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Review

Exosomes in cancer immunoediting and immunotherapy



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ABSTRACT

As an important means of communication among cells, exosomes are being studied more and more widely, especially in the context of cancer immunotherapy. In the phase of tumor immunoediting, exosomes derived from tumor cells and different immune cells have complex and changeable physiological functions, because they carry different proteins and nucleic acid from the source cells. Based on the role of exosomes in the communication between different cells, cancer treatment methods are also under continuous research. This review briefly introduces the molecular composition of exosomes, which is closely related to their secretion mechanism. Subsequently, the role of exosomes encapsulating different information molecules is summarized. The role of exosomes in the three phases of tumor immunoediting is introduced in detail, and the relevant literature of exosomes in the tumor immune microenvironment is summarized by using a novel framework for extracting relevant documents. Finally, it summarizes the various exosome-based immunotherapies currently proposed, as well as the challenges and future prospects of exosomes in tumor immunotherapy.

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1. Introduction

Exosomes, a kind of nano-scale exovesicles secreted by cells, have now been widely studied as one of the communication methods among cells [1–3]. They can be derived from almost all cells and can be detected in biological fluids. The exosomes released to the outside of the cell wrap the information, such as protein and nucleic acid of the source cell, transmitted to neighboring or remote cells [4,5]. Therefore, exosomes are widely involved in physiological and pathological processes, especially in tumorigenesis and treatment [6].

American tumor biologist R.D Schreiber first proposed a hypothesis called "cancer immunoediting" in 2002 [7], which believed that the immune system not only has the ability to exclude tumor cells but also promotes tumor growth. Specifically, the three phases of cancer immunoediting, elimination, equilibrium, and escape [8], are the process of development after the tumor and the immune system fight against each other. In the past two decades, this hypothesis has been gradually refined and widely confirmed [9–12]. Under this concept, there are relatively few discussions about the complex communication of exosomes between tumor

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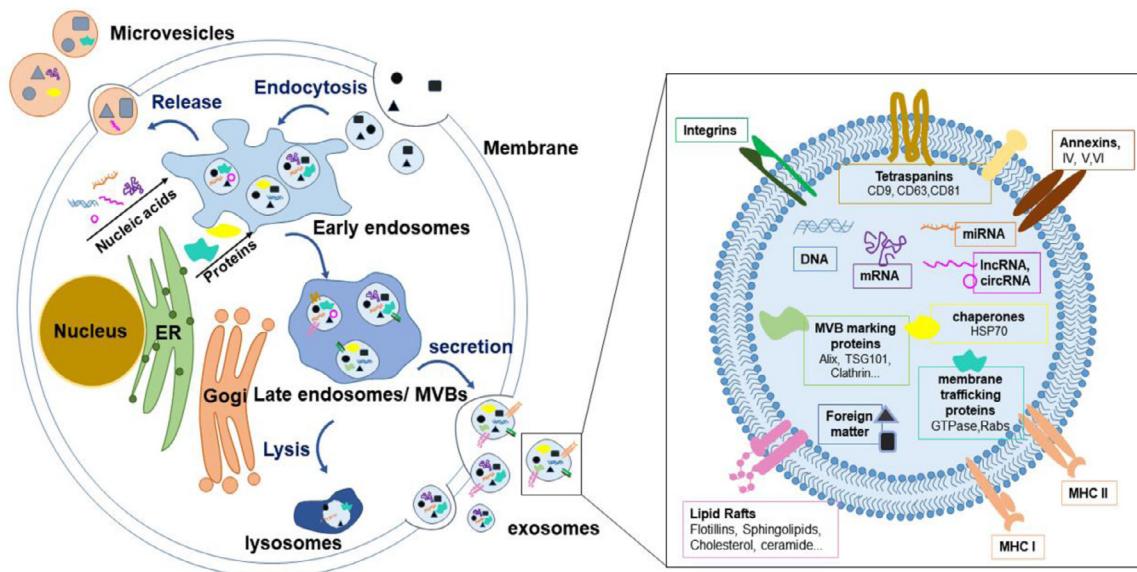


Fig. 1 – The secretion mechanism and composition of exosomes. Exosomes originate from endosomes formed by cell membrane invagination. Early endosomes gradually develop into late endosomes containing multiple vesicles after material sorting and transportation. The small vesicles finally released to the outside through membrane fusion are exosomes. Small extracellular vesicles formed by plasma membrane budding may also be considered exosomes. Exosomes have a phospholipid bilayer and rich in tetraspanins, annexins, integrins, chaperones, lipids and nucleic acids.

cells and immune cells. Therefore, this review will use it as a key point to introduce the role of exosomes in cancer immunoediting and applications related to immunotherapy.

