## Antimicrobial Susceptibilities of 1,730 *Haemophilus influenzae* Respiratory Tract Isolates in Spain in 1998-1999

FRANCESC MARCO,<sup>1</sup> JUAN GARCÍA-DE-LOMAS,<sup>2</sup> CÉSAR GARCÍA-REY,<sup>3</sup> EMILIO BOUZA,<sup>4</sup> LORENZO AGUILAR,<sup>3\*</sup> CARLOS FERNÁNDEZ-MAZARRASA,<sup>5</sup> AND THE SPANISH SURVEILLANCE GROUP FOR RESPIRATORY PATHOGENS<sup>†</sup>

Microbiology Department, IDIBAPS, Hospital Cliníc, Barcelona<sup>1</sup>; Instituto Valenciano de Microbiología, Valencia<sup>2</sup>; Medical Department, GlaxoSmithKline, Tres Cantos, Madrid<sup>3</sup>; Microbiology Department, Hospital General Universitario Gregorio Marañón, Madrid<sup>4</sup>; and Microbiology Department, Residencia Marqués de Valdecilla, Santander,<sup>5</sup> Spain

Received 28 December 2000/Returned for modification 25 June 2001/Accepted 8 August 2001

A  $\beta$ -lactamase prevalence of 23% was found among 1,730 *Haemophilus influenzae* isolates. Ampicillin susceptibility was 70%, and 12% of  $\beta$ -lactamase-negative strains presented diminished susceptibility to ampicillin (BLNAR phenotype). Susceptibility of 90% was found for cefaclor and clarithromycin, whereas it was nearly 100% for cefotaxime, cefixime, azithromycin, and cefuroxime. Ciprofloxacin-resistant (0.1%) and  $\beta$ -lactamase-positive amoxicillin/clavulanate-resistant (BLPACR) phenotypes (0.1%) are anecdotal so far.

The aim of the present study is to describe the current susceptibility patterns of *Haemophilus influenzae* to first-line antibiotics used for respiratory tract infections, as well as to describe the prevalence of the new phenotypes of antibiotic resistance in Spain.

A total of 1,730 consecutive clinical isolates of *H. influenzae* obtained from community-acquired respiratory tract infections, collected between November 1998 and October 1999, were included in this prospective surveillance study involving 17 hospitals selected on the basis of geographic location. Strains were sent to a central laboratory (Instituto Valenciano de Microbiología, Valencia, Spain) to confirm the identification by colony morphology, Gram staining, growth in chocolate but not in blood agar, catalase test, and X and V factor requirements. In addition, a  $\beta$ -lactamase test (nitrocefin; Becton Dickinson) was performed (9). Susceptibility testing was per-

formed following the National Committee for Clinical Laboratory Standards (NCCLS) 1999 guidelines using a semiautomated microdilution (7) method (Sensititre; Trek Diagnostics Inc., Westlake, Ohio) against antimicrobials commonly used as empiric therapy of respiratory tract infections in Spain. These were penicillin, amoxicillin, ampicillin, amoxicillin-clavulanate, cefaclor, cefuroxime, cefixime, cefotaxime, erythromycin, clarithromycin, azithromycin, and ciprofloxacin.

H. influenzae ATCC 49247, H. influenzae ATCC 49766, and Escherichia coli ATCC 35218 were used as quality control strains following NCCLS recommendations (7). BLNAR strains were defined as those strains failing to detect β-lactamase production and presenting with either a high ( $\geq 4 \,\mu g/ml$ ) or intermediate (2 µg/ml) ampicillin MIC (3). On the other hand, B-lactamase-positive amoxicillin-clavulanate-resistant (BLPACR) strains were defined as those with an MIC to amoxicillin-clavulanate of  $\geq 8 \ \mu g/ml$  that rendered positive with the nitrocefin disk test (3). Differences between  $\beta$ -lactamase-positive and  $\beta$ -lactamase-negative strains with respect to the prevalence of susceptibility were calculated using comparison of proportions with the chi square test. Estimation of 95% confidence intervals (95%CI) for population proportions was made by the exact binomial method. Epi-Info version 6.04 was used for statistical calculations.

