



Published in final edited form as:

ACS Sens. 2021 August 27; 6(8): 2787–2801. doi:10.1021/acssensors.1c01133.

State of Sweat: Emerging Wearable Systems for Real-Time, Non-Invasive Sweat Sensing and Analytics

Roozbeh Ghaffari^{1,2,3}, Da Som Yang¹, Joohee Kim¹, Amer Mansour⁴, John A. Wright³, Jeffrey B. Model^{1,3}, Donald E. Wright³, John A. Rogers^{1,2,3,5,6}, Tyler R. Ray^{7,8,*}

¹Querrey Simpson Institute for Bioelectronics, Northwestern University, Evanston, IL, 60202 USA

²Department of Biomedical Engineering, Northwestern University, Evanston, IL, 60202 USA.

³Epicore Biosystems, Inc., Cambridge, MA, 02139 USA

⁴Division of Biological Sciences, The University of Chicago, Chicago, IL, 60637 USA.

⁵Departments of Materials Science and Engineering, Mechanical Engineering, Electrical and Computer Engineering, Chemistry, Northwestern University, Evanston, IL, 60202 USA

⁶Department of Neurological Surgery, Northwestern University Feinberg School of Medicine, Chicago, IL, 60611 USA.

⁷Department of Mechanical Engineering, University of Hawai'i at M noa, Honolulu, HI 96822 USA.

⁸Department of Cell and Molecular Biology, John A. Burns School of Medicine, University of Hawai'i at M noa, Honolulu, HI 96813 USA.

Abstract

Skin-interfaced wearable systems with integrated colorimetric assays, microfluidic channels, and electrochemical sensors offer powerful capabilities for non-invasive, real-time sweat analysis. This review details recent progress in the development and translation of novel wearable sensors for personalized assessment of sweat dynamics and biomarkers, with precise sampling and real-time analysis. Sensor accuracy, system ruggedness, and large-scale deployment in remote environments represent key opportunity areas, enabling broad deployment in the context of field studies, clinical trials, and recent commercialization. On-body measurements in these contexts show good agreement compared to conventional laboratory based sweat analysis approaches. These device demonstrations highlight the utility of biochemical sensing platforms for personalized assessment of performance, wellness, and health across a broad range of applications.

Graphical Abstract

*Corresponding Author Tyler R. Ray, raytyler@hawaii.edu.



Keywords

health monitoring; wearable sensors; epidermal microfluidics; lab-on-chip; flexible electronics; biosensors; sweat analysis; eccrine sweat

Introduction:

The shifting paradigm in clinical practice to an evidence-based care underscores the critical need for an expanded suite of capabilities for the rapid and continuous assessment of digital and metabolic biomarkers relevant to human health^{1,2}. Traditional evidence-based approaches to patient care, which relied on the informed opinions of medical practitioners for selection of a course of therapy, have yielded to evidence-based clinical strategies that employ quantitative metrics to inform therapeutic interventions and treatment efficacy³. Although recent studies demonstrate the power of this approach in assessing therapeutic benefit (e.g., surgical interventions^{4,5}, off-label drug use⁶⁻⁸), evidence-based medicine remains, by nature, reactive—capable of supporting treatments for an active, symptomatic disease state. Extending evidence-based approaches that enable proactive interventions during periods of healthy living and early onset of disease require the advent of new digital health tools and analytics that not only track physiological health status but also alert to subtle perturbations.

Skin-interfaced wearable systems offer multiparameter sensing capabilities to address these limitations by monitoring the diverse range of signals arising from natural physiological processes⁹. Novel instruments that track the biochemical (i.e. electrolytes, metabolites, hormones), biophysical (i.e. temperature, biopotentials, hemodynamics), and kinematic (e.g. movement, posture, gait) signals from the body, provide critically valuable information about overall health status¹⁰. Conventional wearable systems support the quantitative assessment of select physiological parameters via wrist-worn (e.g. smart watches), chest-strapped (e.g. heart-rate monitors), and apparel-integrated (e.g. sun exposure monitors) device form-factors. Continuous glucose monitors (CGMs) have been commercialized and widely adopted, highlighting the enormous potential for real time biochemical sensing of glucose levels for diabetics. For devices worn continuously, the ubiquitous nature of such systems can yield important health insights from a limited range of health markers¹¹. Nevertheless, these conventional platforms typically lack the ability to non-invasively

characterize multiple biomarkers and the underpinning metabolic processes essential to overall health.

Blood-based analysis is the primary approach to monitoring body chemistry via invasive sampling (blood draw) and expensive, centralized laboratory equipment¹². Biofluids such as tears, interstitial fluid, and sweat are attractive alternatives for non-invasive sampling and analysis.¹³ Of these alternative biofluids, eccrine sweat is of particular interest^{12,14} on account of rich composition of biochemical information including micronutrients (electrolytes), metabolites, hormones, proteins, nucleic acids, and exogenous agents¹⁵⁻²⁰ and suitability for facile, non-invasive collection. Emerging classes of skin-interfaced wearable platforms harness recent advances in soft microfluidics, flexible/stretchable electronics, and electrochemical sensing technologies to support the continuous or intermittent assessment of sweat composition in a variety of conditions or settings^{15,21-24}. The resulting time-dynamic insight these platforms offer into metabolic activity is critical for creating a comprehensive understanding of health, nutrition, stress, and wellness status.

This perspective offers an overview of the current state-of-the-art for wearable sweat biosensors, with particular emphasis on the application use-cases for these sensors. The nascent field is of considerable interest with recent reviews^{11,14,20,25-54} contextualizing the progress of wearable sweat sensors within the scope of skin-interfaced devices^{9,15,18,23,24,55}, sensing technologies^{13,22,24,56-63}, specific applications^{10,17,19,21,57,61,64,65}, material systems^{66,67}, and fabrication methods⁶⁸. By contrast, this perspective highlights the most advanced translational embodiments spanning the fundamental use cases for these platforms in relationship to sensing targets. A short introductory section summarizes key considerations in terms of sweat collection and the sensing architectural constructs that form the foundation of these wearable systems. The section that follows broadly classifies the application targets according to athletic performance and clinical diagnostics with representative examples of the current approaches. The perspective concludes with a discussion of efforts to expand overall utility of these sensors for diagnostic applications, in which clinical validation of sensor technologies will be critically important for commercialization.

Sweat analysis: sampling methods and analytical approaches

Wearable, sweat-based platforms must address sweat collection for a diverse range of applications including passive sweat in fragile infants to intense physical exertion in athletes and warfighters. These sensors must function in arid, hot temperatures, under high humidity conditions, and even during aquatic activities. Across all use cases these platforms must establish and maintain a conformal, intimate interface with the epidermis to support robust sweat collection and analysis. Soft, wearable microfluidic devices utilize biocompatible, low-modulus elastomeric (poly(dimethylsiloxane), PDMS) substrates and hypoallergenic silicone adhesives to support a robust, watertight interface for the pristine capture and clean storage of sweat. Activated eccrine sweat glands excrete sweat at a natural pressure sufficient to route sweat through networks of microfluidic channels and reservoirs⁶⁹. As detailed in recent reviews^{15,24,43,45,62,70}, the integration of optical (e.g. colorimetric^{22,71-80}, fluorescent⁸¹⁻⁸⁴ assays) and electrochemical^{22,23,30,45,85} sensors, either singularly or in

tandem, enable quantitative analysis of sweat biocomposition. Constraints from operating conditions, body-interfacing locations, and time-dynamic biochemical variations in sweat composition necessitate sophisticated lab-on-chip design strategies to obtain high-quality measurements. These competing requirements define the chemical sensor performance specifications for precision, sensitivity, selectivity, operational stability, operational lifespan, methodology of data transfer, and power requirements.

Simple, adequate stimulation of sweat remains a longstanding⁹⁶ and significant⁹⁷⁻⁹⁹ challenge for sweat-based analytical platforms. Intense physical activity, exposure to heat stress, and localized chemical inducement are the core methods for generating sufficient microliter volumes of sweat for biochemical analysis with suitability defined by target application^{16,100-104}. Whereas exercise-based stimulation serves as the primary means for athletic performance monitoring⁸⁶ (Fig. 1A), clinical diagnostics support on-demand analysis through the transcutaneous delivery of a cholinergic agonist via iontophoretic stimulation⁸⁷ (Fig. 1B) to activate localized sweat glands. For a given sensing application, a key consideration in tandem with mode of deployment (ambulatory vs. stationary individual) is the dependence of both the rate of sweat production¹⁰⁵ and biochemical composition^{100,106,107} on the stimulation method. Additionally, these methods are not amenable to applications that require frequent, repeated stimulation events (as comparable to blood glucose measurements). Recent efforts to support daily health assessments demonstrate the potential for collection of sweat at a consistent flow rate¹⁰⁸ generated either during showering⁸⁸ (Fig 1C) or by natural perspiration processes¹⁰⁹⁻¹¹¹ (Fig 1D). By virtue of the passive nature and circumvention of resource and exertion requirements, these alternative stimulation methods may significantly expand the breadth of potential applications for sweat analytics.

Emerging from early device designs⁷¹ of simple networks of microfluidic channels and reservoirs, current wearable microfluidic platforms employ a suite of sophisticated design strategies to collect and route sweat. Valves are a key component to many fluidic platforms and thus permit the direct capture and routing of sweat from the epidermis to target regions of a device in a programmatic manner. Most demonstrations⁹⁰ (Fig. 2A) are passive in nature (i.e. battery-free) relying upon fluidic resistance changes⁶⁹, one-time chemical reactions (e.g., sodium polyacrylate, a super-absorbent polymer)⁷⁴, or surface functionalization (hydrophobic/hydrophilic surfaces)¹¹² to control fluid flow via a series of irreversible stop-points. A recent device embodiment⁹¹ employs an active valve concept comprising the combination of thermo-responsive poly(N-isopropylacrylamide)-based hydrogel and wireless heating elements to enable dynamic control of sweat transport in response to physical actuation of hydrogel size (Fig. 2B). These valve concepts offer nuanced control over fluid routing⁴², which is key both for accurate sensor performance and for correlating the time-dynamic response of sweat constituents to physiological parameters (e.g., mental state, physical activity). Valves are of particular interest for optical sensing approaches. Colorimetric and fluorescence-based sensors operate by reacting a defined sweat volume with a chemical or molecular assay to generate an optical signal proportional to target analyte concentration. Integration of networks of valves enable fully passive optical sensors to “chronosample” sweat¹¹³ as described in Fig. 2A, either in time or fixed volumes, to provide quantitative measurements at defined intervals. By contrast, electrochemical

sensors, typically employed for continuous sweat monitoring, require constant flux of sweat over the sensor surface to maintain analytical performance^{30,45,49,50,114}. Integration of such sensors with networks of active or passive valves enables discrete activation of sensors, deconvolution of flow-rate effects, and programmed sensing at selected time intervals.

The expanding library of valving approaches, in combination with other emerging design concepts such as integrated mixing systems¹¹⁵, facilitates development of devices capable of high-precision sensing of sweat biomarkers. Research efforts seek expanded sensing capabilities to support the long-term, real-time monitoring of sweat biomarkers in a battery-free manner. Use of smartphone-based image analysis offers a simple, direct mode for biomarker analysis^{75,86,88,116,117}; however, this analytical pathway is ill-suited for assessing time-dynamic information in demanding applications (i.e. during active physical exercise). One recent embodiment⁹² employs sweat-activated galvanic cells to serve as a series of “stopwatches” to establish time stamps for passive colorimetric measurements during an activity period (Fig. 2C). Other approaches harness sweat-based biofuel cells⁹³ to generate sufficient power to record and store measurements from electrochemical sensors during an activity to be retrieved via a wireless data transfer at the conclusion of the testing period (Fig. 2D). Implementation of such strategies¹¹⁸⁻¹²⁰ enables epidermal microfluidic devices to support multiple sensing modes (optical/electrochemical) in a battery-free form factor.

As described elsewhere^{9,13,15,16,24,55,121-123}, eccrine sweat contains a wide range of metabolites, electrolytes, and xenobiotics that offer detailed clinical insight into disease states and valuable information regarding overall health. Many target sweat biomarkers are present only in extremely low concentrations²⁰. Transduction of meaningful signals from these low-concentration species requires careful consideration of strategies to mitigate sample loss, biofouling of sensor surfaces, sample contamination, and deconvolution of interfering factors⁴⁶. Optimizing device and sensor geometries yield powerful advantages in this context. Fig. 2E highlights a recent strategy⁹⁴ to reduce sample loss from device deformation with a device construct that directly integrates impermeable, rigid channels within a soft, compliant polymer matrix. Resistant to deformation from physical impact, the optimized device geometry maintains a robust, conformal interface with the epidermis to support sweat collection and analysis. A similar approach¹²⁴ offers improvements to the robustness of integrated sensors such as in the utilization of novel material designs to circumvent biofouling on the surface of electrochemical sensors. Both examples reduce or eliminate interference effects for devices during operation; however, certain biomarkers (sweat chloride for cystic fibrosis) may require ex situ analysis necessitating consideration of external contamination factors. Eliminating operator interaction through utilization of custom extraction hardware⁹⁵ represents one such strategy for obtaining a “clean” sweat sample free of interfering contaminants (Fig. 2F). In all cases, obtaining meaningful insight from wearable sweat sensors requires operational performance to remain invariant to external environmental factors¹³. To this end, recent efforts¹²⁵ seek to decouple target signals from interfering species, co-dependent biomarkers (e.g., pH, temperature), and other sources of noise (e.g., motion, biophysical signals). Further technical progress necessitates sophisticated sensing strategies and complex device designs to address these expanding challenges of sweat stimulation, fluid-handling, and contamination. Such developments are critical for obtaining meaningful physiological insight from sweat in a variety of

potential use cases. Such considerations must occur in tandem to the demands imposed by application-specific requirements.

