

REVIEW

CD47—SIRP α -targeted therapeutics: status and prospects

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CD47 is a “don’t eat me” signal to phagocytes that is overexpressed on many tumor cells as a potential mechanism for immune surveillance evasion. CD47 and its interaction with signal-regulating protein alpha (SIRP α) on phagocytes is therefore a promising cancer target. Therapeutic antibodies and fusion proteins that block CD47 or SIRP α have been developed and have shown activity in preclinical models of hematologic and solid tumors. Anemia is a common adverse event associated with anti-CD47 treatment, but mitigation strategies—including use of a low ‘priming’ dose—have substantially reduced this risk in clinical studies. While efficacy in single-agent clinical studies is lacking, findings from studies of CD47—SIRP α blockade in combination with agents that increase ‘eat me’ signals or with antitumor antibodies are promising. Magrolimab, an anti-CD47 antibody, is the furthest along in clinical development among agents in this class. Magrolimab combination therapy in phase Ib/II studies has been well tolerated with encouraging response rates in hematologic and solid malignancies. Similar combination therapy studies with other anti-CD47—SIRP α agents are beginning to report. Based on these early clinical successes, many trials have been initiated in hematologic and solid tumors testing combinations of CD47—SIRP α blockade with standard therapies. The results of these studies will help determine the role of this novel approach in clinical practice and are eagerly awaited.

Key words: CD47, SIRP α , magrolimab, macrophage, phagocytosis, innate immunotherapy

INTRODUCTION

Immune surveillance between normal cells, defective cells, and foreign pathogens is regulated by cell-surface receptors, which mediate interactions between immune cells and their targets. These include markers of ‘self’ or “don’t eat me” signals that interact with proteins expressed on the surface of phagocytes to inhibit phagocytosis, such as the tumor cell major histocompatibility complex type 1 component, β_2 -microglobulin, interaction with macrophage leukocyte immunoglobulin (Ig)-like receptor B1 (LILRB1)¹; tumor cell programmed death-ligand 1 (PD-L1) interaction with programmed cell death protein 1 (PD-1) on tumor-associated macrophages²; tumor cell CD24 interaction with sialic-acid-binding Ig-like lectin 10 (Siglec-10) on tumor-associated macrophages³; and CD47, a cell-surface protein that has ubiquitous expression and an array of cellular functions with multiple binding partners.^{4–7} CD47 inhibits phagocytosis through an interaction with signal-regulating protein alpha (SIRP α) on the surface of

phagocytic cells (Figure 1).^{8,9} Interaction with CD47 promotes localization of SIRP α to the phagocytic synapse, which activates Src homology region 2 domain-containing phosphatase-1 (SHP-1) phosphatase, and ultimately inhibits non-muscle myosin IIA accumulation at the cell membrane, preventing engulfment.^{10,11} Blockade of CD47—SIRP α signaling has recently been investigated as a means of activating phagocytic cells, particularly macrophages, for therapeutic purposes.

Basis for targeting CD47 in cancer

Evasion of immune system surveillance is a fundamental step in tumorigenesis.¹² Malignant cells from multiple tumor types express higher levels of CD47 than do normal cells,^{13–18} suggesting that using CD47 overexpression to masquerade as ‘self’ is a common mechanism for cancer cells to escape immune surveillance. Several mechanisms may lead to advantageous overexpression of CD47. CD47 transcription is induced by MYC,¹⁹ a potent oncogene and driver of many malignancies.²⁰ In a hypoxic tumor microenvironment, CD47 is up-regulated by direct binding of hypoxia-inducible factor 1 (HIF-1) to the CD47 promoter.²¹ CD47 transcription is also regulated by tumor-specific enhancers and super enhancers, which can be activated by pro-inflammatory pathways.²² CD47 overexpression may counteract the overexpression of pro-phagocytic ‘eat me’

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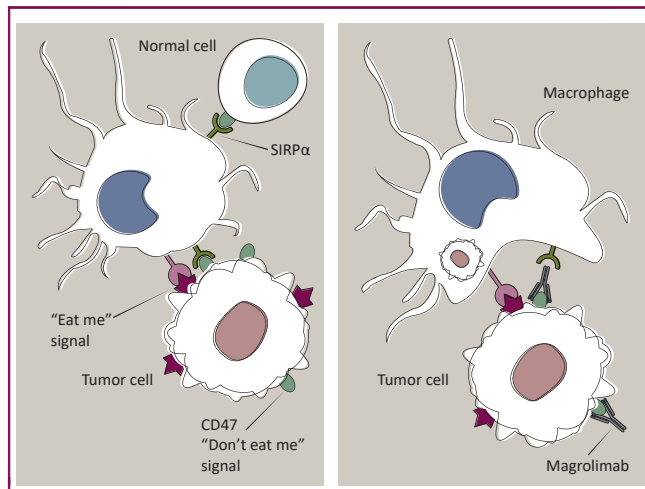


Figure 1. Prevention of phagocytosis by CD47–SIRP α interactions and mechanism of action of CD47–SIRP α therapeutic blockade with magrolimab. CD47 is a “don’t eat me” signal expressed on the cell surface. Interaction of CD47 with SIRP α on phagocytes prevents phagocytic elimination of healthy cells. CD47 is overexpressed on cancer cells to overcome the expression of ‘eat me’ signals and help tumor cells evade macrophage immune surveillance. Blockade of the CD47–SIRP α interaction, as shown with the anti-CD47 antibody (magrolimab) on the right, unmasks the ‘eat me’ signals and promotes phagocytic elimination of tumor cells. Most healthy cells do not express ‘eat me’ signals, and therefore are spared from phagocytosis under CD47–SIRP α blockade. SIRP α , signal-regulating protein alpha. Adapted from Chao et al.¹¹¹

signals that are up-regulated in response to cell stress or because these prophagocytic signals provide a tumorigenic advantage.²³

Therapeutic blockade of CD47–SIRP α interactions: preclinical evidence

Genetic knockdown of CD47 expression renders cells vulnerable to phagocytosis by macrophages *in vitro* and *in vivo*, and indirect reduction of CD47 expression by knockdown of transcription-inducing pathways also increases phagocytosis of tumor cells.^{8,13,21,22,24,25} Monoclonal antibodies to CD47 and SIRP α , and SIRP α fusion proteins, have been developed to block the interaction between tumor cell CD47 and macrophage SIRP α , providing several ways to kill tumor cells, depending on the specific agent used (Figure 2): blocking CD47 or SIRP α removes the “don’t eat me” signal, permitting phagocytosis by macrophages; antibodies may activate Fc-dependent mechanisms, including antibody-dependent cellular cytotoxicity (ADCC), antibody-dependent cellular phagocytosis (ADCP), and complement-dependent cytotoxicity (CDC); antibodies may induce apoptosis directly; and phagocytes may present tumor antigens for CD8+ T-cell activation.²⁶ CDC activation by antibodies to CD47 has not been reported; however, other therapeutic antibodies (e.g. rituximab, daratumumab, and ofatumumab) are known to activate CDC, and this appears to depend on several factors including antibody isotype, binding strength, valence, and epitope; receptor cluster formation; and expression of complement regulatory proteins on tumor cells.^{27–30} Direct induction of apoptosis occurs with some anti-CD47 antibodies but not others, and

with differing potency.^{14,31–35} This process is caspase-independent,³¹ and mechanisms may involve ligation and activation of the CD47 ligand, thrombospondin,³² cytoskeletal reorganization,³¹ and up-regulation of HIF-1 α .³⁴

