



HHS Public Access

Author manuscript

Immunol Rev. Author manuscript; available in PMC 2023 July 01.

Published in final edited form as:

Immunol Rev. 2022 July ; 308(1): 123–148. doi:10.1111/imr.13078.

Role of Hormones in the Pregnancy and Sex-Specific Outcomes to Infections with Respiratory Viruses

Orlando Cervantes^{1,2,*}, Irene Cruz Talavera^{1,3}, Emma Every⁴, Brahm Coler^{2,5}, Miranda Li^{2,6}, Amanda Li^{2,7}, Hanning Li², Kristina Adams Waldorf^{1,2,*}

¹Department of Global Health, University of Washington, Seattle, Washington, United States of America

²Department of Obstetrics and Gynecology, University of Washington, Seattle, Washington, United States of America

³Vaccine and Infectious Disease Division, Fred Hutchinson Cancer Research Center, Seattle, Washington, United States of America

⁴University of Washington School of Medicine, Spokane, Washington, United States of America

⁵Elson S. Floyd College of Medicine, Washington State University, Spokane, Washington, United States of America

⁶Department of Biological Sciences, Columbia University, New York City, New York, United States of America

⁷Case Western Reserve, Cleveland, Ohio, United States of America

Summary

Pregnant women infected with pathogenic respiratory viruses, like influenza A viruses (IAV) and coronaviruses, are at higher risk for mortality, hospitalization, preterm birth and stillbirth. Several factors are likely to contribute to the susceptibility of pregnant individuals to severe lung disease including changes in pulmonary physiology, immune defenses and effector functions of some immune cells. Pregnancy is also a physiologic state characterized by higher levels of multiple hormones that may impact the effector functions of immune cells, such as progesterone, estrogen, human chorionic gonadotropin, prolactin and relaxin. Each of these hormones acts to support a tolerogenic immune state of pregnancy, which helps prevent fetal rejection, but may also contribute to an impaired antiviral response. In this review, we address the unique role of adaptive and innate immune cells in the control of pathogenic respiratory viruses and how pregnancy and specific hormones can impact their effector actions. We highlight viruses with sex-specific differences in infection outcomes and why pregnancy hormones may contribute to fetal protection but aid the virus at the expense of the mother's health.

***Co-Corresponding Authors:** Orlando Cervantes, BS and Kristina Adams Waldorf, MD; University of Washington, Box 358070, 750 Republican Street, Seattle, WA, 98109; Telephone: 206-616-5258; Fax: 206-543-3915; adamsk@uw.edu and ocervant@uw.edu. Note that both co-corresponding authors have the same physical address and telephone numbers.

Conflict of Interest:

There is no conflict of Interest to declare.

Keywords

pregnancy; virus; influenza; COVID-19; pneumonia; estrogen; progesterone

Introduction

Although essential to human survival, pregnancy remains one of the most poorly understood biological and immunological events. High mortality rates of pregnant individuals during the coronavirus disease 2019 (COVID-19) pandemic have underscored how changes in immune function during pregnancy can leave young individuals extremely vulnerable to viral infections.^{1–6} Pregnant people are also vulnerable to pandemic influenza A viruses (IAV) with increased rates of hospitalization, mortality, preterm birth, stillbirth and neonatal death.^{7–17} How sex hormones impact immune function is understudied, but of paramount importance to understanding pregnancy- and sex-specific differences in respiratory infection outcomes. Pregnancy represents an extreme hormone milieu. Compared to the non-pregnant state, a pregnant individual will experience large increases in corticosteroids (20-fold)¹⁸, progesterone (6–8-fold)¹⁹, estrogen (estriol: 1,000-fold; estradiol/estrone: 6–100-fold)^{20,21}, prolactin (10–20-fold)²², relaxin (>10-fold)²³ and human chorionic gonadotropin (hCG, only made during pregnancy). A woman will produce more estrogen in a single pregnancy than during the rest of her entire lifetime. This profound hormonal environment shapes immune cell function and is instrumental in driving both pregnancy- and sex-specific differences in the outcome of respiratory viral infections.

In pregnancy, many sex hormones function to promote immune tolerance to enable the mother to tolerate a semi-allogeneic fetus without rejection.^{24–27} However, this tolerogenic state comes at the expense of antiviral immunity, as pregnant women are more vulnerable to many viral respiratory pathogens versus non-pregnant women. In this review, we explore the intersection between hormones, immunity, and pregnancy to provide insight into both pregnancy- and sex-specific differences in the outcome to respiratory virus infection.^{1–17} First, we address the known pregnancy- and sex-specific differences in the outcome to respiratory virus infections, which may be related in part to differences in sex hormones. Secondly, we address the impact of hormonal changes on distinct immune cell subsets focusing on adaptive immunity (T and B cell subsets) with a more limited discussion of innate immunity. Finally, we discuss how sex steroids can impair adaptive immune function to specific respiratory viruses, focusing on IAV, respiratory syncytial virus (RSV) and the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Throughout the review, we discuss hypotheses linking hormone-mediated changes during pregnancy and the vulnerability of pregnant people to severe disease from respiratory viral infections.

Pregnancy- and Sex-Specific Differences in Outcomes to Respiratory Virus Infection

Many respiratory viral infections are associated with greater mortality and more severe lung disease in pregnant people. In this section, we focus on the pregnancy- and sex-specific differences in outcomes to influenza A viruses and highly pathogenic coronaviruses, for

which there is the most published data. We acknowledge that pregnancy-, sex- and gender-specific differences in health and disease outcomes are the result of numerous factors including sex hormones, genetics, sex-linkage of immune response genes, behaviors (e.g., hand washing), lifestyle, nutritional habits, occupation and perceived stress and pain; these factors may also modify epigenetics that modulates gene expression and biological phenotypes.²⁸ Although the foundation for these differences may be multi-factorial, they establish a framework for focusing on the impact of sex hormones on diverse immune cell populations.

Influenza A Viral Infections: Pregnancy- and Sex-Specific Differences in Outcomes

IAV is an orthomyxovirus, which evades adaptive immunity by rapidly acquiring mutations in its surface-proteins, neuraminidase (NA) and hemagglutinin (HA).²⁹ Changes in mutations can result in antigenic drift, which represent minor changes to the NA and HA proteins that may result in the need to annually revise the seasonal influenza vaccine. Occasionally, antigenic shift occurs, which describes larger changes in the NA and HA proteins occurring through genomic reassortment that can contribute to influenza pandemics. In all IAV pandemics since 1918, pregnant people have been described as a vulnerable population to IAV infection with higher rates of mortality, hospitalization, preterm birth and stillbirth.^{7–17}

The first reports of pregnancy as a high-risk condition for severe IAV disease occurred during the 1918–1919 IAV pandemic ('Spanish flu'). An analysis of death certificates in London found that the mortality rate for all women (15–49 years) was 4.9/1,000, but in pregnancy was 5.3–5.7/1,000.³⁰ As many pregnant women who died did not have their pregnancy noted on their death certificate, this is likely an underestimate. Similarly, in the U.S., reports of pregnancy mortality obtained from a questionnaire of obstetricians surveying 1,350 pregnancies found that 678 of these patients developed pneumonia (50%), 365 died (27%) and there were high rates of stillbirths.³¹ Reports of higher mortality, hospitalization, preterm birth and stillbirth also occurred during the 1957 pandemic ('Asian flu') and the more recent 2009 H1N1 pandemic ('swine flu').^{7,8,11,13–17,28,32–34} The postpartum period after delivery represents a particularly high-risk time for mortality due to IAV.³⁵ This is a time of rapid physiological changes, fluid shifts and a precipitous drop in sex hormones. In a systematic review of more than 610,000 participants identified the women less than four weeks postpartum as one of the few risk factors associated with a significantly increased risk of death from pandemic influenza [odds ratio: 4.43, 95% Confidence Interval (C.I.): 1.24–15.81].³⁵ The impact of changes in sex hormones and other physiological changes on the immunity and health of postpartum women is unknown.

Sex-specific differences in the outcomes to IAV infections have been observed throughout the last pandemics. Young adult women typically have a higher hospitalization rates and mortality to pandemic influenza and avian influenza (H5N1).^{36–40} However, at the extremes of age, men have been reported to have higher IAV-mortality and morbidity. Japanese males younger than 20 and older than 80 were reported to have higher influenza morbidity during the H1N1 2009 pandemic than Japanese females.⁴¹ These sex difference in IAV infection outcomes also translate to the immune response to IAV vaccination. In reproductive age

adults, women tend to develop higher antibody titers than men to the seasonal IAV vaccine, which has been attributed to higher estrogen levels and X-linked immune responses genes (e.g., toll-like receptor 7).^{42,43 44} In summary, sex-specific differences are age dependent and are reflected in both the IAV-associated mortality and vaccination efficacy.

Coronavirus Infections: Pregnancy- and Sex-Specific Differences in Outcomes

Although pregnant people tend to be young and healthy, they are highly vulnerable to severe COVID-19 from an infection with the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), the virus that causes COVID-19.^{1,5,45-49} Although pregnant individuals were identified early in the IAV H1N1 2009 pandemic as a high-risk and vulnerable group in the U.S.^{11,13}, this was not recognized for the first, critical eight months of the COVID-19 pandemic. A broader view of the immunologic state of pregnant individuals, which is shaped in part by sex hormones, would suggest that pregnancy is a state that is likely to be more susceptible to viral infections. Indeed, pregnant individuals have a prolonged COVID-19 symptomatic course in comparison to reported outcomes in young adults; pregnant individuals reported 37 days (median time) until symptom resolution and 25% reported symptoms for more than 8 weeks.⁵⁰ In June of 2020, a population-based study by the U.S. Centers for Disease Control (CDC) found that pregnant patients with SARS-CoV-2 infections were at higher risk for hospitalization, mechanical ventilation and ICU admission, but mortality rates were similar between pregnant and non-pregnant reproductive age women (0.2%).⁵¹ Subsequently, another CDC study including more cases found an increased risk of mortality among pregnant women versus non-pregnant women with SARS-CoV-2⁵², prompting the CDC to revise public health guidance and indicate that pregnant women are at risk for severe COVID-19 disease.⁴⁵ Other population-based studies support that pregnancy is a vulnerable state for SARS-CoV-2 infection and linked to adverse maternal, pregnancy and neonatal outcomes.^{4,6,49,53}

A male bias in COVID-19 mortality and intensive care unit admission has been reported in a meta-analysis of more than 3 million global cases of COVID-19.⁵⁴ Although there was no difference in the proportion of females and males with COVID-19 disease, males had a 2.8-fold higher odds of requiring intensive care and 1.4-fold higher odds of death. To understand how sex differences in COVID-19 outcomes might be explained immunologically, longitudinal blood samples from hospitalized men and women were studied to evaluate immunophenotypes, cytokines and SARS-CoV-2 viral load.⁵⁵ In males, a greater plasma IL-8 and IL-18 level and higher frequencies of activated non-classical monocytes were observed compared to females. However, females had greater T cell activation than males. A patient's age was correlated negatively with a poor T cell response associated with adverse outcomes in males, but not females. However, higher levels of cytokines were linked to worse disease in females, but not in males. Whether these sex-specific differences in immunologic correlates of disease are also linked to sex hormones is unknown.

The Changing Hormonal Landscape Across Pregnancy

The receptors for pregnancy hormones can be found on most immune cells, enabling immunomodulation to be tightly coupled to reproduction and pregnancy.⁵⁶ Dramatic changes to the hormonal milieu during pregnancy are designed to enhance survival and tolerance of the semi-allogeneic fetus. One of the earliest physiologic changes in pregnancy is the rise in the hormone hCG, which acts to quickly promote an immunologic state conducive to fetal survival and growth. hCG is produced initially by the blastocyst and begins to rise at the time of embryo implantation, doubles approximately every 2–3 days reaching peak levels in the first trimester followed by a gradual decline until term. Relaxin, a hormone that follows a similar trajectory to hCG peaks in the first trimester and then slowly declines until term. In contrast, there is a gradual rise in estrogen (90% is estriol), progesterone and prolactin into the third trimester. Progesterone plays two critical roles in pregnancy by preparing the endometrial lining for implantation of the embryo and maintaining uterine quiescence until the time of birth. Progesterone is initially produced by the ovaries and then taken over by the placenta after 10 weeks gestation. During pregnancy, the production of progesterone gradually increases from non-pregnant levels of 1–50 nmol/L (depending on menstrual cycle phase) to serum concentrations between 175 and 811 nmol/L in the third trimester of pregnancy.⁵⁷ Although variation in some hormone levels have been linked to the timing of labor onset (i.e., preterm birth or post-term birth)^{58,59}, whether pregnant individuals at the extremes of sex hormone production might be susceptible to viral infections due to differences in immunomodulation are unknown. The estradiol level significantly increases during pregnancy ranging from 110 to 440 pmol/L during the follicular phase, up to 1800 pmol/L during the first trimester, up to 75,000 pmol/L during the second trimester, and up to 136,000 pmol/L during the third trimester.⁶⁰ In a murine model, the increased morbidity due to influenza infection was shown to correlate with levels of circulating estrogen, which are highest in the third trimester and correlate with the period of greatest disease severity in pregnant women.⁶¹

T cells: Intersection of Hormones, Pregnancy and Immune Function in Distinct T Cell Subsets

T cells provide cross-reactive cellular immunity to various pathogens via recognition of conserved antigens presented by antigen-presenting cells. CD4+ effector T cells, also known as T helper cells, play a major role in mediating cellular immunity and also suppressing immune functions of some cells in their environment to limit tissue damage. Helper T cells are classified into several distinct subsets, only some of which are covered in this review that have key roles in pregnancy tolerance, viral respiratory infections and are affected by sex hormones: Treg, Th17, Th1, and Th2. CD4- and CD8- $\gamma\delta$ T cells are another key cell type that plays important roles in immune protection in mucosal tissues. These T cell subsets are distinguished by their master transcription factors and respective cytokine secretomes.^{62,63} The balance among these subsets is of critical importance to favor fetal tolerance, which requires a modulation of maternal immunity in the periphery and at the maternal-fetal interface. However, a shift in the balance of these subsets, driven in part by sex hormones, may be detrimental to clearing viral infections. Here, we synthesize literature

on the hormonal impact on immunity in pregnancy and how this secondarily may impair antiviral immunity.

Regulatory T Cells

Role of CD4+FoxP3+ Regulatory T cells in Immunity and Pregnancy

Regulatory T cells (Tregs) are essential for self-tolerance, prevention of autoimmunity, regulation of inflammatory immune responses, and for the establishment and maintenance of pregnancy (Fig. 1). Regulatory T cells (Tregs) are a subset of CD4⁺ T cells characterized by their expression of the transcription factor forkhead box P3 (FoxP3) and their ability to regulate inflammatory responses, establish self-tolerance, and suppress effector T cells and other immune cells.^{64,65} FoxP3 is essential for Treg's immunosuppressive function and is the master regulator of Treg development, function, and differentiation.^{66,67} Tregs are critical immune mediators of tolerance, as patients suffering from fatal immune dysregulation polyendocrinopathy enteropathy X-linked syndrome (IPEX) and scurfy mice lacking functional expression of FoxP3 suffer from severe autoimmunity and dysregulated effector T cell responses.^{68,69} Tregs have been most extensively studied in the thymus where they develop, and in peripheral lymphoid organs. However, there is a growing body of work characterizing phenotypically distinct tissue Tregs with unique roles in non-lymphoid tissue compartments such as visceral adipose tissue, skin, muscle, intestines, lungs, and genitourinary tract.⁷⁰⁻⁷⁴ There are two main subsets of Tregs: natural Helios⁺ NRP1⁺ Tregs (nTreg) develop in the thymus and primarily involved in tolerance to self and environmental antigens, whereas induced Helios⁻ Tregs (iTreg) arise in periphery from CD4⁺ FoxP3⁻ T cells in response to high inflammation or after antigen-exposure, often in the context of infection.⁷⁵⁻⁷⁸ The differentiation of iTreg is dependent on many factors, including the strength of TCR signaling, signals from APCs, inflammatory responses, and the local cytokine environment.^{75-77,79} Though distinguishing these subsets *in vivo* and *ex vivo* has proven difficult, as knowing which markers should be used to differentiate them has been unclear.⁸⁰

Tregs utilize various mechanisms of immune suppression to regulate immune responses and control inflammation (Fig. 1). They express anti-inflammatory cytokines, such as TGF- β and IL-10.^{81,82} Tregs can also suppress interleukin 2 (IL-2) T cell consumption through high expression of the IL-2 receptor (CD25) and directly kill target cells in a contact-dependent manner through the expression of granzyme B.^{83,84} Importantly, granzyme B⁺ Tregs have been shown to play a critical role in controlling lung inflammation and tissue damage during acute RSV infection in mice.⁸⁵ In addition to their immunosuppressive capabilities Tregs also contribute to tissue healing and integrity through the expression of epidermal growth factor receptor ligands, such as amphiregulin and keratinocyte growth factor, during viral infection and or tissue injury.⁸⁶⁻⁸⁹ Not only are Tregs essential for self-tolerance, preventing autoimmunity, and regulating inflammatory immune responses, but they are also required at the maternal-fetal interface during implantation and throughout pregnancy.

Changes in Treg Frequency in Response to Pregnancy and Sex Hormones

hCG, produced by the blastocyst and syncytiotrophoblast during pregnancy, has been shown to increase Treg frequency, promote recruitment to the endometrium, and enhance suppressive Treg function in mice and humans^{90–93}. The location and number of Tregs fluctuates first in response to seminal antigens and the blastocyst during implantation, and later due to changing hormonal levels throughout pregnancy with a rapid decrease systemically in the third trimester before birth^{94–96}. Tregs are actively recruited and accumulate locally in the female reproductive tract and draining lymph nodes, as early as first exposure to sperm and male seminal fluid antigens, to mediate tolerance to the fetus.^{97,98} In fact, depletion or decrease of Tregs, locally and systemically, before implantation results in a failure of gestation and immunological rejection of the fetus in mice and humans.^{99,100} The number and frequency of Tregs in peripheral blood also fluctuates throughout the menstrual cycle and peaks just before ovulation, correlating with serum estrogen levels, before dramatically decreasing in the luteal phase.^{99–101} Once pregnant, maternal FoxP3⁺ Tregs greatly increase in circulating blood and locally at the maternal-fetal interface in the placenta during healthy pregnancies^{102,103}. Tregs are known mediators of fetal tolerance crucial for implantation and successful pregnancy.¹⁰⁰

There is increasing evidence that sex hormones induce expansion and differentiation of Tregs and promote their strong immunosuppressive functions. Estrogen enhances the expansion of Tregs, their ability to effectively suppress the activation and proliferation of effector T cells, increases *FoxP3* expression, and decreases production of IL-17 by Th17 cells in mice.^{26,104} Estriol, which is not present in non-pregnant females and produced at high concentrations by the placenta in the first trimester, also increases the frequency of Tregs, suppresses Th17 cell expansion, and enhances the generation of iTreg *in vitro*.¹⁰⁵ Progesterone has been found to promote the differentiation of iTreg with improved stability *in vitro*.¹⁰⁶ However, elevated levels of progesterone during the second and third trimesters have been correlated with reduced activity and expansion of Tregs in humans, though the opposite has been reported in mice.^{27,104,107} Progesterone is also present at high levels in human cord blood where it has been shown to drive a shift towards immunosuppressive Tregs while simultaneously preventing differentiation of Th17 cells.¹⁰⁶ These findings demonstrate the importance of sex-specific hormones in the localization and expansion of Tregs.

The large systemic expansion of Tregs during pregnancy suggests that many may be peripherally induced iTregs. A couple of mouse studies proposed that both nTregs and peripheral iTregs are important in pregnancy, as adoptive transfer of pregnancy induced or TGF- β induced iTregs prevented abortion during allogenic pregnancy in mice.^{108,109} One study found that in normal human pregnancies clonally expanded iTregs are induced by CD14⁺DC-Sign⁺ antigen presenting cells and are the dominant subset located in the decidua. Additionally, this process of iTreg induction was defective in pre-eclampsia cases resulting in poor iTreg cell induction in the decidua.¹¹⁰ Similarly, another research group found that clonally expanded antigen-specific effector memory Tregs were found in the decidua, but not in peripheral blood in humans. Furthermore, these decidual clonally expanded effector Tregs were highest in third trimester in healthy women, and lower in pre-eclampsia

cases.^{111,112} Taken together, these reports indicate that iTreg induction plays a large role in the systemic and local increase of Tregs during pregnancy, and in maintaining tolerance to the fetus.

Tregs and Viral Respiratory Infections

Despite their ability to suppress pro-inflammatory anti-viral responses, Tregs play a crucial role in healing tissue damage, orchestrating antigen-specific responses, and controlling systemic and local inflammatory responses during viral respiratory infections.^{88,113–115} During influenza A infection in mice, amphiregulin-expressing lung Tregs play a major role in tissue repair of cell-mediated damage early in infection resulting in lower lung pathology scores than mice with amphiregulin-deficient Tregs, despite no difference in viral load and clearance.⁸⁸ Interestingly, another study showed that estriol treatment of female mice following influenza A infection resulted in a skew towards Th2 responses and an increase of Treg frequencies in lung tissue and associated with decreased disease severity, though viral replication and clearance was unaffected.¹¹⁶ Tregs have also been shown to promote influenza specific Tfh and germinal center responses by controlling IL-2 availability.^{117,118} Additionally, the Tregs that accumulate in the lung during influenza infection seem to persist in the tissue after resolution as long-lived antigen-specific memory cells.⁷² However, the presence of Tregs does not always correlate with better disease outcomes, as one study observed that the altered increase and expansion of suppressive Tregs following influenza infection in aged mice when compared to young mice, resulted in delayed and decreased antigen-specific CD8 T cell responses.¹¹⁹ Therefore, in addition to gender, age is also an important factor in the function and expansion of Tregs during infection.

