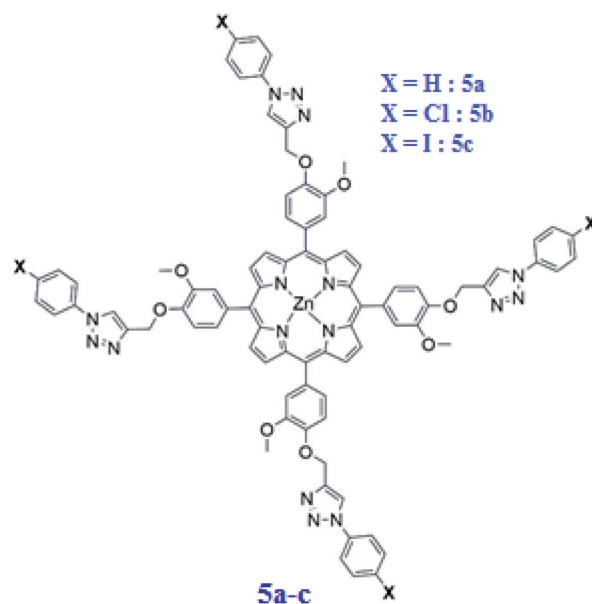
During the past two decades, reports on the enduring carcinogenic and mutagenic characteristics of MG dye have increased.[19] Since 2000, the MG dye has been listed as a priority chemical for carcinogenicity assessment by the U.S. Food and Drug Administration.[20] However, due to its efficacy and low cost, it is still used in many countries. Therefore, many treatment technologies have been applied to decolorize MG from aqueous medium.[21]

Here, we describe the preparation of three new substituted *meso*-arylporphyrins using the "click chemistry" strategy and their zinc(II) corresponding complexes namely: the *meso*-tetrakis(3-methoxy-4-((1-phenyl-1H-1,2,3-triazol-4-yl)methoxy)phenyl)porphyrin (**4a**) [$H_2(T_{AzP}-HVP)$], the *meso*-tetrakis(4-((1-(4-chlorophenyl)-1H-1,2,3-triazol-4-yl)methoxy)-3-methoxyphenyl)porphyrin (**4b**) [$H_2(T_{AzP}-ClVP)$], and the *meso*-tetrakis(4-((1-(4-iodinephenyl)-1H-1,2,3-triazol-4-yl)methoxy)-3-methoxyphenyl)porphyrin (**4c**) [$H_2(T_{AzP}-IVP)$] and the corresponding zinc(II) metalloporphyrins: **5a–c** with the formulas [$Zn(T_{AzP}-HVP)$], [$Zn(T_{AzP}-ClVP)$] and [$Zn(T_{AzP}-IVP)$], respectively (Scheme 2).

All these porphyrin derivatives have been characterized by UV-visible, fluorescence, IR and ¹H NMR spectrometry, as well as by mass spectrometry and cyclic voltammetry. The second part of this paper concerns the study of the chloride and bromide ions receptor by the zinc(II) chloro–triazole and iodine–triazole porphyrin complexes **5b–c**. The other important goal of the present work is to study the efficiency of the free base porphyrins **4a–c** and the corresponding zinc(II) complexes **5a–c** in the adsorption of the malachite green (MG) dye. The catalytic oxidative degradation efficiency of the MG dye using the three triazole *meso*-arylporphyrin zinc(II) compounds **5a–c** have also



Scheme 2  Structure of [$Zn(T_{AzP}-XVP)$] complexes. **5a**: X = H, **5b**: X = Cl and **5c**: X = I.

been described which is the first example of the use of a porphyrinic compound for the degradation of this dye.

## 2.  Results and discussion

### 2.1.  Synthesis

Scheme 3 depicts the overall synthetic route for the preparation of *meso*-tetrakis(3-methoxy-4-((1-phenyl-1H-1,2,3-triazol-4-yl)methoxy)phenyl)porphyrin (**4a**) [$H_2(T_{AzP}-HVP)$], the *meso*-tetrakis(4-((1-(4-chlorophenyl)-1H-1,2,3-triazol-4-yl)methoxy)-3-methoxyphenyl)porphyrin (**4b**) [$H_2(T_{AzP}-ClVP)$] and the *meso*-tetrakis(4-((1-(4-iodinephenyl)-1H-1,2,3-triazol-4-yl)methoxy)-3-methoxyphenyl)porphyrin (**4c**) [$H_2(T_{AzP}-IVP)$], as well as the corresponding zinc(II) metalloporphyrins **5a–c** with the formulas [$Zn(T_{AzP}-HVP)$], [$Zn(T_{AzP}-ClVP)$] and [$Zn(T_{AzP}-IVP)$], respectively (see synthetic procedures in the ESI†). The first step is the preparation of the 3-methoxy-4-(prop-2-ynl-yloxy) benzaldehyde (**2**). This derivative was prepared by reacting the 3-hydroxy-3-methoxy-benzaldehyde (**1**) with $K_2CO_3$ and the propargyl bromide in acetone under reflux for 4 hours. The second step is the synthesis of the three substituted [1,2,3](triazol-4-ylmethoxy)-benzaldehyde: 3-methoxy-4-(3-phenyl-3H-[1,2,3]triazol-4-ylmethoxy)-benzaldehyde (**3a**), the 4-[3-(4-chloro-phenyl)-3H-[1,2,3]triazol-4-ylmethoxy]-3-methoxy-benzaldehyde (**3b**) and the (4-iodine-phenyl)-3H-[1,2,3]triazol-4-ylmethoxy]-3-methoxy-benzaldehyde (**3c**). These compounds were prepared using the so-called "click reaction" (CuAAC) by reacting the 3-methoxy-4-(prop-2-ynl-yloxy)benzaldehyde (**2**) with the 1-azidobenzene or the 1-azido-4-chlorobenzene or the 1-azido-4-iodinebenzene and the diisopropylethylamine (DIPEA) reagents in the presence of copper(I) (CuI) catalyst.[22] The success of the click reaction



Scheme 1  Structure of the malachite green (MG) dye.
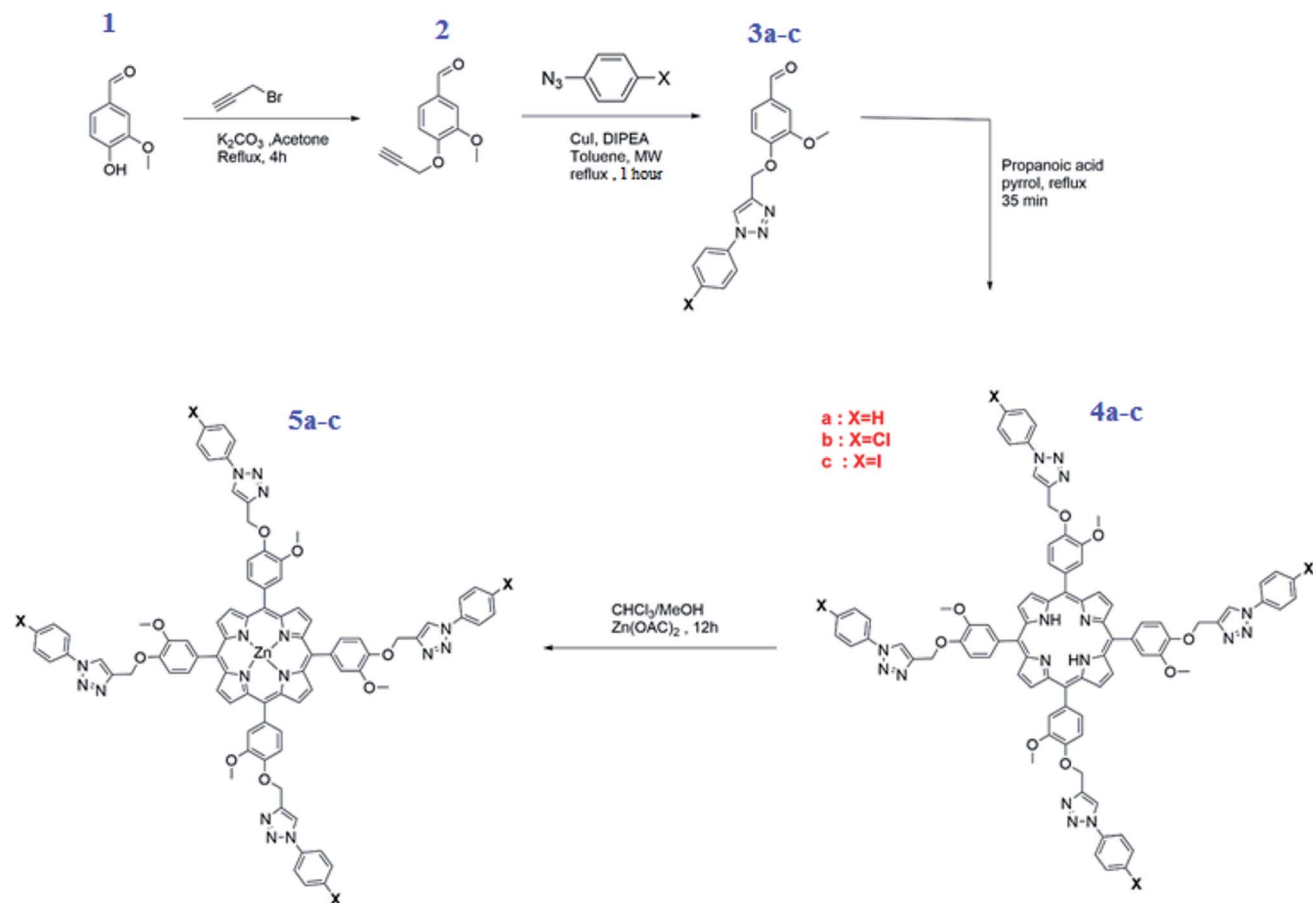
**Scheme 3** Synthesis of aldehydes **2−3a−c**, the free base porphyrins **4a−c** and the zinc(II) porphyrin complexes **5a−c**.

is verified in IR by the disappearance of the two absorption bands of the azido group ($\nu_{as}(N_3) \sim 2130$ and $\sim 2080$ cm$^{-1}$) and the alkyne group of compound **2** and the appearance of a new band at 1726 cm$^{-1}$ assigned to $\nu$(C=N, N=N) of the triazole group (Fig. SI-2†). The three triazole-*meso*-porphyrins **4a−c** were prepared using a slightly modified Adler–Longo procedure.[23] The metalation by the zinc(II) of the three **4a−c** porphyrins were made according to the literature method[24] using the zinc(II) acetate salt Zn(OAc)$_2$·2H$_2$O leading to the three zinc(II) metalloporphyrins [Zn(T$_{AzP}$-HVP)] (**5a**), [Zn(T$_{AzP}$-ClVP)] (**5b**) and [Zn(T$_{AzP}$-IVP)] (**5c**). The three free base porphyrins **4a−c** and the corresponding zinc(II) metalloporphyrins **5a−c** were characterized by UV-visible, fluorescence, IR and $^1$H NMR spectroscopies as well as by mass spectrometry.