Translational Applications

Wearable platforms for real-time sweat analysis represent a significant advancement for providing personalized and actionable insights across a variety of applications spanning athletic performance to daily health monitoring. Integration of advanced sensors and fluid-sampling designs coupled with soft, flexible substrates establishes a powerful foundation for expanding the suite of biochemical markers and physiological signals accessible to the wearer. The sections that follow highlight emerging epidermal microfluidic devices broadly categorized by use for performance health management and clinical diagnostics.

Performance Health Management

Many commercial demonstrations of performance driven wearable devices have focused on monitoring physiological and biomechanical signals during physical activity¹³¹. Initially developed for professional athletes, wide adoption of fitness trackers over the past decade illustrates the growing consumer interest in understanding the activity-dependent response of the human body to physical stress⁶⁵. Such insight is essential for reducing risk of injury, monitoring recovery times, and improving overall well-being. Although capable of assessing the core biophysical and kinematic signals for this purpose, these existing wearable platforms lack the sensing capabilities necessary to monitor metabolic health¹²⁶. This section describes the latest representative skin-interfaced microfluidic devices deployed for ambulatory metabolic health assessment.

Thermoregulatory sweat response is essential for maintaining homeostasis and gives rise to loss of water, electrolytes, and other sweat constituents during physical activity¹⁰². Excessive total sweat fluid and electrolyte losses could impair cognitive and athletic performance or result in severe conditions such as heat stroke or death¹⁰¹. These effects manifest as changes in sweat parameters (rate, composition) and tend to vary widely across individuals¹⁰⁰. Differences in physiology, training, activity-type, physical intensity, and surrounding environment necessitate personalized hydration strategies based on individual sweat profiles to ensure adequate fluid replenishment^{101,132}. Practitioners and athletes typically estimate whole-body sweat loss by recording changes in body mass after physical activity¹⁰³. This approach requires high fidelity measurements through careful adherence to testing protocols and precise accounting of fluid intake and urine loss during the exercise period to obtain meaningful, albeit retrospective, insight. By contrast, regional sweat sampling estimates whole-body sweat loss by collecting sweat from a localized anatomical site via absorbent pads, filter paper, or plastic coils and specialized, wired equipment¹³³. Although more practical, the absence of a standardized assessment method has historically restricted the utility of regional sampling resulting in the generation of only limited physiological insight.

Epidermal microfluidic devices offer powerful capabilities for accurately monitoring sweat dynamics by virtue of the conformal, fluid-tight interface. These devices harvest sweat

directly from sweat glands in a manner that isolates the sample from environmental contaminants thereby enabling precise, real-time characterization of sweat biomarkers. Although the performance of these sensing platforms has been extensively validated for a variety of biomolecular targets and sensor architectures^{23,43}, there is an absence of studies correlating regional measurements from such wearable sensors and the whole-body sweat response. A recent report¹²⁶ (Fig. 3A) represents the first large-scale systematic study (N = 312) correlating regional and whole-body sweat rate and sweat chloride measurements using a soft, flexible microfluidic patch. The device comprises two discrete networks of microfluidic channels which contain either an integrated colorimetric assay for quantifying sweat chloride concentration or a highly-visible dye to facilitate assessment of sweat volume. Use of a smartphone and companion app enables digital image capture and automated measurement of instantaneous sweat rate, sweat chloride, and total sweat loss. Contralateral comparisons of these epidermal microfluidic devices to absorbent patches in combination with whole-body sweat measurements in a controlled laboratory environment demonstrate good agreement between the predicted (from regional measurements) and measured whole-body sweat rate and sweat chloride concentrations (mean absolute error of 14% and 13% respectively), which serves as the basis for actionable hydration feedback.

Establishing a strong correlation between regional and whole-body sweat-based measurements represents a key step for developing new insights into the physiological relevance of sweat biochemical signals. In addition to fluid and chloride loss, the concentration of glucose^{125,134}, lactate^{135,136}, ammonia¹³⁷, and cortisol¹³⁸ in sweat have value for monitoring athletic training and conditioning. Varying dynamically with physiological status (diet, stress, overall health) and activity²⁰, biomarker concentrations also correspond to dynamic variations in instantaneous sweat rate^{125,139}. Recent efforts (Fig. 3B-D) offer the requisite temporal resolution of instantaneous sweat rate to deconvolve this variability with real-time continuous sensing strategies. Representative devices integrate electrical conductivity¹²⁷ (Fig. 3B), capacitive¹²⁸ (Fig. 3C), or temperature¹²⁹ (Fig. 3D) sensors with wireless data transfer and ultrathin batteries to support continuous monitoring of physiologically relevant sweat rates (0 to 5 $\mu\text{L min}^{-1}$). Conductive¹²⁷ or capacitive methods utilize electrode pairs embedded in microfluidic channels to measure the change in conductivity or capacitance across the channel as sweat fills. For the conductive method, a direct contact to sweat and electrodes, whereas the capacitive method relies on non-contact measurement of sweat fill into the device. An alternative non-contact approach measures real-time sweat flow rate using a localized heater embedded between two thermistors. This design architecture can quantify flow rates with high sensitivity and without direct contact within the microfluidic device. Sensing platforms that leverage real-time sweat rate measurements with highly sensitive and selective multiparameter sensors for monitoring low-concentration sweat constituents (e.g. cytokines) may yield further insights for assessing the health status of athletes during activity, recovery, and rest.

A logical progression for performance assessment is the development of bi-directional communication between the device and user upon detection of an anomalous physiological event (e.g. dehydration). Fig. 3E shows a skin-interfaced platform¹³⁰ that circumvents the need for user engagement during wear with the automated delivery of sensory warnings via sweat-triggered chemesthetic agents. The device deploys an effervescent pump to eject

menthol (or capsaicin) onto the epidermis when a dehydration condition is detected due to excessive sweat loss.

The device geometry and reversible visual sweat indicators permit the sensor to be manually reset after rehydration. In aggregate, these representative platforms represent key advances in establishing the compatibility of regional sweat analysis at prescribed anatomical locations with developing holistic personalized hydration strategies or for athletic performance monitoring.

Clinical Diagnostics

Prior to the advent of epidermal microfluidic sensors, few applications existed for clinical utilization of biochemical sweat analysis. Chloride is a critical sweat biomarker used in clinical diagnostics of cystic fibrosis (CF). Diagnosis of CF is perhaps the oldest sweat-based diagnostic based upon recorded instances from the Middle-Ages¹⁴⁷. Established clinically in 1959¹⁴⁸, quantitative evaluation of sweat chloride in neonates remains the only widely available method for confirmatory diagnosis of cystic fibrosis. Conventional clinical diagnostic methods are cumbersome; they utilize wrist-strapped devices to collect sweat from infants that often produce insufficient sweat for analysis. Recent work (Fig. 4A, 4) highlights the immense promise of wearable sweat sensors in mitigating such diagnostic and interfacing challenges. One recent demonstration¹⁴⁰ (Fig. 4A) utilizes a soft elastomeric microfluidic platform and a skin-safe adhesive to maintain conformal integration with the skin to facilitate near perfect efficiency in collecting sufficient sweat volumes for analysis (N = 51, infants to adults). Integration of colorimetric chloride sensors with advanced image processing techniques enables smartphone-based image analysis to quantify sweat chloride levels with an accuracy similar to the established clinical method (coulometric titration) in a limited study (N = 5, adults). Another embodiment¹⁴⁹ integrates a salt-bridge based potentiometric sensor with wireless Bluetooth communications to monitor sweat chloride concentration from a smartphone in real-time during exercise. A small field study highlights performance for adult patients with (N=10) and without (N=10) cystic fibrosis. Although these platforms and others^{150,151} demonstrate immense potential to improving cystic fibrosis diagnostics, substantial expansion of clinical study populations is requisite for establishing operational performance equivalence to current clinical methods¹⁵².

Resulting from recent interest in utilizing sweat as a non-invasive target for metabolic health monitoring, considerable research efforts seek to expand the utility of diagnostic sweat testing from CF and atopic dermatitis to diabetes. Self-testing and frequent assessments of blood glucose concentration are vital components to diabetic health management strategies¹⁵³. Conventional sensing approaches for daily assessment rely on invasive, painful, skin-piercing microneedle sampling (finger prick). Although continuous glucose monitoring systems^{154,155} may mitigate the need for frequent self-testing, development of a non-invasive, pain-free glucose monitoring device remains of intense academic and commercial interest. Sweat represents an attractive biofluid in this context as recent studies demonstrate a linear correlation between sweat and blood glucose levels¹⁵⁶⁻¹⁶⁰. One recent demonstrator device⁸³ (Fig. 4C) employs a ratiometric fluorescence sensing strategy to detect the onset of hyperglycemia during sleep. A simple wearable pad containing

co-immobilized with functionalized dual-fluorescence nanohybrid substrates (luminescent porous silicon nanoparticle/carbon quantum dot structure with bimetallic nanoparticles) and glucose-oxidase measures sweat glucose concentration by monitoring a proportional color shift (red to blue) under UV illumination using a smartphone camera.

A recent paper¹¹¹ (Fig. 4D) reports utilization of Janus-wettability (hydrophobic/hydrophilic) textile band to self-pump microdroplets of sweat from the epidermis to functionalized chronoamperometric sensing electrodes to monitoring concentrations of glucose, lactate, Na⁺, and K⁺ in sweat. Another approach¹¹⁶ (Fig. 4E) achieves wireless, battery-free sweat glucose monitoring during physical exercise from biofuel cell glucose sensors, near-field communication (NFC) technology for data retrieval, and smartphone. The biofuel cell-based glucose sensor generates electrical signals in proportion to the concentration of glucose, which circumvents the need for a potentiostat (as required for amperometric sensors) thereby minimizing overall device size. The integration of colorimetric sensors for sweat chloride and pH in addition to biofuel cell lactate sensors permits simultaneous multiparameter analysis of metabolic activity and overall physiological state.

Other wearable sensor designs seek to harness blood-correlated biomarkers beyond chloride and glucose (e.g. lactate¹⁶¹⁻¹⁶⁵, ethanol¹⁵⁶, cortisol^{166,167}) to address diagnostic challenges related to diabetes and other diseases. Recent examples of wearable electrochemical sensing platforms demonstrate the promise of sweat analytics for monitoring biomolecular changes relevant to diseases such as gout¹⁴² (uric acid, Fig. 4F) or general conditions such as fever¹⁴³ (cytokines, Fig. 4G). Nitrile glove-based system, with integrated electrode sensors¹⁴⁴ (Fig. 4H), provides in situ monitoring of sweat biomarkers including ethanol, Zn, pH, chloride, and vitamin C. The glove creates a local environment that is conducive to passive sweat induction and analysis across multiple biomarkers. To achieve a broad target specificity, a recent study¹⁴⁵ (Fig. 4I) uses flexible plasmonic metasurface designs with surface-enhanced Raman scattering (SERS), whereby the intensities of the biomarkers are measured via Raman spectrometer equipped microscope. Because the SERS spectrum is different across different biomarkers, the sensor showed robust target specificity compared with wearable electrochemical sensors. Another recent demonstration device¹⁴⁶ (Fig. 4J) circumvents the need for aggregate sweat collection or physical activity with a design that integrates hydrophilic wicking materials, an optimized microfluidic channel network, and electrochemical sensors to collect and analyze thermoregulatory sweat at a resting state. Supported by small pilot studies, this platform is capable of monitoring the onset of disease conditions (hypoglycemia) and variations in psychological factors (stress) through changes in sweat rate as well as the time-dynamic variations in concentration of drug therapeutics (Parkinson's) through electrochemical analysis. These and other recent examples^{93,138,143,145,168-173} highlight the powerful capabilities that wearable sensors offer for non-invasive clinical diagnostics and disease management.

Future Opportunities and Commercialization

Rapid manufacturing and process development of wearable sweat sensors has gained significant traction recently, due in part to the convergence of key advances in flexible

electronics, biochemical sensors, and materials science. The initial cohort of epidermal microfluidic sensors established an analytical pathway for obtaining personalized, real-time, continuous assessment of physiological parameters relevant for vastly expanding understanding of human health. The wearable sweat sensing platforms highlighted here represent key technological developments for realizing this significant potential. While these milestones suggest rapid maturation of this class of technology, a few key challenges remain before wide-spread adoption could be achieved.

Continued progress requires technological innovations with particular emphasis on scale up manufacturing and robustness. An important frontier of this research is in the integration of multimodal sensing platforms for monitoring biochemical and biophysical parameters in a continuous, long-term mode of operation. This necessitates consideration of sensor performance within a broad context of power management, wireless communication, and data acquisition of fully-integrated biochemical sensing systems. The recent emergence^{93,169,174-177} of biofuel cell-based self-powered wearable sensors represent a successful pathway to realizing such a fully-integrated platform.

The complex composition of sweat poses some of the most interesting challenges for wearable sweat sensors. In contrast to conventional laboratory-based analytical methods, these sensing platforms must operate in robust, stable manner under dynamic conditions and without the oversight of skilled technicians. Demonstrations of selective and multimodal sensors offer routes towards rapid, repeatable on-body measurements; however, certain constructs exhibit susceptibility to measurement errors caused by biofouling, varying ambient conditions (e.g. temperature or pH fluctuations), and motion artifacts. Although highly multiplexed sensors and nuanced device designs can mitigate such influences, development of new encapsulation materials and packaging strategies that protect against noise factors such as moisture and corrosion, could eliminate deterioration and sources of noise from non-specific binding or cross-talk, particularly for ultralow concentration species (e.g. DNA, RNA), is of critical importance.

