

HHS Public Access

Arch Dermatol Res. Author manuscript; available in PMC 2023 December 01.

Published in final edited form as:

Author manuscript

Arch Dermatol Res. 2022 December; 314(10): 953-959. doi:10.1007/s00403-021-02319-7.

Uncommon variants in FLG2 and TCHHL1 are associated with remission of atopic dermatitis in a large longitudinal US cohort

Ronald Berna, BS¹, Nandita Mitra, PhD², Ole Hoffstad, MA^{1,2}, Bradley Wubbenhorst³, Katherine L. Nathanson, MD³, David J. Margolis, MD PhD^{1,2}

¹ Department of Dermatology, Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania

² Department of Biostatistics, Epidemiology, and Informatics, Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania

³ Department of Medicine, Division of Translational Medicine and Human Genetics, Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania

Abstract

Atopic dermatitis (AD) is a relapsing inflammatory skin disease; filaggrin (FLG) variation has been consistently associated with its pathogenesis. Filaggrin-2 (FLG2) and trichohyalin-like-1 (TCHHL1) are members of the same protein family (S100 fused type proteins), are similar in structure to FLG, and may be involved in AD pathogenesis. We sought to evaluate the association between variation in FLG2, TCHHL1 and AD remission. We sequenced FLG2 and TCHHL1 in a longitudinal AD cohort using targeted capture based massively parallel sequencing. Association between individual alleles and AD remission was evaluated with generalized estimating equations for binary outcomes. Association between groups of alleles and AD remission was evaluated using a genetic algorithm to group alleles. We identified 2 loss-of-function (LoF) mutations in FLG2 (Ser2377Ter,Arg2207Ter) and 2 LoF mutations in TCHHL1 (Gln656Ter,Gln294Ter), none of which were associated with AD remission. Common (MAF>5%) alleles in FLG2 were similarly unassociated with AD. No common alleles in TCHHL1 were associated with AD remission after multiple testing correction. Among self-described whites, a group of 34 uncommon alleles in FLG2 were associated with increased AD remission (OR 7.64e17; 95%CI 4.41e17–1.32e18; adjusted p<1.0e-16). 12 uncommon alleles in TCHHL1 were associated with increased AD remission (OR 23.46; 95% CI 7.07-77.89; adjusted p=0.064). Among self-described African

Code availability: R code for this project is available upon request from the corresponding author.

Corresponding author: Ronald Berna BS, Perelman School of Medicine, University of Pennsylvania, ronald.berna@pennmedicine.upenn.edu, cell: 609-304-6024.

Publisher's Disclaimer: This AM is a PDF file of the manuscript accepted for publication after peer review, when applicable, but does not reflect post-acceptance improvements, or any corrections. Use of this AM is subject to the publisher's embargo period and AM terms of use. Under no circumstances may this AM be shared or distributed under a Creative Commons or other form of open access license, nor may it be reformatted or enhanced, whether by the Author or third parties. See here for Springer Nature's terms of use for AM versions of subscription articles: https://www.springernature.com/gp/open-research/policies/accepted-manuscript-terms

Conflicts of interest: David Margolis is a consultant for Pfizer with respect to studies of atopic dermatitis and serves on an advisory board for the National Eczema Association. No other authors state financial conflicts of interest with respect to this investigation

IRB approval: The PEER study was approved by the Institutional Review Board of the University of Pennsylvania, and written informed consent was obtained from all participants from the participant or his or her caregiver.

Americans, 13 uncommon *FLG2* alleles were associated with increased AD remission (OR 21.01; 95%CI 11.90–37.09; adjusted p<1.0e-16). No *TCHHL1* uncommon allele groups were associated with AD remission among African Americans. Our study supports the role of uncommon alleles in *FLG2* and *TCHHL1* in AD pathogenesis.

Keywords

atopic dermatitis; epidemiology; epidermal biology; epidermal barrier function; filaggrin 2; trichohyalin-like-1; population genetics; S100 fused type proteins

INTRODUCTION

Atopic dermatitis (AD) is a common chronic inflammatory skin disease which typically presents with red itchy patches on the flexural parts of the extremities [1,2,3,4]. AD is common, affecting up to 20% of children and 3% of adults in the industrialized world [5]. As with many common diseases, the pathogenesis of AD likely varies among individuals, and is influenced by both genetic and environmental factors [4,6,7].