### 2.2. Mass spectrometry

All synthetic compounds were characterized by ESI or MALDI-TOF mass spectrometry (see ESI†). The MALDI-TOF spectra of our three zinc(II) metalloporphyrins [Zn(T$_{AzP}$-HVP)], [Zn(T$_{AzP}$-ClVP)] and [Zn(T$_{AzP}$-IVP)] (**5a−c**), shown in Fig. SI-1,† were recorded in the tetrahydrofuran solvent using the *trans*-2-[3-(4-*tert*-butylphenyl)-2-propenylidene]malonitrile (DCTB) as matrix. For all three zinc(II) porphyrin species, the [M]$^+$ and [M + nH]$^{n+}$

fragments were observed which is a clear indication of the stability of the **5a−c** compounds in THF solutions.

### 2.3. IR and $^1$H NMR investigation

The IR spectra of the **4a−c** free base porphyrins and the corresponding zinc(II) metallated species **5a−c** confirm the formation
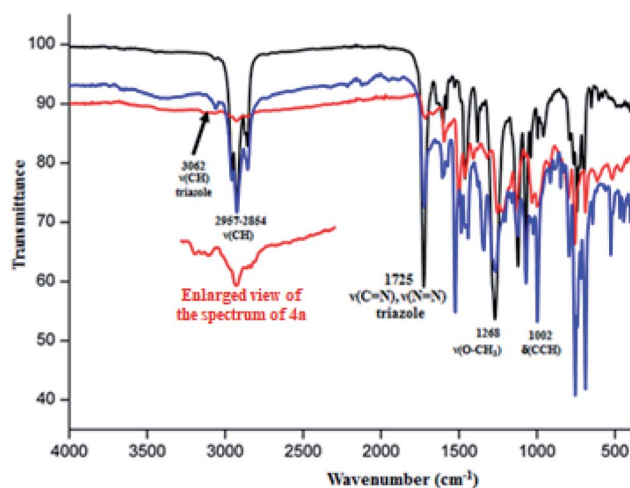


**Fig. 1** IR spectra (solid state) of [Zn(T$_{AzP}$-HVP)] (**5a**) (red color), [Zn(T$_{AzP}$-ClVP)] (**5b**) (black color) and [Zn(T$_{AzP}$-IVP)] (**5c**) (blue color).

**Table 1** UV-visible data of the three triazole *meso*-arylporphyrins (**4a–c**) and a selection of several *meso*-arylporphyrins and zinc(II) tetra-coordinated metalloporphyrins. The solvent used is the dichloromethane

Free base *meso*-arylporphyrins

| Compound | $\lambda_{max}$ (nm) ($\varepsilon \times 10^{-3}$ L mmol$^{-1}$ cm$^{-1}$) | | | | | $E_{gap}$-opt (eV) | Ref. |
|---|---|---|---|---|---|---|---|
| | Soret band | | | Q bands | | | |
| H$_2$(TPP)[a] | 416(419) | 513(20) | 550(20) | 590(6) | 646(6) | 1.89 | 26 |
| H$_2$(TTP)[b] | 420 | 516 | 552 | 594 | 640 | 1.86 | 27 |
| H$_2$(TPBP)[c] | 420(513) | 516(17) | 552(7) | 591(5) | 646(4) | 1.82 | 4 |
| H$_2$(TEBOP)[d] | 422(295) | 517(9) | 554(8) | 593(5) | 651(7) | 1.85 | 18 |
| H$_2$(T$_{AzP}$-HVP) **4a** | 420(565) | 517(42) | 554(19) | 593(15) | 650(13) | 1.88 | This work |
| H$_2$(T$_{AzP}$-ClVP) **4b** | 422(551) | 518(35) | 555(26) | 594(15) | 651(17) | 1.87 | This work |
| H$_2$(T$_{AzP}$-IVP) **4c** | 424(576) | 520(46) | 555(29) | 595(24) | 652(18) | 1.86 | This work |

Zinc(ii) *meso*-arylporphyrin complexes

| Compound | $\lambda_{max}$ (nm) ($\varepsilon \times 10^{-3}$ L mmol$^{-1}$ cm$^{-1}$) | | | $E_{gap}$-opt (eV) | Ref. |
|---|---|---|---|---|---|
| | Soret band | Q bands | | | |
| [Zn(TPP)][a] | 421(524) | 550(21) | 591(25) | 1.91 | 28 |
| [Zn(TMP)][e] | 420 | 550 | 586 | — | 29 |
| [Zn(TPBP)][c] | 425(546) | 554(24) | 596(9) | 1.99 | 24 |
| [Zn(TEBOP)][d] | 424(219) | 552(11) | 594(5) | 2.03 | 18 |
| [Zn(T$_{AzP}$-HVP)] **5a** | 424(530) | 551(26) | 592(10) | 2.04 | This work |
| [Zn(T$_{AzP}$-ClVP)] **5b** | 426(523) | 552(31) | 594(15) | 2.02 | This work |
| [Zn(T$_{AzP}$-IVP)] **5c** | 426(532) | 558(29) | 597(12) | 2.01 | This work |

[a] TPP = *meso*-tetraphenylporphyrinato. [b] TTP = *meso*-tetratolylporphyrinato. [c] TPBP = *meso*-tetrakis-[4-(benzoyloxy)phenyl]porphyrinato. [d] TEBOP = *meso*-tetrakis(ethyl-4(4-butyryl)oxyphenyl)porphyrinato. [e] TMP = *meso*-tetrakis(2,4,6-trimethylphenyl)porphyrinato.

of the triazole *meso*-arylporphyrins as well as the corresponding zinc(II) coordination compound. Indeed, the values of the $\nu$(CH) stretching frequencies of the C—H bonds of the triazole and the aryl groups of H$_2$(T$_{AzP}$-HVT) (**4a**), H$_2$(T$_{AzP}$-ClVT) (**4b**) and H$_2$(T$_{AzP}$-IVT) (**4c**) are ~3070 cm$^{-1}$ and in the [2955–2854] cm$^{-1}$ range, respectively (Fig. SI-3†). For the zinc(II) corresponding complexes **5a–c** (Fig. 1), the $\nu$(CH) stretching frequencies values are practically the same as those of the free base porphyrins **4a–c**. The presence of the triazole fragment is confirmed by a strong absorption band at ~1725 cm$^{-1}$ for both free base porphyrins **4a–c** and the corresponding zinc(II) complexes **5a–c**. This band is attributed to $\nu$(N=N) and $\nu$(C=N) of the triazole group.[25] The metallation of **4a–c** porphyrins are confirmed by (i) the disappearance of the absorption band correspondent to the $\nu$(N—H) stretching frequency of the free base porphyrins (~3280 cm$^{-1}$) and (ii) by the shift of the deformation frequency $\delta$(CCH) from ~965 cm$^{-1}$ (**4a–c**) to ~1000 cm$^{-1}$ for the zinc(II) complexes **5a–c** (Fig. SI-3† and 1).

The proton NMR spectra of the three zinc(II) complexes **5a–c** are shown in Fig. SI-4-6.† The β-pyrrolic protons of the three porphyrinates in complexes **5a–c** resonate at about 9 ppm while the phenylic protons resonate between 7.19 and 7.71 ppm. For these three **5a–c** species, the chemical shift values of the triazole protons are ~8 ppm and those of the O–CH$_2$-triazole groups are ~5.4 ppm. The methoxy groups for **5a–c** presents singlets at about 4.0 ppm.

## 2.4. Photophysical properties

The electronic absorption spectra of the free base porphyrins **4a–c** and the corresponding zinc(II) metallated porphyrin species (**5a–c**) are depicted in Fig. SI-7 and SI-8† while the UV-visible data are given in Table 1. The $\lambda_{max}$ values of the Soret band of the free base porphyrins **4a–c** are ~420 nm while those
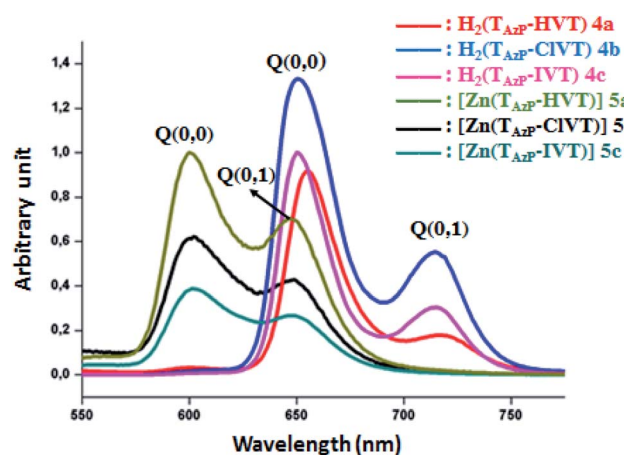


**Fig. 2** The emission spectra of compounds **4a–c** and **5a–c**. The spectra were recorded in dichloromethane solvent with a concentration ~10$^{-6}$ M. The excitation wavelength value is 430 nm.

