

Salivary IgA against sporozoite-specific embryogenesis-related protein (TgERP) in the study of horizontally transmitted toxoplasmosis via *T. gondii* oocysts in endemic settings

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SUMMARY

The aim of this study was to contribute to the better understanding of the relative epidemiological importance of different modes of infection with respect to horizontal transmission of *Toxoplasma gondii* in endemic settings. We investigated the prevalence of salivary IgA against a sporozoite-specific embryogenesis-related protein (TgERP) in a highly endemic area for toxoplasmosis in Brazil in order to pinpoint parasite transmission via oocysts. Prevalence calculated by salivary IgA specific to TgERP was compared to the prevalence calculated by serum IgG against both TgERP and tachyzoites (in conventional serological tests). Prevalence calculated by different serological and salivary parameters varied in the studied age groups. However, for the 15–21 years age group, values for *T. gondii* prevalence estimated by conventional serological tests and by anti-TgERP salivary IgA were similar; i.e. 68·7% and 66·6% or 66·7%, respectively, using two different cut-off parameters for salivary IgA anti-TgERP. Furthermore, salivary IgA anti-TgERP for this age group presented the highest specificity (93·33%), sensitivity (93·94%), and likelihood (14·09) compared to all the other age groups. These data demonstrate the importance of age for salivary IgA investigation against TgERP to estimate the mode of *T. gondii* transmission in endemic settings.

Key words: Diagnosis, oocysts, salivary IgA, *Toxoplasma gondii*, waterborne infections.

INTRODUCTION

Toxoplasmosis infection caused by the protozoan *Toxoplasma gondii* is widely distributed in humans in many parts of the world [1]. In Brazil, its prevalence ranks among the highest in the world [2]. A broad

range of potential routes of *T. gondii* transmission has been developed by the parasite in the course of its evolution [3]. Congenital toxoplasmosis in humans results from vertical transmission of the parasite during pregnancy and constitutes a very dangerous mode of infection to the fetus, causing stillbirth or a variety of disease syndromes, from chronic infection without apparent clinical symptoms, to blindness and mental retardation in children [4, 5]. Besides congenital infection, humans become infected through ingestion of tissue cysts in undercooked or uncooked

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meat, or by ingesting food or water contaminated with sporulated oocysts (containing the infective sporozoite forms) from infected feline faeces [6–12]. The relative epidemiological importance of different modes of infection with respect to horizontal transmission of *T. gondii*, namely by ingestion of tissue cysts in undercooked or uncooked meat, or by ingesting food or water contaminated with sporulated oocysts from cat faeces [13–16], remains unknown. The ability to discriminate between the two modes of infection depends, among other factors, on serological tests capable of discriminating both modes of infection. In epidemiological terms, using conventional serology, which recognizes antigens from tachyzoites, it is not possible to determine the predominant route (oocysts or tissue cysts) of transmission [17]. Serological diagnosis pinpointing *T. gondii* cysts or oocysts as the mode of transmission by means of an ELISA using a recombinant protein, i.e. the sporozoite-specific embryogenesis-related protein (TgERP) [18], and more recently the CCp5A recombinant protein [19], has allowed differentiation between the two modes of infection.

IgA antibodies together with IgM and IgG against antigens derived from tachyzoite forms of *T. gondii* are frequently detected in sera of patients in the early and chronic phase of toxoplasmosis [20]. The presence of IgG and IgA anti-*T. gondii* in saliva and other body fluids has also been reported [21–25]. However, the specific role of IgA against *T. gondii* in the human oral cavity remains elusive. The isolation and/or detection of viable *T. gondii* from human saliva has also been reported [21, 26, 27], thus the presence of IgA against sporulated *T. gondii* oocyst antigens in the saliva of patients exposed to the risk of *T. gondii* infection is plausible and an open question for investigation. In the present study *T. gondii* prevalence using conventional and TgERP ELISAs was investigated in the sera and saliva of patients exposed to the risk of *T. gondii* infection via oocyst infection [20]. It was found that salivary IgA against TgERP is an important antibody to be considered in the study of horizontal *T. gondii* infection transmitted by oocysts.

