

Review Article

Molecular Mechanisms of Hawthorn Extracts in Multiple Organs Disorders in Underlying of Diabetes: A Review

Izadpanah Gheitasi ¹, Feryal Savari ², Ghaidafeh Akbari ¹, Jamshid Mohammadi ¹,
Ali Reza Fallahzadeh ³ and Hossein Sadeghi ¹

¹Medicinal Plants Research Center, Yasuj University of Medical Sciences, Yasuj, Iran

²Department of Basic Sciences, Shoushtar Faculty of Medical Sciences, Shoushtar, Iran

³Cellular and Molecular Research Center, Yasuj University of Medical Sciences, Yasuj, Iran

Correspondence should be addressed to Ghaidafeh Akbari; ghaidafehakbari@yahoo.com and Jamshid Mohammadi; j.mohammadi.4554@gmail.com

Received 7 February 2022; Accepted 8 May 2022; Published 7 June 2022

Academic Editor: Nelli Giribabu

Copyright © 2022 Izadpanah Gheitasi et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Diabetes mellitus (DM) is one of the most important metabolic disorders associated with chronic hyperglycemia and occurs when the body cannot manage insulin secretion, insulin action, or both. Autoimmune destruction of pancreatic beta cells and insulin resistance are the major pathophysiological factors of types 1 and 2 of DM, respectively. Prolonged hyperglycemia leads to multiple organs dysfunctions, including nephropathy, neuropathy, cardiomyopathy, gastropathy, and micro- and macrovascular disorders. The basis of the metabolic abnormalities in carbohydrate, fat, and protein in diabetes is insufficient action of insulin on various target tissues. Medicinal plants are rich sources of bioactive chemical compounds with therapeutic effects. The beneficial effects of leaves, fruits, and flowers extracts of *Crataegus oxyacantha*, commonly called hawthorn, belonging to the Rosaceae family, are widely used as hawthorn-derived medicines. Data in this review have been collected from the scientific articles published in databases such as Science Direct, Scopus, PubMed, Web of Science, and Scientific Information Database from 2000 to 2021. Based on this review, hawthorn extracts appear both therapeutic and protective effects against diabetic-related complications in various organs through molecular mechanisms, such as decreasing triglyceride, cholesterol, very low density lipoprotein and increasing the antioxidant activity of superoxide dismutase, catalase, glutathione peroxidase, total antioxidant capacity, decreasing malondialdehyde level, and attenuating tumor necrosis factor alpha, interleukin 6 and sirtuin 1/AMP-activated protein kinase (AMPK)/nuclear factor kappa B (NF- κ B) pathway and increasing the phosphorylation of glucose transporter 4, insulin receptor substrate 1, AKT and phosphoinositide 3-kinases, and attenuating blood sugar and regulation of insulin secretion, insulin resistance, and improvement of histopathological changes in pancreatic beta cells. Collectively, hawthorn can be considered as one new target for the research and development of innovative drugs for the prevention or treatment of DM and related problems.

1. Introduction

1.1. Diabetes Mellitus. Diabetes mellitus (DM) is one of the most common metabolic disorders marked by chronic hyperglycemia and occurs when the body is not able to properly manage insulin secretion, insulin function, or both [1, 2]. Type 1 diabetes mellitus (T1DM) accounts for only 5–10% of those with diabetes [3]. Autoimmune destruction of pancreatic beta cells in T1DM leads to intensive insulin deficiency [4, 5]. Type 2 diabetes mellitus (T2DM) is a

common type of DM in adults and accounts for approximately 90–95% of all diabetic individuals worldwide [3]. T2DM is manifested by insulin insensitivity because of insulin resistance, attenuating insulin production, and eventually pancreatic beta-cells dysfunction [6]. Insulin resistance, as the main pathophysiologic factor of T2DM, is manifested by disability of cells in glucose utilization [4, 7]. The basis of the metabolic abnormalities in carbohydrate, fat, and protein in diabetes is insufficient action of insulin in various target tissues [3]. Overweight, unhealthy dietary

patterns, sedentary lifestyle, genetic background, aging, and stress are known to be implicated in the progression of T2DM [8].

Diabetic-related complications including hypertension, atherosclerosis, blindness, kidney failure, and stroke increase the mortality rate [9, 10]. DM is associated with typical diabetic symptoms such as polydipsia, polyuria, polyphagia, and weight loss [11]. Furthermore, impairment of gastrointestinal function, numbness in lower extremities, and neuronal dysfunction may be accompanied with chronic hyperglycemia [12]. Hyperglycemia leads to the disruption of metabolic processes in multiple organs and results in retinopathy, nephropathy, and neuropathy [13, 14]. Oxidative stress and activation of proinflammatory reactions are other factors which contributed to the tissue damage following chronic hyperglycemia [15, 16].

1.2. Hawthorn. Medicinal plants as rich sources of bioactive chemical compounds have beneficial therapeutic effects [17]. *Crataegus oxyacantha*, commonly called hawthorn, belongs to the Rosaceae family. It is a fruit-bearing plant with green, yellow, orange, red, and black berries that grows mostly in Europe, Asia, North America, and Africa [18, 19]. The genus of *Crataegus* has more than 250 different species. The common name of hawthorn is currently used for all plant species of this genus [17, 19]. Hawthorn has gained growing attention due to its low toxicity and minimal unwanted side effects, which highlights its beneficial health effects and makes it easy to candidate as an alternative traditional medicine therapy. Leaves, fruits, and flowers extracts are widely used as the hawthorn-derived medicines [20, 21].

The leaves, flowers, and berries of hawthorn contain a variety of bioflavonoid-like complexes. Biflavonoids found in hawthorn plant include oligomeric procyanidins, vitexin, quercetin, and hyperoside. Other chemical constituents include vitamin C, saponins, tannins, cardiogenic amines (phenylethylamine, tyramine, isobutylamine, O-methoxyphenylethylamine, choline, and acetylcholine), purine derivatives (adenosine, adenine, guanine, caffeine, and amygdalin), triterpene acids, and ursolic acid [22]. Among these chemical bioactive molecules, flavonoids and procyanidins are recognized to be the key responsible components for most of the observed therapeutic effects [23, 24].

1.3. Toxicology. *Crataegus* has low toxicity, with an LD50 of 25 mg/kg [25]. Administration of excessive dosing of hawthorn flower extract (600 mg/kg/day; flavonoids) over 30 days in rats showed unremarkable adverse effects. In humans, the acute oral toxicity of hawthorn was 6 g/kg [26].

2. Biological Activities

It is well known that *Crataegus* species (hawthorn) have been used traditionally as a drug or supplement to promote antioxidant [27], anti-inflammatory [28], antimicrobial [29], hypoglycemic [24, 30], hypolipidemic [24], and hepatoprotective effects [31]. Treatment of atherosclerosis, urinary retention, hypertension, intestinal disorders, and

brain and heart diseases have also been attributed to hawthorn [17, 18]. Hawthorn also has shown promise in the treatment of mild-to-moderate heart failure [32]. Nowadays, hawthorn is frequently used as an antidiabetic therapeutic agent in various metabolic disorders [30, 33]. The major findings suggest that the protective effects of *Crataegus* extract against diabetic-related complications most likely involve blood glucose level lowering, hypolipidemic effect, and antioxidant activity [34] and its ability to normalize insulin secretion [30, 35].

Fruit extract of hawthorn has been reported to possess lipid lowering effect. Other study showed that hawthorn leaf extract has hypolipidemic effect in diabetic models [36]. In addition, hawthorn leaf extract has been shown to have valuable hypoglycemic effect in streptozotocin- (STZ-) induced diabetic model [37] and antioxidant properties of hawthorn has been considered responsible for this beneficial effect [30]. Altogether, the underlying mechanisms of these pharmacological agents may be related to intestinal α -glucosidase inhibition [34, 38], decreased hepatic gluconeogenesis [39, 40], improving the lipid metabolism [39, 41], and insulin sensitivity restoration [35]. Clinically, hawthorn may be one of the future optimizing medicinal plants that shows dual effect against hyperglycemia and hyperlipidemia among human diabetic type 2 patients [42]. Data of this review have been collected from the scientific articles published in databases such as Science Direct, Scopus, PubMed, Web of Science, and Scientific Information Database from 2000 to 2021. The aim of this study is basic mechanistic information about diabetes mellitus and its complications as well as the ways which hawthorn extracts can modulate the adverse effects of diabetes.

2.1. Effect of Hawthorn Extracts on Diabetic-Induced Pancreatic Injury. DM is a prolonged disturbance related to irreversible destruction of islet β -cells insulin production, which is determined by hyperglycemia and altered fat metabolism [43]. β -cells in the islets of Langerhans are major location of synthesis and secretion of insulin [44]. In most individuals, a normal blood glucose concentration or normoglycemia is preserved by an increasing in insulin secretion. However, in persons predisposing to develop T2DM [45], β -cells unable to compensate of insulin resistance, which leads to glucose intolerance, fasting blood sugar increase, and eventually overt diabetes [46].

Three main transcription factors, including pancreas/duodenum homeobox protein 1 (PDX1), neurogenin 3 (Neurog 3), and v-maf musculoaponeurotic fibrosarcoma oncogene homolog A (MAFA), participated in early pancreatic progenitor formation, endocrine lineage specification and differentiation, and maturation of β -cells in the late stage, respectively [47, 48]. They contributed to reprogram of multiple cell types into insulin-producing cells for treatment of T1DM [49]. In this regard, it has been demonstrated that overexpression of PDX1, Neurog 3, and/or MAFA can induce insulin biosynthesis in various non- β -cells [50].

PDX1 is also essential for the differentiation of pancreatic lineages and maintenance of mature β -cells function

[51] and also preserves pancreas against apoptosis [52]. Neurog 3 belongs to the basic helix-loop-helix transcription factor and participates in the nervous system and embryonic pancreas progress [49]. MAFA protein is a subgroup of MAF transcription family that specifically binds to insulin enhancer element RIPE3b and stimulates insulin gene expression [53].

Neurog 3 and MAFA are responsible for islet improvement. Insulin 1 (Ins⁻¹) and insulin 2 (Ins⁻²) promote the synthesis of proinsulin [54]. The upregulation of Ins⁻¹ or Ins⁻² may be triggered by the increased expression levels of MAFA, PDX1, and Neurog 3. It has been shown that the protein expression of MAFA, PDX1, and Neurog 3 significantly attenuated in the diabetic pancreatic tissue and treatment with *Crataegus* flavonoids (CF) at the dose of 200 mg/kg/orally for four weeks increased the protein expression pattern of MAFA, PDX1, and Neurog 3. Therefore, CF may restore the islet cells function by regulating the expression of the previously mentioned factors. CF also can ameliorate glucose intolerance and decline diet and water intake in the diabetic mice [55].