**Table 2**  Emission parameter values of several *meso*-arylporphyrins and a selection of zinc(ii) *meso*-metalloporphyrins

| Compound | $\lambda_{max}$ (nm) | | $\Phi_f{}^a$ | $\tau_f{}^b$ (ns) | Ref. |
|---|---|---|---|---|---|
| | Q(0, 0) band | Q(0, 1) band | | | |
| **Free base meso-arylporphyrins** | | | | | |
| $H_2(T_{AzP}$-HVP) **4a** | 655 | 719 | 0.091 | 8.82 | This work |
| $H_2(T_{AzP}$-ClVP) **4b** | 651 | 715 | 0.076 | 7.18 | This work |
| $H_2(T_{AzP}$-IVP) **4c** | 650 | 714 | 0.071 | 7.12 | This work |
| $H_2(TPP)^c$ | 656 | 717 | 0.09 | — | 33 |
| $H_2(TPP)^c$ | 653 | 712 | 0.12 | 9.6 | 34 |
| $H_2(TMPP)^d$ | 656 | 719 | 0.082 | 7.16 | 35 |
| $H_2(TClPP)^e$ | 651 | 714 | 0.089 | 7.42 | 26 |
| **Tetracoordinated zinc(ii) *meso*-metalloporhyrins** | | | | | |
| $[Zn(T_{AzP}$-HVP)] **5a** | 601 | 649 | 0.051 | 1.96 | This work |
| $[Zn(T_{AzP}$-ClVP)] **5b** | 600 | 648 | 0.035 | 1.78 | This work |
| $[Zn(_{TAzP}$-IVP)] **5c** | 599 | 647 | 0.032 | 1.51 | This work |
| $[Zn(TPP)]^c$ | 597 | 647 | 0.033 | 1.90 | 35 |
| $[Zn(TTP)]^f$ | 600 | 648 | 0.030 | 1.60 | 36 |

$^a$ $\Phi_f$ = fluorescence quantum yield. $^b$ $\tau_f$ = fluorescent lifetime. $^c$ $H_2(TPP)$ = *meso*-tetraphenylporphyrin. $^d$ $H_2(TMPP)$ = *meso*-tetra(*para*-methoxy)phenylporphyrin. $^e$ $H_2(TClPP)$ = *meso*-tetra(*para*-chlorophenyl)porphyrin. $^f$ TTP = *meso*-tetratolylporphyrinato.

of the Q bands are ∼518, ∼550, ∼590 and ∼650 nm which are typical for *meso*-arylporphyrins. For the tetracoordinated zinc(ii) coordination compounds **5a–c** type [Zn(Porph)] (Porph = $T_{AzP}$-HVP or $T_{AzP}$-ClVP or $T_{AzP}$-IVP porphyrinates), the Soret and the Q bands of these metalloporphyrins are, as expected, slightly redshifted compared to the corresponding free base porphyrins.

Notably, the UV-visible data of our three free bases and the corresponding zinc metallated porphyrins are very close to those of the reported zinc(ii) related species (Table 1). The optical gap values ($E_g$-op) of our synthetic compounds **4a–c** and **5a–c** were determined using the Tauc's relation (eqn (1)):[30,31]

$$(\alpha h\nu)^2 = A[h\nu - E_g\text{-op}] \qquad (1)$$

where $A$ is a constant parameter depending on transition probability, $h\nu$ is the incident photon energy and $\alpha$ is the optical absorption coefficient deduced from absorbance data. The intercepts between the solid lines and *x*-axis allow the determination of $E_g$-op values (Fig. SI-9†). The optical gap energy values of the free bases **4a–c** are 1.88, 1.87 and 1.86 eV, respectively, while those of the corresponding zinc(ii) complexes **5a–c** are 2.05, 2.02 and 2.01 eV, respectively. The fact that the $E_g$-op values of the metallated porphyrins are higher than those of the free bases is mainly due to the higher flexibility of the non metallated porphyrins which leads to the destabilization of the HOMO–LUMO orbital energies. Thus, the net result is the reduction of the gap energy values of these *meso*-porphyrins. However, the zinc metallation of these tetradentate ligands decreases the distortion of the porphyrin core. Therefore, the values of the optical energy of the metalloporphyrins are usually higher than those of the non metallated porphyrins.[32] We notice that both **4a–c** porphyrins and **5a–c** Zn(ii) metalloporphyrins present $E_g$-op values very close to the related *meso*-arylporphyrins and zinc(ii) *meso*-arylporphyrins. Fig. 2 illustrates the fluorescence spectra of **4a–c** and **5a–c**, while the

photoluminescence data of these porphyrin species are given in Table 2.

Importantly, it should be noticed that porphyrin derivatives exhibit interesting photophysical properties especially due to the significant aromaticity of the porphyrin macrocycle. Indeed, these species present one very weak emission transition $S_2 \rightarrow S_0$ of the Soret band between the second excited singlet state $S_2$ and the ground state $S_0$. The second emission transition of porphyrin and metalloporphyrins, which is much stronger than the first transition, is the $S_1 \rightarrow S_0$ of the Q bands between the first excited singlet state $S_1$ to the ground state $S_0$. Therefore, only the following two $S_1 \rightarrow S_0$ transitions are observed for the
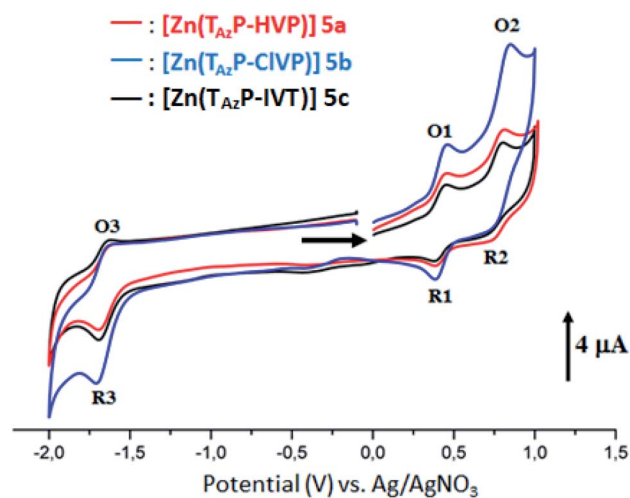


**Fig. 3**  Cyclic voltammograms of complexes **5a–c**. The solvent is a mixture of dichloromethane and acetonitrile (4/1) and the concentration is *ca.* $10^{-3}$ M in 0.1 M TBAPF$_6$, 100 mV s$^{-1}$, vitreous carbon working electrode (∅ = 2 mm).

Table 3 Electrochemical data[a] for the three free base triazole *meso*-arylporphyrins **4a–c** and the corresponding zinc(II) complexes **5a–c** and a selection of several related porphyrins species

| Compound | Oxidations | | Reductions | | |
| | First oxid. | Second oxid. | First red | Second red. | |
| | (O1, R1) | (O2, R2) | (R3, O3) | (R4, O4) | |
| | $E_{1/2}$[b] | $E_{1/2}$ | $E_{1/2}$ | $E_{1/2}$ | Ref. |
|---|---|---|---|---|---|
| $H_2(T_{AzP}\text{-HVP})$ **4a** | 0.86 | 1.09 | −0.95 | −1.36 | This work |
| $H_2(T_{AzP}\text{-ClVP})$ **4b** | 0.85 | 1.08 | −0.76 | −1.40 | This work |
| $H_2(T_{AzP}\text{-IVP})$ **4c** | 0.78 | 1.13 | −0.82 | −1.37 | This work |
| $H_2(TPP)$[c] | 1.02 | 1.26 | −1.20 | −1.55 | 40 |
| $H_2(TBPP)$[d] | 0.95 | 1.36 | −1.12 | −1.53 | 24 |
| $[Zn(T_{AzP}\text{-HVP})]$ **5a** | 0.72 | 1.08 | −1.36 | — | This work |
| $[Zn(T_{AzP}\text{-ClVP})]$ **5b** | 0.71 | 1.09 | −1.37 | — | This work |
| $[Zn(T_{AzP}\text{-IVP})]$ **5c** | 0.72 | 1.05 | −1.37 | — | This work |
| $[Zn(TPP)]$[c] | 0.89 | 1.18 | −1.26 | −1.70 | 41 |
| $[Zn(TBPP)]$[d] | 0.81 | 1.11 | −1.48 | −1.76 | 24 |
| $[Zn(DTPP)]$[e] | 0.72 | 1.15 | −1.42 | — | 40 |

[a] The potentials are reported *versus* SCE. [b] $E_{1/2}$ = half wave potential. [c] TPP = *meso*-tetraphenylporphyrinato. [d] $H_2$TBPP = *meso*-{tetrakis-[4-(benzoyloxy)phenyl]porphyrin. [e] DTPP = 5,15-*p*-ditolyl-10-phenylporphyrinato.

free base porphyrins and metalloporphyrin complexes: $S_1[Q(0, 0)] \rightarrow S_0$ and $S_1[Q(0, 1)] \rightarrow S_0$. For the three free base porphyrin species **4a–c**, the emission spectra show two bands: the weak band at $\lambda_{max} \sim 715$ nm is assigned as Q(0, 1), and the stronger band at $\lambda_{max} \sim 650$ nm is assigned as Q(0,0) (Fig. 2). The values of each $\lambda_{max}$ of our three derivatives are very close to those of the reported *meso*-arylporphyrins (Table 2). The hypochromic shifts of the Q(0, 0) and the Q(0, 1) of the zinc(II) metalloporphyrins **5a–c** compared to those of the corresponding free bases **4a–c** are important for consideration. The $\lambda_{max}$ values of the Q(0, 0) band are ∼600 nm, while the $\lambda_{max}$ values of the Q(0, 1) bands are ∼650 nm. These shifts are mainly due to the metallation of the porphyrins.[37] The Stocks shifts values of the free bases **4a–c** and the **5a–c** zinc(II) metalloporphyrins are small, indicating a fluorescence emission with no significant conformational change between the fundamental and excited states.

The fluorescence properties were studied by both steady-state and time-resolved fluorescence techniques. As indicated in Table 2, the fluorescence quantum yield ($\Phi_f$) value of the $H_2T_{AzP}$-HVP (**4a**) is slightly higher than those of the chloro–triazole and iodine–triazole free base derivatives $H_2T_{AzP}$-ClVP (**4b**) and $H_2T_{AzP}$-IVP (**4c**) with $\Phi_f$ values of 9.1%, 7.6% and 7.1%, respectively, which could be attributed to the higher molar mass of the iodine (in **4c**) and chlorine (in **4b**) atoms compared to that of the hydrogen atom (in **4a**). Notably, the $\Phi_f$ value of the **4a** porphyrin is comparable with that of the *meso*-tetraphenylporphyrin $H_2$TPP ($\Phi_f \sim 10$) (Table 2). As expected the $\Phi_f$ values of our three zinc(II) porphyrins **5a–c** which are 5.1%, 3.5 and 3.1%, respectively, are quite smaller than those of the related free base porphyrins **4a–c**. We also notice that the iodine–triazole and chloro–triazole zinc(II) derivatives exhibit slightly higher fluorescence quantum yields than that of the H-triazole porphyrin species. The lifetime of singlet excited state ($\tau_f$) was measured

by the single photon counting technique, and the fluorescence decays were fitted to single exponentials. The representative fluorescence decays ($\tau_f$) of these four derivatives are similar (see Fig. SI-10†). Our $[Zn(Porph)]$ complexes (**5a–c**) display, as expected, $\tau_f$ values between 7.2 and 8.82 ns which are much higher than the corresponding free bases (**4a–c**) with values in the range 1.51 and 1.96 ns.