2. Exosomal composition

Exosomes, single-layer membrane vesicles with a size of 30–150 nm, are one of the exovesicles secreted by cells, which are heterogeneous and have no independent replication ability [13,14]. Since there is no consensus on the specific markers of several vesicles secreted by cells, according to MISEV2018, the vesicles below 200 nm are classified as small external vesicles, and those above 200 nm are medium or large external vesicles [15]. The exosomes in this review mainly discuss small extracellular vesicles including exosomes. Almost all types of cells secrete exosomes, such as epithelial cells [16], stem cells [17], immune cells [18] and various tumor cells [19]. The molecular composition of exosomes is closely related to their secretion mechanism, as shown in Fig. 1. Exosomes originate from plasma membrane or endosomal membrane budding, although many reports believe that exosomes originate from endosomal membrane budding, ignoring plasma membrane budding and considering them as microvesicles [13,20]. For exosomes originating from late endosomes, they first undergo plasma membrane invagination to form early endosomes [21]. As the endosomal membranes gradually form multivesicular bodies, the exosomes are finally released to the outside of the cell after fusion with the plasma membrane. This formation process is proved by proteomics research [22]. The rich lipid components in exosomes also imply the involvement of endosomal and plasma membranes in the whole process [23].

The cargo sorting mechanism involved in this is complicated. The endosome sorting complex required for transport (ESCRT), consisting of ESCRT-0, ESCRT-I, ESCRT-II, ESCRT-III and vacuolar protein sorting 4(VPS4)-vesicle transport 1 and some auxiliary proteins [24]. For example, it is composed of ALG-2 interacting protein X (Alix), which participates in cargo sorting and the process of endosomal membrane folding and multivesicular body (MVB) formation [25]. However, some studies have shown that eliminating the function of VPS4 has no effect on the exosome secretion of CD63 and other exosomal markers [26], which implies the existence of an exosome secretion mechanism that does not rely on the ESCRT pathway. For example, the work of Xu et al. proved that exosomes secreted by macrophages transfer phagocytic antigens to dendritic cells (DCs) in a ceramide-dependent manner [27]. Many other researchers and them have also used the neutral sphingomyelinase inhibitor, GW4869, to inhibit the secretion of exosomes, implying the fact that exosomes are secreted through the ceramide pathway. Wei et al. determined that RAB31 mediates the formation of intraluminal vesicles through flotillin protein and inhibits the lysosomal degradation of MVB, thereby promoting the release of exosomes [28]. Analysis of the composition of exosomes showed that the exosomes contained abundant types of proteins, such as tetraspanins [29], annexins [30], integrins [31], chaperones [32]. Importantly, exosomes do not contain any nuclear, mitochondrial, endoplasmic reticulum or Golgi-derived proteins, and lysosomal proteases [33,34]. Exosomes are also rich in lipids [35], DNA [36,37], mRNA [38] and non-coding RNA, including miRNA [38], lncRNA [39], circRNA [40]. MISEV2018 didn't specifically defined molecular markers that can specifically characterize

each extracellular vesicle subtype, but still proposed three types of protein markers that need to be analyzed in the preparation of extracellular vesicles [15]. The first category is the transmembrane protein or GPI-anchored protein used to prove the membrane structure, such as the most commonly used tetraspanins CD63 [41], CD81 [42] and CD9 [43]. Although in addition to exosomes, they were also found in other large or small extracellular vesicles(ectosomes) [44,45]. The second category is cytoplasmic proteins, that can bind to the cytoplasmic sequence of membrane or transmembrane proteins, such as TSG101 [46] and Alix [43]. The third category is negative marker proteins that exclude contamination, such as apolipoproteins A1/ and B and albumin, which are easily separated from extracellular vesicles [15,47]. For functional molecules, the types and contents of exosomes from different cell sources are different. For example, the miRNAs in exosomes derived from tumor cells are different from those in normal cells [48].

