



Humoral response to viral vector COVID-19 vaccine in hemodialysis patients

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Background: The coronavirus disease 2019 (COVID-19) vaccine is not readily available in many countries where dosing interval is spaced more than ideal. Patients with chronic kidney disease, especially those on maintenance hemodialysis, have a tendency for a reduced immune response. This study was undertaken to demonstrate the distinct humoral immune response to the viral vector COVID-19 vaccine in patients with kidney failure receiving maintenance hemodialysis.

Methods: The study was carried out with two cohorts: 1) patients receiving maintenance hemodialysis and 2) healthcare workers from the same dialysis center as controls, each group with 72 subjects. Participants received a dose of Covishield ChAdOx1 nCoV-19 coronavirus vaccine. The humoral immunological response was determined using electrochemiluminescence immunoassay which quantitatively measures antibodies to the severe acute respiratory syndrome coronavirus 2 spike protein receptor-binding domain.

Results: All study subjects in the control group developed a humoral response (antibody titer of ≥ 0.8 U/mL), while only 64 of 72 in the dialysis group (88.9%) were responders. Age ($p = -0.234$, $p = 0.04$) and sodium level ($p = 0.237$, $p = 0.04$) correlated with low antibody titer in bivariate analysis. In multivariate analysis, only age (odds ratio, 1.10; 95% confidence interval, 1.01–1.22; $p = 0.045$) was associated with nonresponders.

Conclusion: Our study demonstrated a weak antibody response of hemodialysis patients to the viral vector COVID-19 vaccine. Older age was associated with nonresponders. Evaluation of both humoral and cellular immunity after the second vaccine dose and serial antibody titers can help determine the need for booster shots.

Keywords: Antibody formation, Secondary immunization, Chronic kidney failure, COVID-19, Hemodialysis

Introduction

The coronavirus disease 2019 (COVID-19) pandemic has severely affected countries worldwide. Vaccination of the majority of the population is an effective method to end the pandemic. However, vaccine availability remains unfeasible in many parts of the world, especially in developing

nations. Although in many countries, including India, the replication-defective viral vector vaccine, ChAdOx1 nCoV-19 (Oxford-AstraZeneca), was administered initially as two doses 4 weeks apart, it was later spaced to 12 to 16 weeks for better coverage of the population with at least a single dose and due to improved efficacy with the increased interval between doses [1]. The delay of the second dose

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might be appropriate for the general population; however, patients with chronic kidney disease (CKD), especially those receiving maintenance hemodialysis, tend to have a reduced immune response, as evidenced with the hepatitis B virus vaccine [2] and other vaccinations [3,4].

Furthermore, these patients suffer from high morbidity and mortality due to severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) [5,6] and are at high risk of contracting the infection due to continuous visits to medical facilities and exposure to other people, even during peak times of the COVID-19 pandemic [7]. This study was undertaken to demonstrate the distinct humoral immune response in patients with kidney failure receiving maintenance hemodialysis. The antibody titers against the SARS-CoV-2 spike (S) protein were evaluated in maintenance hemodialysis patients and compared with those in healthcare workers in the same center as controls.

Methods

Study design

Study participants

This study was conducted in our hemodialysis center with two cohorts: 1) patients receiving maintenance hemodialy-

sis and 2) healthcare workers from the same dialysis center as controls. Participants received a dose of Covishield Ch-AdOx1 nCoV-19 coronavirus vaccine (recombinant) developed by AstraZeneca (Cambridge, United Kingdom), and manufactured by Serum Institute of India Pvt Ltd. (Pune, India) from COVID-19 vaccination centers deputed by the government. All patients received the standard dose (0.5 mL) of the vaccine containing 5×10^{10} viral particles [8]. A total of 141 patients was receiving maintenance hemodialysis in our center. At the time of the study, 20 patients had been vaccinated for approximately 6 to 7 weeks and were waiting for their scheduled second dose at 12 to 16 weeks, and 72 patients had been vaccinated 4 weeks prior. Twenty-one patients had a history of COVID-19 infection, 22 were not vaccinated at the time of the study, and six refused to get vaccinated. Participant selection is shown in Fig. 1.

Seventy-two patients included in the study were receiving maintenance hemodialysis in our center and had been vaccinated with the first dose 4 weeks prior. Subjects who were younger than 18 years, had a history of COVID-19 infection, were vaccinated previously at 6 weeks or more, or had an acute illness at the time of the study were excluded. In addition, 72 healthcare workers from the same dialysis center were enrolled as controls for the study.