METHODS

Study population

The subjects enrolled in the present study were from a previously studied endemic area for toxoplasmosis

[28]. In terms of socioeconomic status, they correspond to the lower and lower/middle-class income groups [28]. For sample size calculation (confidence level of 95% and margin of error of 5%) it was considered the seroprevalence reference of 73.1%, which is the expected frequency for lower and lower/middle-class income groups as previously described [28, 29]; the required sample size was 302 subjects. A total of 476 individuals associated (as students, school staff, students' parents) with 10 public schools from Campos dos Goytacazes, located in the north of Rio de Janeiro state (Table 1), were tested in the current study. Schools were representative of the city in terms of geographical locations (urban, suburban, or rural areas) [28]. Individuals were invited to participate in the study; consent was sought and granted (self or guardian) after individuals were informed about the project. Five millilitres of blood was collected from each individual for serological assay. Saliva was collected under stimulation by rubbing the gums with a Salivette sampler sponge (Sarstedt AG & Co., Germany) for ~1 min. After centrifugation, saliva samples were separated in aliquots prior to storage. Each clinical sample (sera and saliva) was labelled to preserve individual identification codes and kept at –20 °C until use. In a group of 96 individuals aged >28 years, members of the adult school staff, only 5 ml of blood sample was collected (no saliva sample was collected) (Table 1). Standard serological tests [Vidas Toxo IgG and IgM kits (bioMérieux, France) and Architect Toxo IgG and IgM kits (Abbott Diagnostics Division, USA)] that are able to detect previous *T. gondii* exposure without discriminating the route of infection were performed to evaluate acute and chronic *T. gondii* human infection. Subjects included in control groups were those that tested negative for conventional ELISA against *T. gondii* (IgG and or IgM) and varied according to age group: 0–7 years ($n = 104$ subjects), 8–14 years ($n = 99$), 15–21 years ($n = 15$), and 22–28 years ($n = 15$).

ELISA against TgERP

The sera and saliva of 380 subjects were analysed for reactivity against TgERP [18]. Briefly, testing for reactivity to TgERP was performed by ELISA; TgERP (uncleaved with factor Xa) was diluted to a concentration of 2 µg/ml in 0.1 M carbonate buffer (pH 9.6). In sequence, plates were washed three times with wash buffer [50 mM Tris (pH 7.4),

Table 1. *Toxoplasma gondii* reactivity positivity in the study population

Study population <i>T. gondii</i> seropositivity, <i>n</i> (%) by conventional ELISA (IgG and IgM from sera) and by TgERP IgG (from sera) and IgA (from saliva)										
Coventional ELISA					TgERP ELISA*					
Age group, yr	No. tested	Male/Female	IgG only	IgM only	IgG and IgM	IgG and/or IgM	IgG	IgA	IgG	IgA
0-7	116	70/46	13 (11.2)	5 (4.3)	3 (2.6)	21 (18.1)	9 (7.75)*	18 (15.5)*	36 (31)†	38 (32.8)†
8-14	156	73/83	59 (37.8)	4 (2.5)	3 (1.9)	66 (42.3)	20 (12.8)*	31 (19.9)*	75 (48.1)†	73 (46.8)†
15-21	48	14/34	33 (68.7)	1 (2.1)	1 (2.1)	35 (72.9)	13 (27.1)*	32 (66.6)*	34 (70.8)†	32 (66.7)†
22-28	60	4/56	45 (75)	2 (3.3)	2 (3.3)	49 (81.6)	31 (51.7)*	39 (65)*	45 (75)†	40 (66.7)†
>28	96	16/80	75 (78.1)	3 (3.1)	3 (3.1)	78 (81.2)	ND	ND	ND	ND
Total	476	177/299	225 (3.1)	15 (3.1)	12 (2.5)	249 (52.3)	73 (19.2)‡	120 (31.6)‡	190 (50)‡	183 (48.1)‡

TgERP, Sporozoite-specific embryogenesis-related protein; ND, not done.

* Values and percentages considering s.d. cut-off.

† Values and percentages considering ROC cut-off.

‡ The total subjects tested for these groups is 380 and corresponds to those tested against TgERP.