Key to the widespread adoption of wearable sweat-sensors is the comprehensive validation of the systems. Although sweat offers enormous potential for noninvasive physiological monitoring, it has remained relatively unexplored in comparison to traditional biofluids such as blood. The emergence of novel physiologically-relevant sweat constituents, such as cortisol, lactate, and ethanol, is the direct result of the interest in non-invasive monitoring and rapid advances in the development of wearable sensing platforms. Continued progress requires extensive, large-scale, multi-center validation studies and formalized clinical trials. Such efforts could yield critical insights into the correlations with blood and urine analytes (and associated time-scales) requisite for establishing a comprehensive profile of sweat-based biochemical markers with physiological relevance. Moreover, such testing could, in turn, validate device performance beyond the research prototype stage of development.

Another important factor driving the demand for wearable sweat-sensor technologies is the development of multiparameter, long duration biochemical and biophysical sensing capabilities. One recent demonstration¹⁷⁸ achieves long-term sensing via on-demand iontophoretic stimulation at defined intervals (Fig. 5A) to monitor sweat biomarkers

with integrated electrochemical sensors. Another recent example¹⁷⁹ integrates a suite of sensor constructs within a single wearable platform to obtain multiparameter measurements of haemodynamic and metabolic biomarkers simultaneously throughout daily activities. Commercialization efforts around these multimodal systems tend to be costly⁷⁰, requiring novel manufacturing tooling and test strategies for large-scale production at high yield¹⁸⁰ (Fig. 5B). It is only within the last few years that the first commercial consumer wearable sweat sensors became widely available to consumers. Developed by Epicore Biosystems and The Gatorade Company and clinically validated in blinded studies¹⁸¹ (the G^x Sweat Patch, Fig. 5C), these microfluidic devices measure regional and whole-body sweat loss, sweat rate and electrolyte parameters, which are relevant to athletic performance and hydration. The G^x Sweat Patch employs colorimetric dyes and assays, along with real-time image processing via a smartphone application to compute results and actionable feedback in real time. Integration of this class of microfluidic technology with electronic modules enables continuous biochemical sensing and real-time alerts. These electronics-enabled epifluidic solutions rely on advances in energy storage, wireless communication, and memory storage as part of the full-integrated system. Large-scale clinical validation studies in sports and industrial safety are underway for the Connected Hydration System (Fig. 5D), and other representative examples of this technology.

ACKNOWLEDGMENT

We would like to acknowledge funding support provided by the Querrey Simpson Institute for Bioelectronics at Northwestern University. This publication was supported by the National Institute on Aging of the National Institutes of Health (R43AG067835). T.R.R. acknowledges additional support as a Junior Investigator from the National Institute of General Medical Sciences of the National Institutes of Health (P20GM113134) and the Hawai'i Community Foundation (Robert C. Perry Fund).

Funding Sources

R.G., J.A.R., and T.R.R. are inventors on patents and patent applications related to epidermal microfluidics. R.G., J.B.M., and J.A.R. are cofounders of Epicore Biosystems, a company that develops epidermal microfluidic devices. J.A.W. and D.E.W. are employees of Epicore Biosystems. T.R.R. has a consulting and advisory relationship with Epicore Biosystems.

REFERENCES

- (1). Djulbegovic B; Guyatt GH Progress in Evidence-Based Medicine: A Quarter Century On. *The Lancet* 2017, 390 (10092), 415–423. 10.1016/S0140-6736(16)31592-6.
- (2). Menendez ME; Jawa A; Haas DA; Warner JJP Orthopedic Surgery Post COVID-19: An Opportunity for Innovation and Transformation. *Journal of Shoulder and Elbow Surgery* 2020, 29 (6), 1083–1086. 10.1016/j.jse.2020.03.024. [PubMed: 32312643]
- (3). Fouad Y; Elwakil R; Elsahhar M; Said E; Bazeed S; Gomaa AA; Hashim A; Kamal E; Mehrez M; Attia D The NAFLD-MAFLD Debate: Eminence vs Evidence. *Liver International* 2021, 41 (2), 255–260. 10.1111/liv.14739. [PubMed: 33220154]
- (4). Mulimani PS Evidence-Based Practice and the Evidence Pyramid: A 21st Century Orthodontic Odyssey. *American Journal of Orthodontics and Dentofacial Orthopedics* 2017, 152 (1), 1–8. 10.1016/j.ajodo.2017.03.020. [PubMed: 28651753]
- (5). Thorborg K; Reiman MP; Weir A; Kemp JL; Serner A; Mosler AB; Hölmich P Clinical Examination, Diagnostic Imaging, and Testing of Athletes With Groin Pain: An Evidence-Based Approach to Effective Management. *J Orthop Sports Phys Ther* 2018, 48 (4), 239–249. 10.2519/jospt.2018.7850. [PubMed: 29510653]

- (6). Rakedzon S; Khoury Y; Rozenberg G; Neuberger A Hydroxychloroquine and Coronavirus Disease 2019: A Systematic Review of a Scientific Failure. *Rambam Maimonides Med J* 2020, 11 (3). 10.5041/RMMJ.10416.
- (7). Cohen MS Hydroxychloroquine for the Prevention of Covid-19 — Searching for Evidence. *New England Journal of Medicine* 2020, 383 (6), 585–586. 10.1056/NEJMe2020388. [PubMed: 32492298]
- (8). Jorge A Hydroxychloroquine in the Prevention of COVID-19 Mortality. *The Lancet Rheumatology* 2021, 3 (1), e2–e3. 10.1016/S2665-9913(20)30390-8. [PubMed: 33521667]
- (9). Ray TR; Choi J; Bandodkar AJ; Krishnan S; Gutruf P; Tian L; Ghaffari R; Rogers JA Bio-Integrated Wearable Systems: A Comprehensive Review. *Chem. Rev* 2019, 119 (8), 5461–5533. 10.1021/acs.chemrev.8b00573. [PubMed: 30689360]
- (10). Shrivastava S; Trung TQ; Lee N-E Recent Progress, Challenges, and Prospects of Fully Integrated Mobile and Wearable Point-of-Care Testing Systems for Self-Testing. *Chem. Soc. Rev* 2020, 49 (6), 1812–1866. 10.1039/C9CS00319C. [PubMed: 32100760]
- (11). Powers R; Etezadi-Amoli M; Arnold EM; Kianian S; Mance I; Gibiansky M; Trietsch D; Alvarado AS; Kretlow JD; Herrington TM; Brillman S; Huang N; Lin PT; Pham HA; Ullal AV Smartwatch Inertial Sensors Continuously Monitor Real-World Motor Fluctuations in Parkinson’s Disease. *Science Translational Medicine* 2021, 13 (579). 10.1126/scitranslmed.abd7865.
- (12). Heikenfeld J; Jajack A; Feldman B; Granger SW; Gaitonde S; Begtrup G; Katchman BA Accessing Analytes in Biofluids for Peripheral Biochemical Monitoring. *Nat Biotechnol* 2019, 37 (4), 407–419. 10.1038/s41587-019-0040-3. [PubMed: 30804536]
- (13). Sempionatto JR; Jeerapan I; Krishnan S; Wang J Wearable Chemical Sensors: Emerging Systems for On-Body Analytical Chemistry. *Anal. Chem* 2020, 92 (1), 378–396. 10.1021/acs.analchem.9b04668. [PubMed: 31626731]
- (14). Sempionatto JR; Jeerapan I; Krishnan S; Wang J Wearable Chemical Sensors: Emerging Systems for On-Body Analytical Chemistry. *Anal. Chem* 2020, 92 (1), 378–396. 10.1021/acs.analchem.9b04668. [PubMed: 31626731]
- (15). Choi J; Ghaffari R; Baker LB; Rogers JA Skin-Interfaced Systems for Sweat Collection and Analytics. *Sci. Adv* 2018, 4 (2), eaar3921. 10.1126/sciadv.aar3921. [PubMed: 29487915]
- (16). Katchman BA; Zhu M; Blain Christen J; Anderson KS Eccrine Sweat as a Biofluid for Profiling Immune Biomarkers. *Prot. Clin. Appl* 2018, 12 (6), 1800010. 10.1002/prca.201800010.
- (17). Lin S; Yu W; Wang B; Zhao Y; En K; Zhu J; Cheng X; Zhou C; Lin H; Wang Z; Hojaiji H; Yeung C; Milla C; Davis RW; Emaminejad S Noninvasive Wearable Electroactive Pharmaceutical Monitoring for Personalized Therapeutics. *Proc Natl Acad Sci USA* 2020, 117 (32), 19017–19025. 10.1073/pnas.2009979117. [PubMed: 32719130]
- (18). Mondal S; Zehra N; Choudhury A; Iyer PK Wearable Sensing Devices for Point of Care Diagnostics. *ACS Appl. Bio Mater* 2020, aacsabm.0c00798. 10.1021/acsabm.0c00798.
- (19). Samson C; Koh A Stress Monitoring and Recent Advancements in Wearable Biosensors. *Front. Bioeng. Biotechnol* 2020, 8, 1037. 10.3389/fbioe.2020.01037. [PubMed: 32984293]
- (20). Brasier N; Eckstein J Sweat as a Source of Next-Generation Digital Biomarkers. *Digit Biomark* 2019, 3 (3), 155–165. 10.1159/000504387. [PubMed: 32095774]
- (21). Yang Y; Gao W Wearable and Flexible Electronics for Continuous Molecular Monitoring. *Chem. Soc. Rev* 2019, 48 (6), 1465–1491. 10.1039/C7CS00730B. [PubMed: 29611861]
- (22). Ghaffari R; Choi J; Raj MS; Chen S; Lee SP; Reeder JT; Aranyosi AJ; Leech A; Li W; Schon S; Model JB; Rogers JA Soft Wearable Systems for Colorimetric and Electrochemical Analysis of Biofluids. *Adv. Funct. Mater* 2019, 1907269. 10.1002/adfm.201907269.
- (23). Bandodkar AJ; Jeang WJ; Ghaffari R; Rogers JA Wearable Sensors for Biochemical Sweat Analysis. *Annual Rev. Anal. Chem* 2019, 12 (1), 1–22. 10.1146/annurev-anchem-061318-114910.
- (24). Brothers MC; DeBrosse M; Grigsby CC; Naik RR; Hussain SM; Heikenfeld J; Kim SS Achievements and Challenges for Real-Time Sensing of Analytes in Sweat within Wearable Platforms. *Acc. Chem. Res* 2019, 52 (2), 297–306. 10.1021/acs.accounts.8b00555. [PubMed: 30688433]

- (25). Wang R; Wang X Sensing of Inorganic Ions in Microfluidic Devices. *Sensors and Actuators B: Chemical* 2021, 329, 129171. 10.1016/j.snb.2020.129171.
- (26). Upasham S; Churcher NKM; Rice P; Prasad S Sweating Out the Circadian Rhythm: A Technical Review. *ACS Sens.* 2021, 6 (3), 659–672. 10.1021/acssensors.0c02622. [PubMed: 33645964]
- (27). Song Y; Mukasa D; Zhang H; Gao W Self-Powered Wearable Biosensors. *Acc. Mater. Res* 2021, 2 (3), 184–197. 10.1021/accountsmr.1c00002.
- (28). Sharma A; Badea M; Tiwari S; Marty JL Wearable Biosensors: An Alternative and Practical Approach in Healthcare and Disease Monitoring. *Molecules* 2021, 26 (3), 748. 10.3390/molecules26030748. [PubMed: 33535493]
- (29). Park M; Heo YJ Biosensing Technologies for Chronic Diseases. *BioChip J* 2021, 15 (1), 1–13. 10.1007/s13206-021-00014-3.
- (30). Min J; Sempionatto JR; Teymourian H; Wang J; Gao W Wearable Electrochemical Biosensors in North America. *Biosensors and Bioelectronics* 2021, 172, 112750. 10.1016/j.bios.2020.112750. [PubMed: 33129072]
- (31). Mercuri M; Fernandez Rivas D Challenges and Opportunities for Small Volumes Delivery into the Skin. *Biomicrofluidics* 2021, 15 (1), 011301. 10.1063/5.0030163. [PubMed: 33532017]
- (32). Hernández-Rodríguez JF; Rojas D; Escarpa A Electrochemical Sensing Directions for Next-Generation Healthcare: Trends, Challenges, and Frontiers. *Anal. Chem* 2021, 93 (1), 167–183. 10.1021/acs.analchem.0c04378. [PubMed: 33174738]
- (33). Bhide A; Ganguly A; Parupudi T; Ramasamy M; Muthukumar S; Prasad S Next-Generation Continuous Metabolite Sensing toward Emerging Sensor Needs. *ACS Omega* 2021, 6 (9), 6031–6040. 10.1021/acsomega.0c06209. [PubMed: 33718694]
- (34). Ates HC; Brunauer A; Stetten F; Urban GA; Güder F; Merkoçi A; Früh SM; Dincer C Integrated Devices for Non-Invasive Diagnostics. *Adv. Funct. Mater* 2021, 31 (15), 2010388. 10.1002/adfm.202010388.
- (35). Baldo TA; de Lima LF; Mendes LF; de Araujo WR; Paixão TRLC; Coltro WKT Wearable and Biodegradable Sensors for Clinical and Environmental Applications. *ACS Appl. Electron. Mater* 2021, 3 (1), 68–100. 10.1021/acsaelm.0c00735.
- (36). Zanfognini B; Pigani L; Zanardi C Recent Advances in the Direct Electrochemical Detection of Drugs of Abuse. *J Solid State Electrochem* 2020, 24 (11–12), 2603–2616. 10.1007/s10008-020-04686-z.
- (37). Ye S; Feng S; Huang L; Bian S Recent Progress in Wearable Biosensors: From Healthcare Monitoring to Sports Analytics. *Biosensors* 2020, 10 (12), 205. 10.3390/bios10120205.
- (38). Yang Y; Chen Y; Tang H; Zong N; Jiang X Microfluidics for Biomedical Analysis. *Small Methods* 2020, 4 (4), 1900451. 10.1002/smt.201900451.
- (39). Xu C; Yang Y; Gao W Skin-Interfaced Sensors in Digital Medicine: From Materials to Applications. *Matter* 2020, 2 (6), 1414–1445. 10.1016/j.matt.2020.03.020. [PubMed: 32510052]
- (40). Wen F; He T; Liu H; Chen H-Y; Zhang T; Lee C Advances in Chemical Sensing Technology for Enabling the Next-Generation Self-Sustainable Integrated Wearable System in the IoT Era. *Nano Energy* 2020, 78, 105155. 10.1016/j.nanoen.2020.105155.
- (41). Shao Y; Ying Y; Ping J Recent Advances in Solid-Contact Ion-Selective Electrodes: Functional Materials, Transduction Mechanisms, and Development Trends. *Chem. Soc. Rev* 2020, 49 (13), 4405–4465. 10.1039/C9CS00587K. [PubMed: 32458836]
- (42). Narayanamurthy V; Jeroish ZE; Bhuvaneshwari KS; Bayat P; Premkumar R; Samsuri F; Yusoff MM Advances in Passively Driven Microfluidics and Lab-on-Chip Devices: A Comprehensive Literature Review and Patent Analysis. *RSC Adv.* 2020, 10 (20), 11652–11680. 10.1039/D0RA00263A. [PubMed: 35496619]
- (43). Qiao L; Benzigar MR; Subramony JA; Lovell NH; Liu G Advances in Sweat Wearables: Sample Extraction, Real-Time Biosensing, and Flexible Platforms. *ACS Appl. Mater. Interfaces* 2020, 12 (30), 34337–34361. 10.1021/acsaami.0c07614. [PubMed: 32579332]
- (44). Moonen EJM; Haakma JR; Peri E; Pelssers E; Mischi M; den Toonder MJM Wearable Sweat Sensing for Prolonged, Semicontinuous, and Nonobtrusive Health Monitoring. *View* 2020, 1 (4), 20200077. 10.1002/VIW.20200077.

