



Published in final edited form as:

Exp Dermatol. 2022 May ; 31(5): 781–788. doi:10.1111/exd.14521.

The disturbed expression of vitamin D and retinoic acid-related orphan receptors α and γ and of megalin in inflammatory skin diseases

Anna A Brozyna¹, Michał A mijewski², Kinga Linowiecka¹, Tae-Kang Kim³, Radomir M Slominski^{3,4}, Andrzej T Slominski^{3,5}

¹Department of Human Biology, Institute of Biology, Faculty of Biological and Veterinary Sciences, Nicolaus Copernicus University, Toruń, Poland.

²Faculty of Medicine, Department of Histology, Medical University of Gdańsk, Gdańsk, Poland.

³Department of Dermatology, University of Alabama at Birmingham, Birmingham, Alabama, USA.

⁴Graduate Biomedical Sciences Program, University of Alabama at Birmingham, Birmingham, Alabama, USA.

⁵Laboratory Service, VA Medical Center at Birmingham, Birmingham, Alabama, USA.

Abstract

The pathogenesis of inflammatory skin diseases is associated with the abnormal activity of keratinocytes and immune cells infiltrate. Vitamin D₃ deficiency can correlate with the increased incidence, severity, and duration of inflammatory skin disorders. The exact mechanism on how vitamin D₃ influences inflammatory skin diseases still requires clarification. However, it can be associated with the disturbances in transmembrane glycoprotein - LRP2/megalín, which is implicated in vitamin D₃ transport to the cell, and defects in vitamin D-signaling through the nuclear receptors. Therefore, by using immunohistochemistry, we analyzed the expression of LRP2/megalín, VDR, ROR α and ROR γ in allergic contact dermatitis, lichen simplex chronicus, sarcoidosis and psoriasis in comparison to the normal skin. We observed decreased expression of LRP2/megalín in all inflammatory lesions in comparison to the normal skin. Significant differences were also noticed in VDR, ROR α and ROR γ levels between inflammatory lesions and normal skin. Our research indicates disturbed expression of LRP2/megalín, VDR, ROR α and ROR γ in inflammatory skin lesions in comparison to normal skin. Therefore, we suggest that changes in the activity of these proteins may play role in pathogenesis of inflammatory skin disorders. Furthermore, we suggest that LRP2/megalín, VDR, ROR α and ROR γ may serve as targets in therapy of these diseases.

Corresponding authors: anna.brozyna@umk.pl (AAB) and aslominski@uabmc.edu (ATS).

Authors' contributions

AAB - designed the study, performed experiments, evaluated immunohistochemically stained samples, interpreted the data, and wrote the paper; M , KL, RMS - took part in data collection/analysis and preparation of the manuscript; A.T.S. designed the study, provided the material for the experiments, coordinated the experiments, analyzed data, worked on the manuscript, and secured the funding. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Keywords

vitamin D; nuclear receptors; megalin; sarcoidosis; lichen simplex chronicus; psoriasis; allergic contact dermatitis

Background

The skin serves as a barrier between the environment and the inner part of the organism, and plays a crucial role in the defense against pathogens, chemicals, and/or physical stressors, as well as maintaining the body's homeostasis¹. The skin is also a significant part of immune system². It is engaged in the development of anti-microbial resistance, autoimmunity and inflammatory skin conditions, allergies or tumor immunity³. The cutaneous immune surveillance includes innate and adaptive immune responses⁴.

There is increased evidence that the impaired regulation of immune skin functions underlies the pathogenesis of wide range of cutaneous disorders. For example, psoriasis is one of the most commonly diagnosed inflammatory skin diseases⁵ with worldwide prevalence of about 2%⁶. This disorder manifests as inflammatory lesions with epidermal hyperplasia and immune cells infiltration⁷. One commonly used treatment for psoriasis, ultraviolet B phototherapy, inhibits the inflammatory responses, while stimulating vitamin D formation in the skin⁸. Vitamin D₃ is activated by two-step hydroxylation catalyzed by cytochrome P450 enzymes including CYP2R1 and CYP27A1 with final C-1 α hydroxylation by CYP27B1, to produce 1,25(OH)₂D₃, or by sequential hydroxylation of the side chain by CYP11A1^{9, 10}. These pathways are also expressed in the skin^{11, 12} and immune system¹³.

About 40% of the European population suffer from vitamin D₃ deficiency¹⁴, which can be linked to autoimmune disorders¹⁵. Vitamin D₃ has immunomodulatory effects by regulation of T cells^{16–18} and is implicated in the regulation of both the innate and adaptive immunity¹⁶. Several studies have shown a relationship between decreased levels of vitamin D₃ and incidence of autoimmune skin disorders^{19–21}, severity of psoriasis^{22–24} and atopic dermatitis²⁵. In addition, polymorphism in vitamin D receptor (VDR) gene has been implicated in both severity²⁶ and susceptibility^{27,28} to psoriasis.