In respiratory syncytial virus (RSV), disease severity and tissue damage of the lungs has been linked to functional and phenotypic differences in the Treg population. In RSV infected mice, CTLA-4⁺ Tregs rapidly accumulate and proliferate in draining lymph nodes and lungs. Furthermore, *in vivo* depletion of Tregs prior to infection resulted in delayed viral clearance, exacerbated disease, delayed recruitment of antigen specific T cells to the lung, and increased frequency of IFN- γ and TNF (Tumor Necrosis Factor)- α expressing cytotoxic T cells.¹¹⁵ In another study, Treg depleted mice had enhanced viral clearance but also experienced increased weight loss, increased cytokine and chemokine levels, and increased T cell activation and cellular infiltration of the lungs. In the same study, when Treg numbers and activity was boosted using IL-2 immune complexes, inflammation was decreased and lung Tregs from infected mice expressed granzyme B. Furthermore, selective loss of granzyme B expression in Tregs resulted in higher cell infiltration into the lungs.⁸⁵ In humans, children with severe RSV infection have significantly reduced activated Tregs in peripheral blood and decreased IL-33 protein in nasal washes, a cytokine crucial in maintaining Tregs in mucosal tissues.¹²⁰

Like in RSV infection, disease severity and lung pathology in SARS-CoV2 have been associated with dysregulated immune responses, including a deficit of Tregs at the site of infection. A recent study showed that an increased frequency of activated Tregs in peripheral blood was associated with severe disease in hospitalized COVID-19 patients, but not in RSV or influenza patients. Furthermore, they found that increased levels of

CXCL10 and CXCL9 in peripheral blood might retain CXCR3⁺ Tregs in the periphery from homing to the lungs.¹²¹ Another study also reported that COVID-19 patients had decreased frequency and number of Tregs in peripheral blood samples that were negatively correlated with CD4 and CD8 T cell frequencies.¹²² Similarly, others revealed that an increased frequency of circulating Tregs characterized by overexpression of immunosuppressive markers and reduced amphiregulin expression correlated with severe disease and predicted mortality.^{123,124} Others have shown that severe COVID-19 airways have reduced Treg frequencies compared to healthy airways.¹²⁵ Additionally, elevated pre-infection Treg frequencies at steady-state correlated with better protection and lower viral load upon SARS-CoV2 infection in a mouse model.¹²⁶ Therefore, it is reasonable to speculate that defective trafficking, localization, and reduced tissue healing function in Tregs correlates with worse disease outcomes during viral respiratory infections.

During pregnancy, there is a general attenuation of the maternal immune system skewing towards anti-inflammatory responses. Dysregulation of the Th1/Th2 and Th17/Treg balance and an increase in pro-inflammatory responses during viral respiratory infection is associated with worse disease outcomes and complications in pregnant people.¹²⁷ A mouse study found that pregnant mice infected with IAV 2009 pH1N1 had increased mortality and displayed signs of reduced epithelial regeneration and impaired lung repair, as well as elevated levels of Tregs in the lungs despite no significant increase in viral load. There was an increased infiltration of pulmonary neutrophils and macrophages strongly correlated with elevated mortality, as well as increased nitrites and several proinflammatory cytokines involved in cell recruitment to the lungs.¹²⁸ It is reasonable to consider that this dysregulated inflammatory immune response is likely what caused increased mortality in pregnant mice and recruited an already expanded Treg population to the lungs. In the case of SARS-CoV2 infection in pregnancy, several recent studies have shown that COVID-19 patients have a significant reduction in Treg numbers and frequencies, causing an imbalance in the Treg/Th17 ratio.^{129–131} Therefore, severe disease outcomes in pregnant people during viral respiratory infection could be due to uncontrolled systemic inflammation and a disruption of the Treg/Th17 balance rather than a Treg-driven inhibition of anti-viral immune responses.

Sex Differences in the Treg Population in Non-pregnant Women and Men

In humans, the fact that females have a lower frequency of circulating Tregs than men has been suggested as a reason for an increased predisposition to autoimmune disease.¹³² Androgens have been implicated in modulating Treg responses in the context of autoimmune disease. One study showed that testosterone replacement therapy in male mice inhibited the development of autoimmune orchitis, had an immunosuppressive effect on the immune system, significantly reduced the number of CD4 conventional T cells, and induced a significant increase in the number of Tregs in the testis. Additionally, *in vitro* testosterone treatment of naïve T cells led to the expansion and differentiation of an iTreg subset with suppressive functions.¹³³ The same research group later showed that androgen stimulation of Tregs leads to increased transcription and expression of FoxP3 through modification of histone H4 acetylation. This androgen-dependent increase in FoxP3 expression occurred in T cells from ovulating women but not from men, indicating sex-specific androgen signaling.¹³⁴ Another recent study showed that testosterone significantly increased FoxP3

expression in Tregs from females with systemic lupus erythematosus and that plasma concentrations of testosterone positively correlated with higher FoxP3 Treg expression.¹³⁵ In male mice, visceral adipose tissue was found to be enriched for Tregs in comparison to females. Additionally, increased inflammation in male visceral adipose tissue facilitated recruitment and expansion of Tregs cells via the CCL21-CCR2 axis and an androgen-regulated IL-33 stromal cell population, respectively.¹³⁶ Interestingly this phenomenon is different in obesity, wherein in visceral adipose tissue the obese male mice have reduced Tregs, and obese female mice have increased Tregs. This increase in female visceral adipose tissue Tregs was abrogated in ovariectomized obese mice but restored upon estrogen supplementation, suggesting that estrogen may attenuate obesity-associated chronic inflammation. Furthermore, chemokine expression revealed a different localization system for female visceral adipose tissue Tregs which trafficked to the tissue via CCL24, CCR6, and CXCR3 signaling.¹³⁷

In summary, Tregs control pro-inflammatory effector T cell responses, aid in preventing tissue injury, and are essential mediators of tolerance to self and fetal-alloantigens. Their localization, expansion, and immunosuppressive functions are influenced by fluctuating hormone levels during pregnancy. Importantly, the expansion of Tregs locally and systemically is required for a healthy and successful pregnancy. Dysregulated Treg responses or an imbalance in the Treg/Th17 ratio may be a significant factor in pregnancy complications and adverse outcomes to viral respiratory infections.

Th17 Cells

Th17 Cells and the Influence of Sex Hormones

Th17 cells are a CD4+ T helper lineage that are characterized by retinoic acid receptor-related orphan receptor transcription factors (ROR γ t and ROR α) and the production of pro-inflammatory cytokines (IL-17, IL-21, IL-22; Fig. 2).^{138,139} Naive CD4+ T cells differentiate into Th17 cells after stimulation with IL-6 and TGF- β . However, there has also been a subpopulation of CD39+ CD73+ Th17 with immunosuppressive functions known as supTh17. The supTh17 cells help produce adenosines from extracellular nucleotides with ectonucleotidases.^{140,141} Progesterone has been found to promote the proliferation of supTh17, which may play a role in lung repair in influenza-infected mice.¹⁴²

Hormones crucial for a successful pregnancy impact the function and proliferation of Th17 cells. Work with progesterone showed a suppressive role in inflammation and as an element for milder disease outcomes. Within the umbilical cord, progesterone can bias the naive CD4+ T cells towards differentiation into Tregs and away from their natural bias towards Th17.^{106,143} Progestogens can downregulate the secretion of pro-inflammatory cytokines tied to a Th1/Th17 response whilst supporting Th2 cytokine secretion in peripheral blood from patients with unexplained recurrent spontaneous miscarriages, which already have a pro-inflammatory immune environment.¹⁴⁴ In pregnant cows, progesterone decreased the expression of ROR γ t and IL-17, but upregulated IL-4 (Th2 differentiation) creating conditions necessary for a successful pregnancy.¹⁴⁵ Medroxyprogesterone acetate, a synthetic progestogen, suppresses the expression of ROR γ t, IL-22, and IL-17, all key to Th17 function.^{144,146} Estradiol at estrus levels in mice was enough to downregulate the

Th17 immune response to extracellular pathogens and sperm.^{147,148} However, it's possible that both estradiol and progesterone may synergistically promote IL-17 and IL-23 secretion, especially in the context of a runaway Th2 immune environment.¹⁴⁹ A possible mechanism for this could be dendritic cells stimulated by estradiol secrete IL-1 which in turn can signal for the proliferation of Th17 cells to combat infection.¹⁵⁰ Estradiol at pregnancy levels suppresses the differentiation of Th17 cells from CD4+ via a PD-1 signaling pathway.¹⁵¹ Estrogen deficiency can have deleterious effects in the nervous system as Th17 cells can promote aneurysm formation and rupture.¹⁵² Estrogen can thus have a regulatory role in Th17 localization.¹⁵³ Pregnancy and pregnancy-like levels of estrogen can downregulate Th17 transcription factors and cytokines and upregulate Treg/Th2 transcription factors.¹⁵⁴ Estriol, at pregnancy-like levels, dampens Th17 responses by increasing the Treg/Th17 ratio and IL-10 secretion while inhibiting IL-17 secretion and neutrophil function.^{105,155} There is also evidence that estradiol can enhance the population of tissue resident memory-like Th17 in the female reproductive tract during an antiviral immune response.¹⁵⁶ Also, deficiencies in estrogen receptor alpha (ER α) reduce Th17 differentiation which correlates with IL-17 and IL-23 secretion reduction.¹⁵⁷ However, estradiol can trigger the formation of an ER α -repressor complex which represses expression of ROR γ t.¹⁵⁸

Regulation of the Treg/Th17 Balance During Pregnancy

Studying the “seesaw” between Treg/Th17 frequencies has yielded new knowledge about how recurrent pregnancy loss and other complications can arise that put the wellbeing of both parent and fetus at risk. In healthy pregnancies, the Treg/Th17 favors the proliferation of Tregs and the development of a tolerogenic immune profile, though Th17 levels may be proportionally higher in the decidua than in the parent's peripheral blood.¹⁵⁹ In patients with pre-eclampsia, a pregnancy complication featuring high blood pressure and organ dysfunction in the liver or kidneys, the ratio is skewed in favor of Th17.¹⁰³ A similar phenomenon has been observed in the peripheral blood and decidua of patients with unexplained recurrent pregnancy loss when compared to healthy pregnancies¹⁶⁰. It should be noted that Th17 cells also fulfill an important part in protecting pregnancies from infection hence the observed role that pregnancy-specific glycoprotein 1a plays in stimulating dendritic cells to enrich the circulation of Th17, among other lymphocytes.¹⁶¹ Another potential effect of healthy Th17 presence is that IL-17 has been shown to stimulate trophoblasts towards the production of progesterone *in vitro*.¹⁶² Th17 cells can also be recruited by decidual stromal cells and prevent trophoblast apoptosis in the first trimester of healthy pregnancies.¹⁶³ However, chronic infusion of IL-17 in pregnant rats yielded physiological complications, hypertension and impaired uterine artery reactivity, and increased cytotoxic NK cell activity.¹⁶⁴ IL-17 and Th17 cells were also suggested to be in part responsible for maternal immune activation and subsequent fetal cortical abnormalities in line with an autism phenotype.¹⁶⁵

Th17 as a Mediator of Viral Immunity

The role of Th17 cells and their effectors in influenza, especially in the context of pregnancy, is an evolving field of study. In a murine model of H3N2 infection during pregnancy, IL-17 was elevated in circulation and ROR γ t levels were upregulated in the intestines in parallel with maternal pulmonary inflammation and dam weight loss.¹⁶⁶ In

line with this, higher levels of IL-17 and Th17-linked cytokines (IL-6, IL-8) were found in human patients infected with the 2009 H1N1 strain.¹⁶⁷ Also, mice who were also infected with this strain were relieved of acute lung injury reducing the circulation of IL-17. Another study suggested that elevation in the Th17 cytokine profile (IL-17, IL-6, IL-8, IL-9) is a hallmark of early, severe influenza disease.¹⁶⁸ IL-17RA deficient mice were found to have better survival, less weight loss, and lower levels of oxidized phospholipids, particularly relevant as an agent of lung injury during influenza infection.¹⁶⁹ On the other hand, in a study of an IAV H5N1 model of infection in mice, IL-17 knockout led to worse disease outcomes and impaired B cell localization to the lung over the course of infection.¹⁷⁰ As Th17 is typically considered as a crucial component of the immune defense against extracellular pathogens, research suggests that influenza A infection triggers interferon-mediated repression of Th17 immunity which impairs the clearance of subsequent bacterial co-infection.¹⁷¹ Th17 immunity against influenza may also be impaired during pregnancy because of the bias towards IL-10+ Tregs.¹⁷² Of note as well is that Th17/Tc17 frequencies and IL-17 concentrations in circulation were depressed in humans infected with H7N9, an avian influenza strain of concern.¹⁷³ All these works come together to suggest that appropriate Th17 responses are crucial to overcoming influenza disease. However, conditions of pregnancy can result in a dampening of Th17 immunity, possibly alleviating lung injury and prolonging the disease course.

RSV, while predominantly a disease of concern for children, has yielded insights into Th17 behavior during viral respiratory infections. Nasal washes from infants with RSV infection revealed that IL-8 levels were lower than normal in these infants, which may tie into Th17 disruption, as neutrophil recruitment is a major downstream effect of their activity.¹⁷⁴ IL-27 downregulation may be at play during RSV infection and can lead to elevated IL-17 circulation, leading to an imbalance in the Th1/Th2 dynamic.¹⁷⁵ RSV infection in mice has suggested that Th17 are promoted during infection as signaling pathways leading to their differentiation were upregulated in the lungs.¹⁷⁶ Rat infants infected with RSV showed elevated presence of Th17 biomarkers and lower levels of Treg biomarkers in their lungs.¹⁷⁷ A possible mechanism for the skewering of the Th17/Treg seesaw could be that dysfunctional dendritic cells may be secreting more IL-6 and creating the conditions for Th17 polarization during RSV infection.¹⁷⁸ Furthermore, elevation of circulating levels of IL-17 may exacerbate symptoms and lower levels of IL-17 actually leads to milder disease outcomes and a stronger RSV-specific CD8+ T cell response.¹⁷⁹ RSV infection needs to be studied further to discern whether infections during pregnancy, especially if concurrent with offspring delivery, result in a pathogenic Th17 response.

Numerous studies have sought to establish Th17 cells as crucial mediators of severe COVID-19. Previous reviews have speculated that adverse pregnancy outcomes in people with COVID-19 and pregnant people's susceptibility to respiratory illnesses could be in part explained by a Treg/Th17 imbalance contributing to a cytokine storm.¹⁸⁰ In the context of severely ill patients, researchers found high frequencies of Th17 cells, cytotoxic CD8+ cells, and levels of IL-17, suggesting a dysregulated pro-inflammatory Th17 response.^{131,181,182} This phenomenon correlates to a decrease in the frequency of Tregs and expression of their biomarkers (TGF- β , IL-10) which could be disastrous in the context of pregnancy.¹¹³ These findings are in agreement with earlier studies of Middle Eastern Respiratory Syndrome,

an earlier coronavirus epidemic.¹⁸³ Even after viral clearance, tissue resident memory-like Th17 cells were found in the respiratory and circulatory systems of patients, possibly playing a role in disease severity and lung injury. However, a single cell RNAseq study found that Th17 cells were underrepresented among CD4+ T cells taken from COVID-19 patients. Ultimately, the research in this area is ongoing, but most research points to a Th17 dysregulation that contributes to COVID-19's characteristic cytokine storm.

Th17 cells are a key mediator of the inflammatory response to pathogens by recruiting neutrophils to sites of inflammation. However, a strong Th17 response can be harmful to both the pulmonary and placental tissues. Estrogen and progesterone can downregulate the proliferation and function of Th17 cells to avoid this potential for harm during pregnancy. Furthermore, Th17 cells maintain a delicate ratio with Tregs, a ratio that may develop into a detriment in the initial immune response towards a viral respiratory infection. This leads to potential future research work into studying the interactions between the Treg/Th17 ratio, sex hormones, and the need to maintain an immunological landscape favorable for pregnancy.

$\gamma\delta$ T cells

$\gamma\delta$ T cells, Sex Hormones and the Impact on Respiratory Viral Infections

$\gamma\delta$ T cells make up only 5–10% of peripheral T cells, but are most concentrated at mucosal tissues (i.e., skin, intestine, uterus). They fulfill an innate-like function between the innate and adaptive responses due to their localization (Fig. 3).¹⁸⁴ They can act to facilitate innate immunity, but also can persist as pathogen-specific T cells. Aside from their function, the major distinguishing feature of this group is that they come in combinations of V γ 1–9 chains with either V γ 1 or V γ 2 chains for their T cell receptors, in comparison to the alpha-beta chains of the rest of their T cell cousins. Regarding their role in innate immunity, there are two recognized groups of $\gamma\delta$ T cells: those that produce IFN- γ and others that produce IL-17 (Fig 4).¹⁸⁵ The fate of T cells depends on either strong TCR signaling for IFN-+ or cytokine stimulation (IL-1 β , IL-23, TGF- β) for IL-17+ T cells.^{186,187} IL-17+ $\gamma\delta$ T cells secrete some of the same cytokines as Th17 cells (IL-21, IL-22).¹⁸⁷ During early acute infection, IL-17+ $\gamma\delta$ cells make up the primary source of IL-17 secretion, more so than the CD4+ Th17 cells.^{188–190} In mucosal tissue, the frequency of $\gamma\delta$ T cells is almost 40% and most of these display the V δ 1 chain, in contrast to the peripheral blood $\gamma\delta$ T cells which typically display V δ 2.^{191,192}

Sex Hormones Influence $\gamma\delta$ T Cell Frequencies

Sex hormones influence $\gamma\delta$ T cell functions and localization in females.¹⁹³ Peripheral $\gamma\delta$ T cell frequencies increase in healthy pregnancies and over 90% express progesterone receptors (PR), which connects to secretion of progesterone-induced blocking factor (PIBF), a protein that aids immune evasion of the developing embryo, and IL-10 secretion.¹⁹⁴ In patients with unexplained recurrent spontaneous abortion, PR+ decidual $\gamma\delta$ T cells decreased significantly, correlating with a decrease in PIBF and co-stimulatory molecules. A high enough level of progesterone is necessary for high secretion of PIBF among $\gamma\delta$ T cells, suggesting a hormone-mediated regulation of decidual $\gamma\delta$ T cell function.¹⁹³ In

an interesting study using sheep, progesterone was found to prevent xenograft rejection and increase $\gamma\delta$ T cell frequency in the uterus.¹⁹⁵ As progesterone levels increase during pregnancy, endometrial stromal cells recruit V δ 1+ $\gamma\delta$ T cells to the endometrium, shifting the V δ 1/V $\gamma\delta$ 2 ratio away from V δ 2+ $\gamma\delta$ T cells, possible bad actors in unexplained recurrent spontaneous abortion.¹⁹⁶ Estrogen seems to selectively regulate IL-17+ $\gamma\delta$ T cell migration from lymphoid tissue; possibly allowing a greater frequency of them to be diverted towards the uterine tissue during pregnancy.^{197,198} Estradiol stimulates IL-17 secretion by $\gamma\delta$ T cells in female reproductive tissue, which is then necessary to prime an adequate Th17 response.¹⁹⁹

$\gamma\delta$ T cells and Their Functions During Pregnancy

During pregnancy, $\gamma\delta$ subsets maintain a delicate balance between those primed for responding to pathogens and those assisting a successful pregnancy. It has been shown that at the start of pregnancy, there is a pro-inflammatory bias among the uterine $\gamma\delta$ T cell population, but as the pregnancy progresses, there arises a tolerogenic bias in the decidual $\gamma\delta$ T cell population.^{200–202} Furthermore, V δ 1 $\gamma\delta$ T cells maintain cell-to-cell communication with trophoblasts, which may help, dictate expansion in their population in the later trimester of pregnancy.^{203,204} Decidual $\gamma\delta$ T cells produce TGF- β and IL-10, key cytokines for maternal tolerance of alloantigens, and produce little to no inflammatory cytokines such as TNF- α and IFN- γ . In addition, decidual $\gamma\delta$ T cells may promote trophoblast proliferation and suppress apoptosis in trophoblasts, supporting early pregnancy.²⁰⁵ Researchers have also found that decidual $\gamma\delta$ T cells expressed memory cell markers (CD45RO) and cytotoxic markers, suggesting an early role in preventing unwanted infections.²⁰⁶ Among the $\gamma\delta$ T cells of the uterus and placenta/decidua there is a large frequency of IL-17+ $\gamma\delta$ T cells, which may be critical for preparing the uterus for implantation after insemination.^{205,207,208} These IL-17+ $\gamma\delta$ T cells present a V γ 6 chain and may peak in frequency sometime in the second trimester.²⁰⁹

As opposed to other tissues, IFN- γ + $\gamma\delta$ T cells may be at lower frequencies in maternal tissues as their secreted cytokines may be harmful for pregnancy.^{208,210} Furthermore, they can infiltrate the fetal nervous system after maternal immune inflammation.²¹¹ It is also worth noting, there is evidence to suggest that inflammatory IL-17+ $\gamma\delta$ T cells may also be responsible for negative pregnancy outcomes for patients with recurrent pregnancy loss.²¹² An overabundance of cytotoxic $\gamma\delta$ T cells during early pregnancy would be dangerous, as they are linked to patients with recurrent implantation failure.^{213,214} Interestingly, work done in studying correlations between $\gamma\delta$ chain combinations and pregnancy outcomes. V γ 9V δ 2 T cells were predominant in patients with recurrent spontaneous abortions, while V γ 1V δ 1 were more frequent in healthy pregnancies, probably because of their preponderance to produce IL-10 and suppress cytotoxic NK activity.^{215,216} V δ 2 $\gamma\delta$ T cells in peripheral blood of recurrent spontaneous abortion patients expressed more CD107 and IL-17, leading to the idea that this phenotype is in part responsible for recurrent spontaneous abortion.²¹⁷ Provocatively, a study in late gestation sows found that CD8+ IFN- γ + cytotoxic $\gamma\delta$ T cells were prevalent in maternal peripheral blood and gestational tissues.²¹⁸

Role of $\gamma\delta$ T cells in Viral Respiratory Infections

As a component of the innate immune response, $\gamma\delta$ T cells secrete pro-inflammatory cytokines and their population expands, and the same functions hold true during respiratory viral infections. IL-17+ $\gamma\delta$ T cells are recruited at high frequencies to the upper respiratory tract as part of innate immunity against influenza virus infection.²¹⁹ These IL-17+ T cells express the V γ 4 chain, which can also be found in $\gamma\delta$ T cells in the uterus and become tissue resident memory-like cells in the aftermath of other respiratory infections.^{208,219,220} This relation suggests that the IL-17+ $\gamma\delta$ T cell population may be impacted if influenza infection occurred during pregnancy. IL-21 plays a role in a negative feedback loop for the IL-17+ $\gamma\delta$ T cells in the respiratory tract during influenza infection.²¹⁸ Interestingly, IL-17+ $\gamma\delta$ T cells have a protective function for the lungs of neonates infected with influenza, which suggests this phenomenon may also occur during *in utero* infection.^{169,221} V γ 9V δ 2 T cells fulfill a cytotoxic role, eliminating influenza-infected lung epithelial cells and monocyte-derived macrophages of the respiratory system.^{222–225} Lung inflammation decreases when V γ 9V δ 2 T cells are stimulated thereby decreasing influenza disease severity, for both avian and seasonal human influenza.²²⁶ V γ 9V δ 2, during influenza infection, secrete Th1 cytokines and chemokines and are classified as IFN- γ + $\gamma\delta$ T cells.^{222,227,228} This subset also strongly relies on CD137 co-stimulation for ideal effector function and expresses high levels of CD69 activation marker in the context of influenza infection.^{228–230}

In other respiratory viral infections, $\gamma\delta$ T cells also form a fundamental portion of the antiviral innate immune response. During RSV infection, IL-17 expression is impaired in neonates and V γ 4+ $\gamma\delta$ T cells adopt an IFN- γ +, proinflammatory cytokine secretome.^{231,232} However, it has not been confirmed if there is a specific $\gamma\delta$ T cell subset that handles IL-17 secretion during RSV infection. Increased frequencies of V δ 2+ $\gamma\delta$ T cells were detected in neonates born from mothers with ongoing SARS-CoV-2 infection along with a cytokine secretion profile most likely derived from the maternal immune activation event.²³³ Earlier work with SARS-CoV-1 identified that IFN- γ + V γ 9V δ 2 T cells were an important portion of the effector memory T cell response to SARS-CoV-1.²³⁴ This subset may also form a part of the immune response to SARS-CoV-2, localizing to the lung from peripheral blood and persist as tissue resident memory-like cells.²³⁵ However, a study has shown that $\gamma\delta$ T cell populations decrease during SARS-CoV-2 infection, possibly a result of exhaustion.²³⁶

However, most studies outline a transitional role for $\gamma\delta$ T cell subsets as they aid in implantation via inflammation and must quickly adopt a support role in maternal immune tolerance. However, $\gamma\delta$ T cells can also participate in a strong inflammatory response during a viral respiratory infection that can be harmful to the mother. An interesting research direction would be to investigate $\gamma\delta$ T cell subset localization to the lung during pregnancy or under the influence of sex hormones. Understanding how $\gamma\delta$ T cell subset frequencies change in these special immunological circumstances could be valuable in understanding the pulmonary host response to respiratory viral infection during pregnancy.