Blockade of the CD47–SIRP α interaction with anti-CD47 antibodies has anticancer effects in preclinical models.^{14,16–18,35–45} Magrolimab (formerly Hu5F9-G4) is a humanized anti-CD47 antibody that binds CD47 with low nanomolar affinity and is based on an Ig G4 scaffold to minimize Fc-mediated effector toxicity for non-tumor cells expressing CD47.³⁵ Magrolimab binding is sufficient to induce phagocytosis of cancer cells by macrophages *in vitro*, but it does not activate ADCC or CDC, or directly induce apoptosis.³⁵ In immunodeficient [NOD/SCID/IL-2Rgnull (NSG)] models, magrolimab shows strong monotherapy activity against human hematological malignancies³⁵; in ovarian cancer cell lines and patient-derived xenograft models, including taxane-resistant tumors; and in orthotopic xenografts of five aggressive pediatric brain tumor types, without evidence of harm to other central nervous system cells.^{44,46} CC-90002 elicited phagocytosis of hematologic and solid tumor cells *in vitro* and showed significant dose-dependent antitumor effects in multiple myeloma (MM) cell line xenograft models *in vivo*. Significant tumor regression was also observed in cell line- and patient-derived triple-negative breast cancer (TNBC) and acute myeloid leukemia (AML) HL-60 xenograft models.⁴⁷ Lemzoparlimab (TJC4) monotherapy completely eradicated Raji cell tumors in a xenograft model and extended survival in AML xenografted mice.³⁹ SRF231 induced phagocytosis of human hematopoietic tumor cell lines by human macrophages; interaction of SRF231 Fc domain with Fc γ R induced apoptosis and ADCP; and significant antitumor activity with sustained tumor regression was observed across several xenograft models of hematologic malignancies.⁴² AO-176 led to apoptosis selectively in tumor cells, not normal cells including activated T cells, and promoted dose-dependent phagocytosis of several hematologic and solid tumor cell types. Tumor growth inhibition was observed in xenograft lymphoma, TNBC, ovarian and gastric carcinoma models.⁴⁰ In one MM xenograft model, AO-176 induced complete remission lasting up to 120 days in all treated mice.⁴⁸ Interestingly, most anti-SIRP α antibodies and non-Fc CD47-targeting agents have not shown single-agent activity in the preclinical setting,^{49–52} suggesting that engaging Fc receptors contribute a key component of efficacy in preclinical models.

Activity in rational therapeutic combinations

Targeted antibodies to tumor cell markers are a mainstay of cancer treatment and are thought to act through Fc effector mechanisms including ADCC, ADCP, CDC, and induction of apoptosis.^{27,53} Co-treatment with CD47–SIRP α -blocking agents may synergize with tumor-targeting antibodies by enhancing the potential of phagocytes, particularly macrophages, to execute these effector functions (Figure 3). Indeed, CD47–SIRP α blockade combined with

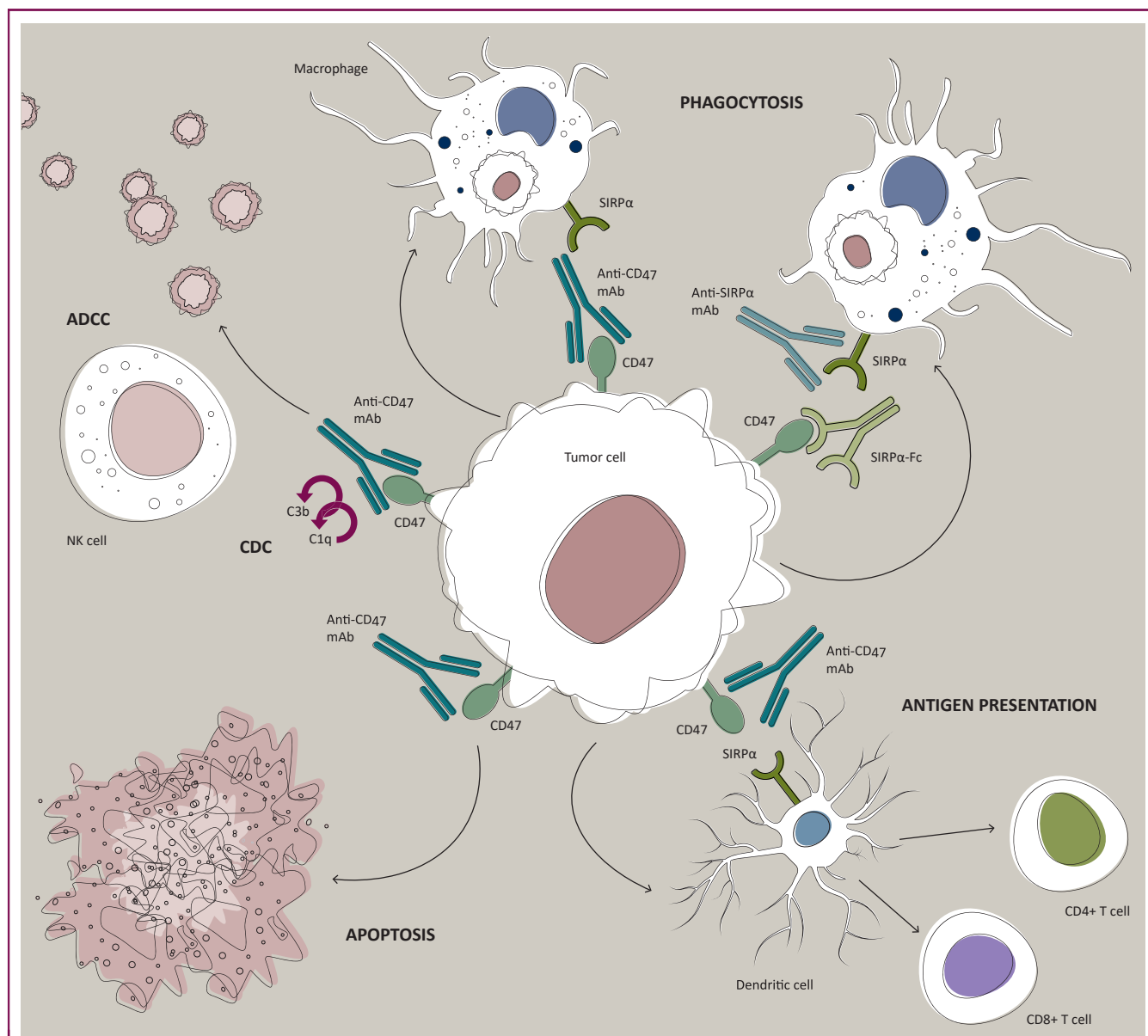


Figure 2. Mechanisms of targeting the CD47–SIRP α pathway in cancer.