It has been observed that STZ administration leads to the islet β -cells toxicity and a significant reduction in the size of islets of Langerhans. The partly protective effects of CF on the islet β -cells are attributed to the improvement of histopathological changes of the mentioned cells [55]. Furthermore, STZ-induced diabetes is associated with deterioration of the acinar cells of the pancreas, insult to lobules, and edema. Treatment with hydroalcoholic extract of *Crataegus monogyna* at the different doses of 100, 200, and 400 mg/kg intraperitoneally for the duration of three weeks alleviated the previously mentioned histological alterations in diabetic rats [55].

2.2. Effect of Hawthorn Extracts on Diabetic-Induced Hepatic Injury. The liver is responsible for about 80% of endogenous glucose synthesis [56] and plays an important function in regulating of glucose homeostasis via two major mechanisms including glycogenolysis and gluconeogenesis [57]. The binding of insulin to its receptor phosphorylates PI3K/AKT pathway then regulates glycogen synthesis through inhibiting of glycogen synthase as a major enzyme in glycogenolysis. Furthermore, phosphorylation of PI3K/AKT via forkhead box protein O1 (FOXO1) inhibits phosphoenolpyruvate carboxykinase (PEPCK) and glucose 6-phosphatase (G6Pase), as key rate limiting enzymes in gluconeogenesis [58].

It has been shown, in T2DM, that hepatic insulin receptors 1A (IR-1A) deficiency is associated with an increase in gluconeogenesis, suppressing of glycogen synthesis, elevating the levels of hemoglobin A1c (HbA_{1c}), and upregulating of glucose transporter 2 (GLUT2) and G6Pase mRNAs and downregulating of glycerol kinase (GK) mRNA. In addition, the activities of the glutathione (GSH) and superoxide dismutase (SOD) were decreased. Furthermore, the levels of inflammatory cytokines including tumor necrosis factor alpha (TNF- α) and interleukin 6 (IL-6) and thiobarbituric acid reactive substances (TBARS) in hepatic

tissue were increased. The serum levels of triglyceride (TG), total cholesterol (TC), low density lipoprotein (LDL), and very low density lipoprotein (VLDL) were increased in T2DM. As indicated in Table 1, *Crataegus aronia* application at the dose of 500 mg/kg/orally for 60 days significantly reversed all aforementioned alterations to near normal, evidently through hawthorn-mediated antioxidant and anti-inflammatory actions in diabetic rats [59].

On the other hand, the hypoglycemic and hypolipidemic actions of hawthorn administration at different doses of 0.2, 0.5, and 1.0 g/kg for four weeks by oral administration regulated the expression of hepatic gluconeogenesis and lipogenesis associated genes including sterol regulatory element binding protein-1c (SREBP-1c) and fatty acid synthase (FAS). Furthermore, blood glucose lowering effect of hawthorn may be attributed to attenuating of gluconeogenesis in the liver tissue through downregulating of PEPCK gene expression following AMP-activated protein kinase (AMPK) activation [39].

Peroxisome proliferator-activated receptor α (PPAR α), as a nuclear receptor, has a pivotal role in lipid metabolism [60], especially in hepatic lipid metabolism through the induction of various genes such as fatty acid transport proteins and fatty acid oxidation cascades [61]. PPAR α also has an essential function in SREBP-mediated regulation of lipogenic genes [62]. Furthermore, PPAR α reduces the serum levels of TG, while it elevates high-density lipoprotein-cholesterol (HDL-C) in patients with dyslipidemia [63]. In this regard, it has been shown that hawthorn effectively increases the mRNA expression levels of PPAR α in the liver tissue of high fat diet mice model, whereas it inhibits the expression of SREBP1c and SREBP2 contributed to the triacylglycerol and total cholesterol synthesis, respectively [39].

2.3. Effect of Hawthorn Extracts on Diabetic-Induced Gastric Injury. Gastroparesis is explained as a syndrome specified by dysfunction in stomach resulting in delayed gastric emptying without mechanical obstruction [64]. Diabetic gastroparesis (DGP) is a well-established chronic side effects of diabetes. It has been reported that 30–50% of patients with T1DM or T2DM suffer from gastroparesis [65]. Furthermore, the risk of developing DGP, being over 7 and 30-times for T2DM and T1DM, respectively. DGP is a highly symptomatic disorder determined by fullness, anorexia [66], early satiety, abdominal pain [67], weight loss, abdominal distension, nausea and vomiting [68], gastric dysrhythmia, antral hypomotility, and/or delayed gastric emptying [65].

Numerous factors such as autonomic system neuropathy, disturbance in neurotransmission [69], hyperglycemia, neurological disorders, alterations in gastrointestinal (GI) hormones, microvascular and GI smooth muscle abrasions, and structural and practical irregularity of interstitial cells of Cajal (ICC) are involved in the pathogenesis of DGP [68]. Hyperglycemia-induced oxidative stress is a major mechanism for various complications of diabetes including DGP and leads to cellular malfunction and tissue damage [70]. The role of three main antioxidant enzymes such as SOD,

TABLE 1: Effects of hawthorn extracts on diabetic-induced injury in several organs.

Organ	Hawthorn extract	Dose(s)/route/ duration/animal	Effects	Reference (s)
Stomach	Ethyl acetate extract hawthorn seeds	3, 6, and 12 mg/kg/oral/4 weeks/rat	↓ BS&MDA SOD and GP _x ↑ ↑ plasma level and ghrelin gastric expression ↑ nNOS gastric expression c-kit gastric expression ↑ Gastric emptying and small intestinal ↑ propulsion ↑ Body weight	[68]
Liver	<i>Crataegus aronia</i>	500 mg/kg/60 days/oral/rat	↓ BS, HbA _{1c} , OGTT, TG, TC, LDL, VLDL, TBARS, TNF-α&IL-6 ↑ GSH, SOD, HDL, and hepatic glycogen ↓ GLUT-2 and G6Pase hepatic mRNA _s ↑ IR-A and GK hepatic mRNA _s	[59]
Liver	<i>Crataegus pinnatifida</i> Bge. var. <i>major</i> N.E. Br.	0.2, 0.5, 1.0 g/kg/4 weeks/oral/rat	↑ AMPK phosphorylation& HDL-C PPARs expression↑ ↓ PEPCCK and glucose production ↓ SREBP-1c, SREBP2, and FAS	[39]
Pancrease	<i>Crataegus monogyna</i> extract	100, 200, and 400 mg/kg/ip/3 weeks/rat	↓ BS and MDA TAC↑ ↓ histopathological alterations	[55]
Pancrease	<i>Crataegus flavonoids</i>	200 mg/kg/oral/4 weeks/mice	↑ Serum insulin levels ↑ PDX-1, MafA and Neurog 3, Ins-1, and Ins-2 ↓ Appetite ↓ Water intake ↓ OGTT ↓ Histopathological alterations	[55]
Heart	Hawthorn leaf flavonoids	50, 100, and 200 mg/kg/oral/16 weeks/rat	↓ BS and MDA ↓ TNF-α and NF-κB proteins expression ↓ mRNA expression of PKC-α & PKC-α protein ↓ Histopathological changes SOD ↑	[81]
Heart	<i>Crataegus oxyacantha</i> extract	100 mg/kg/oral/10 weeks/rat	↑ Body weight ↑ GPx ↓ FBS, CK-MB, and LDH ↓ MPO	[88]
Aorta	Hawthorn polyphenol extract	300 mg/kg/oral/4 weeks/rat	↓ TC, TG, FBS, OGTT, and LPS levels ↑ Insulin level and body weigh ↑ Phosphorylation of GLUT4, IRS1, AKT, and PI3K in liver ↓ TNF-α and IL-6 in serum ↑ Phosphorylation of IRS1 and AKT in skeletal muscle ↓ Protein expression of TNF-α and IL-6 in liver and skeletal muscle ↓ Protein expression of MCP-1 in the liver Repair of aortic injury via SIRT1/NF-κB/Wnt2/β-catenin pathway ↓ Inflammation and IR (liver): SIRT1/AMPK/NF-κB pathway ↓ Inflammation & IR (skeletal muscle): SIRT1/NF-κB pathway	[95]
Kidney	Hawthorne leaf flavonoids	200 mg/kg/oral/12 weeks/rat	↑ Body weight ↑ SOD and NO ↓ Urine protein concentration ↓ BUN, Cr, TG& MDA levels ↓ p38MPAK protein expression ↓ Histopathological changes	[116]
Brain	<i>Crataegus</i> extract	100, 300, and 1000 mg/kg/oral/2 weeks/rat	↓ Cholesterol, TG, and BS ↑ HDL Improvement of PAL	[24]

catalase (CAT), and glutathione peroxidase (GP_x), as the first line defense systems, are important in animals [71]. The beneficial effect of these antioxidants on GI disorder was reported [72] and may be also effective for DGP [70].

It is revealed that the antioxidant activity of SOD and GP_x decreased and the level of malondialdehyde (MDA), as

a major indicator of stress oxidative, enhanced significantly in underlying of DGP. As shown in Figure 1, the oral administration of ethyl acetate extract hawthorn seeds (HSEAE) at the doses of 3, 6, and 12 mg/kg for four weeks reverses all these changes in diabetic rats. This implies that administration of HSEAE can beneficially suppress oxidative stress [68].

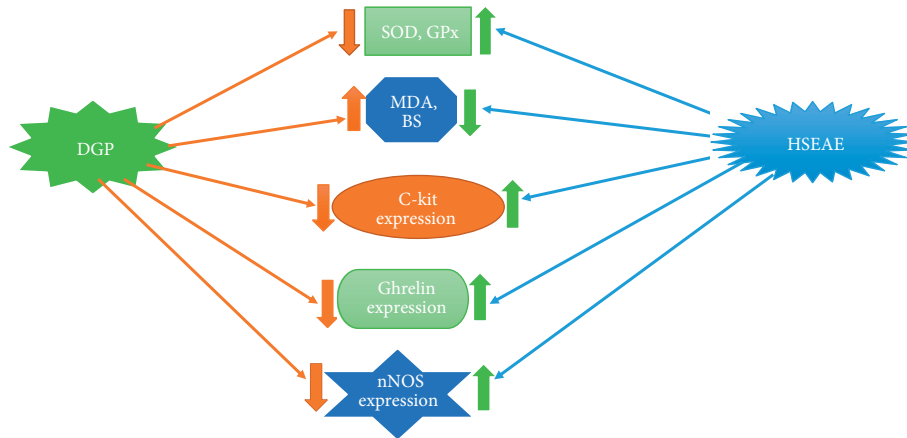


FIGURE 1: Schematic presentation of HSEAE on DGP and possible mechanisms.