Th1 and Th2 Cells

Impact of Sex Hormones on Th1/Th2 Balance

A balance of Th1/Th2 subsets was an early immune mechanism proposed to explain tolerance of the semi-allogeneic fetus. Many studies followed demonstrating that maternal T cell mediated tolerance and T cell suppression as critical for a successful pregnancy. While compelling, many of these studies that support the Th1/Th2 paradigm were limited by use of experimental animal models, small human sample size, and a focus on recurrent spontaneous abortions, which in sum are not representative of the typical healthy human pregnancy.^{201,237–239} Human pregnancy is now understood as a state of maternal-fetal immunomodulation, where hormones upregulate the innate and humoral immune response while downregulating the cell-mediated immune response, a stage-dependent shift in the maternal immune response that is largely mediated by Th1 and Th2 cells.⁷ First, Th1 and Th2 cells play a key role in establishing and maintaining pregnancy (Fig. 4). During implantation, a Th1 cytokine profile dominates, producing a proinflammatory response during implantation and releasing TNF- α , which inhibits trophoblast cell mobility and adhesion.²⁴⁰ Together with IFN- γ , these cytokines promote angiogenesis and uterine tissue remodeling to control trophoblast invasion at the implantation site.²⁴⁰ Th1 cells also release IL-2, which stimulates the chorion to release PGE2, a prostaglandin critical for pregnancy maintenance.²⁴¹ After implantation, the maternal immune response shifts to a Th2 dominated profile. Uterine dendritic cells induce naïve CD4+ T cells to differentiate into Th2 cells, and the decidua releases cytokines to recruit Th2 cells to the maternal-fetal interface.^{240,242} Th2 cells then promote a largely humoral immune response for the rest of the pregnancy.²⁴³

A shift from a Th1 to a Th2 profile in pregnancy was proposed to explain how the maternal immune response is sufficiently immunosuppressed to tolerate a semi-allogeneic fetus, while maintaining sufficient vigilance to defend both the mother and fetus from infection.^{240,242,244} Indeed, the Th1/Th2 profile in human maternal blood has been demonstrated to decrease throughout the course of gestation in tandem with rising hCG, estrogen and progesterone sex hormones. At low concentrations, estradiol promotes Th1 cell-mediated immune response by enhancing IFN- γ transcription.²⁴⁵ At high concentrations, estradiol promotes Th2-led humoral immunity.^{246,247} Estrogen contributes to the Th2 immune profile by stimulating IL-4 secretion.²⁴⁵ By decreasing levels of TNF- α , hCG also inhibits the Th1 pathway.^{243,248,249} Progesterone also promotes a Th2 profile as shown *in vitro* and *in vivo*. In primary CD4+ T cells isolated from the PBMCs of non-pregnant women, progesterone suppressed CD4+ T cell activation, and downregulated gene targets associated with the STAT1 and STAT3 pathways including IL-2, IL-12b, IL-10.^{250,251} Progesterone inhibits the Th1 pathway by decreasing TNF- α and IL-6 release, and stimulates the Th2 pathway by increasing IL-4 and IL10 release.^{243,252} High concentrations of progesterone also stimulate lymphocytes to synthesize progesterone-induced binding factor (PIBF).²⁵³ PIBF in turn promotes naïve CD4+ T cell differentiation into Th2 cells, which then secrete high levels of anti-inflammatory cytokines.²⁵² Additionally, progesterone directly stimulates T cell activation: modulating TCR signaling, blocking PIBF-induced degranulation, and inhibiting cellular cytotoxicity.²⁵⁴

Overall, multiple sex hormones act on Th1 and Th2 cells to shift their ratios during pregnancy to favor fetal tolerance.

Impact of Pregnancy Hormones on Th1/Th2 Response to Respiratory Viral Pathogens

The maternal immune response during pregnancy is thus not simply suppressed but rather biased towards an antibody-based, anti-inflammatory profile that shapes disease pathogenesis and manifestation. In non-pregnant individuals, SARS-CoV-2 disease progression is characterized by a cytokine storm of pro-inflammatory, Th1 cytokines that include IL-2, IFN- γ , IL-1, and TNF- α .²⁵⁵ Higher levels of IFN- γ were linked to a higher risk of mortality, suggesting that an ineffective Th1 response prevented successful elimination of the virus.²⁵⁵ A reduction in Th1 responses during pregnancy may partially explain why pregnant women are more susceptible to severe SARS-Cov-2 and IAV disease.²⁵⁶ In pregnant Sprague-Dawley rats, IAV infection further shifted the ratio of Th1/Th2 to favor Th2, which led to more severe disease.²⁵⁷ Pregnant BALB/c mice infected with the 2009 pandemic IAV H1N1 virus exhibited lower levels of peripheral blood T cells and antibody responses than their non-pregnant counterparts.^{257,258} Notably, a change in Th1/Th2 profiles in pregnant women does not seem to affect the efficacy of the inactivated influenza vaccine,²⁵⁹ suggesting that more work must be done to characterize the impact of pregnancy hormones on the Th1/Th2 cytokine profiles, vaccine efficacy and outcome to respiratory viral infections.

Taken together, key pregnancy hormones regulate Th1 and Th2 activity to favor tolerance of the semi-allogeneic fetus. Severe maternal disease associated with respiratory viral infections has been attributed, in part, to the Th2 dominance in pregnancy, which is less adept at clearing viral infections. Much of the existing literature evaluates this relationship by measuring cytokine levels characteristic of either Th1 or Th2 cells in pregnant animal models and human subjects. However, to fully elucidate the regulatory role that pregnancy hormones like progesterone and estradiol play in the Th1/Th2 immune response, more studies are needed that directly target the reciprocal interactions between pregnancy hormones and effector lymphocytes.

B Cells

Immunologic Functions of B Cells

B cells serve numerous immunologic functions which include (1) displaying diverse immunoglobulin (Ig) surface receptors to bind, phagocytose, and present foreign antigens for cellular or complement activation, (2) differentiating into plasma cells that produce immunoglobulins (Ig) of unique affinity, specificity, and function, and (3) releasing cytokines (e.g., IL-6, IFN- γ , TNF- α , IL-10, IL-13, etc.) for coordination with other host immune cells including other antigen presenting cells (e.g., dendritic cells, macrophages, Langerhans cells) and T lymphocytes.²⁶⁰ There are two subsets of B lymphocytes: B1 and B2 cells.^{261,262} B2 cells are essential to the adaptive arm of the immune system, initially displaying IgM surface receptors for both antigen sampling and co-stimulation with splenic T lymphocytes.^{261,262} Antigen-bound IgM surface receptors undergo internalization and subsequent Ig class-switching, somatic hypermutation, and affinity maturation.²⁶⁰ Further

signaling stimulates B2 cells to differentiate into plasma cells and memory B cells capable of producing secretory Ig that underlie high-affinity, high-specificity antibody-mediated responses.²⁶² B1 cells, in contrast, produce ‘natural antibodies’ which are generated without antigenic stimulation and initially accumulate during fetal development to provide lower affinity and lower specificity Ig for broader neutralization of exogenous proteins or apoptotic self-cells.^{263–265} The antibody-mediated protection conferred by B lymphocytes is tightly regulated to allow these cells to differentiate “self” from “nonself”.

The Influence of Sex Hormones on B Cells

Sex steroids (e.g., estrogens, progesterone, and androgens) have been well established as potent regulators of humoral immune responses, notably contributing to upregulation, downregulation, and dysregulation of adaptive immune responses.^{245,264–269} This complex interplay is best demonstrated through the actions of estrogen; elevated estrogen levels have been well established in the pathogenesis of certain autoimmune disorders (e.g., systemic lupus erythematosus, multiple sclerosis) while in tandem contributing to robust immune responses to infection or vaccination as well as diminished immune responses in the setting of marked hormonal fluctuations (e.g., pregnancy).^{245,267,268,270} The potentially opposing impacts of estrogen on B cells comprise an area of ongoing research, however, the current literature highlights that estrogen confers up- and down-regulation of B cell production, maturity, and stimulation through interactions with unique receptors (estrogen receptors (ER): ER α or ER β). Binding of ER shapes downstream gene expression and subsequent cell-to-cell interactions, most notably within the context of differing physiologic states (e.g., pregnancy, menopause, puberty).^{245,266,268} The immunomodulation of B cell production and differentiation by sex hormones is best exemplified in pregnancy where sex hormones directly affect B cell differentiation and proliferation²⁷¹.

B Cells in Pregnancy

During pregnancy, the fetus expresses both maternal and paternal human leukocyte antigen (HLA) genes, presenting a mosaic of MHC receptors recognized as partially foreign by circulating maternal B cells. Activation of the maternal immune system against a semi-allogeneic fetus could have catastrophic consequences (e.g., pregnancy loss) in the absence of immunomodulation. Further downregulation of inflammatory and immunologic responses to infection are needed to prevent fetal injury due to infection.²⁷² Multiple mechanisms involving B cell subsets act to suppress maternal immune activation throughout pregnancy. Protective antibodies, produced by B2 cells, identify the paternal antigens present on the fetus and subsequently downregulate cytotoxic lymphocyte responses by trophoblastic cells in the placenta.²⁶⁴ Regulatory B cells (Breg) further modulate the maternal immune response by secreting IL-10, an anti-inflammatory cytokine that suppresses pro-inflammatory responses by maternal effector T cells and activates anti-inflammatory cell lines.^{264,272–274} Breg levels rise steadily throughout the first trimester of pregnancy; emerging research on estrogen and mechanisms of immunomodulation has identified hCG and estrogen receptors on Breg cells, implicating hormonal fluctuations in the Breg elevations characteristic to early pregnancy.²⁷⁴ Through increased IL-10 signaling, levels of proinflammatory Th1 cells are gradually eclipsed by anti-inflammatory T helper

2 (Th2) cells which further upregulate IL-10 production and perpetuate maternal immune suppression.²⁶⁴

Although Breg subsets appear to increase in the first trimester, studies characterizing circulating B cell populations in pregnancy have shown that nearly all other B cell subsets are significantly diminished by the third trimester and throughout parturition, compared with non-pregnant or postpartum individuals.²⁶³ Immature B cells are particularly affected due to suppressed lymphopoiesis in the bone marrow. A gradual rise in estradiol reduces IL-7 cytokine expression, which is critical for B cell differentiation and proliferation and contributes to lymphopoiesis (Fig. 5).^{275,276} In effect, rising estrogen levels throughout pregnancy cause a marked decrease in B cell precursor cells and a gradual increase in specific subsets of mature B cells which are elevated throughout the third trimester and parturition.^{263,274} B cell counts among postpartum and non-pregnant women are largely similar, indicating that lymphopenia in the third trimester resolves following delivery as estradiol levels decrease and B cell maturation resumes, and broader B cell populations rise to near-normal levels.²⁶³ This unique finding was further confirmed by diminished levels of CD23, a marker of B cell differentiation from naïve to memory B cells, throughout the transition from third trimester into parturition with gradual returns to baseline in the postpartum period.²⁶³ Despite these fluctuations, B cell levels remain within standard reference ranges throughout pregnancy, delivery, and the postpartum period. A selective increase in naïve mature B cells may be explained by the effects of progesterone, which reduces further differentiation to plasma cells or memory B cells.²⁶³ Progesterone acts to suppress maternal immune responses by stimulating IL-10 production and modulating dendritic and natural killer cell functions to reduce interactions with B cells, dampening broader immune activation.^{277,278} Studies employing murine models have identified that progesterone suppresses antigen presentation among B cells through the diminished expression of critical surface co-stimulatory receptors CD80 and CD86.²⁵⁴ It is plausible that a steep decline in estrogen and progesterone levels following parturition may underlie increased lymphopoiesis and B cell differentiation to restore baseline mature B cell populations.

B cells serve a paramount role in protecting both the mother and fetus from infectious pathogens. Fetal B1 lymphocytes produce natural antibodies to supplement maternal immune responses.^{261,279} Transplacental transfer of maternal IgG antibodies to the fetus, starting as early as 13 weeks, augments the newborn's own developing immune system. This mechanism of passive immunity underlies an added benefit of maternal vaccination and presents a key area of further study to identify how maternal antibodies can be both beneficial and potentially harmful by way of the maternal-fetal interface.

Pregnancy Influences on B Cell Responses to Viral Infections

Respiratory syncytial virus (RSV) is a major cause of lower respiratory tract infections among infants; pulmonary infiltrates consisting of various B cell subsets reflect robust neonatal adaptive immune responses to the viral pathogen. RSV particles bind and infect fetal Breg in lung tissues, inducing the upregulation of Breg CX3CR1 chemokine receptors that interact with RSV G glycoproteins to stimulate IL-10 secretion and downstream anti-

inflammatory responses.^{280,281} Profuse IL-10 release has been correlated with diminished T helper 1 (Th1) priming or activation. In conjunction with increased RSV-infected Breg cell frequencies, IL-10 elevations may be predictive for more severe outcomes in the setting of RSV infections during pregnancy.²⁸¹ Transplacental transfer of maternally produced IgG significantly contributes to fetal immunity against RSV and potential vaccines to induce maternal antibody responses are being evaluated.

In pregnancy, the maternal adaptive immune response is attenuated through the decreased production of specific cytokines (IFN- α and IFN- γ) as well as the impaired maturation of antibodies critical to neutralizing pathogenic respiratory viruses, like IAV.²⁷¹ Whether increased morbidity is due to prolonged viral insult or damage from dysregulated maternal immune activation is often uncertain. The inflammatory state that follows IAV infection, however, has been closely linked with the development of neurocognitive disorders in children.²⁸² Although further research is needed to evaluate immune responses to influenza in pregnancy, an inactivated influenza vaccine has been well established as a safe method of protecting pregnant mothers against infection. The vaccine has been shown to promote robust serologic immune responses in pregnancy and maternal vaccination primes the fetus to elicit its own B cell response.^{283,284}

B cell-mediated antibody responses have been shown to assist in neutralization and clearing of SARS-CoV-2 as with most other viral infections in pregnancy.²⁸⁵ Estrogen- and progesterone-mediated actions on lymphopoiesis reduces the population of maternal B cells capable of responding to SARS-CoV-2 infection in pregnancy and pregnant women are less likely to have detectable antibody titers against SARS-CoV-2 compared to non-pregnant counterparts. Correlational analysis of differentially expressed genes representing cellular activity identified diminished expression of IL-6 and elevations in gene expression among naive B cell or memory B cell populations for IL-1 β among pregnant women compared with non-pregnant women²⁸⁶. The cytokine storm characteristic of COVID-19 disease is partially produced through the broad antibody and cytokine release by activated B cells; this cascade of immune activation appears to be attenuated during pregnancy.^{287,288}

Hormonal Influences on B Cells Beyond Pregnancy

One of the most significant immunological modifications during pregnancy is the increase of B cell responses due to the increased levels of progesterone and estrogen during pregnancy, which both reach their peak levels in the third trimester of gestation (Fig. 5).²⁴⁷ Outside of obstetrical models, the hormonal influence over B cell production and function is further exemplified through androgen-derived immunomodulation. Like estrogen and progesterone, testosterone reduces B cell lymphopoiesis in the bone marrow through attenuation of stromal cell differentiation.^{266,289,290} One mechanism for this androgenic marrow immunosuppression is exemplified in androgen receptor knock-out murine models wherein the absence of androgen receptors on osteoblast cells led to increased B cell precursor subsets, implicating osteoblasts in androgenic influences over B cell precursor production.²⁸⁹ These knock-out studies, however, found no significant change in splenic B cell counts, despite an established role of testosterone in suppressing splenic B cell populations.²⁸⁹ An alternative mechanism of testosterone-

regulated splenic immune suppression has been suggested to involve the cytokine BAFF, which serves as a critical survival factor for splenic B cells and is negatively regulated by testosterone.^{266,289,290 266,289,290}

B cells comprise a critical arm of the adaptive immune response, contributing both a myriad of receptor-mediated antibody responses and regulatory anti-inflammatory functions to the modulation of healthy immunity. Pregnancy represents a dynamic hormonal milieu, whereby the specific impacts of sex hormones on B cell function can be best evaluated. Estrogen and progesterone contribute significantly to the upregulation of specific mature B cell lines (e.g., Breg) and the suppression of numerous other B cell populations which aids the attenuation of maternal immune responses against fetal antigenicity. This modulation of maternal immunity further impacts maternal adaptive immune function in the setting of viral respiratory infections, contributing to higher morbidity and mortality risks among pregnant individuals.

Innate Lymphoid Cells

Innate lymphoid cells (ILC) refer to a broad category encompassing a variety of lymphoid lineage cells that respond rapidly to pathogen-associated molecular patterns (PAMPs) rather than specific antigens. Many subtypes with a wide variety of functions are classified within this broad category, but this review will focus on those that are involved in cytokine secretion during pregnancy and respiratory infections and that may respond to sex hormones. Here we highlight ILC subtypes and decidual natural killer (dNK) cells, as they have either a prominent role in pregnancy, defense against viral respiratory pathogens or are known to be influenced by sex hormones.

ILCs and Sex Hormones

A major class of cytokine secreting ILCs are helper ILCs. Helper ILCs are divided into 3 groups: ILC1s, ILC2s, and ILC3s, which are analogous to T-helper (Th) cell subsets Th1, Th2, and Th17, respectively.^{291–293} They are often tissue resident cells, providing a broad first line of innate immune defense. Lymphoid tissue inducer cells are considered part of the ILC3 group but can be considered separately concerning respiratory virus response, as they are not functionally linked to antiviral immune responses and, instead, largely involved in lymphoid organ development.²⁹⁴ Another subtype of ILC is the decidual natural killer cell (dNK). dNK cells are considered ILCs because they do not respond to specific antigens. Despite their name, dNK cells are non-cytotoxic, instead producing cytokines and chemokines like Th cells do. These cells become tissue residents within the uterus during pregnancy.²⁹⁵

Although the recent discovery of ILCs means that there is not much information on their response to pregnancy hormones, there are known sex differences in ILC numbers. Much of this early research has been done on mouse models, and the response of human ILCs to sex hormones is an area that remains to be explored. In mice, the ILC3 and ILC1 subgroups are not shown to express any sex hormone receptors, while ILC2 expresses androgen receptors, mediating the effects of testosterone and dihydrotestosterone.²⁹⁶ Female mice have greater quantities of ILC2, and human studies have shown higher numbers of

ILC2 in women with asthma than in men with asthma.²⁹⁶ Furthermore, mouse models demonstrate decreased ILC2 and eosinophil counts, both of which are implicated in respiratory inflammation, in response to testosterone.²⁹⁷ The majority of ILC2 do not express estrogen receptors, but uterine ILC2 express both *Esr1* and *Esr2*.²⁹⁸ The effect of pregnancy hormones on ILCs is also not yet well researched, but some early findings suggest that the ability of ILC3 to present self-antigens on HLA is hormonally controlled. hCG downregulated HLA production in murine models, while VEGF, progesterone, and estradiol did not have any effect.^{298,299} The effect of pregnancy hormones on ILCs is unknown, but some early findings suggest that the ability of ILC3 to present self-antigens on HLA is hormonally controlled. hCG downregulated HLA production in murine models, while VEGF, progesterone, and estradiol did not have any effect.^{298,299}

Sex hormones and dNK cell function are tightly linked. Prolactin and IL-15 increase after ovulation in response to progesterone, inducing the proliferation of dNK.²⁹⁵ These hormones continue to rise as the pregnancy continues. As the function of dNK cells changes throughout the pregnancy, so do the hormones they respond to. For example, dNK cells migrate to the spiral arteries to direct remodeling specifically in early pregnancy. This timing is associated with a rise in estrogen and progesterone.³⁰⁰ Another sex hormone, estradiol, induces CXCL10 and CXCL11 expression in the endometrium, both of which are ligands for CXCR3 on dNKs.³⁰¹ The association between sex hormones and the increased numbers of dNK cells and dNK ligands suggests that the hormones may induce their migration and proliferation. Interestingly, though dNK cells respond to progesterone, they do not express a progesterone receptor, meaning that the effects of this sex hormone are mediated indirectly.³⁰²

ILCs in Pregnancy and in Viral Respiratory Infections

All ILCs have been found in the endometrium of the non-pregnant uterus. Though ILC2 is initially only present in low numbers, a shift in ILC expression occurs during pregnancy.³⁰³ All helper ILC subtypes are also present within the decidual tissue in a pregnant person, with ILC2 representing the most abundant subtype and ILC1 making up a minor proportion of the uterine ILCs.³⁰⁴ In general, ILCs produce the same cytokines in the decidua as they do in other tissues, though the relative numbers of cytokines produced can change as the fetal microenvironment changes.³⁰⁴ For example, in preterm labor, ILC3 located in the decidua produce higher levels than ILC2 of cytokines, like IL-22. The amniotic fluid is another important location for ILC defense, as it protects the fetus from infection. Here, ILC3 predominates, producing IL-17 to fight amniotic infection.³⁰⁵ The relative abundances of each ILC subtype, like the proportions of the cytokines, change throughout gestation, such as ILC3 levels remaining high until the second trimester of pregnancy, when they begin to wane.³⁰⁵ Though the exact function of the maternal ILCs has not yet been elucidated, it is thought that they may play a role in both fetal tolerance and immune defense against pathogens.