Vitamin D₃ is transported by the group-specific component vitamin D binding protein (GC/DBP)²⁹. GC/DBP is a ligand for LRP2/megalyn (low density lipoprotein-related protein 2), which is a transmembrane glycoprotein³⁰. LRP2/megalyn is involved in reabsorption and further metabolism of chemicals and small proteins, including vitamins transfer proteins³¹. Previous research has indicated an unambiguous relationship between LRP2/megalyn and vitamin D₃ endocytosis³² and activation of genomic and non-genomic responses³³. LRP2/megalyn is implicated in regulation of 25(OH)D uptake by kidneys³². It is therefore likely that there is connection between LRP2/megalyn expression and vitamin D in psoriasis development.

Questions addressed

To exert the phenotypic activities, active forms of vitamin D₃ must interact with VDR^{11, 34}. More recently, it was reported that retinoic acid-related orphan receptors α and γ (ROR α

and ROR γ , coded by *RORA* and *RORC* genes, respectively) are targets for regulation by vitamin D₃ hydroxyderivatives³⁴, acting as inverse agonists on ROR α and ROR γ . ROR α and ROR γ are expressed both in normal and pathological skin^{34, 35}. RORs are implicated in wide variety of cell functions³⁶, and can play a role in inflammatory skin disorders³⁷. Therefore, to gain an insight into vitamin D endocrine system in inflammatory skin diseases, we analyzed the expression of LRP2/megalin, VDR, ROR α and ROR γ in psoriasis, allergic contact dermatitis (ACD), lichen simplex chronicus (LSC) and sarcoidosis.

Experimental design

For immunohistochemistry skin samples of ACD, LSC, sarcoidosis, psoriasis and normal skin was included into this study (Table 1). The authors declare that this investigation was carried out following the rules of the Declaration of Helsinki of 1975 (revised in 2008) and this study was approved by the Institutional Review Board of the UAB under IRB-940831016 (OCCC Tissue Procurement CORE Facility) and IRB-00000726 (Title E150427002, Dr. A. Slominski PI). The IRB of the UAB, which gave their permission for conducting this study, waived the requirement to obtain Patients' informed consent for this research.

The samples were stained for LRP2/megalin, VDR, ROR α and ROR γ antigens as previously described^{34, 35, 38, 39}. Detailed data are presented in the Supplementary file and Supplementary Table 1. The immunostained sections were assessed using semiquantitative scoring systems using both the percentage of immunoreactive cells (IR) and the staining intensity (SI) from 0 to 3 arbitrary units (A.U.) according to the following formula $SQ = \text{mean}(\text{IR} \times \text{SI})/100$. Statistical comparison of subgrouped data were performed with GraphPad Prism software (version 5.0; GraphPad Software, La Jolla, CA, USA).

Testing of the expression of VDR, ROR α and ROR γ in HaCaT keratinocytes was performed using western blot according to the procedure presented in Supplementary file.

The analysis of the expression of VDR, ROR α , ROR γ and LRP2/megalin on mRNA level followed public genomics data repository (<https://www.ebi.ac.uk/gxa/home>) and project characterized in ⁴⁰ (for details see Supplementary file). The comparisons between the psoriatic individuals and healthy controls were performed using t-test with $P < 0.05$ considered to indicate a statistically significant difference.

Results

The expression of all tested markers was detected using immunohistochemistry in skin compartments. VDR showed predominant nuclear immunostaining, ROR α and ROR γ showed strong nuclear and weak cytoplasmic immunostaining, and LRP2/megalin showed cytoplasmic location. The representative images of immunostained sections are in Fig. 1. The strong nuclear stain vs weak cytoplasmic was further validated by western blot using antibodies against VDR, ROR α and ROR γ (Supplementary Figure 1).

In ACD, the reduced levels of VDR, ROR α and ROR γ and LRP2/megalin in lesional skin were observed in comparison to normal skin (Fig. 2). The expression levels of analyzed receptors in lymphocytes was higher than in lesional keratinocytes (Fig. 2A).

In LSC the lower levels of VDR, ROR α , ROR γ and LRP2/megalin were found in comparison to normal skin (Fig. 2A). In lymphocytes VDR levels were low and comparable to that observed in lesional epidermis, while ROR γ expression was as high as in normal epidermis (Fig. 1).

In sarcoidosis, the expression of ROR α and ROR γ in lesional skin and lymphocytes was as high as in normal tissue, while in granulomas ROR α and ROR γ levels were reduced when compared to lymphocytes. VDR and LRP2/megalin levels were significantly reduced in pathological cells and granulomas (Fig. 1). VDR levels in lymphocytes were also reduced in comparison to normal skin.

Levels of VDR and ROR α were similar in psoriatic and normal (control) skin (Fig. 2A). ROR γ levels in psoriatic keratinocytes were comparable to normal epidermis, being elevated in lymphocytes (Fig. 1 2A). However, LRP2/megalin was significantly reduced in lesional in comparison to the normal skin (Fig. 2A). Analysis of the data from Expression Atlas (<https://www.ebi.ac.uk/gxa/home>) has shown similar expression of *VDR* mRNA in psoriasis in comparison to normal skin, with expression of *RORA* and *RORC* and *LRP2/megalin* mRNA being lower in lesional than in normal skin (Fig. 2B). The gene expression pattern for the *VDR* and *LRP2/megalin* was consistent with the corresponding protein expression levels.