150 mM NaCl, 5% skim milk, 1.0% Triton X-100]. The plates were then incubated with 100 µl/well of pre-diluted sera or saliva (1:100 and 1:10, respectively) in wash buffer for 30 min at room temperature. After another cycle of three washes, 100 µl of rabbit immunoglobulin anti-human IgG (for sera samples) or anti-human IgA (for saliva samples) conjugated to peroxidase (Kirkegaard & Perry Laboratories Inc., USA) pre-diluted 1:1000 in wash buffer was added to sera and saliva tests, respectively. The plates were incubated for 30 min at room temperature and then washed again three times with wash solution and incubated with 100 µl/well of developing solution. The reading was performed in an ELISA reader (VMax ELISA reader, Molecular Devices Inc., USA) at 405 nm. Reference positive and negative controls were included on each plate for cut-off calculations.

Cut-off calculations for ELISA against TgERP

The cut-off calculation based on standard deviation (s.d.) was established from a positive cut-off as the mean value of optical density (OD) of seronegative samples in conventional ELISA, plus three times the s.d. of seronegative samples as described previously [18]. Cut-off calculation was based on the receiver-operating characteristic (ROC) curve: the ROC curve data provided the accuracy of each test with area under the curve (AUC) measurements.

$$\text{sensitivity} = \frac{\text{true positives}}{\text{true positives} + \text{false negatives}} \times 100,$$

$$\text{specificity} = \frac{\text{true negatives}}{\text{true negatives} + \text{false positives}} \times 100,$$

$$\text{likelihood ratio} = \left[\frac{\text{sensitivity}}{1 - \text{specificity}} \right]$$

were also calculated for each immunoglobulin reactivity pattern.

Statistical analysis

Statistical analyses were performed by GraphPad Prism v. 6.0 software (Graphpad Software, USA) for all statistical analyses (ROC curve and Spearman analyses).

Ethical standards

The study protocol complied with the Declaration of Helsinki and was approved by the local ethics

committee (protocol nos. 164525/027/07 and 716-147/2014).

RESULTS

Study population characteristics and prevalence of anti-*T. gondii* IgM and IgG by conventional serology

The mean age of patients was 17.89 years (s.d. = 14.79, 95% confidence interval 16.56–19.23, range 0–75 years); 177 (37.2%) were males and 299 (62.8%) females (Table 1). The overall seroprevalence of *T. gondii* to IgG and/or IgM antibodies tested by conventional ELISA was 52.3% (249/476). Of this, IgG antibodies were detected in 49.8% (237/476) while IgM antibodies were present in 5.7% (27/476). Twelve (2.5%) participants tested positive for both IgG and IgM antibodies. Fifteen (3.1%) participants tested positive for only IgM antibodies while 47.3% (225/476) participants tested positive for only IgG antibodies. The remaining 47.7% (227/476) of participants tested negative for all antibodies. A progressively increased statistically significant ($P < 0.0001$) anti-*T. gondii* seropositivity (IgG) was observed as age increased (Table 1). For IgM, however, the prevalence was not associated with age ($P = 0.9284$).

ROC curves for anti-TgERP IgG and IgA reactivity

The ability of anti-TgERP IgG in sera samples and IgA in saliva samples to discriminate seropositive populations (by conventional ELISA) was analysed by ROC curves (Fig. 1*a, b*). The 1:100 serum dilution was selected to measure IgG, whereas 1:10 saliva dilution was the best for discriminatory analysis of IgA. IgA reactivity was the most promising biomarker for discrimination between positive and negative individuals and those who were negative for toxoplasmosis by conventional ELISA. The IgA and IgG analysis by ROC curves presented good performances in general for all age groups. However, for the 0–7 years age group the cut-off values established by the ROC curve from saliva IgA is below the level of the ELISA's technical sensitivity capacity (OD = 0.074). A good performance was observed for IgG sera samples for the 15–21 years age group (Se = 90.91%, Sp = 73.33%, AUC = 0.8596 and LR+ = 3.409) (Fig. 1*a, b*). However, the best performance for the 15–21 years age group was found for IgA saliva samples (Se = 93.94%, Sp = 93.33%, AUC = 0.9677 and LR+ = 14.09) compared to all other age groups.