## 2.5. Cyclic voltammetry

Cyclic voltammograms (CV) of our porphyrin derivatives **4a–c** and **5a–c** were recorded at room temperature in a mixture of dichloromethane and acetonitrile (4/1) under an argon atmosphere with tetrabutylammonium hexafluorophosphate (TBAPF$_6$) as the supporting electrolyte (0.1 M). The CV of the free base porphyrins **4a–c** are illustrated in Fig. SI-11,† while those of the corresponding zinc(II) complexes **5a–c** are depicted in Fig. 3. The electrochemical data of our synthetic porphyrin species and several related compounds are given in Table 3. The free bases **4a–c** exhibit two reversible one-electron reduction waves and two reversible one-electron oxidation waves, which correspond to the reduction and the oxidation of the porphyrin ring. The half potential values ($E_{1/2}$) of the three triazole *meso*-arylporphyrins **4a–c** are much smaller than those of the unsubstituted *meso*-tetraphenylporphyrin (H$_2$TPP) (Table 3). Neya *et al.*,[38] reported that the second reduction of the porphyrin ring in metalloporphyrins, with non-redox-active cationic metal centers such as Zn(II) and Mg(II), is usually not observed in the electrochemical window of the solvent which is the case for our zinc(II) derivatives **5a–c**. It was also reported that the zinc metallation of the *meso*-arylporphyrins leads to the shifts of the characteristic reduction and oxidation potentials to more negative values.[29,39] In our case, only the $E_{1/2}$ values of the

first reduction waves for [Zn(T$_{azP}$-HVP)], [Zn(T$_{azP}$-ClVP)] and [Zn(T$_{azP}$-IVP)] (**5a–c**) are shifted to more negative values (Table 3).

In short, the CV investigation on our synthetic **4a–c** and **5a–c** porphyrin species show that the nature of the substituted groups on the phenyls of the *meso*-arylporphyrins do not have a very significant effect on the electrochemical properties of this type of species.

Ghosh *et al.*,[42] estimated the electrochemical gap energy ($E_g$-el) as the difference between the potential of the first oxidation potential and the first reduction potential of the porphyrin. The estimated energy values of the HOMO and LUMO orbitals as well as the value of the $E_g$-el can be calculated using the following equations (eqn (2)–(4)):[43]

$$E_{HOMO} = -(V_{onset\text{-}ox} - V_{ref} + 4.8) \text{ eV} \qquad (2)$$

$$E_{LUMO} = -(V_{onset\text{-}red} - V_{ref} + 4.8) \text{ eV} \qquad (3)$$

$$E_g\text{-el} = (E_{LUMO} - E_{HOMO}) \text{ eV} \qquad (4)$$

The $E_{HOMO}$ and the $E_{LUMO}$ are also known as the ionization potential (IP) and the electron affinity (EA), respectively. The $V_{onset\text{-}ox}$ and the $V_{onset\text{-}red}$ are the oxidation onset and the reduction onset, respectively and the $V_{ref}$ is the reference half-wave potential. The values of $E_{HOMO}$, $E_{LUMO}$ and $E_g$-el were calculated using equations eqn (2)–(4) and are reported in Table 4 along with those of several related compounds. The energy level of the HOMO and LUMO orbitals as well as the $E_g$-el of **4a–c** and **5a–c** are illustrated in Fig. SI-12.† From Table 4 we can deduce: (i) except for the case of the H$_2$(TPBP) porphyrin, the electrochemical gap energy values of the zinc metalloporphyrins are higher than those of the free base porphyrins; (ii) as expected, the values of the $E_g$-el of **4a–c** and **5a–c** are higher than those of the $E_g$-op (optical gap energy) of the same species, which is also the case for all known *meso*-

arylporphyrins and the corresponding [Zn(Porph)] complexes[24] and (iii) the iodine–triazole fee base porphyrin **4c** exhibits the smallest $E_g$-el gap value (1.77 eV) compared to those of **4a–b** (1.95 and 1.87 eV, respectively) while the three zinc(II) metallated porphyrins **5a–c** presents very close values of $E_g$-el ∼ 2.14 eV.

## 2.6.  Anion binding studies

The zinc(II) complexes **5b–c** with the chloro–triazole and the iodine–triazole *meso*-porphyrinates were tested as Cl$^-$ and Br$^-$ anions detector by UV-visible titration. The two halide ions titrations, carried out in dichloromethane solvent, revealed significant perturbation of the Soret and the Q bands of the zinc(II) **5b–c** receptors as a function of the concentration of the *tert*-butyl ammonium chloride and bromide salts (TBACl and TBABr). In the literature, several UV-visible titrations involving the tetracoordinated zinc(II) *meso*-porphyrins type [Zn(Porph)] (Porph = *meso*-arylporphyrin) and neutral N-donor ligands are reported.[18] These investigations indicate the presence of a redshift of the Soret and the Q bands, upon addition of the axial ligand, of about 10 nm and 5 nm for the two bands, respectively, and the formation of a 1 : 1 coordination complex type [Zn(Porph)(L)] (L = N-neutral axial ligand).

The UV-visible titration spectra of [Zn(T$_{AzP}$-ClVP)] (**5b**) (concentration about 10$^{-6}$ M) with Cl$^-$, in the Soret region, are shown in Fig. 4 and 5. The gradual addition of chloride to **5b** leads to a bathochromic shift of the Soret band from 426 nm to 436 nm ($\Delta\lambda_{max}$ = 10 nm) with one distinct isosbestic point at 431 nm. In the Q region, the titration of the same complex **5b** with Cl$^-$ exhibits also a redshift of the Q(0, 0) and Q(0, 1) bands. The titration of the same complex **5b** with the bromide anion, using the same solvent and concentration as the Cl$^-$ titration, gives a comparable result with a redshift of the Soret and the Q bands (Fig. 4). The isosbestic points are 431 nm for the Soret

**Table 4** Values of the HOMO, LUMO orbitales energies and the electrochemical gap energy for a selection of porphyrins species

| Compound | $E_{HOMO}$ (eV) | $E_{LUMO}$ (eV) | $E_g$-el (eV) | Ref. |
|---|---|---|---|---|
| **Free base *meso*-arylporphyrins** | | | | |
| H$_2$(TPP)[a] | −5.50 | −3.49 | 2.01 | 44 |
| H$_2$(TPBP)[b] | −5.87 | −3.73 | 2.14 | 24 |
| H$_2$(T$_{AzP}$-HVP) **4a** | −5.40 | −3.45 | 1.95 | This work |
| H$_2$(T$_{AzP}$-ClVP) **4b** | −5.53 | −3.66 | 1.87 | This work |
| H$_2$(T$_{AzP}$-IVP) **4c** | −5.32 | −3.56 | 1.77 | This work |
| | | | | |
| **Zinc(II) *meso*-arylporphyrin tetracoordinated complexes** | | | | |
| [Zn(TPP)][a] | −5.58 | −3.46 | 2.12 | 45 |
| [Zn(TPBP)][b] | −5.64 | −3.27 | 2.37 | 24 |
| [Zn(T$_{AzP}$-HVP)] **5a** | −5.24 | −3.09 | 2.15 | This work |
| [Zn(T$_{AzP}$-ClVP)] **5b** | −5.23 | −3.09 | 2.14 | This work |
| [Zn(T$_{AzP}$-IVP)] **5c** | −5.24 | −3.11 | 2.13 | This work |

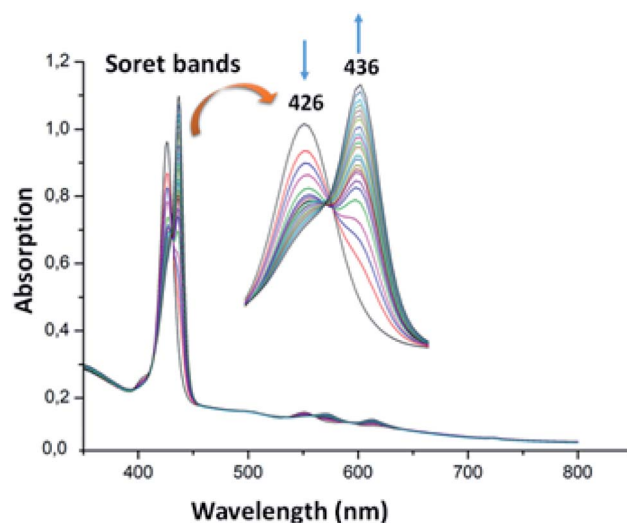[a] TPP = *meso*-tetraphenylporphyrinato.  [b] TBPP = *meso*-tetrakis-[4-(benzoyloxy)phenyl]porphyrinato.



**Fig. 4**  Changes in the absorption spectra in the Soret band region of complex **5b** (∼10$^{-6}$ M) recorded in dichloromethane, upon addition of Cl$^-$ (0–500 equiv.). The inset shows enlarged view.

region and 561 and 594 nm for the Q bands region. These values are practically the same as those of the Cl⁻ titration.

For the zinc iodine–triazole complex [Zn(T$_{AzP}$-IVP)] (**5c**), the titrations with both the Cl⁻ and Br⁻ halide ions are very similar to each other and to those of the zinc chloro–triazole complex **5b** (Fig. 6 and 7). As mentioned above, the presence of isosbestic points is an indication of formation of a 1 : 1 receptor/anion adduct, *i.e.*, leading to a pentacoordinate complex type [Zn(Porph)(L)]. The redshift of the absorption bands of metalloporphyrins is due to the narrowing of the HOMO–LUMO and the energy gap is explained by the deformation of the porphyrin core.[46] This bathochromic effect is also related to the electron-withdrawing groups at the *meso* and β-pyrrolic positions of the porphyrin macrocycle and to the nature of the metal ion.[47,48]

In order to compare the Cl⁻ and Br⁻ detecting properties of our zinc complexes **5a–b** with those of the unsubstituted zinc(II)-*meso*-tetraphenylporphyrin ([Zn(TPP)]) we also performed a UV-visible titration in the Soret band region of this complex with Cl⁻ and Br⁻ halide ions. The spectra concerning these titrations are shown in Fig. SI-13.† The association constants values of the 1 : 1 complexes **5a–b** : X (X = Cl⁻ and Br⁻) as well as those of the [Zn(TPP)X] compounds are reported in Table SI-1.† To calculate these associate constants $K_{as}$ (and log $K_{as}$) we use the so-called "strong interactions" method[49] (see the ESI† for details).