Genetic studies of AD have suggested that both barrier dysfunction and immunodysregulation are key in disease pathogenesis [4,8]. Genetic variation in cytokines involved in the TH2 response, such as thymic stromal lymphopoietin (TSLP), and epidermal surface barrier proteins, primarily filaggrin (FLG), have been associated with variation in AD onset and persistence [5,8,15]. Of these, the most consistently associated and widely studied is FLG. Loss-of-function mutations in *FLG* lead decreased protein levels and are thought to produce a barrier defect which predisposes an individual to AD [17].

FLG protein belongs to a family of S100 fused-type proteins (SFTPs), many of which are part of the development of the skin's cornified envelope [5,9,23]. The genes cluster in a region of chromosome 1 called the epidermal differentiation complex (EDC) [5,9,23]. Other proteins in this family include filaggrin-2 and trichohyalin-like 1 protein [5,9,10,11]. Due to these proteins' structural and functional similarity to FLG, it is possible that variation in any of the SFTP genes could be associated with variation in skin barrier function and thus increase AD susceptibility and persistence [5,9,10,11,17,23].

Filaggrin-2 is an SFTP encoded for by the FLG2 gene. It is located very close to FLG. Previous studies suggest that the FLG2 protein is involved in maintenance of the epidermal barrier and cell-cell adhesion in the cornified layers of skin [11,13]. Decreased expression of FLG2 has been associated with a thinner epidermis; decreased levels of filaggrin-2 have been observed in the lesional skin of patients with AD and psoriasis [5,10,11,12]. Study of FLG2 variation in AD has been limited and largely involved studies in African Americans. In 2014, Margolis et al. identified three notable FLG2 variants in African American individuals, two of which were associated with increased AD persistence [10]. In contrast, in 2015, Pellerin et al. found numerous single nucleotide polymorphisms (SNPs) in FLG2, but found no variants associated with the presence of AD [5].

Trichohyalin-like 1 protein is encoded for by the *TCHHL1* gene. Similarly to filaggrin-2, trichohyalin-like 1 protein is an SFTP located near FLG in the EDC. This gene, and its protein's function, is not well-understood [18,19]. A previous investigation, from a small subset of the present cohort, identified loss of function (LoF) mutations in *TCHHL1*, but did not find an association with AD [20]. Another report identified mutations in *TCHHL1* associated with AD, but the sample size was small with no correction for multiple comparisons [21]. Due to *TCHHL1*'s proximity to FLG and limited prior studies, further studies are needed to determine if variation in *TCHHL1* is associated with AD.

Our goal was to examine genetic variation in two filaggrin-like proteins, previously known to have loss-of-function variants, in a longitudinal cohort of individuals with AD, and to better appreciate the relationship between variation in filaggrin-2, trichohyalin-like 1, and AD remission.

METHODS

Genetic data were obtained from a subset of the Pediatric Eczema Elective Registry (PEER) study for which DNA samples were available. Both the overall PEER cohort and the subset with genetic data have been previously described [7,8]. Analyses were conducted on both the overall cohort, and after stratification by ancestry. Determination of ancestry was based on self-report. Self-described ancestry previously has been determined to strongly correlate with genetic markers of ancestry within this cohort [15]. Written informed consent was obtained from all participants. This study was approved by the institutional review board of the University of Pennsylvania.

DNA was collected with Oragene DNA collection kits (DNA Genotek, Ottawa, Canada). Sequencing of *FLG2* and *TCHHL1* was conducted using targeted capture based Massively Parallel Sequencing (MPS) on PEER individuals with sufficient DNA. Sequencing read alignment and variant calling were accomplished as previously described [8,22].

Disease clearance was defined using a self-reported outcome of whether or not a child's skin was symptom-free during the previous six months. As children in PEER could be followed for up to 10 years, PEER participants can have multiple reports of this outcome over time. The association between these outcomes and individual SNPs were evaluated with generalized estimating equations (GEE) for binary outcomes, assuming an exchangeable working correlation structure with empirical standard errors [8]. This GEE model incorporates all individual reports to provide a single point estimate of the likelihood of AD healing over time, which we interpret as a measure of AD remission. An OR>1 indicates an association with greater AD remission, whereas an OR<1 indicates an association with greater AD remission, whereas an OR<1 indicates an association form the R package geepack 1.2–1.