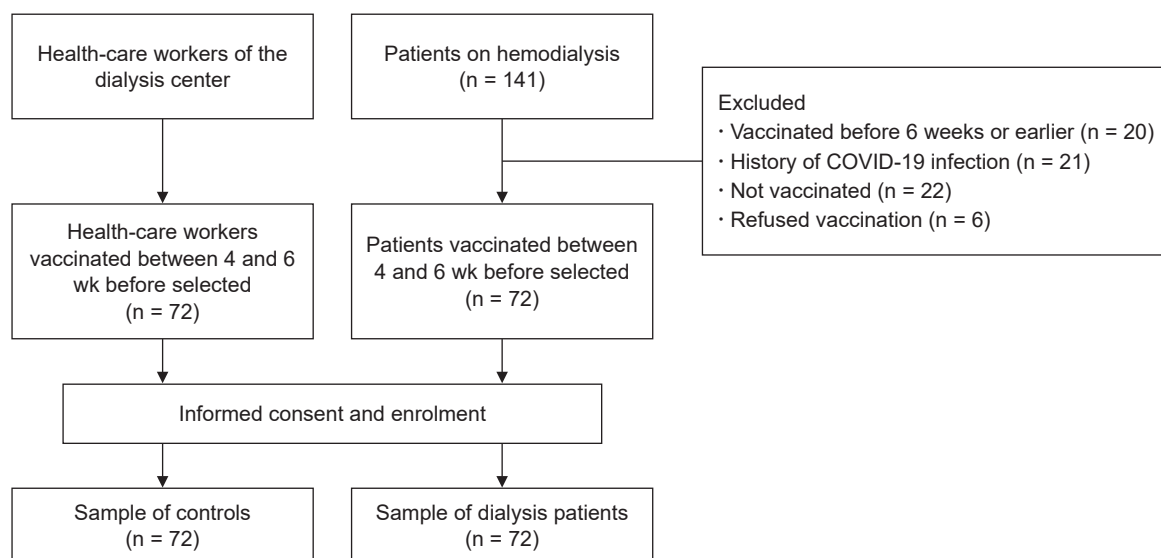


Figure 1. Study participant selection. All participants, including the controls, provided blood samples in the 5th week after vaccination. COVID-19, coronavirus disease 2019.

Antibody measurement

The humoral immunological response was determined by measuring antibody titers using electrochemiluminescence immunoassay with Elecsys Anti-SARS-CoV-2 S on a Cobas 6000 analyzer e 601 module (Roche Diagnostics, Rotkreuz, Swiss) [9]. This test is an immunoassay for *in vitro* quantitative determination of antibodies (including immunoglobulin G [IgG]) to the SARS-CoV-2 S protein receptor-binding domain (RBD) in human serum and plasma. In previous studies, although full-length and fragments of S protein induced specific antibodies with neutralizing activity, the RBD was suggested as a major target for eliciting highly potent neutralizing antibodies with protective efficacy [10]. This test has a linear range of 0.4 to 250 U/mL. Titers of <0.8 U/mL were considered nonreactive and ≥ 0.8 U/mL reactive.

Data and sample collection

Informed consent was obtained from the participants including controls for drawing blood samples and participation in the study. Demographic details, medical history, and adverse events following vaccination of the patients and controls were collected with a proforma. Venous blood samples were taken from the patients during their dialysis visits before the initiation of the session.

Although the SARS-CoV-2 antibodies against S antigen and its subunits can be detected as early as 1 to 3 weeks after an infection or vaccination, the test used attains maximum sensitivity by the 5th week [9]. Therefore, all samples were collected between 28 and 35 days after the first dose of the vaccine for both patients and controls. Body mass index (BMI) was defined as dry weight in kilograms divided by the square of the height in meters.

Statistical analyses

Statistical analysis was performed using the IBM SPSS ver. 23.0 (IBM Corp., Armonk, NY, USA). All categorical variables were presented as frequency and percentage and continuous variables as either mean (standard deviation) or median (interquartile range). The shape of the data distribution of antibody titers was assessed using the histogram/Shapiro-Wilk test. The chi-square test and Fisher exact test were used to determine the difference between the dialysis and control groups based on proportion values,

and an independent sample t test or Mann-Whitney U test was used to determine the difference among the groups based on continuous values. Spearman correlation was used to determine the relationship between antibody titers and demographic and laboratory parameters. Multivariate logistic regression was used to determine the most important predictors of nonreactive antibody titers. BMI was classified based on Asian BMI classification guidelines. A p-value of <0.05 was considered statistically significant.