A close inspection of Table SI-1† indicates that the iodine–triazole zinc derivative **5c** presents a better binding affinity than the chloro–triazole zinc metalloporphyrin **5b** for both Cl⁻ and Br⁻ anions. This could be explained by the higher donor effect of the chlorine in the *para* position of the T$_{AzP}$-ClVP porphyrinate than that of the iodine atom in the T$_{AzP}$-IVP moiety leading to a higher electron density on the zinc(II) center metal for **5b**. Therefore, the bonding affinity of zinc(II) in **5b** for the two halides is smaller than that of the iodine–triazole zinc T$_{AzP}$-IVP receptor. In contrast, the value of the log $K_{as}$ for the **5b** receptor for Cl⁻ is higher than that for Br⁻ which are 4.0245 and 3.5512, respectively. This is not the case for the **5c** receptor for which log $K_{as}$ value, in the case of the Br⁻ anion, is higher than that for the Cl⁻ anion; *i.e.* log $K_{as}$ is 4.6179 and 5.0566, respectively.

Notably, our two receptors present much better affinity for Cl⁻ and Br⁻ than the zinc(II) unsubstituted *meso*-tetraphenylporphyrin complex ([Zn(TPP)]) used as reference (Table SI-1†).

Beer *et al.*,[12] reported the use of a zinc(II) cage porphyrin complex [Zn(PC)] (PC = *meso*-tetrayltetrakis(carbonyloxy-methanediyl-1*H*-1,2,3-triazole-4,1-diylmethanediyl)tetraphenyl)-porphyrin) as receptor of several anions (such as F⁻, Cl⁻, Br⁻, I⁻ and SO$_4^{2-}$). This later complex presents a high binding affinity for Cl⁻ (log $K_{as}$ = 4.0860), but a very small association constant $K_{as}$ value (<50) for Br⁻. This can be explained by the small size of the porphyrin cage which is not accommodated to host the highly diffuse bromine ion. It is interesting to note (i) that our zinc(II) chlorine porphyrin derivative (**5b**) has a similar bonding affinity for Cl⁻ ion as the [Zn(PC)] related receptor with very close log $K_{as}$ values (4.0224 and 4.0860, respectively) and (ii) our iodine–triazole zinc derivative (**5c**) exhibits much better bonding affinity for Cl⁻ anion than the zinc-porphyrin cage

species ([Zn(PC)] with log $K_{as}$ value of 4.6179 for our **5c** complex compared to log $K_{as}$ value of 4.0860 [Zn(PC)].

## 2.7. Adsorption

The adsorption process is generally due to several physico-chemical forces that occur at the solid–liquid interface such as van der Waals forces,[50,51] hydrogen bonds[52] and hydrophobic interactions.[53,54] To determine the efficiency of adsorption of the MG dye on the **4a–c** free base porphyrins and the corresponding zinc(II) complexes **5a–c**, we carried out a kinetic study using our synthetic porphyrin derivatives as adsorbents. The adsorption spectrum of MG dye as a function of time in presence of **4a–c** and **5a–c** species are shown in Fig. SI-14.† The adsorption efficiency (R%) of the MG dye on **4a–c** and **5a–c** porphyrin compounds are depicted in Fig. 8. The R% values for the **4a–c** free base porphyrins are 35.5, 37.25 and 36.5%, while those for the **5a–c** complexes are 29, 31.75 and 30.75%, respectively. The fact that the adsorption efficiency of the free base porphyrins are higher than those of the corresponding [Zn(Porph)] complexes **5a–c** could be related to the pyrrole hydrogen atoms in **4a–c** leading to hydrogen bond type interactions with the nitrogens of the MG dye molecules. Notably, the chloro–triazole and iodine–triazole **4b–c** free base *meso*-arylporphyrin derivatives present better adsorption than that of the H-triazole T$_{AzP}$-HVP species (**4a**). This trend is also present in the corresponding zinc(II) porphyrin complexes (**5a–c**). This might result from the fact that the chlorine and iodine atoms in the case of the **4b–c** and **5b–c** species are involved in a n–π type interactions with the MG molecules, which is not possible in the case of the H-triazole **4a** and the chloro–triazole **5a** porphyrin species (Fig. SI-15†).[55] It should also be noticed that all **4a–c** and **5a–c** porphyrin compounds present π–π type interactions with the MG dye molecules (Fig. SI-16†). Furthermore, the nitrogen atom of the –(CH$_3$)$_2$N– group of the malachite green molecule is most likely coordinated to the center Zn$^{2+}$ ion as in the case of N-donor ligands (L) which react with the tetra-coordinated [Zn(Porph)] species leading to penta-coordinated complexes type [Zn(Porph)(L)].

## 2.8. Adsorption kinetic

Adsorption kinetic determines the time required to reach equilibrium between the solute and the adsorbent and it also gives an idea of the adsorption mechanism and the mode of transfer between the liquid and solid phases. Several kinetic models have been developed to describe the adsorption kinetics and to specify the nature of the interactions at the solid–liquid interface. In this work, four kinetic models have been selected to study the kinetic behavior of the MG dye at the surface of our porphyrinic compounds, namely: the pseudo first order kinetic model, the pseudo second order kinetic model, the intraparticle diffusion model and the Elovich model.[56] The adsorption capacity curves as function of time for our six porphyrinic compounds (**4a–c** and **5a–c**) are given in Fig. 9.

Fig. SI-17† illustrates the experimental data of the adsorption of MG dye on the **4a–c** and **5a–c** compounds. The data were fitted using the four kinetic models and the results are given in
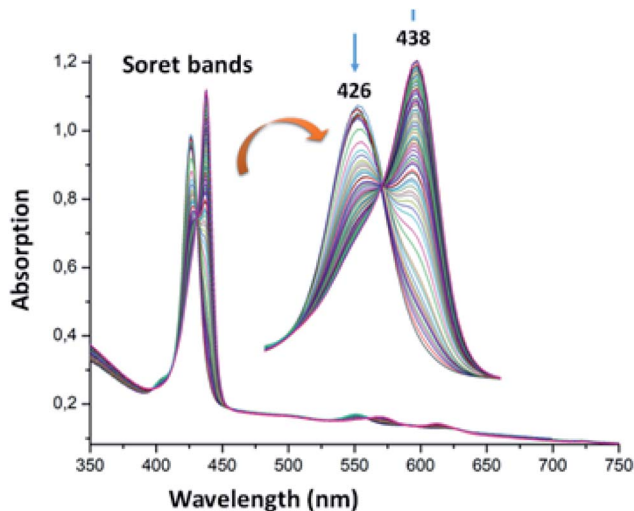
Fig. 5  Changes in the absorption spectra in the Soret band region of complex **5b** ($\sim 10^{-6}$ M) recorded in dichloromethane, upon addition of Br$^-$ (0–600 equiv.). The inset shows enlarged view.
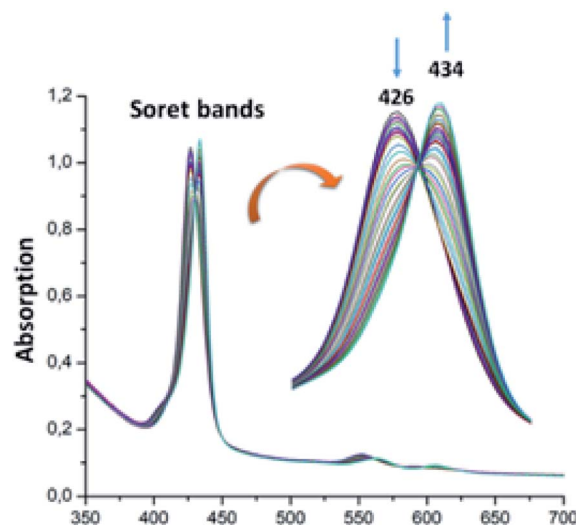


Fig. 7  Changes in the absorption spectra in the Soret band region of complex **5c** ($\sim 10^{-6}$ M) recorded in dichloromethane, upon addition of Br$^-$ (0–30 equiv.). The inset shows enlarged view.

Table SI-2.† The values of the amount of MG adsorption calculated from the pseudo second order model are the closest to those determined experimentally for all **4a–c** and **5a–c** porphyrin species, which indicates the adequacy of the use of this model to describe the adsorption phenomenon. The correlation coefficient ($R^2$) values of the pseudo second order model are close to 0.99. In the case of the pseudo first order model, the $R^2$ parameters are between 0.86 and 0.90 for all compounds. For the intraparticle diffusion model, the correlation coefficient does not exceed 0.91 and in the case of Elovich model, the $R^2$ values are between 0.93 and 0.97. These data indicate that the pseudo second order kinetic model is the most appropriate for describing the adsorption of MG dye by our porphyrin derivatives.

The second order model equation used is as follows (eqn (5)):

$$\frac{t}{q_t} = \frac{1}{(q_e^2 \times k_2)} + \frac{t}{q_e} \tag{5}$$

where, $q_t$ and $q_e$ represent the amounts of the adsorbent adsorbed at time $t$ and equilibrium, respectively, while $k_2$ is the rate constant for the pseudo second order model (g mg$^{-1}$ min$^{-1}$). The values of $k_2$ and $q_{cal}$ were calculated for each of the six porphyrinic compounds from the slope and the intersection of the corresponding plot, respectively.