ILCs have a major role in innate, first-line defense against respiratory viral pathogens due to their location at mucosal sites like the respiratory tract. The predominate ILC helper subtype in the respiratory tract is ILC3, and its production of IL-17 and IL-22 is key in

mediating immune defense against respiratory pathogens and allergens.^{291,306} One study demonstrated that infection in mouse models of mouse adapted H1N1 influenza induced ILC response within the lungs and ILC depletion reduced lung function in the setting of influenza infection.³⁰⁷ Studies of respiratory syncytial virus (RSV) and severe acute respiratory syndrome-coronavirus-2 (SARS-CoV-2) have shown similar ILC2 induction in the lungs.^{308,309}

Though the research in this area is early and mostly correlative, these ILCs are thought to increase inflammation and contribute to lung remodeling, potentially leading to severe lung disease following respiratory virus infection.³⁰⁶ One study showed that RSV activation of ILC2 leads to inflammation and eosinophilia in the lungs.³¹⁰ Though the research in this area is early and mostly correlative, these ILCs are thought to increase inflammation and contribute to lung remodeling, potentially leading to severe lung disease following respiratory virus infection.³⁰⁶ One study showed that RSV activation of ILC2 leads to inflammation and eosinophilia in the lungs.³¹⁰ The inflammatory role of ILC2 in the respiratory system is also evident from the marked increase in ILC2 associated with non-viral pulmonary pathologies like asthma and chronic obstructive pulmonary disease.³¹¹ Alternatively, ILCs also appear to be important in clearing respiratory viruses and reducing respiratory symptoms. Early studies of ILCs in SARS-CoV-2 patients showed an association between ILC2 reduction with increased severity of COVID-19 illness.³⁰⁹ In this way, ILCs modulate a balance between respiratory immune defense and damage modulated by hyper-inflammation.

Essential Role for Decidual NK Cells in Pregnancy and the Influence of Sex Hormones

Another subtype of ILC that are important in pregnancy are decidual natural killer cells (dNKs). dNKs are uniquely implicated in both fetal development in a normal pregnancy and immune defense against pathogen infected decidual cells. dNK cells are the main lymphocyte in the decidua basalis.^{295,312} In a healthy pregnancy, their main functions are to promote angiogenesis and vascular remodeling by secreting vascular endothelial growth factor (VEGF), direct migration of the fetal trophoblast by secreting chemokines IL-8 and IP-10 and induce fetal tolerance.^{312,313} The presence of dNK cells has been shown to be essential for spiral artery modification in mouse models, and though their role in human angiogenesis is less clear, they remain in proximity to human spiral arteries, suggesting a relationship.³¹⁴ The invasive fetal trophoblast expresses CXCR1 and CXCR3, which are receptors for ligands IL-8 and IP-10, respectively. These chemoattractants direct the migration of the trophoblast by interacting with the fetal chemokine receptors.³¹⁵ Finally, since trophoblasts do not express the classic “self” labels like HLA-A/B that protect them from immune defense, they must have another mechanism inducing tolerance. Though the exact mechanism has not yet been elucidated, dNK cells are associated with decidual CD14+ cells that induce Tregs, which are known to have a role in fetal tolerance. Furthermore, spontaneous fetal abortion, potentially via maternal immune rejection, is associated with a reduced number of dNKs and decidual Tregs.³¹⁶

dNK cells are not located in the lung tissue, so there is no currently known role in defense against respiratory viral pathogens. They have been shown to play a part in defending

against pathogens that infect the decidua, such as cytomegalovirus (CMV).²⁹⁵ Studies have shown that when exposed to CMV, dNK cells exhibit plasticity and enhance lytic function to become cytotoxic and induce apoptosis in infected cells.³¹⁷ Studies with pregnancies infected with *Listeria monocytogenes* shows a similar ability to become cytotoxic in the presence of pathogens but notes that the mechanism of killing is not like the inflammatory perforin and granzyme secretion that peripheral NK cells use. Instead, dNK cells selectively deliver antimicrobial granulysin to infected cells via direct contact, thereby killing the *Listeria* without harming the trophoblast.³¹⁸ However, they are not currently known to have a direct role against preventing *in utero* infection by respiratory viruses. SARS-CoV-2 shows only rare vertical transmission, suggesting that there is some maternal-fetal barrier immune activity against the virus. Preliminary research documents an increased number of third trimester dNK cells, as well as T cells and macrophages, in those with symptomatic COVID-19.³¹⁹

ILCs are a bridge between the innate immune response and the adaptive response to respiratory viral infections mounting a first line inflammatory response to viruses in the respiratory tract. dNK cells proliferate in response to sex hormones like prolactin to maintain healthy pregnancies and have a potential role in defending against *in utero* infections. Future directions for research on ILCs may include investigating ILC response to sex hormones that they exhibit receptors for, such as androgens and estrogens. This may help determine if uterine ILCs have a role in protecting the fetus from pathogens. Further research could also be done on dNK cells to determine the mechanism for the increased dNK cell count in pregnancies with symptomatic COVID-19 to examine whether they have a role in preventing vertical transmission. The potential transition of dNK cells from one facilitating immune tolerance to active immune defense is underexplored.

Conclusions

Pregnant people infected with pathogenic respiratory viruses, like IAV and SARS-CoV-2, are at a higher risk for hospitalization, preterm birth, stillbirth, Cesarean section and maternal mortality due to severe lung disease.^{1-3,5-8,10-12,17,34,46,320,321} In our review, we summarize the literature regarding the effect that sex hormones have on distinct lymphoid immune cell populations, the ways these immune cells behave during pregnancy, and how they act over the course of viral respiratory infections. The picture that emerges is that there is a tradeoff between the hormonal and immune environment needed to promote fetal tolerance to support a successful pregnancy and an immunologic state that is optimally equipped to clear a pathogenic respiratory virus. Immune cell populations are in balance with one another to control the trajectory of the immune response. A combination of higher Treg and Breg populations, impaired B cell lymphopoiesis and Th17 differentiation are ideal modifications to prioritize immune tolerance of the fetus but leave the pregnant individual disadvantaged to mount a rapid immune response. However, the area of confluence between sex hormones of pregnancy, such as estrogens and progestogens, and immune responses to viral respiratory infections remains understudied. This is in part due to a lack of animal models that recapitulate the natural progression of human IAV disease, endocrinology, and pregnancy. Nonhuman primates are an excellent model of pregnancy³²² and viral respiratory infections³²²⁻³²⁸, but most have differences in the hormonal cascade associated

with parturition (labor onset). A combination of *in vitro* and *in vivo* models is necessary to dissect the complex interactions between sex hormones, pregnancy and the pathogenesis of viral respiratory infections.

Acknowledgements:

We would like to thank Ms. Mindi Raker for her assistance with the graphic design of the figures. This work was supported by funding from the National Institutes of Health grants R01AI133976, R01AI145890, R01AI143265, R01HD098713 and R01AI164588 to K.A.W and the University of Washington Department of Obstetrics and Gynecology to O.C. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

References:

1. Stock SJ, Carruthers J, Calvert C, et al. SARS-CoV-2 infection and COVID-19 vaccination rates in pregnant women in Scotland. *Nat Med.* 2022.
2. Lokken EM, Huebner EM, Taylor GG, et al. Disease Severity, Pregnancy Outcomes and Maternal Deaths among Pregnant Patients with SARS-CoV-2 Infection in Washington State. *American journal of obstetrics and gynecology.* 2021.
3. Metz TD, Clifton RG, Hughes BL, et al. Association of SARS-CoV-2 Infection With Serious Maternal Morbidity and Mortality From Obstetric Complications. *JAMA : the journal of the American Medical Association.* 2022.
4. Villar J, Ariff S, Gunier RB, et al. Maternal and Neonatal Morbidity and Mortality Among Pregnant Women With and Without COVID-19 Infection: The INTERCOVID Multinational Cohort Study. *JAMA Pediatrics.* 2021.
5. Delahoy MJ, Whitaker M, O'Halloran A, et al. Characteristics and Maternal and Birth Outcomes of Hospitalized Pregnant Women with Laboratory-Confirmed COVID-19 - COVID-NET, 13 States, March 1-August 22, 2020. *MMWR Morb Mortal Wkly Rep.* 2020;69(38):1347-1354. [PubMed: 32970655]
6. Kasehagen L, Byers P, Taylor K, et al. COVID-19-Associated Deaths After SARS-CoV-2 Infection During Pregnancy - Mississippi, March 1, 2020-October 6, 2021. *MMWR Morb Mortal Wkly Rep.* 2021;70(47):1646-1648. [PubMed: 34818319]
7. Centers for Disease C, Prevention. 2009 pandemic influenza A (H1N1) in pregnant women requiring intensive care - New York City, 2009. *MMWR Morb Mortal Wkly Rep.* 2010;59(11):321-326. [PubMed: 20339343]
8. Freeman DW, Barno A. Deaths from Asian influenza associated with pregnancy. *American journal of obstetrics and gynecology.* 1959;78:1172-1175. [PubMed: 13824729]
9. Dodds L, McNeil SA, Fell DB, et al. Impact of influenza exposure on rates of hospital admissions and physician visits because of respiratory illness among pregnant women. *CMAJ : Canadian Medical Association journal = journal de l'Association medicale canadienne.* 2007;176(4):463-468.
10. Nishiura H Excess risk of stillbirth during the 1918-1920 influenza pandemic in Japan. *Eur J Obstet Gynecol Reprod Biol.* 2009;147(1):115. [PubMed: 19679387]
11. Jamieson DJ, Honein MA, Rasmussen SA, et al. H1N1 2009 influenza virus infection during pregnancy in the USA. *Lancet.* 2009;374(9688):451-458. [PubMed: 19643469]
12. Centers for Disease C, Prevention. Maternal and infant outcomes among severely ill pregnant and postpartum women with 2009 pandemic influenza A (H1N1)--United States, April 2009-August 2010. *MMWR Morb Mortal Wkly Rep.* 2011;60(35):1193-1196. [PubMed: 21900872]
13. Centers for Disease C, Prevention. Novel influenza A (H1N1) virus infections in three pregnant women - United States, April-May 2009. *MMWR Morb Mortal Wkly Rep.* 2009;58(18):497-500. [PubMed: 19444154]
14. Jimenez MF, El Beitune P, Salcedo MP, Von Ameln AV, Mastalir FP, Braun LD. Outcomes for pregnant women infected with the influenza A (H1N1) virus during the 2009 pandemic in Porto Alegre, Brazil. *Int J Gynaecol Obstet.* 2010;111(3):217-219. [PubMed: 20801449]

15. Siston AM, Rasmussen SA, Honein MA, et al. Pandemic 2009 influenza A(H1N1) virus illness among pregnant women in the United States. *JAMA : the journal of the American Medical Association*. 2010;303(15):1517–1525. [PubMed: 20407061]
16. Hardy JM, Azarowicz EN, Mannini A, Medearis DN Jr., Cooke RE. The effect of Asian influenza on the outcome of pregnancy, Baltimore, 1957–1958. *Am J Public Health Nations Health*. 1961;51:1182–1188. [PubMed: 13711529]
17. Mosby LG, Rasmussen SA, Jamieson DJ. 2009 pandemic influenza A (H1N1) in pregnancy: a systematic review of the literature. *American journal of obstetrics and gynecology*. 2011;205(1):10–18. [PubMed: 21345415]
18. Solano ME, Arck PC. Steroids, Pregnancy and Fetal Development. *Front Immunol*. 2019;10:3017. [PubMed: 32038609]
19. Tal R, Taylor HS. Endocrinology of Pregnancy. In: Feingold KR, Anawalt B, Boyce A, et al. , eds. South Dartmouth, Massachusetts: MDText.com, Inc.: <https://www.ncbi.nlm.nih.gov/books/NBK278962/>. Accessed February 14, 2022.
20. Soldin OP, Guo T, Weiderpass E, Tractenberg RE, Hilakivi-Clarke L, Soldin SJ. Steroid hormone levels in pregnancy and 1 year postpartum using isotope dilution tandem mass spectrometry. *Fertility and sterility*. 2005;84(3):701–710. [PubMed: 16169406]
21. Trotter A, Maier L, Grill H-J, Wudy SA, Pohlandt F. 17 β -Estradiol and Progesterone Supplementation in Extremely Low-Birth-Weight Infants. *Pediatric Research*. 1999;45(4):489–493. [PubMed: 10203139]
22. Ostrom KM. A review of the hormone prolactin during lactation. *Prog Food Nutr Sci*. 1990;14(1):1–43. [PubMed: 2092340]
23. Haning RV Jr., Goldsmith LT, Seifer DB, et al. Relaxin secretion in in vitro fertilization pregnancies. *American journal of obstetrics and gynecology*. 1996;174(1 Pt 1):233–240. [PubMed: 8572013]
24. Muzzio D, Zygmunt M, Jensen F. The role of pregnancy-associated hormones in the development and function of regulatory B cells. *Front Endocrinol (Lausanne)*. 2014;5:39. [PubMed: 24744750]
25. Polanczyk MJ, Carson BD, Subramanian S, et al. Cutting edge: estrogen drives expansion of the CD4+CD25+ regulatory T cell compartment. *J Immunol*. 2004;173(4):2227–2230. [PubMed: 15294932]
26. Polanczyk MJ, Hopke C, Huan J, Vandenbark AA, Offner H. Enhanced FoxP3 expression and Treg cell function in pregnant and estrogen-treated mice. *J Neuroimmunol*. 2005;170(1–2):85–92. [PubMed: 16253347]
27. Mao G, Wang J, Kang Y, et al. Progesterone increases systemic and local uterine proportions of CD4+CD25+ Treg cells during midterm pregnancy in mice. *Endocrinology*. 2010;151(11):5477–5488. [PubMed: 20844003]
28. Kim HM, Kang YM, Song BM, Kim HS, Seo SH. The 2009 pandemic H1N1 influenza virus is more pathogenic in pregnant mice than seasonal H1N1 influenza virus. *Viral Immunol*. 2012;25(5):402–410. [PubMed: 22985287]
29. Peteranderl C, Herold S, Schmoldt C. Human Influenza Virus Infections. *Semin Respir Crit Care Med*. 2016;37(4):487–500. [PubMed: 27486731]
30. Bourne AW. Influenza: pregnancy, labour, the puerperium, and diseases of women. In: Crookshank FG, ed. *Influenza: essays by several authors*. London: Heinemann; 1922:433–443.
31. Harris JW. Influenza occurring in pregnant women: a statistical study of thirteen hundred and fifty cases. *JAMA : the journal of the American Medical Association*. 1919;72:978–980.
32. Pierce M, Kurinczuk JJ, Spark P, Brocklehurst P, Knight M, Ukoss. Perinatal outcomes after maternal 2009/H1N1 infection: national cohort study. *BMJ*. 2011;342:d3214. [PubMed: 21672992]
33. Louie JK, Acosta M, Jamieson DJ, Honein MA, California Pandemic Working G. Severe 2009 H1N1 influenza in pregnant and postpartum women in California. *N Engl J Med*. 2010;362(1):27–35. [PubMed: 20032319]
34. Ribeiro AF, Pellini ACG, Kitagawa BY, et al. Severe influenza A(H1N1)pdm09 in pregnant women and neonatal outcomes, State of Sao Paulo, Brazil, 2009. *PLoS one*. 2018;13(3):e0194392. [PubMed: 29579099]

35. Mertz D, Kim TH, Johnstone J, et al. Populations at risk for severe or complicated influenza illness: systematic review and meta-analysis. *BMJ*. 2013;347:f5061. [PubMed: 23974637]
36. Serfling RE, Sherman IL, Houseworth WJ. Excess pneumonia-influenza mortality by age and sex in three major influenza A2 epidemics, United States, 1957–58, 1960 and 1963. *Am J Epidemiol*. 1967;86(2):433–441. [PubMed: 6058395]
37. Klein SL, Passaretti C, Anker M, Olukoya P, Pekosz A. The impact of sex, gender and pregnancy on 2009 H1N1 disease. *Biology of Sex Differences*. 2010;1(1):5. [PubMed: 21208468]
38. Klein S, Pekosz A, Passaretti C, Anker M, Olukoya P. Sex, gender and influenza. Geneva: World Health Organization. 2010:1–58.
39. Update: WHO-confirmed human cases of avian influenza A (H5N1) infection, November 2003–May 2008. *Wkly Epidemiol Rec*. 2008;83(46):415–420. [PubMed: 19009716]
40. Kumar A, Zarychanski R, Pinto R, et al. Critically ill patients with 2009 influenza A(H1N1) infection in Canada. *JAMA : the journal of the American Medical Association*. 2009;302(17):1872–1879. [PubMed: 19822627]
41. Eshima N, Tokumaru O, Hara S, et al. Sex- and age-related differences in morbidity rates of 2009 pandemic influenza A H1N1 virus of swine origin in Japan. *PloS one*. 2011;6(4):e19409. [PubMed: 21559366]
42. Potluri T, Fink AL, Sylvia KE, et al. Age-associated changes in the impact of sex steroids on influenza vaccine responses in males and females. *NPJ Vaccines*. 2019;4:29. [PubMed: 31312529]
43. Engler RJ, Nelson MR, Klote MM, et al. Half- vs full-dose trivalent inactivated influenza vaccine (2004–2005): age, dose, and sex effects on immune responses. *Arch Intern Med*. 2008;168(22):2405–2414. [PubMed: 19064822]
44. Fink AL, Engle K, Ursin RL, Tang WY, Klein SL. Biological sex affects vaccine efficacy and protection against influenza in mice. *Proc Natl Acad Sci U S A*. 2018;115(49):12477–12482. [PubMed: 30455317]
45. Data on COVID-19 during Pregnancy. 2020; <https://www.cdc.gov/coronavirus/2019-ncov/cases-updates/special-populations/pregnancy-data-on-covid-19.html>. Accessed November 4, 2020.
46. Woodworth KR, O'Malley Olsen E, Neelam V, et al. Birth and Infant Outcomes Following Laboratory-Confirmed SARS-CoV-2 Infection in Pregnancy — SET-NET, 16 Jurisdictions, March 29–October 14, 2020. *MMWR Morb Mortal Wkly Rep*. 2020;69.
47. Panagiotakopoulos L, Myers TR, Gee J, et al. SARS-CoV-2 Infection Among Hospitalized Pregnant Women: Reasons for Admission and Pregnancy Characteristics - Eight U.S. Health Care Centers, March 1-May 30, 2020. *MMWR Morb Mortal Wkly Rep*. 2020;69(38):1355–1359. [PubMed: 32970660]
48. Pierce-Williams RAM, Burd J, Felder L, et al. Clinical course of severe and critical COVID-19 in hospitalized pregnancies: a US cohort study. *Am J Obstet Gynecol MFM*. 2020.
49. Lokken EM, Huebner EM, Taylor GG, et al. Disease Severity, Pregnancy Outcomes and Maternal Deaths among Pregnant Patients with SARS-CoV-2 Infection in Washington State. *American journal of obstetrics and gynecology*. 2021. DOI: 10.1016/j.ajog.2020.12.1221.
50. Afshar Y, Gaw SL, Flaherman VJ, et al. Clinical Presentation of Coronavirus Disease 2019 (COVID-19) in Pregnant and Recently Pregnant People. *Obstet Gynecol*. 2020.
51. Ellington S, Strid P, Tong VT, et al. Characteristics of Women of Reproductive Age with Laboratory-Confirmed SARS-CoV-2 Infection by Pregnancy Status - United States, January 22–June 7, 2020. *MMWR Morb Mortal Wkly Rep*. 2020;69(25):769–775. [PubMed: 32584795]
52. Zambrano LD, Ellington S, Strid P, et al. Update: Characteristics of Symptomatic Women of Reproductive Age with Laboratory-Confirmed SARS-CoV-2 Infection by Pregnancy Status - United States, January 22–October 3, 2020. *MMWR Morb Mortal Wkly Rep*. 2020;69(44):1641–1647. [PubMed: 33151921]
53. DeSisto CL, Wallace B, Simeone RM, et al. Risk for Stillbirth Among Women With and Without COVID-19 at Delivery Hospitalization - United States, March 2020–September 2021. *MMWR Morb Mortal Wkly Rep*. 2021;70(47):1640–1645. [PubMed: 34818318]
54. Peckham H, de Grijter NM, Raine C, et al. Male sex identified by global COVID-19 meta-analysis as a risk factor for death and ITU admission. *Nature Communications*. 2020;11(1):6317.

55. Takahashi T, Ellingson MK, Wong P, et al. Sex differences in immune responses that underlie COVID-19 disease outcomes. *Nature*. 2020;588(7837):315–320. [PubMed: 32846427]
56. Littauer EQ, Esser ES, Antao OQ, Vassilieva EV, Compans RW, Skountzou I. H1N1 influenza virus infection results in adverse pregnancy outcomes by disrupting tissue-specific hormonal regulation. *PLoS Pathog*. 2017;13(11):e1006757. [PubMed: 29176767]
57. Shah NM, Lai PF, Imami N, Johnson MR. Progesterone-Related Immune Modulation of Pregnancy and Labor. *Frontiers in Endocrinology*. 2019;10.
58. McLean M, Bisits A, Davies J, Woods R, Lowry P, Smith R. A placental clock controlling the length of human pregnancy. *Nat Med*. 1995;1(5):460–463. [PubMed: 7585095]
59. Smith R Corticotropin-releasing hormone and the fetoplacental clock: an Australian perspective. *American journal of obstetrics and gynecology*. 1999;180(1 Pt 3):S269–271. [PubMed: 9914632]
60. Gabriel G, Arck PC. Sex, Immunity and Influenza. *J Infect Dis*. 2014;209(suppl 3):S93–S99. [PubMed: 24966196]
61. Pazos MA, Kraus TA, Munoz-Fontela C, Moran TM. Estrogen mediates innate and adaptive immune alterations to influenza infection in pregnant mice. *PLoS One*. 2012;7(7):e40502. [PubMed: 22792357]
62. Beová K, Hancková M, Ko i K, Kúdelová M, Betáková T. T cells and their function in the immune response to viruses. *Acta Virol*. 2020;64(02):131–143. [PubMed: 32551782]
63. Nussing S, Sant S, Koutsakos M, Subbarao K, Nguyen THO, Kedzierska K. Innate and adaptive T cells in influenza disease. *Front Med*. 2018;12(1):34–47. [PubMed: 29352371]
64. Sakaguchi S, Yamaguchi T, Nomura T, Ono M. Regulatory T cells and immune tolerance. *Cell*. 2008;133(5):775–787. [PubMed: 18510923]
65. Smigiel KS, Srivastava S, Stolley JM, Campbell DJ. Regulatory T-cell homeostasis: steady-state maintenance and modulation during inflammation. *Immunol Rev*. 2014;259(1):40–59. [PubMed: 24712458]
66. Fontenot JD, Gavin MA, Rudensky AY. Foxp3 programs the development and function of CD4+CD25+ regulatory T cells. *Nat Immunol*. 2003;4(4):330–336. [PubMed: 12612578]
67. Williams LM, Rudensky AY. Maintenance of the Foxp3-dependent developmental program in mature regulatory T cells requires continued expression of Foxp3. *Nat Immunol*. 2007;8(3):277–284. [PubMed: 17220892]
68. Bennett CL, Christie J, Ramsdell F, et al. The immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome (IPEX) is caused by mutations of FOXP3. *Nat Genet*. 2001;27(1):20–21. [PubMed: 11137993]
69. Brunkow ME, Jeffery EW, Hjerrild KA, et al. Disruption of a new forkhead/winged-helix protein, scurfy, results in the fatal lymphoproliferative disorder of the scurfy mouse. *Nat Genet*. 2001;27(1):68–73. [PubMed: 11138001]
70. Sanchez Rodriguez R, Pauli ML, Neuhaus IM, et al. Memory regulatory T cells reside in human skin. *J Clin Invest*. 2014;124(3):1027–1036. [PubMed: 24509084]
71. Li C, DiSpirito JR, Zemmour D, et al. TCR Transgenic Mice Reveal Stepwise, Multi-site Acquisition of the Distinctive Fat-Treg Phenotype. *Cell*. 2018;174(2):285–299 e212. [PubMed: 29887374]
72. Brincks EL, Roberts AD, Cookenham T, et al. Antigen-specific memory regulatory CD4+Foxp3+ T cells control memory responses to influenza virus infection. *J Immunol*. 2013;190(7):3438–3446. [PubMed: 23467933]
73. Hadis U, Wahl B, Schulz O, et al. Intestinal tolerance requires gut homing and expansion of FoxP3+ regulatory T cells in the lamina propria. *Immunity*. 2011;34(2):237–246. [PubMed: 21333554]
74. Soerens AG, Da Costa A, Lund JM. Regulatory T cells are essential to promote proper CD4 T-cell priming upon mucosal infection. *Mucosal Immunol*. 2016;9(6):1395–1406. [PubMed: 27007674]
75. Zheng SG, Wang JH, Gray JD, Soucier H, Horwitz DA. Natural and induced CD4+CD25+ cells educate CD4+CD25- cells to develop suppressive activity: the role of IL-2, TGF-beta, and IL-10. *J Immunol*. 2004;172(9):5213–5221. [PubMed: 15100259]
76. Josefowicz SZ, Niec RE, Kim HY, et al. Extrathymically generated regulatory T cells control mucosal TH2 inflammation. *Nature*. 2012;482(7385):395–399. [PubMed: 22318520]

77. Shevach EM, Thornton AM. tTregs, pTregs, and iTregs: similarities and differences. *Immunol Rev.* 2014;259(1):88–102. [PubMed: 24712461]
78. Thornton AM, Korty PE, Tran DQ, et al. Expression of Helios, an Ikaros transcription factor family member, differentiates thymic-derived from peripherally induced Foxp3+ T regulatory cells. *J Immunol.* 2010;184(7):3433–3441. [PubMed: 20181882]
79. Pratama A, Schnell A, Mathis D, Benoist C. Developmental and cellular age direct conversion of CD4+ T cells into RORgamma+ or Helios+ colon Treg cells. *J Exp Med.* 2020;217(1).
80. Szurek E, Cebula A, Wojciech L, et al. Differences in Expression Level of Helios and Neuropilin-1 Do Not Distinguish Thymus-Derived from Extrathymically-Induced CD4+Foxp3+ Regulatory T Cells. *PLoS one.* 2015;10(10):e0141161. [PubMed: 26495986]
81. Li MO, Wan YY, Flavell RA. T cell-produced transforming growth factor-beta1 controls T cell tolerance and regulates Th1- and Th17-cell differentiation. *Immunity.* 2007;26(5):579–591. [PubMed: 17481928]
82. Rubtsov YP, Rasmussen JP, Chi EY, et al. Regulatory T cell-derived interleukin-10 limits inflammation at environmental interfaces. *Immunity.* 2008;28(4):546–558. [PubMed: 18387831]
83. Gondek DC, Lu LF, Quezada SA, Sakaguchi S, Noelle RJ. Cutting edge: contact-mediated suppression by CD4+CD25+ regulatory cells involves a granzyme B-dependent, perforin-independent mechanism. *J Immunol.* 2005;174(4):1783–1786. [PubMed: 15699103]
84. Pandiyan P, Zheng L, Ishihara S, Reed J, Lenardo MJ. CD4+CD25+Foxp3+ regulatory T cells induce cytokine deprivation-mediated apoptosis of effector CD4+ T cells. *Nat Immunol.* 2007;8(12):1353–1362. [PubMed: 17982458]
85. Loebbermann J, Thornton H, Durant L, et al. Regulatory T cells expressing granzyme B play a critical role in controlling lung inflammation during acute viral infection. *Mucosal Immunol.* 2012;5(2):161–172. [PubMed: 22236998]
86. Burzyn D, Kuswanto W, Kolodin D, et al. A special population of regulatory T cells potentiates muscle repair. *Cell.* 2013;155(6):1282–1295. [PubMed: 24315098]
87. Dial CF, Tune MK, Doerschuk CM, Mock JR. Foxp3(+) Regulatory T Cell Expression of Keratinocyte Growth Factor Enhances Lung Epithelial Proliferation. *Am J Respir Cell Mol Biol.* 2017;57(2):162–173. [PubMed: 28296468]
88. Arpaia N, Green JA, Moltedo B, et al. A Distinct Function of Regulatory T Cells in Tissue Protection. *Cell.* 2015;162(5):1078–1089. [PubMed: 26317471]
89. Varanasi SK, Rajasagi NK, Jaggi U, Rouse BT. Role of IL-18 induced Amphiregulin expression on virus induced ocular lesions. *Mucosal Immunol.* 2018;11(6):1705–1715. [PubMed: 30087443]
90. Huang X, Cai Y, Ding M, Zheng B, Sun H, Zhou J. Human chorionic gonadotropin promotes recruitment of regulatory T cells in endometrium by inducing chemokine CCL2. *J Reprod Immunol.* 2020;137:102856. [PubMed: 31809964]
91. Liu X, Ma D, Wang W, et al. Intrauterine administration of human chorionic gonadotropin improves the live birth rates of patients with repeated implantation failure in frozen-thawed blastocyst transfer cycles by increasing the percentage of peripheral regulatory T cells. *Arch Gynecol Obstet.* 2019;299(4):1165–1172. [PubMed: 30659362]
92. Diao LH, Li GG, Zhu YC, et al. Human chorionic gonadotropin potentially affects pregnancy outcome in women with recurrent implantation failure by regulating the homing preference of regulatory T cells. *American journal of reproductive immunology (New York, NY : 1989).* 2017;77(3).
93. Schumacher A, Heinze K, Witte J, et al. Human chorionic gonadotropin as a central regulator of pregnancy immune tolerance. *J Immunol.* 2013;190(6):2650–2658. [PubMed: 23396945]
94. Zhao JX, Zeng YY, Liu Y. Fetal alloantigen is responsible for the expansion of the CD4(+)CD25(+) regulatory T cell pool during pregnancy. *J Reprod Immunol.* 2007;75(2):71–81. [PubMed: 17686527]
95. Huang N, Chi H, Qiao J. Role of Regulatory T Cells in Regulating Fetal-Maternal Immune Tolerance in Healthy Pregnancies and Reproductive Diseases. *Front Immunol.* 2020;11:1023. [PubMed: 32676072]
96. Jorgensen N, Persson G, Hviid TVF. The Tolerogenic Function of Regulatory T Cells in Pregnancy and Cancer. *Front Immunol.* 2019;10:911. [PubMed: 31134056]

97. Robertson SA, Prins JR, Sharkey DJ, Moldenhauer LM. Seminal fluid and the generation of regulatory T cells for embryo implantation. *American journal of reproductive immunology* (New York, NY : 1989). 2013;69(4):315–330.
98. Guerin LR, Moldenhauer LM, Prins JR, Bromfield JJ, Hayball JD, Robertson SA. Seminal fluid regulates accumulation of FOXP3+ regulatory T cells in the preimplantation mouse uterus through expanding the FOXP3+ cell pool and CCL19-mediated recruitment. *Biol Reprod*. 2011;85(2):397–408. [PubMed: 21389340]
99. Zhou J, Wang Z, Zhao X, Wang J, Sun H, Hu Y. An increase of Treg cells in the peripheral blood is associated with a better in vitro fertilization treatment outcome. *American journal of reproductive immunology* (New York, NY : 1989). 2012;68(2):100–106.
100. Shima T, Sasaki Y, Itoh M, et al. Regulatory T cells are necessary for implantation and maintenance of early pregnancy but not late pregnancy in allogeneic mice. *J Reprod Immunol*. 2010;85(2):121–129. [PubMed: 20439117]
101. Arruvito L, Sanz M, Banham AH, Fainboim L. Expansion of CD4+CD25+and FOXP3+ regulatory T cells during the follicular phase of the menstrual cycle: implications for human reproduction. *J Immunol*. 2007;178(4):2572–2578. [PubMed: 17277167]
102. Sasaki Y, Darmochwal-Kolarz D, Suzuki D, et al. Proportion of peripheral blood and decidual CD4(+) CD25(bright) regulatory T cells in pre-eclampsia. *Clin Exp Immunol*. 2007;149(1):139–145. [PubMed: 17459078]
103. Santner-Nanan B, Peek MJ, Khanam R, et al. Systemic increase in the ratio between Foxp3+ and IL-17-producing CD4+ T cells in healthy pregnancy but not in preeclampsia. *J Immunol*. 2009;183(11):7023–7030. [PubMed: 19915051]
104. Mjosberg J, Svensson J, Johansson E, et al. Systemic reduction of functionally suppressive CD4dimCD25highFoxp3+ Tregs in human second trimester pregnancy is induced by progesterone and 17beta-estradiol. *J Immunol*. 2009;183(1):759–769. [PubMed: 19535629]
105. Shirshv SV, Nekrasova IV, Gorbunova OL, Orlova EG. Effect of Estriol, Chorionic Gonadotropin, and Oncostatin M on the Expression of Recombinase RAG-1 in Regulatory T Lymphocyte Subpopulations. *Bull Exp Biol Med*. 2019;167(1):57–61. [PubMed: 31177451]
106. Lee JH, Ulrich B, Cho J, Park J, Kim CH. Progesterone promotes differentiation of human cord blood fetal T cells into T regulatory cells but suppresses their differentiation into Th17 cells. *J Immunol*. 2011;187(4):1778–1787. [PubMed: 21768398]
107. Engler JB, Kursawe N, Solano ME, et al. Glucocorticoid receptor in T cells mediates protection from autoimmunity in pregnancy. *Proc Natl Acad Sci U S A*. 2017;114(2):E181–E190. [PubMed: 28049829]
108. Qiu T, Teng Y, Wang Y, Xu L. Adoptive transfer of transforming growth factor- β -induced CD4+CD25+ regulatory T cells prevents immune response-mediated spontaneous abortion. *Reprod Fertil Dev*. 2015.
109. Wang WJ, Liu FJ, Xin L, et al. Adoptive transfer of pregnancy-induced CD4+CD25+ regulatory T cells reverses the increase in abortion rate caused by interleukin 17 in the CBA/JxBALB/c mouse model. *Human reproduction* (Oxford, England). 2014;29(5):946–952.
110. Hsu P, Santner-Nanan B, Dahlstrom JE, et al. Altered decidual DC-SIGN+ antigen-presenting cells and impaired regulatory T-cell induction in preeclampsia. *Am J Pathol*. 2012;181(6):2149–2160. [PubMed: 23063509]
111. Tsuda S, Zhang X, Hamana H, et al. Clonally Expanded Decidual Effector Regulatory T Cells Increase in Late Gestation of Normal Pregnancy, but Not in Preeclampsia, in Humans. *Front Immunol*. 2018;9:1934. [PubMed: 30197648]
112. Tsuda S, Nakashima A, Shima T, Saito S. New Paradigm in the Role of Regulatory T Cells During Pregnancy. *Front Immunol*. 2019;10:573. [PubMed: 30972068]
113. Sadeghi A, Tahmasebi S, Mahmood A, et al. Th17 and Treg cells function in SARS-CoV2 patients compared with healthy controls. *J Cell Physiol*. 2021;236(4):2829–2839. [PubMed: 32926425]
114. León B, Bradley JE, Lund FE, Randall TD, Ballesteros-Tato A. FoxP3+ regulatory T cells promote influenza-specific Tfh responses by controlling IL-2 availability. *Nature Communications*. 2014;5(1):3495.

115. Fulton RB, Meyerholz DK, Varga SM. Foxp3+ CD4 regulatory T cells limit pulmonary immunopathology by modulating the CD8 T cell response during respiratory syncytial virus infection. *J Immunol.* 2010;185(4):2382–2392. [PubMed: 20639494]
116. Vermillion MS, Ursin RL, Attreed SE, Klein SL. Estriol Reduces Pulmonary Immune Cell Recruitment and Inflammation to Protect Female Mice From Severe Influenza. *Endocrinology.* 2018;159(9):3306–3320. [PubMed: 30032246]
117. Leon B, Bradley JE, Lund FE, Randall TD, Ballesteros-Tato A. FoxP3+ regulatory T cells promote influenza-specific Tfh responses by controlling IL-2 availability. *Nat Commun.* 2014;5:3495. [PubMed: 24633065]
118. Lu C, Chen W. Influenza virus infection selectively triggers the accumulation and persistence of more potent Helios-expressing Foxp3(+) regulatory T cells in the lungs. *Immunol Cell Biol.* 2021;99(10):1011–1025. [PubMed: 34251701]
119. Williams-Bey Y, Jiang J, Murasko DM. Expansion of regulatory T cells in aged mice following influenza infection. *Mech Ageing Dev.* 2011;132(4):163–170. [PubMed: 21414341]
120. Christiaansen AF, Syed MA, Ten Eyck PP, et al. Altered Treg and cytokine responses in RSV-infected infants. *Pediatr Res.* 2016;80(5):702–709. [PubMed: 27486703]
121. Vick SC, Frutoso M, Mair F, et al. A regulatory T cell signature distinguishes the immune landscape of COVID-19 patients from those with other respiratory infections. *Sci Adv.* 2021;7(46):eabj0274. [PubMed: 34757794]
122. Gao M, Liu Y, Guo M, et al. Regulatory CD4(+) and CD8(+) T cells are negatively correlated with CD4(+)/CD8(+) T cell ratios in patients acutely infected with SARS-CoV-2. *J Leukoc Biol.* 2021;109(1):91–97. [PubMed: 32930458]
123. Galvan-Pena S, Leon J, Chowdhary K, et al. Profound Treg perturbations correlate with COVID-19 severity. *Proc Natl Acad Sci U S A.* 2021;118(37).
124. Harb H, Benamar M, Lai PS, et al. Notch4 signaling limits regulatory T-cell-mediated tissue repair and promotes severe lung inflammation in viral infections. *Immunity.* 2021;54(6):1186–1199 e1187. [PubMed: 33915108]
125. Szabo PA, Dogra P, Gray JI, et al. Longitudinal profiling of respiratory and systemic immune responses reveals myeloid cell-driven lung inflammation in severe COVID-19. *Immunity.* 2021;54(4):797–814 e796. [PubMed: 33765436]
126. Graham JB, Swarts JL, Leist SR, et al. Baseline T cell immune phenotypes predict virologic and disease control upon SARS-CoV infection in Collaborative Cross mice. *PLoS Pathog.* 2021;17(1):e1009287. [PubMed: 33513210]
127. Foo SS, Cambou MC, Mok T, et al. The systemic inflammatory landscape of COVID-19 in pregnancy: Extensive serum proteomic profiling of mother-infant dyads with in utero SARS-CoV-2. *Cell Rep Med.* 2021;2(11):100453. [PubMed: 34723226]
128. Marcelin G, Aldridge JR, Duan S, et al. Fatal outcome of pandemic H1N1 2009 influenza virus infection is associated with immunopathology and impaired lung repair, not enhanced viral burden, in pregnant mice. *Journal of virology.* 2011;85(21):11208–11219. [PubMed: 21865394]
129. Wang F, Hou H, Luo Y, et al. The laboratory tests and host immunity of COVID-19 patients with different severity of illness. *JCI Insight.* 2020;5(10).
130. Qin C, Zhou L, Hu Z, et al. Dysregulation of immune response in patients with COVID-19 in Wuhan, China. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America.* 2020.
131. De Biasi S, Meschiari M, Gibellini L, et al. Marked T cell activation, senescence, exhaustion and skewing towards TH17 in patients with COVID-19 pneumonia. *Nat Commun.* 2020;11(1):3434. [PubMed: 32632085]
132. Afshan G, Afzal N, Qureshi S. CD4+CD25(hi) regulatory T cells in healthy males and females mediate gender difference in the prevalence of autoimmune diseases. *Clin Lab.* 2012;58(5–6):567–571. [PubMed: 22783590]
133. Fijak M, Schneider E, Klug J, et al. Testosterone replacement effectively inhibits the development of experimental autoimmune orchitis in rats: evidence for a direct role of testosterone on regulatory T cell expansion. *J Immunol.* 2011;186(9):5162–5172. [PubMed: 21441459]

134. Walecki M, Eisel F, Klug J, et al. Androgen receptor modulates Foxp3 expression in CD4+CD25+Foxp3+ regulatory T-cells. *Mol Biol Cell*. 2015;26(15):2845–2857. [PubMed: 26063731]
135. Singh RP, Bischoff DS. Sex Hormones and Gender Influence the Expression of Markers of Regulatory T Cells in SLE Patients. *Front Immunol*. 2021;12:619268. [PubMed: 33746959]
136. Vasanthakumar A, Chisanga D, Blume J, et al. Sex-specific adipose tissue imprinting of regulatory T cells. *Nature*. 2020;579(7800):581–585. [PubMed: 32103173]
137. Ishikawa A, Wada T, Nishimura S, et al. Estrogen regulates sex-specific localization of regulatory T cells in adipose tissue of obese female mice. *PLoS One*. 2020;15(4):e0230885. [PubMed: 32240221]
138. Park H, Li Z, Yang XO, et al. A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. *Nat Immunol*. 2005;6(11):1133–1141. [PubMed: 16200068]
139. Ivanov II, McKenzie BS, Zhou L, et al. The orphan nuclear receptor ROR γ directs the differentiation program of proinflammatory IL-17+ T helper cells. *Cell*. 2006;126(6):1121–1133. [PubMed: 16990136]
140. Chalmin F, Mignot G, Bruchard M, et al. Stat3 and Gfi-1 transcription factors control Th17 cell immunosuppressive activity via the regulation of ectonucleotidase expression. *Immunity*. 2012;36(3):362–373. [PubMed: 22406269]
141. Longhi MS, Moss A, Bai A, et al. Characterization of human CD39+ Th17 cells with suppressor activity and modulation in inflammatory bowel disease. *PloS one*. 2014;9(2):e87956. [PubMed: 24505337]
142. Hall OJ, Limjunyawong N, Vermillion MS, et al. Progesterone-Based Therapy Protects Against Influenza by Promoting Lung Repair and Recovery in Females. *PLoS Pathog*. 2016;12(9):e1005840. [PubMed: 27631986]
143. Black A, Bhaumik S, Kirkman RL, Weaver CT, Randolph DA. Developmental regulation of Th17-cell capacity in human neonates. *Eur J Immunol*. 2012;42(2):311–319. [PubMed: 22101893]
144. AbdulHussain G, Azizieh F, Makhseed M, Raghupathy R. Effects of Progesterone, Dydrogesterone and Estrogen on the Production of Th1/Th2/Th17 Cytokines by Lymphocytes from Women with Recurrent Spontaneous Miscarriage. *J Reprod Immunol*. 2020;140:103132. [PubMed: 32380371]
145. Maeda Y, Ohtsuka H, Tomioka M, Oikawa M. Effect of progesterone on Th1/Th2/Th17 and regulatory T cell-related genes in peripheral blood mononuclear cells during pregnancy in cows. *Vet Res Commun*. 2013;37(1):43–49. [PubMed: 23203561]
146. Piccinni MP, Lombardelli L, Logiodice F, Kullolli O, Maggi E, Barkley MS. Medroxyprogesterone Acetate Decreases Th1, Th17, and Increases Th22 Responses via AHR Signaling Which Could Affect Susceptibility to Infections and Inflammatory Disease. *Front Immunol*. 2019;10:642. [PubMed: 31001262]
147. Lasarte S, Elsner D, Guia-Gonzalez M, et al. Female sex hormones regulate the Th17 immune response to sperm and *Candida albicans*. *Human reproduction (Oxford, England)*. 2013;28(12):3283–3291.
148. Relloso M, Aragonese-Fenoll L, Lasarte S, et al. Estradiol impairs the Th17 immune response against *Candida albicans*. *J Leukoc Biol*. 2012;91(1):159–165. [PubMed: 21965175]
149. Newcomb DC, Cephus JY, Boswell MG, et al. Estrogen and progesterone decrease let-7f microRNA expression and increase IL-23/IL-23 receptor signaling and IL-17A production in patients with severe asthma. *J Allergy Clin Immunol*. 2015;136(4):1025–1034 e1011. [PubMed: 26242299]
150. Anipindi VC, Bagri P, Roth K, et al. Estradiol Enhances CD4+ T-Cell Anti-Viral Immunity by Priming Vaginal DCs to Induce Th17 Responses via an IL-1-Dependent Pathway. *PLoS Pathog*. 2016;12(5):e1005589. [PubMed: 27148737]
151. Garnier L, Laffont S, Lelu K, Yogev N, Waisman A, Guery JC. Estrogen Signaling in Bystander Foxp3(neg) CD4(+) T Cells Suppresses Cognate Th17 Differentiation in Trans and Protects from Central Nervous System Autoimmunity. *J Immunol*. 2018;201(11):3218–3228. [PubMed: 30355786]