Conclusions & perspectives

In the present study, we investigated the expression of VDR, ROR α and ROR γ and LRP2/megalin in inflammatory skin diseases to determine the expression pattern for these receptors and their potential to serve as a predictive targets or markers for vitamin D-based therapies. On the protein level, we observed the reduced expression of LRP2/megalin in all analyzed skin lesions, the reduced ROR α and ROR γ level in ACD and LCS, the reduced VDR level in lichen simplex chronicus, ACD and sarcoidosis. The immunostaining pattern in all analyzed lesions and normal skin for VDR, ROR α and ROR γ and LRP2/megalin was similar. Up to now, there is shortage of information on the expression of analyzed proteins in the skin samples of inflammatory cutaneous diseases. To the best of our knowledge, our data is the first time that describes the expression of ROR α and LRP2/megalin in inflammatory skin diseases and ROR γ in skin samples of these diseases.

Some studies reported that psoriasis patients have reduced vitamin D3 levels^{24, 41}. Vitamin D can regulate the proliferation and growth of keratinocytes^{17, 42–44}, and it was also successfully incorporated as an adjuvant treatment for psoriasis^{26, 45}. The therapeutic action of the active forms of vitamin D is mediated by its receptor, VDR. Additionally, the *VDR* polymorphism is linked to psoriasis susceptibility^{27,28} and susceptibility of psoriatic patients to the vitamin D-based treatment. The reduced VDR levels in psoriatic skin are related to the reduced tight junctions⁴⁶. It has been suggested that such disturbances can

affect the maintenance of skin homeostasis⁴⁶. *VDR* polymorphisms can also be considered as genetic risk factors for sarcoidosis⁴⁷ with no effects of *VDR* SNPs on severity of sarcoidosis⁴⁸, oral lichen planus⁴⁹ and atopic dermatitis⁵⁰. The expression of *VDR* in inflammatory skin diseases is altered, but some study showed contradictory data. Kim et al⁵¹ observed the gradual decrease of *VDR* expression from normal skin to atopic dermatitis, from perilesional skin to psoriatic lesion. However, recent study showed that *VDR* is present in psoriatic skin with its predominantly strong expression⁵². The differences between these studies could result from the different antibodies used for the *VDR* detection. This consideration is supported by similar results of our study with the study by Milde et al⁵³, who used 9A7 clone of *VDR* antibody and showed comparable level of *VDR* in normal and psoriatic skin.

The expression of *ROR* γ in inflammatory skin diseases is usually reported in peripheral blood samples since *ROR* γ t is a crucial receptor for the differentiation of Th17 cells⁵⁴, that play essential role in the pathogenesis of psoriasis and other chronic inflammatory processes of the skin⁵⁵. Ecoeur et al⁵⁶ showed that selective *ROR* γ t inhibitor, Cpd A, inhibited Th17 pathway and the production of pro-inflammatory cytokines by T-cells and reduced IL-17-induced responses in keratinocytes. It has been proposed that *ROR* γ t can be molecular target for the psoriasis treatment⁵⁷. In mouse models of atopic dermatitis and acute irritant dermatitis, synthetic *ROR* α / γ inverse agonist - SR1001, exerted anti-inflammatory effects, restored epidermal barrier affecting multiple cell types in the skin⁵⁸. Our previous study showed that *ROR* γ and *ROR* α are expressed in human skin and can serve as receptors for vitamin D derivatives³⁴. In this study we found that *ROR* α and *ROR* γ are present in inflammatory skin diseases, however their expression was disturbed, suggesting that impaired *ROR* α and *ROR* γ expression can be involved in pathogenesis of inflammation.

LRP2/megalin may be crucial for proper cell development and differentiation⁵⁹. Its expression was detected in normal skin, hair follicles⁶⁰ and melanoma cells⁶¹. Our data showed the reduced *LRP2/megalin* levels in lesional cells in comparison to normal skin. Therefore, we suggest that the reduced expression of *LRP2/megalin* can be involved in pathogenesis of psoriasis and other inflammatory skin disorders through reduction of endocytosis and intracellular vitamin D3 signaling in affected skin cells.

In summary, the expression of *VDR*, *ROR* α and *ROR* γ and *LRP2/megalin* is aberrant in inflammatory skin diseases. We suggest that the disturbances in their expression are related to the pathogenesis of ACD, LSC, sarcoidosis and psoriasis. In addition, the presence of these receptors in the skin should allow to target them for vitamin-D based therapies of the inflammatory skin diseases providing a background for future clinical trials.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgement

Writing of this letter was in part supported by the NIH grants 1R01AR073004, R01AR071189, R21AI149267-01A1 and VA merit 1I01BX004293-01A1 to ATS and the Cancer Center Core grant (P30CA13148).