Our analysis plan is demonstrated in Figure 1. *FLG2* and *TCHHL1* were evaluated independently, and analyses were conducted in both the pooled cohort and race-stratified cohorts. Association of variants differed depending on whether the SNPs had MAF 5%. MAF <5% is a commonly used cutoff for distinguishing common from low-frequency and

rare alleles, and has been used by our group in previous work [8,22,26]. We *a priori* decided that SNPs with MAF 5% would be independently assessed for association with AD remission. SNPs with MAF < 5% were considered to have insufficient data for reliable GEE estimates when evaluated independently, so these variables were grouped according to an algorithm previously developed and validated [22]. Thus, groups of variants with MAF < 5%, rather than individual variants, were evaluated for association with AD remission. All analyses were implemented in R, version 3.6.1.

RESULTS

We obtained genotyping of *FLG2* and *TCHHL1* on 705 PEER individuals; 326 were self-described African Americans and 379 were self-described whites. As of June 2020, over 12,000 follow-up surveys were completed within the PEER study, following participants for a mean of 102 months. Demographic information for the PEER genetic cohort is provided in Table 1.

163 unique variants were identified in *FLG2*, of which three were LoF. 120 unique variants were identified in the African American cohort, and 99 unique variants were identified in the white cohort. Of these, 23 variants in the pooled cohort had MAF 5%. Supplementary Table S1 presents the unadjusted association of these *FLG2* SNPs with AD persistence. No variants reached significance at a p-threshold of 0.05. Two LoF variants were identified in our cohort; they are presented in Table 2. The previously studied SNP rs12568784 was not associated with AD persistence (OR, 0.98, 95% CI, 0.82–1.16, unadjusted p-value=0.790). Independent GEE estimates for rs377218292 (NC_000001.10:g.152323643G>A) could not be obtained because of insufficient sample size. One stop-loss variant was identified in our sample, NP_001014364.1:p.Ter2392Ser (rs150529054). This variant was only present in three individuals, so it was not studied further.

60 unique variants were identified in *TCHHL1*; there were 41 unique variants in the African American cohort and 26 unique variants in the white cohort. Of these, only three variants had MAF 5% in the pooled population. Supplementary Table S2 presents the unadjusted association of these TCHHL1 SNPs with AD persistence. None were statistically significantly associated with AD remission after correction for multiple comparisons. Two loss-of-function mutations were identified; they are presented in Table 2. Neither of the LoF variants in *TCHHL1* (rs150014958 and rs61749316) were present at a high enough frequency to assess for association with AD persistence independently.

The rare variant algorithm was run independently on rare variants in *FLG2* and *TCHHL1*, and independently run on white and African American individuals. All p-values were corrected for 250,000 independent tests. Within the white population, thirty-four rare variants were identified in *FLG2*; having any of these was associated with increased AD remission (OR, 7.64e17, 95% CI 4.41e17–1.32e18, adjusted p-value < 1.0e-16). 26 individuals, representing 6.9% of the study population, carried at least one of these variants. These variants are presented in Table 3. Within the self-described African American population, 13 uncommon FLG2 alleles were associated with increased AD remission (OR 21.01; 95%CI 11.90–37.09; adjusted p<1.0e-16). These variants are presented in Table 3.

Page 5

In *TCHHL1*, in the white population, twelve rare variants were identified, which together trended towards association with increased AD remission (OR, 23.46, 95% CI 7.07–77.89, adjusted p-value 0.064). These variants were identified in 13 individuals, representing 3.4% of the study population. These variants are presented in Table 3. Within the African American population, no TCHHL1 uncommon allele groups were associated with AD remission.