3. Advantages of exosomal communication

There is not only one way in which exosomes are taken up by recipient cells. It may be through endocytosis, pinocytosis, phagocytosis, or fusion with the cell membrane that their contents are internalized by the proximal or distal recipient cells [49]. This is the way exosomes serve as a means of intercellular communication. This feature of exosomes, coupled with their own nanometer-scale particle size, enables them to be used as one of the research methods of excellent materials for drug delivery [50]. Based on the consistency of structure and function, the composition and content of cargo carried by exosomes are different, which also makes them have specific cell communication functions in different environments. What the proteins highly expressed in tumor-derived exosomes (TEs) help to promote the proliferation and metastasis of tumor cells [47,51]. For example, TEs have been reported to carry a tetraspanin, Tspan8, which is taken up by recipient endothelial cells and effectively induces tumor angiogenesis [19]. High levels of integrins have been detected in exosomes derived from tumor cells, which have been proven to be one of the most effective substances for tumor metastasis [51]. Exosomes have been determined to contribute to the formation of niches before tumor metastasis. They can be taken up by cells far away from the primary tumor and eventually metastasize [49]. miRNAs change is often observed in tumor cells used as molecular markers for disease diagnosis [52]. For example, Fan et al. proposed a new platform for rapid capture of TEs to test the types and levels of miRNA in exosomes, and analyzed the specific changes of miR-21 derived from exosomes in the serum of patients with lung cancer, liver cancer and breast cancer [53]. Another study showed that the levels of miR-21, miR-27a and miR-375 can be used as potential markers for clinical diagnosis of breast cancer [54]. Similarly, Hong et al. was found that miR-4435 is a potential circulating miRNA biomarker for colorectal cancer [55].

TEs were found to carry a variety of tumor antigens and were presented by DCs to trigger cytotoxic T lymphocytes

(CTls) response [56]. As early as 1998, studies have shown that exosomes derived from DCs can trigger specific CTls to eliminate tumors. It may be because the exosomes derived from DCs obtain abundant major histocompatibility complex (MHC) molecules from the parent cells, which activate immunity after being taken up by CTls [57]. Additionally, DCs-derived exosomes (DEs) stimulate a stronger cytotoxic response and anti-tumor immunity than TEs, which may be closely related to the communication function of DEs to promote antigen presentation [58]. The role of immune cell-derived exosomes in resisting tumor proliferation cannot be ignored. Perforin was detected in exosomes derived from natural killer cells (NK cells), and it seems to be effective in inducing cell death after being taken up by tumor cells as receptors [59]. Exosomes derived from CTls are also collected and used to study their potential effects [60].

However, the battle between the tumor and the immune system is not a simple process of ebb and flow, but a very complicated one. In this environment where cell-to-cell communication occurs frequently, exosomes are indispensably involved in the process. The physiological functions of exosomes in tumor immunoediting and their role in cancer immunotherapy will be described in detail below.

4. Exosomes in tumor immunoediting

Tumor immunoediting was a hypothesis first put forward by American scholar S in 2002, which was that the immune system has contradictory dual effects on tumors [7]. The dual role is reflected in three phases: elimination, equilibrium and escape [61]. In the elimination phase, the rapid growth of tumor tissues promotes the production of new blood vessels, and antigenic tumor cells are recognized and eliminated by immune effector cells [62]. However, some tumor cells with genetic mutations have lower immunogenicity and cytotoxicity to avoid being eliminated by immune cells, which is the equilibrium phase [63]. By reducing the expression of related antigens or up-regulating immunosuppressive receptors/ligands to destroy T cell responses, or even secreting immunosuppressive factors and stimulating the activation of immunosuppressive cells to change the microenvironment, tumor cells gradually proliferate and even metastasize, and develop to the escape phase [63]. In the three phases of tumor immunoediting, the communication between tumor tissues and the immune system mediated by exosomes is frequent, especially in the most studied tumor escape phase [64–68].

In order to accurately collect reports on exosomes and tumor immunity, we propose a novel related literature extraction framework (Fig. 2), which is based on a hybrid biomedical knowledge network and includes a variety of different knowledge and information sources to extract literature in PubMed. Entities and relationships are represented and managed in structural knowledge such as ontology, semantic web and knowledge graph, and non-structural knowledge such as research literature, medical case reports, and textbooks. We use “exosome or small extracellular vesicle”, “tumor or cancer” and “immune microenvironment” as input keywords and get 726 results through PubMed FTP server. We select some of the words