152. Hoh BL, Rojas K, Lin L, et al. Estrogen Deficiency Promotes Cerebral Aneurysm Rupture by Upregulation of Th17 Cells and Interleukin-17A Which Downregulates E-Cadherin. *J Am Heart Assoc.* 2018;7(8).
153. Andersson A, Stubelius A, Karlsson MN, et al. Estrogen regulates T helper 17 phenotype and localization in experimental autoimmune arthritis. *Arthritis Res Ther.* 2015;17:32. [PubMed: 25888974]
154. Haghmorad D, Amini AA, Mahmoudi MB, Rastin M, Hosseini M, Mahmoudi M. Pregnancy level of estrogen attenuates experimental autoimmune encephalomyelitis in both ovariectomized and pregnant C57BL/6 mice through expansion of Treg and Th2 cells. *J Neuroimmunol.* 2014;277(1–2):85–95. [PubMed: 25457839]
155. Nekrasova I, Shirshv S. Estradiol in regulation of cell-mediated immune reactions in multiple sclerosis. *J Neuroimmunol.* 2020;349:577421. [PubMed: 33032016]
156. Bagri P, Ghasemi R, McGrath JJC, et al. Estradiol Enhances Antiviral CD4(+) Tissue-Resident Memory T Cell Responses following Mucosal Herpes Simplex Virus 2 Vaccination through an IL-17-Mediated Pathway. *Journal of virology.* 2020;95(1).
157. Fuseini H, Cephus JY, Wu P, et al. ERalpha Signaling Increased IL-17A Production in Th17 Cells by Upregulating IL-23R Expression, Mitochondrial Respiration, and Proliferation. *Front Immunol.* 2019;10:2740. [PubMed: 31849948]
158. Chen RY, Fan YM, Zhang Q, et al. Estradiol inhibits Th17 cell differentiation through inhibition of RORgammaT transcription by recruiting the ERalpha/REA complex to estrogen response elements of the RORgammaT promoter. *J Immunol.* 2015;194(8):4019–4028. [PubMed: 25769926]
159. Nakashima A, Ito M, Yoneda S, Shiozaki A, Hidaka T, Saito S. Circulating and decidual Th17 cell levels in healthy pregnancy. *American journal of reproductive immunology (New York, NY : 1989).* 2010;63(2):104–109.
160. Wang WJ, Hao CF, Yi L, et al. Increased prevalence of T helper 17 (Th17) cells in peripheral blood and decidua in unexplained recurrent spontaneous abortion patients. *J Reprod Immunol.* 2010;84(2):164–170. [PubMed: 20106535]
161. Martinez FF, Knubel CP, Sanchez MC, Cervi L, Motran CC. Pregnancy-specific glycoprotein 1a activates dendritic cells to provide signals for Th17-, Th2-, and Treg-cell polarization. *Eur J Immunol.* 2012;42(6):1573–1584. [PubMed: 22678910]
162. Pongcharoen S, Supalap K. Interleukin-17 increased progesterone secretion by JEG-3 human choriocarcinoma cells. *American journal of reproductive immunology (New York, NY : 1989).* 2009;61(4):261–264.
163. Wu HX, Jin LP, Xu B, Liang SS, Li DJ. Decidual stromal cells recruit Th17 cells into decidua to promote proliferation and invasion of human trophoblast cells by secreting IL-17. *Cell Mol Immunol.* 2014;11(3):253–262. [PubMed: 24633013]
164. Travis OK, White D, Pierce WA, et al. Chronic infusion of interleukin-17 promotes hypertension, activation of cytolytic natural killer cells, and vascular dysfunction in pregnant rats. *Physiol Rep.* 2019;7(7):e14038. [PubMed: 30963715]
165. Choi GB, Yim YS, Wong H, et al. The maternal interleukin-17a pathway in mice promotes autism-like phenotypes in offspring. *Science (New York, NY).* 2016;351(6276):933–939.
166. Antonson AM, Kenney AD, Chen HJ, Corps KN, Yount JS, Gur TL. Moderately pathogenic maternal influenza A virus infection disrupts placental integrity but spares the fetal brain. *Brain Behav Immun.* 2021;96:28–39. [PubMed: 33989741]
167. Li C, Yang P, Sun Y, et al. IL-17 response mediates acute lung injury induced by the 2009 pandemic influenza A (H1N1) virus. *Cell Res.* 2012;22(3):528–538. [PubMed: 22025253]
168. Bermejo-Martin JF, Ortiz de Lejarazu R, Pumarola T, et al. Th1 and Th17 hypercytokinemia as early host response signature in severe pandemic influenza. *Crit Care.* 2009;13(6):R201. [PubMed: 20003352]
169. Crowe CR, Chen K, Pociask DA, et al. Critical role of IL-17RA in immunopathology of influenza infection. *J Immunol.* 2009;183(8):5301–5310. [PubMed: 19783685]

170. Wang X, Chan CC, Yang M, et al. A critical role of IL-17 in modulating the B-cell response during H5N1 influenza virus infection. *Cell Mol Immunol*. 2011;8(6):462–468. [PubMed: 21946434]
171. Kudva A, Scheller EV, Robinson KM, et al. Influenza A inhibits Th17-mediated host defense against bacterial pneumonia in mice. *J Immunol*. 2011;186(3):1666–1674. [PubMed: 21178015]
172. McKinstry KK, Strutt TM, Buck A, et al. IL-10 deficiency unleashes an influenza-specific Th17 response and enhances survival against high-dose challenge. *J Immunol*. 2009;182(12):7353–7363. [PubMed: 19494257]
173. Bao J, Cui D, Wang X, et al. Decreased Frequencies of Th17 and Tc17 Cells in Patients Infected with Avian Influenza A (H7N9) Virus. *J Immunol Res*. 2019;2019:1418251. [PubMed: 31061831]
174. Assefa D, Amin N, Dozor AJ, Parton LA. Attenuated interleukin-8/leukocyte immunoresponse in preterm infants compared with term infants hospitalized with respiratory syncytial virus bronchiolitis: a pilot study. *Hum Immunol*. 2011;72(9):708–711. [PubMed: 21683109]
175. de Almeida Nagata DE, Demoor T, Ptaschinski C, et al. IL-27R-mediated regulation of IL-17 controls the development of respiratory syncytial virus-associated pathogenesis. *Am J Pathol*. 2014;184(6):1807–1818. [PubMed: 24726498]
176. Qin L, Qiu K, Hu C, Wang L, Wu G, Tan Y. Respiratory syncytial virus promoted the differentiation of Th17 cells in airway microenvironment through activation of Notch-1/Delta3. *J Med Microbiol*. 2019;68(4):649–656. [PubMed: 30843783]
177. Gao M, Liu LX, Wu FL, et al. The Changes of Th17/Treg and Related Cytokines: IL-17, IL-23, IL-10, and TGF-beta in Respiratory Syncytial Virus Bronchiolitis Rat Model. *Iran J Allergy Asthma Immunol*. 2017;16(5):386–395. [PubMed: 29149778]
178. Reed M, Morris SH, Owczarczyk AB, Lukacs NW. Deficiency of autophagy protein Map1-LC3b mediates IL-17-dependent lung pathology during respiratory viral infection via ER stress-associated IL-1. *Mucosal Immunol*. 2015;8(5):1118–1130. [PubMed: 25669150]
179. Mukherjee S, Lindell DM, Berlin AA, et al. IL-17-induced pulmonary pathogenesis during respiratory viral infection and exacerbation of allergic disease. *Am J Pathol*. 2011;179(1):248–258. [PubMed: 21703407]
180. Muyayalo KP, Huang DH, Zhao SJ, Xie T, Mor G, Liao AH. COVID-19 and Treg/Th17 imbalance: Potential relationship to pregnancy outcomes. *American journal of reproductive immunology (New York, NY : 1989)*. 2020;84(5):e13304.
181. Xu Z, Shi L, Wang Y, et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *Lancet Respir Med*. 2020;8(4):420–422. [PubMed: 32085846]
182. Tiwari-Heckler S, Rauber C, Longhi MS, et al. Dysregulated Host Response in Severe Acute Respiratory Syndrome Coronavirus 2-Induced Critical Illness. *Open Forum Infect Dis*. 2021;8(3):ofab019. [PubMed: 33778090]
183. Mahallawi WH, Khabour OF, Zhang Q, Makhdoum HM, Suliman BA. MERS-CoV infection in humans is associated with a pro-inflammatory Th1 and Th17 cytokine profile. *Cytokine*. 2018;104:8–13. [PubMed: 29414327]
184. Born WK, Reardon CL, O'Brien RL. The function of gammadelta T cells in innate immunity. *Curr Opin Immunol*. 2006;18(1):31–38. [PubMed: 16337364]
185. Papotto PH, Ribot JC, Silva-Santos B. IL-17(+) gammadelta T cells as kick-starters of inflammation. *Nat Immunol*. 2017;18(6):604–611. [PubMed: 28518154]
186. Sumaria N, Grandjean CL, Silva-Santos B, Pennington DJ. Strong TCRgammadelta Signaling Prohibits Thymic Development of IL-17A-Secreting gammadelta T Cells. *Cell Rep*. 2017;19(12):2469–2476. [PubMed: 28636936]
187. Sutton CE, Lalor SJ, Sweeney CM, Brereton CF, Lavelle EC, Mills KH. Interleukin-1 and IL-23 induce innate IL-17 production from gammadelta T cells, amplifying Th17 responses and autoimmunity. *Immunity*. 2009;31(2):331–341. [PubMed: 19682929]
188. Lockhart E, Green AM, Flynn JL. IL-17 production is dominated by gammadelta T cells rather than CD4 T cells during Mycobacterium tuberculosis infection. *J Immunol*. 2006;177(7):4662–4669. [PubMed: 16982905]

189. Umemura M, Yahagi A, Hamada S, et al. IL-17-mediated regulation of innate and acquired immune response against pulmonary *Mycobacterium bovis* bacille Calmette-Guerin infection. *J Immunol.* 2007;178(6):3786–3796. [PubMed: 17339477]
190. Shibata K, Yamada H, Hara H, Kishihara K, Yoshikai Y. Resident Vdelta1+ gammadelta T cells control early infiltration of neutrophils after *Escherichia coli* infection via IL-17 production. *J Immunol.* 2007;178(7):4466–4472. [PubMed: 17372004]
191. Deusch K, Luling F, Reich K, Classen M, Wagner H, Pfeffer K. A major fraction of human intraepithelial lymphocytes simultaneously expresses the gamma/delta T cell receptor, the CD8 accessory molecule and preferentially uses the V delta 1 gene segment. *Eur J Immunol.* 1991;21(4):1053–1059. [PubMed: 1826884]
192. Parker CM, Groh V, Band H, et al. Evidence for extrathymic changes in the T cell receptor gamma/delta repertoire. *J Exp Med.* 1990;171(5):1597–1612. [PubMed: 2185330]
193. Liang Q, Tong L, Xiang L, et al. Correlations of the expression of gammadelta T cells and their co-stimulatory molecules TIGIT, PD-1, ICOS and BTLA with PR and PIBF in the peripheral blood and decidual tissues of women with unexplained recurrent spontaneous abortion. *Clin Exp Immunol.* 2021;203(1):55–65. [PubMed: 33017473]
194. Polgar B, Barakonyi A, Xynos I, Szekeres-Bartho J. The role of gamma/delta T cell receptor positive cells in pregnancy. *American journal of reproductive immunology (New York, NY : 1989).* 1999;41(4):239–244.
195. Majewski AC, Hansen PJ. Progesterone inhibits rejection of xenogeneic transplants in the sheep uterus. *Horm Res.* 2002;58(3):128–135. [PubMed: 12218378]
196. Cai D, Tang Y, Yao X. Changes of gammadelta T cell subtypes during pregnancy and their influences in spontaneous abortion. *J Reprod Immunol.* 2019;131:57–62. [PubMed: 30710888]
197. Andersson A, Grahnemo L, Engdahl C, et al. IL-17-producing gammadelta T cells are regulated by estrogen during development of experimental arthritis. *Clin Immunol.* 2015;161(2):324–332. [PubMed: 26423309]
198. Mikolajewicz K, Chodaczek G. Going deeper: three-dimensional study of gammadelta T cells in mouse reproductive tract using tissue clearing methods. *Immunol Cell Biol.* 2019;97(1):104–111. [PubMed: 30218458]
199. Anipindi VC, Bagri P, Dizzell SE, et al. IL-17 Production by gammadelta(+) T Cells Is Critical for Inducing Th17 Responses in the Female Genital Tract and Regulated by Estradiol and Microbiota. *Immunohorizons.* 2019;3(7):317–330. [PubMed: 31356161]
200. Arck PC, Ferrick DA, Steele-Norwood D, et al. Murine T cell determination of pregnancy outcome. *Cell Immunol.* 1999;196(2):71–79. [PubMed: 10527558]
201. Clark DA, Croitoru K. TH1/TH2,3 imbalance due to cytokine-producing NK, gammadelta T and NK-gammadelta T cells in murine pregnancy decidua in success or failure of pregnancy. *Am J Reprod Immunol.* 2001;45(5):257–265. [PubMed: 11432400]
202. Terzieva A, Dimitrova V, Djerov L, et al. Early Pregnancy Human Decidua is Enriched with Activated, Fully Differentiated and Pro-Inflammatory Gamma/Delta T Cells with Diverse TCR Repertoires. *International journal of molecular sciences.* 2019;20(3).
203. Barakonyi A, Kovacs KT, Miko E, Szereday L, Varga P, Szekeres-Bartho J. Recognition of nonclassical HLA class I antigens by gamma delta T cells during pregnancy. *J Immunol.* 2002;168(6):2683–2688. [PubMed: 11884433]
204. Meeusen E, Fox A, Brandon M, Lee CS. Activation of uterine intraepithelial gamma delta T cell receptor-positive lymphocytes during pregnancy. *Eur J Immunol.* 1993;23(5):1112–1117. [PubMed: 8477805]
205. Fan DX, Duan J, Li MQ, Xu B, Li DJ, Jin LP. The decidual gamma-delta T cells up-regulate the biological functions of trophoblasts via IL-10 secretion in early human pregnancy. *Clin Immunol.* 2011;141(3):284–292. [PubMed: 21873118]
206. Mincheva-Nilsson L, Hammarstrom S, Hammarstrom ML. Human decidual leukocytes from early pregnancy contain high numbers of gamma delta+ cells and show selective down-regulation of alloreactivity. *J Immunol.* 1992;149(6):2203–2211. [PubMed: 1381400]
207. Song ZH, Li ZY, Li DD, et al. Seminal plasma induces inflammation in the uterus through the gammadelta T/IL-17 pathway. *Sci Rep.* 2016;6:25118. [PubMed: 27109934]

208. Pinget GV, Corpuz TM, Stolp J, et al. The majority of murine gammadelta T cells at the maternal-fetal interface in pregnancy produce IL-17. *Immunol Cell Biol.* 2016;94(7):623–630. [PubMed: 27241697]
209. Polese B, Grیدهlet V, Perrier d’Hauterive S, et al. Accumulation of IL-17(+) Vgamma6(+) gammadelta T cells in pregnant mice is not associated with spontaneous abortion. *Clin Transl Immunology.* 2018;7(1):e1008. [PubMed: 29484185]
210. Miko E, Szereday L, Barakonyi A, Jarkovich A, Varga P, Szekeres-Bartho J. Immunoactivation in preeclampsia: Vdelta2+ and regulatory T cells during the inflammatory stage of disease. *J Reprod Immunol.* 2009;80(1–2):100–108. [PubMed: 19395088]
211. Lewis EL, Tulina N, Anton L, Brown AG, Porrett PM, Elovitz MA. IFNgamma-Producing gamma/delta T Cells Accumulate in the Fetal Brain Following Intrauterine Inflammation. *Front Immunol.* 2021;12:741518. [PubMed: 34675929]
212. Talukdar A, Rai R, Aparna Sharma K, Rao DN, Sharma A. Peripheral Gamma Delta T cells secrete inflammatory cytokines in women with idiopathic recurrent pregnancy loss. *Cytokine.* 2018;102:117–122. [PubMed: 28802663]
213. Huang C, Zhang Y, Xiang Z, et al. Granzyme B-expressing gammadelta-T and NK cells as a predictor of clinical pregnancy failure in patients with unexplained repeated implantation failure. *J Reprod Immunol.* 2021;144:103269. [PubMed: 33540297]
214. Mincheva-Nilsson L, Nagaeva O, Sundqvist KG, Hammarstrom ML, Hammarstrom S, Baranov V. gammadelta T cells of human early pregnancy decidua: evidence for cytotoxic potency. *International immunology.* 2000;12(5):585–596. [PubMed: 10784604]
215. Zhao Y, Dai ZP, Lv P, Gao XM. Phenotypic and functional analysis of human T lymphocytes in early second- and third-trimester fetuses. *Clin Exp Immunol.* 2002;129(2):302–308. [PubMed: 12165087]
216. Barakonyi A, Polgar B, Szekeres-Bartho J. The role of gamma/delta T-cell receptor-positive cells in pregnancy: part II. *American journal of reproductive immunology (New York, NY : 1989).* 1999;42(2):83–87.
217. Yu L, Zhang Y, Xiong J, et al. Activated gammadelta T Cells With Higher CD107a Expression and Inflammatory Potential During Early Pregnancy in Patients With Recurrent Spontaneous Abortion. *Front Immunol.* 2021;12:724662. [PubMed: 34484234]
218. Stas MR, Koch M, Stadler M, et al. NK and T Cell Differentiation at the Maternal-Fetal Interface in Sows During Late Gestation. *Front Immunol.* 2020;11:582065. [PubMed: 33013937]
219. Palomino-Segura M, Latino I, Farsakoglu Y, Gonzalez SF. Early production of IL-17A by gammadelta T cells in the trachea promotes viral clearance during influenza infection in mice. *Eur J Immunol.* 2020;50(1):97–109. [PubMed: 31777067]
220. Misiak A, Wilk MM, Raverdeau M, Mills KH. IL-17-Producing Innate and Pathogen-Specific Tissue Resident Memory gammadelta T Cells Expand in the Lungs of Bordetella pertussis-Infected Mice. *J Immunol.* 2017;198(1):363–374. [PubMed: 27864475]
221. Guo XJ, Dash P, Crawford JC, et al. Lung gammadelta T Cells Mediate Protective Responses during Neonatal Influenza Infection that Are Associated with Type 2 Immunity. *Immunity.* 2018;49(3):531–544 e536. [PubMed: 30170813]
222. Qin G, Liu Y, Zheng J, et al. Phenotypic and functional characterization of human gammadelta T-cell subsets in response to influenza A viruses. *J Infect Dis.* 2012;205(11):1646–1653. [PubMed: 22457284]
223. Li H, Xiang Z, Feng T, et al. Human Vgamma9Vdelta2-T cells efficiently kill influenza virus-infected lung alveolar epithelial cells. *Cell Mol Immunol.* 2013;10(2):159–164. [PubMed: 23353835]
224. Qin G, Mao H, Zheng J, et al. Phosphoantigen-expanded human gammadelta T cells display potent cytotoxicity against monocyte-derived macrophages infected with human and avian influenza viruses. *J Infect Dis.* 2009;200(6):858–865. [PubMed: 19656068]
225. Schwaiger T, Sehl J, Karte C, et al. Experimental H1N1pdm09 infection in pigs mimics human seasonal influenza infections. *PLoS One.* 2019;14(9):e0222943. [PubMed: 31539406]