A total of 1,534 samples (88.6%) were collected from the lower respiratory tract (sputum and bronchoalveolar lavage), 26 (1.5%) were from hemoculture, and 170 (9.8%) were from middle ear exudate. The vast majority of isolates (97.3%) were not typeable, whereas serogroups b, d, e, and f represented just 1.2, 0.2, 0.2, and 1%, respectively. The different production of  $\beta$ -lactamase in hemoculture (30.7%) in comparison with middle ear samples (23.5%) and lower respiratory tract samples (22.8%) did not show a significant statistical difference.

Prevalence of susceptibility, and MICs for 50 and 90% of strains ( $MIC_{50}$  and  $MIC_{90}$ ) are shown in Table 1. According to NCCLS breakpoints, all antibiotics tested exhibited susceptibility rates equal to or higher than 99% except for ampicillin (70%), clarithromycin (89%), and cefaclor (91%). The antibi-

<sup>\*</sup> Corresponding author. Mailing address: Medical Department, GlaxoSmithKline, C/Severo Ochoa, 2, 28760 Tres Cantos, Madrid, Spain. Phone: 34 91 807 5912. Fax: 34 91 807 0596. E-mail: lorenzo .aguilar-alfaro@gsk.com.

<sup>†</sup> Members of the Spanish Surveillance Group for Respiratory Pathogens are: E. Cercenado, Hospital Gregorio Marañón, Madrid; T. Jiménez de Anta, Hospital Clínic, Barcelona; A. García-Perea, S. García, and M. Güeni, Hospital La Paz, Madrid; E. Pérez-Trallero and J. Larruskain, Complejo Hospitalario Donostia, San Sebastián; J. Barrón and L. López, Hospital de Cruces, Baracaldo; C. Rubio and C. García, Hospital Clínico Universitario, Zaragoza; J. A. García-Rodríguez (Study Coordinator) and I. Trujillano, Hospital Clínico Universitario, Salamanca; J. Ruiz and E. Simarro, Hospital Virgen de la Arrixaca, Murcia; C. García-Riestra, B. Regueiro, A. Jato, and M. Prieto, Hospital Clínico Universitario, Santiago de Compostela; J. M. Nogueira, Hospital Dr. Peset, Valencia; C. Latorre and A. Gené, Hospital Sant Joan de Deu, Barcelona; M. de la Rosa, Hospital Virgen de las Nieves, Granada; E. Perea and L. Martínez, Hospital Virgen de la Macarena, Sevilla; A. M. Martín and F. Cañas, Hospital Insular, Las Palmas; M. Casal and A. Ibarra, Hospital Reina Sofía, Córdoba; D. Romero and M. González, Hospital Nuestra Señora de Alarcos, Ciudad Real; F. Baquero (Study Coordinator), Hospital Ramón y Cajal, Madrid; J. J. Granizo, Fundación Jiménez Díaz, Madrid; L. López and C. Gimeno, Instituto Valenciano de Microbiología, Valencia; and R. Dal-Ré, GlaxoSmithKline, Tres Cantos, Madrid, Spain.

Overall $(n = 1,730)$ $\beta$ -Lactamase positive $(n = 399)$ $\beta$ -Lactamase negative $(n = 399)$	Lactamase negativ	(n = 1,331)
Antimicrobial MIC (μg/ml) No. (%) of strains MICs (μg/ml) No. (%) of strains MICs (μg/ml) No. (%) of strains	No.	(%) of strains
$\frac{1}{MIC_{50}} \frac{1}{MIC_{90}} \frac{1}{MIC_{50}} \frac{1}{MIC_{50}} \frac{1}{MIC_{50}} \frac{1}{MIC_{90}} \frac{1}{MIC_{50}} \frac{1}$	0 Susceptible	ntermediate Resistan
Penicillin 1 ≥16 ≤0.015–≥16 NA NA NA ≥16 ≥16 NA NA NA 0.5 4 NA	NA	NA NA
Amoxicillin $1 \ge 16 \le 0.015 \ge 16$ NA NA NA $\ge 16 \ge 16$ NA NA NA $1$ 4 NA	NA	NA NA
$ \text{Ampicillin} \qquad \leq 0.5 \geq 8 \qquad \leq 0.5 - \geq 8 \qquad 1,209 \ (69.9) \qquad 96 \ (5.5) \qquad 425 \ (24.6) \geq 8 \qquad \geq 8 \qquad 39 \ (9.7) \qquad 9 \ (2.3) \qquad 351 \ (88) \qquad \leq 0.5 \qquad 2 \qquad 1,170 \ (87.9) \qquad 8 \qquad = 1,170 \ (87.9) \qquad 1,170 \ (87.9) \ $	1,170(87.9)	$87 (6.5)^b$ 74 $(5.6)^b$
$Amox-clav^{c} 1 2 \leq 0.015-8 1.722 (99.5) NA 8 (0.5) 1 4 397 (99.5) NA 2 (0.5) 0.5 2 1.325 (99.5)$	1,325 (99.5)	NA 6 (0.5)
Cefaclor 4 8 $\leq 1-\geq 64$ 1,581 (91.4) 99 (5.7) 50 (2.9) 8 $\geq 64$ 281 (70.4) 75 (18.8) 43 (10.8) 4 8 1,699 (97.7) 2 (18.8) (10.8) 4 (10.8)	1,699 (97.7)	24 (1.8) 7 (0.5)
Cefuroxime 1 4 $\leq 0.25-8$ 1,718 (99.3) 12 (0.7) 0 1 4 392 (98.2) 7 (1.8) 0 1 4 1,725 (99.6)	1,725 (99.6)	5(0.4) 0
Cefixime ≤0.25 ≤0.25 ≤0.25 ≥8 1,727 (99.8) NA NA ≤0.25 ≤0.25 397 (99.5) NA NA ≤0.25 ≤0.25 1,330 (99.9)	5 1,330 (99.9)	NA NA
Cefotaxime ≤0.25 ≤0.25 ≤0.25 ≤ 1,729 (99.9) NA NA ≤0.25 ≤0.25 399 (100) NA NA ≤0.25 ≤0.25 1,330 (99.9)	5 1,330 (99.9)	NA NA
Erythromycin 4 8 $\leq 0.12 - \geq 64$ NA NA NA 4 8 NA NA NA 4 8 NA	NA	NA NA
$Clarithromycin 4 16 \leq 0.25 \geq 64 1.546 (89.4) 159 (9.2) 25 (1.4) 8 16 353 (88.5) 40 (10) 6 (1.5) 4 16 1.193 (89.7) 15 (10.5) 16 1.193 (10.5) $	1,193 (89.7)	119(8.9) 19(1.4)
Azithromycin $0.5$ 2 $\leq 0.12 - \geq 64$ 1,712 (98.9) NA NA $0.5$ 2 393 (98.5) NA NA $0.5$ 2 1,319 (99.1)	1,319(99.1)	NTA NTA
Ciprofloxacin $\leq 0.5 \leq 0.5 \leq 0.5.4 $ 1,728 (99.9) NA NA $\leq 0.5 \leq 0.5.399$ (100) NA NA $\leq 0.5 \leq 0.5.1,329$ (99.8)	1,329 (99.8)	NA NA

otics with lowest intrinsic potency (as measured by  $MIC_{90}$ ) happened to be penicillin, amoxicillin, ampicillin, cefaclor, erythromycin, and clarithromycin, all of them displaying an  $MIC_{90}$  of  $\geq 8 \ \mu g/ml$ . The remaining antibiotics for which there are NCCLS breakpoints presented a far better intrinsic potency, and as expected, third-generation cephalosporins and ciprofloxacin showed the lowest  $MIC_{90}$ .

Nearly one of every four isolates (399 of 1,730) was a  $\beta$ -lactamase producer (23%; 95%CI, 21 to 25%), a figure very much like that found in our previous surveillance study (25.7%) (4), but significantly lower than that reported in Spain by other national (10) and multinational (11) studies, around 30 to 35%. Considering geographical differences in our 17 centers,  $\beta$ -lactamase production ranged from 15.8 to 30.4%, but no statistical significance was reached. The  $\beta$ -lactamase production status of the isolates did not influence their susceptibility to the antibiotics tested except for ampicillin (9.7% for  $\beta$ -lactamase positive versus 87.9% for  $\beta$ -lactamase negative; P < 0.0001) and cefaclor (70.4% for  $\beta$ -lactamase positive versus 97.7% for  $\beta$ -lactamase negative; P < 0.0001). Cefaclor was the sole oral cephalosporin for which  $\beta$ -lactamase production influenced the prevalence of susceptibility.

Concerning  $\beta$ -lactamase-negative isolates, 87 (6.5%) and 74 (5.5%) strains were found to be intermediate and resistant to ampicillin, respectively, and were therefore categorized as BLNAR. This means that 9.3% of the whole *H. influenzae* population of the study were BLNAR (95% CI, 8 to 10.7%), suggesting an increase compared with previous reports in Spain of around 5% (3,10). Again, no significant differences in the prevalence of BLNAR strains were seen on a geographical basis.

Macrolide nonsusceptibility was around 1% for azithromycin and 10% for clarithromycin, most of this being nonsusceptibility categorized as intermediate resistance. In any case, clarithromycin was significantly less active than azithromycin in terms of  $MIC_{90}$  values (16 µg/ml versus 2 µg/ml).

Regarding the extremely rare ciprofloxacin resistance phenotype, we found only two isolates (0.1%) with MICs of 2 and 4 µg/ml, which is in accordance with other authors (1). Likewise, only two isolates (0.1%) were found to be BLPACR, which is less than reported elsewhere (3).

*H. influenzae* is one of the most prevalent isolates in community-acquired respiratory tract infections. Multicenter surveillances have had key importance in ascertaining  $\beta$ -lactamase production (2,4,5,8,10), decreasing susceptibility to certain macrolides (4), and increasing intrinsic resistance to ampicillin mediated by altered penicillin-binding proteins (PBP) (6), the so-called  $\beta$ -lactamase-negative ampicillin resistance (BLNAR). Likewise, surveys have revealed novel resistance phenotypes as very rare ciprofloxacin-resistant (1) and  $\beta$ -lactamase-positive and amoxicillin-clavulanate-resistant (BLPACR) isolates (3).

Our survey shows that  $\beta$ -lactamase production in Spain (23%) is homogeneously distributed and seems to be decreasing compared with other reports, but ampicillin susceptibility hardly changes (69.9%; 95% CI, 67.7 to 72%). However, the prevalence of the BLNAR phenotype seems to be increasing (9.3%), and sharp differences in the activity of macrolides are also evident. Both BLPACR and ciprofloxacin-resistant phenotypes constitute to date just anecdotal events (0.1%), but

they must be monitored. Regarding BLNAR strains and following NCCLS, the question is open as to what extent ampicillin-intermediate strains should be considered resistant to the remaining  $\beta$ -lactams antibiotics within the family.

Differences in susceptibility patterns between different surveillances over time due to the increasing prevalence of certain phenotypes of resistance or the emergence of new ones make national multicenter surveys a must. Most of the multicenter surveillances of resistance carried out so far have as a main criticism that they make their estimations based either on a relatively small number of isolates from a single country or on a large number of isolates from many different countries. Both attitudes cannot but lead to a somewhat inadequate representation of the complex reality of the resistance dynamics for a given country. That is why extensive national multicenter surveillances provide more reliable information, ensuring a better estimation for the country whose specific information on resistance is sought.