- (45). Mohan AMV; Rajendran V; Mishra RK; Jayaraman M Recent Advances and Perspectives in Sweat Based Wearable Electrochemical Sensors. *TrAC Trends in Analytical Chemistry* 2020, 131, 116024. 10.1016/j.trac.2020.116024.
- (46). Liu C; Xu T; Wang D; Zhang X The Role of Sampling in Wearable Sweat Sensors. *Talanta* 2020, 212, 120801. 10.1016/j.talanta.2020.120801. [PubMed: 32113563]
- (47). Ling Y; An T; Yap LW; Zhu B; Gong S; Cheng W Disruptive, Soft, Wearable Sensors. *Adv. Mater* 2020, 32 (18), 1904664. 10.1002/adma.201904664.
- (48). Fan R; Andrew TL Perspective—Challenges in Developing Wearable Electrochemical Sensors for Longitudinal Health Monitoring. *J. Electrochem. Soc* 2020, 167 (3), 037542. 10.1149/1945-7111/ab67b0.
- (49). Falk M; Psotta C; Cirovic S; Shleev S Non-Invasive Electrochemical Biosensors Operating in Human Physiological Fluids. *Sensors* 2020, 20 (21), 6352. 10.3390/s20216352.
- (50). Arroyo-Currás N; Dauphin-Ducharme P; Scida K; Chávez JL From the Beaker to the Body: Translational Challenges for Electrochemical, Aptamer-Based Sensors. *Anal. Methods* 2020, 12 (10), 1288–1310. 10.1039/D0AY00026D.
- (51). Dervisevic M; Alba M; Prieto-Simon B; Voelcker NH Skin in the Diagnostics Game: Wearable Biosensor Nano- and Microsystems for Medical Diagnostics. *Nano Today* 2020, 30, 100828. 10.1016/j.nantod.2019.100828.
- (52). Seshadri DR; Li RT; Voos JE; Rowbottom JR; Alfes CM; Zorman CA; Drummond CK Wearable Sensors for Monitoring the Physiological and Biochemical Profile of the Athlete. *npj Digital Medicine* 2019, 2 (1), 1–16. 10.1038/s41746-019-0150-9. [PubMed: 31304351]
- (53). Mahmud MS; Fang H; Carreiro S; Wang H; Boyer EW Wearables Technology for Drug Abuse Detection: A Survey of Recent Advancement. *Smart Health* 2019, 13, 100062. 10.1016/j.smhl.2018.09.002.
- (54). Chung M; Fortunato G; Radacsi N Wearable Flexible Sweat Sensors for Healthcare Monitoring: A Review. *J. R. Soc. Interface* 2019, 16 (159), 20190217. 10.1098/rsif.2019.0217. [PubMed: 31594525]
- (55). Zhao J; Guo H; Li J; Bandodkar AJ; Rogers JA Body-Interfaced Chemical Sensors for Noninvasive Monitoring and Analysis of Biofluids. *Trends in Chemistry* 2019, 1 (6), 559–571. 10.1016/j.trechm.2019.07.001.
- (56). Harris J; Bickford J; Cho P; Coppock M; Farrell ME; Holthoff EL; Ratcliff E Approaching Single Molecule Sensing: Predictive Sweat Sensor Design for Ultra-Low Limits of Detection. In *Chemical, Biological, Radiological, Nuclear, and Explosives (CBRNE) Sensing XX*; Guicheteau JA, Howle CR, Eds.; SPIE: Baltimore, United States, 2019; p 15. 10.1117/12.2518543.
- (57). Klimuntowski M; Alam MM; Singh G; Howlader MMR Electrochemical Sensing of Cannabinoids in Biofluids: A Noninvasive Tool for Drug Detection. *ACS Sens.* 2020, 5 (3), 620–636. 10.1021/acssensors.9b02390. [PubMed: 32102542]
- (58). Manjakkal L; Dervin S; Dahiya R Flexible Potentiometric PH Sensors for Wearable Systems. *RSC Adv.* 2020, 10 (15), 8594–8617. 10.1039/D0RA00016G. [PubMed: 35496561]
- (59). Martín Vázquez PE; Brunel F; Raimundo J-M Recent Electrochemical/Electrical Microfabricated Sensor Devices for Ionic and Polyionic Analytes. *ACS Omega* 2020, 5 (10), 4733–4742. 10.1021/acsomega.9b04331. [PubMed: 32201758]
- (60). Lyu Y; Gan S; Bao Y; Zhong L; Xu J; Wang W; Liu Z; Ma Y; Yang G; Niu L Solid-Contact Ion-Selective Electrodes: Response Mechanisms, Transducer Materials and Wearable Sensors. *Membranes* 2020, 10 (6), 128. 10.3390/membranes10060128.
- (61). Teymourian H; Parrilla M; Sempionatto JR; Montiel NF; Barfidokht A; Van Echelpoel R; De Wael K; Wang J Wearable Electrochemical Sensors for the Monitoring and Screening of Drugs. *ACS Sens.* 2020, 5 (9), 2679–2700. 10.1021/acssensors.0c01318. [PubMed: 32822166]
- (62). Wang Z; Shin J; Park J; Lee H; Kim D; Liu H Engineering Materials for Electrochemical Sweat Sensing. *Adv. Funct. Mater* 2021, 31 (12), 2008130. 10.1002/adfm.202008130.
- (63). Yun SM; Kim M; Kwon YW; Kim H; Kim MJ; Park Y-G; Park J-U Recent Advances in Wearable Devices for Non-Invasive Sensing. *Applied Sciences* 2021, 11 (3), 1235. 10.3390/app11031235.

- (64). Li S; Ma Z; Cao Z; Pan L; Shi Y Advanced Wearable Microfluidic Sensors for Healthcare Monitoring. *Small* 2020, 16 (9), 1903822. 10.1002/sml.201903822.
- (65). Ray T; Choi J; Reeder J; Lee SP; Aranyosi AJ; Ghaffari R; Rogers JA Soft, Skin-Interfaced Wearable Systems for Sports Science and Analytics. *Current Opinion in Biomedical Engineering* 2019, 9, 47–56. 10.1016/j.cobme.2019.01.003.
- (66). Ferrari LM; Keller K; Burtcher B; Greco F Temporary Tattoo as Unconventional Substrate for Conformable and Transferable Electronics on Skin and Beyond. *Multifunct. Mater* 2020, 3 (3), 032003. 10.1088/2399-7532/aba6e3.
- (67). Sreenilayam SP; Ahad IU; Nicolosi V; Acinas Garzon V; Brabazon D Advanced Materials of Printed Wearables for Physiological Parameter Monitoring. *Materials Today* 2020, 32, 147–177. 10.1016/j.mattod.2019.08.005.
- (68). Lin J; Zhu Z; Cheung CF; Yan F; Li G Digital Manufacturing of Functional Materials for Wearable Electronics. *J. Mater. Chem. C* 2020, 8 (31), 10587–10603. 10.1039/D0TC01112F.
- (69). Choi J; Xue Y; Xia W; Ray TR; Reeder JT; Bandodkar AJ; Kang D; Xu S; Huang Y; Rogers JA Soft, Skin-Mounted Microfluidic Systems for Measuring Secretory Fluidic Pressures Generated at the Surface of the Skin by Eccrine Sweat Glands. *Lab Chip* 2017, 17 (15), 2572–2580. 10.1039/C7LC00525C. [PubMed: 28664954]
- (70). Ghaffari R; Rogers JA; Ray TR Recent Progress, Challenges, and Opportunities for Wearable Biochemical Sensors for Sweat Analysis. *Sensors and Actuators B: Chemical* 2021, 332, 129447. 10.1016/j.snb.2021.129447. [PubMed: 33542590]
- (71). Koh A; Kang D; Xue Y; Lee S; Pielak RM; Kim J; Hwang T; Min S; Banks A; Bastien P; Manco MC; Wang L; Ammann KR; Jang K-I; Won P; Han S; Ghaffari R; Paik U; Slepian MJ; Balooch G; Huang Y; Rogers JA A Soft, Wearable Microfluidic Device for the Capture, Storage, and Colorimetric Sensing of Sweat. *Science Translational Medicine* 2016, 8 (366), 366ra165–366ra165. 10.1126/scitranslmed.aaf2593.
- (72). Kim SD; Koo Y; Yun Y A Smartphone-Based Automatic Measurement Method for Colorimetric PH Detection Using a Color Adaptation Algorithm. *Sensors* 2017, 17 (7), 1604. 10.3390/s17071604.
- (73). A. Piriya VS; Joseph P; K. Daniel SCG; Lakshmanan S; Kinoshita T; Muthusamy S Colorimetric Sensors for Rapid Detection of Various Analytes. *Materials Science and Engineering: C* 2017, 78, 1231–1245. 10.1016/j.msec.2017.05.018. [PubMed: 28575962]
- (74). Kim SB; Zhang Y; Won SM; Bandodkar AJ; Sekine Y; Xue Y; Koo J; Harshman SW; Martin JA; Park JM; Ray TR; Crawford KE; Lee K-T; Choi J; Pitsch RL; Grigsby CC; Strang AJ; Chen Y-Y; Xu S; Kim J; Koh A; Ha JS; Huang Y; Kim SW; Rogers JA Super-Absorbent Polymer Valves and Colorimetric Chemistries for Time-Sequenced Discrete Sampling and Chloride Analysis of Sweat via Skin-Mounted Soft Microfluidics. *Small* 2018, 14 (12), 1703334. 10.1002/sml.201703334.
- (75). Choi J; Bandodkar AJ; Reeder JT; Ray TR; Turnquist A; Kim SB; Nyberg N; Hourlier-Fargette A; Model JB; Aranyosi AJ; Xu S; Ghaffari R; Rogers JA Soft, Skin-Integrated Multifunctional Microfluidic Systems for Accurate Colorimetric Analysis of Sweat Biomarkers and Temperature. *ACS Sens.* 2019, 4 (2), 379–388. 10.1021/acssensors.8b01218. [PubMed: 30707572]
- (76). Jain V; Ochoa M; Jiang H; Rahimi R; Ziaie B A Mass-Customizable Dermal Patch with Discrete Colorimetric Indicators for Personalized Sweat Rate Quantification. *Microsyst Nanoeng* 2019, 5 (1), 29. 10.1038/s41378-019-0067-0. [PubMed: 31240108]
- (77). Xiao J; Liu Y; Su L; Zhao D; Zhao L; Zhang X Microfluidic Chip-Based Wearable Colorimetric Sensor for Simple and Facile Detection of Sweat Glucose. *Anal. Chem* 2019, 91 (23), 14803–14807. 10.1021/acs.analchem.9b03110. [PubMed: 31553565]
- (78). Tu E; Pearlmutter P; Tiangco M; Deroose G; Begdache L; Koh A Comparison of Colorimetric Analyses to Determine Cortisol in Human Sweat. *ACS Omega* 2020, 5 (14), 8211–8218. 10.1021/acsomega.0c00498. [PubMed: 32309731]
- (79). Vaquer A; Barón E; de la Rica R Wearable Analytical Platform with Enzyme-Modulated Dynamic Range for the Simultaneous Colorimetric Detection of Sweat Volume and Sweat Biomarkers. *ACS Sens.* 2021, 6 (1), 130–136. 10.1021/acssensors.0c01980. [PubMed: 33371672]