Ethics statement

The Institutional Ethics Committee in our hospital requires approval only for original research with trial/experimental interventions (procedure/drugs) administered patients (approval No. DHR Reg.No.EC/NEW/INST/2020/484). These patients were not treated with any trial/experimental therapy. All procedures followed the guidance of the Declaration of Helsinki. Written informed consent was obtained from the patients for participation in the study and for performing investigations.

Results

The dialysis and control groups each included 72 participants. The majority of patients in the dialysis group were males (80.6%) and in the control group females (66.7%). The average age in the dialysis group was higher (46.14 ± 8.89 years) than in the control group (40.35 ± 6.65 years). Mean BMI was lower in the dialysis group (20.87 ± 4.26 kg/m²).

The majority of patients (n = 48) had hypertension (66.7%), 20 (27.8%) had diabetes mellitus, four (5.6%) had dilated cardiomyopathy, four (5.6%) had a history of cerebrovascular accident, and two (2.8%) had hypothyroidism. Among patients in the dialysis group, 36 (50.0%) had hepatitis C virus (HCV) infection at the time of the study, and 26 (36.1%) were receiving treatment. On average, the dialysis duration was 8.65 ± 6.68 months; four patients (5.6%) had permanent catheters, and the remaining patients had arteriovenous fistula for hemodialysis access. None of the study participants were on or received immunosuppressive agents at least 3 months prior to the study.

Diabetic kidney disease was the most common native kidney disease, observed in 18 patients (25.0%), followed by chronic glomerulonephritis in 14 (19.4%), chronic in-

terstitial nephritis in six (8.3%), obstructive uropathy in five (6.9%), hypertensive nephrosclerosis in three (4.2%), and autosomal dominant polycystic kidney disease in two (2.8%). Native kidney disease was not known in 24 (33.3%) patients because many patients in India present with advanced stages of CKD. Among control subjects, four (5.6%) had diabetes mellitus, and two (2.8%) had hypertension. Demographic details, clinical characteristics, and laboratory results of the study subjects are shown in [Table 1](#).

Lower humoral response in the dialysis group

All study subjects in the control group developed a positive humoral response defined based on a value of ≥ 0.8 U/mL compared with only 64 of 72 (88.9%) in the dialysis group ([Table 1](#)). All eight patients with nonreactive antibody titers were older (greater than the mean age of the dialysis group), four were HCV infected, and two had diabetes mellitus.

Adverse events

Fever and myalgia were significantly more common in the control subjects ([Table 2](#)). Other adverse events such as pain at the local site, pruritus, or abdominal pain were observed in only a few patients. Serious adverse events were not observed in any study subjects.

Factors affecting antibody titers

In the dialysis group, age negatively correlated with antibody titer ([Table 3](#)); titer was lower with increasing age (Spearman correlation $\rho = -0.234$, $p = 0.04$). Dialysis dose (Kt/V) did not correlate with antibody titer ($\rho = 0.085$, $p = 0.75$). Similarly, hemoglobin ($\rho = 0.003$, $p = 0.98$) and albumin levels ($\rho = 0.147$, $p = 0.22$) did not correlate with titer. Bivariate analysis of laboratory results, sex, age, and BMI with antibody titers is shown in [Table 3](#).

In the dialysis group, age and sodium level correlated with antibody titer in bivariate analysis; titers were lower in patients with higher age (Spearman correlation $\rho = -0.234$, $p = 0.048$) and lower sodium level ($\rho = 0.237$, $p = 0.045$). Statistically significant correlation was not observed between antibody titers and other lab parameters.

Fisher exact test ($p > 0.05$) showed no significant difference in the proportion of reactive titer group subjects and

nonreactive titer group subjects regarding HCV infection, diabetes mellitus, BMI, dialysis duration, lymphocyte count, and adverse events such as fever and myalgia ([Table 4](#)).

A multivariate logistic regression model was used to construct the statistical model to predict the risk of nonreactive antibody titers using the most significant two factors correlating with low antibody titers identified based on bivariate analysis, age, and sodium levels ([Table 5](#)). Only age ($p = 0.045$) was a statistically significant independent variable associated with nonreactive antibody titers.