References

1. Slominski AT, Zmijewski MA, Skobowiat C, Zbytek B, Slominski RM, Steketee JD. Sensing the environment: regulation of local and global homeostasis by the skin's neuroendocrine system. *Adv Anat Embryol Cell Biol* 2012;212:v, vii, 1–115. doi:10.1007/978-3-642-19683-6_1
2. Shimada S, Katz SI. The skin as an immunologic organ. *Arch Pathol Lab Med* 1988;112(3):231–4. [PubMed: 2449876]
3. Richmond JM, Harris JE. Immunology and skin in health and disease. *Cold Spring Harb Perspect Med* 2014;4(12):a015339. doi:10.1101/cshperspect.a015339 [PubMed: 25452424]
4. Kupper TS, Fuhlbrigge RC. Immune surveillance in the skin: mechanisms and clinical consequences. *Nat Rev Immunol* 2004;4(3):211–22. doi:10.1038/nri1310 [PubMed: 15039758]
5. Guttman-Yassky E, Nograles KE, Krueger JG. Contrasting pathogenesis of atopic dermatitis and psoriasis--part I: clinical and pathologic concepts. *J Allergy Clin Immunol* 2011;127(5):1110–8. doi:10.1016/j.jaci.2011.01.053 [PubMed: 21388665]
6. Nestle FO, Kaplan DH, Barker J. Psoriasis. *N Engl J Med* 2009;361(5):496–509. doi:10.1056/NEJMra0804595 [PubMed: 19641206]
7. Rendon A, Schakel K. Psoriasis Pathogenesis and Treatment. *Int J Mol Sci* 2019;20(6)doi:10.3390/ijms20061475
8. Wong T, Hsu L, Liao W. Phototherapy in psoriasis: a review of mechanisms of action. *J Cutan Med Surg* 2013;17(1):6–12. doi:10.2310/7750.2012.11124 [PubMed: 23364144]
9. Tuckey RC, Cheng CYS, Slominski AT. The serum vitamin D metabolome: What we know and what is still to discover. *J Steroid Biochem Mol Biol* 2019;186:4–21. doi:10.1016/j.jsbmb.2018.09.003 [PubMed: 30205156]
10. Slominski AT, Chairprasongsuk A, Janjetovic Z, et al. Photoprotective Properties of Vitamin D and Lumisterol Hydroxyderivatives. *Cell Biochem Biophys* 2020;78(2):165–180. doi:10.1007/s12013-020-00913-6 [PubMed: 32441029]
11. Bikle DD. Vitamin D metabolism, mechanism of action, and clinical applications. *Chem Biol* 2014;21(3):319–29. doi:10.1016/j.chembiol.2013.12.016 [PubMed: 24529992]
12. Slominski RM, Raman C, Elmets C, Jetten AM, Slominski A, Tuckey RC. The significance of CYP11A1 expression in skin physiology and pathology. *Mol Cell Endocrinol* 2021; Available online 12 March 2021, 111238:111238. doi:10.1016/j.mce.2021.111238
13. Slominski RM, Tuckey RC, Manna PR, et al. Extra-adrenal glucocorticoid biosynthesis: implications for autoimmune and inflammatory disorders. *Genes Immun* 2020;21(3):150–168. doi:10.1038/s41435-020-0096-6 [PubMed: 32203088]
14. Cashman KD, Dowling KG, Skrabakova Z, et al. Vitamin D deficiency in Europe: pandemic? *Am J Clin Nutr* 2016;103(4):1033–44. doi:10.3945/ajcn.115.120873 [PubMed: 26864360]
15. Cantorna MT. Mechanisms underlying the effect of vitamin D on the immune system. *Proc Nutr Soc* 2010;69(3):286–9. doi:10.1017/S0029665110001722 [PubMed: 20515520]
16. Chambers ES, Hawrylowicz CM. The impact of vitamin D on regulatory T cells. *Curr Allergy Asthma Rep* 2011;11(1):29–36. doi:10.1007/s11882-010-0161-8 [PubMed: 21104171]
17. Gniadecki R, Gajkowska B, Hansen M. 1,25-dihydroxyvitamin D3 stimulates the assembly of adherens junctions in keratinocytes: involvement of protein kinase C. *Endocrinology* 1997;138(6):2241–8. doi:10.1210/endo.138.6.5156 [PubMed: 9165007]
18. Konijeti GG, Arora P, Boylan MR, et al. Vitamin D Supplementation Modulates T Cell-Mediated Immunity in Humans: Results from a Randomized Control Trial. *J Clin Endocrinol Metab* 2016;101(2):533–8. doi:10.1210/jc.2015-3599 [PubMed: 26653112]
19. Kuang Y, Xiao Y, Fang Z, et al. Association of Serum Vitamin D With Psoriasis and Effect Modification by Central Obesity. *Front Med (Lausanne)* 2020;7:236. doi:10.3389/fmed.2020.00236 [PubMed: 32626717]
20. Marahatta S, Agrawal S, Khan S. Study on Serum Vitamin D in Alopecia Areata Patients. *J Nepal Health Res Counc* 2019;17(1):21–25. doi:10.33314/jnhrc.1475 [PubMed: 31110371]