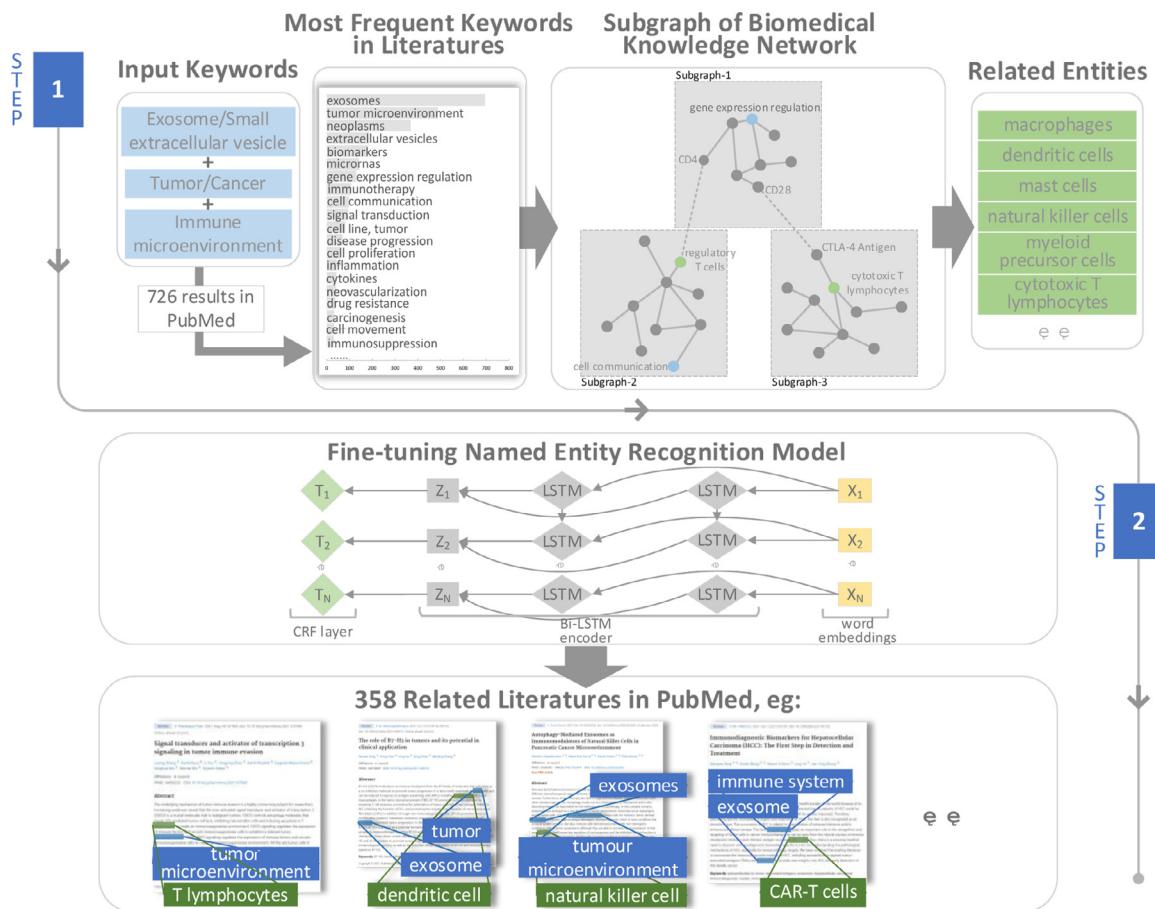


Fig. 2 – The framework is used to explore relevant literature on exosomes and tumor immunity. Step 1: input keywords, search all in PubMed, and count the frequency of keywords. Our keywords are “exosome/small extracellular vesicle”, “tumor/cancer” and “tumor microenvironment”. After screened, 726 articles were obtained. Based on the biomedical network and MORM model to expand and search for related entities, finding relevant entities such as “macrophages”, “dendritic cells”, “mast cells”, “natural killer cells”, “myeloid-derived suppressor cells” and “cytotoxicity T lymphocytes”; Step 2: Fine-tune the named entity recognition model to search for the most relevant documents describing the relationship between keywords and entities. In the end, we obtained 358 target documents.

according to the frequency of keywords and MeSH terms in these 726 articles. Based on the biomedical knowledge network and relationships extraction model, we will expand the entities which are related to the keywords and limit the type of related entities to cell. Multi ontology relatedness model (MORM) [69] is applied to explore indirect or implicit relationships between biological concepts. We use a scoring function to normalize the weights of each entity relation between (0–1) [70]. The scoring function is shown below:

$$\text{score}_i = \frac{W_i}{\max(W_i)}$$

where W_i is the credibility of entity relationship. In this paper, MORM includes 225 cell entities and 18,910 relatedness links between cells and other biomedical entities such as phenotype, disease and chemicals. Secondly, we fine-tuning a named entity recognition model based on Bi-LSTM encoder [71], which has pre-trained on 100 million PubMed abstracts and PMC full papers, to recognize the related entities

accurately. We use a standard CRF layer at the end of encoder, where Z_N is denoted as the output of Bi-LSTM encoder. Given the entity label $Y = \{y_1, y_2, y_3, \dots, y_N\}$. The probability of the sequence label is calculated by:

$$P(Y|Z) = \frac{\exp(\sum_i (W_{y_i} Z_i + b_{(y_{i-1}, y_i)}))}{\sum_{y'} \exp(\sum_i (W_{y'_i} Z_i + b_{(y'_{i-1}, y'_i)}))}$$

where y' denotes label sequence. W_{y_i} and $b_{(y_{i-1}, y_i)}$ are weight matrix of training parameters. The final output of our framework is to return the related literature which describe the relationships between input keywords and related entities.