Therapeutic targeting of the CD47–SIRP α pathway can cause elimination of cancer cells through multiple mechanisms. Firstly, inhibition of the CD47–SIRP α interaction with a blocking anti-CD47 antibody, a blocking anti-SIRP α antibody, or a recombinant SIRP α protein (depicted here as a bivalent Fc-fusion protein) leads to phagocytic uptake of tumor cells by macrophages. Secondly, an anti-CD47 antibody can eliminate tumor cells through traditional antibody Fc-dependent mechanisms including natural killer cell-mediated ADCC and CDC. Thirdly, anti-CD47 antibody may directly stimulate apoptosis of tumor cells through a caspase-independent mechanism. Fourthly, anti-CD47 antibody may enable phagocytic uptake of tumor cells by dendritic cells and subsequent antigen presentation to CD4 and CD8 T cells, thereby stimulating an antitumor adaptive immune response.

ADCC, antibody-dependent cellular cytotoxicity; CDC, complement-dependent cytotoxicity; mAb, monoclonal antibody; NK, natural killer; SIRP α , signal-regulating protein alpha. Reprinted from Chao et al.,²⁶ with permission from Elsevier.

tumor-targeting antibodies rituximab, cetuximab, panitumumab, trastuzumab, daratumumab, and dinutuximab has had additive or synergistic efficacy in preclinical models, including cancer lines selected to be resistant to ADCC.^{12,35,39,46,54–63} Individual tumor-targeting antibodies have been shown to activate multiple Fc effector functions,^{30,64–66} broadening the therapeutic potential of these combinations.

Calreticulin (CRT), an ‘eat me’ signal, is up-regulated on tumor cell surfaces by cytotoxic stimuli, including anthracyclines and inhibitors of protein phosphatase 1/GADD34

(tautomycin, calyculin A, and salubrinal), suggesting potential synergy between cytotoxic agents and CD47–SIRP α blockade (Figure 3).⁶⁷ Azacitidine has also increased the expression of CRT on the surface of AML cells.^{68,69} Because tumor cells express relatively high levels of CRT as well as CD47, and most normal cells express low levels of CD47 without CRT, combining CD47–SIRP α blockade and cytotoxic agents can selectively target tumor cells.²³ Magrolimab combined with azacitidine increased phagocytosis of HL60 cells by macrophages *in vitro* significantly beyond the level observed with either agent alone.⁶⁸ Growth of HL60

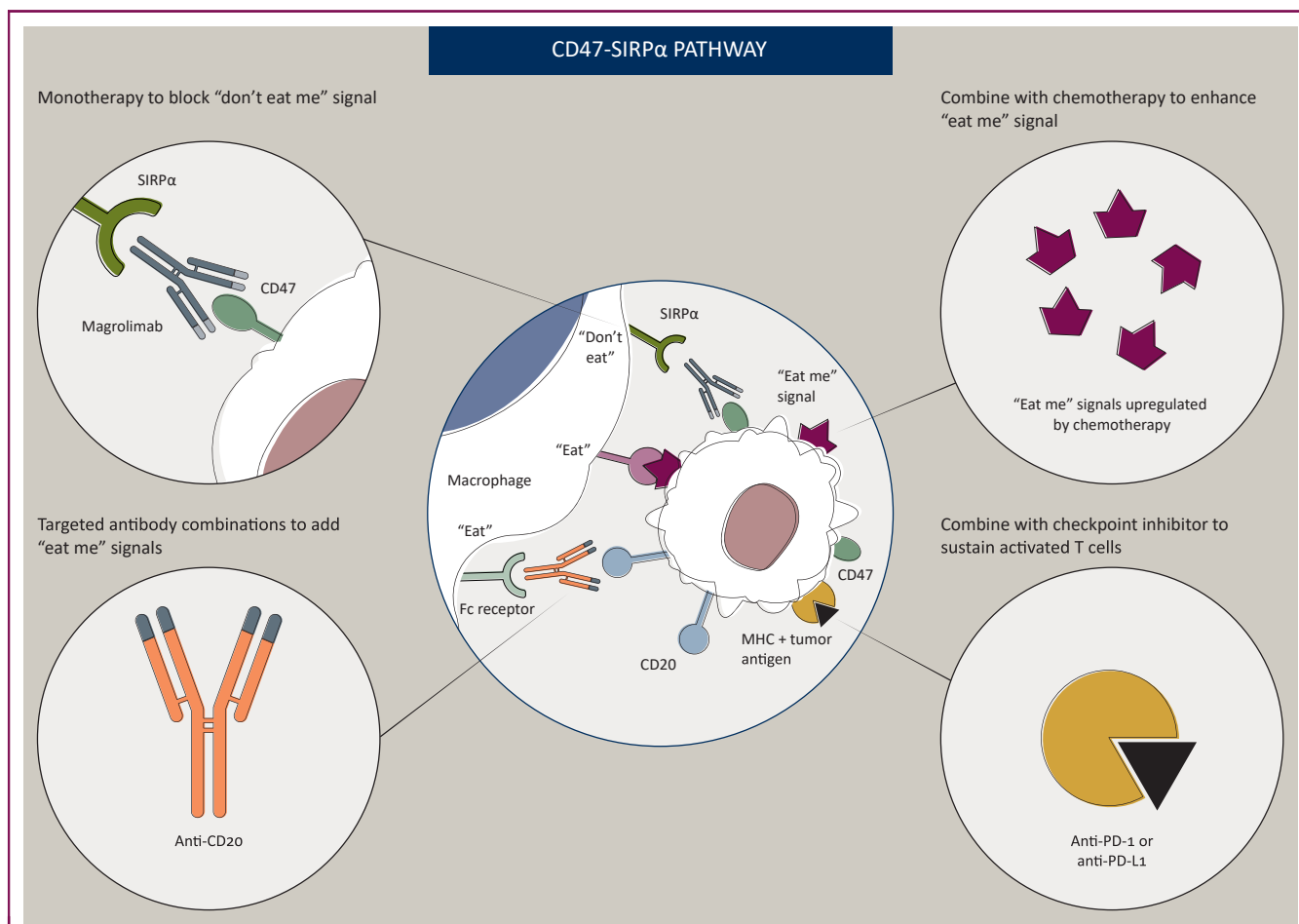


Figure 3. Potential synergistic combinations with anti-CD47 treatment.