- (80). Zhang K; Zhang J; Wang F; Kong D Stretchable and Superwetable Colorimetric Sensing Patch for Epidermal Collection and Analysis of Sweat. *ACS Sens.* 2021, 6 (6), 2261–2269. 10.1021/acssensors.1c00316. [PubMed: 34048231]
- (81). Sekine Y; Kim SB; Zhang Y; Bandodkar AJ; Xu S; Choi J; Irie M; Ray TR; Kohli P; Kozai N; Sugita T; Wu Y; Lee K; Lee K-T; Ghaffari R; Rogers JA A Fluorometric Skin-Interfaced Microfluidic Device and Smartphone Imaging Module for *in Situ* Quantitative Analysis of Sweat Chemistry. *Lab Chip* 2018, 18 (15), 2178–2186. 10.1039/C8LC00530C. [PubMed: 29955754]
- (82). Ardalan S; Hosseini M; Vosough M; Golmohammadi H Towards Smart Personalized Perspiration Analysis: An IoT-Integrated Cellulose-Based Microfluidic Wearable Patch for Smartphone Fluorimetric Multi-Sensing of Sweat Biomarkers. *Biosensors and Bioelectronics* 2020, 168, 112450. 10.1016/j.bios.2020.112450. [PubMed: 32877780]
- (83). Cui Y; Duan W; Jin Y; Wo F; Xi F; Wu J Ratiometric Fluorescent Nanohybrid for Noninvasive and Visual Monitoring of Sweat Glucose. *ACS Sens.* 2020, 5 (7), 2096–2105. 10.1021/acssensors.0c00718. [PubMed: 32450686]
- (84). Kim SB; Lee B; Reeder JT; Seo SH; Lee S-U; Shin J; Sekine Y; Jeong H; Oh YS; Aranyosi J; Lee SP; Model JB; Lee G; Seo M-H; Soo S; Jo S; Park G; Han S; Park I; Jung H-I; Koo J; Braun PV; Rogers JA Soft, Skin-Interfaced Sweat Microfluidic Systems with Integrated Lateral Flow 2 Immunoassays, Fluorometric Sensors and Impedance-Based Measurement Capabilities. 26.
- (85). Teymourian H; Parrilla M; Sempionatto JR; Montiel NF; Barfidokht A; Van Echelpoel R; De Wael K; Wang J Wearable Electrochemical Sensors for the Monitoring and Screening of Drugs. *ACS Sens.* 2020, 5 (9), 2679–2700. 10.1021/acssensors.0c01318. [PubMed: 32822166]
- (86). Reeder JT; Choi J; Xue Y; Gutruf P; Hanson J; Liu M; Ray T; Bandodkar AJ; Avila R; Xia W; Krishnan S; Xu S; Barnes K; Pahnke M; Ghaffari R; Huang Y; Rogers JA Waterproof, Electronics-Enabled, Epidermal Microfluidic Devices for Sweat Collection, Biomarker Analysis, and Thermography in Aquatic Settings. *Sci. Adv* 2019, 5 (1), eaau6356. 10.1126/sciadv.aau6356. [PubMed: 30746456]
- (87). Kim J; Sempionatto JR; Imani S; Hartel MC; Barfidokht A; Tang G; Campbell AS; Mercier PP; Wang J Simultaneous Monitoring of Sweat and Interstitial Fluid Using a Single Wearable Biosensor Platform. *Adv. Sci* 2018, 5 (10), 1800880. 10.1002/advs.201800880.
- (88). Zhang Y; Guo H; Kim SB; Wu Y; Ostojich D; Park SH; Wang X; Weng Z; Li R; Bandodkar AJ; Sekine Y; Choi J; Xu S; Quaggin S; Ghaffari R; Rogers JA Passive Sweat Collection and Colorimetric Analysis of Biomarkers Relevant to Kidney Disorders Using a Soft Microfluidic System. *Lab Chip* 2019, 19 (9), 1545–1555. 10.1039/C9LC00103D. [PubMed: 30912557]
- (89). Saha T; Fang J; Mukherjee S; Dickey MD; Velez OD Wearable Osmotic-Capillary Patch for Prolonged Sweat Harvesting and Sensing. *ACS Appl. Mater. Interfaces* 2021, 13 (7), 8071–8081. 10.1021/acsaami.0c22730. [PubMed: 33587589]
- (90). Choi J; Kang D; Han S; Kim SB; Rogers JA Microfluidic Networks: Thin, Soft, Skin-Mounted Microfluidic Networks with Capillary Bursting Valves for Chrono-Sampling of Sweat (*Adv. Healthcare Mater.* 5/2017). *Adv. Healthcare Mater* 2017, 6 (5). 10.1002/adhm.201770023.
- (91). Lin H; Tan J; Zhu J; Lin S; Zhao Y; Yu W; Hojaiji H; Wang B; Yang S; Cheng X; Wang Z; Tang E; Yeung C; Emaminejad S A Programmable Epidermal Microfluidic Valving System for Wearable Biofluid Management and Contextual Biomarker Analysis. *Nat Commun* 2020, 11 (1), 4405. 10.1038/s41467-020-18238-6. [PubMed: 32879320]
- (92). Bandodkar AJ; Choi J; Lee SP; Jeang WJ; Agyare P; Gutruf P; Wang S; Sponenburger RA; Reeder JT; Schon S; Ray TR; Chen S; Mehta S; Ruiz S; Rogers JA Soft, Skin-Interfaced Microfluidic Systems with Passive Galvanic Stopwatches for Precise Chronometric Sampling of Sweat. *Adv. Mater* 2019, 31 (32), 1902109. 10.1002/adma.201902109.
- (93). Yu Y; Nassar J; Xu C; Min J; Yang Y; Dai A; Doshi R; Huang A; Song Y; Gehlhar R; Ames AD; Gao W Biofuel-Powered Soft Electronic Skin with Multiplexed and Wireless Sensing for Human-Machine Interfaces. *Sci. Robot* 2020, 5 (41), eaaz7946. 10.1126/scirobotics.aaz7946. [PubMed: 32607455]
- (94). Choi J; Chen S; Deng Y; Xue Y; Reeder JT; Franklin D; Oh YS; Model JB; Aranyosi AJ; Lee SP; Ghaffari R; Huang Y; Rogers JA Skin-Interfaced Microfluidic Systems That Combine Hard and Soft Materials for Demanding Applications in Sweat Capture and Analysis. *Adv. Healthcare Mater* 2020, 2000722. 10.1002/adhm.202000722.

- (95). Aranyosi AJ; Model JB; Zhang MZ; Lee SP; Leech A; Li W; Seib MS; Chen S; Reny N; Wallace J; Shin MH; Bandodkar AJ; Choi J; Paller AS; Rogers JA; Xu S; Ghaffari R Rapid Capture and Extraction of Sweat for Regional Rate and Cytokine Composition Analysis Using a Wearable Soft Microfluidic System. *Journal of Investigative Dermatology* 2020, S0022202X20316857. 10.1016/j.jid.2020.05.107.
- (96). Gorvoy JD; Acs H; Stein ML The Hazard of Induction of Sweating in Cystic Fibrosis of the Pancreas. *Pediatrics* 1960, 25 (6), 977–982. [PubMed: 13851354]
- (97). Liu C; Xu T; Wang D; Zhang X The Role of Sampling in Wearable Sweat Sensors. *Talanta* 2020, 212, 120801. 10.1016/j.talanta.2020.120801. [PubMed: 32113563]
- (98). Hussain JN; Mantri N; Cohen MM Working Up a Good Sweat – The Challenges of Standardising Sweat Collection for Metabolomics Analysis. 22.
- (99). LeGrys VA; Moon TC; Laux J; Accurso F; Martiniano SA A Multicenter Evaluation of Sweat Chloride Concentration and Variation in Infants with Cystic Fibrosis. *J Cyst Fibros* 2019, 18 (2), 190–193. 10.1016/j.jcf.2018.12.006. [PubMed: 30583934]
- (100). Baker LB; Wolfe AS Physiological Mechanisms Determining Eccrine Sweat Composition. *Eur J Appl Physiol* 2020, 120 (4), 719–752. 10.1007/s00421-020-04323-7. [PubMed: 32124007]
- (101). Baker LB; De Chavez PJD; Ungaro CT; Sopena BC; Nuccio RP; Reimel AJ; Barnes KA Exercise Intensity Effects on Total Sweat Electrolyte Losses and Regional vs. Whole-Body Sweat [Na+], [Cl-], and [K+]. *European Journal of Applied Physiology* 2019, 119 (2), 361–375. 10.1007/s00421-018-4048-z. [PubMed: 30523403]
- (102). Baker LB Physiology of Sweat Gland Function: The Roles of Sweating and Sweat Composition in Human Health. *Temperature (Austin)* 2019, 6 (3), 211–259. 10.1080/23328940.2019.1632145. [PubMed: 31608304]
- (103). Baker LB; Ungaro CT; Sopena BC; Nuccio RP; Reimel AJ; Carter JM; Stofan JR; Barnes KA Body Map of Regional vs. Whole Body Sweating Rate and Sweat Electrolyte Concentrations in Men and Women during Moderate Exercise-Heat Stress. *Journal of Applied Physiology* 2018, 124 (5), 1304–1318. 10.1152/jappphysiol.00867.2017. [PubMed: 29420145]
- (104). Goulet EDB; Asselin A; Gosselin J; Baker LB Measurement of Sodium Concentration in Sweat Samples: Comparison of Five Analytical Techniques. *Appl Physiol Nutr Metab.* 2017, 42 (8), 861–868. 10.1139/apnm-2017-0059. [PubMed: 28407476]
- (105). Souza SL; Graça G; Oliva A Characterization of Sweat Induced with Pilocarpine, Physical Exercise, and Collected Passively by Metabolomic Analysis. *Skin Res Technol* 2018, 24 (2), 187–195. 10.1111/srt.12412. [PubMed: 29131416]
- (106). Klous L; De Ruyter C; Alkemade P; Daanen H; Gerrett N Sweat Rate and Sweat Composition during Heat Acclimation. *Journal of Thermal Biology* 2020, 93, 102697. 10.1016/j.jtherbio.2020.102697. [PubMed: 33077118]
- (107). Steijlen ASM; Bastemeijer J; Groen P; Jansen KMB; French PJ; Bossche A A Wearable Fluidic Collection Patch and Ion Chromatography Method for Sweat Electrolyte Monitoring during Exercise. *Anal. Methods* 2020, 12 (48), 5885–5892. 10.1039/D0AY02014A. [PubMed: 33290448]
- (108). Shay T; Saha T; Dickey MD; Velev OD Principles of Long-Term Fluids Handling in Paper-Based Wearables with Capillary–Evaporative Transport. *Biomicrofluidics* 2020, 14 (3), 034112. 10.1063/5.0010417. [PubMed: 32566070]
- (109). Yokus MA; Saha T; Fang J; Dickey MD; Velev OD; Daniele MA Towards Wearable Electrochemical Lactate Sensing Using Osmotic-Capillary Microfluidic Pumping. In *2019 IEEE SENSORS*; IEEE: Montreal, QC, Canada, 2019; pp 1–4. 10.1109/SENSORS43011.2019.8956651.
- (110). Zhao FJ; Bonmarin M; Chen ZC; Larson M; Fay D; Runnoe D; Heikenfeld J Ultra-Simple Wearable Local Sweat Volume Monitoring Patch Based on Swellable Hydrogels. *Lab Chip* 2020, 20 (1), 168–174. 10.1039/C9LC00911F. [PubMed: 31796944]
- (111). He X; Yang S; Pei Q; Song Y; Liu C; Xu T; Zhang X Integrated Smart Janus Textile Bands for Self-Pumping Sweat Sampling and Analysis. *ACS Sens.* 2020, 5 (6), 1548–1554. 10.1021/acssensors.0c00563. [PubMed: 32466645]