21. Upala S, Sanguankeo A. Low 25-hydroxyvitamin D levels are associated with vitiligo: a systematic review and meta-analysis. *Photodermatol Photoimmunol Photomed* 2016;32(4):181–90. doi:10.1111/phpp.12241 [PubMed: 27005676]
22. Ingram MA, Jones MB, Stonehouse W, et al. Oral vitamin D3 supplementation for chronic plaque psoriasis: a randomized, double-blind, placebo-controlled trial. *J Dermatolog Treat* 2018;29(7):648–657. doi:10.1080/09546634.2018.1444728 [PubMed: 29480035]
23. Adiguna MS, Rusyati LMM, Sudarsa PSS. Correlation of plasma vitamin d receptors with the severity of psoriasis vulgaris. *Bali Medical Journal* 2020;9(3):668–671. doi:DOI: 10.15562/bmj.v9i3.2013
24. Filoni A, Vestita M, Congedo M, Giudice G, Tafuri S, Bonamonte D. Association between psoriasis and vitamin D: Duration of disease correlates with decreased vitamin D serum levels: An observational case-control study. *Medicine (Baltimore)* 2018;97(25):e11185. doi:10.1097/MD.00000000000011185
25. Raj KAP, Handa S, Narang T, Sachdeva N, Mahajan R. Correlation of serum vitamin D levels with severity of pediatric atopic dermatitis and the impact of vitamin D supplementation on treatment outcomes. *J Dermatolog Treat* 2020:1–4. doi:10.1080/09546634.2020.1818677
26. Barrea L, Savanelli MC, Di Somma C, et al. Vitamin D and its role in psoriasis: An overview of the dermatologist and nutritionist. *Rev Endocr Metab Disord* 2017;18(2):195–205. doi:10.1007/s11154-017-9411-6 [PubMed: 28176237]
27. Lee YH. Vitamin D receptor ApaI, TaqI, BsmI, and FokI polymorphisms and psoriasis susceptibility: an updated meta-analysis. *Clin Exp Dermatol* 2019;44(5):498–505. doi:10.1111/ced.13823 [PubMed: 30474246]
28. Liu J, Wang W, Liu K, et al. Vitamin D receptor gene polymorphisms are associated with psoriasis susceptibility and the clinical response to calcipotriol in psoriatic patients. *Exp Dermatol* 2020;29(12):1186–1190. doi:10.1111/exd.14202 [PubMed: 32997398]
29. Haddad JG, Matsuoka LY, Hollis BW, Hu YZ, Wortsman J. Human plasma transport of vitamin D after its endogenous synthesis. *J Clin Invest* 1993;91(6):2552–5. doi:10.1172/JCI116492 [PubMed: 8390483]
30. Saito A, Pietromonaco S, Loo AK, Farquhar MG. Complete cloning and sequencing of rat gp330/”megalin,” a distinctive member of the low density lipoprotein receptor gene family. *Proc Natl Acad Sci U S A* 1994;91(21):9725–9. doi:10.1073/pnas.91.21.9725 [PubMed: 7937880]
31. Christensen EI, Birn H, Storm T, Weyer K, Nielsen R. Endocytic receptors in the renal proximal tubule. *Physiology (Bethesda)* 2012;27(4):223–36. doi:10.1152/physiol.00022.2012 [PubMed: 22875453]
32. Nykjaer A, Dragun D, Walther D, et al. An endocytic pathway essential for renal uptake and activation of the steroid 25-(OH) vitamin D3. *Cell* 1999;96(4):507–15. doi:10.1016/s0092-8674(00)80655-8 [PubMed: 10052453]
33. Zmijewski MA, Carlberg C. Vitamin D receptor(s): In the nucleus but also at membranes? *Exp Dermatol* 2020;29(9):876–884. doi:10.1111/exd.14147 [PubMed: 32654294]
34. Slominski AT, Kim TK, Takeda Y, et al. RORalpha and ROR gamma are expressed in human skin and serve as receptors for endogenously produced noncalcemic 20-hydroxy- and 20,23-dihydroxyvitamin D. *FASEB J* 2014;28(7):2775–89. doi:10.1096/fj.13-242040 [PubMed: 24668754]
35. Brozyna AA, Jozwicki W, Skobowiat C, Jetten A, Slominski AT. RORalpha and RORgamma expression inversely correlates with human melanoma progression. *Oncotarget* 2016;7(39):63261–63282. doi:10.18632/oncotarget.11211 [PubMed: 27542227]
36. Jetten AM. Retinoid-related orphan receptors (RORs): critical roles in development, immunity, circadian rhythm, and cellular metabolism. *Nucl Recept Signal* 2009;7:e003. doi:10.1621/nrs.07003 [PubMed: 19381306]
37. Jetten AM, Takeda Y, Slominski A, Kang HS. Retinoic acid-related Orphan Receptor gamma (RORgamma): connecting sterol metabolism to regulation of the immune system and autoimmune disease. *Curr Opin Toxicol* 2018;8:66–80. doi:10.1016/j.cotox.2018.01.005 [PubMed: 29568812]