226. Tu W, Zheng J, Liu Y, et al. The aminobisphosphonate pamidronate controls influenza pathogenesis by expanding a gammadelta T cell population in humanized mice. *J Exp Med*. 2011;208(7):1511–1522. [PubMed: 21708931]
227. Qin G, Liu Y, Zheng J, et al. Type 1 responses of human Vgamma9Vdelta2 T cells to influenza A viruses. *J Virol*. 2011;85(19):10109–10116. [PubMed: 21752902]
228. Dong P, Ju X, Yan Y, et al. gammadelta T Cells Provide Protective Function in Highly Pathogenic Avian H5N1 Influenza A Virus Infection. *Front Immunol*. 2018;9:2812. [PubMed: 30564234]
229. Pei Y, Wen K, Xiang Z, et al. CD137 costimulation enhances the antiviral activity of Vgamma9Vdelta2-T cells against influenza virus. *Signal Transduct Target Ther*. 2020;5(1):74. [PubMed: 32488072]
230. Lu Y, Li Z, Ma C, et al. The interaction of influenza H5N1 viral hemagglutinin with sialic acid receptors leads to the activation of human gammadelta T cells. *Cell Mol Immunol*. 2013;10(6):463–470. [PubMed: 23912782]
231. Dodd J, Riffault S, Kodituwakku JS, Hayday AC, Openshaw PJ. Pulmonary V gamma 4+ gamma delta T cells have proinflammatory and antiviral effects in viral lung disease. *J Immunol*. 2009;182(2):1174–1181. [PubMed: 19124761]
232. Huang H, Saravia J, You D, Shaw AJ, Cormier SA. Impaired gamma delta T cell-derived IL-17A and inflammasome activation during early respiratory syncytial virus infection in infants. *Immunol Cell Biol*. 2015;93(2):126–135. [PubMed: 25267484]
233. Gee S, Chandiramani M, Seow J, et al. The legacy of maternal SARS-CoV-2 infection on the immunology of the neonate. *Nat Immunol*. 2021;22(12):1490–1502. [PubMed: 34616036]
234. Poccia F, Agrati C, Castilletti C, et al. Anti-severe acute respiratory syndrome coronavirus immune responses: the role played by V gamma 9V delta 2 T cells. *J Infect Dis*. 2006;193(9):1244–1249. [PubMed: 16586361]
235. Cerapio JP, Perrier M, Pont F, et al. Single-Cell RNAseq Profiling of Human gammadelta T Lymphocytes in Virus-Related Cancers and COVID-19 Disease. *Viruses*. 2021;13(11).
236. Lei L, Qian H, Yang X, et al. The phenotypic changes of gammadelta T cells in COVID-19 patients. *J Cell Mol Med*. 2020;24(19):11603–11606. [PubMed: 32864865]
237. Chaouat G, Ledee-Bataille N, Dubanchet S, Zourbas S, Sandra O, Martal J. TH1/TH2 paradigm in pregnancy: paradigm lost? Cytokines in pregnancy/early abortion: reexamining the TH1/TH2 paradigm. *Int Arch Allergy Immunol*. 2004;134(2):93–119. [PubMed: 15153791]
238. Jenkins C, Roberts J, Wilson R, MacLean MA, Shilito J, Walker JJ. Evidence of a T(H) 1 type response associated with recurrent miscarriage. *Fertility and sterility*. 2000;73(6):1206–1208. [PubMed: 10856484]
239. Piccinni MP, Beloni L, Livi C, Maggi E, Scarselli G, Romagnani S. Defective production of both leukemia inhibitory factor and type 2 T-helper cytokines by decidual T cells in unexplained recurrent abortions. *Nat Med*. 1998;4(9):1020–1024. [PubMed: 9734394]
240. Wang W, Sung N, Gilman-Sachs A, Kwak-Kim J. T Helper (Th) Cell Profiles in Pregnancy and Recurrent Pregnancy Losses: Th1/Th2/Th9/Th17/Th22/Tfh Cells. *Front Immunol*. 2020;11:2025. [PubMed: 32973809]
241. Zhang L, Zhao Z, Mi H, Liu B, Wang B, Yang L. Modulation of Helper T Cytokines in Thymus during Early Pregnancy in Ewes. *Animals (Basel)*. 2019;9(5).
242. Tsuda H, Michimata T, Hayakawa S, et al. A Th2 chemokine, TARC, produced by trophoblasts and endometrial gland cells, regulates the infiltration of CCR4(+) T lymphocytes into human decidua at early pregnancy. *American Journal of Reproductive Immunology*. 2002;48(1):1–8. [PubMed: 12322891]
243. Morelli SS, Mandal M, Goldsmith LT, Kashani BN, Ponzio NM. The maternal immune system during pregnancy and its influence on fetal development. *Research and Reports in Biology*. 2015;6:171–119 p.
244. Wegmann TG, Lin H, Guilbert L, Mosmann TR. Bidirectional cytokine interactions in the maternal-fetal relationship: is successful pregnancy a TH2 phenomenon? *Immunol Today*. 1993;14(7):353–356. [PubMed: 8363725]

245. Khan D, Ansar Ahmed S. The Immune System Is a Natural Target for Estrogen Action: Opposing Effects of Estrogen in Two Prototypical Autoimmune Diseases. *Front Immunol.* 2015;6:635. [PubMed: 26779182]
246. Robinson DP, Klein SL. Pregnancy and pregnancy-associated hormones alter immune responses and disease pathogenesis. *Horm Behav.* 2012;62(3):263–271. [PubMed: 22406114]
247. Straub RH. The complex role of estrogens in inflammation. *Endocr Rev.* 2007;28(5):521–574. [PubMed: 17640948]
248. Kalinski P Regulation of immune responses by prostaglandin E2. *J Immunol.* 2012;188(1):21–28. [PubMed: 22187483]
249. Kirkwood JM, Butterfield LH, Tarhini AA, Zarour H, Kalinski P, Ferrone S. Immunotherapy of cancer in 2012. *CA Cancer J Clin.* 2012;62(5):309–335. [PubMed: 22576456]
250. Hellberg S, Raffetseder J, Rundquist O, et al. Progesterone Dampens Immune Responses in In Vitro Activated CD4(+) T Cells and Affects Genes Associated With Autoimmune Diseases That Improve During Pregnancy. *Front Immunol.* 2021;12:672168. [PubMed: 34054852]
251. Miyaura H, Iwata M. Direct and indirect inhibition of Th1 development by progesterone and glucocorticoids. *Journal of Immunology.* 2002;168(3):1087–1094.
252. Szekeres-Bartho J, Wegmann TG. A progesterone-dependent immunomodulatory protein alters the Th1/Th2 balance. *J Reprod Immunol.* 1996;31(1–2):81–95. [PubMed: 8887124]
253. Szekeres-Bartho J, Polgar B. PIBF: the double edged sword. Pregnancy and tumor. *Am J Reprod Immunol.* 2010;64(2):77–86. [PubMed: 20367622]
254. Shah NM, Lai PF, Imami N, Johnson MR. Progesterone-Related Immune Modulation of Pregnancy and Labor. *Front Endocrinol (Lausanne).* 2019;10:198. [PubMed: 30984115]
255. Gadotti AC, de Castro Deus M, Telles JP, et al. IFN-gamma is an independent risk factor associated with mortality in patients with moderate and severe COVID-19 infection. *Virus Res.* 2020;289:198171. [PubMed: 32979474]
256. Karlsson EA, Marcelin G, Webby RJ, Schultz-Cherry S. Review on the impact of pregnancy and obesity on influenza virus infection. *Influenza Other Respir Viruses.* 2012;6(6):449–460. [PubMed: 22335790]
257. Gu X, Li P, Liu H, Li N, Li S, Sakuma T. The effect of influenza virus A on th1/th2 balance and alveolar fluid clearance in pregnant rats. *Exp Lung Res.* 2011;37(7):445–451. [PubMed: 21777148]
258. Chan KH, Zhang AJ, To KK, et al. Wild type and mutant 2009 pandemic influenza A (H1N1) viruses cause more severe disease and higher mortality in pregnant BALB/c mice. *PLoS one.* 2010;5(10):e13757. [PubMed: 21060798]
259. Yamaguchi K, Hisano M, Isojima S, et al. Relationship of Th1/Th2 cell balance with the immune response to influenza vaccine during pregnancy. *J Med Virol.* 2009;81(11):1923–1928. [PubMed: 19774681]
260. Fettke F, Schumacher A, Canellada A, et al. Maternal and Fetal Mechanisms of B Cell Regulation during Pregnancy: Human Chorionic Gonadotropin Stimulates B Cells to Produce IL-10 While Alpha-Fetoprotein Drives Them into Apoptosis. *Front Immunol.* 2016;7:495. [PubMed: 28008329]
261. Montecino-Rodriguez E, Dorshkind K. B-1 B cell development in the fetus and adult. *Immunity.* 2012;36(1):13–21. [PubMed: 22284417]
262. Rodriguez-Zhurbenko N, Quach TD, Hopkins TJ, Rothstein TL, Hernandez AM. Human B-1 Cells and B-1 Cell Antibodies Change With Advancing Age. *Front Immunol.* 2019;10:483. [PubMed: 30941130]
263. Lima J, Martins C, Leandro MJ, et al. Characterization of B cells in healthy pregnant women from late pregnancy to post-partum: a prospective observational study. *BMC pregnancy and childbirth.* 2016;16(1):139. [PubMed: 27267973]
264. Muzzio D, Zenclussen AC, Jensen F. The Role of B Cells in Pregnancy: the Good and the Bad. *American Journal of Reproductive Immunology.* 2013;69(4):408–412. [PubMed: 23351028]
265. Reyneveld GI, Savelkoul HFJ, Parmentier HK. Current Understanding of Natural Antibodies and Exploring the Possibilities of Modulation Using Veterinary Models. A Review. *Front Immunol.* 2020;11:2139. [PubMed: 33013904]

266. Ellis TM, Moser MT, Le PT, Flanigan RC, Kwon ED. Alterations in peripheral B cells and B cell progenitors following androgen ablation in mice. *International Immunology*. 2001;13(4):553–558. [PubMed: 11282994]
267. Gubbels Bupp MR, Jorgensen TN. Androgen-Induced Immunosuppression. *Front Immunol*. 2018;9:794. [PubMed: 29755457]
268. Hill L, Jeganathan V, Chinnasamy P, Grimaldi C, Diamond B. Differential Roles of Estrogen Receptors α and β in Control of B-Cell Maturation and Selection. *Molecular Medicine*. 2011;17(3–4):211–220. [PubMed: 21107497]
269. Grimaldi CM, Jeganathan V, Diamond B. Hormonal regulation of B cell development: 17 beta-estradiol impairs negative selection of high-affinity DNA-reactive B cells at more than one developmental checkpoint. *J Immunol*. 2006;176(5):2703–2710. [PubMed: 16493025]
270. Ysraelit MC, Correale J. Impact of sex hormones on immune function and multiple sclerosis development. *Immunology*. 2019;156(1):9–22. [PubMed: 30222193]
271. Swieboda D, Littauer EQ, Beaver JT, et al. Pregnancy Downregulates Plasmablast Metabolic Gene Expression Following Influenza Without Altering Long-Term Antibody Function. *Front Immunol*. 2020;11:1785. [PubMed: 32922392]
272. Fink NR, Chawes B, Bønnelykke K, et al. Levels of Systemic Low-grade Inflammation in Pregnant Mothers and Their Offspring are Correlated. *Scientific Reports*. 2019;9(1).
273. Bouaziz JD, Calbo S, Maho-Vaillant M, et al. IL-10 produced by activated human B cells regulates CD4(+) T-cell activation in vitro. *Eur J Immunol*. 2010;40(10):2686–2691. [PubMed: 20809522]
274. Guzman-Genuino RM, Diener KR. Regulatory B Cells in Pregnancy: Lessons from Autoimmunity, Graft Tolerance, and Cancer. *Front Immunol*. 2017;8:172. [PubMed: 28261223]
275. Medina KL, Smithson G, Kincade PW. Suppression of B lymphopoiesis during normal pregnancy. *J Exp Med*. 1993;178(5):1507–1515. [PubMed: 8228804]
276. Bosco N, Ceredig R, Rolink A. Transient decrease in interleukin-7 availability arrests B lymphopoiesis during pregnancy. *Eur J Immunol*. 2008;38(2):381–390. [PubMed: 18203141]
277. Xu Y, He H, Li C, et al. Immunosuppressive effect of progesterone on dendritic cells in mice. *J Reprod Immunol*. 2011;91(1–2):17–23. [PubMed: 21856019]
278. Arruvito L, Giulianelli S, Flores AC, et al. NK cells expressing a progesterone receptor are susceptible to progesterone-induced apoptosis. *J Immunol*. 2008;180(8):5746–5753. [PubMed: 18390760]
279. Palma J, Tokarz-Deptuła B, Deptuła J, Deptuła W. Natural antibodies – facts known and unknown. *Central European Journal of Immunology*. 2018;43(4):466–475. [PubMed: 30799995]
280. Laubreton D, Drajac C, Eléouët J-F, et al. Regulatory B Lymphocytes Colonize the Respiratory Tract of Neonatal Mice and Modulate Immune Responses of Alveolar Macrophages to RSV Infection in IL-10-Dependant Manner. *Viruses*. 2020;12(8):822.
281. Zhivaki D, Lemoine S, Lim A, et al. Respiratory Syncytial Virus Infects Regulatory B Cells in Human Neonates via Chemokine Receptor CX3CR1 and Promotes Lung Disease Severity. *Immunity*. 2017;46(2):301–314. [PubMed: 28228284]
282. Jacobsen H, Walendy-Gnirß K, Tekin-Bubenheim N, et al. Offspring born to influenza A virus infected pregnant mice have increased susceptibility to viral and bacterial infections in early life. *Nature Communications*. 2021;12(1).
283. Kay AW, Bayless NL, Fukuyama J, et al. Pregnancy Does Not Attenuate the Antibody or Plasmablast Response to Inactivated Influenza Vaccine. *Journal of Infectious Diseases*. 2015;212(6):861–870. [PubMed: 25740957]
284. Rastogi D B cell priming in utero to influenza vaccination*1. *Journal of Allergy and Clinical Immunology*. 2004;113(2):S50.
285. Laidlaw BJ, Ellebedy AH. The germinal centre B cell response to SARS-CoV-2. *Nature Reviews Immunology*. 2022;22(1):7–18.
286. Sherer ML, Lei J, Creisher PS, et al. Pregnancy alters interleukin-1 beta expression and antiviral antibody responses during severe acute respiratory syndrome coronavirus 2 infection. *Am J Obstet Gynecol*. 2021;225(3):301 e301–301 e314. [PubMed: 33798476]

287. Chen R, Zhang S, Su S, Ye H, Shu H. Interactions Between Specific Immune Status of Pregnant Women and SARS-CoV-2 Infection. *Front Cell Infect Microbiol.* 2021;11:721309. [PubMed: 34458162]
288. Chen G, Zhang Y, Zhang Y, et al. Differential immune responses in pregnant patients recovered from COVID-19. *Signal Transduction and Targeted Therapy.* 2021;6(1).
289. Wilhelmson AS, Lantero Rodriguez M, Stubelius A, et al. Testosterone is an endogenous regulator of BAFF and splenic B cell number. *Nature Communications.* 2018;9(1).
290. Sakiani S, Olsen NJ, Kovacs WJ. Gonadal steroids and humoral immunity. *Nat Rev Endocrinol.* 2013;9(1):56–62. [PubMed: 23183675]
291. Panda SK, Colonna M. Innate Lymphoid Cells in Mucosal Immunity. *Front Immunol.* 2019;10:861. [PubMed: 31134050]
292. Vivier E, Artis D, Colonna M, et al. Innate Lymphoid Cells: 10 Years On. *Cell.* 2018;174(5):1054–1066. [PubMed: 30142344]
293. Mazzurana L, Rao A, Van Acker A, Mjosberg J. The roles for innate lymphoid cells in the human immune system. *Semin Immunopathol.* 2018;40(4):407–419. [PubMed: 29948108]
294. Strober W The LT_i cell, an immunologic chameleon. *Immunity.* 2010;33(5):650–652. [PubMed: 21094460]
295. Jabrane-Ferrat N Features of Human Decidual NK Cells in Healthy Pregnancy and During Viral Infection. *Front Immunol.* 2019;10:1397. [PubMed: 31379803]
296. Jacquelot N, Luong K, Seillet C. Physiological Regulation of Innate Lymphoid Cells. *Front Immunol.* 2019;10:405. [PubMed: 30915072]
297. Cephus JY, Stier MT, Fuseini H, et al. Testosterone Attenuates Group 2 Innate Lymphoid Cell-Mediated Airway Inflammation. *Cell Rep.* 2017;21(9):2487–2499. [PubMed: 29186686]
298. Blanquart E, Laffont S, Guery JC. Sex hormone regulation of innate lymphoid cells. *Biomedical journal.* 2021;44(2):144–156. [PubMed: 33888441]
299. Einkenkel R, Ehrhardt J, Hartmann K, Kruger D, Muzzio DO, Zygmunt M. Hormonally controlled ILC antigen presentation potential is reduced during pregnancy. *Reproduction.* 2020;160(1):155–169. [PubMed: 32130203]
300. Zhang X, Wei H. Role of Decidual Natural Killer Cells in Human Pregnancy and Related Pregnancy Complications. *Front Immunol.* 2021;12:728291. [PubMed: 34512661]
301. Sentman CL, Meadows SK, Wira CR, Eriksson M. Recruitment of uterine NK cells: induction of CXC chemokine ligands 10 and 11 in human endometrium by estradiol and progesterone. *J Immunol.* 2004;173(11):6760–6766. [PubMed: 15557169]
302. Gibson DA, Greaves E, Critchley HO, Saunders PT. Estrogen-dependent regulation of human uterine natural killer cells promotes vascular remodelling via secretion of CCL2. *Human reproduction (Oxford, England).* 2015;30(6):1290–1301.
303. Miller D, Motomura K, Garcia-Flores V, Romero R, Gomez-Lopez N. Innate Lymphoid Cells in the Maternal and Fetal Compartments. *Front Immunol.* 2018;9:2396. [PubMed: 30416502]
304. Xu Y, Romero R, Miller D, et al. Innate lymphoid cells at the human maternal-fetal interface in spontaneous preterm labor. *Am J Reprod Immunol.* 2018;79(6):e12820. [PubMed: 29457302]
305. Mendes J, Areia AL, Rodrigues-Santos P, Santos-Rosa M, Mota-Pinto A. Innate Lymphoid Cells in Human Pregnancy. *Front Immunol.* 2020;11:551707. [PubMed: 33329512]
306. Yang D, Guo X, Huang T, Liu C. The Role of Group 3 Innate Lymphoid Cells in Lung Infection and Immunity. *Front Cell Infect Microbiol.* 2021;11:586471. [PubMed: 33718260]
307. Monticelli LA, Sonnenberg GF, Abt MC, et al. Innate lymphoid cells promote lung-tissue homeostasis after infection with influenza virus. *Nat Immunol.* 2011;12(11):1045–1054. [PubMed: 21946417]
308. Stier MT, Bloodworth MH, Toki S, et al. Respiratory syncytial virus infection activates IL-13-producing group 2 innate lymphoid cells through thymic stromal lymphopoietin. *J Allergy Clin Immunol.* 2016;138(3):814–824 e811. [PubMed: 27156176]
309. Kumar A, Cao W, Endrias K, Kuchipudi SV, Mittal SK, Sambhara S. Innate lymphoid cells (ILC) in SARS-CoV-2 infection. *Mol Aspects Med.* 2021;80:101008. [PubMed: 34399986]

310. Wu YH, Lai AC, Chi PY, et al. Pulmonary IL-33 orchestrates innate immune cells to mediate respiratory syncytial virus-evoked airway hyperreactivity and eosinophilia. *Allergy*. 2020;75(4):818–830. [PubMed: 31622507]
311. Fonseca W, Lukacs NW, Elesela S, Malinczak CA. Role of ILC2 in Viral-Induced Lung Pathogenesis. *Front Immunol*. 2021;12:675169. [PubMed: 33953732]
312. Liu Y, Gao S, Zhao Y, Wang H, Pan Q, Shao Q. Decidual Natural Killer Cells: A Good Nanny at the Maternal-Fetal Interface During Early Pregnancy. *Front Immunol*. 2021;12:663660. [PubMed: 34054831]
313. Jabrane-Ferrat N, Siewiera J. The up side of decidual natural killer cells: new developments in immunology of pregnancy. *Immunology*. 2014;141(4):490–497. [PubMed: 24256296]
314. Croy BA, Esadeg S, Chantakru S, et al. Update on pathways regulating the activation of uterine Natural Killer cells, their interactions with decidual spiral arteries and homing of their precursors to the uterus. *J Reprod Immunol*. 2003;59(2):175–191. [PubMed: 12896821]
315. Hanna J, Goldman-Wohl D, Hamani Y, et al. Decidual NK cells regulate key developmental processes at the human fetal-maternal interface. *Nat Med*. 2006;12(9):1065–1074. [PubMed: 16892062]
316. Vacca P, Cantoni C, Vitale M, et al. Crosstalk between decidual NK and CD14+ myelomonocytic cells results in induction of Tregs and immunosuppression. *Proc Natl Acad Sci U S A*. 2010;107(26):11918–11923. [PubMed: 20547831]
317. Siewiera J, El Costa H, Tabiasco J, et al. Human cytomegalovirus infection elicits new decidual natural killer cell effector functions. *PLoS Pathog*. 2013;9(4):e1003257. [PubMed: 23592985]
318. Crespo AC, Mulik S, Dotiwala F, et al. Decidual NK Cells Transfer Granulysin to Selectively Kill Bacteria in Trophoblasts. *Cell*. 2020;182(5):1125–1139 e1118. [PubMed: 32822574]
319. Juttukonda LJ, Wachman EM, Boateng J, Jain M, Benarroch Y, Taglauer ES. Decidual immune response following COVID-19 during pregnancy varies by timing of maternal SARS-CoV-2 infection. In: Cold Spring Harbor Laboratory; 2021.
320. Lokken EM, Walker CL, Delaney S, et al. Clinical characteristics of 46 pregnant women with a severe acute respiratory syndrome coronavirus 2 infection in Washington State. *American journal of obstetrics and gynecology*. 2020;223(6):911 e911–911 e914. [PubMed: 32439389]
321. Pierce-Williams RAM, Burd J, Felder L, et al. Clinical course of severe and critical COVID-19 in hospitalized pregnancies: a US cohort study. *Am J Obstet Gynecol MFM*. 2020:100134. [PubMed: 32391519]
322. Li M, Brokaw A, Furuta AM, et al. Non-human Primate Models to Investigate Mechanisms of Infection-Associated Fetal and Pediatric Injury, Teratogenesis and Stillbirth. *Front Genet*. 2021;12(1054).
323. Mayer AE, Parks GD. An AGM model for changes in complement during pregnancy: neutralization of influenza virus by serum is diminished in late third trimester. *PLoS one*. 2014;9(11):e112749. [PubMed: 25409303]
324. Weinfurter JT, Brunner K, Capuano SV 3rd, et al. Cross-reactive T cells are involved in rapid clearance of 2009 pandemic H1N1 influenza virus in nonhuman primates. *PLoS Pathog*. 2011;7(11):e1002381. [PubMed: 22102819]
325. Marriott AC, Dennis M, Kane JA, et al. Influenza A Virus Challenge Models in Cynomolgus Macaques Using the Authentic Inhaled Aerosol and Intra-Nasal Routes of Infection. *PLoS one*. 2016;11(6):e0157887. [PubMed: 27311020]
326. Short SJ, Lubach GR, Karasin AI, et al. Maternal influenza infection during pregnancy impacts postnatal brain development in the rhesus monkey. *Biol Psychiatry*. 2010;67(10):965–973. [PubMed: 20079486]
327. Go JT, Belisle SE, Tchitchek N, et al. 2009 pandemic H1N1 influenza virus elicits similar clinical course but differential host transcriptional response in mouse, macaque, and swine infection models. *BMC Genomics*. 2012;13:627. [PubMed: 23153050]
328. Kobasa D, Jones SM, Shinya K, et al. Aberrant innate immune response in lethal infection of macaques with the 1918 influenza virus. *Nature*. 2007;445(7125):319–323. [PubMed: 17230189]