- (112). Zhang Y; Chen Y; Huang J; Liu Y; Peng J; Chen S; Song K; Ouyang X; Cheng H; Wang X Skin-Interfaced Microfluidic Devices with One-Opening Chambers and Hydrophobic Valves for Sweat Collection and Analysis. *Lab Chip* 2020, 20 (15), 2635–2645. 10.1039/D0LC00400F. [PubMed: 32555915]
- (113). Choi J; Kang D; Han S; Kim SB; Rogers JA Thin, Soft, Skin-Mounted Microfluidic Networks with Capillary Bursting Valves for Chrono-Sampling of Sweat. *Adv. Healthcare Mater* 2017, 6 (5), 1601355. 10.1002/adhm.201601355.
- (114). Schmidt-Speicher LM; Länge K Microfluidic Integration for Electrochemical Biosensor Applications. *Current Opinion in Electrochemistry* 2021, 100755. 10.1016/j.coelec.2021.100755.
- (115). Lin H; Hojaiji H; Lin S; Yeung C; Zhao Y; Wang B; Malige M; Wang Y; King K; Yu W; Tan J; Wang Z; Cheng X; Emaminejad S A Wearable Electrofluidic Actuation System. *Lab Chip* 2019, 19 (18), 2966–2972. 10.1039/C9LC00454H. [PubMed: 31397462]
- (116). Bandodkar AJ; Gutruf P; Choi J; Lee K; Sekine Y; Reeder JT; Jeang WJ; Aranyosi AJ; Lee SP; Model JB; Ghaffari R; Su C-J; Leshock JP; Ray T; Verrillo A; Thomas K; Krishnamurthi V; Han S; Kim J; Krishnan S; Hang T; Rogers JA Battery-Free, Skin-Interfaced Microfluidic/Electronic Systems for Simultaneous Electrochemical, Colorimetric, and Volumetric Analysis of Sweat. *Sci. Adv* 2019, 5 (1), eaav3294. 10.1126/sciadv.aav3294. [PubMed: 30746477]
- (117). Kim SB; Koo J; Yoon J; Hourlier-Fargette A; Lee B; Chen S; Jo S; Choi J; Oh YS; Lee G; Won SM; Aranyosi AJ; Lee SP; Model JB; Braun PV; Ghaffari R; Park C; Rogers JA Soft, Skin-Interfaced Microfluidic Systems with Integrated Enzymatic Assays for Measuring the Concentration of Ammonia and Ethanol in Sweat. *Lab Chip* 2020, 20 (1), 84–92. 10.1039/C9LC01045A. [PubMed: 31776526]
- (118). Song Y; Min J; Yu Y; Wang H; Yang Y; Zhang H; Gao W Wireless Battery-Free Wearable Sweat Sensor Powered by Human Motion. *Sci. Adv* 2020, 6 (40), eaay9842. 10.1126/sciadv.aay9842. [PubMed: 32998888]
- (119). Bandodkar AJ; Lee SP; Huang I; Li W; Wang S; Su C-J; Jeang WJ; Hang T; Mehta S; Nyberg N; Gutruf P; Choi J; Koo J; Reeder JT; Tseng R; Ghaffari R; Rogers JA Sweat-Activated Biocompatible Batteries for Epidermal Electronic and Microfluidic Systems. *Nat Electron* 2020, 3 (9), 554–562. 10.1038/s41928-020-0443-7.
- (120). Lu Y; Jiang K; Chen D; Shen G Wearable Sweat Monitoring System with Integrated Micro-Supercapacitors. *Nano Energy* 2019, 58, 624–632. 10.1016/j.nanoen.2019.01.084.
- (121). Kaya T; Liu G; Ho J; Yelamarthi K; Miller K; Edwards J; Stannard A Wearable Sweat Sensors: Background and Current Trends. *Electroanalysis* 2019, 31 (3), 411–421. 10.1002/elan.201800677.
- (122). McCaul M; Glennon T; Diamond D Challenges and Opportunities in Wearable Technology for Biochemical Analysis in Sweat. *Current Opinion in Electrochemistry* 2017, 3 (1), 46–50. 10.1016/j.coelec.2017.06.001.
- (123). Qiao L; Benziger MR; Subramony JA; Lovell NH; Liu G Advances in Sweat Wearables: Sample Extraction, Real-Time Biosensing, and Flexible Platforms. *ACS Appl. Mater. Interfaces* 2020, 12 (30), 34337–34361. 10.1021/acsami.0c07614. [PubMed: 32579332]
- (124). Pal A; Nadiger VG; Goswami D; Martinez RV Conformal, Waterproof Electronic Decals for Wireless Monitoring of Sweat and Vaginal PH at the Point-of-Care. *Biosensors and Bioelectronics* 2020, 160, 112206. 10.1016/j.bios.2020.112206. [PubMed: 32339147]
- (125). Wiorek A; Parrilla M; Cuartero M; Crespo GA Epidermal Patch with Glucose Biosensor: PH and Temperature Correction toward More Accurate Sweat Analysis during Sport Practice. *Anal. Chem* 2020, 92 (14), 10153–10161. 10.1021/acs.analchem.0c02211. [PubMed: 32588617]
- (126). Baker LB; Model JB; Barnes KA; Anderson ML; Lee SP; Lee KA; Brown SD; Reimel AJ; Roberts TJ; Nuccio RP; Bonsignore JL; Ungaro CT; Carter JM; Li W; Seib MS; Reeder JT; Aranyosi AJ; Rogers JA; Ghaffari R Skin-Interfaced Microfluidic System with Personalized Sweating Rate and Sweat Chloride Analytics for Sports Science Applications. *Sci. Adv* 2020, 6 (50), eaabe3929. 10.1126/sciadv.abe3929. [PubMed: 33310859]
- (127). Francis J; Stamper I; Heikenfeld J; Gomez EF Digital Nanoliter to Milliliter Flow Rate Sensor with *in Vivo* Demonstration for Continuous Sweat Rate Measurement. *Lab Chip* 2019, 19 (1), 178–185. 10.1039/C8LC00968F.

- (128). Choi D-H; Gonzales M; Kitchen GB; Phan D-T; Searson PC A Capacitive Sweat Rate Sensor for Continuous and Real-Time Monitoring of Sweat Loss. *ACS Sens.* 2020, 5 (12), 3821–3826. 10.1021/acssensors.0c01219. [PubMed: 33263987]
- (129). Kwon K; Kim JU; Deng Y; Krishnan SR; Choi J; Jang H; Lee K; Su C-J; Yoo I; Wu Y; Lipschultz L; Kim J-H; Chung TS; Wu D; Park Y; Kim T; Ghaffari R; Lee S; Huang Y; Rogers JA An On-Skin Platform for Wireless Monitoring of Flow Rate, Cumulative Loss and Temperature of Sweat in Real Time. *Nat Electron* 2021. 10.1038/s41928-021-00556-2.
- (130). Reeder JT; Xue Y; Franklin D; Deng Y; Choi J; Prado O; Kim R; Liu C; Hanson J; Ciraldo J; Bandodkar AJ; Krishnan S; Johnson A; Patnaude E; Avila R; Huang Y; Rogers JA Resettable Skin Interfaced Microfluidic Sweat Collection Devices with Chemesthetic Hydration Feedback. *Nat Commun* 2019, 10 (1), 5513. 10.1038/s41467-019-13431-8. [PubMed: 31797921]
- (131). Seshadri DR; Li RT; Voos JE; Rowbottom JR; Alfes CM; Zorman CA; Drummond CK Wearable Sensors for Monitoring the Internal and External Workload of the Athlete. *npj Digital Medicine* 2019, 2 (1), 1–18. 10.1038/s41746-019-0149-2. [PubMed: 31304351]
- (132). Villiger M; Stoop R; Vetsch T; Hohenauer E; Pini M; Clarys P; Pereira F; Clijisen R Evaluation and Review of Body Fluids Saliva, Sweat and Tear Compared to Biochemical Hydration Assessment Markers within Blood and Urine. *Eur J Clin Nutr* 2018, 72 (1), 69–76. 10.1038/ejcn.2017.136. [PubMed: 28853743]
- (133). Baker LB; Nuccio RP; Reimel AJ; Brown SD; Ungaro CT; De Chavez PJD; Barnes KA Cross-validation of Equations to Predict Whole-body Sweat Sodium Concentration from Regional Measures during Exercise. *Physiol Rep* 2020, 8 (15). 10.14814/phy2.14524.
- (134). Zhao J; Lin Y; Wu J; Nyein HYY; Bariya M; Tai L-C; Chao M; Ji W; Zhang G; Fan Z; Javey A A Fully Integrated and Self-Powered Smartwatch for Continuous Sweat Glucose Monitoring. *ACS Sens.* 2019, 4 (7), 1925–1933. 10.1021/acssensors.9b00891. [PubMed: 31271034]
- (135). Vinoth R; Nakagawa T; Mathiyarasu J; Mohan AMV Fully Printed Wearable Microfluidic Devices for High-Throughput Sweat Sampling and Multiplexed Electrochemical Analysis. *ACS Sens.* 2021, 6 (3), 1174–1186. 10.1021/acssensors.0c02446. [PubMed: 33517662]
- (136). Shitanda I; Mitsumoto M; Loew N; Yoshihara Y; Watanabe H; Mikawa T; Tsujimura S; Itagaki M; Motosuke M Continuous Sweat Lactate Monitoring System with Integrated Screen-Printed MgO-Templated Carbon-Lactate Oxidase Biosensor and Microfluidic Sweat Collector. *Electrochimica Acta* 2021, 368, 137620. 10.1016/j.electacta.2020.137620.
- (137). Renner E; Lang N; Langenstein B; Struck M; Bertsch T Validating Sweat Ammonia as Physiological Parameter for Wearable Devices in Sports Science*. In 2020 42nd Annual International Conference of the IEEE Engineering in Medicine & Biology Society (EMBC); IEEE: Montreal, QC, Canada, 2020; pp 4644–4647. 10.1109/EMBC44109.2020.9175434.
- (138). Lee H-B; Meeseepong M; Trung TQ; Kim B-Y; Lee N-E A Wearable Lab-on-a-Patch Platform with Stretchable Nanostructured Biosensor for Non-Invasive Immunodetection of Biomarker in Sweat. *Biosensors and Bioelectronics* 2020, 156, 112133. 10.1016/j.bios.2020.112133. [PubMed: 32174559]
- (139). Baker LB Sweating Rate and Sweat Sodium Concentration in Athletes: A Review of Methodology and Intra/Interindividual Variability. *Sports Med* 2017, 47 (1), 111–128. 10.1007/s40279-017-0691-5. [PubMed: 28332116]
- (140). Ray TR; Ivanovic M; Curtis PM; Franklin D; Guventurk K; Jeang WJ; Chafetz J; Gaertner H; Young G; Rebollo S; Model JB; Lee SP; Ciraldo J; Reeder JT; Hourlier-Fargette A; Bandodkar AJ; Choi J; Aranyosi AJ; Ghaffari R; McColley SA; Haymond S; Rogers JA Soft, Skin-Interfaced Sweat Stickers for Cystic Fibrosis Diagnosis and Management. *Sci. Transl. Med* 2021, 13 (587), eabd8109. 10.1126/scitranslmed.abd8109. [PubMed: 33790027]
- (141). Choi D-H; Kitchen GB; Jennings MT; Cutting GR; Searson PC Out-of-Clinic Measurement of Sweat Chloride Using a Wearable Sensor during Low-Intensity Exercise. *npj Digit. Med* 2020, 3 (1), 49. 10.1038/s41746-020-0257-z. [PubMed: 32258431]
- (142). Yang Y; Song Y; Bo X; Min J; Pak OS; Zhu L; Wang M; Tu J; Kogan A; Zhang H; Hsiai TK; Li Z; Gao W A Laser-Engraved Wearable Sensor for Sensitive Detection of Uric Acid and Tyrosine in Sweat. *Nat Biotechnol* 2020, 38 (2), 217–224. 10.1038/s41587-019-0321-x. [PubMed: 31768044]

- (143). Jagannath B; Lin K; Pali M; Sankhala D; Muthukumar S; Prasad S Temporal Profiling of Cytokines in Passively Expressed Sweat for Detection of Infection Using Wearable Device. *Bioeng Transl Med* 2021. 10.1002/btm2.10220.
- (144). Bariya M; Li L; Ghattamaneni R; Ahn CH; Nyein HYY; Tai L-C; Javey A Glove-Based Sensors for Multimodal Monitoring of Natural Sweat. *Sci. Adv* 2020, 6 (35), eabb8308. 10.1126/sciadv.abb8308. [PubMed: 32923646]
- (145). Wang Y; Zhao C; Wang J; Luo X; Xie L; Zhan S; Kim J; Wang X; Liu X; Ying Y Wearable Plasmonic-Metasurface Sensor for Noninvasive and Universal Molecular Fingerprint Detection on Biointerfaces. *Science Advances* 2021, 7 (4), eabe4553. 10.1126/sciadv.abe4553. [PubMed: 33523953]
- (146). Nyein HYY; Bariya M; Tran B; Ahn CH; Brown BJ; Ji W; Davis N; Javey A A Wearable Patch for Continuous Analysis of Thermoregulatory Sweat at Rest. *Nat Commun* 2021, 12 (1), 1823. 10.1038/s41467-021-22109-z. [PubMed: 33758197]
- (147). Mishra A; Greaves R; Massie J The Relevance of Sweat Testing for the Diagnosis of Cystic Fibrosis in the Genomic Era. *Clin Biochem Rev* 2005, 26 (4), 135–153. [PubMed: 16648884]
- (148). Gibson LE; Cooke RE A Test for Concentration of Electrolytes in Sweat in Cystic Fibrosis of the Pancreas Utilizing Pilocarpine by Iontophoresis. *Pediatrics* 1959, 23 (3), 545–549. [PubMed: 13633369]
- (149). Choi D-H; Thaxton A; cheol Jeong I; Kim K; Sosnay PR; Cutting GR; Searson PC Sweat Test for Cystic Fibrosis: Wearable Sweat Sensor vs. Standard Laboratory Test. *Journal of Cystic Fibrosis* 2018, 17 (4), e35–e38. 10.1016/j.jcf.2018.03.005. [PubMed: 29580829]
- (150). Emaminejad S; Gao W; Wu E; Davies ZA; Yin Yin Nyein H; Challa S; Ryan SP; Fahad HM; Chen K; Shahpar Z; Talebi S; Milla C; Javey A; Davis RW Autonomous Sweat Extraction and Analysis Applied to Cystic Fibrosis and Glucose Monitoring Using a Fully Integrated Wearable Platform. *Proc Natl Acad Sci USA* 2017, 114 (18), 4625–4630. 10.1073/pnas.1701740114. [PubMed: 28416667]
- (151). Hauke A; Oertel S; Knoke L; Fein V; Maier C; Brinkmann F; Jank MPM Screen-Printed Sensor for Low-Cost Chloride Analysis in Sweat for Rapid Diagnosis and Monitoring of Cystic Fibrosis. *Biosensors* 2020, 10 (9), 123. 10.3390/bios10090123.
- (152). Rock M; LeGrys V The CF Quantum Sweat Test: Not Ready For Clinical Use. *Clin Lab Sci* 2020, ascls.119.002105. 10.29074/ascls.119.002105.
- (153). Teymourian H; Barfidokht A; Wang J Electrochemical Glucose Sensors in Diabetes Management: An Updated Review (2010–2020). *Chem. Soc. Rev* 2020, 49 (21), 7671–7709. 10.1039/D0CS00304B. [PubMed: 33020790]
- (154). Lee I; Probst D; Klonoff D; Sode K Continuous Glucose Monitoring Systems - Current Status and Future Perspectives of the Flagship Technologies in Biosensor Research -. *Biosensors and Bioelectronics* 2021, 181, 113054. 10.1016/j.bios.2021.113054. [PubMed: 33775474]
- (155). Juska VB; Pemble ME A Critical Review of Electrochemical Glucose Sensing: Evolution of Biosensor Platforms Based on Advanced Nanosystems. *Sensors* 2020, 20 (21), 6013. 10.3390/s20216013.
- (156). Hauke A; Simmers P; Ojha YR; Cameron BD; Ballweg R; Zhang T; Twine N; Brothers M; Gomez E; Heikenfeld J Complete Validation of a Continuous and Blood-Correlated Sweat Biosensing Device with Integrated Sweat Stimulation. *Lab Chip* 2018, 18 (24), 3750–3759. 10.1039/C8LC01082J. [PubMed: 30443648]
- (157). Jajack A; Brothers M; Kasting G; Heikenfeld J Enhancing Glucose Flux into Sweat by Increasing Paracellular Permeability of the Sweat Gland. *PLOS ONE* 2018, 13 (7), e0200009. 10.1371/journal.pone.0200009. [PubMed: 30011292]
- (158). La Count TD; Jajack A; Heikenfeld J; Kasting GB Modeling Glucose Transport From Systemic Circulation to Sweat. *Journal of Pharmaceutical Sciences* 2019, 108 (1), 364–371. 10.1016/j.xphs.2018.09.026. [PubMed: 30273561]
- (159). Karpova EV; Shcherbacheva EV; Galushin AA; Vokhmyanina DV; Karyakina EE; Karyakin AA Noninvasive Diabetes Monitoring through Continuous Analysis of Sweat Using Flow-Through Glucose Biosensor. *Anal. Chem* 2019, 91 (6), 3778–3783. 10.1021/acs.analchem.8b05928. [PubMed: 30773009]