38. Brozyna AA, Jozwicki W, Slominski AT. Decreased VDR expression in cutaneous melanomas as marker of tumor progression: new data and analyses. *Anticancer Res* 2014;34(6):2735–43. [PubMed: 24922634]
39. Jozwicki W, Brozyna AA, Siekiera J, Slominski AT. Expression of Vitamin D Receptor (VDR) Positively Correlates with Survival of Urothelial Bladder Cancer Patients. *Int J Mol Sci* 2015;16(10):24369–86. doi:10.3390/ijms161024369 [PubMed: 26501255]
40. Li B, Tsoi LC, Swindell WR, et al. Transcriptome analysis of psoriasis in a large case-control sample: RNA-seq provides insights into disease mechanisms. *J Invest Dermatol* 2014;134(7):1828–1838. doi:10.1038/jid.2014.28 [PubMed: 24441097]
41. Grassi T, Panico A, Bagordo F, et al. Direct detection of free vitamin D as a tool to assess risk conditions associated with chronic plaque psoriasis. *J Prev Med Hyg* 2020;61(3):E489–E495. doi:10.15167/2421-4248/jpmh2020.61.3.1482 [PubMed: 33150238]
42. Tang JY, Fu T, Lau C, Oh DH, Bikle DD, Asgari MM. Vitamin D in cutaneous carcinogenesis: part II. *J Am Acad Dermatol* 2012;67(5):817 e1–11; quiz 827–8. doi:10.1016/j.jaad.2012.07.022 [PubMed: 23062904]
43. Tang JY, Fu T, Lau C, Oh DH, Bikle DD, Asgari MM. Vitamin D in cutaneous carcinogenesis: part I. *J Am Acad Dermatol* 2012;67(5):803 e1–12, quiz 815–6. doi:10.1016/j.jaad.2012.05.044 [PubMed: 23062903]
44. Bikle DD. Vitamin D and the skin: Physiology and pathophysiology. *Rev Endocr Metab Disord* 2012;13(1):3–19. doi:10.1007/s11154-011-9194-0 [PubMed: 21845365]
45. Disphanurat W, Viarasilpa W, Chakkavittumrong P, Pongcharoen P. The Clinical Effect of Oral Vitamin D2 Supplementation on Psoriasis: A Double-Blind, Randomized, Placebo-Controlled Study. *Dermatol Res Pract* 2019;2019:5237642. doi:10.1155/2019/5237642
46. Visconti B, Paolino G, Carotti S, et al. Immunohistochemical expression of VDR is associated with reduced integrity of tight junction complex in psoriatic skin. *J Eur Acad Dermatol Venereol* 2015;29(10):2038–42. doi:10.1111/jdv.12736 [PubMed: 25220655]
47. Niimi T, Tomita H, Sato S, et al. Vitamin D receptor gene polymorphism in patients with sarcoidosis. *Am J Respir Crit Care Med* 1999;160(4):1107–9. doi:10.1164/ajrccm.160.4.9811096 [PubMed: 10508794]
48. Stjepanovic MI, Mihailovic-Vucinic V, Spasovski V, et al. Genes and metabolic pathway of sarcoidosis: identification of key players and risk modifiers. *Arch Med Sci* 2019;15(5):1138–1146. doi:10.5114/aoms.2018.79682 [PubMed: 31572458]
49. Shen H, Liu Q, Huang P, et al. Vitamin D receptor genetic polymorphisms are associated with oral lichen planus susceptibility in a Chinese Han population. *BMC Oral Health* 2020;20(1):26. doi:10.1186/s12903-020-1002-3 [PubMed: 32000758]
50. Zhang L, Zhang S, He C, Wang X. VDR Gene Polymorphisms and Allergic Diseases: Evidence from a Meta-analysis. *Immunol Invest* 2020;49(1–2):166–177. doi:10.1080/08820139.2019.1674325 [PubMed: 31752548]
51. Kim SK, Park S, Lee ES. Toll-like receptors and antimicrobial peptides expressions of psoriasis: correlation with serum vitamin D level. *J Korean Med Sci* 2010;25(10):1506–12. doi:10.3346/jkms.2010.25.10.1506 [PubMed: 20890434]
52. Chandra R, Roesyanto-Mahadi ID, Yosi A. Pilot study: immunohistochemistry expressions of vitamin D receptor associated with severity of disease in psoriasis patients. *Int J Dermatol* 2020;59(9):1092–1097. doi:10.1111/ijd.15018 [PubMed: 32592616]
53. Milde P, Hauser U, Simon T, et al. Expression of 1,25-dihydroxyvitamin D3 receptors in normal and psoriatic skin. *J Invest Dermatol* 1991;97(2):230–9. doi:10.1111/1523-1747.ep12480255 [PubMed: 1649228]
54. Annunziato F, Cosmi L, Santarlasci V, et al. Phenotypic and functional features of human Th17 cells. *J Exp Med* 2007;204(8):1849–61. doi:10.1084/jem.20070663 [PubMed: 17635957]
55. Bergallo M, Accorinti M, Galliano I, et al. Expression of miRNA 155, FOXP3 and ROR gamma, in children with moderate and severe atopic dermatitis. *G Ital Dermatol Venereol* 2020;155(2):168–172. doi:10.23736/S0392-0488.17.05707-8 [PubMed: 29249119]
56. Ecoeur F, Weiss J, Kaupmann K, Hintermann S, Orain D, Guntermann C. Antagonizing Retinoic Acid-Related-Orphan Receptor Gamma Activity Blocks the T Helper 17/Interleukin-17