CD47–SIRP α pathway blockade in combination with therapies that increase the ‘eat me’ signals on tumor cells has the potential for synergistic clinical efficacy. Some types of chemotherapy and other cytotoxic agents increase the expression of ‘eat me’ signals on tumor cells. Similarly, tumor-targeted antibodies present Fc regions to the Fc receptors on phagocytes, triggering ADCP. Phagocytosis of tumor cells by macrophages or dendritic cells can lead to tumor cell antigen presentation to T cells, activating antitumor T-cell responses; therefore, combination of CD47–SIRP α pathway blockade with T-cell checkpoint inhibitors may also produce synergistic efficacy. ADCP, antibody-dependent cellular phagocytosis; MHC, major histocompatibility complex; PD-1, programmed cell death protein 1; PD-L1, programmed death-ligand 1; SIRP α , signal-regulating protein alpha.

cells engrafted into NSG mice was also inhibited with magrolimab + azacitidine treatment as early as day 10, and growth elimination with 100% survival was maintained up to study termination 255 days post-engraftment.⁶⁸ Another anti-CD47-blocking antibody increased the sensitivity of high CD47-expressing hepatocellular carcinoma (HCC) cells to doxorubicin or cisplatin *in vitro*, and the combination was synergistic in patient-derived HCC xenografts.⁷⁰

Combinations of anti-CD47–SIRP α with T-cell checkpoint inhibitors can independently activate both innate and adaptive immune responses, and, based on the ability of phagocytes to cross-present antigens to T cells, have the potential for a synergistic antitumor immune response (Figure 3).^{71,72} In fact, evidence suggests that the effects of CD47 blockade in some cancer models require cross-priming of T cells.⁷¹ Preclinically, anti-mouse CD47 nanobody treatment (lacking single-agent activity) significantly enhanced response to anti-PD-L1 in mouse melanoma cells induced to express PD-L1; and in a syngeneic mouse

melanoma model, the combination significantly delayed tumor growth and prolonged survival.⁷³ Similar tumor-specific effects of anti-CD47 and anti-PD-L1 antibodies in mouse pancreatic cancer models suggest that the efficacy of a dual anti-CD47-SIRP α /PD-L1 approach will depend on tumor type and microenvironment.^{74,75} In combination with anti-cytotoxic T-lymphocyte associated protein 4 (CTLA4) antibodies, anti-CD47 antibodies significantly suppressed tumor growth, extended survival, and increased CD8+ T-cell proliferation in tumor-draining lymph nodes in a mouse model of pancreatic cancer, suggesting another potential avenue for clinical development.⁷⁶

Collectively, preclinical studies of CD47–SIRP α blockade across hematologic and solid tumor models suggest potential for clinical efficacy, particularly in combination regimens. Hematologic tumors may be especially susceptible, because macrophage clearance of those cells is a natural part of their life cycle. Questions remain, including the role of prophagocytic signals such as CRT expressed on tumor cells versus macrophage-secreted CRT, which binds to

asialoglycans on target cells and CD91 on macrophages and helps mediate phagocytosis.^{77,78} Tumor and other cells also express sialoglycoproteins on their cell surfaces; hyper-sialylation could be another means of immune surveillance escape.⁷⁹ Regulation of tumor cell expression of CRT and sialoglycoproteins may offer additional therapeutic avenues.

Safety of targeting the CD47–SIRP α axis

The ubiquitous expression of CD47 is unique among current immunotherapy targets, sparking concerns regarding on-target toxicity resulting from phagocytosis of normal CD47-expressing cells after treatment with either CD47- or SIRP α -blocking drugs.⁷² In particular, red blood cells (RBCs) express CD47 as a regulator of their lifespan by macrophages and are therefore vulnerable to anti-CD47 antibodies late in their lives.⁸ Other blood cells and many other cell types also express CD47, which broadens the potential range of on-target toxicities. In addition, the Fc region of some anti-CD47 and anti-SIRP α antibodies has the potential to activate ADCC and CDC, which may be toxic to normal cells.^{4,5,28,80} IgG1 and IgG3 are the most potent activators of ADCC and CDC, but IgG4 is not associated with either²⁸; therefore, isotype scaffold may define the toxicity profile of antibody-based therapies.

Evaluation of potential toxicities in mouse models is limited by lack of cross-reactivity between most humanized or fully human antibodies and fusion proteins with mouse proteins on sensitive cell types.^{35,38,40} Nonhuman primates (NHPs) closely recapitulate human physiology and protein sequences.⁸¹ In particular, cynomolgous monkey CD47 differs from human CD47 in only three amino acids, which are outside of the SIRP α -binding domain, so this NHP has been used routinely in pharmacokinetic and toxicology studies involving CD47.^{35,37-40,45} These studies have confirmed that RBCs are quite sensitive to CD47–SIRP α blockade; while others, including white blood cells, are surprisingly resistant against any measurable toxic effects. Other than RBCs, normal cells do not express prophagocytic signals, which likely protects them from phagocytosis under CD47–SIRP α blockade.²³ Multiple *in vitro* studies have confirmed that, in contrast to tumor cells and RBCs, normal cells are not eliminated in the presence of an anti-CD47 antibody.^{14,23}

RBC depletion

Aging RBCs are naturally cleared from circulation by macrophages.⁸² CD47 on RBCs inhibits phagocytic clearance and, conversely, RBCs that lack CD47 are rapidly cleared from the circulation.⁸ As RBCs age, they gradually lose CD47 expression and increase expression of prophagocytic signals, which increases their susceptibility to phagocytosis.^{25,83} On-target dose-dependent anemia, with a parallel increase in reticulocytosis, was observed in NHPs 5-7 days after magrolimab dosing, and hemoglobin spontaneously returned to baseline levels \sim 2 weeks after dosing.³⁵

Despite high CD47 expression, white blood cells and platelets were unaffected, and there was no evidence of intravascular hemolysis. Although anemia occurred in all NHPs who received magrolimab, the degree of anemia varied among individuals at the same dose. Similar findings in NHPs have been reported for other anti-CD47 antibodies.⁸⁴ While not fully understood, this variance may be driven by differences in expression of pro-phagocytic signals by RBCs. Further study of this phenomenon could help to identify clinical biomarkers that may be associated with more severe anemia in patients.

Safety and mitigation strategies for RBC depletion with monotherapy

Recognition of potential on-target anemia with CD47–SIRP α blockade has led to a broad variety of efforts to develop therapeutic strategies that avoid anemia. The antibody Fc scaffold used may also influence the clinical toxicity profile of anti-CD47–SIRP α agents. Data on NHPs are available for some agents; clinical data are preliminary and based on conference presentations for all but magrolimab.