- (160). Sempionatto JR; Moon J-M; Wang J Touch-Based Fingertip Blood-Free Reliable Glucose Monitoring: Personalized Data Processing for Predicting Blood Glucose Concentrations. *ACS Sens.* 2021, acssensors.1c00139. 10.1021/acssensors.1c00139.
- (161). Currano LJ; Sage FC; Hagedon M; Hamilton L; Patrone J; Gerasopoulos K Wearable Sensor System for Detection of Lactate in Sweat. *Sci Rep* 2018, 8 (1), 15890. 10.1038/s41598-018-33565-x. [PubMed: 30367078]
- (162). Gillan L; Teerinen T; Suhonen M; Kivimäki L; Alastalo AT Simultaneous Multi-Location Wireless Monitoring of Sweat Lactate Trends. *Flex. Print. Electron* 2021. 10.1088/2058-8585/ac13c4.
- (163). Xuan X; Pérez-Ràfols C; Chen C; Cuartero M; Crespo GA Lactate Biosensing for Reliable On-Body Sweat Analysis. *ACS Sens.* 2021. 10.1021/acssensors.1c01009.
- (164). Yu M; Li Y-T; Hu Y; Tang L; Yang F; Lv W-L; Zhang Z-Y; Zhang G-J Gold Nanostructure-Programmed Flexible Electrochemical Biosensor for Detection of Glucose and Lactate in Sweat. *Journal of Electroanalytical Chemistry* 2021, 882, 115029. 10.1016/j.jelechem.2021.115029.
- (165). Karpova EV; Laptev AI; Andreev EA; Karyakina EE; Karyakin AA Relationship Between Sweat and Blood Lactate Levels During Exhaustive Physical Exercise. *ChemElectroChem* 2020, 7 (1), 191–194. 10.1002/celec.201901703.
- (166). Sekar M; Pandiaraj M; Bhansali S; Ponpandian N; Viswanathan C Carbon Fiber Based Electrochemical Sensor for Sweat Cortisol Measurement. *Sci Rep* 2019, 9 (1), 403. 10.1038/s41598-018-37243-w. [PubMed: 30674991]
- (167). Torrente-Rodríguez RM; Tu J; Yang Y; Min J; Wang M; Song Y; Yu Y; Xu C; Ye C; IsHak WW; Gao W Investigation of Cortisol Dynamics in Human Sweat Using a Graphene-Based Wireless MHealth System. *Matter* 2020, 2 (4), 921–937. 10.1016/j.matt.2020.01.021. [PubMed: 32266329]
- (168). Cheng C; Li X; Xu G; Lu Y; Low SS; Liu G; Zhu L; Li C; Liu Q Battery-Free, Wireless, and Flexible Electrochemical Patch for in Situ Analysis of Sweat Cortisol via near Field Communication. *Biosensors and Bioelectronics* 2021, 172, 112782. 10.1016/j.bios.2020.112782. [PubMed: 33157409]
- (169). Sun M; Gu Y; Pei X; Wang J; Liu J; Ma C; Bai J; Zhou M A Flexible and Wearable Epidermal Ethanol Biofuel Cell for On-Body and Real-Time Bioenergy Harvesting from Human Sweat. *Nano Energy* 2021, 86, 106061. 10.1016/j.nanoen.2021.106061.
- (170). Tai L-C; Ahn CH; Nyein HYY; Ji W; Bariya M; Lin Y; Li L; Javey A Nicotine Monitoring with a Wearable Sweat Band. *ACS Sens.* 2020, 5 (6), 1831–1837. 10.1021/acssensors.0c00791. [PubMed: 32429661]
- (171). Lin S; Yu W; Wang B; Zhao Y; En K; Zhu J; Cheng X; Zhou C; Lin H; Wang Z; Hojaiji H; Yeung C; Milla C; Davis RW; Emaminejad S Noninvasive Wearable Electroactive Pharmaceutical Monitoring for Personalized Therapeutics. *Proc Natl Acad Sci USA* 2020, 117 (32), 19017–19025. 10.1073/pnas.2009979117. [PubMed: 32719130]
- (172). Tai L-C; Gao W; Chao M; Bariya M; Ngo QP; Shahpar Z; Nyein HYY; Park H; Sun J; Jung Y; Wu E; Fahad HM; Lien D-H; Ota H; Cho G; Javey A Methylxanthine Drug Monitoring with Wearable Sweat Sensors. *Advanced Materials* 2018, 30 (23), 1707442. 10.1002/adma.201707442.
- (173). Xu G; Cheng C; Liu Z; Yuan W; Wu X; Lu Y; Low SS; Liu J; Zhu L; Ji D; Li S; Chen Z; Wang L; Yang Q; Cui Z; Liu Q Battery-Free and Wireless Epidermal Electrochemical System with All-Printed Stretchable Electrode Array for Multiplexed In Situ Sweat Analysis. *Advanced Materials Technologies* 2019, 4 (7), 1800658. 10.1002/admt.201800658.
- (174). Jeerapan I; Sempionatto JR; Wang J On-Body Bioelectronics: Wearable Biofuel Cells for Bioenergy Harvesting and Self-Powered Biosensing. *Adv. Funct. Mater* 2020, 30 (29), 1906243. 10.1002/adfm.201906243.
- (175). Jin X; Bandothkar AJ; Fratus M; Asadpour R; Rogers JA; Alam MA Modeling, Design Guidelines, and Detection Limits of Self-Powered Enzymatic Biofuel Cell-Based Sensors. *Biosensors and Bioelectronics* 2020, 168, 112493. 10.1016/j.bios.2020.112493. [PubMed: 32889394]

- (176). Sun M; Xin T; Ran Z; Pei X; Ma C; Liu J; Cao M; Bai J; Zhou M A Bendable Biofuel Cell-Based Fully Integrated Biomedical Nanodevice for Point-of-Care Diagnosis of Scurvy. *ACS Sens.* 2021, 6 (1), 275–284. 10.1021/acssensors.0c02335. [PubMed: 33356148]
- (177). Yin L; Moon J-M; Sempionatto JR; Lin M; Cao M; Trifonov A; Zhang F; Lou Z; Jeong J-M; Lee S-J; Xu S; Wang J A Passive Perspiration Biofuel Cell: High Energy Return on Investment. *Joule* 2021, 5 (7), 1888–1904. 10.1016/j.joule.2021.06.004.
- (178). Hojajji H; Zhao Y; C. Gong M; Mallajosyula M; Tan J; Lin H; M. Hojajji A; Lin S; Milla C; M. Madni A; Emaminejad S An Autonomous Wearable System for Diurnal Sweat Biomarker Data Acquisition. *Lab on a Chip* 2020, 20 (24), 4582–4591. 10.1039/D0LC00820F. [PubMed: 33052990]
- (179). Sempionatto JR; Lin M; Yin L; De la paz E; Pei K; Sonsa-ard T; de Loyola Silva AN; Khorshed AA; Zhang F; Tostado N; Xu S; Wang J An Epidermal Patch for the Simultaneous Monitoring of Haemodynamic and Metabolic Biomarkers. *Nat Biomed Eng* 2021. 10.1038/s41551-021-00685-1.
- (180). Nyein HYY; Bariya M; Kivimäki L; Uusitalo S; Liaw TS; Jansson E; Ahn CH; Hangasky JA; Zhao J; Lin Y; Happonen T; Chao M; Liedert C; Zhao Y; Tai L-C; Hiltunen J; Javey A Regional and Correlative Sweat Analysis Using High-Throughput Microfluidic Sensing Patches toward Decoding Sweat. *Sci. Adv* 2019, 5 (8), eaaw9906. 10.1126/sciadv.aaw9906. [PubMed: 31453333]
- (181). Epicore Biosystems <http://www.epicorebiosystems.com/> (accessed 2021 –05 –31).

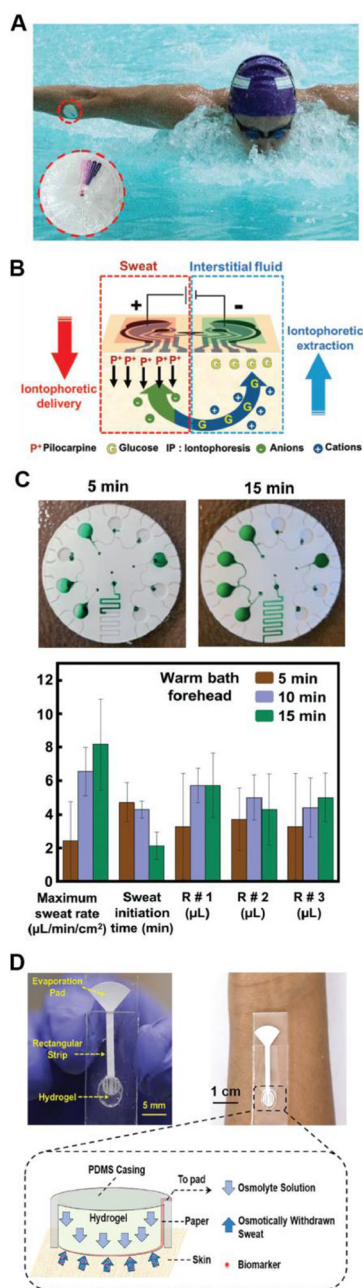


Figure 1.

Methods for sweat stimulation. Typical methods for sweat stimulation include (A) physical activity⁸⁶ or (B) pharmacological stimulation⁸⁷. Adapted with permission from ref (⁸⁶), Copyright 2019 American Association for the Advancement of Science, and ref (⁸⁷), Copyright 2018 John Wiley and Sons, respectively. Alternative approaches seek to collect sweat passively using (C) thermal stimulation via showering⁸⁸ or (D) wicking materials⁸⁹. Adapted with permission from ref (⁸⁸), Copyright 2019 The Royal Society of Chemistry, and ref (⁸⁹), Copyright 2021 American Chemical Society, respectively.