Several hormones secreted by the GI system participate in regulating of GI motor activity [73]. Ghrelin is an endogenous ligand of growth hormone releasing hormone receptor, released into the bloodstream from the stomach. Ghrelin activates growth hormone and gastric acid secretion via the hypothalamus and accelerates GI peristalsis and food intake [74]. It is presented that plasma and gastric ghrelin expression in DGP significantly were downregulated but dramatically upregulated following administration of HSEAE [68].

Earlier study showed that reduction of neuronal nitric oxide synthase (nNOS) activity or protein expression in the gastric neurons of DGP results in a decrease in the nitric oxide (NO) production, which decelerates gastric activity [75]. In this regard, one experimental study showed that the administration of extract of hawthorn seeds increased the decreased protein expression of nNOS in the gastric of DGP subject, which may be advantage for facilitating gastric motility [68]. ICCs are the pacemaker for gut motility. They have beneficial effects on contractile activity of stomach muscle and eventually promote the gastric emptying [76]. ICCs are identified by mature molecular marker c-kit. Downregulation of c-kit expression critically influences the normal function, growth, and maturity of ICCs and declines their number and disturbances ultrastructural [66]. It has been revealed that the expression of c-kit protein in the DGP subjects was significantly downregulated and upregulated after the administration of HSEAE [68].

The molecular mechanism underlying ethyl acetate extract hawthorn seeds (HSEAE, 3, 6, and 12 mg/kg/oral/4 weeks/rat) on diabetic gastroparesis (DGP) may be related to elevating the antioxidant enzymes activity of superoxide dismutase (SOD), catalase (CAT), upregulating expression of C-kit, ghrelin, nitric oxide synthase (nNOS), and also attenuating the blood glucose (BS) and malondialdehyde (MDA) levels.

2.4. Effect of Hawthorn Extracts on Diabetic-Induced Cardiac Injury. Diabetic cardiomyopathy (DCM) is determined by a direct damage to cardiac muscle and subsequently structural and practical alterations in myocardium in absence of

hypertension and other cardiovascular problems. Hyperglycemia has a pivotal role in the pathogenesis of DCM. Cardiac hypertrophy, ventricular electrophysiological disturbances, and heart failure are the most important complications of DCM [77].

It has been reported that DCM is accompanying with oxidative stress, expression levels of inflammatory proteins, apoptosis, and accumulation of extracellular matrix [78]. Hyperglycemia results in reactive oxygen species (ROS) overproduction that led to the oxidative stress-related cardiac injury and major changes in myocardial cells, and consequently DCM. Antioxidants can eliminate a little amount of ROS in the normal physiological situation, but in pathological conditions for example in diabetes, because of the excessive ROS production, antioxidants cannot remove ROS and therefore induce insult to heart cells [79]. It is revealed that the activity of SOD, one of the most significant antioxidant enzymes [80], markedly decreased following DCM. Pretreatment with doses of 50, 100, and 200 mg/kg of hawthorn leaf flavonoids (HLF) for 16 weeks significantly increased the activity of SOD and decreased the increased levels of MDA in underlying DCM [81].

Protein kinase C (PKC) plays a fundamental role in regulation of multiple cellular processes [82]. Evidence indicated that the PKC activation can be involved in the cardiovascular injury through increasing the synthesis of extracellular matrix, regulating the calcium ion metabolism of cardiac cells, stimulating angiotensin II, and inducing ROS generation and inflammatory agents [83, 84]. The mRNA and protein expression of PKC- α were evaluated following experimental DCM. It is observed that the mRNA and protein expression of PKC- α in the cytoplasm and cytomembrane of cardiac cells dramatically increased. Pretreatment with HLF significantly downregulates these parameters in underlying DCM [81].

Inflammation participates in the progress of many cardiovascular disorders including hypertension, atherosclerosis, ischemic heart diseases, and congestive heart failure [85]. Furthermore, inflammatory cytokines have a key role in cardiac disturbance following myocardial infarction [86]. The correlation between TNF- α and nuclear factor kappa B (NF- κ B) has been reported in myocardial

dysfunction in underlying diabetes. The stimulated NF- κ B can dramatically upregulate the expression of TNF- α in left ventricular myocardium of diabetic subjects; however, TNF- α activates overexpression of NF- κ B [87]. Experimental evidence illustrated that HLF treatment significantly downregulates the overexpression of NF- κ Bp65 and TNF- α in cardiac tissue after induction of diabetes [81].

The cardioprotective actions of *Crataegus oxyacantha* extract on cardiac ischemia/reperfusion (I/R) damage in underlying diabetes has been investigated. The advantage effects may be related partly to antioxidant effects of hawthorn [88]. Lactate dehydrogenase (LDH) and creatine kinase myocardial band isoenzyme (CK-MB) are the most sensitive and specific indices for the evaluation of cardiac insult. Myocardial cells death leads to a pivotal increase in the serum levels of these enzymes [89]. Scientific evidence showed that the serum levels of LDH and CK-MB significantly increased after cardiac I/R injury associated with diabetic induction. Moreover, this study suggested that the previously mentioned indicators dramatically reduce following the oral administration of *Crataegus oxyacantha* extract at the dose of 100 mg/kg for 10 weeks in diabetic rats [88].

Oxidative stress contributed in I/R injury development, and these effects aggravated in diabetes [90]. Myeloperoxidase (MPO) as an oxidative stress parameter leads to ROS generation. Combination treatment of *Crataegus oxyacantha* extract with resistance training significantly attenuated the increased levels of MPO following cardiac I/R insult in underlying diabetes [88]. Experimental study indicated that diabetes induced-oxidative stress reduces the activity of GPx, which leads to the aggregation of hydrogen peroxide levels [91]. GPxas, a major ROS scavenger, accelerates the oxidation of glutathione via cumene hydroperoxide. It is reported that *Crataegus oxyacantha* extract, along with resistance training, significantly increased the decreased activity of GPx [88].

2.5. Effect of Hawthorn Extracts on Diabetic-Induced Vascular Injury. Insulin resistance (IR) is a major etiology and pathogenesis for T2DM. Development of IR is mostly associated with oxidative stress and/or inflammatory reactions induced by numerous proinflammatory agents such as interleukin-1 beta (IL-1 β), IL-6, and TNF- α [92]. Oxidative stress results in a mismatch between the ROS generation and antioxidant system. Pancreatic β -cells, adipocytes, and peripheral tissues are more susceptible to the destruction effects of oxidative stress [93]. Oxidative stress leads to impairment of insulin secretion in β -cells and IR progress in adipocytes and peripheral tissues that eventually accelerates postprandial hyperglycemia and overt T2DM. As shown in Figure 2, both postprandial hyperglycemia and T2DM act as feedback loop for the occurrence of oxidative stress [92]. It has been indicated the potential therapeutic or preventive effects of polyphenols as free radical scavengers in diabetes and its complications [94].

These events are associated with impairment of insulin secretion in β -cells and development of insulin resistance in

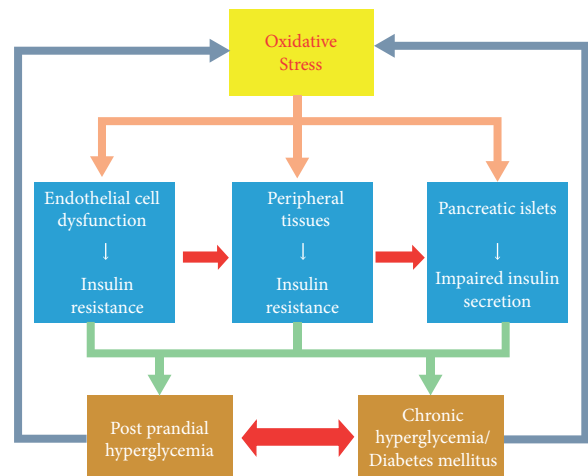


FIGURE 2: Schematic presentation of correlation between oxidative stress and DM. Oxidative stress displays dangerous effects on β -cells of pancreatic islets, adipocytes, and peripheral tissues.

adipocytes and peripheral tissues, which lead to the progress of postprandial hyperglycemia and overt T2DM, both of which also operate as feedback loop for the occurrence of oxidative stress [92].

It has been shown that hawthorn polyphenols extract (HPE) at the dose of 300 mg/kg for four weeks can decrease the progression of T2DM by attenuating ROS and increasing the CAT, GPx, and total antioxidant capacity (TAC) in serum and colonic tissue of diabetic rats model. This study also confirmed that HPE can effectively regulate altering parameters such as fasting blood sugar (FBS), oral glucose tolerance test (OGTT), TG, TC, insulin, lipopolysaccharide (LPS) serum levels, and body weight [95]. GLUTs facilitate absorption of glucose from the circulation into the cells. GLUT4 is essentially expressed in skeletal muscles, adipocytes, and cardiomyocytes. GLUT4 particularly stimulates glucose uptake into the muscle and adipose cells [96].

Insulin receptor substrate (IRS) family has six proteins (IRS1-IRS6). IRS1 and IRS2 are essential in insulin signaling pathways [97]. IRS-1 plays a pivotal role in activation of phosphatidylinositol 3-kinase (PI3K) as the active center of most insulin metabolic actions [98]. It is well established that attachment of insulin-to-insulin receptors is associated with tyrosine phosphorylation of IRS-1, which advances signal transduction via the IRS/PI3K signaling pathway and leads to glucose and fat metabolism [99]. Furthermore, stimulation of the PI3K/AKT pathway increases insulin secretion from pancreatic β -cells [100]. In this regard, it has been shown that PI3K can enhance AKT kinase (protein kinase B or PKB) activity by stimulating AKT tyrosine phosphorylation and finally provoke glucose transport, lipogenesis, glycogen synthesis, and repression of gluconeogenesis [98].