DISCUSSION

FLG LoF variation is an important risk factor for AD and AD persistence. Several genes that code for filaggrin-like proteins (S100FTPs) cluster on chromosome 1 in the EDC region. However, there have been very few studies examining *FLG2* and *TCHHL1*, and prior studies focused on African American individuals. This report is the largest sequencing study of *FLG2* and *TCHHL1* in AD to date, and includes both African American and white individuals. This study is further distinguished by the use of *remission*, rather than presence of AD, as the primary outcome measure.

Taken together, our analyses of both *FLG2* and *TCHHL1* imply that LoF mutations in S100FTPs are not necessarily associated with AD persistence in either whites or African Americans. Since LoF mutations are generally thought to significantly decrease protein production, and since these LoF mutations in *FLG2* and *TCHHL1* are not associated with more persistent AD, it is tempting to speculate that the barrier function defect found in AD is relatively specific—that is, only specific types of skin-barrier alteration, namely those induced by filaggrin loss-of-function mutations, influence AD pathogenesis. Conversely, recent research into filaggrin suggests that filaggrin's contribution to keratohyalin granules, and the skin's barrier function, is dependent upon liquid-liquid phase separation, and that only relatively small amounts of protein are necessary for adequate granule formation and function [24,25]. Extending this hypothesis to *FLG2* and *TCHHL1*, it is possible that the identified loss-of-function mutations within these genes do not sufficiently decrease protein quantity to alter their function.

That said, missense variants have been found to be important in other genes associated with AD remission, so it is reasonable to hypothesize that other variation in *FLG2* and *TCHHL1* may be associated with AD remission [8]. We identified no relatively common variants (MAF 5%) in *FLG2* and *TCHHL1* associated with AD remission after correction for multiple comparisons. This finding, coupled with the analysis of LoF variants, suggests that *individual, common* variants in FLG2 and TCHHL1 are not significantly associated with AD remission.

Unlike common variants, uncommon alleles, particularly in *FLG2*, were associated with AD remission. We found 34 rare variants in the white population that, when analyzed as a group, are associated with increased AD remission. This represents the first association of *FLG2* variation with AD in white individuals, and the first association of *FLG2* non-LoF variants with AD remission. These variants represent 6.9% of the study population, a considerable number of individuals for a disease with a prevalence of up to 20% of the general population [5]. Similarly, in African Americans, 13 uncommon FLG2 alleles were associated with

increased AD remission. In *TCHHL1*, when analyzed together, 12 rare variants trended towards association with AD remission. These variants were only present in 3.4% of the study population, and the association did not reach statistical significance after adjustment for multiple testing. Future investigations, not subject to such a stringent correction factor, may identify a significant association here.

The primary limitation of our study was that we only studied African American and white individuals, so our results may not apply to AD in patients of differing ancestries. Further studies in diverse populations will be needed to confirm these findings. The incidence of several LoF variants in both genes was very small (<5%), so we were unable to evaluate independent associations between these genes and AD remission.

In this investigation, we report results from the largest sequencing study of *FLG2* and *TCHHL1* in whites with AD. In contrast with previous studies, we observed no common *FLG2* variants associated with AD severity. We also found no common variants in *TCHHL1* associated with AD severity. However, we did find that uncommon allele composites within both *FLG2* and *TCHHL1* were associated with AD remission, implying that these genes may be relevant in certain subsets of AD individuals. This study provides further insight into the genetics of AD, demonstrating that uncommon alleles within non-FLG skin barrier proteins may be involved in AD pathogenesis.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Funding:

This work was supported in part by a grant from the National Institutes for Health NIAMS R01-AR070873 (PI: Margolis). The PEER study is funded as the Atopic Dermatitis Registry by Valeant Pharmaceuticals International (PI: Margolis).

Data availability:

The PEER data (source) is not currently publically available. The PEER study is an ongoing study sponsored by Valeant in response to a post marketing commitment with the FDA. A BED file of the probes used for targeted capture is available upon request from the corresponding author.