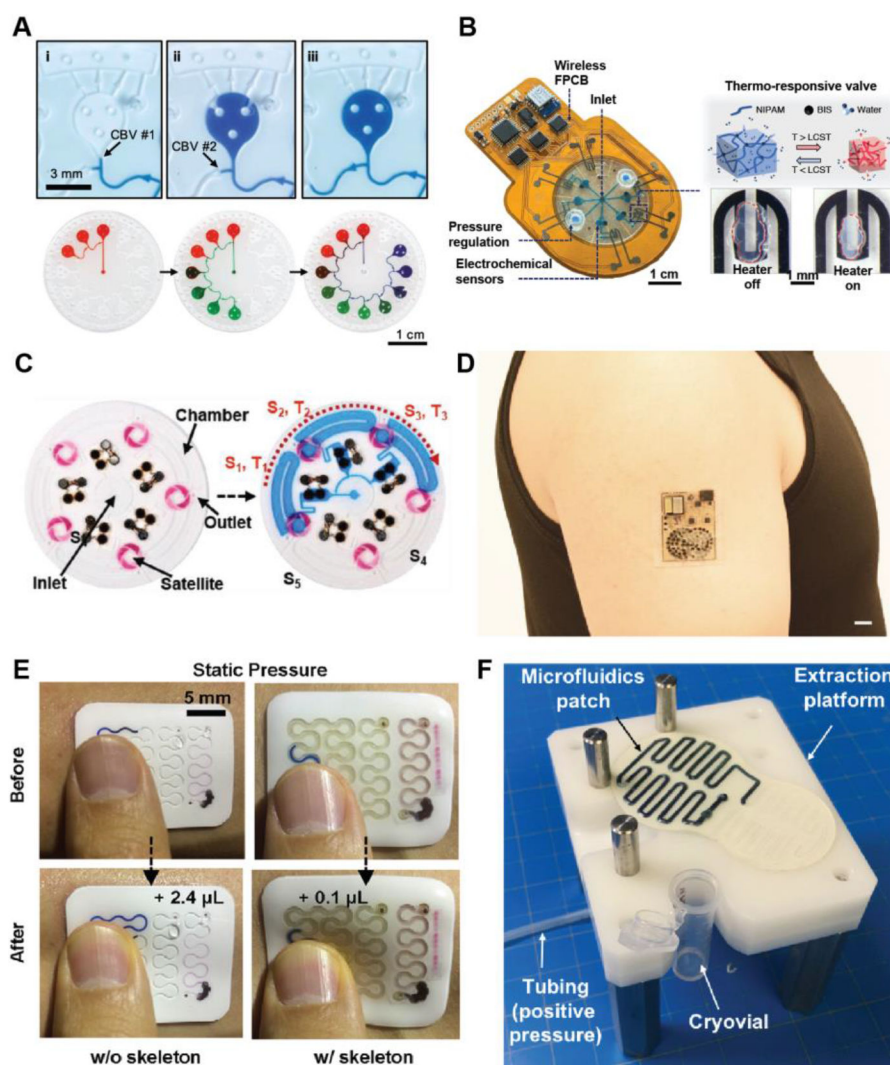


Figure 2. Technology foundations for wearable sweat sensing. Fluid Handling. Networks of (A) passive⁹⁰ or (B) active valves⁹¹ enable sophisticated routing of harvested sweat in a programmatic manner. Adapted with permission from ref (⁹⁰), Copyright 2017 John Wiley and Sons, and ref (⁹¹), Copyright 2020 Springer Nature, respectively Timing. Nuanced designs integrate sensing features such as sweat-activated galvanic cells shown in (C) to enable temporal analysis of sweat constituents⁹². Adapted with permission from ref (⁹²). Copyright 2019 John Wiley and Sons. In large arrays (D) such biofuel cells support battery-free electrochemical sensing of sweat⁹³. Adapted with permission from ref (⁹³). Copyright 2020 American Association for the Advancement of Science. Advanced designs. Optimization of mechanical properties (E) address sensing challenges in high-impact environments⁹⁴. Adapted with permission from ref (⁹⁴). Copyright 2020 John Wiley and Sons. Utilization of customized extraction hardware (F) assists in reducing sample contamination⁹⁵. Adapted with permission from ref (⁹⁵). Copyright 2021 Elsevier B.V.

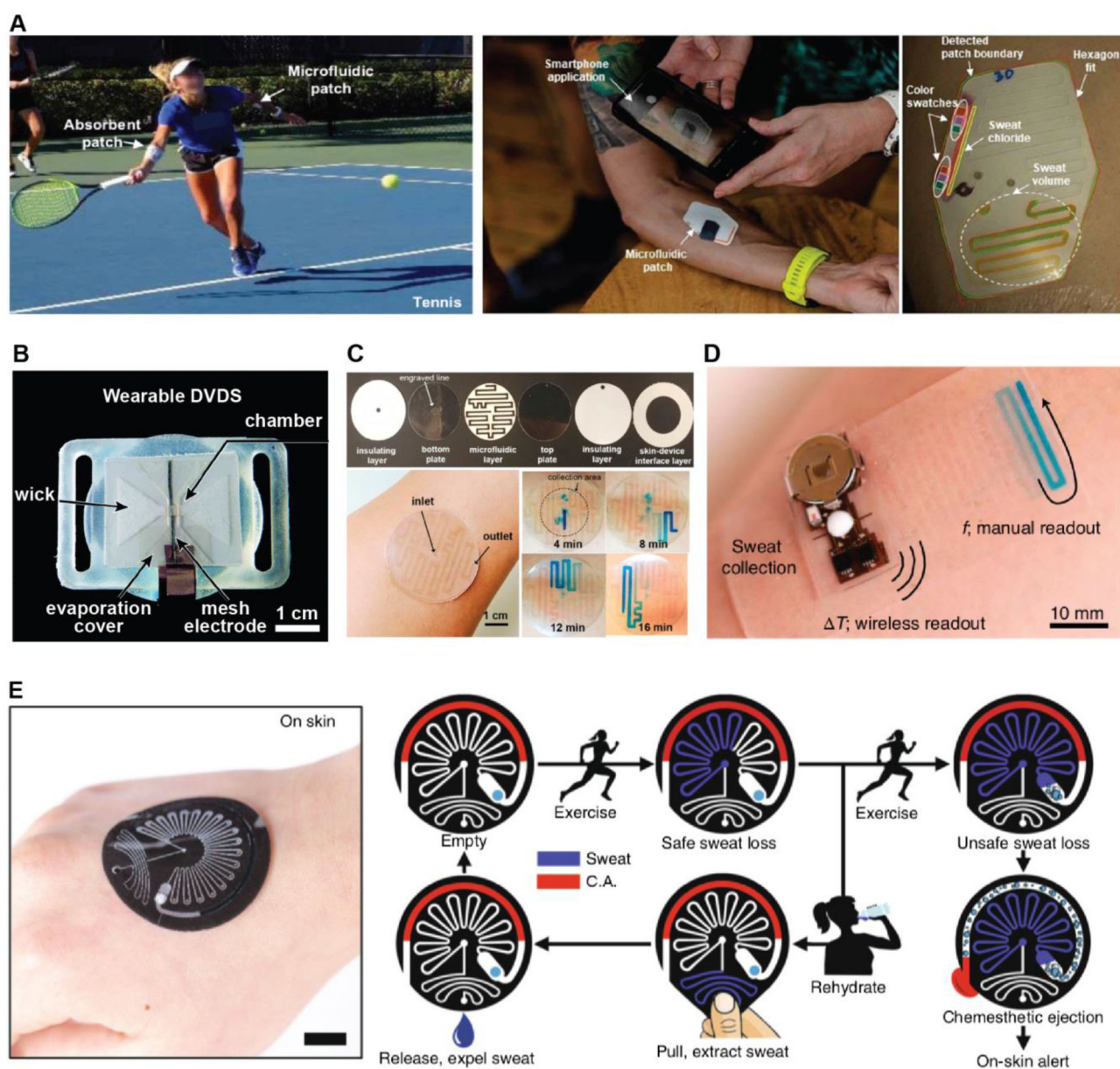


Figure 3. Performance health management. (A) A recent large-scale study validates the performance of a wearable microfluidic patch for estimating whole-body sweat parameters from regional sweat analysis¹²⁶. Many sweat biomarkers exhibit a concentration dependence on rate of sweat loss. Adapted with permission from ref (¹²⁶). Copyright 2020 American Association for the Advancement of Science. Recent embodiments utilize (B) conductivity¹²⁷, (C) capacitive¹²⁸, or (D) thermal¹²⁹ sensing strategies to continuously measure real-time sweat rate. Adapted with permission from ref (¹²⁷), Copyright 2019 The Royal Society of Chemistry, ref (¹²⁸), Copyright 2020 American Chemical Society, ref (¹²⁹), Copyright 2021 Springer Nature, respectively. (E) Emerging device architectures integrate chemesthetic sensors and user-activated valves to alert wearers to anomalous physiological conditions

during exercise¹³⁰. Adapted with permission from ref (¹³⁰). Copyright 2019 Springer Nature.

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript



Figure 4.

Clinical diagnostics. Sweat chloride. Sweat chloride is a longstanding clinically validated diagnostic biomarker used for confirmatory diagnosis of cystic fibrosis (CF). Recent reports demonstrate (A) the first large-scale study of a soft, flexible epidermal platform (“sweat sticker”)¹⁴⁰ for clinical diagnosis and (B) use of wearable sweat sensors for monitoring sweat chloride levels outside of a clinical setting¹⁴¹. Adapted with permission from ref (140), Copyright 2021 American Association for the Advancement of Science, and ref (141), Copyright 2020 Springer Nature, respectively. Emerging sweat biomarkers. Use of sweat glucose as a noninvasive replacement for blood glucose monitoring in diabetes management is of academic and commercial interest with recent efforts demonstrating sensors for monitoring sweat glucose levels (C) at rest⁸³ and (D) during exercise¹¹¹. Adapted with permission from ref (83), Copyright 2020 American Chemical Society, and ref (111), Copyright 2020 American Chemical Society, respectively. (E) One embodiment demonstrates glucose monitoring during exercise in wireless, battery-free form factor.¹¹⁶ Adapted with permission from ref (116). Copyright 2019 American Association for the Advancement of Science. Other targets of interest include the concentration of (F) uric acid in sweat¹⁴² (for gout), (G) various cytokines¹⁴³ (inflammation, fever), (H) vitamins¹⁴⁴ (nutrition monitoring), and (I) illicit drugs¹⁴⁵. Adapted with permission from ref (142), Copyright 2020 Springer Nature; ref (143), Copyright 2021 John Wiley and Sons; ref (144), Copyright 2020 American Association for the Advancement of Science; ref (145), Copyright 2021 American Association for the Advancement of Science, respectively. (J) Device designs exploiting wicking materials enable passive (i.e., absence of active

sweating) multiparameter monitoring of disease biomarkers or the concentration of drug therapeutics¹⁴⁶. Adapted with permission from ref (¹⁴⁶). Copyright 2021 Springer Nature.

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

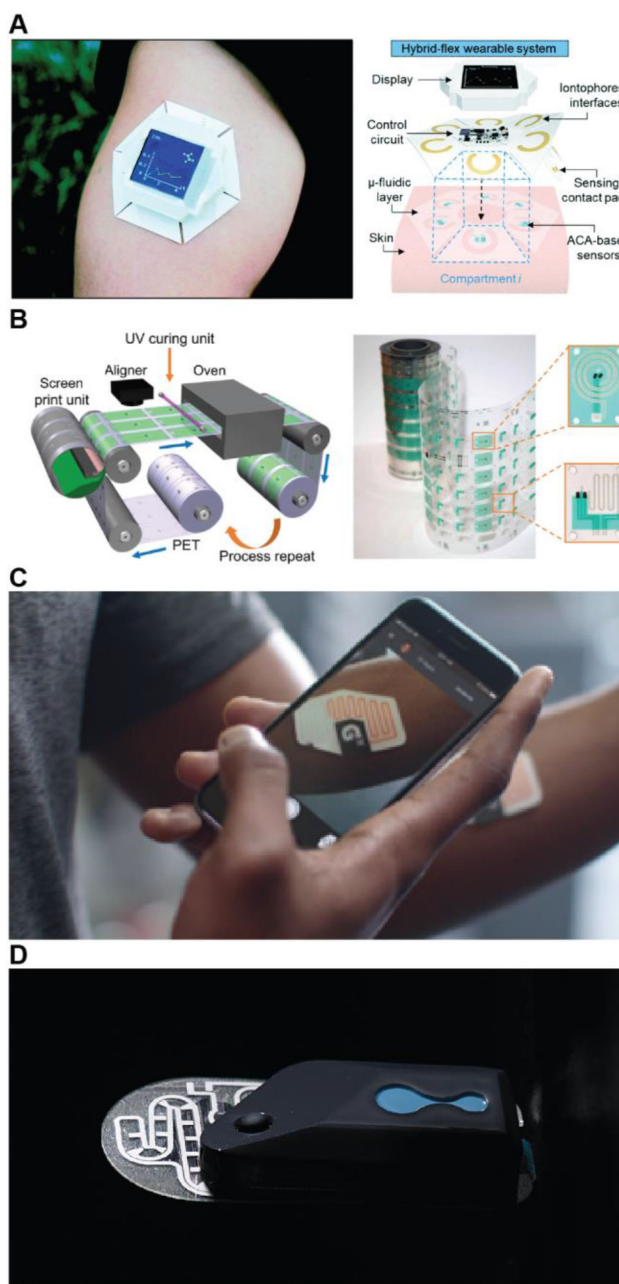


Figure 5. Integrated devices and commercially available systems. (A) An integrated device strategy for long-term sweat analysis via on-demand sweat stimulation¹⁷⁸. Adapted with permission from ref (¹⁷⁸). Copyright 2020 The Royal Society of Chemistry. (B) A recent effort details a strategy to utilize roll-to-roll manufacturing to produce epidermal microfluidic sensors in a scalable manner suitable for mass manufacture¹⁸⁰. Implementation of such fabrication strategies enables further maturation of wearable sweat sensing platforms and offers opportunities for broad consumer adoption. Adapted with permission from ref (¹⁸⁰). Copyright 2019 American Association for the Advancement of Science. (C) Gx Sweat Patch and (D) Connected Hydration System, both developed by Epicore Biosystems¹⁸¹, represent

the vanguard of the emerging commercial sensing platforms. Reprinted with permission from ref (¹⁸¹). Copyright 2021 Epicore Biosystems.

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript