



REVIEW

Epigenetic mechanisms regulate sex-specific bias in disease manifestations

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Received: 30 April 2022 / Revised: 2 June 2022 / Accepted: 20 June 2022 / Published online: 29 June 2022
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Abstract

Sex presents a vital determinant of a person's physiology, anatomy, and development. Recent clinical studies indicate that sex is also involved in the differential manifestation of various diseases, affecting both clinical outcome as well as response to therapy. Genetic and epigenetic changes are implicated in sex bias and regulate disease onset, including the inactivation of the X chromosome as well as sex chromosome aneuploidy. The differential expression of X-linked genes, along with the presence of sex-specific hormones, exhibits a significant impact on immune system function. Several studies have revealed differences between the two sexes in response to infections, including respiratory diseases and COVID-19 infection, autoimmune disorders, liver fibrosis, neuropsychiatric diseases, and cancer susceptibility, which can be explained by sex-biased immune responses. In the present review, we explore the input of genetic and epigenetic interplay in the sex bias underlying disease manifestation and discuss their effects along with sex hormones on disease development and progression, aiming to reveal potential new therapeutic targets.

Key messages

- Sex is involved in the differential manifestation of various diseases.
- Epigenetic modifications influence X-linked gene expression, affecting immune response to infections, including COVID-19.
- Epigenetic mechanisms are responsible for the sex bias observed in several respiratory and autoimmune disorders, liver fibrosis, neuropsychiatric diseases, and cancer.

Keywords X-inactivation · Epigenetics · Sex bias · Hormones · COVID-19 · X-linked genes · Autoimmune disorders · Liver fibrosis · Neuropsychiatric diseases · Cancer

Abbreviations

ACE2 Angiotensin converting enzyme 2
AD Alzheimer's disease
ASD Autism spectrum disorder
ATRX ATRX chromatin remodeler
CD40L Cluster of differentiation 40 ligand
CXCL1 C-X-C motif chemokine ligand 1
CXCL2 C-X-C motif chemokine ligand 2
COPD Chronic obstructive pulmonary disease

CNKSR2 Connector enhancer of kinase suppressor of Ras 2
Cyp Cytochrome P family
DDX3X DEAD-box helicase 3 X-linked
Ephx2 Epoxide hydrolase 2
ER Estrogen receptor
ERE Estrogen response element
FOXP3 Forkhead box P3
H3K27me3 Histone 3 lysine 27 trimethylation
HCC Hepatocellular carcinoma
H19 H19 imprinted maternally expressed transcript
IGF2 Insulin like growth factor 2
IL-17A Interleukin 17 alpha
IFRD1 Interferon related developmental regulator 1

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KRT77	Keratin 77
KDM5C	Lysine demethylase 5C
lncRNAs	Long non-coding RNAs
LRTIs	Lower respiratory tract infections
MAGEC3	MAGE family member C3
MDD	Major depressive disorder
miR	microRNA
mKDM6A	Mutated lysine demethylase 6 alpha
NLGN4	Neurologin 4 X-linked
NOS-3	Nitric oxide synthase-3
PRC2	Polycomb repressive complex 2
PAH	Pulmonary arterial hypertension
PCDH11XY	Protocadherin 11 X/Y-linked
SIRPD	Signal regulatory protein delta
Sult	Sulfotransferase
XCL	X-C motif chemokine ligand 1

Introduction

Gene expression is heavily influenced by epigenetic DNA changes, as well as post-translational modifications (PTMs) of histones which affect chromatin accessibility and structure [1–7]. DNA exists in a condensed form in the nucleus, wrapped around the nucleosomes, which are the functional units of chromatin, consisting of histone octamers (a pair of each H3, H4, H2A, H2B). Histone PTMs are mainly located at the N-terminal part of H3 and H4 and include methylation, acetylation, phosphorylation, ubiquitination, and sumoylation. The entirety of histone PTMs at different sites at each specific time point comprise the “histone code” and play a crucial role in the fate of gene expression by altering chromatin dynamics, including accessibility and transcription factor binding [8–12]. In this way, gene expression can be altered without any changes in the underlying nucleotide sequence, thus creating a link between genotype and phenotype. During development, a cross talk between epigenetic mechanisms, PTMs and hormones, such as estrogens and androgens acting upstream, have been shown to enhance a cascade of events that regulate mechanisms of gene expression inside the nucleus [13–16].

During embryogenesis in mammals, a single female X chromosome undergoes silencing, forming the Barr body. This developmental imprinting event ensures the proper dosage compensation of expression in the X-linked genes between males and females. Recent studies have revealed several genes which can evade X silencing process and are important for sex determination, immune response as well as developmental growth [17–19].

The choice of the silenced X chromosome creates a mosaic pattern, called lyonization in female cells. Although the inactivation of the X chromosome is random in humans, once the X chromosome gets inactivated, it remains inactive

throughout the cell’s lifetime. On the contrary, the X inactivation process in marsupials and in mice is taking place exclusively at the paternally derived X chromosome [17–21].

Epigenetic mechanisms are highly implicated in gene regulation during the X-chromosome inactivation. In order to achieve proper dosage of X-chromosome–encoded proteins, a well-controlled fine-tuning of gene expression in females has been developed [19, 22]. During female mammal development, X chromosome inactivation (XCI) occurs through the action of Xist (X-inactive specific transcript), a long non-coding RNA (lncRNA) that is transcribed from the X chromosome. Each X chromosome contains an X inactivation center which contains both the Xist locus and its antisense transcription unit, Tsix [23]. Once XCI starts, Xist covers almost the entirety of one of the two X chromosomes and induces its silencing through Xist-mediated recruitment of chromatin-modifying, transcriptional-silencing, as well as other RNA-binding proteins [24]. In turn, this leads to epigenetic and structural modifications of the X chromosome, giving rise to one condensed Barr body covered by the Xist RNA [23]. In more detail, Xist promotes the aggregation of supramolecular complexes (SMACs) which include many copies of the Msx2-interacting protein (SPEN), critical for transcriptional repression. SMACs also favor the epigenetic regulators, polycomb group proteins (PcG) deposition inducing chromatin compaction. In this way, increased SMACs levels around genes propagate gene silencing along the X chromosome [24].

The polycomb repressive complex 2 (PRC2), one of the two PcG classes that is recruited by Xist, deposits H3K27me3 histone marks leading to further downregulation of target gene expression. Male cells contain one less chromosome X compared to females. This process aims to nullify the increased expression of X chromosome genes in females compared to males, in the absence of XCI. In combination, differential DNA methylation at CpG islands of the X-linked genes mediates the fine tuning of their expression [17–19]. This imprinting event in females is established through epigenetic mechanisms including DNA and histone methylation (especially H3K27 trimethylation, H3K27me3), mediated by the action of PRC2. It has been shown that the PRC2 subunit binds to a non-coding RNA (ncRNA) within the Xist RNA and is thereby targeted to the female X chromosome, mediating its inactivation [2, 20, 25–28].

Subsequently, the PRC2 complex plays an important role in establishing an inactive status of many genes on the X inactive chromosomes during development. The PRC2 group consists of four core components—the enhancer of zeste (EZH1/2 in mammals, E(z) in *Drosophila*); extra sex-combs (Eed in mammals, Esc in *Drosophila*); suppressor of zeste 12 (Suz12 in mammals, Su(z)12 in *Drosophila*); and a nucleosome remodeling factor (Rbbp7/4 in mammals, Nurf55 in *Drosophila*). The methyltransferases EZH1 and

2 are responsible for establishing and maintaining the inactive histone mark H3K27me3 on the female X chromosome [20, 25–28].

It is therefore evident that a better understanding of the epigenetic mechanisms that underlie the dosage compensation in different sexes may explain the differential response in disease progression such as chronic inflammation, autoimmune diseases, and liver fibrosis (Fig. 1).

Differential manifestation of infectious diseases in male and female patients

It has long been observed that males and females harbor differences in their immune responses against most bacterial, viral, and parasitic infections, such as influenza, hepatitis, AIDS, tuberculosis, and malaria. The underlying pathways, which promote these sex-based immunologic differences in response to infectious diseases, have only recently been investigated. Among them, the influence of sex hormones on pathways regulating the immune system, along with the effect of X-chromosome inactivation on X-related immune genes, appears to play a central role.