REFERENCES

- 1. Bieber T Atopic dermatitis. N Engl J Med. 2008; 358(14):1483-1494. [PubMed: 18385500]
- 2. Abramovits W Atopic dermatitis. J Am Acad Dermatol. 2005;53(1)(suppl 1):S86–S93. [PubMed: 15968268]
- Akdis CA, Akdis M, Bieber T, Bindslev-Jensen C, Boguniewicz M, Eigenmann P, et al. Diagnosis and treatment of atopic dermatitis in children and adults: European Academcy of Allerology and Clinical Immunology/American Academy of Allergy, Asthma and Immunology.PRACTALL Consensus Report. Allergy. 2006; 61:969–987. [PubMed: 16867052]
- 4. Leung DY, Bieber T. Atopic dermatitis. Lancet. 2003; 361(9352):151-160. [PubMed: 12531593]

- Pellerin L, Henry J, Hsu C-Y, Balica S, Jean-Decoster C, Mechin M-C, et al. Defects of filaggrin-like proteins in both lesional and nonlesional atopic skin. J Allergy Clin Immunol. 2013;131(4):1094–1102. [PubMed: 23403047]
- 6. Berna R, Mitra N, Hoffstad O, Wan J, Margolis DJ. Identifying phenotypes of atopic dermatitis in a longitudinal US cohort using unbiased statistical clustering. J Invest Dermatol. Article in Press.
- Margolis JS, Abuabara K, Bilker W, Hoffstad O, Margolis DJ. Persistence of mild to moderate atopic dermatitis. JAMA Dermatol. 2014a; 150(6):593–600. [PubMed: 24696036]
- Berna R, Mitra N, Lou C, Hoffstad O, Wubbenhorst B, D'Andrea K, et al. Thymic Stromal Lymphopoietin and IL7R Variants are Associated with Persistent Atopic Dermatitis. J Invest Dermatol. Article in Press.
- Wu Z, Latendorf T, Meyer-Hoffert U, and Schroder J-M. Identification of trichohyalin-like 1, an S100 fused-type protein selectively expressed in hair follicles. J Invest Dermatol. 2011; 131(8):1761–1763. [PubMed: 21614011]
- Margolis DJ, Gupta J, Apter AJ, Ganguly T, Hoffstad O, Papadopoulos M, et al. Filaggrin-2 variation is associated with more persistent atopic dermatitis in African American subjects. J Invest Dermatol. 2014b; 133(3):784–789.
- Pendaries V, Le Lamer M, Cau L, Hansmann B, Malaisse J, Kezic S, et al. In a three-dimensional reconstructed human epidermis filaggrin-2 is essential for proper cornification. Cell Death Dis. 2015; 6(2):e1656. [PubMed: 25695608]
- Makino T, Mizawa M, Yamakoshi T, Takaishi M, and Shimizu T. Expression of filaggrin-2 protein in the epidermis of human skin diseases: a comparative analysis with filaggrin. Biochem Biophys Res Comm. 2014; 449(1):100–106. [PubMed: 24813994]
- Mohamad J, Sarig O, Godsel LM, Peled A, Malchin N, Bochner R, et al. Filaggrin 2 deficiency results in abnormal cell-cell adhesion in the cornified cell layers and causes peeling skin syndrome type A. J Invest Dermatol. 2018; 138(8):1736–1743. [PubMed: 29758285]
- Bolling MC, Jan SZ, Pasmooij AMG, Lemmink HH, Franke LH, Yenamandra VK, et al. Generalized ichthyotic peeling skin syndroem due to FLG2 mutations. J Invest Dermatol. 2018; 138(8):1881–1884. [PubMed: 29505760]
- Lou C, Mitra N, Wubbenhorst B, D'Andrea K, Hoffstad O, Kim BS, et al. Association between fine mapping thymic stromal lymphopoietin and atopic dermatitis onset and persistence. Ann Allergy Asthma Immunol. 2019;123(6):595–601. [PubMed: 31491540]
- Li H, Durbin R. Fast and accurate long-read alignment with Burrows-Wheeler Transform. Bioinformatics. 2010; 26(5):589–595. [PubMed: 20080505]
- Margolis DJ, Gupta J, Apter AJ, Hoffstad O, Papadopoulos M, Rebbeck TR, et al. Exome sequencing of filaggrin and related genes in African-American children with atopic dermatitis. J Invest Dermatol. 2014c; 134:2272–2274. [PubMed: 24608987]
- Yamakoshi T, Makino T, Rehman MU, Yoshihisa Y, Sugimori M, and Shimizu T. Trichohyalin-like 1 protein, a member of fused S100 proteins, is expressed in nromal and pathologic human skin. Biochem Biophys Res Commun. 2013; 432:66–72. [PubMed: 23376073]
- Wu Z, Latendorf T, Meyer-Hoffert U, and Schroder J-M. Identification of trichohyalin-like 1, an S100 fused-type protein selectively expressed in hair follicles. J Invest Dermatol. 2011; 131:1761– 1763. [PubMed: 21614011]
- Margolis DJ, Gupta J, Apter AJ, Hoffstad O, Papadopoulos M, Rebbeck TR, et al. Exome sequencing of filaggrin and related genes in African-American children with atopic dermatitis. J Invest Dermatol. 2014c; 134:2272–2274. [PubMed: 24608987]
- Pigors M, Common JEA, Wong XFC, Malik S, Scott CA, Tabarra N, et al. Exome sequencing and rare variant analysis reveals multiple filaggrin mutations in Bangladeshi families with atopic eczema and additional risk genes. J Invest Dermatol. 2018; 138:2674–2677. [PubMed: 29857066]
- 22. Berna R, Mitra N, Hoffstad O, Wubbenhorst B, Nathanson K, Margolis DJ. Using a machine learning approach to identify low-frequency and rare filaggrin alleles associated with remission of atopic dermatitis. JID Innov. Article in Press.
- 23. Mischke D, Korge BP, Marenholz I, Volz A, Ziegler A. Genes encoding structural proteins of epidermal cornification and S100 calcium-binding proteins form a gene complex ("Epidermal