### 2.9.   Degradation of MG dye

**2.9.1.   Effect of the initial MG concentration.** In order to obtain the optimal condition of the initial MG dye concentration, only the **5a** derivative was tested. The degradation
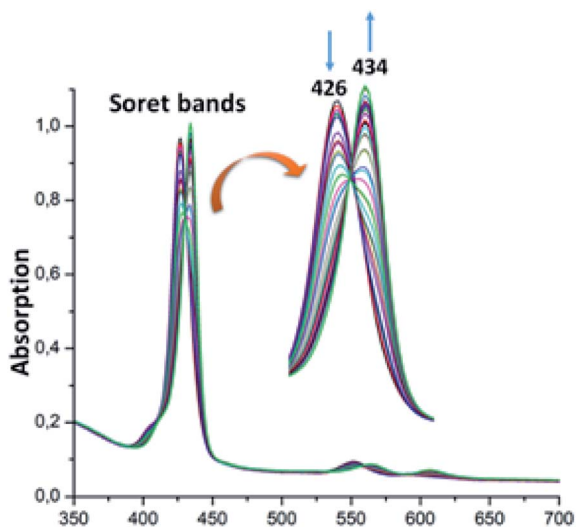


Fig. 6  Changes in the absorption spectra in the Soret band region of complex **5c** ($\sim 10^{-6}$ M) recorded in dichloromethane, upon addition of Cl$^-$ (0–30 equiv.). The inset shows enlarged view.
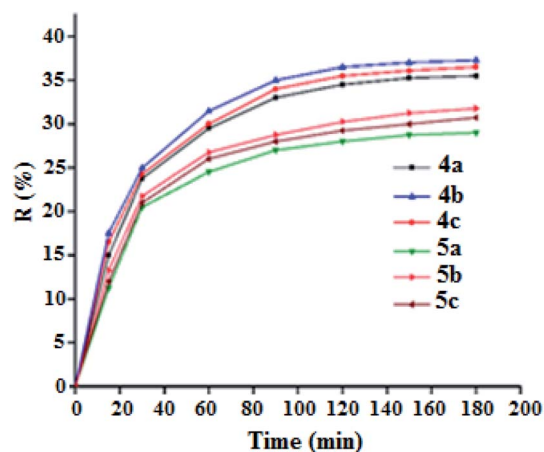


Fig. 8  Adsorption efficiency curves of malachite green dye ($C_0 = 20$ mg L$^{-1}$) as a function of time of the porphyrin compounds **4a–c** and **5a–c** ($m = 5$ mg, pH = 8).
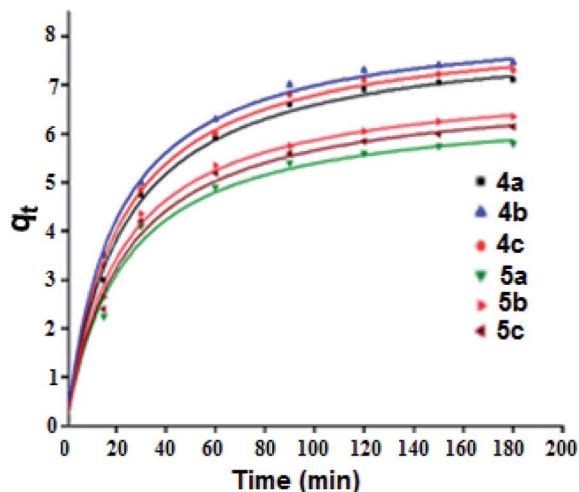
Fig. 9 Adsorption capacity of the MG dye ($C_0 = 20$ mg L$^{-1}$) as a function of time for the **4a–c** and **5a–c** porphyrin species ($m = 5$ mg, pH $= 8$). Dots represents the experimental data and the curves where plotted using the pseudo second-order kinetic model.



Fig. 11 Change in the degradation efficiency of the MG dye on varying initial H$_2$O$_2$ concentration using **5a** triazole zinc(II) derivative as a catalyst.

procedure of the MG dye consists of reacting the dye with H-triazole *meso*-arylporphyrin zinc(II) (**5a**) ($m = 5$ mg) in the presence of an aqueous solution of H$_2$O$_2$ (6 mg L$^{-1}$) at pH $= 8$. Fig. 10 shows the evolution of the color removal against initial MG dye concentration which shows that the degradation kinetics of MG is slow when the initial dye concentration is increased. This indicates that the increase of the initial concentration of the dye leads to an increase in the number of malachite green molecules, while the number of hydroxyl radicals remains constant. As a consequence, there is a decrease in the kinetics of the degradation reaction as well as the discoloration efficiency. These trends are consistent with those reported in the literature.[57]

**2.9.2. Effect of the initial H$_2$O$_2$ concentration.** The **5a** triazole zinc(II) derivative was also used to obtain the optimal condition for the initial hydrogen peroxide concentration.
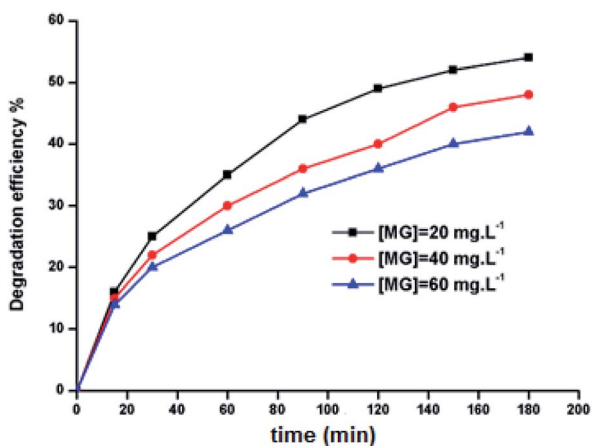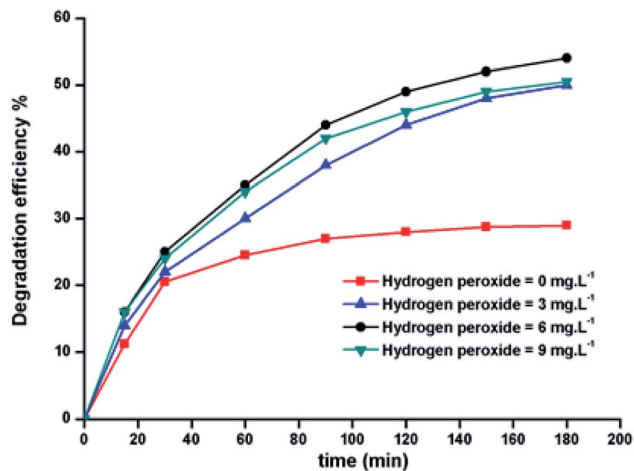
Thus, the effect of the initial hydrogen peroxide concentration $C_o$ on the malachite green dye degradation was studied using an initial MG concentration of 20 mg L$^{-1}$ at room temperature using **5a** compound as a catalyst. The $C_o$ initial H$_2$O$_2$ aqueous concentrations used were 0, 3, 6 and 9 mg L$^{-1}$. The increase in H$_2$O$_2$ concentration from 0 to 6 mg L$^{-1}$ leads to an increase in the degradation efficiency (Fig. 11). However, for a $C_o$ concentration of 9 mg L$^{-1}$, the discoloration efficiency of the MG decreases. This phenomenon could be explained by the fact that at high concentration, hydrogen peroxide is a powerful OH$^{\bullet}$ scavenger[58] (Equations (eqn (6)–(8)).

$$H_2O_2 + OH^{\bullet} \rightarrow HO_2^{\bullet} + H_2O \tag{6}$$

$$H_2O_2 + OH_2^{\bullet} \rightarrow OH^{\bullet} + O_2 + H_2O \tag{7}$$

$$HO_2^{\bullet} + OH^{\bullet} \rightarrow H_2O + O_2 \tag{8}$$



Fig. 10 Evolution of the color removal against initial dye concentration. The hydrogen peroxide concentration, $C_o$ is 6 mg L$^{-1}$, **5a** catalyst weight $m = 5$ mg, pH $= 8$ and the MG mass, $m_o$ is 5 mg.
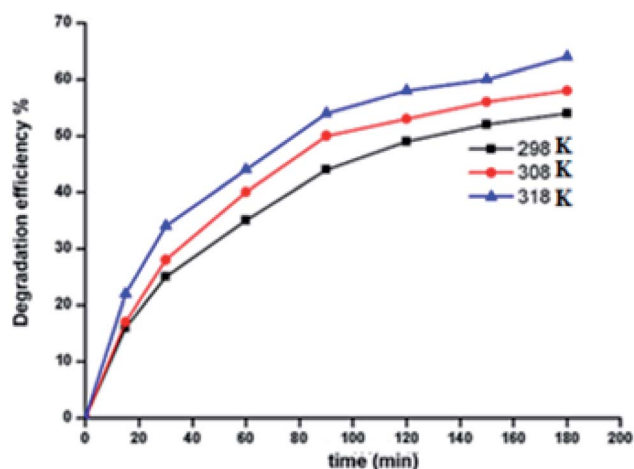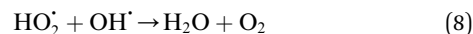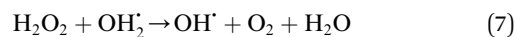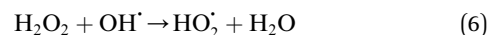


Fig. 12 Change of the degradation efficiency of the MG solution as function of time using **5a** used as catalyst at 298, 308 and 318 K.

**2.9.3. Effects of pH.** To investigate the effects of pH on the degradation of the MG dye by complexes **5a–c**, aqueous solutions with pH from 4 to 8 were adjusted by 0.1 M HCl and 0.1 NaOH. The influence of pH on the dye oxidation was studied using five solutions with pH values determined initially (4, 6, 8, 10 and 12) and without any modifications or control of the pH during the process. The results obtained for dye removal as a function of the initial pH of the solution at various reaction times are presented in Fig. SI-18.† The maximum conversion was achieved after 3 h from the start of the reaction for a pH of 8.

**2.9.4. Degradation with optimal conditions.** The degradation study of the MG dye was carried out using an aqueous $H_2O_2$ solution under optimal conditions, found for the **5a** derivative, and applied to the two other species **5b–c** (Fig. SI-19b and c†).