Achieving magrolimab serum levels in NHPs associated with efficacy in xenograft models required doses of 10-30 mg/kg to saturate the ‘antigen sink’ of CD47 on non-tumor cells; therefore, a low priming dose was tested based on the hypothesis that it could serve to remove the most vulnerable RBCs and activate reticulocytosis before initiating a higher maintenance dose. With the use of a priming dose, there was no further decrease in hemoglobin with maintenance doses and hemoglobin levels returned to the normal range.³⁵ This provided rationale for the clinical priming/maintenance dosing strategy to mitigate anemia in magrolimab trials. Clinically, anemia with increased need for RBC transfusion was observed in a dose-escalation trial of magrolimab in relapsed/refractory (R/R) AML.⁸⁵ In contrast, with a magrolimab priming/maintenance dosing schedule, mild transient anemia in \sim 25%-57% of patients with compensatory reticulocytosis was observed in subsequent clinical trials.^{58,69,86-88} In these trials, hemoglobin decreased by a mean of 0.4-0.9 g/dl (maximum 2.5 g/dl) after the first dose, and then returned to baseline; therapy continued to improve hemoglobin levels and reduce the need for RBC transfusion.^{69,87} Evidence from *ex vivo* studies of bone marrow and peripheral blood samples from patients with solid tumors (NCT02216409) or AML (NCT02678338) shows that CD47 protein is ‘pruned’ from RBCs but not white blood cells or AML blasts by magrolimab treatment which, in addition to replacement of RBCs with a younger cell population by the priming dose, leads to lack of further IgG4-driven elimination of RBCs, given the remaining RBCs are CD47 negative and not bound by magrolimab.⁸⁹ These findings further support the use of a priming/maintenance dose to mitigate on-target anemia with magrolimab use. No data have been published regarding this phenomenon for other CD47 agents, but the degree or kinetics of pruning

may differ between them. If so, this may underlie some observed differences in the specific toxicity profiles of different drugs targeting this pathway.

Trillium (TTI)-621 consists of a fusion of the N-terminal V domain of human SIRP α with the IgG1 Fc domain. This 'decoy receptor' was designed to bind to CD47 on tumor cells and activate phagocytosis and Fc effector functions for maximum efficacy. TTI-621 binds to CD47 on a variety of hematologic cells and causes anemia in NHPs but exhibits minimal binding to human RBCs, purportedly because it binds to clustered CD47 in the cell membrane but not distributed CD47 associated with the spectrin cytoskeleton in human RBCs.³⁷ During the dose-escalation period of a phase I study of IgG1-based TTI-621, thrombocytopenia was observed in 30% (22% grade ≥ 3) of 214 patients with R/R non-Hodgkin's lymphoma (NHL).⁹⁰ Results from 25 patients in a dose-finding trial of the related TTI-622 in R/R lymphoma suggest a similar safety profile, with thrombocytopenia in 16% (grade 4 in 1) and neutropenia in 12% (all grade ≥ 3).⁹¹

ALX148 (ALX Oncology, South San Francisco, CA) is a combination of the N-terminal D1 CD47-binding domain of SIRP α engineered to provide picomolar affinity, fused to a modified human IgG1 which is 'Fc dead' to prevent ADCC, CDC, and targeting normal cells for ADCP. ALX148 had no reported effects on hematologic parameters in NHP studies.⁴⁵ A bispecific antibody, TG-1801 (TG Therapeutics, New York, NY; formerly NI-1701 from Novimmune), has also been developed on an IgG1 Fc scaffold; it binds weakly to CD47 and with a higher affinity to CD19, which is widely expressed on malignant B cells.³⁸ This targets antibody binding preferentially to B cells, avoiding RBCs and other CD47+ cells. No clinical monotherapy data have been presented on either compound.

AO-176 (Arch Oncology, Brisbane, CA) is an anti-CD47 antibody with an IgG2 Fc domain that binds selectively to CD47 on tumor cells but not on other cells, and does not activate ADCC but does have a direct cell-killing effect; the mechanisms for selective tumor cell binding and direct killing effect are unknown. AO-176 had minimal effect on hematologic parameters in NHPs, and transient anemia was not seen⁴⁰; however, interim data from a phase I/II trial of AO-176 showed thrombocytopenia in 33% (grade 3 brief thrombocytopenia in 22%) and anemia in 22%.⁹²

Like magrolimab, anti-CD47 antibodies lemozoparlimab, IBI188, SRF231, CC-90002, and AK117 are based on an IgG4 scaffold.^{42,59,93-96} Lemozoparlimab (I-Mab Biopharma, Shanghai, China) binds in such a way that glycosylation near the binding epitopes on RBC CD47 'shields' the RBC from lemozoparlimab binding. Lemozoparlimab had minimal and transient effects on RBCs in NHPs.³⁹ However, despite this characterized mechanism of binding, anemia was observed in 30% of patients treated with lemozoparlimab in a phase I study, with an average transient decrease in hemoglobin of 1.5 g/dl, similar to magrolimab.⁹⁵ Interim phase I data in patients with R/R solid tumors or lymphoma treated with IBI188 (using a priming and maintenance dosing strategy) show anemia in 15% (3/20) of patients and 1 patient having

a grade 4 platelet count decrease.⁹³ SRF231 was associated with fatigue (43%), headache (35%), and pyrexia (30%); anemia was not reported in the abstract.⁹⁴ In the CC-90002 monotherapy trial in patients with R/R AML or high-risk myelodysplastic syndromes (MDS), the expected anemia was observed in 32% of patients; however, thrombocytopenia also occurred in 39% of patients, suggesting an unknown mechanism independent of ADCC and CDC activation.⁹⁷ Among the first 15 patients treated with AK117 in a phase I study, grade 2 anemia and grade 1 thrombocytopenia occurred in 1 patient.⁹⁶ These spectrum of adverse events (AEs) highlight the possibility that even anti-CD47 agents with a similar IgG4 scaffold may have distinct safety profiles.

Antibodies to SIRP α , which is not expressed on RBCs, have also been developed—including ADU-1805 (Aduro Biotech, Berkeley, CA) based on IgG2 and BI 765063 (OSE-172; Boehringer Ingelheim, Ingelheim am Rhein, Germany) based on IgG4—to antagonize this pathway while avoiding the RBC binding of anti-CD47 drugs.^{50,98} The most common AEs in patients with advanced solid tumors who completed dose escalation with BI 765063 ($n = 50$) were infusion-related reactions, fatigue, headache, arthralgia, and diarrhea; as expected, anemia was not observed.⁹⁹

These monotherapy studies provide more questions than answers regarding the role of IgG isotype scaffold and specific mechanisms of target engagement in the safety profiles of CD47—SIRP α -blocking therapies. Importantly, AEs associated with T-cell immune checkpoint inhibitors (colitis, pneumonitis, and hypothyroidism) have not been observed with these macrophage checkpoint inhibitors.¹⁰⁰

Clinical safety in combination therapy studies

Combinations of CD47—SIRP α -blocking therapies or tumor-opsionizing antibodies with other treatments that increase expression of 'eat me' signals have shown promising results in early-stage clinical trials and were well tolerated. A phase Ib/II trial of magrolimab + rituximab in 115 patients with R/R B-cell lymphoma showed the expected mild, transient anemia following the priming dose that resolved on maintenance dose therapy. Most AEs occurred in the first month, only 7% of patients discontinued for AEs, and no late-emerging safety signals were seen up to 24 months.^{58,101} In a phase Ib trial of magrolimab + azacitidine in patients with untreated AML ineligible for intensive chemotherapy or untreated higher-risk MDS, the combination was well tolerated, with no significant immune-related AEs or increases in infections or cytopenia observed, 0% (MDS) and 4.7% (AML) of patients discontinuing for treatment-related AEs, and improvement in cytopenias on therapy.^{69,87} Long-term treatment with magrolimab (up to 25 months) in patients with AML has not shown any late-emerging toxicities.⁶⁹

A small phase Ib study of magrolimab with the PD-L1 inhibitor avelumab in patients with platinum-R/R ovarian cancer or advanced solid tumors ($n = 34$) showed a safety profile consistent with PD-L1 therapy and

magrolimab-related anemia, and discontinuation of either drug because of AEs in 15% of patients.⁸⁸

Clinical efficacy of CD47–SIRP α blockade

Clinical efficacy in monotherapy studies. Monotherapy phase I clinical trials of anti-CD47 antibodies have generally yielded limited signs of efficacy compared to combination strategies. Magrolimab monotherapy was evaluated in 62 heavily pretreated patients with solid tumors or lymphoma, with objective partial responses (PRs) observed in 2 patients with ovarian cancer and a mixed response in 1 patient with diffuse large B-cell lymphoma (DLBCL).⁸⁶ CC-90002 monotherapy in AML and high-risk MDS was terminated during the dose-escalation stage because of an insufficiently promising clinical profile; best response was stable disease (SD) in two patients, and anti-drug antibodies were observed in most patients regardless of dose.^{97,102} Interim data from dose escalation of TTI-621 in R/R NHL ($n = 214$) showed objective responses in 20% of 71 patients.⁹⁰ Similarly, interim results of intra-lesional administration of TTI-621 in cutaneous T-cell lymphoma showed Composite Assessment of Index Lesion Severity response (decrease of $\geq 50\%$) in 34% of 29 patients.¹⁰³ Interim results on SRF231 in 37 patients with R/R solid tumors indicated no complete response (CR) or PR, although prolonged SD was reported.⁹⁴ AO-176 monotherapy produced 1 PR and 7 SD responses in an interim analysis of 27 patients with advanced solid tumors expressing high levels of CD47.⁹² Finally, monotherapy with BI 765063, which binds to SIRP α , led to clinical benefit in 45% of patients with advanced solid tumors, including one PR, during dose escalation.⁹⁹ These studies highlight the limitations of preclinical cancer models for predicting efficacy in human malignancies.

Clinical efficacy in combination therapy studies. Rational combinations of CD47–SIRP α blockade with treatments that increase the presence of phagocytic markers have produced encouraging preliminary data, and many additional studies are underway (Table 1). Magrolimab + rituximab has shown benefit in R/R NHL to at least two prior lines of therapy in a phase Ib study [$n = 22$, 15 with DLBCL, and 7 with follicular lymphoma (FL)]; overall response rates (ORRs) and CR rates, respectively, in DLBCL were 40% and 33%, and in FL were 71% and 43%.⁵⁸ More mature data from the phase Ib/II expansion ($n = 115$, 70 with DLBCL and 45 with indolent lymphoma), including 85% who were rituximab-refractory, show ORR and CR rate, respectively, of 36% and 15% in patients with DLBCL, and 61% and 24% in patients with indolent lymphoma, with a median time to response of 1.8 months and duration of response (DOR) not reached. Similar responses were observed across multiple DLBCL subtypes and primary refractory patients, irrespective of prior lines of therapy.¹⁰¹ Early results from clinical studies in NHL with other anti-CD47–SIRP α agents also suggest efficacy, although patient numbers were small and populations differed. CC-90002 + rituximab in 24 patients with R/R NHL showed an

ORR of 13% with 1 durable CR and 2 patients with PR. DOR was 3.9 months.⁵⁹ In the subset of phase I study, patients with B-cell NHL who received TTI-621 + rituximab ($n = 35$), ORR was 23% (9% CR, 14% PR).⁶¹ Preliminary analysis of data on ALX148 + rituximab yielded a 40.9% ORR (4 CR, 5 PR, 6 SD) in 22 patients on 10 mg/kg, and 63.6% ORR (3 CR, 4 PR, 1 SD, $n = 11$ total) in 11 patients on 15 mg/kg.⁶⁰

Data from phase Ib clinical trials of magrolimab + azacitidine in patients with untreated higher-risk MDS and untreated AML who were ineligible for intensive chemotherapy have been reported. In 33 assessable patients with MDS (of 39 total, 64% high or very high risk); ORR was 91% and CR was 42%, which increased to 56% after 6 months on therapy. In four patients with *TP53* mutations, ORR was 75% and CR was 50%. Median time to response was 1.9 months and median DOR was not reached. Twenty percent of responders were minimal residual disease (MRD) negative after a median of 5.8 months of follow-up.⁸⁷ The AML trial ($n = 64$) included 70% with poor cytogenetic risk and 73% with *TP53* mutations. In patients assessable for efficacy ($n = 43$), ORR was 63% and CR was 42% (69% and 45%, respectively, in patients with *TP53* mutations). MRD negativity was achieved in 35% and 29% of the overall and *TP53*-mutant populations. Median time to response was 1.95 months, median DOR 9.6 months, and median overall survival (OS) 18.9 months in patients with *TP53*-wild-type and 12.9 months in patients with *TP53*-mutant AML.⁶⁹ Several other anti-CD47 agents have also begun clinical testing in combination with azacitidine, but no results have been published.

Early clinical data are also emerging on combinations of anti-CD47–SIRP α agents with targeted antibodies and T-cell checkpoint inhibitors in solid tumors. Magrolimab was combined with cetuximab in patients with advanced colorectal cancer in a phase Ib/II study, with PR achieved in 6% and SD in 44% of patients with *KRAS*-wild-type disease, and SD in 38% of those with *KRAS*-mutant disease. The investigators noted low tumor epidermal growth factor receptor expression at baseline, which may have limited synergy with CD47 blockade.¹⁰⁴ Similarly, a phase Ib study of magrolimab with avelumab in ovarian cancer ($n = 24$ assessable for efficacy) found SD in 42%; the only patient with documented tumor cell PD-L1 expression had an unconfirmed PR.⁸⁸ Investigators from both studies suggested that alternative strategies to enhance phagocytic signals are needed in the respective tumor types. In a phase I study of ALX148 with pembrolizumab [with or without 5-fluorouracil (5-FU) + platinum] in head and neck squamous cell carcinoma (HNSCC) or trastuzumab (with or without ramucirumab + paclitaxel) in human epidermal growth factor receptor 2-positive (HER2+) gastric/gastroesophageal cancer (GC), checkpoint inhibitor-naïve patients with HNSCC had an ORR of 40% with a median progression-free survival (PFS) of 4.6 months and OS not reached ($n = 10$), and those with GC had an ORR of 20% with a median PFS of 2.2 months and median OS of 8.1 months.¹⁰⁵