Differentiation Complex") on human chromosome 1q21. J Invest Dermatol. 1996; 106(5):989–992. [PubMed: 8618063]

- 24. Quiroz FG, Fiore VF, Levorse J, Polak L, Wong E, Pasolli HA, et al. Liquid-liquid phase separation drives skin barrier formation. Science. 2020;367(6483):eaax9554.
- 25. Chua Avecilla AR, Quiroz FG, Cracking the skin barrier: liquid-liquid phase separation shines under the skin, JID Innovations (2021)
- 26. Bomba L, Walter K, Soranzo N. The impact of rare and low-frequency genetic variants in common disease. Genome Biol. 2017 Apr 27;18(1):77. [PubMed: 28449691]



Figure 1: Schematic of analysis flow.

Participant Demographics

~	
Ð	
q	
Ta	

	Whole cohort, n(%)	African American, n(%)	White, n (%)
Number	705	326	379
Age of AD onset, mean (SD)	1.89 (2.65)	2.14 (2.85)	1.84 (2.65)
Sex: male, n (%)	332 (47.1)	136 (42.0)	196 (51.7)
Asthma, n (%)	382 (54.1)	182 (56.2)	200 (52.8)
Seasonal Allergies, n (%)	494 (70.1)	228 (70.4)	266 (70.2)
		-	

-
_
_
<u> </u>
–
_
\sim
\mathbf{O}
_
_
_
~
<u> </u>
$\boldsymbol{\omega}$
B
B
an
anu
anu
anu
anus
anus
anus
anusc
anusci
anuscr
anuscri
anuscri
anuscrip
anuscrip
anuscript

÷	
H	
· Ĕ	
S	
.н	
E .	
<u>e</u>	
~	
Ц	
\triangleleft	
-	
E	
.2	
-	
g	
·H	
at	
·H	
Ř	
Š	
JS 1	
·=	
g	
ţ	
Ч	
ã	
а	
<u>_</u>	
Ħ	
Η	
IHH	
CHHL	
TCHHL	
d TCHHL	
nd TCHHL	
and TCHHL	
2 and TCHHL	
32 and TCHHL	
JG2 and TCHHL	
FLG2 and TCHHL	
FLG2 and TCHHL	
in FLG2 and TCHHL	
s in FLG2 and TCHHL	
nts in FLG2 and TCHHL	
ants in FLG2 and TCHHL	
riants in FLG2 and TCHHL	
ariants in FLG2 and TCHHL	
variants in FLG2 and TCHHL	
n variants in FLG2 and TCHHL	
ion variants in FLG2 and TCHHL	
tion variants in FLG2 and TCHHL	
nction variants in FLG2 and TCHHL	
unction variants in FLG2 and TCHHL	
function variants in FLG2 and TCHHL	
of function variants in FLG2 and TCHHL	
of function variants in FLG2 and TCHHL	
ss of function variants in FLG2 and TCHHL	
oss of function variants in FLG2 and TCHHL	
Loss of function variants in FLG2 and TCHHL	