Discussion

The compromised immunity in hemodialysis patients is evident by a poor immune response to vaccinations such as hepatitis B [1], pneumococcus [2], or influenza [3]. In the present study, the humoral immune response to COVID-19 vaccination (Covishield) in hemodialysis patients compared with a cohort of healthcare workers in the same center was investigated.

Covishield is a recombinant, monovalent vaccine composed of a replication-deficient chimpanzee adenovirus (ChAdOx1) vector that encodes the S glycoprotein of SARS-CoV-2. The antigen is expressed locally, stimulating neutralizing antibodies and cellular immune responses. The initial schedule of Covishield ChAdOx1 nCoV-19 coronavirus vaccine (recombinant) was two doses of 0.5 mL each, 4 weeks apart, which was later spaced to 12 to 16 weeks for better coverage of a larger population and due to improved efficacy in clinical trials with increased spacing. Due to a short supply of vaccines, a policy of initially vaccinating a larger cohort with a single dose might provide better overall population protection than vaccinating half the number of individuals with two doses in the short term [1].

In our study, the antibody titers were significantly lower in the dialysis group, including eight patients (11.1%) with absent antibody titer (< 0.8 U/mL). This weaker antibody response was comparable with results obtained in studies with messenger RNA vaccines [11,12], in which approximately 10% of dialysis patients did not have an antibody response. Simon et al. [13] studied vaccine response in 86 dialysis patients and found 17 (19.8%) were nonresponders even after the second dose, which was almost double than reported in our study. The study by Simon et al. [13] excluded patients who tested positive for antibodies before

Table 1. Demographic details, clinical characteristics, and laboratory parameters of the study subjects

Factor	Dialysis group	Control group
Demographics		
No. of patients	72	72
Sex		
Male	58 (80.6)	24 (33.3)
Female	14 (19.4)	48 (66.7)
Age (yr)	46.14 ± 8.89	40.35 ± 6.65
Clinical characteristics		
Body mass index (kg/m ²)	20.87 ± 4.26	23.12 ± 3.35
Diabetes mellitus	20 (27.8)	4 (5.6)
Hypertension	48 (66.7)	2 (2.8)
Dilated cardiomyopathy	4 (5.6)	-
Cerebrovascular accident	4 (5.6)	-
Hypothyroidism	2 (2.8)	-
HCV infection	36 (50.0)	-
Blood pressure (mmHg)		
Systolic blood pressure	145.81 ± 15.10	127.56 ± 6.20
Diastolic blood pressure	94.77 ± 10.63	79.12 ± 4.21
Native kidney disease		
Diabetic kidney disease	18 (25.0)	-
Chronic glomerulonephritis	14 (19.4)	-
Chronic interstitial nephritis	6 (8.3)	-
Obstructive uropathy	5 (6.9)	-
Hypertensive nephrosclerosis	3 (4.2)	-
ADPKD	2 (2.8)	-
Unknown	24 (33.3)	-
Dialysis access		
Arteriovenous fistula	68 (94.4)	-
Permanent catheter	4 (5.6)	-
Dialysis adequacy (Kt/V)	1.16 ± 0.17	-
Dialysis duration (mo)	8.65 ± 6.68	-
Laboratory parameters		
Hemoglobin (g/dL)	8.07 ± 1.33	12.69 ± 1.27
White blood cell count (cell/mm ³)	6,944.4 ± 2,770.4	8,204.2 ± 1,991.0
Neutrophils (%)	67.85 ± 10.54	58.11 ± 9.13
Lymphocytes (%)	22.49 ± 14.16	31.49 ± 6.72
Neutrophil-to-lymphocyte ratio	3 (2.09–5.26)	1.79 (1.38–2.57)
Creatinine (mg/dL)	7.15 ± 3.29	0.88 ± 0.17
Sodium (mEq/L)	133.5 ± 5.2	136.5 ± 4.0
Potassium (mEq/L)	4.49 ± 1.05	4.27 ± 0.36
Albumin (g/dL)	3.03 ± 0.70	4.05 ± 0.37
Antibody titer (U/mL)		
Reactive titer, ≥0.8	64 (88.9)	72 (100)
Nonreactive titer, <0.8	8 (11.1)	0 (0)

Data are expressed as number only, number (%), mean ± standard deviation, or median (interquartile range).

ADPKD, autosomal dominant polycystic kidney disease; HCV, hepatitis C virus.

Table 2. Adverse events due to vaccination in dialysis and control groups

Adverse event	Dialysis group (n = 72)	Control group (n = 72)
Fever	12 (16.7)	32 (44.4)
Myalgia	6 (8.3)	32 (44.4)
Pruritus	2 (2.8)	0 (0)
Pain at the local site	4 (5.6)	0 (0)
Abdominal pain	0 (0)	2 (2.8)

Data are expressed as number (%).

Table 3. Spearman rank correlation between pathological parameters, sex, age, BMI, and antibody titers

Variable	Dialysis group	
	ρ	p-value
Sex	-0.131	0.27
Age	-0.234	0.05
BMI	0.023	0.85
Hemoglobin	0.003	0.98
White blood cell count	0.144	0.23
Neutrophils	0.016	0.89
Lymphocytes	-0.117	0.33
NLR	0.123	0.30
Creatinine	0.057	0.64
Sodium	0.237	0.05
Albumin	0.147	0.22
Dialysis dose (Kt/V)	0.085	0.75

BMI, body mass index; NLR, neutrophil-to-lymphocyte ratio; ρ , point-biserial.

vaccination, thereby eliminating subjects who developed antibodies due to subclinical infections. In contrast, all participants in the control group developed a humoral response, with the majority (89.9%) having titer of ≥ 250 U/mL. This comparison clearly demonstrated a weaker antibody response in dialysis patients, indicating this group was susceptible to higher morbidity and mortality following infection.

Overall, the age of dialysis subjects enrolled in the present study was relatively younger (46.14 ± 8.89 years) compared with that of CKD patients. Most of the older patients in our center were vaccinated much earlier than 4 weeks because only older individuals were eligible for the initial round of vaccinations in India, hence, were not eligible for the study. In addition, fewer older patients were receiving hemodialysis in the dialysis unit because many older end-stage renal disease patients refuse dialysis, and even those scheduled for maintenance hemodialysis typically do not

Table 4. Comparison of antibody titer levels in HCV infection, diabetes mellitus, and BMI

Variable	Responder	Nonresponder	p-value
HCV infection			
Positive	32 (50.0)	4 (50.0)	>0.999
Negative	32 (50.0)	4 (50.0)	
Diabetes mellitus			
Positive	18 (28.1)	2 (25.0)	>0.999
Negative	46 (71.9)	6 (75.0)	
BMI ^a			
Underweight	22 (16.2)	2 (25.0)	0.05
Normal	57 (41.9)	6 (75.0)	
Overweight	47 (34.6)	0 (0)	
Obese	10 (7.4)	0 (0)	
Dialysis duration (mo)			
≤ 5	34 (53.1)	4 (50.0)	>0.999
> 5	30 (46.9)	4 (50.0)	
Lymphocytes (%)			
≤ 27	72 (52.9)	6 (75.0)	0.29
> 27	64 (47.1)	2 (25.0)	
Significant adverse event			
Fever			
Present	44 (32.4)	0 (0)	0.11
Absent	92 (67.6)	8 (100)	
Myalgia			
Present	38 (27.9)	0 (0)	0.11
Absent	98 (72.1)	8 (100)	

Data are expressed as number (%).

BMI, body mass index; HCV, hepatitis C virus.

^aWorld Health Organization's Asian BMI classification.

Table 5. Factors associated with nonreactive antibody titers among the dialysis patients based on multivariate logistic regression analysis

Predictor	OR (95% CI)	p-value
Age	1.11 (1.01-1.22)	0.05
Sodium	0.76 (0.71-1.08)	0.06

Dependent variable is antibody titers (1, nonreactive; 0, reactive).

CI, confidence interval; OR, odds ratio.

receive dialysis regularly and are lost to follow-up due to economic burden on the family.

Age, sex, and BMI were distributed unequally between the control and dialysis groups; the majority of subjects were female in the control group and male in the dialysis group. Furthermore, the patients in the control group were younger on average and had higher BMI than subjects in

the dialysis group.

A multivariate analysis was performed to account these confounding factors. Among the variables, only age affected the antibody titers with statistical significance, which was in agreement with various studies [13–16] showing similar relatively poor antibody responses in older age groups. This reinforces the importance of the age factor in the humoral response.

Bivariate analysis showed sodium to correlate with low antibody titer ($\rho = -0.237$, $p = 0.04$) but was not significant in multivariate analysis including age and sodium level as variables. The correlation in bivariate analysis indicated the probable presence of cofounders. Hemodilution due to volume overload could have caused pseudo-low antibody titers because samples were collected before the dialysis session. Most patients in the study were presumed to have achieved dry weight because only subjects receiving hemodialysis for more than 2 months were enrolled in the study. However, volume status was assessed based on clinical examination, and objective methods such as body composition monitors were not used. Furthermore, hyponatremia in dialysis patients can be due to factors other than fluid overload, such as malnutrition and inflammation [17,18]. A detailed assessment is needed of nutritional status and inflammatory markers not tested in the present study, which are study's limitations.

A statistically significant association was not observed between antibody levels and sex, BMI, dialysis duration, dialysis dose, hemoglobin, albumin, total count, lymphocyte count, urea, liver function tests, HCV infection, diabetes mellitus, and adverse events due to vaccines. Grupper et al. [15] demonstrated a correlation between lymphocyte count and the humoral response given their role in adaptive immunity, which was not observed here; however, the authors also did not find an association with BMI, dialysis duration, dialysis dose, or albumin level.

Although the seroconversion rate may be considered relatively good for dialysis patients, it is subpar when compared to normal subjects. These patients are exposed repeatedly to other patients and healthcare workers due to their regular visits for dialysis and have high morbidity and mortality due to comorbidities.

Baseline antibody titers were not measured in the study participants. Although previously infected patients were excluded from our study, seroconversion due to subclinical

infections or minimal symptoms might not have been evident. However, this confounding factor was at least partly addressed by selecting control subjects from the same hemodialysis center.

Neutralization assays such as the plaque reduction neutralization test (PRNT) are the gold standard for serological testing and determining immune protection against COVID-19 [19]. PRNT has several limitations rendering it unsuitable for large-scale studies. The assay used in the present study, Elecsys Anti-SARS-CoV-2 S, is not a neutralization test but has a good correlation with pseudo-virus neutralization assay (safer and more versatile than neutralization assay), with a positive agreement of 92.3% (95% confidence interval [CI], 63.97–99.81) [20]. This assay shows a positive predictive agreement of 96.6% (95% CI, 93.35–98.51) and a negative predictive agreement of 99.98% (95% CI, 99.91–100) [21].

SARS-CoV-2 antibodies against S protein and its subunits can be detected within 1 to 3 weeks after infection [22,23]. However, we tested in the 5th week after vaccination because our test attains maximum sensitivity by that period [9].

The upper limit of detection of Elecsys Anti-SARS-CoV-2 S assay is 250 U/mL, a limitation of our study. A higher upper limit of the assay would be preferable because a better difference in adaptive humoral response between dialysis and the control groups could have been identified. Similarly, the complete humoral response profile after the second dose of the COVID-19 vaccine will provide relevant information but will require a larger sample and control cohorts. This was a single-center study with a limited number of patients, and complete humoral response was not the focus; however, differences were apparent.

Lack of follow-up of the patient titers is another limitation of our study. Even in normal subjects with adequate antibody responses, titers have been shown to decline over time [24]. In some studies, SARS-CoV-2 IgG titers decline substantially in dialysis patients within 3 months after diagnosis [25]. Therefore, these patients should be followed up with titers to monitor the level of immunity, which can help determine the need and timing of booster doses.

In addition to antibody production in response to SARS-CoV-2 infection, host cellular immunity plays an essential role in impeding virus replication and expansion at various stages of COVID-19 disease [26]. CD4 and CD8 T

cells recognize multiple regions of the N protein of SARS-CoV-2 in previously exposed patients. T cells showed strong cross-reactivity to the N protein of SARS-CoV-2 in patients who recovered from SARS (the disease associated with SARS-CoV infection) 17 years after the outbreak in 2003 [27]. Furthermore, patients who had no detectable antibody at 6 months after COVID-19 infection were found to have SARS-CoV-2 antigen-specific T-cell responses [28]. Our study did not attempt to determine the T-cell immune response, which might contribute to protection from COVID-19.

In conclusion, our study results demonstrated a weak antibody response of hemodialysis patients to the viral vector COVID-19 vaccine, and older age was associated with nonresponders. Evaluation of both humoral and cellular immunity after the second dose of COVID-19 vaccine and serial follow-up of antibody titers can help determine the need for booster shots.

Conflicts of interest

All authors have no conflicts of interest to declare.

Authors' contributions

Conceptualization: AR, NR

Data curation: NR, RD

Formal analysis: MR, RD

Investigation: NR, AP

Methodology: AR, NR

Project administration: NR, JA

Visualization: NR

Writing-original draft: NR

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