Susceptibility differences to lower respiratory tract infections between the two sexes have been reported and extensively studied in different mice models. Intranasal inoculation of mice with *Streptococcus pneumoniae*, the most commonly isolated cause of bacterial pneumonia, demonstrated a higher susceptibility and decreased survival in males compared to females [29]. This was attributed to the differential immune response between the two sexes. It has been suggested that increased mortality from infectious diseases in males was related to testosterone-induced

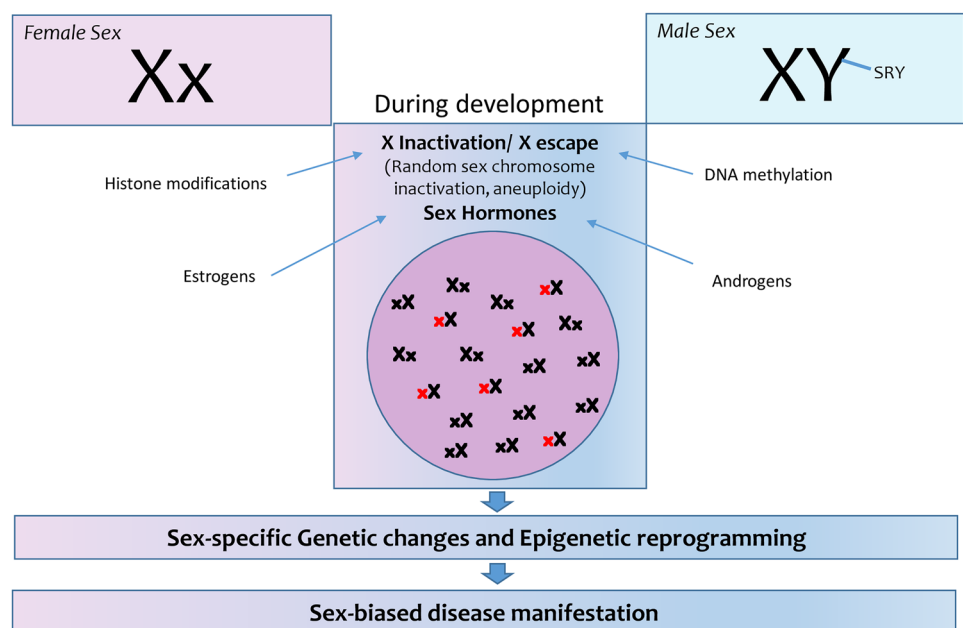
immunosuppression in post-pubertal males [26]. Moreover, a greater number of neutrophils was shown to infiltrate the pulmonary tissues, being accompanied by elevated inflammatory mediators, such as IL-17A, which has been linked to lower levels of female sex hormones [29, 30]. Yang et al. showed that female sex hormones exhibit a protective effect against bacterial pneumonia by enhancing the antimicrobial action of macrophages [31]. This was explained by the estrogen-mediated increase in the expression of nitric oxide synthase-3 (NOS-3), which enhances the killing of bacteria ingested by macrophages. Previous studies have demonstrated an association between estrogen receptors (ERs) with NOS-3 and the lncRNAs HOTAIR and MALAT1 in breast and prostate cancer cells [32]. These two lncRNAs were shown to be regulated by estrogens and further interact with NOS-3 and ERs [32], suggesting a potential role for sex hormones to control the macrophage bactericidal properties in an epigenetic manner.

A similar trend has been reported in mouse models treated with *Mycoplasma pulmonis*, where the mortality of males was increased compared to females and was correlated with dense inflammatory cell infiltrates within the pulmonary alveoli of males [33]. Interestingly, there were no big differences in IgM serum levels between male and female mice.

Effects of COVID-19 infection in male versus female patients

The COVID-19 pandemic has led to numerous deaths worldwide. In a recent study, an increased incidence of COVID-19 without respiratory failure was observed in males (57.7%)

Fig. 1 Genetic changes and epigenetic reprogramming during development underlie the dosage compensation in female and male cells, contributing to sex-biased disease (3.1) manifestation and progression



compared to females (42.3%), but a significantly increased incidence of COVID-19-related severe respiratory failure was detected in males (89.3%) compared to females (10.7%). Moreover, the death rates in the different age groups revealed that males were more likely to die of the infection in age groups below 90 years old [34].

In search of the underlying mechanisms that define the sex bias in infection severity and pathogenicity of the virus, several preliminary studies have proposed different biological mechanisms regulating immune responses in a sex-biased manner, contributing to viral defense mechanisms [29–35].

Studies in mice have shown that male mice demonstrate a greater susceptibility to severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection compared to female mice of the same age, with the difference being far more prominent at increasing age. Males exhibited increased virus titers and vascular leakage, more alveolar edema as well as inflammatory monocyte and neutrophil lung accumulation, whereas no differences between the two sexes were associated with T or B cell activity. The increased susceptibility of male mice to SARS-CoV-2 infection was mainly attributed to the effect of estrogens and their contribution to viral defense, since ovariectomy or the use of estrogen receptor antagonists led to increased mortality in female mice, highlighting the protective effects of estrogen receptor signaling in SARS-CoV-2 infections [35].

On the other hand, several molecular factors induce the expression of key proteins in viral entry at the epithelial cells, which are also affecting the infectivity rate. One such entry protein expressed in host cells is angiotensin converting enzyme 2 (ACE2). ACE2 is a dipeptidyl carboxy dipeptidase of the angiotensin-converting enzyme family which cleaves angiotensin I into angiotensin 1-9 and angiotensin II into the vasodilator angiotensin 1-7. ACE2 is the entry point of the SARS-CoV-2 virus, and its levels define the magnitude of viral infection. Regulation of this protein is fine-tuned by genetic and epigenetic mechanisms [36–41]. The *Ace2* gene in humans is located on the X chromosome, implying a potential sex-specific gene regulation. The *Ace2* gene is highly expressed in a lot of tissues including the epithelial cells of the lung, heart, and testis. It is an X chromosome inactivation escaper gene and therefore its expression is not regulated by the dosage compensation machinery [42]. Male cells are expressing higher levels of the ACE2 protein, especially in the lungs, evidence that may explain potential sex differences that have been observed during the progression of the COVID-19 infection and its complications [43]. A study of 450 DNA methylation data from 244 fresh human lung tissues has detected a differential methylation profile for the *Ace2* gene at cg23232263 and cg16734967 [44]. It was shown that the male lung tissue had decreased methylation levels of this gene compared to females. Furthermore, males

and females who were smokers or had chronic obstructive pulmonary disease demonstrated increased levels of *Ace2* gene methylation [44]. In addition, DNA methylation levels at a CpG island in the dataset related to the *Ace2* gene exhibited a large degree of variability in both men and women suggesting that DNA methylation of *Ace2* varies by individual. Of note, these datasets did not have comparable metadata for age and the differences in DNA methylation related to *Ace2* may reflect the cell type differences in lung tissues [19, 45–47].

Sex-bias effects in respiratory diseases

Sex-based differences are also prevalent in respiratory diseases, with DNA methylation changes between the two sexes being observed as early as at birth [48]. In respect to asthma, during the early years of childhood, there is an uneven distribution of the disease between males and females, with exceeding incidence in males compared to females [49–52]. This distribution, however, changes at pre-adolescence, when females begin to get asthma more frequently than males, and continues at post-adolescence where the increased incidence in females leads to a “sex-reversal” [53] and a more severe phenotype [54]. Asthma appears to be associated with changes in the DNA methylome [55]. In the study by Patel et al., the DNA methylation patterns were observed in a variety of genes in pre-adolescent and post-adolescent subjects [56]. Their results pointed towards 13 CpG sites which demonstrate sex and age-related changes. They also detected a sex-related effect difference at 9 of those CpG islands, including an increase in CpG site cg03269757 methylation which led to decreased risk of asthma acquisition in males and increased risk in females. In a similar manner, 5 genes which were mapped to CpG islands showed a sex-related response to elevation of DNA methylation, indicating that the effect of DNA methylation was affected by the subject’s sex. Increased methylation of the cg11295724 CpG island at subjects of 10 years of age resulted in increased expression of the signal regulatory protein delta precursor (*SIRPD*) gene which regulates T cell activation in males and its decreased expression in females. Moreover, the interferon related developmental regulator 1 (*IFRDI*) gene, a transcription factor which regulates skeletal muscle differentiation in asthmatic responses of the airway, shows strong sex-specific effects on asthma transition.

Sex-related differences have also been observed in chronic obstructive pulmonary disease (COPD) [57] and many epigenetic pathways have been associated with smoking [58]. Koo et al. investigated tissue samples from adult patients and fetal lungs, as well as cord blood and detected differences in CpG sites, such as cg03691818 where the keratin 77 (*KRT77*) gene is mapped. Of note, keratins are involved in the structural integrity of epithelia cells and this

site was found to be hypomethylated in males compared to females, an effect observed also in other CpG sites [59].

Furthermore, females demonstrated a predisposition for the development of pulmonary arterial hypertension (PAH) which has been partly attributed to the epigenetic regulation of the *Ephx2* gene [60]. This gene codes for the soluble epoxide hydrolase, an enzyme which degrades epoxyeicosatrienic acids (EETs), regulating pulmonary circulation. The increased amount of estrogens observed in females is responsible for the downregulation of this gene expression through an epigenetic pathway, leading to increased EETs and subsequently, high PAH susceptibility [60].

Sex bias in autoimmune diseases

Autoimmune diseases demonstrate a high prevalence, affecting an estimated 5–10% of the population, with a female predominance. However, differences have been observed in female to male ratios of each autoimmune disease. For instance, the sex ratio in inflammatory bowel disease and diabetes mellitus type I exceeds 1:1 by a slight amount, but increases to almost 2:1 in multiple sclerosis (MS), to 3:1 in rheumatoid arthritis (RA), and to 9:1–10:1 in systemic lupus erythematosus (SLE) [22]. The molecular mechanisms underlying these differences are not well-understood. However, recent studies suggest that sex hormones such as estrogens and the X chromosome silencing are important drivers of sex-biased manifestations in autoimmune diseases [61].

Estrogens act through four different pathways, including the classical receptor-mediated, the non-classical, the non-ligand-mediated genomic (nuclear), and the non-genomic (extranuclear) pathway, to influence downstream gene expression, protein modifications, and signaling. The molecular actions of estrogens are mediated through the genomic and non-genomic pathways [62]. In the genomic pathway, estrogens bind to intracellular receptors (estrogen receptor, ER α , and ER β). Upon binding, ERs are dimerized and translocated to the nucleus where they bind to specific motifs, the estrogen response elements (ERE), present in the target DNA. The consensus ERE site is 5'-GGTCAnnnTGACC-3' and is located in promoters as well as in the distal regulatory elements of genes, affecting immune response and cell signaling pathways. On the other hand, in the non-classical genomic pathways, ER directly bound to DNA, cross talks with various transcription factors, or it may act in a tether-mediated manner as a co-factor of transcription factors, including the activating protein 1 (AP-1), NF- κ B, and p300 proteins. In this way, estrogens are able to molecularly fine-tune adaptive and immune cell responsive pathways [62]. For example, activated T cells demonstrate estrogen receptor expression. At the same time, both ER mRNA and protein levels have been found in T and B cells, monocytes, and

dendritic cells showing the importance of the hormone in immune response activation [63].

Sex hormone receptors are widely expressed in immune cells, while androgen and estrogen response elements are present in several genes involved in immune response, indicating a major effect of sex hormones on the inflammatory response pathways. Furthermore, a lot of variations have been detected during puberty, pregnancy, and menopause demonstrating the complex regulation of immunity by sex hormones [64].

The X chromosome contains a wide variety of genes which are related to the immune response [65]. In this way, immune function and dysregulation may stem from abnormal silencing of the X chromosome as well as from major defects on X chromosome genes, such as IL-1R associated kinase 1 (*IRAK1*), IL-2R γ chain, IL-3R α chain, IL-13 α chain, *GATA1*, *TLR7*, *CD40L*, and *FOXP3* [61]. The involvement of the X chromosome in sex-biased immune responses is demonstrated in inherited disorders, including Klinefelter syndrome in males with XXY and Turner syndrome in females with XO, both characterized by hormonal and immune abnormalities [66, 67].

The effects of microRNAs (miRNA) in a sex-specific manner could potentially further promote differences in the immune response between the two sexes. It has been previously shown that miRNAs could play an inflammatory role by controlling the expression of important genes, such as *IL-1* and *TNF- α* which are involved in systemic inflammation [68]. Differences between the two sexes based on the expression of the X-linked miRNA could explain the stronger immune system in females [61]. Despite the vast gap in knowledge concerning the differences between male and female miRNA expression, it has been found that the X chromosome contains nearly 800 miRNAs [69]. Some of the X-linked miRNAs have been associated with immune regulation. More specifically, miR106a, which is located on the X chromosome, has been shown to downregulate the anti-inflammatory cytokine IL-10 [70]. Furthermore, the X chromosome-contained miR-17-92 cluster was shown to be a crucial factor for B and T cell maturation [71, 72]. The other side of this phenomenon is that females are more likely to develop autoimmune diseases [67].

The X chromosome contains the information coding for 10% of the human genome miRNAs versus 2% of miRNAs on chromosome Y. The differential expression of miRNAs (for example, miR-18b, miR-223) from X chromosomes due to female mosaicism, X inactivation, and silencing escape may explain the higher responses of the immune system upon infection that are observed in females compared to males [68, 73].

A recent study by Moeser et al. showed that perinatal sex hormones during the embryonic development are responsible for mast cell-associated disorders later in life.

Naturally, a high level of perinatal androgens is associated to reduced severity of mast cell-mediated anaphylaxis in male mice [74–76]. Mast cells recognized as effector cells of the immune system represent the first line of defense against various toxins. Their production and function are orchestrated by the activity of sex hormones, such as androgens and testosterone, regulating immune responses. Overactivation of mast cells has been connected to immunological disorders, chronic inflammatory diseases, and death. As part of their defense mechanism, mast cells release proteases, histamine, and serotonin [75]. Previous studies have shown that female mast cells produce, store, and release more proteases and amines than males. Thus, female mast cells are more capable to induce aggressive immune responses, leading to inflammatory and autoimmune diseases than the respective male cells [75, 77].

Current research has unraveled the mechanisms of action of perinatal hormones and the way sex may interfere with the severity of anaphylactic responses. In male embryos, high levels of perinatal androgens stimulate mast stem cells in the bone marrow to acquire a “masculinized” phenotype with reduced production and storage of granule mediators. As a result, they produce and release lower levels of inflammatory substances, leading to reduced possibility of anaphylactic responses and manifestation of chronic inflammatory disorders during the adult life. On the other hand, female embryos exposed to perinatal androgens exhibit reduced histamine levels and less-severe anaphylactic responses as adults [75].

Changes in chromatin configuration have been associated with the relaxation of XCI in female patients and the manifestation of autoimmune diseases. Notably, both naive T and B lymphocytes are characterized by an uncommon pattern of the inactive X chromosome at the level of chromosomal organization. Xist RNA and histone repressive marks are delocalized and reversed, leading to a minor biallelic transcription of autoimmune disease-associated X-linked genes, such as *CD40L* and *CXCR3* [78]. Following lymphocyte activation, such epigenetic changes on the X chromosome are normally reversed in females but not in SLE patients. Biallelic expression of genes connected to the manifestation of the disease is higher in female patients compared to healthy individuals [79–81].

The Y chromosome in mammals is responsible for carrying genes contributing to sex differences and especially those related to testis maturation and spermatogenesis. Several studies in mice have revealed the importance of this chromosome in immune responses and manifestations of autoimmune diseases. Gene expression profiling in these mice demonstrated that Y variants control many immune response genes found on other chromosomes, functioning as trans-expression quantitative trait loci (eQTLs) [82].

In females, several genetic variants and X-linked genes are associated to the risk of autoimmune diseases, such as SLE and MS. One of these important genes, forkhead box p3 (*FOXP3a*), encodes a pioneer factor controlling T lymphocyte development into regulatory cells and suppressing self-reactivity [83].

An interesting observation comes from the studies in men with Klinefelter syndrome (XXY) and in females with Turner syndrome (X0). The presence of two X chromosomes increases the incidence of severe autoimmune diseases, indicating that X chromosome imbalance rather than hormonal differences is likely to contribute to immune disorders in a sex-specific manner [84]. This imbalance could come from both epigenetic and genetic factors that misregulate the X inactivation center across the single female chromosomes. Supporting this hypothesis, two escape genes, *TLR7* and *CD40L*, are characterized by elevated expression in XX females, as well as XXY males versus XY males and X0 females [85].

Epigenetic mechanisms regulating liver fibrosis in a sex-specific manner

Transcriptional silencing of targeted genes is mainly epigenetically regulated through specific histone post-translational modifications, changes in DNA chemical modifications, and interplay with miRNAs that influence chromatin accessibility and gene expression patterns. Dysregulation of epigenetic mechanisms is often linked to certain diseases, such as cancer, liver fibrosis, and autoimmune disorders [86].

One of the most important protein complexes mediating gene silencing is PRC2 and its components EZH1 and EZH2 which deposit H3K27me₃, inducing chromatin compaction. EZH1 and EZH2 possess complementary functions. EZH1 is mainly expressed in adult tissues and EZH2 dominates in embryonic cells, while both enzymes regulate a small set of the same genes through their activity [87–90].

Although the importance of chromatin states in the sex-dependent regulation of genes in female and male mouse livers was previously reported [91], the recent study of Lau-Corona et al. elucidated the sex-biased action of the two methyltransferases EZH1 and EZH2 in liver fibrosis and liver metabolism [92]. They detected that the trimethylation of histone H3K27 (H3K27me₃) is a significant sex-biased repressive mark localized on many female-specific expressed genes, including cytochrome P450 (*Cyp* genes) and sulfotransferase (*Sult* genes) of male mouse livers, but not at male-biased genes of female mouse livers. In support to these findings, RNAseq data showed that various female-biased genes were upregulated (derepressed) in E1/E2-KO male livers and only a few male-biased genes in E1/E2-KO female livers [91, 92].

Sex-biased expression of related liver genes could explain why males exhibit increased susceptibility to liver fibrosis and showed increased male incidence, as well as progression of HCC [91]. Thus, several studies evaluated the implementation of the PRC2 complex in the progression of the disease. By using double knock-out mice for *Ezh1* and *Ezh2*, they evaluated the epigenetic effects of these enzymes on a genome-wide level and the differences between the two sexes in the liver. As a result, more HCC and fibrosis-related genes were found to be dysregulated in E1/E2-KO male versus E1/E2-KO female liver. Thirty-two (32) sex-independent genes were identified as more highly upregulated in E1/E2-KO female than in E1/E2-KO male mouse livers. Moreover, in the double knock out mice, three genes which are strictly associated with HCC, including the lncRNA gene *H19*, *Igf2*, and *miR675*, were further characterized by accumulation of active marks on their promoters and open chromatin configurations that led to their overexpression in female mice [91].

Sex bias in neuropsychiatric disorders

Genes usually upregulated in autism spectrum disorder (ASD) were found to be located in hypermethylated regions in females. Hypermethylation could be responsible for decreased ASD-associated gene expression in females compared to males, explaining the difference in prevalence between the two sexes [93]. On the other hand, genes that are downregulated in major depressive disorder (MDD) were also found to be hypermethylated in females, which could explain the higher disease predisposition in females [93].

XY homologous genes escape X-inactivation, and the homolog genes of the Y-chromosome non-recombining region are of special interest in males. Differences in the X and Y homolog gene pairs contribute to sex differences, for example, in the ages of onset in psychosis. One example is the *KDM5C* gene which escapes X-inactivation and the *NLGN4* gene which has been linked to autism, as well as the *Protocadherin11 XY* gene-pair which has been associated with psychosis [94].

Sex chromosomes also seem to contribute to sex differences in Alzheimer's disease (AD) [95]. An additional X chromosome may result in a protective effect against AD by promoting the transcription of genes that would normally escape X inactivation. Around 15% of X-chromosome genes avoid this inactivation, leading to their increased expression in females compared to males [96]. For example, loss-of-function mutations of *KDM6A* which encodes for a histone demethylase have been associated with intellectual disability [97]. In this context, a recent study demonstrated that adding an extra X chromosome to mice makes them resilient to AD, likely due to increased *KDM6A* expression [98]. Another example is protocadherin 11 X-linked (*PCDH11X*), which

also escapes X inactivation via epigenetic mechanisms [99]. A single nucleotide polymorphism in the *PCDH11X* gene (rs5984894) has been linked to higher AD risk in women [100]. Moreover, men with higher levels of loss of chromosome Y (LOY), the most common acquired somatic mutation associated with aging, had greater risks of AD. It has been suggested that LOY might cause a dysfunctional immune system and contribute to abnormal neuronal differentiation as well as early cell death [101, 102]. Moreover, hypomethylated CpG islands were found in the *aurora kinase C* gene promoter in males with AD, which were hypermethylated in female AD patients. Females are also characterized by reduced histone deacetylase 2 levels, when compared to males [103] and long noncoding RNA levels were also found to differ between the two sexes in AD [104].

Cancer and sex biases rely on epigenetics

Recent studies have linked the gene dosage imbalance on the X chromosome with the manifestation of cancer in a sex-biased mode. Several associations and whole genome studies have shown that the X chromosome contains a wide variety of genes which demonstrate differential expression between the two sexes. Moreover, XCI-evading genes have been correlated with cancer in humans. More specifically, six genes which belong to the tumor suppressor family that evade the XCI (i.e., *CNKSR2*, *ATRX*, *DDX3X*, *KDM6A*, *KDM5C*, and *MAGEC3*) have been shown to play a protective role in females when loss-of-function mutations are present, and partially explain the increased incidence of 23 cancer types in males compared to females [105]. Among them, *KDM6A* codes for a histone lysine demethylase which demonstrates functional variations from its Y homolog. A recent pioneer study in mice has associated the effects of this enzyme with the manifestation of bladder cancer, uncovering genetic and epigenetic alterations from hormonal effects associated with cancer [106].

Another interesting example is the case of increased male to female ratio in the incidence and mortality of glioblastoma which contradicts the increased mutation burden observed in females, but a higher incidence and mortality in men.

Many genes that are mutated in cancer cells are located on the X chromosome which has been shown to contain several areas which possess tumor suppressor genes [107]. This means that males are more susceptible to single hits in these loci, leading to increased propensity for cancer. These loci, however, are also commonly mutated in female cancers, such as breast and ovarian cancer [108, 109]. The alpha thalassemia/mental retardation syndrome X-linked (*ATRX*) gene which is responsible for DNA repair is also frequently found to be mutated [110]. The expression of this gene is tightly regulated in females by the Xist RNA and repressive

Table 1 Epigenetic mechanisms and sex bias in various diseases

Disease	Epigenetic mechanism	Affected gene	References
Respiratory diseases			
LRTIs	Increased susceptibility and mortality of males, since estrogens regulate lncRNAs, which interact with NOS-3 that enhances macrophage-mediated killing of bacteria		[32]
COVID-19	Increased susceptibility and mortality of males, due to estrogen contribution to viral defense and decreased gene methylation in males	<i>ACE2</i>	[44]
Asthma	Increased incidence in pre-adolescent males and post-adolescent females, due to sex and age-related changes in gene methylation	<i>SIRPD, IFRD1</i>	[55, 56]
COPD	Decreased gene methylation in males	<i>KRT77</i>	[59]
PAH	Increased susceptibility in females, due to estrogen-mediated increase in gene methylation	<i>Ephx2</i>	[60]
Autoimmune diseases			
IBD, MS, RA, SLE	Increased susceptibility in females, due to estrogen-mediated control of immune responses, abnormal silencing of X-linked genes and X-linked miRNA expression (miR106A, miR-17-92, miR-18b, miR-223)	<i>IL-1R/2R/3R, IL-13, TLR7, IRAK1, FOXP3, GATA1, CD40L, IL-10, CXCR3</i>	[61]
Gastrointestinal diseases			
HCC and liver fibrosis	Increased susceptibility in males due to increased histone methylation in males, causing gene silencing	<i>Cyp, Sult, H19, Igf2, miR675</i>	[91]
Neuropsychiatric diseases			
ASD	Increased incidence in males due to: Decreased ASD-related gene methylation and thus increased expression		[93]
	Expression of Y-linked genes escaping inactivation	<i>KDM5C, NLGN4</i>	[94]
MDD	Increased incidence in females, due to increased gene methylation and thus decreased gene expression of anti-MDD genes		[93]
AD	Increased incidence in males due to: Decreased gene inactivation in females Hypomethylation of genes in males Reduced HDAC2 levels in females	<i>KDM6A, PCDH11X</i> <i>Aurora kinase C</i>	[98] [103] [103]
Cancer			
43 cancer types including glioblastoma	Decreased incidence and mortality of some cancer types, e.g., glioblastoma in females due to tumor suppressor genes escaping XCI	<i>CNKSR2, ATRX, DDX3X, KDM6A, KDM5C, MAGEC3</i>	[105]
Breast and ovarian cancer	Increased susceptibility in females due to dysregulation of the Xist RNA expression and repressive marks, causing gene expression imbalances	<i>ATRX</i>	[111]
Stomach and esophageal cancer	Increased susceptibility in females due to gene mutations, hindering interaction with ATM protein kinase to ensure efficient DNA damage response and repair	<i>EP400</i>	[112]

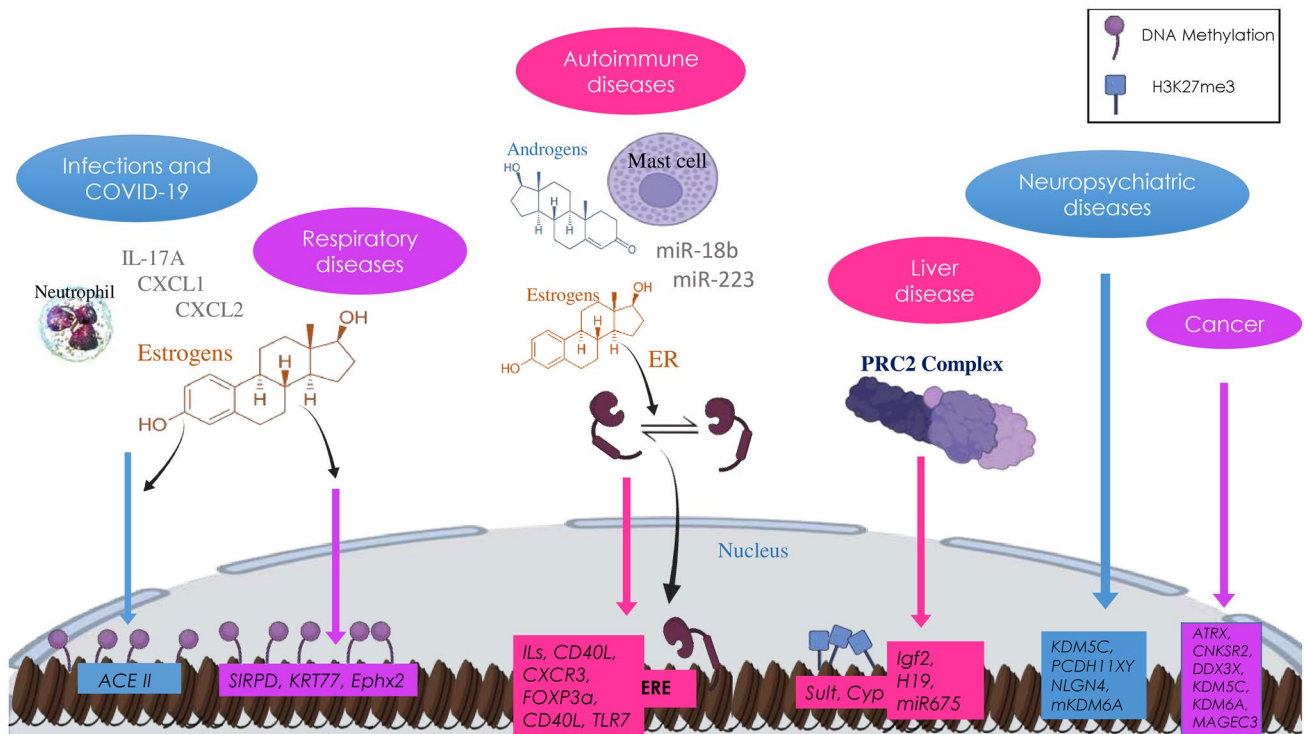


Fig. 2 Sex-bias epigenetic mechanisms underlie the manifestation of various diseases. Women and men differ in the susceptibility and manifestations of various diseases due to epigenetic mechanisms reg-

ulating the effects of sex hormones, as well as the differential expression of X-chromosome-encoded genes

epigenetic marks, to achieve a monoallelic expression and proper dosage balance. Upon misregulation of the Xist RNA expression, incomplete DNA repair and tumor progression occur. Interestingly, this gene escapes the X inactivation center (XIC) only in the brain of female individuals, resulting in its biallelic expression, protecting females against brain tumors, and more specifically glioblastomas [111].

Moreover, the E1A-binding protein p400, EP400, a component of the nucleosome-acetyltransferase of histone H4 (NuA4) complex, is responsible for acetylating histones H4 and H2A, leading to transcription and high expression of targeted genes. It interacts with the Myc protein to regulate target gene expression during proliferation. Recently, a functional interaction of EP400 with the ataxia-telangiectasia mutated (ATM) protein kinase has been reported, which works to achieve efficient DNA damage response and repair [112]. Taken together, these findings shed light to the importance of mutations on EP400 as tumor drivers. A study of male and female patients with stomach and esophageal cancers has revealed an association between the EP400 mutation and an increased mutation load in female-derived samples when compared to male ones [113].

Lastly, there is a hypothesis in the field of cancer biology that chromatin configuration and genome architecture could be manipulated in a sex-specific manner. This change can

further contribute to the manifestation of several diseases, including cancer. Supporting this working model, recent studies have shown that two major factors of genome topology and genome organization (i.e., COHESIN and CTCF) bind via a sex-related manner to chromatin and that mutated binding sites of CTCF may induce tumorigenic effects [79, 114, 115]. Moreover, sex-specific chromatin quaternary structure and accessibility could influence the efficacy or efficiency of the DNA damage response. This evidence provides a better understanding of the mechanistic aspect of the DNA damage response and its effect in the sex bias observed in tumor development. Further research addressing both genetic as well as epigenetic alterations in normal and precancerous tissues is expected to shed more light into the mechanisms underlying sex-specific differences in genomic mutations and their repair.

Conclusion—future perspectives

Taken all together, it is evident that several disease manifestations are attributed to sex bias mediated by epigenetics mechanisms and are not always related to sex hormones (Table 1, Fig. 2). The unique genomic and epigenomic profiling of the two sexes could further define the respective

immune response. One of the main drivers for this differentiation is the dosage compensation machinery between the two sexes. Through X inactivation and chromosomal organization, which occur very early in development, female cells regulate a series of critical genes in their lifespan. Misregulation of X inactivation process leads to a sex-biased manifestation of several immunological disorders. The underlying regulatory mechanism is based on epigenetic factors, such as DNA and histone methylation, which drives gene silencing on the inactive female X chromosome.

Several studies are currently elucidating the critical role of epigenetic enzymes, such as EZH2 and respective histone modifications to the sex-biased manifestation of diseases. Recently, COVID-19 susceptibility and progression were connected to epigenetic factors and mechanisms that act in a different way between the two sexes. Further studies are, however, needed to fully uncover the sex-biased epigenome in different disease states, opening a new window to personalized medicine and treatment, with the use of drugs targeting important epigenetic mechanisms.

Author contribution SC, MM, and DS performed the literature survey and wrote the first draft of the manuscript. CP conceived, revised, and edited the final manuscript. All authors read and approved the final manuscript.

Data availability Not applicable.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication Not applicable.

Competing interests The authors declare no competing interests.

References

- Greer EL, Shi Y (2012) Histone methylation: a dynamic mark in health, disease and inheritance. *Nat Rev Genet* 13:343–357. <https://doi.org/10.1038/nrg3173>
- Trotman JB, Calabrese JM (2020) How to silence an X chromosome. *Nature* 578:365–366
- Orphanides G, Reinberg D (2002) A unified theory of gene expression. *Cell* 108:439–451. [https://doi.org/10.1016/s0092-8674\(02\)00655-4](https://doi.org/10.1016/s0092-8674(02)00655-4)
- Bannister AJ, Kouzarides T (2011) Regulation of chromatin by histone modifications. *Cell Res* 21:381–395. <https://doi.org/10.1038/cr.2011.22>
- Allfrey VG, Faulkner R, Mirsky AE (1964) Acetylation and methylation of histones and their possible role in the regulation of RNA synthesis. *Proc Natl Acad Sci U S A* 51:786–794. <https://doi.org/10.1073/pnas.51.5.786>
- Luger K, Mäder AW, Richmond RK et al (1997) Crystal structure of the nucleosome core particle at 2.8 Å resolution. *Nature* 389:251–260. <https://doi.org/10.1038/38444>
- Xhemalce B, Dawson MA, Bannister AJ (2011) Histone modifications. *Rev Cell Biol Mol Med*
- Kouzarides T (2007) Chromatin modifications and their function. *Cell* 128:693–705. <https://doi.org/10.1016/j.cell.2007.02.005>
- Tchasovnikarova IA, Kingston RE (2018) Beyond the histone code: a physical map of chromatin states. *Mol Cell* 69:5–7. <https://doi.org/10.1016/j.molcel.2017.12.018>
- Peterson CL, Lanier M-A (2004) Histones and histone modifications. *Curr Biol* 14:R546–51. <https://doi.org/10.1016/j.cub.2004.07.007>
- Ng MK, Cheung P (2015) A brief histone in time: understanding the combinatorial functions of histone PTMs in the nucleosome context. *Biochem Cell Biol* 94:33–42. <https://doi.org/10.1139/bcb-2015-0031>
- Henikoff S, Shilatifard A (2011) Histone modification: cause or cog? *Trends Genet* 27:389–396. <https://doi.org/10.1016/j.tig.2011.06.006>
- Snyder EM, Small CL, Li Y, Griswold MD (2009) Regulation of gene expression by estrogen and testosterone in the proximal mouse reproductive tract. *Biol Reprod* 81:707–716. <https://doi.org/10.1095/biolreprod.109.079053>
- Aranda A, Pascual A (2001) Nuclear hormone receptors and gene expression. *Physiol Rev* 81:1269–1304. <https://doi.org/10.1152/physrev.2001.81.3.1269>
- Parker MG (1998) Transcriptional activation by oestrogen receptors. *Biochem Soc Symp* 63:45–50
- Dilworth FJ, Chambon P (2001) Nuclear receptors coordinate the activities of chromatin remodeling complexes and coactivators to facilitate initiation of transcription. *Oncogene* 20:3047–3054. <https://doi.org/10.1038/sj.onc.1204329>
- Galupa R, Heard E (2018) X-chromosome inactivation: a crossroads between chromosome architecture and gene regulation. *Annu Rev Genet* 52:535–566. <https://doi.org/10.1146/annurev-genet-120116-024611>
- Żylicz JJ, Bousard A, Žumer K et al (2019) The implication of early chromatin changes in X chromosome inactivation. *Cell* 176:182–197.e23. <https://doi.org/10.1016/j.cell.2018.11.041>
- Tukiainen T, Villani A-C, Yen A et al (2017) Landscape of X chromosome inactivation across human tissues. *Nature* 550:244–248. <https://doi.org/10.1038/nature24265>
- Żylicz JJ, Heard E (2020) Molecular mechanisms of facultative heterochromatin formation: an X-chromosome perspective. *Annu Rev Biochem* 89:255–282. <https://doi.org/10.1146/annurev-biochem-062917-012655>
- Wang X, Soloway PD, Clark AG (2010) Paternally biased X inactivation in mouse neonatal brain. *Genome Biol* 11:R79–R79. <https://doi.org/10.1186/gb-2010-11-7-r79>
- Invernizzi P, Pasini S, Selmi C et al (2009) Female predominance and X chromosome defects in autoimmune diseases. *J Autoimmun* 33:12–16. <https://doi.org/10.1016/j.jaut.2009.03.005>
- Fang H, Distèche CM, Berletch JB (2019) X inactivation and escape: epigenetic and structural features. *Front Cell Dev Biol* 7:219. <https://doi.org/10.3389/fcell.2019.00219>
- Markaki Y, Gan Chong J, Wang Y et al (2021) Xist nucleates local protein gradients to propagate silencing across the X chromosome. *Cell* 184:6174–6192.e32. <https://doi.org/10.1016/j.cell.2021.10.022>
- Collombet S, Ranisavljevic N, Nagano T et al (2020) Parental-to-embryo switch of chromosome organization in early embryogenesis. *Nature* 580:142–146. <https://doi.org/10.1038/s41586-020-2125-z>
- Loda A, Heard E (2019) Xist RNA in action: past, present, and future. *PLoS Genet* 15:e1008333–e1008333. <https://doi.org/10.1371/journal.pgen.1008333>
- Okamoto I, Patrat C, Thépot D et al (2011) Eutherian mammals use diverse strategies to initiate X-chromosome inactivation

- during development. *Nature* 472:370–374. <https://doi.org/10.1038/nature09872>
28. Kalantry S, Magnuson T (2006) The polycomb group protein EED is dispensable for the initiation of random X-chromosome inactivation. *PLOS Genet* 2:e66
 29. Kadioglu A, Cuppone AM, Trappetti C et al (2011) Sex-based differences in susceptibility to respiratory and systemic pneumococcal disease in mice. *J Infect Dis* 204:1971–1979. <https://doi.org/10.1093/infdis/jir657>
 30. Tyagi AM, Srivastava K, Mansoori MN et al (2012) Estrogen deficiency induces the differentiation of IL-17 secreting Th17 cells: a new candidate in the pathogenesis of osteoporosis. *PLoS One* 7:e44552–e44552. <https://doi.org/10.1371/journal.pone.0044552>
 31. Yang Z, Huang Y-CT, Koziel H et al (2014) Female resistance to pneumonia identifies lung macrophage nitric oxide synthase-3 as a therapeutic target. *Elife* 3:e03711. <https://doi.org/10.7554/eLife.03711>
 32. Aiello A, Bacci L, Re A et al (2016) MALAT1 and HOTAIR long non-coding RNAs play opposite role in estrogen-mediated transcriptional regulation in prostate cancer cells. *Sci Rep* 6:38414. <https://doi.org/10.1038/srep38414>
 33. Yancey AL, Watson HL, Cartner SC, Simecka JW (2001) Gender is a major factor in determining the severity of mycoplasma respiratory disease in mice. *Infect Immun* 69:2865–2871. <https://doi.org/10.1128/IAI.69.5.2865-2871.2001>
 34. Li Y, Jerkic M, Slutsky AS, Zhang H (2020) Molecular mechanisms of sex bias differences in COVID-19 mortality. *Crit Care* 24:405. <https://doi.org/10.1186/s13054-020-03118-8>
 35. Channappanavar R, Fett C, Mack M et al (2017) Sex-based differences in susceptibility to severe acute respiratory syndrome coronavirus infection. *J Immunol* 198:4046–4053. <https://doi.org/10.4049/jimmunol.1601896>
 36. Kim D, Lee J-Y, Yang J-S et al (2020) The architecture of SARS-CoV-2 transcriptome. *Cell* 181:914–921.e10. <https://doi.org/10.1016/j.cell.2020.04.011>
 37. Kim J-M, Chung Y-S, Jo HJ et al (2020) Identification of coronavirus isolated from a patient in Korea with COVID-19. *Osong public Heal Res Perspect* 11:3–7. <https://doi.org/10.24171/j.phrp.2020.11.1.02>
 38. Jia HP, Look DC, Shi L et al (2005) ACE2 receptor expression and severe acute respiratory syndrome coronavirus infection depend on differentiation of human airway epithelia. *J Virol* 79:14614–14621. <https://doi.org/10.1128/JVI.79.23.14614-14621.2005>
 39. Verdecchia P, Cavallini C, Spanevello A, Angeli F (2020) The pivotal link between ACE2 deficiency and SARS-CoV-2 infection. *Eur J Intern Med* 76:14–20. <https://doi.org/10.1016/j.ejim.2020.04.037>
 40. Wan Y, Shang J, Graham R et al (2020) Receptor recognition by the novel coronavirus from Wuhan: an analysis based on decade-long structural studies of SARS coronavirus. *J Virol* 94:e00127–20. <https://doi.org/10.1128/JVI.00127-20>
 41. Li F (2016) Structure, function, and evolution of coronavirus spike proteins. *Annu Rev Virol* 3:237–261. <https://doi.org/10.1146/annurev-virology-110615-042301>
 42. Gemmati D, Bramanti B, Serino ML, et al (2020) COVID-19 and individual genetic susceptibility/receptivity: role of ACE1/ACE2 genes, immunity, inflammation and coagulation. Might the double X-chromosome in females be protective against SARS-CoV-2 compared to the single X-chromosome in males? *Int J Mol Sci* 21:3474. <https://doi.org/10.3390/ijms21103474>
 43. Zhao Y, Zhao Z, Wang Y et al (2020) Single-cell RNA expression profiling of ACE2, the receptor of SARS-CoV-2. *Am J Respir Crit Care Med* 202:756–759. <https://doi.org/10.1164/rccm.202001-0179LE>
 44. Beacon TH, Delcuve GP, Davie JR (2020) Epigenetic regulation of ACE2, the receptor of the SARS-CoV-2 virus. *Coronavirus Relat Res Collect* 1:386–399. <https://doi.org/10.1139/gen-2020-0124@cfac.issue1>
 45. Corley MJ, Ndhlovu LC (2020) DNA methylation analysis of the COVID-19 host cell receptor, angiotensin I converting enzyme 2 gene (ACE2) in the respiratory system reveal age and gender differences. Preprints. <https://doi.org/10.20944/preprints202003.0295.v1>
 46. Sawalha AH, Zhao M, Coit P, Lu Q (2020) Epigenetic dysregulation of ACE2 and interferon-regulated genes might suggest increased COVID-19 susceptibility and severity in lupus patients. *Clin Immunol* 215:108410. <https://doi.org/10.1016/j.clim.2020.108410>
 47. Chlamydas S, Papavassiliou AG, Piperi C (2021) Epigenetic mechanisms regulating COVID-19 infection. *Epigenetics* 16:263–270. <https://doi.org/10.1080/15592294.2020.1796896>
 48. Mukherjee N, Arathimos R, Chen S et al (2021) DNA methylation at birth is associated with lung function development until age 26 years. *Eur Respir J*. <https://doi.org/10.1183/13993003.03505-2020>
 49. Arathimos R, Granell R, Henderson J et al (2017) Sex discordance in asthma and wheeze prevalence in two longitudinal cohorts. *PLoS One* 12:e0176293
 50. Fuseini H, Newcomb DC (2017) Mechanisms driving gender differences in asthma. *Curr Allergy Asthma Rep* 17:19. <https://doi.org/10.1007/s11882-017-0686-1>
 51. Postma DS (2007) Gender differences in asthma development and progression. *Gend Med* 4 Suppl B:S133–46. [https://doi.org/10.1016/s1550-8579\(07\)80054-4](https://doi.org/10.1016/s1550-8579(07)80054-4)
 52. Hohmann C, Keller T, Gehring U et al (2019) Sex-specific incidence of asthma, rhinitis and respiratory multimorbidity before and after puberty onset: individual participant meta-analysis of five birth cohorts collaborating in MeDALL. *BMJ open Respir Res* 6:e000460–e000460. <https://doi.org/10.1136/bmjresp-2019-000460>
 53. Vink NM, Postma DS, Schouten JP et al (2010) Gender differences in asthma development and remission during transition through puberty: the TRacking Adolescents' Individual Lives Survey (TRAILS) study. *J Allergy Clin Immunol* 126:496–498. <https://doi.org/10.1016/j.jaci.2010.06.018>
 54. Zein JG, Erzurum SC (2015) Asthma is different in women. *Curr Allergy Asthma Rep* 15:28. <https://doi.org/10.1007/s11882-015-0528-y>
 55. den Dekker HT, Burrows K, Felix JF et al (2019) Newborn DNA-methylation, childhood lung function, and the risks of asthma and COPD across the life course. *Eur Respir J* 53:1801795. <https://doi.org/10.1183/13993003.01795-2018>
 56. Patel R, Solatikia F, Zhang H et al (2021) Sex-specific associations of asthma acquisition with changes in DNA methylation during adolescence. *Clin Exp Allergy J Br Soc Allergy Clin Immunol* 51:318–328. <https://doi.org/10.1111/cea.13776>
 57. DeMeo DL, Ramagopalan S, Kavati A et al (2018) Women manifest more severe COPD symptoms across the life course. *Int J Chron Obstruct Pulmon Dis* 13:3021–3029. <https://doi.org/10.12147/COPD.S160270>
 58. McCartney DL, Stevenson AJ, Hillary RF et al (2018) Epigenetic signatures of starting and stopping smoking. *EBioMedicine* 37:214–220. <https://doi.org/10.1016/j.ebiom.2018.10.051>
 59. Koo H-K, Morrow J, Kachroo P et al (2021) Sex-specific associations with DNA methylation in lung tissue demonstrate smoking interactions. *Epigenetics* 16:692–703. <https://doi.org/10.1080/15592294.2020.1819662>
 60. Huang A, Kandhi S, Sun D (2021) Roles of genetic predisposition in the sex bias of pulmonary pathophysiology, as a function of estrogens : sex matters in the prevalence of lung diseases.

- Adv Exp Med Biol 1303:107–127. https://doi.org/10.1007/978-3-030-63046-1_7
61. Taneja V (2018) Sex hormones determine immune response. *Front Immunol* 9:1931. <https://doi.org/10.3389/fimmu.2018.01931>
 62. Lai Y-J, Yu D, Zhang JH, Chen G-J (2017) Cooperation of genomic and rapid nongenomic actions of estrogens in synaptic plasticity. *Mol Neurobiol* 54:4113–4126. <https://doi.org/10.1007/s12035-016-9979-y>
 63. Ortona E, Pierdominici M, Rider V (2019) Editorial: sex hormones and gender differences in immune responses. *Front Immunol* 10:1076. <https://doi.org/10.3389/fimmu.2019.01076>
 64. Kadel S, Kovats S (2018) Sex hormones regulate innate immune cells and promote sex differences in respiratory virus infection. *Front Immunol* 9:1653. <https://doi.org/10.3389/fimmu.2018.01653>
 65. Bianchi I, Lleo A, Gershwin ME, Invernizzi P (2012) The X chromosome and immune associated genes. *J Autoimmun* 38:J187–92. <https://doi.org/10.1016/j.jaut.2011.11.012>
 66. Abramowitz LK, Olivier-Van Stichelen S, Hanover JA (2014) Chromosome imbalance as a driver of sex disparity in disease. *J genomics* 2:77–88. <https://doi.org/10.7150/jgen.8123>
 67. Quintero OL, Amador-Patarroyo MJ, Montoya-Ortiz G et al (2012) Autoimmune disease and gender: plausible mechanisms for the female predominance of autoimmunity. *J Autoimmun* 38:J109–19. <https://doi.org/10.1016/j.jaut.2011.10.003>
 68. Pinheiro I, Dejager L, Libert C (2011) X-chromosome-located microRNAs in immunity: might they explain male/female differences? The X chromosome-genomic context may affect X-located miRNAs and downstream signaling, thereby contributing to the enhanced immune response of females. *Bioessays* 33:791–802. <https://doi.org/10.1002/bies.201100047>
 69. Guo X, Su B, Zhou Z, Sha J (2009) Rapid evolution of mammalian X-linked testis microRNAs. *BMC Genomics* 10:97. <https://doi.org/10.1186/1471-2164-10-97>
 70. Sharma A, Kumar M, Aich J et al (2009) Posttranscriptional regulation of interleukin-10 expression by hsa-miR-106a. *Proc Natl Acad Sci U S A* 106:5761–5766. <https://doi.org/10.1073/pnas.0808743106>
 71. Ventura A, Young AG, Winslow MM et al (2008) Targeted deletion reveals essential and overlapping functions of the miR-17 through 92 family of miRNA clusters. *Cell* 132:875–886. <https://doi.org/10.1016/j.cell.2008.02.019>
 72. Xiao C, Srinivasan L, Calado DP et al (2008) Lymphoproliferative disease and autoimmunity in mice with increased miR-17-92 expression in lymphocytes. *Nat Immunol* 9:405–414. <https://doi.org/10.1038/ni1575>
 73. Khalifa O, Pers Y-M, Ferreira R et al (2016) X-linked miRNAs associated with gender differences in rheumatoid arthritis. *Int J Mol Sci* 17:1852. <https://doi.org/10.3390/ijms17111852>
 74. Rubtsova K, Marrack P, Rubtsov AV (2015) Sexual dimorphism in autoimmunity. *J Clin Invest* 125:2187–2193. <https://doi.org/10.1172/JCI78082>
 75. Mackey E, Thelen KM, Bali V et al (2020) Perinatal androgens organize sex differences in mast cells and attenuate anaphylaxis severity into adulthood. *Proc Natl Acad Sci U S A* 117:23751–23761. <https://doi.org/10.1073/pnas.1915075117>
 76. Klein SL, Flanagan KL (2016) Sex differences in immune responses. *Nat Rev Immunol* 16:626–638. <https://doi.org/10.1038/nri.2016.90>
 77. Gupta K, Harvima IT (2018) Mast cell-neural interactions contribute to pain and itch. *Immunol Rev* 282:168–187. <https://doi.org/10.1111/imr.12622>
 78. Wang J, Syrett CM, Kramer MC et al (2016) Unusual maintenance of X chromosome inactivation predisposes female lymphocytes for increased expression from the inactive X. *Proc Natl Acad Sci U S A* 113:E2029–E2038. <https://doi.org/10.1073/pnas.1520113113>
 79. Credendino SC, Neumayer C, Cantone I (2020) Genetics and epigenetics of sex bias: insights from human cancer and autoimmunity. *Trends Genet* 36:650–663. <https://doi.org/10.1016/j.tig.2020.06.016>
 80. El-Osta A (2004) Introduction: understanding the consequences of epigenetic mechanisms and its effects on transcription in health and disease. *Cancer Biol Ther* 3:816–818. <https://doi.org/10.4161/cbt.3.9.1100>
 81. Long H, Yin H, Wang L et al (2016) The critical role of epigenetics in systemic lupus erythematosus and autoimmunity. *J Autoimmun* 74:118–138. <https://doi.org/10.1016/j.jaut.2016.06.020>
 82. Case LK, Wall EH, Dragon JA et al (2013) The Y chromosome as a regulatory element shaping immune cell transcriptomes and susceptibility to autoimmune disease. *Genome Res* 23:1474–1485. <https://doi.org/10.1101/gr.156703.113>
 83. Hori S, Sakaguchi S (2004) Foxp3: a critical regulator of the development and function of regulatory T cells. *Microbes Infect* 6:745–751. <https://doi.org/10.1016/j.micinf.2004.02.020>
 84. Syrett CM, Anguera MC (2019) When the balance is broken: X-linked gene dosage from two X chromosomes and female-biased autoimmunity. *J Leukoc Biol* 106:919–932. <https://doi.org/10.1002/JLB.6RI0319-094R>
 85. Sarmiento L, Svensson J, Barchetta I et al (2019) Copy number of the X-linked genes TLR7 and CD40L influences innate and adaptive immune responses. *Scand J Immunol* 90:e12776. <https://doi.org/10.1111/sji.12776>
 86. Murata M (2018) Inflammation and cancer. *Environ Health Prev Med* 23:50. <https://doi.org/10.1186/s12199-018-0740-1>
 87. Schuettengruber B, Bourbon H-M, Di Croce L, Cavalli G (2017) Genome regulation by polycomb and trithorax: 70 years and counting. *Cell* 171:34–57. <https://doi.org/10.1016/j.cell.2017.08.002>
 88. Steffen PA, Ringrose L (2014) What are memories made of? How polycomb and trithorax proteins mediate epigenetic memory. *Nat Rev Mol Cell Biol* 15:340–356. <https://doi.org/10.1038/nrm3789>
 89. Piunti A, Shilatifard A (2016) Epigenetic balance of gene expression by polycomb and COMPASS families. *Science* 352:aad9780. <https://doi.org/10.1126/science.aad9780>
 90. Geisler SJ, Paro R (2015) Trithorax and polycomb group-dependent regulation: a tale of opposing activities. *Development* 142:2876–2887. <https://doi.org/10.1242/dev.120030>
 91. Sugathan A, Waxman DJ (2013) Genome-wide analysis of chromatin states reveals distinct mechanisms of sex-dependent gene regulation in male and female mouse liver. *Mol Cell Biol* 33:3594–3610. <https://doi.org/10.1128/MCB.00280-13>
 92. Lau-Corona D, Bae WK, Hennighausen L, Waxman DJ (2020) Sex-biased genetic programs in liver metabolism and liver fibrosis are controlled by EZH1 and EZH2. *PLoS Genet* 16:e1008796–e1008796. <https://doi.org/10.1371/journal.pgen.1008796>
 93. Xia Y, Dai R, Wang K et al (2021) Sex-differential DNA methylation and associated regulation networks in human brain implicated in the sex-biased risks of psychiatric disorders. *Mol Psychiatry* 26:835–848. <https://doi.org/10.1038/s41380-019-0416-2>
 94. Crow TJ (2015) Is psychosis a disorder of XY epigenetics?. *eBioMedicine* 2:794–795. <https://doi.org/10.1016/j.ebiom.2015.06.015>
 95. Guo L, Zhong MB, Zhang L et al (2022) Sex differences in Alzheimer's disease: insights from the multiomics landscape. *Biol Psychiatry* 91:61–71. <https://doi.org/10.1016/j.biopsych.2021.02.968>
 96. Carrel L, Willard HF (2005) X-inactivation profile reveals extensive variability in X-linked gene expression in females. *Nature* 434:400–404. <https://doi.org/10.1038/nature03479>

97. Miyake N, Mizuno S, Okamoto N et al (2013) KDM6A point mutations cause Kabuki syndrome. *Hum Mutat* 34:108–110. <https://doi.org/10.1002/humu.22229>
98. Davis EJ, Broestl L, Abdulai-Saiku S et al (2020) A second X chromosome contributes to resilience in a mouse model of Alzheimer's disease. *Sci Transl Med* 12:eaa5677. <https://doi.org/10.1126/scitranslmed.aaz5677>
99. Lopes AM, Ross N, Close J et al (2006) Inactivation status of PCDH11X: sexual dimorphisms in gene expression levels in brain. *Hum Genet* 119:267–275. <https://doi.org/10.1007/s00439-006-0134-0>
100. Carrasquillo MM, Zou F, Pankratz VS et al (2009) Genetic variation in PCDH11X is associated with susceptibility to late-onset Alzheimer's disease. *Nat Genet* 41:192–198. <https://doi.org/10.1038/ng.305>
101. Dumanski JP, Lambert J-C, Rasi C et al (2016) Mosaic loss of chromosome Y in blood is associated with Alzheimer disease. *Am J Hum Genet* 98:1208–1219. <https://doi.org/10.1016/j.ajhg.2016.05.014>
102. Mendivil-Perez M, Velez-Pardo C, Kosik KS et al (2019) iPSCs-derived nerve-like cells from familial Alzheimer's disease PSEN1 E280A reveal increased amyloid-beta levels and loss of the Y chromosome. *Neurosci Lett* 703:111–118. <https://doi.org/10.1016/j.neulet.2019.03.032>
103. Mahady L, Nadeem M, Malek-Ahmadi M et al (2019) HDAC2 dysregulation in the nucleus basalis of Meynert during the progression of Alzheimer's disease. *Neuropathol Appl Neurobiol* 45:380–397. <https://doi.org/10.1111/nan.12518>
104. Cao M, Li H, Zhao J et al (2019) Identification of age- and gender-associated long noncoding RNAs in the human brain with Alzheimer's disease. *Neurobiol Aging* 81:116–126. <https://doi.org/10.1016/j.neurobiolaging.2019.05.023>
105. Dunford A, Weinstock DM, Savova V et al (2017) Tumor-suppressor genes that escape from X-inactivation contribute to cancer sex bias. *Nat Genet* 49:10–16. <https://doi.org/10.1038/ng.3726>
106. Kaneko S, Li X (2018) X chromosome protects against bladder cancer in females via a KDM6A-dependent epigenetic mechanism. *Sci Adv* 4:eaar5598–eaar5598. <https://doi.org/10.1126/sciadv.aar5598>
107. Liu R, Kain M, Wang L (2012) Inactivation of X-linked tumor suppressor genes in human cancer. *Future Oncol* 8:463–481. <https://doi.org/10.2217/fon.12.26>
108. Buekers TE, Lallas TA, Buller RE (2000) Xp22.2-3 loss of heterozygosity is associated with germline BRCA1 mutation in ovarian cancer. *Gynecol Oncol* 76:418–422. <https://doi.org/10.1006/gyno.1999.5713>
109. Sirchia SM, Ramoscelli L, Grati FR et al (2005) Loss of the inactive X chromosome and replication of the active X in BRCA1-defective and wild-type breast cancer cells. *Cancer Res* 65:2139–2146. <https://doi.org/10.1158/0008-5472.CAN-04-3465>
110. Ren W, Medeiros N, Warneford-Thomson R et al (2020) Disruption of ATRX-RNA interactions uncovers roles in ATRX localization and PRC2 function. *Nat Commun* 11:2219. <https://doi.org/10.1038/s41467-020-15902-9>
111. Zhang H, Liao J, Zhang X et al (2019) Sex difference of mutation clonality in diffuse glioma evolution. *Neuro Oncol* 21:201–213. <https://doi.org/10.1093/neuonc/nyy154>
112. Smith RJ, Savoian MS, Weber LE, Park JH (2016) Ataxia telangiectasia mutated (ATM) interacts with p400 ATPase for an efficient DNA damage response. *BMC Mol Biol* 17:22. <https://doi.org/10.1186/s12867-016-0075-7>
113. Li CH, Haider S, Shiah Y-J et al (2018) Sex differences in cancer driver genes and biomarkers. *Cancer Res* 78:5527–5537. <https://doi.org/10.1158/0008-5472.CAN-18-0362>
114. Liu EM, Martinez-Fundichely A, Diaz BJ et al (2019) Identification of cancer drivers at CTCF insulators in 1,962 whole genomes. *Cell Syst* 8:446–455.e8. <https://doi.org/10.1016/j.cels.2019.04.001>
115. Matthews BJ, Waxman DJ (2020) Impact of 3D genome organization, guided by cohesin and CTCF looping, on sex-biased chromatin interactions and gene expression in mouse liver. *Epigenetics Chromatin* 13:30. <https://doi.org/10.1186/s13072-020-00350-y>

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