4.1. Elimination phase: immune activation

4.1.1. Promote antigen presentation

Raposo et al. firstly proved that exosomes released by B lymphocytes bound to MHC class II and played the role of

Table 1 – Cancer immunotherapy of different types of cancer with modified exosomal carriers.

Cancer type	Exosomes	Source cells	Modification/ Effective cargo	Affected cells	Refs.
Melanoma	CpG-SAV-Exo	B16BL6	CpG DNA-modified exosomes expressing SAV-LA	DC cells	[152]
	CIITA-Exo	B16F1	MHC class II molecules and CD86	DCs/ CD4+ T cells	[160]
Breast cancer	SMART-Exo	Expi293	anti-human CD3 and anti-human HER2 antibodies	SK-BR-3, HCC, MDA-MB-468, Human PBMCs	[161]
	Dox @ Exo-PH20-FA	HEK 293T	human hyaluronidase, folic acid, doxorubicin	PC3, 4T1, HK-2, M2 macrophages	[151]
Colon carcinoma	CAR-Exo-CTX or CAR-Exo-TTZ	CAR-T	cetuximab scFv or trastuzumab scFv	MCF-7 EGFR or MCF-7 HER2	[168]
	PTX-M1-Exo	M1 macrophages	Paclitaxel	4T1	[170]
Lung cancer	mTEx	MC38	murine IL-12	Immature DCs	[158]
	Exo/IL-18	LS-174T	AdhIL-18	DCs/CD8+ T cells	[153]
Lymphoma	CD40l-Exo	3LL Lewis lung cancer cell	CD40 ligand gene	BMDCs	[165]
Hepatocellular carcinoma	Exosomes	A549	Rab27a overexpression vector	DCs/ CD4+ T cells	[147]
	Exo/IL-2	E.G7-OVA	IL-2	Th1/CTLs	[135]
Pancreatic ductal adenocarcinoma	iExo-OXA	bone marrow mesenchymal stem cells	Galectin-9 siRNA	M2-TAMs, CTLs	[154]
Multiple myeloma	Exo-TNF- α	J558	TNF- α	CD8+ T cells	[162]
Renal cell carcinoma	Exo / IL-12	RC2	G250, Glycolipid anchored IL-12 (GPI-IL-12)	CTLs	[155]
Leukemia glioblastoma	RDE	RenCa cells	GM-CSF,IL-12	CD8+ T cells	[163]
	CTX-poly I:C-Exo	L1210	CTX, poly I:C	spleen cells	[156]
	DEXCRCL-GL261	DCs	Cell Lysate	CTLs	[131]

antigen presentation [72]. The exosomes derived from the famous antigen-presenting cell DCs are detected to promote the antigen presentation mechanism and they are quickly used as one of the strategies of cancer immunotherapy. The therapy strategy is arranged in Part 5 in detail. Andre et al. proved that DEs with MHC class I to stimulate CD8+ T cells in vitro [73]. Lysosomal-associated membrane protein 2 may help DCs internalized antigens to be enriched in exosomes to obtain high immunogenicity [74]. Except for MHC molecules, costimulatory molecules (CD86) and cell adhesion molecules (ICAM-1) have also been shown to be delivered to immature DCs, T cells or B cells through DEs [75]. In more detail, the antigen presentation effect of extracellular vesicles (including exosomes) derived from antigen presenting cells was shown in the work of Lindenbergh et al. [76]. Additionally, Exosomes carrying tumor antigens were isolated and detected from tumor cell culture medium cultured *in vitro* or ascites and blood of tumor patients [77,78]. Lymphocyte cytosolic protein 1, a membrane-associated antigen that stimulates immune responses, has been detected in exosomes secreted by chronic lymphocytic leukemia cells as well [79]. These exosomes carrying MHC-antigen peptide complexes are recognized by CD4+ T cells or CD8+ T cells, and then activate the immune response [80]. Alternatively, they have also been found to stimulate the initial DCs to become mature DCs

and indirectly promote antigen presentation [81,82]. More over some researchers have observed that immature DCs show higher exosomal internalization ability than mature DCs [82].

4.1.2. Enhance the cytotoxicity of immune cells

In addition to indirectly enhancing immune effects by promoting antigen presentation, heat shock protein 70 carried on the surface of TEs may interact with Bag-4 ligand to directly stimulate the activation of NK cells [83]. Not only activate NK cells, further, Bag-6 expressed on TEs will bind to the surface receptor NKP30 and induce the up-regulation of granzyme B, Interleukin 2 (IL-2), tumor necrosis factor-alpha (TNF- α), interferon-gamma (IFN- γ) in NK cells, triggering NK cells-mediated cytotoxicity [84,85].

DEs carry high levels of intercellular adhesion molecule 1 (ICAM-1) enhancing their capture by other immune cells [86]. In addition to ICAM-1, MHC peptide complexes and T cell costimulatory molecules found on DEs may induce antigen-dependent T cell immune responses [87]. The ability of exosomes secreted by mature DCs to induce antigen-specific T cell activation *in vitro* is 50 to 100 times higher than that of immature DCs [88]. More importantly, TNF- α , FasL and TNF-related apoptosis-inducing ligand (TRAIL) expressed on the surface of DEs directly induce apoptosis of

tumor cells [89]. Therefore, the enhanced immune-mediated cytotoxicity by exosomes may be a potential motivation for tumor immunotherapy.

4.2. Equilibrium phase

In the equilibrium phase of immunoediting, tumor cells with weakened antigenicity will neither be recognized by the immune system, nor will they overgrow due to the strong elimination of the previous phase [63]. Therefore, it seems impossible to detect a visible tumor in the patient's body [62]. At this phase, there has been no discussion about exosomes yet. This balance between the tumor and immune system is not static. Tumor cells may undergo genetic mutations under the pressure of the immune system, which is the critical moment for immunoediting. When the accumulation of this gene mutation reaches a certain level, the immune system will be in a weak position and enter the phase of immune escape [90].

4.3. Escape phase: immunosuppression

After the equilibrium phase, the mutation effect accumulated by tumor cells for a long time makes it exhibit low immunogenicity and express specific immune ligands, so as to escape the surveillance of the immune system. TEs secreted more also transmit more complex information communication at this phase than normal cells [91]. It has been proved that exosomes are involved in inhibiting the differentiation and maturation of DCs, limiting the cytotoxicity of NK cells, inhibiting the proliferation of B cells [92–98], and possessing the immune regulation ability of CTLs [99–101]. In short, TEs may be an important way to mediate tumor immunosuppression in the escape phase.

4.3.1. Regulatory proteins or cytokines suppress immune effects

In the process of tumor development, tumor-associated antigens and specific antigens are lost under the pressure of the immune system and the recognizable proteins are down-regulated, which reduce immune recognition and clearance. The expression of transforming growth factor- β (TGF- β) in TEs induces the phenotypic differentiation of myeloid-derived suppressor cells (MDSCs) and inhibits the maturation of DCs and T lymphocyte proliferation [102]. In addition to cytokines, the exosomes of prostate cancer cells carrying miR-212-3p inhibit the expression of MHC II and induce immune tolerance in DCs [94]. The exosomes of pancreatic cancer not only mediate the down-regulation of TNF- α and IL-12 in DCs by miR-203, but also deliver miR-23a to target CD107a in NK cells to exert immunosuppressive effects [103,104]. The exosomes encapsulating miR-92b transfer from liver cancer cells to NK cells, causing the down-regulation of CD69 and the inhibition of NK cell-mediated cytotoxicity [83]. Exosomes derived from nasopharyngeal carcinoma cells have been shown to significantly down-regulate IL-2, IFN- γ and IL-17 and inhibit the proliferation and differentiation of T cells [105]. TEs express apoptosis activating factors FasL and TRAIL, which

directly induce CD8+ T cell apoptosis expect for inhibiting immune cell function indirectly [100].

4.3.2. Up-regulation of inhibitory immune checkpoint molecules

Tumor cells overexpress some specific immune checkpoint molecules to inhibit the activation of T cells [106], such as cytotoxic T lymphocyte-associated antigen 4 (CTLA-4) [107], programmed death 1 (PD-1) [108], T cell membrane protein 3 (TIM-3) [109] and lymphocyte activation gene 3 (LAG-3) [110], etc. Studies have shown that these inhibitory immune checkpoint molecules are also expressed on the surface of TEs [111–113]. This evidence provides a possible conviction for the immunosuppressive effect of exosomes on T cells. NKG2D is a cytotoxic receptor shared by NK cells and T cells. It has been detected that NKG2D ligand is expressed on the surface of exosomes produced by mesothelioma cells [114], and its specific binding with NKG2D to confuse NK cells and T cells and mediate tumor immune escape [115].