Table 1. Ongoing and recruiting trials of anti-CD47 and anti-SIRP α agents (by estimated study completion date, grouped by agent)

Agent	Company	Regimen	Population	Estimated enrollment, <i>n</i>	Estimated study completion	NCT identifier	Phase	Status
AO-176 (anti-CD47 IgG2 mAb)	Arch Oncology	AO-176 OR AO-176 + paclitaxel	Advanced solid tumors	183	March 2023	NCT03834948	I/II	Recruiting
		OR AO-176 + pembrolizumab AO-176 OR AO-176 + either dexamethasone OR dexamethasone + bortezomib	R/R MM	102	March 2023	NCT04445701	I/II	Recruiting
HX009 (anti-CD47/PD-1 bifunctional antibody)	Waterstone Hanxbio Pty Ltd	HX009 monotherapy	R/R advanced malignant tumors	21	September 2021	NCT04097769	I	Active, not recruiting
		HX009 monotherapy	Unresectable locally advanced/metastatic solid tumors	210	February 2023	NCT04886271	II	Recruiting
TTI-621 (SIRPaFc)	Trillium Therapeutics Inc.	TTI-621 OR TTI-621 + either rituximab OR nivolumab	R/R hematological malignancies and selected solid tumors (PTCL, CTCL)	260	December 2021	NCT02663518	I	Recruiting
TTI-622 (SIRPaFc)		TTI-622 OR TTI-622 + either azacitidine OR azacitidine + venetoclax OR carfilzomib + dexamethasone	R/R lymphoma or MM	150	December 2022	NCT03530683	I	Recruiting
IBI188 (anti-CD47 mAb)	Innovent Biologics (Suzhou) Co. Ltd.	IBI188 OR IBI188 + rituximab	Solid tumors and lymphomas	49	January 2022	NCT03717103	I	Active, not recruiting
		IBI188 + azacitidine	Newly diagnosed high-risk MDS	12	February 2022	NCT04485065	I	Recruiting
		IBI188 + azacitidine	AML	126	May 2022	NCT04485052	I/II	Recruiting
		IBI188 monotherapy	Advanced malignant tumors and lymphomas	42	August 2022	NCT03763149	I	Active, not recruiting
BI-765063/OSE172 (anti-SIRP α Mab)	Boehringer Ingelheim	IBI188 + GM-CSF + cisplatin/ carboplatin + bevacizumab + sintilimab + pemetrexed	Advanced malignancies	120	October 2022	NCT04861948	I	Recruiting
		BI-765063 OR BI-765063 + BI-754091 (a PD-1 receptor antagonist)	Japanese adults w/ advanced solid tumors	36	June 2022	NCT04653142	I	Recruiting
SL-172154	Shattuck Labs, Inc.	BI-765063 OR BI-765063 + BI-754091 (a PD-1 receptor antagonist)	Advanced solid tumors (NSCLC, TNBC, pancreatic cancer, melanoma, HNSCC, RCC, UC, SCL, gastric cancer, CRC, and OC)	116	December 2022	NCT03990233	I	Recruiting
		SL-172154 monotherapy	Unresectable, locally advanced/metastatic ovarian, primary peritoneal, or fallopian tube cancer	40	July 2022	NCT04406623	I	Recruiting
ALX148 (CD47 antagonist)	ALX Oncology, Inc.	SL-172154 monotherapy	Cutaneous SCC or HNSCC	18	July 2022	NCT04502888	I	Recruiting
		ALX148 OR ALX148 + either pembrolizumab OR trastuzumab OR rituximab OR pembrolizumab + 5-FU + cisplatin OR trastuzumab + ramucirumab + paclitaxel	Advanced/metastatic solid tumor malignancy; or R/R NHL	174	December 2022	NCT03013218	I	Active, not recruiting
		ALX148 + azacitidine	Previously untreated or R/R higher-risk MDS	173	December 2023	NCT04417517	I/II	Recruiting
		ALX148 + venetoclax and azacitidine	Newly diagnosed or R/R AML	97	December 2023	NCT04755244	I/II	Recruiting
		ALX148 + pembrolizumab + cisplatin/carboplatin + 5-FU	Advanced HNSCC	112	October 2024	NCT04675333	II	Recruiting
ALX148 + pembrolizumab	Advanced HNSCC	111	October 2024	NCT04675294	II	Recruiting		

Continued

Table 1. Continued

Agent	Company	Regimen	Population	Estimated enrollment, <i>n</i>	Estimated study completion	NCT identifier	Phase	Status
IBI322 (recombinant anti-human CD47/PD-L1 bispecific antibody)	Innovent Biologics (Suzhou) Co. Ltd.	IBI322 monotherapy	Advanced malignant tumors	45	December 2022	NCT04338659	I	Not yet recruiting
		IBI322 monotherapy	Hematologic malignancy that failed standard treatment	230	November 2023	NCT04795128	I	Recruiting
		IBI322 monotherapy	Locally advanced, unresectable or metastatic tumors	218	December 2023	NCT04328831	Ia/Ib	Recruiting
IMC-002 (IgG4 CD47 mAb)	ImuneOncia Therapeutics Inc.	IMC-002 monotherapy	Metastatic or locally advanced solid tumors and R/R lymphomas	24	December 2022	NCT04306224	I	Recruiting
TQB2928 (blocks CD47 and SIRPa)	Chia Tai Tianqing Pharmaceutical Group Co., Ltd.	TQB2928 monotherapy	Locally advanced unresectable/metastatic solid tumors, hematological malignancies, or lymphoma	20	December 2022	NCT04854681	I	Not yet recruiting
TG-1801 (NI-1701) (CD47 and CD19 antibody)	TG Therapeutics, Inc.	TG-1801 OR TG-1801 + ublituximab	B-cell lymphoma	16	December 2022	NCT03804996	I	Recruiting
		TG-1801 OR TG-1801 + ublituximab	B-cell lymphoma or CLL	60	December 2023	NCT04806035	Ib	Recruiting
AK117 (anti-CD47 mAb)	Akeso	AK117 monotherapy	R/R advanced solid tumor, NHL (including R/R transformed lymphoma)	162	January 2023	NCT04728334	I	Recruiting
		AK117 monotherapy	NHL	159	September 2023	NCT04349969	I	Not yet recruiting
GS-189 (FSI-189) (anti-SIRPα mAb)	Gilead Sciences/Forty Seven Inc.	FSI-189 OR FSI-189 + rituximab	R/R NHL	75	August 2023	NCT04502706	I	Recruiting
Magrolimab (anti-CD47 mAb) Magrolimab (anti-CD47 mAb)	Gilead Sciences Gilead Sciences	Magrolimab + daratumumab + pomalidomide + dexamethasone + bortezomib	R/R MM	153	September 2023	NCT04892446	II	Not yet recruiting
		Magrolimab + azacitidine + venetoclax OR magrolimab + mitoxantrone + etoposide + cytarabine OR magrolimab + CC-486	Myeloid malignancies	164	March 2024	NCT04778410	II	Not yet recruiting
	Stanford University, Merck Sharp & Dohme Corp.	Magrolimab OR magrolimab + azacitidine	R/R AML, MDS (monotherapy); untreated or R/R AML, MDS (with azacitidine)	287	February 2025	NCT03248479	I	Recruiting
		Magrolimab + azacitidine OR venetoclax + azacitidine OR 7+3	TP53-mutant AML	346	November 2024	NCT04778397	III	Recruiting
		Magrolimab + pembrolizumab + 5-FU + platinum OR magrolimab + docetaxel	HNSCC	233	December 2024	NCT04854499	II	Recruiting
		Magrolimab + azacitidine	Treatment-naive higher-risk MDS	520	February 2025	NCT04313881	III	Recruiting
		Magrolimab + docetaxel	Solid tumors (mNSCLC, mSCLC)	116	March 2025	NCT04827576	II	Recruiting
		Magrolimab + rituximab OR rituximab + gemcitabine + oxaliplatin	R/R B-cell NHL	422	August 2026	NCT02953509	I/II	Recruiting
Magrolimab + pembrolizumab	Hodgkin's lymphoma, or R/R Hodgkin's lymphoma	24	May 2026	NCT04788043	II	Not yet recruiting		