Prevalence of anti-TgERP IgG and IgA according to cut-off determination and Spearman's correlation between sera IgG and saliva IgA anti-TgERP

The seroprevalence by conventional ELISA IgG and TgERP ELISA (IgG from sera and IgA from saliva) are shown in Table 1 and Figure 2(*a, b*). The prevalence of IgG measured by antibodies against TgERP reached 75% in individuals aged 22–28 years; the prevalence measured by conventional ELISA in the same age group is 81.6% (Table 1). In Figure 2(*a, b*), the prevalence curves are shown as a function of the age group population. The seroprevalence indicated in the TgERP ELISA was calculated considering two cut-off criteria: (1) the threefold standard deviation (s.d. cut-off) values of samples negative in conventional ELISA as published previously [18, 30] and (2) by ROC curve (ROC cut-off). The prevalence curves for conventional ELISA IgG and TgERP IgG and IgA presented a generally similar shape even when considering s.d. cut-off or ROC cut-off for TgERP ELISA (Fig. 2*a, b*) with diverging points in all age groups with exception of the 15–21 years age group. For this age group the prevalence values were very similar: for IgG anti-*T. gondii* conventional ELISA (68.7%), and for IgA anti-TgERP ELISA (66.6% and 66.7%) when considering s.d. cut-off and ROC curve cut-off, respectively (Table 1). This age group is included in the section of the age prevalence curve where the slope is highest for both conventional ELISA (IgG) and TgERP ELISA (IgG and IgA) (Fig. 2*a, b*, respectively). For all other age groups the *T. gondii* prevalence curves estimated by TgERP ELISA using s.d. cut-off criteria for IgG and IgA measurements were lower than the prevalence estimated by ROC cut-off criteria (Table 1). The very low prevalence of IgM estimated by conventional ELISA detected in the present study is shown by a dotted line in Figure 2(*a, b*). Spearman's correlation test between ELISA OD values of anti-TgERP IgG from sera and anti-TgERP IgA from saliva was used to estimate strength of the link between systemic IgG and salivary IgA, both against TgERP. The best significant correlation parameters were observed for the 15–21 years age group ($r = 0.56$ and $P < 0.0001$). Considering prevalence calculated by IgG anti-TgERP (70.8%) using ROC cut-off values (shown in Table 1) in this age group, it is probable that the cut-off values for TgERP based on ROC curves may be better than the s.d. cut-off for ages >7 years.

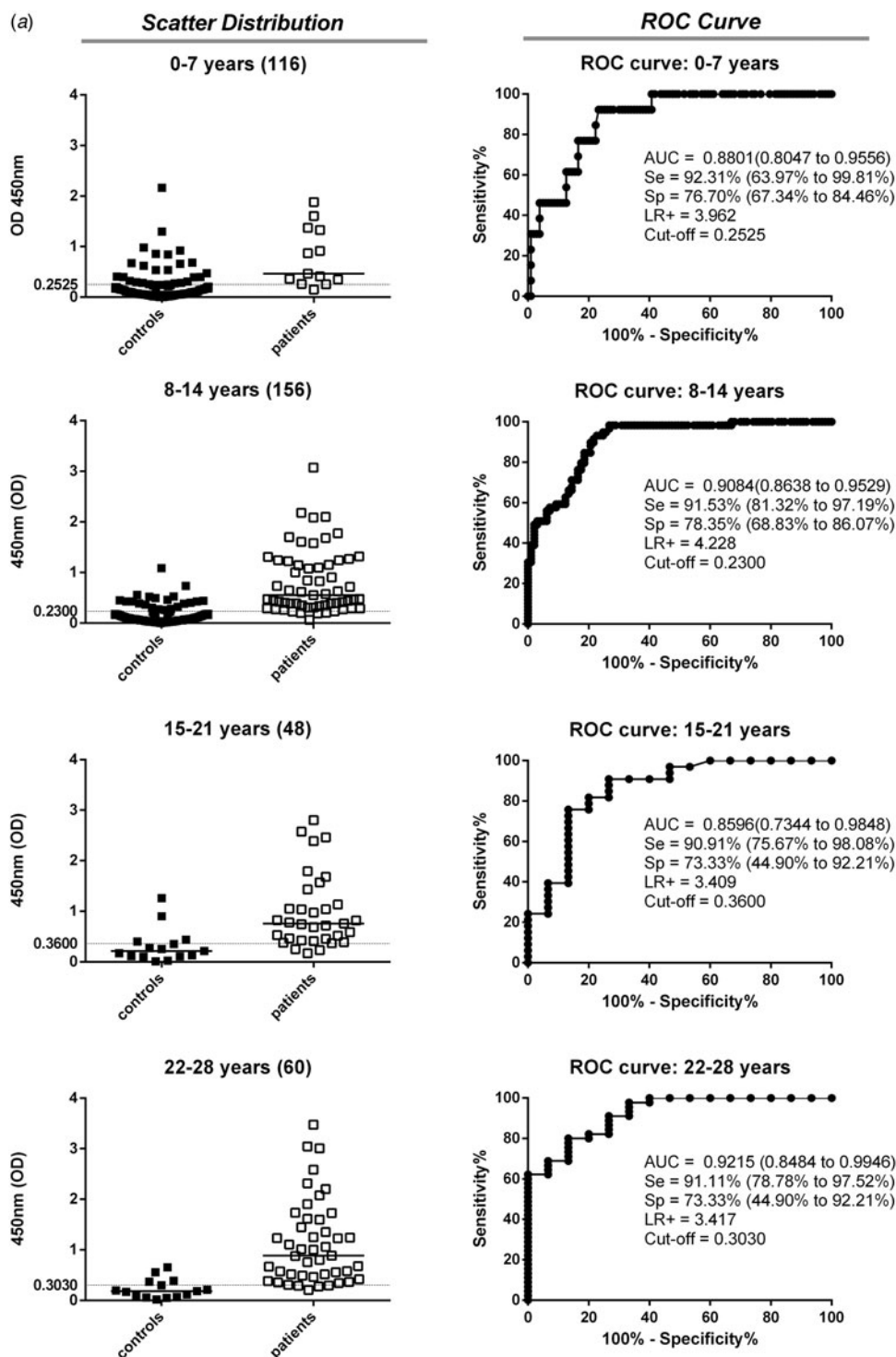


Fig. 1. Anti-*Toxoplasma gondii* immunoglobulins detected by conventional ELISA (IgG) and sporozoite-specific embryogenesis-related protein (TgERP) ELISA (IgG and IgA) to discriminate controls (■) from patients (□) in the different age groups for the individuals enrolled in the study. Data are expressed as scattering distribution of optical density (OD) for all immunoglobulins. ROC curve analysis was used to establish the best cut-off edge to segregate immunoglobulin reactivity in the groups. The cut-off edges and the performance indices (Se, Sp, AUC and LR+) are provided. (a) IgG reactivity assessed in sera samples for TgERP ELISA. (b) IgA reactivity assessed in saliva samples for TgERP ELISA. The number of individuals in each group is denoted within parentheses. The subjects included in control groups were those that tested negative for conventional ELISA against *T. gondii* (IgG and/or IgM) and varied according to age group: 0–7 years ($n = 104$ subjects), 8–14 years ($n = 99$), 15–21 years ($n = 15$), and 22–28 years ($n = 15$).

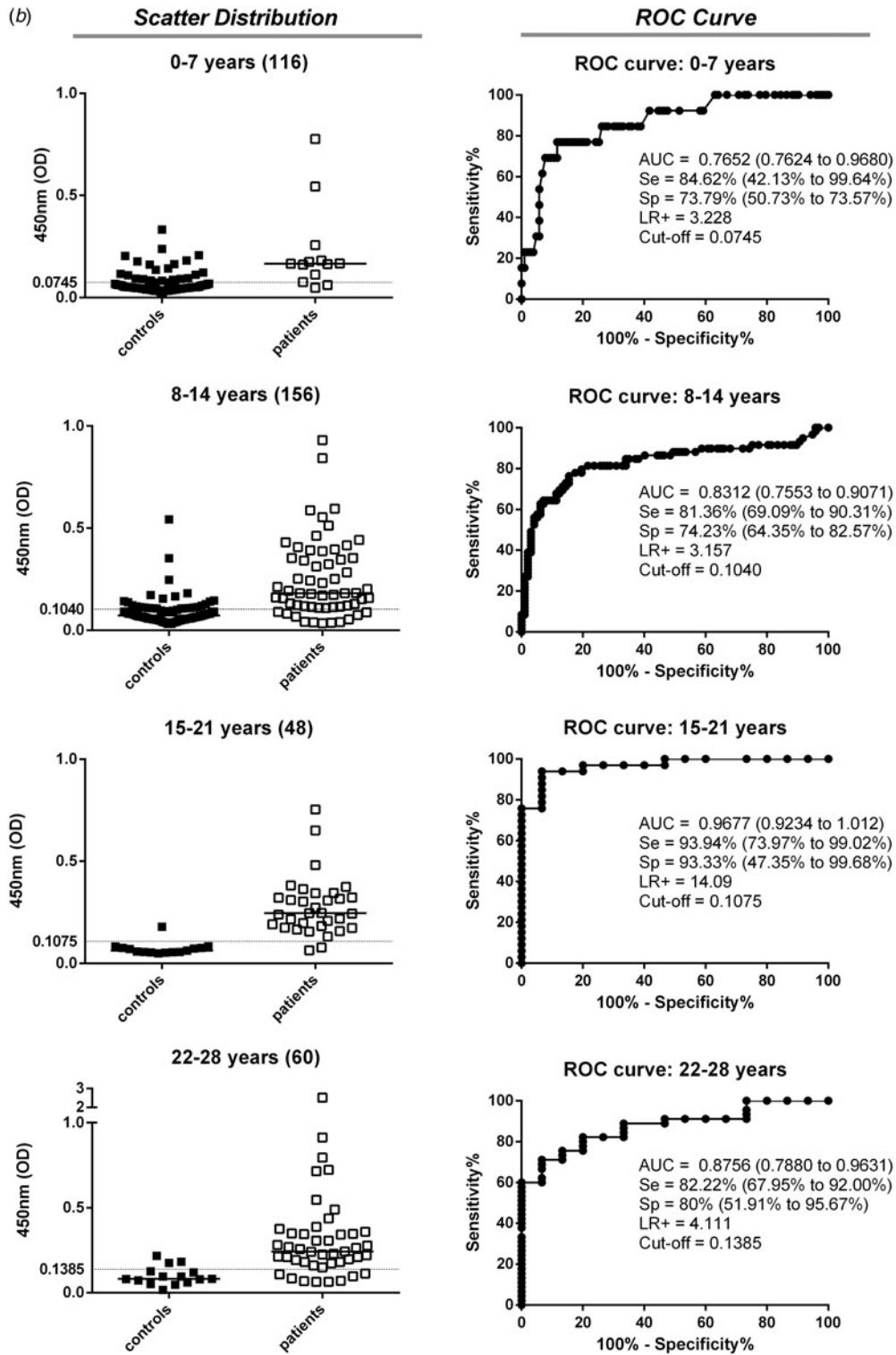


Fig. 1b (cont.)

DISCUSSION

The aim of this study was to contribute to the better understanding of the relative epidemiological importance of different modes of infection with respect to

horizontal transmission of *T. gondii*, namely by ingestion of tissue cysts in undercooked or uncooked meat, or by ingestion of food or water contaminated with sporulated oocysts (containing the infective sporozoite

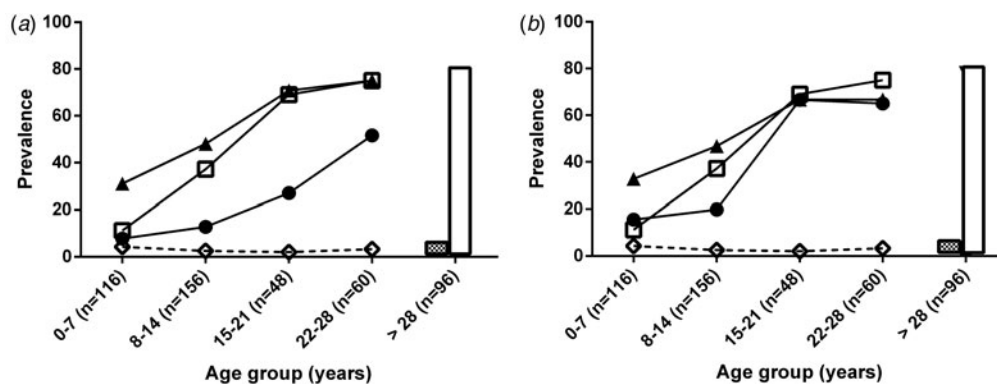


Fig. 2. *T. gondii* seroprevalence according to age, as determined by conventional ELISA and sporozoite-specific embryogenesis-related protein (TgERP) ELISA for all 476 individuals enrolled in this study. Data are expressed as percentages for all curves. (a) Seroprevalences according to age assessed in sera samples by IgG anti-*T. gondii* conventional ELISA (□), IgG TgERP s.d. cut-off (●), IgG TgERP ROC cut-off (▲) and IgM anti-*T. gondii* conventional ELISA (◇). The bars represent the seroprevalence of IgM (black bars) and IgG (white bars) in the >28 years age group. (b) Seroprevalence according to age assessed in saliva samples by IgG anti-*T. gondii* conventional ELISA (□), IgA TgERP s.d. cut-off (●), IgA TgERP ROC cut-off (▲) and IgM anti-*T. gondii* conventional ELISA (◇). The bars represent the seroprevalences of IgM (white bars with black circles) and IgG (white bars) in the >28 years age group. The number of individuals in each group is denoted within parentheses. The subjects included in control groups were those that tested negative for conventional ELISA against *T. gondii* (IgG and or IgM) and varied according to age group: 0–7 years ($n = 104$ subjects), 8–14 years ($n = 99$), 15–21 years ($n = 15$), and 22–28 years ($n = 15$).

forms) from cat faeces. The present findings will be discussed from an epidemiological perspective highlighting the potential value of salivary IgA against TgERP as a tool to investigate the predominant transmission route (oocysts or tissue cysts) of *T. gondii* infection in epidemiological surveys.

The disparities of prevalence rates found for each studied age group employing two different cut-off parameters shown in Table 1 provide evidence that more population-based studies employing *T. gondii* sporozoite antigens must be conducted to improve our understanding. With the exception of the prevalence calculated for the 15–21 years age group employing salivary IgA using both s.d. and ROC cut-offs and IgG employing sera samples using the s.d. cut-off (Table 1), all other prevalences calculated using positivity against TgERP presented considerable disparities with prevalence calculated by IgG (alone or together with IgM) detected by commercial kits. Nonetheless, the present study indeed confirms the importance of oocyst infection contributing to the overall prevalence of toxoplasmosis in endemic settings as shown in Table 1 and Figure 2(a, b) and reported previously [30]. To the best of our knowledge this is the first report on salivary IgA against sporulated oocyst antigens.

Despite the increasing interest and the potential use of antibodies (mainly IgG and IgA) in oral fluid

(saliva), in view of its advantage over serum as a medium for antibody detection due to non-invasive sampling, the use of saliva samples for community-based epidemiological surveys, and for clinical diagnosis is in part complex and restricted to certain conditions. A comprehensive and detailed review on the physiological, immunological and technical aspects of the salivary antibodies' capacity to reproduce reliably both mucosal and systemic immunity highlights the dimension of the complexity regarding this matter [31]. Our data illustrate in part such complexity and point to the importance of the age-range parameter for saliva sampling in population-based studies on *T. gondii* infections. The age of patients was the determining factor in our study for data interpretation on the relative contribution of both *T. gondii* cysts and oocyst transmission. In fact, age has been shown to be an important variable in studies of immunoglobulins in oral fluids in general [32–34] and this phenomenon can be in part due to the slow development of the salivary IgA system [32].

In this study, because there is no serological parameter in the gold standard category for Ig recognition of *T. gondii* sporozoite antigens, we assumed as infected with *T. gondii* those patients presenting positive serology to commercial conventional kits that detected antibodies (IgG and or IgM against *T. gondii*) in order to analyse and compare the serology and

salivary Ig recognition profile against *T. gondii* sporozoite antigens. Therefore, it is not possible to rule out the possibility of patients presenting as negative in conventional serology to *T. gondii*, but presenting as positive against TgERP, being infected with *T. gondii*. In fact, using the threefold s.d. criteria to establish positivity and negativity to TgERP in the present study we did not identify patients with such serological profile, namely negative to conventional serology and positive to TgERP. However, when our group used the same threefold s.d. criteria to evaluate serological IgG positivity against TgERP in a previous study, we identified patients presenting that profile [30]. We have proposed a possible explanation for that based on the vast genetic diversity of parasites in South America compared to the lower genetic diversity of parasites reported in North America and Europe [35]. Moreover, considering that commercial kits have been prepared with antigens derived from strains prevalent in North America and Europe there is in theory, a possibility of the existence of patients infected with parasites able to induce an immune response that will not recognize antigens present in commercial kits [30].

Based on threefold s.d. and ROC curve criteria that were employed to attempt to establish good cut-off values for the TgERP ELISA system in the present study it was possible to identify an age group that was shown to be suitable for studying the specific salivary antibodies against sporozoite antigens. That age group was 15–21 years, which, beyond presenting the highest values for sensitivity, specificity, and likelihood compared to all other age groups (Fig. 1), was in the prevalence curve (Fig. 2). The section of the curve where the slope is greatest is classically considered in epidemiology to be the point at which the incidence of transmission is highest. Furthermore, for that age group the best value was observed for Spearman's correlation (0.56, $P < 0.0001$) between sera IgG and saliva IgA against TgERP, showing that IgA from saliva against sporozoite antigens probably reflects the systemic IgG response against the same antigens in those aged 15–21 years. We did not evaluate salivary IgG nor serological IgA against TgERP in the present study due to scarcity of material (respectively saliva and sera) from some younger age groups. Nonetheless, it can be evaluated in further studies and will be of help to contribute to the better understanding of the mucosal, salivary glands, and systemic immune response dynamics against *T. gondii* sporozoite antigens in humans. The very close

prevalence rates identified using TgERP ELISAs by both cut-off criteria, namely 66.6% by threefold s.d. criteria or 66.7% by ROC curve criteria, and 68.7% using the conventional ELISA system, reinforces the importance of the 15–21 years age group for studying the mode of transmission of *T. gondii* infection. However, it is important to note that it is likely that the age where the slope is highest in prevalence curves for toxoplasmosis is the real point of importance and studies in other areas may demonstrate that this age range can vary. The meaning of the coincidence of prevalence rates in this age group using both TgERP and tachyzoite antigens remains to be investigated. Nonetheless, it has been demonstrated that IgG against TgERP declines 8 months after primary oocyst exposure in the context of infections occurring in outbreaks and that tachyzoite immune responses detected by conventional ELISA are concomitant during this period [18]. The coincident prevalence observed using both Ig against TgERP and against tachyzoites can be explained by the possibility of a recent infection by oocysts (at least within the time-frame of 8 months). This result emphasizes the importance of studying the mode of *T. gondii* infection transmission in this age group or alternatively in the section of the age prevalence curve where the highest slope is observed.

In this study, a very low frequency of IgM-positive individuals from all age groups was observed. However, low IgM prevalence is a common factor observed for individuals living in endemic areas (including the recently infected young) for parasitic diseases [36, 37].

In conclusion, we present evidence that salivary IgA against sporozoite antigens is important in studying cysts vs. oocyst-induced *T. gondii* infection in endemic settings. The major areas of importance are: (a) the highest scores (sensitivity sensibility and likelihood) compared to serological IgG for all age groups studied; (b) important and significant Spearman correlation value to systemic IgG ($r = 0.56$, $P < 0.0001$); and (c) its identification as an IgG revealing *T. gondii* prevalence at the point along the age prevalence curve where the slope is the highest (for ages 14–21 years in the current study) in coincidence with systemic IgG against TgERP and IgG against *T. gondii* tachyzoite antigens. We propose that other endemic areas should be studied with comparable parameters as those investigated in this study with a view to clarifying if the phenomenon we describe in this study can also be observed in other endemic settings for *T. gondii* infection. Such

replica studies would be very important for the better understanding of the relative epidemiological importance of different modes of infection with respect to horizontal transmission of *T. gondii*, which in a previous study [30], and in this study was >50% of the total prevalence when measured by antibodies (IgG from sera or IgA from saliva) against TgERP. Equally important in such population-based studies is the potential contribution they can provide towards the recommendation of using salivary IgA to investigate prevalence of toxoplasmosis, with regard to its advantage over serum as a medium for antibody detection due to its non-invasive nature. Furthermore, studies of this nature are of fundamental importance to define better and more effective primary prevention strategies to minimize *T. gondii* transmission mainly in endemic settings.

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DECLARATION OF INTEREST

None.

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