			Pooled cohort (n = 705)			African American (n =	= 379)		White (n = 379)		
Gene	RSID	Location	Odds of Clearing (95% CI)	raw p value	u	Odds of Clearing (95% CI)	raw p value	u	Odds of Clearing (95% CI)	raw p value	u
FLG2	rs12568784*	NC_000001.10: g.152323132G>T	0.977 (0.822,1.161)	0.790	281	0.944 (0.768,1.16)	0.583	161	1.092 (0.792,1.507)	0.592	120
FLG2	rs377218292	NC_000001.10: g.152323643G>A	***	***	1	***	***	1	***	***	0
TCHHL1	rs150014958	NP_001008536.1: p.Gln656Ter	***	***	12	***	***	4	***	***	×
TCHHL1	rs61749316**	NP_001008536.1: p.Gln294Ter	***	***	16	***	***	9	***	***	10
*											

^{*}Previously identified in Margolis et al. 2014b.

** Previously identified in Margolis et al. 2014c.

*** Could not be estimated, or insufficient sample size for estimation. Author Manuscript

Author Manuscript

Berna	et	al.	

FLG2, white	8		FLG2, Afric	an Americans		TCHHL1, w	hites	
Location	Nucleotide change	Amino acid change	Location	Nucleotide change	Amino acid change	Location	Nucleotide change	Amino acid change
152328471	C>T	p.G597G	152322250	G>A	intronic	152057347	G>A	
152329677	G>A	p.D195D	152322365	T>G	intronic	152059461	G>T	p.Q233K
152331902	G>C		152323269	A>G	p.H2331H	152057769	A>G	H797H
152331264	G>A	p.L33L	152324043	A>G	p.H2073H	152058358	A>G	p.P600P
152330789	G>A		152325368	G>A	p.H1632Y	152058563	C>T	p.G532E
152329534	C>T	p.G243E	152325410	C>T	p.G1618R	152058633	A>T	p.S509T
152329530	C>G	p.L244F	152325444	A>G	p.H1606H	152058648	G>T	p.P504T
152329370	A>G	p.C298R	152325742	C>T	p.S1507N	152059278	G>A	p.Q294X
152328924	T>C	p.V446V	152326154	G>A	p.H1370Y	152059570	T>A	p.1196I
152328475	T>C	p.H596R	152326696	A>G	p.F1189S	152059638	C>T	p.E174K
152328308	A>G	p.S652P	152327295	C>G	р.Q989Н	152061454	C>T	
152328190	G>A	p.S691F	152330196	C>T	intronic	152061533	C>T	
152327955	G>A	p.S769S	152330801	C>T	intronic			
152326902	C>T	p.S1120S						
152326715	C>A	p.V1183L						
152326620	T>C	p.Q1214Q						
152325965	G>A	p.H1433Y						
152325911	C>T	p.G1451R						
152325766	G>T	p.S1499Y						
152325522	A>G	p.T1580T						
152325455	C>T	p.G1603R						
152324325	A>G	p.A1979A						
152324316	G>T	p.H1982Q						
152324315	A>G	p.Y1983H						
152324312	G>C	p.P1984A						
152324311	G>C	p.P1984R						

Manuscript	Author	
script	Manus	
	script	

cript	
Auth	
nor Manuscript	

Au	
thor	
Mai	
nuso	
cript	

FLG2, white	es		FLG2, Afric	an Americans		TCHHL1, w	hites	
Location	Nucleotide change	Amino acid change	Location	Nucleotide change	Amino acid change	Location	Nucleotide change	Amino acid change
152324146	T>G	р.Н2039Р						
152324114	C>T	p.A2050T						
152324109	A>G	p.H2051H						
152324106	G?A	p.G2052G						
152323176	C>T	p.G2362G						
152322465	C>T							
152322324	C>T							
152322292	G>A							