4.3.3. Induction of differentiation and proliferation of immunosuppressive cells

TEs may induce bone marrow-derived monocytes to differentiate into MDSCs and prevent differentiation into DCs and macrophages [116]. HSP70 highly expressed on the surface of exosomes participate in the activation of MDSC through TLR2, one of the toll-like receptor family members [117,118]. The phenotype of T lymphocytes classified as immunosuppressive regulatory T cells (Tregs) may be induced by TEs [119,120]. This upregulation of Tregs mediated by TEs is also one of the potential pathways for tumor immune escape [121]. In hepatocellular carcinoma immune escape studies, TEs also induced an increase in the number of B regulatory cells and significantly inhibited the activity of CD8+ T cells [122].

4.3.4. Inducing polarization of macrophages

TEs are involved in the remodeling of the tumor microenvironment including the induction of macrophage polarization [123]. As an important part of the tumor microenvironment, tumor-associated macrophages (TAMs) are divided into inflammatory macrophages (M1) and anti-inflammatory macrophages (M2), which M2 phenotype is conducive to the immunosuppressive tumor microenvironment [124–126]. TEs mediate ncRNA to regulate the M2 polarization of TAMs, such as miR-21, miR-222 [127], lncRNA TUC339 [128], etc., which helps reduce the release of pro-inflammatory factors to enhance tumor proliferation and metastasis [123].

In short, exosomes from different sources affect the differentiation and maturation of a variety of immune cells, potentially affecting immune activation and immune suppression in the process of tumor immunoediting. They have been shown to mediate the delivery and function realization of different cargoes, which is extremely attractive for the consideration of tumor immunotherapy strategies.

5. Exosome-based tumor immunotherapy strategies

5.1. Cancer vaccine

Cancer vaccines that awaken the human immune system against cancer through tumor cell-related antigens are widely used in tumor treatment research [129]. In the clearance phase of tumor immunoediting, exosomes played an excellent role in immune activation. Since exosomes can cause immune activation, whether TEs carrying tumor-associated antigens or exosomes derived from DCs presenting antigens, they can be used in the field of anti-cancer vaccines [6]. After capturing the antigen, DCs seem to pack the MHC antigen peptide complex with immunostimulatory factors into exosomes [130]. These exosomes transfer MHC antigen-peptide complexes to inactive DCs that in the lymph nodes and help these DCs acquire the ability to stimulate CTLs. Exosomes from DCs may also be loaded with chaperone-rich cell lysates produced by tumor cells [131]. Under this concept, DCs-derived exosomal vaccines have been tested in phase I clinical trials in patients with advanced non-small cell lung cancer and metastatic melanoma [132]. As congenital phagocytes, macrophages are educated in the tumor microenvironment to become anti-inflammatory phenotype promoting tumor proliferation and migration. TAMs-derived exosomes have the potential as vaccine adjuvants as well. A study using exosomes derived from pro-inflammatory macrophages to act on melanoma showed that these exosomes can be used as vaccine adjuvants to enhance the response of CTLs [133]. TEs can also be studied as one of the potential anti-tumor vaccines for cancer immunotherapy [134]. They can be engineered to directly cause specific immunity as an acellular vaccine [135], or can be a cell vaccine co-incubation with DCs [136]. Compared with exosomes derived from tumor cells, exosomes derived from DCs stimulate a stronger CD8+ response and anti-tumor immunity because TEs can also cause immunosuppression [136,137]. The study of exosomes as cancer vaccines has attracted more and more attention from researchers, and has broad prospects in tumor immunotherapy. However, for clinical trials, the feasibility of large-scale production of exosomes and the safety of in vivo administration of exosomes are still emphasized.

5.2. Drug delivery vehicles for tumor immunotherapy

As a natural biological carrier, exosomes have drug delivery advantages over other non-biological nanocarriers [138–140]. Exosomes have been fully demonstrated for cargo delivery and regulation of different cellular processes in the tumor immunoediting. Due to the widespread expression of transmembrane proteins [141], it can prevent exosomes from being phagocytosed by mononuclear macrophages in the circulation, and significantly reduce the blood clearance rate than other vectors [142]. What the important points are that exosomes show excellent ability to penetrate tissue barriers due to nanoscale size and cargo delivery advantage [143–145]. Based on these advantages, exosomes, as endogenous nanocarriers, have received great attention