- Pathway Leading to Attenuated Pro-inflammatory Human Keratinocyte and Skin Responses. *Front Immunol* 2019;10:577. doi:10.3389/fimmu.2019.00577 [PubMed: 30972071]
57. Tang L, Yang X, Liang Y, Xie H, Dai Z, Zheng G. Transcription Factor Retinoid-Related Orphan Receptor γ : A Promising Target for the Treatment of Psoriasis. *Front Immunol* 2018;9:1210. doi:10.3389/fimmu.2018.01210 [PubMed: 29899748]
58. Dai J, Choo MK, Park JM, Fisher DE. Topical ROR Inverse Agonists Suppress Inflammation in Mouse Models of Atopic Dermatitis and Acute Irritant Dermatitis. *J Invest Dermatol* 2017;137(12):2523–2531. doi:10.1016/j.jid.2017.07.819 [PubMed: 28774591]
59. Akour AA, Kennedy MJ, Gerk P. Receptor-mediated endocytosis across human placenta: emphasis on megalin. *Mol Pharm* 2013;10(4):1269–78. doi:10.1021/mp300609c [PubMed: 23438198]
60. Adly MA. Expression of the carrier protein transthyretin and its receptor megalin in human skin: preliminary findings. *Br J Dermatol* 2010;162(1):213–5. doi:10.1111/j.1365-2133.2009.09519.x [PubMed: 19886883]
61. Andersen RK, Hammer K, Hager H, et al. Melanoma tumors frequently acquire LRP2/megalín expression, which modulates melanoma cell proliferation and survival rates. *Pigment Cell Melanoma Res* 2015;28(3):267–80. doi:10.1111/pcmr.12352 [PubMed: 25585665]

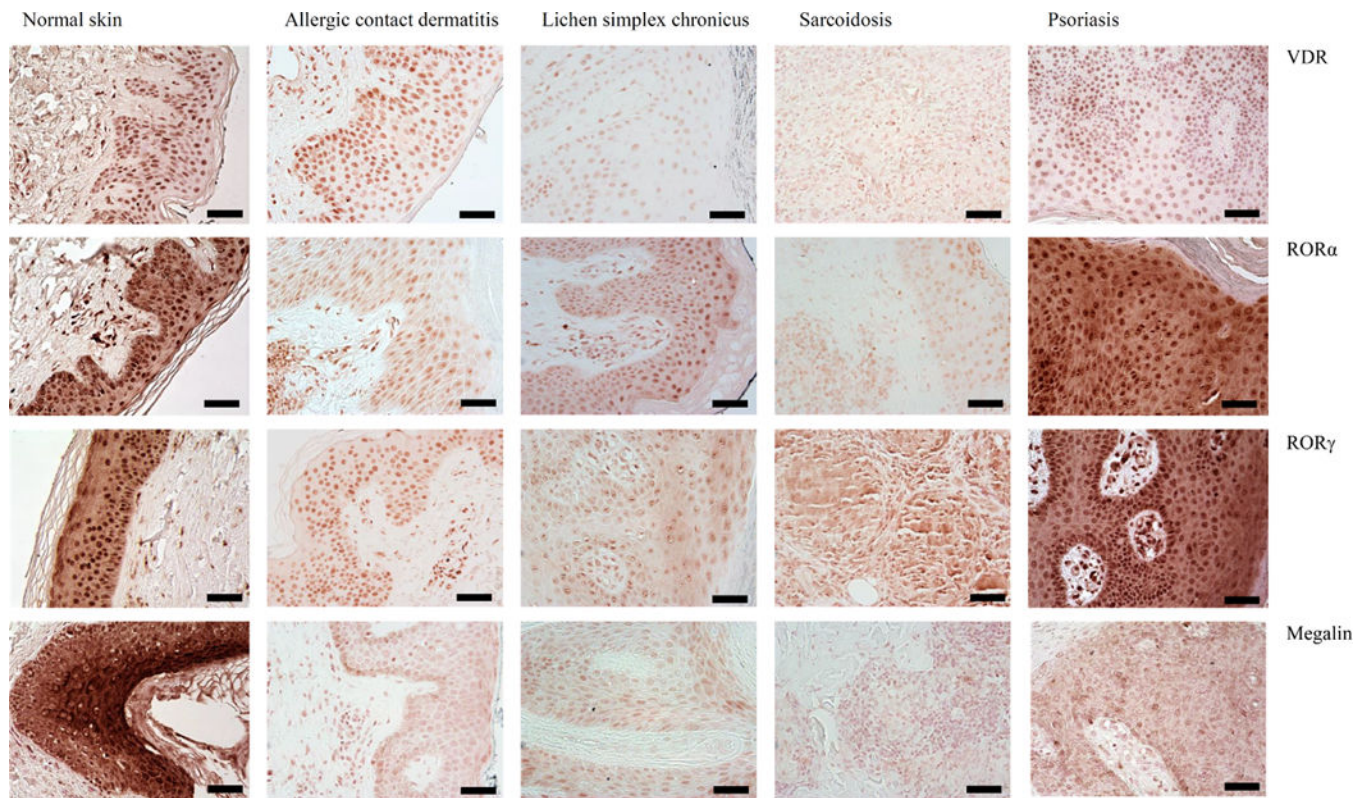


Figure 1. Representative images of immunostaining of VDR, ROR α and ROR γ and LRP2/megalin in allergic contact dermatitis, lichen simplex chronicus, sarcoidosis and psoriasis. Scale bars: = 50 μ m.

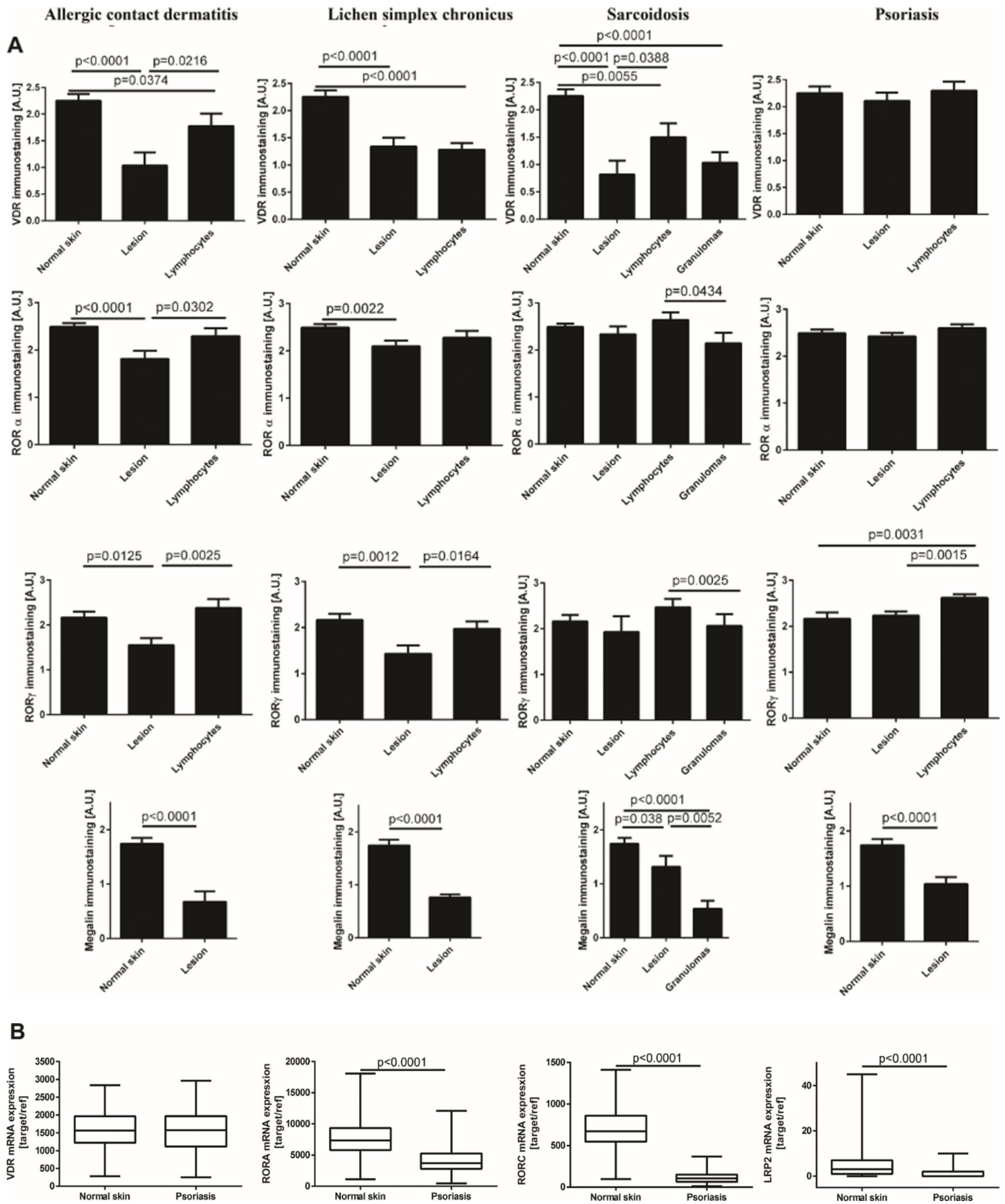


Figure 2.

Expression levels of VDR, ROR α and ROR γ and LRP2/megalin. A) The expression measured by immunohistochemistry in psoriasis, allergic contact dermatitis, lichen simplex chronicus and sarcoidosis. Statistically significant differences are denoted with p values as determined by Student's *t*-test. B). The expression measured by RNA-seq in psoriasis and normal skin (<https://www.ebi.ac.uk/gxa/home>; ⁴⁰).

Table 1.

Characteristic of normal skin (control) and lesional skin samples.

	Normal skin (control)	Psoriasis	Atopic contact dermatitis	Lichen simplex chronicus	Sarcoidosis
Number of cases	36	26	10	18	8
Sex (F/M)	17/9*	12/14	2/8	3/13**	5/1****
Age (mean [range])	44.3 (20–68)*	52.7 (20–86)	50.6 (20–62)	51.1 (34–70)	#
Location	*			**	****
<i>Head (forehead, jaw, chin, temple, scalp)</i>	2	1		6	
<i>Neck</i>	1			4	
<i>Leg</i>	2	4	3	1	1
<i>Arm</i>		7			4
<i>Hand</i>	1		1		
<i>Breast</i>	6			2	
<i>Back</i>	1	6	1	1	1
<i>Abdomen</i>	11	6			
<i>Foreskin</i>	2				
<i>Other</i>		2	5	2	
Immunostaining					
<i>VDR</i>	34	26	10	17	7
<i>LRP2/megalin</i>	26	26	10	18	8
<i>RORα</i>	33	26	10	18	7
<i>RORγ</i>	31	26	10	18	7

* data available for 26 cases

** data available for 16 cases

*** data available for 6 cases

lack of the data

Number of samples stained for VDR, megalin and RORs differs due to lack of representative of lesional areas in the sections.