Fig. SI-19a† represents the absorption curve of the MG dye in presence of $H_2O_2$ without catalyst (blank experiment) *versus* time where it can be seen that the absorption remains practically the same for a period of 180 min. By using the same mass of the three catalysts **5a–c**, the obtained degradation yields are 54.0, 52.1 and 48.9% for **5a**, **5b** and **5c**, respectively (Fig. SI-20a†). The degradation reactions were achieved after 180 min. Fig. SI-20b† shows the degradation removal efficiency for complexes **5a–c** using the same number of moles ($3.35 \times 10^{-6}$ mol.) for the three species. The obtained degradation yields are 54.0, 53.88 and 53.94% for **5a**, **5b** and **5c**, respectively. These results show that using the same number of moles for our three **5a–c** catalysts, the degradation efficiency is virtually the same ($\sim$54%). This confirms the fact that the central metal is the main site of degradation of the dye[58] and that the nature of the substituent X (X = H, Cl and I) on the *para*-position of the

**Table 5** Selection of several methods used for MG degradation with the optimal reaction conditions and yields

| Method of degradation | Degradation system and optimal reacting conditions | Degradation yield, time reaction | Ref. |
|---|---|---|---|
| Chemical method: Fenton's reagent | Fe(II)/aqueous $H_2O_2$, pH = 3.4, $[H_2O_2]_o = 0.50$ mM, $[Fe^{2+}] = 0.10$ mM, $[MG]_o = 20$ mg $L^{-1}$, 30 °C | 99.25% (60 min) | 60 |
| Chemical method: mesoporous carbon adsorbent | Mesoporous silica (MCM-48), pH = 8.5, [silica] = $5.10^{-3}$ g/25 mL, $[MG]_o$ = 50 mg $L^{-1}$, $T$ = 25 °C | 98.2% (30 min) | 61 |
| Photocatalysis method | Using naked niobium oxide ($Nb_2O_5$) as a photocatalyst. weight of catalyst = 0.1 g, $[MG]_o$ = 5 ppm; $T$ = 25 °C, 3 mV cm$^{-2}$ light intensity, pH = 6 | 53.25% (60 min) | 62 |
| Photocatalysis method | Ag doped $TiO_2$, pH = 9, $[MG]_o = 5 \times 10^{-6}$ M, [catalyst] = 0.12 g, 60 mW cm$^{-2}$ light intensity | 99.5% | 63 |
| Biodegradation method | System used: bacterium *Enterobacter asburiae* strain XJUHX-4TM, semi-synthetic medium having the following composition was used: MG: 0.100–1.00 g $L^{-1}$, $KH_2PO_4$: 1 g $L^{-1}$, KCl: 0.5 g $L^{-1}$, $NaNO_3$: 2 g $L^{-1}$, $MgSO_4 \cdot 7 H_2O$: 0.5 g $L^{-1}$, $FeSO_4 \cdot 7H_2O$: 0.01 g $L^{-1}$, sucrose: 15 g $L^{-1}$, and beef extract: 3 g $L^{-1}$ | Maximum decolorization (>98%) in semi-synthetic media | 64 |
| Biodegradation method | System used: *Pseudomonas* sp. strain DY1, $[MG_o]$ = 100–1000 mg $L^{-1}$, pH = 6.6, $T$ = 28–30 °C | Effective decolorization: 78.9–84.3% | 65 |

Use of our three zinc(II) complexes **5a–c**

| Complex | System used | Yield |
|---|---|---|
| $[Zn(T_{AzP}\text{-HVP})]$ **5a** | $[H_2O_2] = C_o = 6$ mg $L^{-1}$, [MG] = 20 mg $L^{-1}$, weight of catalyst (**5a**) = $m_o$ = 5 mg, pH = 8, $T$ = 25 °C | 54.0% (180 min) |
| $[Zn(T_{AzP}\text{-ClVP})]$ **5b** | $[H_2O_2] = C_o = 6$ mg $L^{-1}$, [MG] = 20 mg $L^{-1}$, weight of catalyst (**5b**) = $m_o$ = 5 mg, pH = 8, $T$ = 25 °C | 52.0% (180 min) |
| $[Zn(T_{AzP}\text{-IVP})]$ **5c** | $[H_2O_2] = C_o = 6$ mg $L^{-1}$, [MG] = 20 mg $L^{-1}$, weight of catalyst (**5c**) = $m_o$ = 5 mg, pH = 8, $T$ = 25 °C | 48.9% (180 min) |

terminal phenyl of the three triazole *meso*-aryporphyrins **4a–c**, practically has no effect on this degradation.

**2.9.5. Effect of temperature.** For the zinc(II) porphyrin derivative **5a**, the effect of temperature on the degradation of MG dye has been investigated. The temperatures used were 25, 35 and 45 °C (Fig. 12). As expected, the degradation efficiency of MG dye increased from 54.0% to 64.2% as a consequence of increasing the temperature from 25 to 45 °C. This is due to the fact that higher temperatures increase the reaction rate between hydrogen peroxide and the zinc metal center. This in turn increases the rate of generation of the OH˙ radical oxidizing species.[59]

**2.9.6. Comparison of our procedure with other reported methods.** Many investigations concerning the MG degradation have previously been reported in the literature.[19] The most important of which are: the chemical methods, the photo-catalysis methods and the biodegradation methods (Table 5).

As depicted in Table 5, the degradation yields of the MG dye obtained using our zinc(II) **5a–c** (∼50%) porphyrin zinc(II) complexes are lower than those obtained using the other methods shown in this table. It is expected that the use of other metal ions instead of Zn(II), such as Cu(II) or Fe(II)[60] with our three triazole porphyrins in the presence of $H_2O_2$ will lead to an increase in the degradation efficiency of the dye. This is due to the fact that ferrous and cuprous ions are more effective than $Zn^{2+}$ ion in giving $HO_2^˙$ and OH˙ radicals.

Nevertheless, the obtained yields are quite acceptable for a first reported investigation on the MG degradation using a porphyrin derivative. Thus, these encouraging results suggest that our zinc(II) **5a–c** can be further improved and can be concomitantly used with other degradation techniques such as the photodegradation where the expected yields are higher than those reported here.

**2.9.7. Activation energy and thermodynamic variables.** To better understand the degradation phenomenon, the pseudo first order model was used to calculate the kinetic parameters. Arrhenius law (eqn (9)) was used to determine the activation energy ($E_a$). The thermodynamic activation parameters of enthalpy ($\Delta H^*$), entropy ($\Delta S^*$) and Gibbs free energy ($\Delta G^*$) were calculated using Eyring's equation (eqn (10) and (11)).[66]

$$\ln k_o = \ln A_o - \frac{E_a}{RT} \qquad (9)$$

$$\ln\left(\frac{k_o}{T}\right) = \ln\left(\frac{k}{h}\right) + \frac{\Delta S^*}{R} - \frac{\Delta H^*}{RT} \qquad (10)$$

$$\Delta G^* = \Delta H^* - T\,\Delta S^* \qquad (11)$$

where $k_o$ is the kinetic rate, $h$ and $k$ are the Plank's and Boltzmann's constants, respectively. $A_o$ is the Arrhenius constant and $R$ is the universal gas constant (8.314 J mol$^{-1}$ K$^{-1}$). The obtained thermodynamic parameters and activation energy values are summarized in Table 6. From these results, it can be seen that the free energy is positive in all the cases. This indicates that the degradation of MG dye using $H_2O_2$ is a non-spontaneous reaction regardless of temperature. The free enthalpy is positive,

**Table 6** Thermodynamic parameters ($\Delta S^*$, $\Delta H^*$, $\Delta G^*$) and activation energy ($E_a$) for complex **5a**

| $T$ (°C) | 25 | 35 | 45 |
|---|---|---|---|
| $k_o$ (min$^{-1}$) | 0.0042 | 0.0046 | 0.0054 |
| $E_a$ (kJ mol$^{-1}$) | 9.867 | 9.867 | 9.867 |
| $\Delta S^*$ (J mol$^{-1}$ K$^{-1}$) | −265.983 | −265.983 | −265.983 |
| $\Delta H^*$ (kJ mol$^{-1}$) | 7.308 | 7.308 | 7.308 |
| $\Delta G^*$ (kJ mol$^{-1}$) | 86.570 | 89.230 | 91.890 |

meaning the degradation process is endothermic and a higher temperature facilitates degradation. The negative value of the entropy suggests the diminution of the disorder during the degradation process. The activation energy value is 9.867 kJ mol$^{-1}$. Since the thermodynamic parameters found for both **5b–c** complexes are very close to those of complex **5a**, only the results founds for the latter zinc(II) derivative are reported.

## 3. Conclusion

We have successfully synthesized three novel triazole *meso*-arylporphyrins, the: H-triazole, chloro–triazole and iodine–triazole *meso*-arylporphyrins with the respective formulas $H_2$($T_{AzP}$-HVP) (**4a**), $H_2$($T_{AzP}$-ClVP) (**4b**) and $H_2$($T_{AzP}$-IVP) (**4c**) as well as their corresponding zinc(II) porphyrins **5a–c**. The chloride and bromide binding properties of the chloro–triazole and the iodine–triazole zinc(II) triazole *meso*-arylporphyrins derivatives **5b–c** have been monitored by UV-visible technique. Both **5b–c** receptors exhibit strong Cl$^-$ and Br$^-$ anion binding affinities, forming 1 : 1 stoichiometric complexes. The adsorption efficiency of the malachite green dye (MG) on all three base porphyrins and their corresponding zinc(II) coordination compounds revealed that: (i) the adsorption efficiency of the free base porphyrins **4a–c** are higher than those of the corresponding zinc(II) porphyrin complexes **5a–c**, (ii) the free base chloro–triazole and iodine–triazole **4b–c** porphyrin derivatives show better adsorption than that of the H-triazole $T_{AzP}$-HVP species (**4a**), which is also the case for the corresponding zinc(II) porphyrin complexes (**5a–c**) and (iii), the pseudo second order kinetic model is the most appropriate to describe the adsorption of MG dye by our free porphyrin compounds. The degradation investigation of the MG dye, using complexes **5a–c** as catalysts, carried out using an aqueous $H_2O_2$ solution under optimal conditions, shows that the degradation yield using the H-triazole *meso*-arylporphyrin zinc(II) derivative **5a** is higher than those of the chloro–triazole and the iodine–triazole *meso*-arylporphyrin zinc(II) complexes **5b–c**. The degradation yields using our **5a–c** species are ∼54% which are much smaller than several reported MG dye degradations using other methods. Nevertheless, this is an encouraging result as this is the first time a porphyrin derivative has been used for the degradation of this dye and studies are underway to test our zinc(II) derivatives in photochemical degradations of the malachite green and other dyes. The thermodynamic parameters were determined and show that the free energy is positive in all the cases indicating that the degradation of MG dye using $H_2O_2$ is a non-

spontaneous, temperature-dependent reaction. Furthermore, the enthalpy is positive which implies that the degradation process is endothermic, while the negative value of the entropy suggests the diminution of disorder during the degradation process.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

## References

1 M. Pineiro, A. L. Carvalho, M. M. Pereira, A. M. d'A. R. Gonsalves, L. G. Arnaut and S. J. Formosinho, *Chem.–Eur. J.*, 1998, 2299–2307.

2 X. I. Liang, X. Li and X. Yue, *Angew. Chem., Int. Ed.*, 2011, **50**, 11622–11627.

3 S. Lin, C. S. Diercks, Y.-B. Zhang, N. Kornienko, E. M. Nichols, Y. Zhao, A. R. Paris, D. Kim, P. Yang, O. M. Yaghi and C. J. Chang, *Science*, 2015, **349**, 1208–1213.

4 H. Imahori, S. Hayashi, H. Hayashi, A. Oguro, S. Eu, T. Umeyama and Y. Matano, *J. Phys. Chem. C*, 2009, **113**, 18406–18413.

5 M. Florescu and M. David, *Sensors*, 2017, **17**, 1–16.

6 A. Colombelli, M. G. Manera, V. Borovkovb, G. Giancanec, L. Valli and R. Rell, *Sens. Actuators, B*, 2017, **246**, 1039–1048.

7 E. Aguilar-Ortíz, N. Lévaray, M. Vonlanthen, E. G. Morales-Espinoza, Y. Rojas-Aguirre, X. X. Zhu and E. Rivera, *Dyes Pigm.*, 2016, **132**, 110–120.

8 B. M. J. M. Suijkerbuijk and R. J. M. Gebbink, *Angew. Chem., Int. Ed. Engl.*, 2008, **47**, 7396–7421.

9 W. Auwärter, D. Écija, F. Klappenberger and J. V. Barth, *Nat. Chem.*, 2015, **7**, 105–120.

10 J. M. Gottfried, *Surf. Sci. Rep.*, 2015, **70**, 259–279.

11 H. C. Kolb, M. G. Finn and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2001, **40**, 2004–2021.

12 L. C. Gilday, N. G. White and P. D. Beer, *Dalton Trans.*, 2012, **41**, 7092–7097.

13 L. C. Gilday, N. G. White and P. D. Beer, *Dalton Trans.*, 2013, **42**, 15766–15773.

14 M. Ismail, K. Akhtar, M. I. Khan, T. Kamal, M. A. Khan, A. M. Asiri, J. Seo and S. B. Khan, *Curr. Pharm. Des.*, 2019, **25**, 3653–3671.

15 K. Su Min, R. Satish Kumar, J.-H. Lee, K. S. Kim, S. G. Lee and Y.-A. Son, *Dyes Pigm.*, 2019, **160**, 37–47.

16 Y. Li, L. Wang, Y. Gao, W. Yang, Y. Lia and C. Guo, *RSC Adv.*, 2018, **8**, 7330–7339.

17 M. Guergueb, S. Nasri, J. Brahmi, F. Loiseau, F. Molton, T. Roisnel, V. Guerineau, I. Turowska-Tyrk, K. Aouadi and H. Nasri, *RSC Adv.*, 2020, **10**, 6900–6918.

18 R. Soury, M. Jabli, T. A. Saleh, W. S. Abdul-Hassan, E. Saint-Aman, F. Loiseau, C. Philouze and H. Nasri, *RSC Adv.*, 2018, **8**, 20143–20156.

19 P. N. P. Raval, P. U. Shah and N. K. Shah, *Appl. Water Sci.*, 2017, **7**, 3407–3445.

20 S. Srivastava, R. Sinha and D. Roy, *Aquat. Toxicol.*, 2004, **66**, 319–329.

21 S.-Y. An, S.-K. Min, I.-H. Cha, Y.-L. Choi, Y.-S. Cho, C.-H. Kim and Y.-C. Lee, *Biotechnol. Lett.*, 2002, **24**, 1037–1040.

22 J. Brahmi, K. Aouadi, M. Msaddek, J.-P. Praly and S. Vidal, *C. R. Chim.*, 2016, **19**, 933–935.

23 A. D. Adler, F. R. Longo, J. D. Finarelli, J. Goldmacher, j. Assour and L. J. Korsakoff, *Org. Chem.*, 1967, **32**, 476.

24 S. Nasri, I. Zahou, I. Turowska-Tyrk, T. Roisnel, F. Loiseau, E. Saint-Amant and H. Nasri, *Eur. J. Inorg. Chem.*, 2016, 5004–5019.

25 K. Görlitzer, S. Huth and P. G. Jones, *Pharmazie*, 2005, **60**, 269–272.

26 K. Ezzayani, Z. Denden, S. Najmudin, C. Bonifácio, E. Saint-Aman, F. Loiseau and H. Nasri, *Eur. J. Inorg. Chem.*, 2014, 5348–5361.

27 C. C. Mak, N. Bampos and J. K. M. Sanders, *Angew Chem. Int. Ed.*, 1998, **37**, 3020–3023.

28 H. M. Zeyada, M. M. Makhlouf and M. A. Ali, *Jpn. J. Appl. Phys., Part 1*, 2016, **55**, 022601.

29 C.-I. Lin, M.-Y. Fang and S.-H. Cheng, *J. Electroanal. Chem.*, 2002, **531**, 155–162.

30 P. Tao, A. Viswanath, L. S. Schadler, B. C. Benicewicz and R. W. Siegel, *ACS Appl. Mater. Interfaces*, 2011, **3**, 3638–3645.

31 P. Singh, A. Kaushal and D. Kaur, *J. Alloys Compd.*, 2009, **471**, 11–15.

32 K. M. Barkigia, M. D. Berber, J. Fajer, C. J. Medforth, M. W. Renner and K. M. Smith, *J. Am. Chem. Soc.*, 1990, **112**, 8851–8854.

33 J. Zhang, P. Zhang, Z. Zhang and X. J. Wei, *J. Phys. Chem. A*, 2009, **113**, 5367–5374.

34 M. Gouterman, *J. Mol. Spectrosc.*, 1961, **6**, 138–163.

35 C. C. Mak, N. Bampos and J. K. M. Sanders, *Angew. Chem., Int. Ed.*, 1998, **37**, 3020–3022.

36 Z. Denden, K. Ezzayani, E. Saint-Aman, F. Loiseau, S. Najm, Bonifácio, J.-C. Daran and H. Nasri, *Eur. J. Inorg. Chem.*, 2015, 2596–2610.

37 D. Kim and E. J. Shin, *Bull. Korean Chem. Soc.*, 2003, **24**, 1490–1494.

38 S. Neya and N. Funasaki, *Tetrahedron Lett.*, 2002, **43**, 1057–1058.

39 D. J. Quimby and F. R. Longo, *J. Am. Chem. Soc.*, 1975, **97**, 5111–5117.

40 K. M. Kadish and M. M. Morrison, *J. Am. Chem. Soc.*, 1976, **98**, 3326–3328.

41 C.-I. Lin, M.-Y. Fang and S.-H. Cheng, *J. Electroanal. Chem.*, 2002, **531**, 155–162.

42 A. Ghosh, S. M. Mobin, R. Fröhlich, R. J. Butcher, D. K. Maity and M. Ravikanth, *Inorg. Chem.*, 2010, **49**, 8287–8297.

43 J. L. Bredas, R. Silbey, D. S. Bordeaux and R. R. Chance, *J. Am. Chem. Soc.*, 1983, **105**, 6555–6559.

44 D. M. Lyons, J. Kesters, W. Maes, W. Christopher, J. Bielawski and L. Sessler, *Synth. Met.*, 2013, **178**, 56–61.

45 K. M. Kadish, P. Chen, Y. Yu. Enakieva, S. E. Nefedov, Y. G. Gorbunova, A. Y. Tsivadze, A. Bessmertnykh-Lemeune, C. Stern and R. Guilard, *J. Electroanal. Chem.*, 2011, **656**, 61–71.

46 K. M. Barkigia, L. Chantranupong, K. M. Smith and J. Fajer, *J. Am. Chem. Soc.*, 1988, **110**, 7566–7567.

47 J. A. Shelnutt, in *The Porphyrin Handbook*, ed. K. M. Kadish, K. M. Smith and R. Guilard, Academic Press, San Diego, 2000, vol. 7, pp. 167–223.

48 A. Ozarowski, H. M. Lee and A. L. Balch, *J. Am. Chem. Soc.*, 2003, **125**, 12606–12614.

49 J. Polster and H. Lachmann *Spectrometric Titrations*, Verlag Chemie, Weinheim, 1989, p. 292.

50 S. U. Khan, *Fundamental aspects of pollution control and environmental Science*, Elsevier, New York, 1980.

51 N. Senesi and Y. Chen, *Ecological Studies*, ed. Z. Gerstl, Y. Chen, U. Mingelgril and B. Yaron, Sringer verglag, Berlin, 1989, pp. 37–90.

52 S. Chowdhury and P. Das, *Sep. Sci. Technol.*, 2011, **46**, 1966–1976.

53 D. A. Dzombak and R. G. Luthy, *Soil Sci.*, 1984, **137**, 292–308.

54 C. T. Chiou, L. J. Peters and V. H. Freed, *Science*, 1979, **206**, 831–832.

55 N. B. Swan and M. A. A. Zaini, *Ecol. Chem. Eng. S*, 2019, **26**, 119–132.

56 H. Qui, L. Lv, B.-C. Pan, Q.-J. Zhang, W.-M. Zhang and Q.-X. Zhang, *J. Zhejiang Univ., Sci., A*, 2009, **10**, 716–724.

57 B. H. Hameed and T. W. Lee, *J. Hazard. Mater.*, 2009, **164**, 468–472.

58 R. Aravindhan, N. N. Fathima, J. R. Rao and B. U. Nair, *J. Hazard. Mater.*, 2006, **B138**, 152–159.

59 J. H. Sun, S. P. Sun, G. L. Wang and L. P. Qiao, *Dyes Pigm.*, 2007, **74**, 647–652.

60 B. H. Hameed and T. W. Lee, *J. Hazard. Mater.*, 2009, **164**, 468–472.

61 M. Anbia and A. Ghaffari, *J. Iran. Chem. Soc.*, 2012, **8**, 67–76.

62 A. S. Hussein and N. Y. Fairooz, *Journal of University of Babylon for Pure and Applied Sciences*, 2016, **9**, 2510–2518.

63 H. R. Khan, M. A. Choudhary, M. A. Mirza, Z. Ahmed and Y. Khan, *Asian J. Chem.*, 2017, **29**, 257–260.

64 T. Mukherjee and M. Das, *Clean: Soil, Air, Water*, 2014, **42**, 849–856.

65 L.-N. Du, S. Wang, G. Li, B. Wang, X.-M. Jia, Y.-H. Zhao and Y.-L. Chen, *Ecotoxicology*, 2011, **20**, 438–446.

66 B. A. Fil, M. T. Yilmaz, S. Bayar and M. T. Elkoca, *Braz. J. Chem. Eng.*, 2014, **31**, 171–182.