in the research of tumor immunotherapy. As mentioned above, exosomes without modification can trigger potent antigen-specific antitumor immune responses [146–148], or use exosomes secreted by mesenchymal stem cells to induce the anti-tumor response of NKT cells [149]. The engineering modification of exosomes as a drug delivery vehicle is a common research hotspot. The first common method is to carry out structural engineering of the exosomal membrane, mainly through targeted modification of surface ligands for the delivery of drugs [150–153]. Exosomes can also be applied as the carriers of both chemotherapy drugs and siRNA, which were also beneficial to improve the drug delivery efficiency [154–156]. Table 1 lists the immunotherapy studies of some exosomes in different types of tumors. We briefly classified and summarized the strategy of the articles on engineered exosomes listed in the table, the engineered exosomes secreted by genetically modified cells can more effectively induce anti-tumor immune responses, such as co-delivery of tumor antigens and adjuvants [152,157–160], add different types of surface-displayed monoclonal antibodies [158,161], overexpress cytokines [162,163], etc. Some genetically modified membrane vesicles can also play a similar immunogenic role by specifically expressing viral antigen as a versatile antigen-delivery system [164,165]. Engineered exosomes help to enhance DCs antigen presentation [159] and promote cellular immune response based on T cells [166,167]. For example, Lu et al. verified that exosomes derived from DCs expressing alpha-fetoprotein can activate antigen-specific cellular immunity and promote the release of immune cytokines in three murine models of hepatocellular carcinoma [146]. T cells expressing chimeric antigen receptors (CAR-T) are a very promising method of cancer immunotherapy. However, CAR-T cells are prone to cause a cytokine storm in the body and cannot accumulate in the tumor microenvironment. This may lead to poor treatment effect. Report that CAR-T cells release exosomes, which carry chimeric antigen receptors (CAR) on their surface. CAR-exosomes do not express PD-1, and recombinant PD-L1 treatment will not reduce their anti-tumor effects compared with CAR-T treatment [168]. More types of cell-derived exosomes are collected and modified for tumor immunotherapy attempts, such as macrophages [133,169–172], NK cells [173,174]. Importantly, exosomes can be obtained from patient serum, which makes it more feasible to apply them to clinical treatment of tumor immunotherapy [100,175].

6. Future directions

Exosomes are heterogeneous extracellular vesicles with a variety of cargo, including lipids, proteins, nucleic acids and carbohydrates. The secretion of exosomes is related to the transport of endosomal and plasma membranes, and the sorting of goods depends on ESCRT or other pathways, such as lipid rafts. The secreted external vesicles are internalized by the recipient cells through endocytosis and receptor binding to mediate information transmission. Exosomes have abundant cell communication functions, including proximal and distal communication. It plays an important role in the

biogenesis and development of tumors. A large number of exosomes secreted by tumor cells seem to have a dual role, which can not only activate immunity, but also inhibit the function of immune cells, promote immune suppression, and form a tumor-promoting immune microenvironment. Based on the concept of tumor immunoediting, we have sorted out the role of exosomes in it, including immune activation that promotes antigen presentation and enhanced response, immune escape that reduces immune clearance by regulating cytokines and immune checkpoint proteins, or stimulates the proliferation of immunosuppressive cells, or promote tumor-related macrophage polarization. Taking advantage of the flexible and rich information transmission functions of exosomes, many researchers have proposed methods for cancer immunotherapy based on exosomes, such as cancer vaccines and various drug delivery vehicles. Exosomes derived from DCs, macrophages, tumor cells and even T cells have been studied for cancer immunotherapy, and there are many promising works. For example, an exciting study proved that exosomes derived from CAR-T cells expressed CAR but not PD-1 and they showed stronger anti-tumor ability and are safer as a cellular vesicles [168]. Despite great achievements in exosome research, challenges remain. The separation and purification technology of exosomes is still a major challenge. The current methods for obtaining exosomes include: ultracentrifugation [176], size exclusion [177], precipitation [178], immunoaffinity capture [177]. However, these methods have different disadvantages, such as high time cost, low yield, and impurity contamination [179]. The recent application of microfluidic technology to the separation of exosomes may be a more promising method because it is more efficient [180]. Related content was recently published in the review by Ding et al. [14]. Another challenge is the complexity of the cargo carried by engineering exosomes. The methods for loading drugs on exosomes include co-incubation [181], electroporation [182], ultrasound [183], liposome-based loading [184] and loading of gene products based on biogenesis [185]. The storage conditions of exosomes are also a point that needs to be considered in research and application. The routine storage is at -80 °C, while lyophilization may be a consideration that does not rely on ultra-low temperature [186]. Overcoming these challenges and standardizing them is the basis for improving the possibility of achieving precise treatment of exosomes in tumor immunity and other diseases. Of course, the exciting thing is that there are more and more researches to try to solve these problems and exosomes still have broad research value and application prospects as biologically derived nanovesicles.

Conflicts of interest

There are no conflicts of interest. The authors alone are responsible for the content and writing of this article.

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