Continued

Table 1. Continued

Agent	Company	Regimen	Population	Estimated enrollment, n	Estimated study completion	NCT identifier	Phase	Status
ZL-1201 (anti-CD47 mAb)	Zai Lab (Shanghai) Co., Ltd.	ZL-1201 monotherapy	Locally advanced unresectable or metastatic solid tumors and lymphomas	66	January 2024	NCT04257617	I	Recruiting
PF-07257876 (CD47/PD-L1 bispecific antibody)	Pfizer	PF-07257876 monotherapy	NSCLC, HNSCC, ovarian cancer	90	July 2024	NCT04881045	I	Not yet recruiting
CPO107 (JMT601) (CD20-CD47 bispecific fusion protein)	Conjupro Biotherapeutics, Inc.	CPO107 monotherapy	CD20-positive NHL	75	December 2024	NCT04853329	I/II	Not yet recruiting
CC-95251 (anti-SIRP α mAb)	Celgene	CC-95251 OR CC-95251 + rituximab OR cetuximab	Advanced solid and hematologic cancers/ neoplasms	230	November 2024	NCT03783403	I	Recruiting
TJ4 (TJ01133, lempoparlimab)	AbbVie	TJ01133 + dexamethasone, carfilzomib, pomalidomide, daratumumab	R/R MM	163	July 2025	NCT04895410	I	Not yet recruiting

5-FU, 5-fluorouracil; AML, acute myeloid leukemia; CLL, chronic lymphocytic leukemia; CRC, colorectal cancer; CTCL, cutaneous T-cell lymphoma; GM-CSF, granulocyte-macrophage colony-stimulating factor; HNSCC, head and neck squamous cell carcinoma; Ig, immunoglobulin; mAb, monoclonal antibody; MDS, myelodysplastic syndrome; MM, multiple myeloma; mNSCLC, metastatic non-small-cell lung cancer; mSCLC, metastatic small-cell lung cancer; NHL, non-Hodgkin's lymphoma; NSCLC, non-small-cell lung cancer; OC, ovarian cancer; PD-1, programmed cell death protein 1; PD-L1, PD-1 ligand; PTCL, peripheral T-cell lymphoma; R/R, relapsed/refractory; RCC, renal cell carcinoma; SCC, squamous cell carcinoma; SCL, small-cell lung cancer; SIRP α , signal-regulating protein alpha; TNBC, triple-negative breast cancer; TTI, trillium; UC, urothelial carcinoma.
Source: [ClinicalTrials.gov](https://clinicaltrials.gov) (updated as of 30 August 2021).

Near-term opportunities for CD47–SIRP α therapeutics

Many clinical trials of anti-CD47–SIRP α agents have been initiated, primarily in combinations with standard therapy (Table 1), including magrolimab, which is currently being tested in multiple trials, some with registrational potential.^{58,69,86,87} While data from trials in solid tumors are eagerly awaited, CD47–SIRP α -targeted therapy has the potential to combine with novel treatment to make a near-term impact in hematological malignancies. For example, in AML, phase III trials of venetoclax combined with azacitidine or low-dose cytarabine in previously untreated AML ineligible for intensive chemotherapy were recently published.^{106,107} Venetoclax is a B Cell Lymphoma 2 (BCL2) inhibitor that induces apoptosis,¹⁰⁸ which increases and redistributes ‘eat me’ signals leading to phagocytic clearance.¹⁰⁹ In experimental studies, the loss of BCL2 in neutrophils signals their disappearance from the blood and tissues, and in mice with enforced expression of neutrophil BCL2 they do not undergo apoptosis; yet at the time normal neutrophils would undergo apoptosis, the BCL2 neutrophils bind CRT and are removed by macrophages.¹¹⁰ CD47–SIRP α blockade thus has potential to synergize with venetoclax. AML clinical trials of IBI188 with azacitidine and ALX148 with venetoclax and azacitidine have been initiated, as have several trials of magrolimab: with venetoclax and azacitidine in newly diagnosed unfit AML; with mitoxantrone, etoposide, and cytarabine in R/R AML; with oral azacitidine as maintenance therapy for patients in complete remission; and a phase III trial of magrolimab + venetoclax versus venetoclax + azacitidine versus 7 + 3 in TP53-mutant AML (Table 1). For high-risk MDS patients, azacitidine and decitabine are the only approved single-agent therapeutics; as responses are generally limited, combination therapies are under investigation. Magrolimab, IBI188, and ALX148 are being combined with azacitidine in treatment-naïve, higher-risk MDS; the ALX148 study will also enroll R/R MDS (Table 1).

CONCLUSIONS

Evidence supporting the CD47–SIRP α interaction as a therapeutic target in cancer is accumulating rapidly, and clinical trials have so far supported key aspects of the preclinical findings. Clinical data, particularly from the hematologic malignancy trials of magrolimab, indicate that CD47–SIRP α blockade in combination with standard treatment is highly efficacious and well tolerated, representing a meaningful advance in patient care. Future studies will determine the ultimate role of anti-CD47 therapy in many cancer indications.

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DISCLOSURE

RM reports employment by Gilead Sciences, Inc., at the time of the study and holds the US20160340397A1 patent and has the US20210115140A1 and US20210147568A1 patents pending. JX reports employment by and stock in Gilead Sciences, Inc. at the time of the study. ILW reports

receiving a percentage of the royalties paid to his institution by Gilead Sciences, Inc.

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