On the other hand, it is indicated that disturbance of translocation of GLUT4, as a downstream target of IRS1/PI3K/AKT signaling pathway, from intracellular to cytomembrane is one of the important etiology of IR in T2DM [101]. Another study presented that IR can improve through the stimulation of IRS1-PI3K-AKT-GLUT4 network in diabetic model [102]. In addition, it has been observed that

HPE effectively enhances the phosphorylation of GLUT4 and IR-associated proteins such as IRS1, AKT, and PI3K in the liver tissue and dramatically increases the expression levels of phosphorylation (p)-IRS1 and p-AKT in the skeletal muscle of diabetic rats [95].

Besides, it has been described that inflammatory markers, including IL-6, TNF- α , and monocyte chemoattractant protein-1 (MCP-1), contributed to IR. These proinflammatory cytokines interfere with insulin signaling networks and influence the insulin signaling kinase action [103]. In addition, chronic inflammation in adipose, liver, and skeletal muscle tissues are involved in the incidence and pathogenesis of T2DM [104]. It is investigated that under action of HPE the serum levels of IL-6 and TNF- α as well as protein expression of TNF- α and IL-6 in skeletal muscle and liver significantly mitigate in the diabetic model. This polyphenol also attenuates the protein expression of MCP-1 in the hepatocytes [95].

NF- κ B is one of the major transcriptional mediated pathways, which participates in numerous inflammatory reactions [105]. It accelerates the expression of downstream target proinflammatory cytokines, such as TNF- α , IL-6, and MCP-1 in the nucleus [95].

The activation of NF- κ B signaling cascade in the aorta of diabetic model has been investigated and reported that phosphorylation of I κ B, as inhibitor of NF- κ B, and breakdown of its lead to NF- κ Bp65 entrance to the nucleus, which induces vascular injury and upregulates proinflammatory markers expression [106]. HPE through decreasing the protein expression of NF- κ Bp65 phosphorylation attenuates the expression of aortic inflammatory agents in the diabetic model [95]. On the other hand, it has been investigated that sirtuin 1 (SIRT1)/AMPK pathway decreases the transcriptional stimulation of NF- κ B [107]. It has been reported that activation of AMPK regulates glucose homeostasis through inhibition of gluconeogenesis and attenuation of glucose levels [108].

SIRT1 is one of most members of sirtuin (SIRT1-7) family whose enzymatic activity is related to nicotinamide adenosine dinucleotide (NAD⁺) cofactor. It is indicated that upregulation of SIRT1 improves insulin sensitivity [109]. Moreover, interaction of SIRT1 with p65 subunit of NF- κ B and its deacetylation inhibits NF- κ B downstream signaling pathway [110]. As shown in Figure 3, HPE attenuates chronic inflammation in the liver and skeletal muscle via the SIRT1/AMPK/NF- κ B and SIRT1/NF- κ B signaling pathways, respectively, thereby suppressing IR in the diabetic model [95].

Diabetic-induced vascular insult is one of the etiologies of mortality rate among patients with diabetes [111]. Wnt/ β -catenin signaling cascades participate in macrovascular injury [112]. The interaction of Wnt ligand with the transmembrane receptors stabilises cytoplasmic β -catenin and ultimately initiates the transcription of downstream target genes of Wnt pathway [113]. It is reported that aberrations of Wnt/ β -catenin signaling pathways are associated with glucose metabolism disorders and increase the risk of T2DM [114]. On the other hand, Wnt/ β -catenin signaling pathway is stimulated by upregulation of SIRT1 [115]. It is

observed that HPE contributed to aortic damage repair via modulating the NF- κ B, downregulating the Wnt/ β -catenin signaling pathway, regulating the expression of its downstream associated proteins, and stimulating the regulatory action together with SIRT1 in diabetic model [95].

Pretreatment with hawthorn polyphenol extract (HPE) via oral administration for four weeks has regulatory effects on oxidant and inflammation agents in serum and colonic tissue and suppresses insulin resistance through inhibition of SIRT1/AMP-activated protein kinase (AMPK)/nuclear factor kappa B (NF- κ B) pathway in the liver and SIRT1/NF- κ B cascade in the skeletal muscles of diabetic rats model. HPE also repairs aortic injury via SIRT1/NF- κ B/Wnt2/ β -catenin pathway.

2.6. Effect of Hawthorn Extracts on Diabetic-Induced Renal Injury. The kidney has a substantial role in glucose handling through gluconeogenesis, glucose filtration, glucose reabsorption, and glucose utilization. Each of these ways can be changed in patients with T2DM [117]. As mentioned earlier, DM is determined by hyperglycemia, polyuria (an excessive urination), polydipsia (a great thirst), polyphagia (an enhanced appetite), and glycosuria [118]. Diabetic kidney disease (DKD) is a danger kidney-related complication of DM, which leads to end-stage renal disease [119]. Multiple factors, such as oxidative stress [120], genetic factors, glucose metabolism disorder, alteration in hemodynamic, and inflammation, participate in the pathogenesis of DKD [121].

Diabetic-induced oxidative stress leads to accumulation of ROS in kidney cells and subsequent stimulated p38 mitogen activated protein kinase (p38MAPK), which is involved in cellular stress, inflammation, and apoptosis through various processes and further exacerbates DKD progression [122]. On the other hand, it has been reported that, following the diabetic nephropathy, renal cell activity of p38 quickly increases in glomeruli, tubules, and renal interstitial cells [123]. Previous study showed that suppression of the p38MAPK signaling pathway can dramatically attenuate the risk of proteinuria in DKD, decrease the secretion of inflammatory mediators, and eventually decline DKD development [124].

Previous literature pointed out that HLF, at the dose of 200 mg/kg for 12 weeks through oral administration, can increase antioxidant capacity and prevent the process of lipid peroxidation in diabetic rats [116]. Another study showed that pretreatment with HLF effectively decreased the increased p38MAPK protein expression in the renal tissue of DKD. As shown in Figure 4, HLF also significantly decreased the elevated serum biomarkers, including blood urea nitrogen (BUN), creatinine (Cr), TG, MDA levels, and urine protein in DKD. Moreover, HLF effectively increased the reduced SOD, NO levels, body weight, and improved histopathological changes of DKD [116].

The possible mechanisms of hawthorn leaf extracts on diabetic nephropathy attributed to increasing SOD and nitric oxide (NO), and decreasing the blood urea nitrogen (BUN), creatinine (Cr), triglyceride (TG), MDA levels, urine protein concentration, 38mitogen activated protein kinase

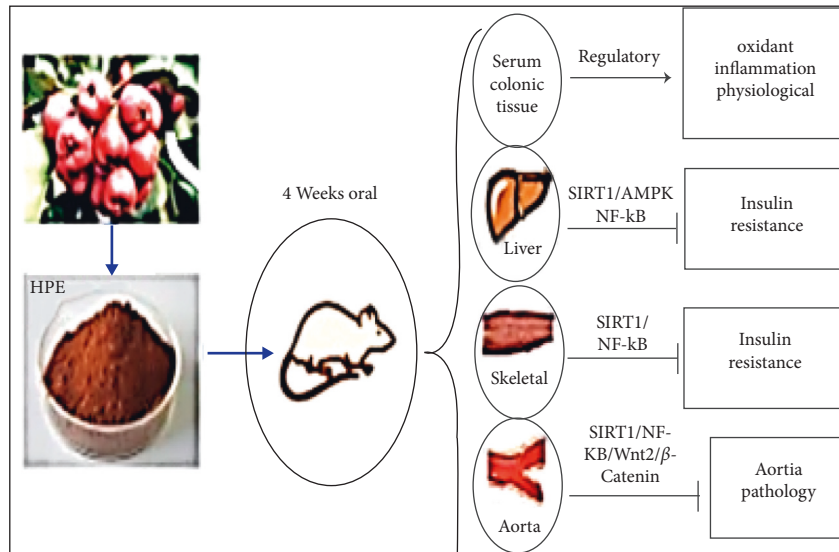


FIGURE 3: Molecular mechanisms of HPE on diabetic-aorta injury [95].

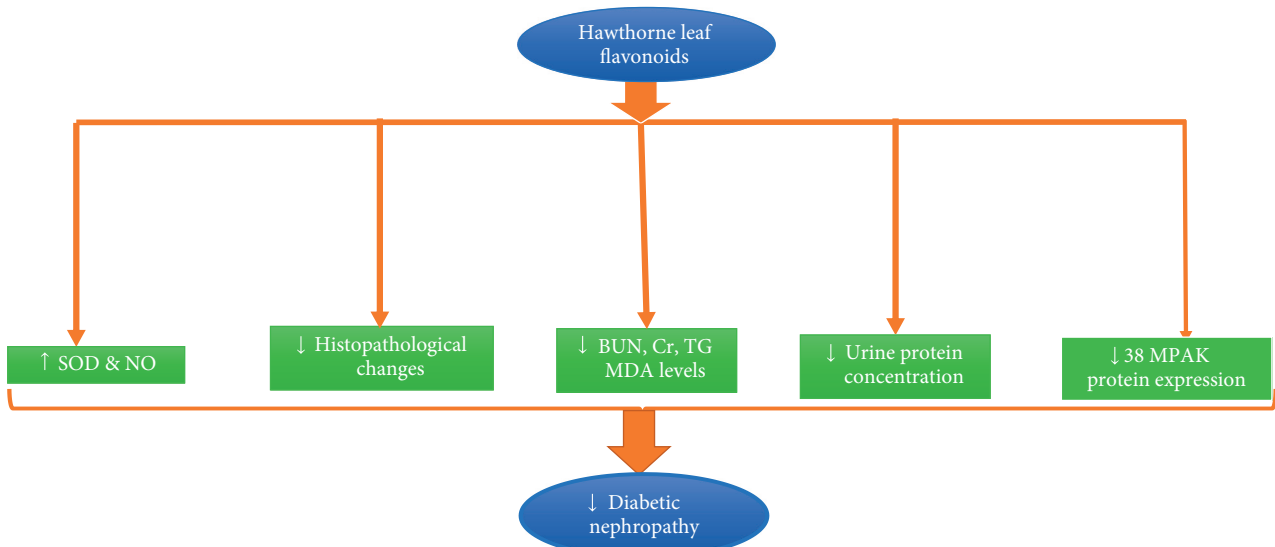


FIGURE 4: Effect of hawthorn extracts (200 mg/kg/oral/12 weeks) on the diabetic nephropathy.

(38MPAK) protein expression, and improving the histopathological changes of kidneys tissue.

2.7. Effect of Hawthorn Extracts on Diabetic-Induced Cerebral Injury. DM is considered as a predisposing factor for cognitive disturbance [125] and the progress of Alzheimer’s disease and dementia [126]. Furthermore, passive avoidance learning (PAL) and memory defective take place in STZ-induced diabetes [127]. The analgesic effects of hawthorn extract on spontaneous locomotors activities and exploratory behaviors have been investigated [128]. In this relation, it is reported that *Crataegus* extract at the doses of 100, 300, and 1000 mg/kg for two weeks by oral administration ameliorated PAL and attenuated time spent in the dark compartment of diabetic rat model [24]. Furthermore,

hawthorn extract and resistance training improved cognitive deficits in STZ -induced diabetic rats [129].

It has been demonstrated that hawthorn extract supplementation decreased the increased BS, obesity-related factors, including TG and cholesterol and increased the decreased density HDL levels in the serum of diabetic model [24]. The effects of hawthorn extract on lipid metabolism may be attributed to the modulating of lipoprotein lipase (LPL) expression through peroxisome proliferator response element cascade [130] and suppressing synergistically 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase and cholesterol assimilation [131]. Another study showed that flavonoids are one of the major components of hawthorn extracts, which operate as an alpha-amylase inhibitor, glycemic management and have beneficial effects on dyslipidemia in diabetic model [132].

3. Conclusion

Based on multiple reports as mentioned in this manuscript, hawthorn extracts supplementation significantly improved diabetic-induced injuries in several organs, such as pancreases, stomach, liver, heart, vessel, kidneys, and brain. The findings suggest that the effects of *Crataegus* extracts against diabetic-related complications are most likely related to improving the physiological functions, including enhancement of body weight, insulin secretion, OGTT, blood glucose level lowering, hypolipidemic action via decreasing TG, cholesterol, and antioxidant effect through increasing the activity of SOD, CAT, GP_x, and TAC and decreasing MDA level and anti-inflammatory effects through attenuating TNF- α , IL-6, and SIRT1/AMPK/NF- κ B pathway. Other mechanisms are increasing the phosphorylation of GLUT4, IRS1, AKT, and PI3K and attenuating the urine protein concentration, p38MAPK protein expression, BUN, and Cr and improving the histopathological changes. Collectively, hawthorn extracts can be regarded as one new target for the research and development of innovative drugs to the prevention or treatment of DM and related complications.

Abbreviations

DM:	Diabetes mellitus
T1DM:	Type 1 diabetes mellitus
T2DM:	Type 2 diabetes mellitus
PDX1:	Pancreas/duodenum homeobox protein 1
Neurog 3:	Neurogenin 3
Ins ⁻¹ :	Insulin ¹
Ins ⁻² :	insulin ²
CF:	Crataegus flavonoids
STZ:	Streptozotocin
FOXO1:	Forkhead box protein O1
PEPCK:	Phosphoenolpyruvate carboxykinase
G6Pase:	Glucose 6-phosphatase
IR-1A:	Insulin receptors 1A
HbA _{1c} :	Hemoglobin A1c
GLUT2:	Glucose transporter 2
GK:	Glycerol kinase
GSH:	Glutathione
SOD:	Superoxide dismutase
TNF- α :	Tumor necrosis factor alpha
IL-6:	Interleukin 6
TBARS:	Thiobarbituric acid reactive substances
TG:	Triglyceride
TC:	Total cholesterol
LDL:	Low density lipoprotein
VLDL:	Very low-density lipoprotein
SREBP-1c:	Sterol regulatory element binding protein-1c
FAS:	Fatty acid synthase
AMPK:	AMP-activated protein kinase
PPAR α :	Peroxisome proliferator-activated receptor α
HDL-C:	High-density lipoprotein-cholesterol
DGP:	Diabetic gastroparesis
GI:	Gastrointestinal
ICC:	Interstitial cells of Cajal
CAT:	Catalase

GPX:	Glutathione peroxidase
MDA:	Malondialdehyde
HSEAE:	Ethyl acetate extract hawthorn seeds
NNOS:	Neuronal nitric oxide synthase
NO:	Nitric oxide
DCM:	Diabetic cardiomyopathy
ROS:	Reactive oxygen species
HLF:	Hawthorn leaf flavonoids
PKC:	Protein kinase C
NF- κ B:	Nuclear factor kappa B
I/R:	Ischemia/reperfusion
LDH:	Lactate dehydrogenase
CK-MB:	Creatine kinase myocardial band isoenzyme
MPO:	Myeloperoxidase
IR:	Insulin resistance
IL-1 β :	Interleukin-1 beta
HPE:	Hawthorn polyphenols extract
TAC:	Total antioxidant capacity
FBS:	Fasting blood sugar
OGTT:	Oral glucose tolerance test
LPS:	Lipopolysaccharide
IRS:	Insulin receptor substrate
PKB:	Protein kinase B
MCP-1:	Monocyte chemoattractant protein-1
SIRT1:	Sirtuin 1
NAD ⁺ :	Nicotinamide adenosine dinucleotide
DKD:	Diabetic kidney disease
p38MAPK:	p38 mitogen activated protein kinase
BUN:	Blood urea nitrogen
Cr:	Creatinine
PAL:	Passive avoidance learning
LPL:	Lipoprotein lipase
HMG-CoA:	3-hydroxy-3-methylglutaryl-coenzyme A.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

The authors thank the support of Yasuj University of Medical Sciences, Yasuj, Iran.

References

- [1] American Diabetes Association, "Diagnosis and classification of diabetes mellitus," *Diabetes Care*, vol. 33, pp. S62–S69, 2010.
- [2] K. Papatheodorou, M. Banach, E. Bekiari, M. Rizzo, and M. Edmonds, "Complications of diabetes 2017," *Journal of Diabetes Research*, vol. 2018, Article ID 3086167, 4 pages, 2018.
- [3] American Diabetes Association, "Diagnosis and classification of diabetes mellitus," *Diabetes Care*, vol. 37, pp. S81–S90, 2014.
- [4] A. A. Oyewande, B. Iqbal, L. F. Abdalla, F. Karim, and S. Khan, "An overview of the pathophysiology of metabolic changes and their sequence of occurrence in obese diabetic females: a narrative review," *Cureus*, vol. 12, no. 10, Article ID e10947, 2020.

- [5] L. A. DiMeglio, C. Evans-Molina, and R. A. Oram, "Type 1 diabetes," *The Lancet*, vol. 391, no. 10138, pp. 2449–2462, 2018.
- [6] C. R. Kahn, "Insulin action, diabetogenes, and the cause of type II diabetes," *Diabetes*, vol. 43, no. 8, pp. 1066–1085, 1994.
- [7] V. Ormazabal, S. Nair, O. Elfeky, C. Aguayo, C. Salomon, and F. A. Zuñiga, "Association between insulin resistance and the development of cardiovascular disease," *Cardiovascular Diabetology*, vol. 17, no. 1, p. 122, 2018.
- [8] M. P. Plummer, M. E. Finnis, L. K. Phillips et al., "Stress induced hyperglycemia and the subsequent risk of type 2 diabetes in survivors of critical illness," *PLoS One*, vol. 11, no. 11, Article ID e0165923, 2016.
- [9] A. A. Siddiqui, S. A. Siddiqui, S. Ahmad, S. Siddiqui, I. Ahsan, and K. Sahu, "Diabetes: mechanism, pathophysiology and management-A review," *International Journal of Drug Development & Research*, vol. 5, no. 2, pp. 1–23, 2013.
- [10] Y.-F. Yang, T.-C. Li, C.-I. Li et al., "Visit-to-Visit glucose variability predicts the development of end-stage renal disease in type 2 diabetes," *Medicine*, vol. 94, no. 44, Article ID e1804, 2015.
- [11] G. B. Stefano, S. Challenger, and R. M. Kream, "Hyperglycemia-associated alterations in cellular signaling and dysregulated mitochondrial bioenergetics in human metabolic disorders," *European Journal of Nutrition*, vol. 55, no. 8, pp. 2339–2345, 2016.
- [12] A. M. Garcia-Serrano and J. M. N. Duarte, "Brain metabolism alterations in type 2 diabetes: what did we learn from diet-induced diabetes models?" *Frontiers in Neuroscience*, vol. 14, p. 229, 2020.
- [13] T. C. van der Pouw Kraan, W. J. Chen, M. Bunck et al., "Metabolic changes in type 2 diabetes are reflected in peripheral blood cells, revealing aberrant cytotoxicity, a viral signature, and hypoxia inducible factor activity," *BMC Medical Genomics*, vol. 8, no. 1, pp. 1–6, 2015.
- [14] J. Mohammadi, H. Delaviz, J. M. Malekzadeh, and A. Roozbehi, "The effect of hydro alcoholic extract of *Juglans regia* leaves in streptozotocin-nicotinamide induced diabetic rats," *Pakistan Journal of Pharmaceutical Sciences*, vol. 25, no. 2, pp. 407–411, 2012.
- [15] M. Monserrat-Mesquida, M. Quetglas-Llabrés, X. Capó et al., "Metabolic syndrome is associated with oxidative stress and proinflammatory state," *Antioxidants*, vol. 9, no. 3, p. 236, 2020.
- [16] S. D. Martin and S. L. McGee, "Metabolic reprogramming in type 2 diabetes and the development of breast cancer," *Journal of Endocrinology*, vol. 237, no. 2, pp. R35–R46, 2018.
- [17] A. Nazhand, M. Lucarini, A. Durazzo et al., "Hawthorn (*Crataegus* spp.): an updated overview on its beneficial properties," *Forests*, vol. 11, no. 5, p. 564, 2020.
- [18] S. A. Ganie, T. Ali Dar, S. Zargar et al., "Crataegus songaricamethanolic extract accelerates enzymatic status in kidney and heart tissue damage in albino rats and its in vitro cytotoxic activity," *Pharmaceutical Biology*, vol. 54, no. 7, pp. 1246–1254, 2016.
- [19] P. R. Venskutonis, "Phytochemical composition and bioactivities of hawthorn (*Crataegus* spp.): review of recent research advances," *Journal of Food Bioactives*, vol. 4, pp. 69–87, 2018.
- [20] A. M. Rababa'h, O. N. Al Yacoub, T. El-Elimat et al., "The effect of hawthorn flower and leaf extract (*Crataegus* Spp.) on cardiac hemostasis and oxidative parameters in Sprague Dawley rats," *Heliyon*, vol. 6, no. 8, Article ID e04617, 2020.
- [21] M. Dianat and G. Akbari, "Protective effect of hydroalcoholic extract of hawthorn fruit on CaCl₂-induced arrhythmias in rat," *Jundishapur Scientific Medical Journal*, vol. 12, no. 6, pp. 693–703, 2014.
- [22] S. K. Verma, V. Jain, D. Verma, and R. Khamesra, "Crataegus oxyacantha-A cardioprotective herb," *Journal of Herbal Medicine and Toxicology*, vol. 1, no. 1, pp. 65–71, 2007.
- [23] S. Dahmer and E. Scott, "Health effects of hawthorn," *American Family Physician*, vol. 81, no. 4, pp. 465–468, 2010.
- [24] A. Pirmoghani, I. Salehi, S. Moradkhani, S. A. Karimi, and S. Salehi, "Effect of Crataegus extract supplementation on diabetes induced memory deficits and serum biochemical parameters in male rats," *IBRO Reports*, vol. 7, no. 7, pp. 90–96, 2019.
- [25] H. Ammon and M. Händel, "Crataegus, Toxikologie und Pharmakologie", ** Teil I: toxizität," *Planta Medica*, vol. 43, no. 10, pp. 105–120, 1981.
- [26] D. Kumar, V. Arya, Z. A. Bhat, N. A. Khan, and D. N. Prasad, "The genus Crataegus: chemical and pharmacological perspectives," *Revista Brasileira de Farmacognosia*, vol. 22, no. 5, pp. 1187–1200, 2012.
- [27] C. Li, H. J. Son, C. Huang, S. K. Lee, J. Lohakare, and M.-H. Wang, "Comparison of Crataegus pinnatifida Bunge var. *typica* Schneider and C. *pinnatifida* Bunge fruits for antioxidant, anti- α -glucosidase, and anti-inflammatory activities," *Food Science and Biotechnology*, vol. 19, no. 3, pp. 769–775, 2010.
- [28] E.-S. Kao, C.-J. Wang, W.-L. Lin, Y.-F. Yin, C.-P. Wang, and T.-H. Tseng, "Anti-inflammatory potential of flavonoid contents from dried fruit of Crataegus pinnatifida in vitro and in vivo," *Journal of Agricultural and Food Chemistry*, vol. 53, no. 2, pp. 430–436, 2005.
- [29] W. Benabderrahmane, M. Lores, O. Benaissa et al., "Polyphenolic content and bioactivities of Crataegus oxyacantha L. (Rosaceae)," *Natural Product Research*, vol. 35, no. 4, pp. 627–632, 2021.
- [30] A. Aierken, T. Buchholz, C. Chen, X. Zhang, and M. F. Melzig, "Hypoglycemic effect of hawthorn in type II diabetes mellitus rat model," *Journal of the Science of Food and Agriculture*, vol. 97, no. 13, pp. 4557–4561, 2017.
- [31] J. Luis Martínez-Rodríguez, R. Gutiérrez-Hernández, C. Araceli Reyes-Estrada et al., "Hepatoprotective, anti-hyperlipidemic and radical scavenging activity of hawthorn (*Crataegus oxyacantha*) and rosemary (*rosmarinus officinalis*) on alcoholic liver disease," *Alternative Therapies in Health & Medicine*, vol. 25, no. 4, pp. 54–63, 2019.
- [32] J. M. Rigelsky and B. V. Sweet, "Hawthorn: pharmacology and therapeutic uses," *American Journal of Health-System Pharmacy*, vol. 59, no. 5, pp. 417–422, 2002.
- [33] S. Dehghani, S. Mehri, and H. Hosseinzadeh, "The effects of Crataegus pinnatifida (Chinese hawthorn) on metabolic syndrome: a review," *Iranian Journal of Basic Medical Sciences*, vol. 22, no. 5, pp. 460–468, 2019.
- [34] J. Miao, X. Li, Y. Fan et al., "Effect of different solvents on the chemical composition, antioxidant activity and alpha-glucosidase inhibitory activity of hawthorn extracts," *International Journal of Food Science and Technology*, vol. 51, no. 5, pp. 1244–1251, 2016.
- [35] J. Liu, H. Zhang, B. Ji et al., "A diet formula of Puerariae radix, Lycium barbarum, Crataegus pinnatifida, and Polygonati rhizoma alleviates insulin resistance and hepatic steatosis in CD-1 mice and HepG2 cells," *Food & Function*, vol. 5, no. 5, pp. 1038–1049, 2014.

- [36] D.-H. Kuo, C.-H. Yeh, P.-C. Shieh, K.-C. Cheng, F.-A. Chen, and J.-T. Cheng, "Effect of shanzha, a Chinese herbal product, on obesity and dyslipidemia in hamsters receiving high-fat diet," *Journal of Ethnopharmacology*, vol. 124, no. 3, pp. 544–550, 2009.
- [37] H. Jouad, A. Lemhadri, M. Maghrani, R. Burcelin, and M. Eddouks, "Hawthorn evokes a potent anti-hyperglycemic capacity in streptozotocin-induced diabetic rats," *Journal of Herbal Pharmacotherapy*, vol. 3, no. 2, pp. 19–29, 2003.
- [38] S. S. Chowdhury, M. N. Islam, H. A. Jung, and J. S. Choi, "In vitro antidiabetic potential of the fruits of *Crataegus pinnatifida*," *Research in Pharmaceutical Sciences*, vol. 9, no. 1, pp. 11–22, 2014.
- [39] C. C. Shih, C. H. Lin, Y. J. Lin, and J. B. Wu, "Validation of the antidiabetic and hypolipidemic effects of hawthorn by assessment of gluconeogenesis and lipogenesis related genes and AMP-activated protein kinase phosphorylation," *Evidence-based Complementary and Alternative Medicine*, vol. 2013, Article ID 597067, 12 pages, 2013.
- [40] W. Xie, D. Xing, Y. Zhao et al., "A new tactic to treat postprandial hyperlipidemia in diabetic rats with gastroparesis by improving gastrointestinal transit," *European Journal of Pharmacology*, vol. 510, no. 1-2, pp. 113–120, 2005.
- [41] T. Wang, Y. An, C. Zhao et al., "Regulation effects of *Crataegus pinnatifida* leaf on glucose and lipids metabolism," *Journal of Agricultural and Food Chemistry*, vol. 59, no. 9, pp. 4987–4994, 2011.
- [42] A. Abdulrahim and M. Al-Shawabkeh, "Effects of Hawthorn on HbA1C and lipids levels in Jordanian diabetic patients (Type2)," *Journal of Chemical and Pharmaceutical Sciences*, vol. 10, no. 2, pp. 822–825, 2017.
- [43] Z.-J. Yang, J. Liu, J.-P. Ge, L. Chen, Z.-G. Zhao, and W.-Y. Yang, "Prevalence of cardiovascular disease risk factor in the Chinese population: the 2007-2008 China national diabetes and metabolic disorders study," *European Heart Journal*, vol. 33, no. 2, pp. 213–220, 2012.
- [44] W. Xie, Y. Zhao, D. Gu, L. Du, G. Cai, and Y. Zhang, "Scorpion in combination with gypsum: novel antidiabetic activities in streptozotocin-induced diabetic mice by up-regulating pancreatic PPAR γ and PDX-1 expressions," *Evidence-based Complementary and Alternative Medicine*, vol. 2011, Article ID 683561, 9 pages, 2011.
- [45] Y. Zheng, S. H. Ley, and F. B. Hu, "Global aetiology and epidemiology of type 2 diabetes mellitus and its complications," *Nature Reviews Endocrinology*, vol. 14, no. 2, pp. 88–98, 2018.
- [46] C. Wysham and J. Shubrook, "Beta-cell failure in type 2 diabetes: mechanisms, markers, and clinical implications," *Postgraduate Medicine*, vol. 132, no. 8, pp. 676–686, 2020.
- [47] E. Conrad, R. Stein, and C. S. Hunter, "Revealing transcription factors during human pancreatic β cell development," *Trends in Endocrinology and Metabolism*, vol. 25, no. 8, pp. 407–414, 2014.
- [48] E. Yoshihara, Z. Wei, C. S. Lin et al., "ERR γ is required for the metabolic maturation of therapeutically functional glucose-responsive β cells," *Cell Metabolism*, vol. 23, no. 4, pp. 622–634, 2016.
- [49] Y. Zhu, Q. Liu, Z. Zhou, and Y. Ikeda, "PDX1, Neurogenin-3, and MAFA: critical transcription regulators for beta cell development and regeneration," *Stem Cell Research & Therapy*, vol. 8, no. 1, pp. 240–247, 2017.
- [50] H. Kaneto, T.-a. Matsuoka, N. Katakami, and M. Matsuhisa, "Combination of MafA, PDX-1 and NeuroD is a useful tool to efficiently induce insulin-producing surrogate 946; -cells," *Current Medicinal Chemistry*, vol. 16, no. 24, pp. 3144–3151, 2009.
- [51] A. M. Holland, M. A. Hale, H. Kagami, R. E. Hammer, and R. J. MacDonald, "Experimental control of pancreatic development and maintenance," *Proceedings of the National Academy of Sciences*, vol. 99, no. 19, pp. 12236–12241, 2002.
- [52] M. M. Sachdeva, K. C. Claiborn, C. Khoo et al., "Pdx1 (MODY4) regulates pancreatic beta cell susceptibility to ER stress," *Proceedings of the National Academy of Sciences*, vol. 106, no. 45, pp. 19090–19095, 2009.
- [53] T.-A. Matsuoka, I. Artner, E. Henderson, A. Means, M. Sander, and R. Stein, "The MafA transcription factor appears to be responsible for tissue-specific expression of insulin," *Proceedings of the National Academy of Sciences*, vol. 101, no. 9, pp. 2930–2933, 2004.
- [54] A. Zaldumbide, F. Carlotti, M. A. Gonçalves et al., "Adenoviral vectors stimulate glucagon transcription in human mesenchymal stem cells expressing pancreatic transcription factors," *PLoS One*, vol. 7, no. 10, Article ID e48093, 2012.
- [55] K. Cui, S. Zhang, X. Jiang, and W. Xie, "Novel synergic antidiabetic effects of *Astragalus polysaccharides* combined with *Crataegus flavonoids* via improvement of islet function and liver metabolism," *Molecular Medicine Reports*, vol. 13, no. 6, pp. 4737–4744, 2016.
- [56] J. E. Gerich, "Role of the kidney in normal glucose homeostasis and in the hyperglycaemia of diabetes mellitus: therapeutic implications," *Diabetic Medicine*, vol. 27, no. 2, pp. 136–142, 2010.
- [57] H. V. Lin and D. Accili, "Hormonal regulation of hepatic glucose production in health and disease," *Cell Metabolism*, vol. 14, no. 1, pp. 9–19, 2011.
- [58] X. Huang, G. Liu, J. Guo, and Z. Su, "The PI3K/AKT pathway in obesity and type 2 diabetes," *International Journal of Biological Sciences*, vol. 14, no. 11, pp. 1483–1496, 2018.
- [59] D. Mostafa, E. Khaleel, and G. Abdel-Aleem, "Inhibition of the hepatic glucose output is responsible for the hypoglycemic effect of *Crataegus aronia* against type 2 diabetes mellitus in rats," *Archives of Biological Sciences*, vol. 70, no. 2, pp. 277–287, 2018.
- [60] A. Peeters and M. Baes, "Role of PPAR in hepatic carbohydrate metabolism," *PPAR Research*, vol. 2010, Article ID 572405, 12 pages, 2010.
- [61] S. Mandard, M. Muller, and S. Kersten, "Peroxisome proliferator-activated receptor a target genes," *Cellular and Molecular Life Sciences*, vol. 61, no. 4, pp. 393–416, 2004.
- [62] R. A. Srivastava, R. Jahagirdar, S. Azhar, S. Sharma, and C. L. Bisgaier, "Peroxisome proliferator-activated receptor- α selective ligand reduces adiposity, improves insulin sensitivity and inhibits atherosclerosis in LDL receptor-deficient mice," *Molecular and Cellular Biochemistry*, vol. 285, no. 1, pp. 35–50, 2006.
- [63] B. Staels and J.-C. Fruchart, "Therapeutic roles of peroxisome proliferator-activated receptor agonists," *Diabetes*, vol. 54, no. 8, pp. 2460–2470, 2005.
- [64] H. P. Parkman, W. L. Hasler, and R. S. Fisher, "American Gastroenterological Association technical review on the diagnosis and treatment of gastroparesis," *Gastroenterology*, vol. 127, no. 5, pp. 1592–1622, 2004.
- [65] J. Yin, J. Chen, and J. D. Z. Chen, "Ameliorating effects and mechanisms of electroacupuncture on gastric dysrhythmia, delayed emptying, and impaired accommodation in diabetic rats," *American Journal of Physiology-Gastrointestinal and Liver Physiology*, vol. 298, no. 4, pp. G563–G570, 2010.

- [66] Y. Chen, H. Wang, H. Li, and S. Liu, "Long-pulse gastric electrical stimulation repairs interstitial cells of cajal and smooth muscle cells in the gastric antrum of diabetic rats," *Gastroenterology Research and Practice*, vol. 2018, Article ID 6309157, 10 pages, 2018.
- [67] T. R. Angeli and G. O'Grady, "Challenges in defining, diagnosing, and treating diabetic gastroparesis," *Journal of Diabetes and Its Complications*, vol. 32, no. 2, pp. 127-128, 2017.
- [68] Z. Niu, M. Yan, X. Zhao, H. Jin, and Y. Gong, "Effect of hawthorn seed extract on the gastrointestinal function of rats with diabetic gastroparesis," *South African Journal of Botany*, vol. 130, pp. 448-455, 2020.
- [69] P. Kashyap and G. Farrugia, "Diabetic gastroparesis: what we have learned and had to unlearn in the past 5 years: Figure 1," *Gut*, vol. 59, no. 12, pp. 1716-1726, 2010.
- [70] A. Hosseini and M. Abdollahi, "Diabetic neuropathy and oxidative stress: therapeutic perspectives," *Oxidative Medicine and Cellular Longevity*, vol. 2013, Article ID 168039, 15 pages, 2013.
- [71] O. M. Ighodaro and O. A. Akinloye, "First line defence antioxidants-superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX): their fundamental role in the entire antioxidant defence grid," *Alexandria journal of medicine*, vol. 54, no. 4, pp. 287-293, 2018.
- [72] G. Akbari, "Role of zinc supplementation on ischemia/reperfusion injury in various organs," *Biological Trace Element Research*, vol. 196, no. 1, pp. 1-9, 2020.
- [73] H. Zhang, T. Han, L.-N. Sun et al., "Regulative effects of essential oil from *Atractylodes lancea* on delayed gastric emptying in stress-induced rats," *Phytomedicine*, vol. 15, no. 8, pp. 602-611, 2008.
- [74] H. Ariga, K. Imai, K. Ludwig, and T. Takahashi, "Gastric emptying, plasma ghrelin and autonomic nerve activity in diabetic rats," *Neuroscience Letters*, vol. 514, no. 1, pp. 77-81, 2012.
- [75] P. R. R. Gangula, S. Mukhopadhyay, K. Ravella et al., "Tetrahydrobiopterin (BH4), a cofactor for nNOS, restores gastric emptying and nNOS expression in female diabetic rats," *American Journal of Physiology - Gastrointestinal and Liver Physiology*, vol. 298, no. 5, pp. G692-G699, 2010.
- [76] A. Pasternak, M. Szura, K. Gil, and A. Matyja, "Interstitial cells of Cajal - systematic review," *Folia morphologica*, vol. 75, no. 3, pp. 281-286, 2016.
- [77] A. Aneja, W. H. W. Tang, S. Bansilal, M. J. Garcia, and M. E. Farkouh, "Diabetic cardiomyopathy: insights into pathogenesis, diagnostic challenges, and therapeutic options," *The American Journal of Medicine*, vol. 121, no. 9, pp. 748-757, 2008.
- [78] R. Mortuza and S. Chakrabarti, "Glucose-induced cell signaling in the pathogenesis of diabetic cardiomyopathy," *Heart Failure Reviews*, vol. 19, 2014.
- [79] M. Joshi, S. R. Kotha, S. Malireddy et al., "Conundrum of pathogenesis of diabetic cardiomyopathy: role of vascular endothelial dysfunction, reactive oxygen species, and mitochondria," *Molecular and Cellular Biochemistry*, vol. 386, no. 1, pp. 233-249, 2014.
- [80] G. Bresciani, I. B. M. da Cruz, and J. González-Gallego, "Manganese superoxide dismutase and oxidative stress modulation," *Advances in Clinical Chemistry*, vol. 68, pp. 87-130, 2015.
- [81] Q. Min, Y. Bai, Y. Zhang et al., "Hawthorn leaf flavonoids protect against diabetes-induced cardiomyopathy in rats via PKC- α signaling pathway," *Evidence-based Complementary and Alternative Medicine*, vol. 2017, Article ID 2071952, 8 pages, 2017.
- [82] A. C. Newton, C. E. Antal, and S. F. Steinberg, "Protein kinase C mechanisms that contribute to cardiac remodeling," *Clinical Science*, vol. 130, no. 17, pp. 1499-1510, 2016.
- [83] Z. Li, C. S. Abdullah, and Z.-Q. Jin, "Inhibition of PKC- θ preserves cardiac function and reduces fibrosis in streptozotocin-induced diabetic cardiomyopathy," *British Journal of Pharmacology*, vol. 171, no. 11, pp. 2913-2924, 2014.
- [84] K. Huynh, B. C. Bernardo, J. R. McMullen, and R. H. Ritchie, "Diabetic cardiomyopathy: mechanisms and new treatment strategies targeting antioxidant signaling pathways," *Pharmacology & Therapeutics*, vol. 142, no. 3, pp. 375-415, 2014.
- [85] L. A. Bienvenu, J. Morgan, A. J. Rickard et al., "Macrophage mineralocorticoid receptor signaling plays a key role in aldosterone-independent cardiac fibrosis," *Endocrinology*, vol. 153, no. 7, 2012.
- [86] S. Bryan, B. Baregzay, D. Spicer, P. K. Singal, and N. Khaper, "Redox-inflammatory synergy in the metabolic syndrome," *Canadian Journal of Physiology and Pharmacology*, vol. 91, no. 1, pp. 22-30, 2013.
- [87] B. D. Shames, H. H. Barton, L. L. Reznikov et al., "Ischemia alone is sufficient to induce TNF- α mRNA and peptide in the myocardium," *Shock*, vol. 17, no. 2, pp. 114-119, 2002.
- [88] K. Ranjbar, E. Zarrinkalam, I. Salehi, A. Komaki, and B. Fayazi, "Cardioprotective effect of resistance training and *Crataegus oxyacantha* extract on ischemia reperfusion-induced oxidative stress in diabetic rats," *Biomedicine & Pharmacotherapy*, vol. 100, pp. 455-460, 2018.
- [89] C.-C. Huang, T. J. Lin, Y. F. Lu, C. C. Chen, C. Y. Huang, and W. T. Lin, "Protective effects of L-arginine supplementation against exhaustive exercise-induced oxidative stress in young rat tissues," *The Chinese Journal of Physiology*, vol. 52, no. 5, pp. 306-315, 2009.
- [90] X. Zeng, H. Wang, X. Xing, Q. Wang, and W. Li, "Dexmedetomidine protects against transient global cerebral ischemia/reperfusion induced oxidative stress and inflammation in diabetic rats," *PLoS One*, vol. 11, no. 3, Article ID e0151620, 2016.
- [91] K. Raedschelders, D. M. Ansley, and D. D. Y. Chen, "The cellular and molecular origin of reactive oxygen species generation during myocardial ischemia and reperfusion," *Pharmacology & Therapeutics*, vol. 133, no. 2, pp. 230-255, 2012.
- [92] K. Rehman and M. S. Akash, "Mechanisms of inflammatory responses and development of insulin resistance: how are they interlinked?" *Journal of Biomedical Science*, vol. 23, no. 1, pp. 1-18, 2016.
- [93] J. L. Evans, I. D. Goldfine, B. A. Maddux, and G. M. Grodsky, "Are oxidative Stress-Activated signaling pathways mediators of insulin resistance and β -cell dysfunction?" *Diabetes*, vol. 52, no. 1, pp. 1-8, 2003.
- [94] S. Banihani, S. Swedan, and Z. Alguraan, "Pomegranate and type 2 diabetes," *Nutrition Research*, vol. 33, no. 5, pp. 341-348, 2013.
- [95] S. Liu, J. Yu, M. Fu, X. Wang, and X. Chang, "Regulatory effects of hawthorn polyphenols on hyperglycemic, inflammatory, insulin resistance responses, and alleviation of aortic injury in type 2 diabetic rats," *Food Research International*, vol. 142, no. 13, Article ID 110239, 2021.
- [96] E. Vargas, V. Podder, and M. A. Sepulveda, *Physiology, Glucose Transporter Type 4*, Statpearls, Treasure Island, FL, USA, 2019.