## REFERENCES

- Biedenbach D. J., and R. N. Jones. 2000. Fluoroquinolone-resistant Haemophilus influenzae: frequency of occurrence and analysis of confirmed strains in the SENTRY antimicrobial surveillance program (North and Latin America). Diagn. Microbiol. Infect. Dis. 36:255–259.
- Cullman, W. 1996. Comparative evaluation of orally active antibiotics against community-acquired pathogens: results of eight European countries. Chemotherapy 42:11–20.
- Doern, G. V., A. B. Brueggemann, G. Pierce, H. Preston Holley, Jr., and A. Rauch. 1997. Antibiotic resistance among clinical isolates of *Haemophilus* influenzae in the United States in 1994 and 1995 and detection of β-lacta-

mase positive strains resistant to amoxicillin-clavulanate: results of a national multicenter surveillance study. Antimicrob. Agents Chemother. **41**:292–297.

- García-Rodríguez, J. A., F. Baquero, J. García-de-Lomas, L. Aguilar and the Spanish Surveillance Group for Respiratory Pathogens. 1999. Antimicrobial susceptibility of 1,422 *Haemophilus influenzae* isolates from respiratory tract infections in Spain: results of a 1-year (1996–97) multicenter surveillance study. Infection 27:265–267.
- Machka, K., I. Braveny, H. Dabernat, K. Dornbush, E. Van Dyck, F. H. Kayser, F. Van Klingeren, H. Mittermayer, E. Perea, and M. Powell. 1988. Distribution and resistance patterns of *Haemophilus influenzae*: a European Cooperative Study. Eur. J. Clin. Microbiol. Infect. Dis. 7:14–24.
- Mendelman, P. M., D. O. Chaffin, T. L. Stull, C. E. Rubens, K. D. Mack, and A. L. Smith. 1984. Characterization of non-β-lactamase-mediated ampicillin resistance in *Haemophilus influenzae*. Antimicrob. Agents Chemother. 26: 235–244.
- National Committee for Clinical Laboratory Standards. 1999. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically— 4th ed.; approved standard. NCCLS document M7–A4. Performance Standards for Antimicrobial Susceptibility Testing; Ninth Informational Supplement. NCCLS document M100–S9. National Committee for Clinical Laboratory Standards, Wayne, Pa.
- Needham, C. S. 1988. *Haemophilus influenzae*: antibiotic susceptibility. Clin. Microbiol. Rev. 1:218–227.
- O'Callaghan, C. H., A. Morris, S. M. Kirby, and A. H. Shingler. 1972. Novel method for detection of β-lactamase by using a chromogenic cephalosporin substrate. Antimicrob. Agents Chemother. 1:283–288.
- Perea, E. J., M. C. García, M. J. Clavijo, G. Piédrola, J. Campos, J. A. García-Rodríguez, E. García-Sánchez, R. Cisterna, M. Alvarez, F. Martín, A. Rodríguez, M. Tejero, B. Regueiro, M. Pérez, R. Martín, R. Verdaguer, J. García-de-Lomas, C. Gimeno, E. Bouza, and M. Rodríguez. 1993. Resistance among *Haemophilus influenzae* in Spain: a second study (1990). Enferm. Infec. Microbiol. Clin. 11:31–40. (In Spanish.)
- Sahm, D. F., M. E. Jones, M. L. Hickey, D. R. Diakun, S. V. Mani, and C. Thornsberry. 2000. Resistance surveillance of *Streptococcus pneumoniae*, *Haemophilus influenzae* and *Moraxella catarrhalis* isolated in Asia and Europe, 1997–1998. J. Antimicrob. Chemother. 45:457–466.