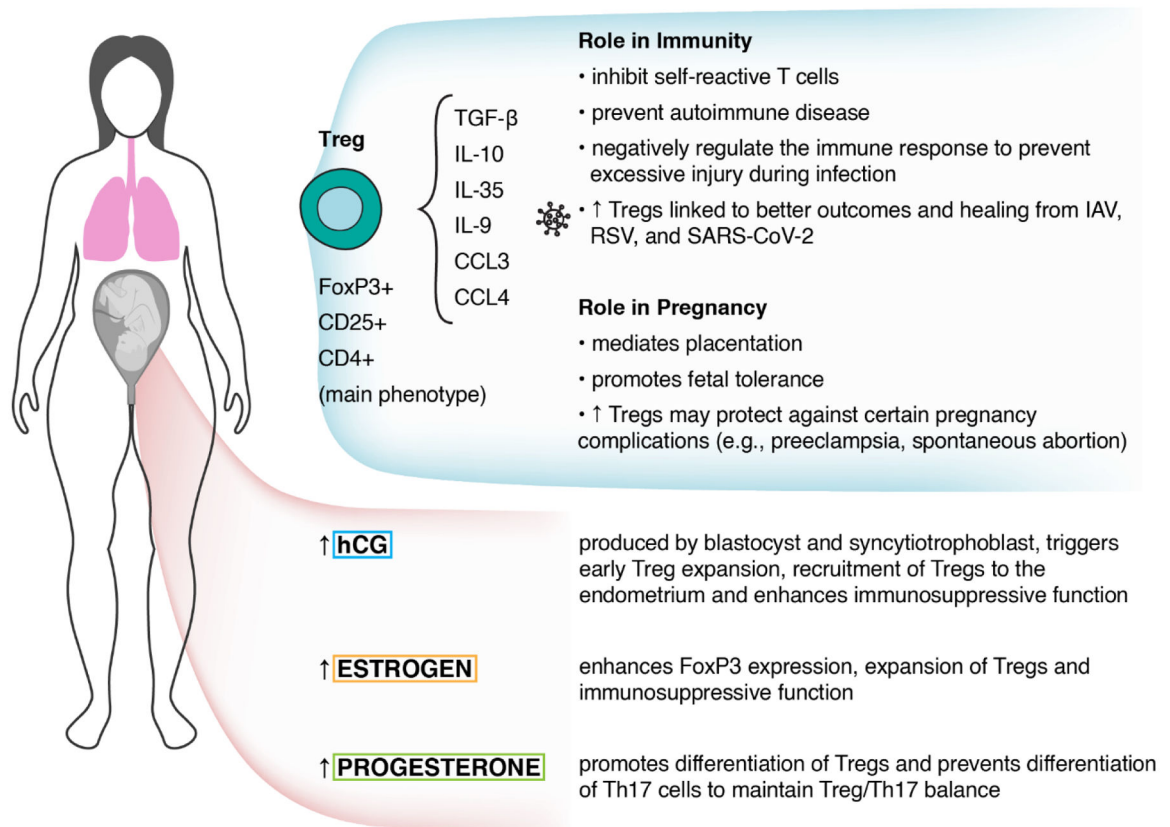


Figure 1. Treg cell role in immunity and pregnancy and the influence of sex hormones.

This figure illustrates the critical role of Tregs in early pregnancy to mediate placentation and promote fetal tolerance. All the major sex hormones in pregnancy support either the proliferation, differentiation, or immunosuppressive function of Tregs. Although there are many Treg phenotypes, the main phenotype is characterized by expression of CD4, CD25 and Forkhead box P3 (FoxP3+). Abbreviations: CD, cluster of differentiation; CD25, also known as the IL-2 receptor alpha chain, CCL3, C-C Motif Chemokine Ligand 3; CCL4, C-C Motif Chemokine Ligand 4; FoxP3, forkhead box P3; IL, interleukin; and TGF- β , transforming growth factor beta.

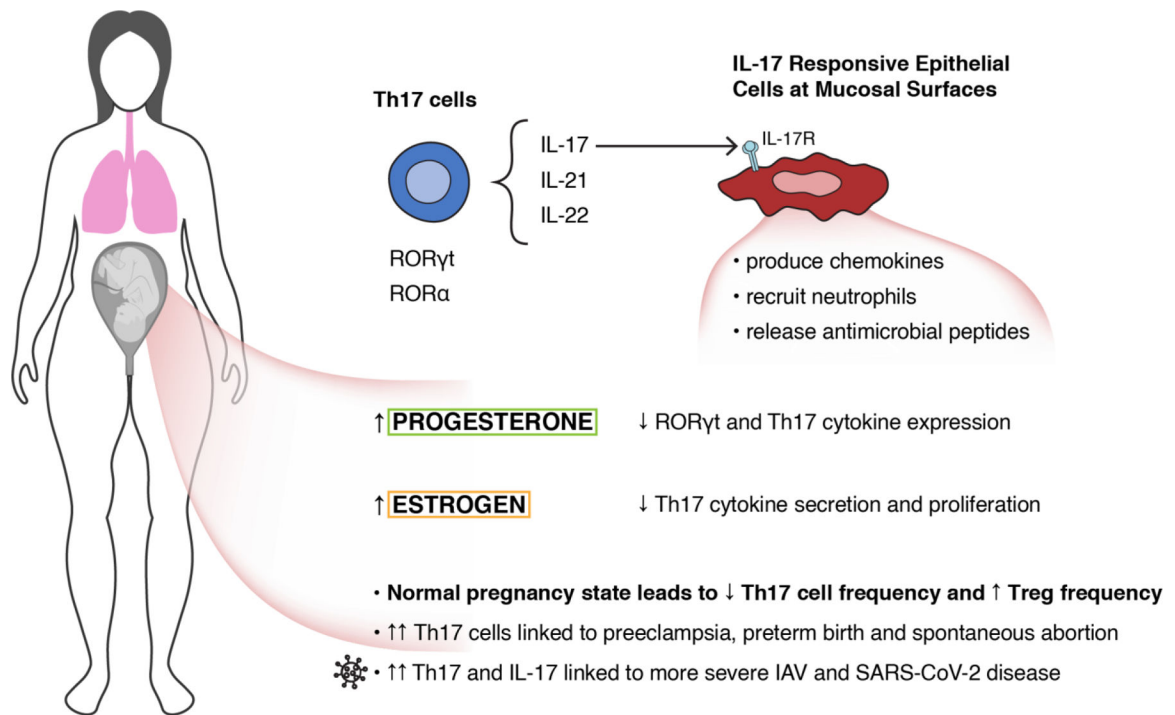


Figure 2. Pro-inflammatory Th17 cells are downregulated during pregnancy.

This figure outlines the pro-inflammatory immune function Th17 cells have in the epithelial and mucosal tissue. Estrogen and progesterone, key sex hormones in pregnancy, dampen the function and decrease the frequency of Th17 cells. Th17 cells are characterized by RAR-related (retinoic acid receptor) orphan receptor (ROR γ t and ROR α) transcription factors and expression of interleukin (IL) cytokines IL-17, IL-21, and IL-22. Abbreviations: IL, interleukin; ROR, RAR-related orphan receptor.

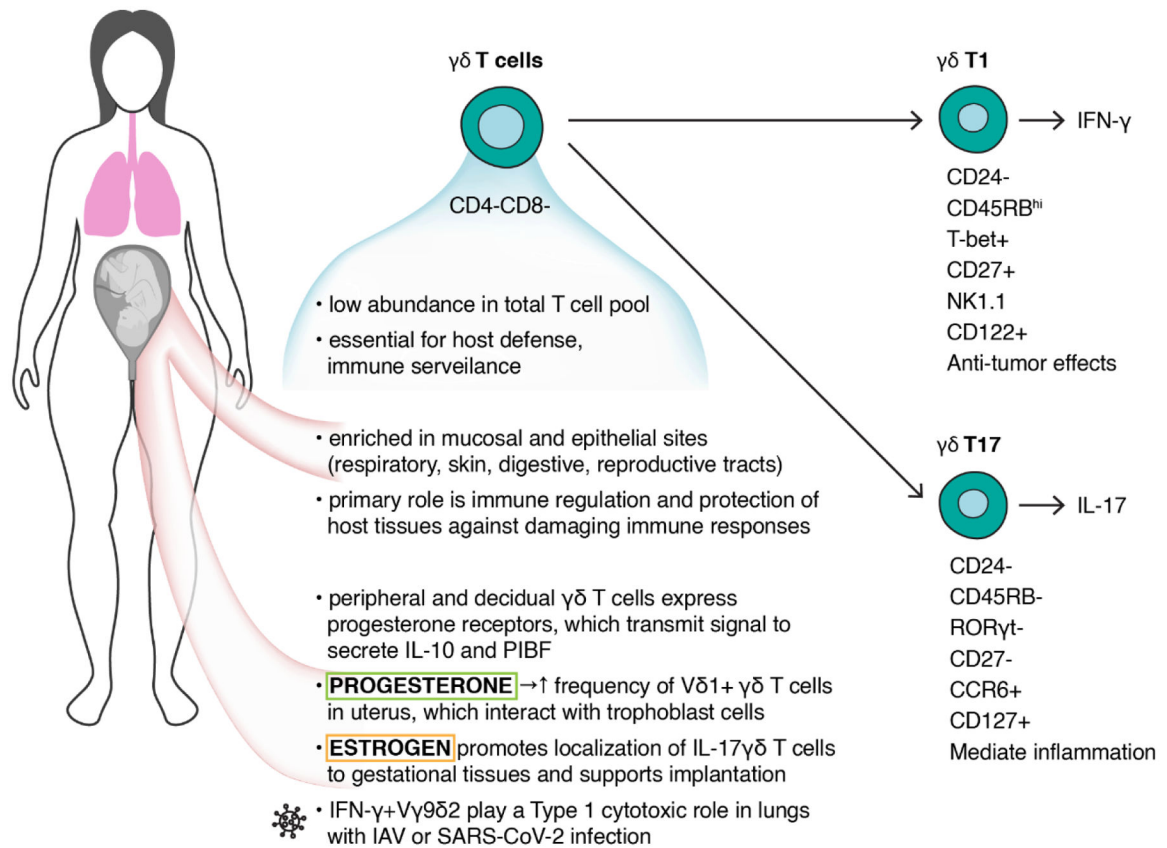


Figure 3. $\gamma\delta$ T cell function and localization are influenced by sex hormones

$\gamma\delta$ T cells are enriched mucosal and epithelial tissue and instrumental in innate immunity and immune regulation. They can be generally divided into IL-17+ and IFN- γ + subsets with distinct functions in immunity and pregnancy. Progesterone stimulates the production of tolerogenic biomolecules such as IL-10 and progesterone induced blocking factor (PIBF). Estrogen and progesterone also promote localization of $V\delta 1$ and IL-17+ $\gamma\delta$ T cells to the gestational tissue. IFN- γ + $\gamma\delta$ T cells, on the other hand, fulfill a Type 1 cytotoxic role during viral respiratory infections that can be detrimental in the gestational environment. Abbreviations: CCR, C-C chemokine receptor; CD, cluster of differentiation; IFN, interferon; IL, interleukin.

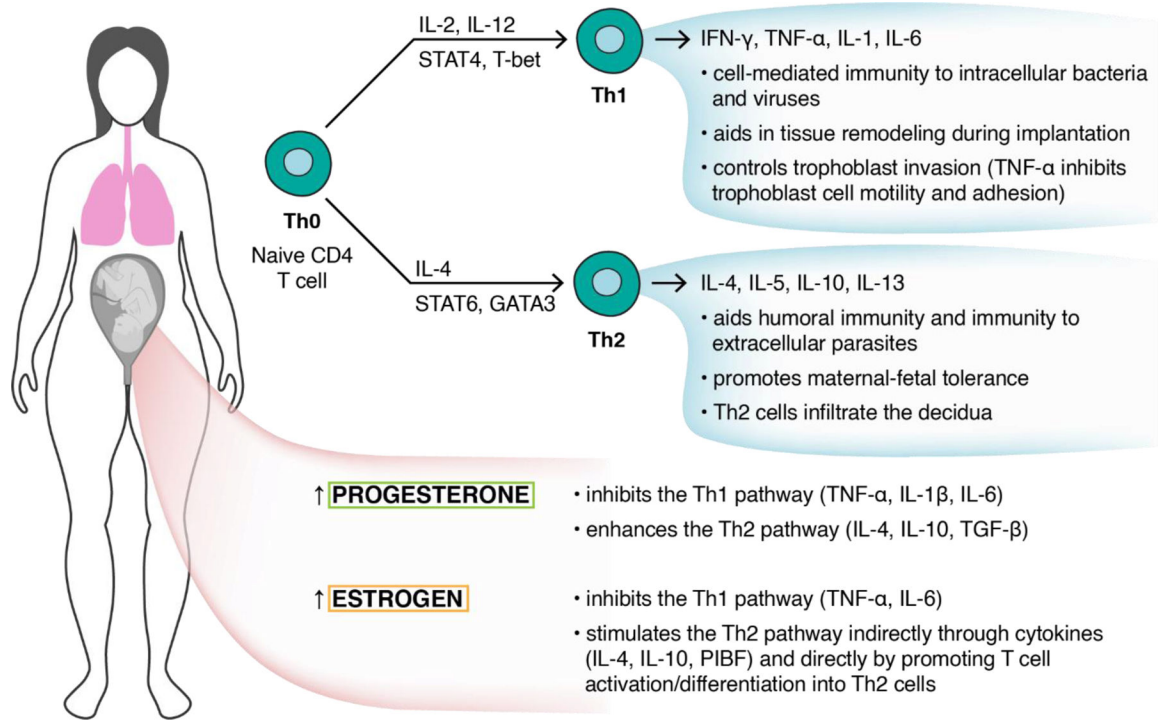


Figure 4. The maternal immune response shifts from a Th1 to a Th2 dominated cytokine profile during pregnancy.

Throughout the course of gestation, Th1 and Th2 cells regulate each other's activity and proliferation via signaling cytokine cascades to establish and maintain a healthy pregnancy. Progesterone and estrogen are key immunomodulators for this process. Abbreviations: CD, cluster of differentiation; GATA, GATA binding protein; IFN, interferon; IL, interleukin; PIBF, progesterone induced blocking factor; STAT, signal transducer and activator of transcription; TGF, transforming growth factor; Th, T helper; TNF, tumor necrosis factor.

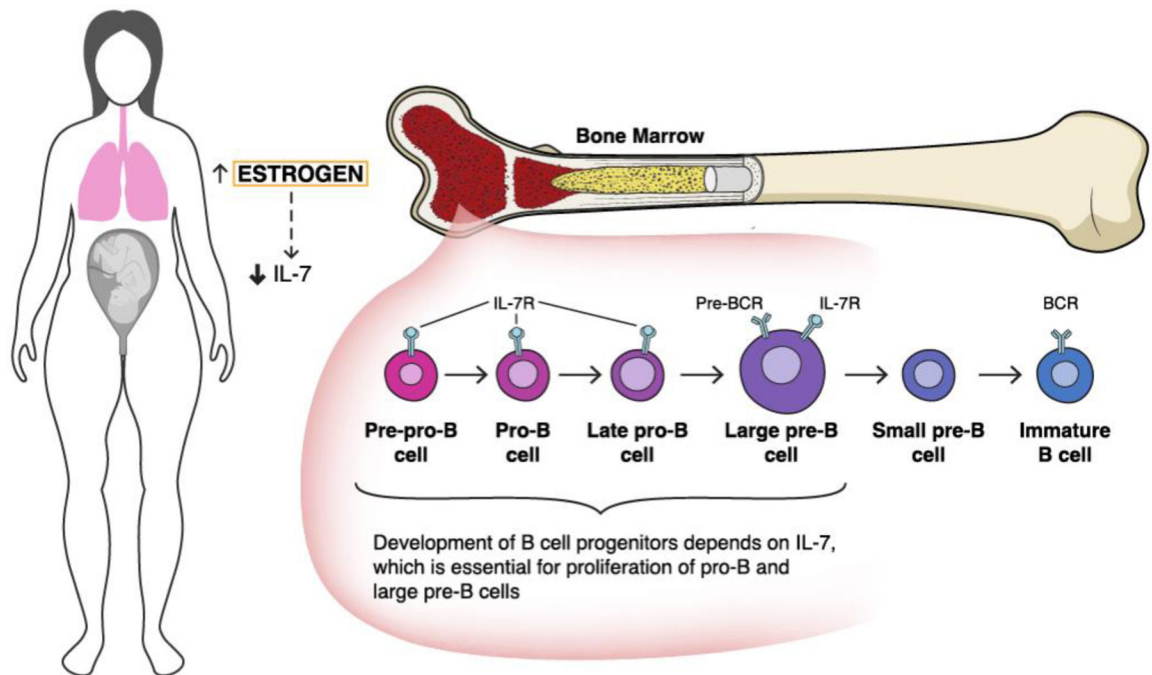


Figure 5. Influences of Sex Hormones on B Cell Production and Maturity.

This figure illustrates the critical role of estrogen in early B cell development to mediate cellular lymphopoiesis. Elevated levels of estrogen in pregnancy modulate B cell proliferation and differentiation through regulating the production and binding of critical cytokines such as IL-7. This serves to attenuate the humoral arm of the adaptive immune response, thereby suppressing overwhelming maternal immunity against an antigenic fetus. Abbreviations: BCR, B Cell Receptor; IL, interleukin.

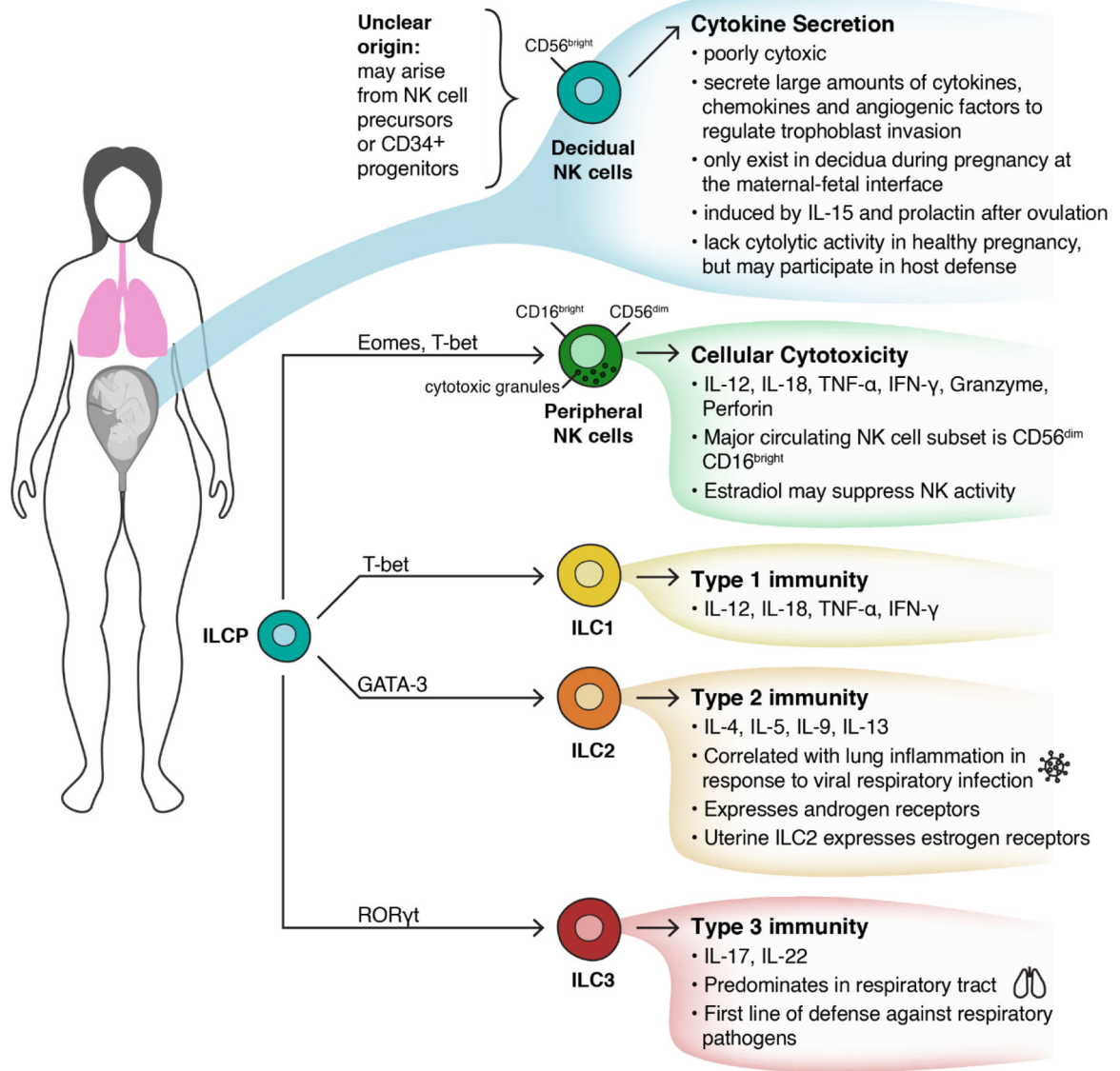


Figure 6. Impact of sex hormones on the function of ILC and dNK cells and their role in pregnancy and respiratory infections.

This figure demonstrates the ILC function of secreting cytokines in response to viral respiratory pathogens and the possible association between ILCs and sex hormones like androgen and estrogen. It also illustrates the role of dNK cells, which secrete cytokines to facilitate trophoblast migration in contrast to pNK cells, which are cytotoxic. dNKs are only found in decidual tissue during pregnancy and are induced by the sex hormone prolactin. Abbreviations: CD, cluster of differentiation; Eomes, eomesodermin; IFN, interferon; IL, interleukin; NK, natural killer; ROR γ t, RAR-related orphan receptor gamma; and TNF- α , tumor necrosis factor alpha.