- [97] D. Cai, S. Dhe-Paganon, P. A. Melendez, J. Lee, and S. E. Shoelson, "Two new substrates in insulin signaling, IRS5/DOK4 and IRS6/DOK5," *Journal of Biological Chemistry*, vol. 278, no. 28, pp. 25323–25330, 2003.
- [98] E. Świdarska, J. Strycharz, A. Wróblewski, J. Szemraj, J. Drzewoski, and A. Śliwińska, "Role of PI3K/AKT pathway in insulin-mediated glucose uptake," *Blood Glucose Levels*, vol. 1, pp. 1–18, 2018.
- [99] W. L. Li, H. C. Zheng, J. Bukuru, and N. De Kimpe, "Natural medicines used in the traditional Chinese medical system for therapy of diabetes mellitus," *Journal of Ethnopharmacology*, vol. 92, no. 1, pp. 1–21, 2004.
- [100] S. Georgia and A. Bhushan, "β cell replication is the primary mechanism for maintaining postnatal β cell mass," *Journal of Clinical Investigation*, vol. 114, no. 7, pp. 963–968, 2004.
- [101] Q. Meng, X. Qi, Y. Chao et al., "IRS1/PI3K/AKT pathway signal involved in the regulation of glycolipid metabolic abnormalities by Mulberry (*Morus alba* L.) leaf extracts in 3T3-L1 adipocytes," *Chinese Medicine*, vol. 15, no. 1, pp. 1–11, 2020.
- [102] P. V. M. Ribeiro, A. Silva, A. P. Almeida, H. H. Hermsdorff, and R. C. Alfenas, "Effect of chronic consumption of pistachios (*Pistacia vera* L.) on glucose metabolism in pre-diabetics and type 2 diabetics: a systematic review," *Critical Reviews in Food Science and Nutrition*, vol. 59, no. 7, pp. 1115–1123, 2019.
- [103] A. J. Stull, K. C. Cash, W. D. Johnson, C. M. Champagne, and W. T. Cefalu, "Bioactives in blueberries improve insulin sensitivity in obese, insulin-resistant men and women," *Journal of Nutrition*, vol. 140, no. 10, pp. 1764–1768, 2010.
- [104] C. Bleau, A. D. Karelis, D. H. St-Pierre, and L. Lamontagne, "Crosstalk between intestinal microbiota, adipose tissue and skeletal muscle as an early event in systemic low-grade inflammation and the development of obesity and diabetes," *Diabetes/metabolism research and reviews*, vol. 31, no. 6, pp. 545–561, 2015.
- [105] T. Liu, L. Zhang, D. Joo, and S. C. Sun, "NF-κB signaling in inflammation," *Signal Transduction and Targeted Therapy*, vol. 2, no. 1, pp. 1–9, 2017.
- [106] S. S. Emadi, F. G. Soufi, A. M. Khamaneh, and M. R. Alipour, "MicroRNA-146a expression and its intervention in NF-κB signaling pathway in diabetic rat aorta," *Endocrine Regulations*, vol. 48, no. 2, pp. 103–108, 2014.
- [107] A. Dihingia, D. Ozah, S. Ghosh et al., "Vitamin K1 inversely correlates with glycemia and insulin resistance in patients with type 2 diabetes (T2D) and positively regulates SIRT1/AMPK pathway of glucose metabolism in liver of T2D mice and hepatocytes cultured in high glucose," *The Journal of Nutritional Biochemistry*, vol. 52, pp. 103–114, 2018.
- [108] Y. C. Long and J. R. Zierath, "AMP-activated protein kinase signaling in metabolic regulation," *Journal of Clinical Investigation*, vol. 116, no. 7, pp. 1776–1783, 2006.
- [109] C. Iside, M. Scafuro, A. Nebbioso, and L. Altucci, "SIRT1 activation by natural phytochemicals: an overview," *Frontiers in Pharmacology*, vol. 11, p. 1225, 2020.
- [110] F. Yeung, J. E. Hoberg, C. S. Ramsey et al., "Modulation of NF-κB-dependent transcription and cell survival by the SIRT1 deacetylase," *The EMBO Journal*, vol. 23, no. 12, pp. 2369–2380, 2004.
- [111] S. Coccheri, "Approaches to prevention of cardiovascular complications and events in diabetes mellitus," *Drugs*, vol. 67, no. 7, pp. 997–1026, 2007.
- [112] W. Matthijs Blankesteyn and K. C. M. Hermans, "Wnt signaling in atherosclerosis," *European Journal of Pharmacology*, vol. 763, pp. 122–130, 2015.
- [113] J. Bian, M. Dannappel, C. Wan, and R. Firestein, "Transcriptional regulation of wnt/β-catenin pathway in colorectal cancer," *Cells*, vol. 9, no. 9, pp. 1–29, 2020.
- [114] S. Dabernat, P. Secrest, E. Peuchant, F. Moreau-Gaudry, P. Dubus, and N. Sarvetnick, "Lack of β-catenin in early life induces abnormal glucose homeostasis in mice," *Diabetologia*, vol. 52, no. 8, pp. 1608–1617, 2009.
- [115] Y. Zhou, Z. Zhou, W. Zhang et al., "SIRT1 inhibits adipogenesis and promotes myogenic differentiation in C3H10T1/2 pluripotent cells by regulating Wnt signaling," *Cell & Bioscience*, vol. 5, no. 1, pp. 61–12, 2015.
- [116] C. Qin, T. Xia, G. Li, Y. Zou, Z. Cheng, and Q. Wang, "Hawthorne leaf flavonoids prevent oxidative stress injury of renal tissues in rats with diabetic kidney disease by regulating the p38 MAPK signaling pathway," *International Journal of Clinical and Experimental Pathology*, vol. 12, no. 9, pp. 3440–3446, 2019.
- [117] A. Mather and C. Pollock, "Glucose handling by the kidney," *Kidney International*, vol. 79, pp. S1–S6, 2011.
- [118] C. O. Eleazu, M. Iroaganachi, K. C. Eleazu, and U. N. Essien, "Ameliorative potentials of cocoyam (*Colocasia esculenta* L.) and unripe plantain (*Musa paradisiaca* L.) on the relative tissue weights of streptozotocin-induced diabetic rats," *Journal of Diabetes Research*, vol. 2013, no. 1, 8 pages, Article ID 160964, 2013.
- [119] C. Guo, F. Han, C. Zhang, W. Xiao, and Z. Yang, "Protective effects of oxymatrine on experimental diabetic nephropathy," *Planta Medica*, vol. 80, no. 04, pp. 269–276, 2014.
- [120] D. K. Singh, P. Winocour, and K. Farrington, "Oxidative stress in early diabetic nephropathy: fueling the fire," *Nature Reviews Endocrinology*, vol. 7, no. 3, pp. 176–184, 2011.
- [121] S. Dronavalli, I. Duka, and G. L. Bakris, "The pathogenesis of diabetic nephropathy," *Nature Clinical Practice Endocrinology & Metabolism*, vol. 4, no. 8, pp. 444–452, 2008.
- [122] A. Ahad, H. Ahsan, M. Mujeeb, and W. A. Siddiqui, "Gallic acid ameliorates renal functions by inhibiting the activation of p38 MAPK in experimentally induced type 2 diabetic rats and cultured rat proximal tubular epithelial cells," *Chemico-Biological Interactions*, vol. 240, pp. 292–303, 2015.
- [123] R. Komers, J. N. Lindsley, T. T. Oyama, D. M. Cohen, and S. Anderson, "Renal p38 MAP kinase activity in experimental diabetes," *Laboratory Investigation*, vol. 87, no. 6, pp. 548–558, 2007.
- [124] T.-F. Tzeng, S.-S. Liou, C. Chang, and I.-M. Liu, "The ethanol extract of *Lonicera japonica* (Japanese honeysuckle) attenuates diabetic nephropathy by inhibiting p-38 MAPK activity in streptozotocin-induced diabetic rats," *Planta Medica*, vol. 80, pp. 121–129, 2014.
- [125] N. S. Chaytor, C. Barbosa-Leiker, C. M. Ryan, L. T. Germine, I. B. Hirsch, and R. S. Weinstock, "Clinically significant cognitive impairment in older adults with type 1 diabetes," *Journal of Diabetes and Its Complications*, vol. 33, no. 1, pp. 91–97, 2019.
- [126] I. H. Salas and B. De Strooper, "Diabetes and Alzheimer's disease: a link not as simple as it seems," *Neurochemical Research*, vol. 44, no. 6, pp. 1271–1278, 2019.
- [127] G. Baydas, V. S. Nedzvetskii, P. A. Nerush, S. V. Kirichenko, and T. Yoldas, "Altered expression of NCAM in hippocampus and cortex may underlie memory and learning deficits in rats with streptozotocin-induced diabetes mellitus," *Life Sciences*, vol. 73, no. 15, pp. 1907–1916, 2003.

- [128] Ö. D. Can, Ü. D. Özkay, N. Öztürk, and Y. Öztürk, "Effects of hawthorn seed and pulp extracts on the central nervous system," *Pharmaceutical Biology*, vol. 48, no. 8, pp. 924–931, 2010.
- [129] E. Zarrinkalam, K. Ranjbar, I. Salehi, N. Kheiripour, and A. Komaki, "Resistance training and hawthorn extract ameliorate cognitive deficits in streptozotocin-induced diabetic rats," *Biomedicine & Pharmacotherapy*, vol. 97, pp. 503–510, 2018.
- [130] C. Fan, J. Yan, Y. Qian, X. Wo, and L. Gao, "Regulation of lipoprotein lipase expression by effect of hawthorn flavonoids on peroxisome proliferator response element pathway," *Journal of Pharmacological Sciences*, vol. 100, no. 1, pp. 51–58, 2006.
- [131] E. K. Al-Hallaq, F. U. Afifi, and S. S. Abdalla, "Evaluation of the hypocholesterolemic effect and phytochemical screening of the hydroethanolic extract of *Crataegus aronia* from Jordan," *Natural Product Communications*, vol. 7, no. 1, pp. 35–38, 2012.
- [132] M. Najafian, A. Ebrahim-Habibi, P. Yaghmaei, K. Parivar, and B. Larijani, "Core structure of flavonoids precursor as an antihyperglycemic and antihyperlipidemic agent: an in vivo study in rats," *Acta Biochimica Polonica*, vol. 57, no. 4, pp. 553–560, 2010.