

Brief Communication



Performance of STANDARD™ M10 SARS-CoV-2 Assay for the Diagnosis of COVID-19 from a Nasopharyngeal Swab

Sin Young Ham ¹, Hyeonju Jeong ¹, Jongtak Jung ¹, Eu Suk Kim ¹,
Kyoung Un Park ², Hong Bin Kim ¹, Jeong Su Park ^{2,*}, and Kyoung-Ho Song ^{1,*}

¹Department of Internal Medicine, Seoul National University Bundang Hospital, Seoul National University College of Medicine, Seongnam, Korea

²Department of Laboratory Medicine, Seoul National University Bundang Hospital, Seoul National University College of Medicine, Seongnam, Korea

OPEN ACCESS

Received: Apr 17, 2022

Accepted: May 2, 2022

Published online: May 13, 2022

Corresponding Authors:

Kyoung-Ho Song, MD, PhD

Department of Internal Medicine, Seoul National University Bundang Hospital, 82, Gumi-ro, 173 Beon-gil, Bundang-gu, Seongnam 13620, Gyeonggi-do, Korea.

Tel: +82-31-787-7044

Fax: +82-31-787-4052

Email: khsongmd@gmail.com

khsongmd@snu.ac.kr

Jeong Su Park, MD, PhD

Department of Laboratory Medicine, Seoul National University Bundang Hospital, 82, Gumi-ro, 173 Beon-gil, Bundang-gu, Seongnam 13620, Gyeonggi-do, Korea.

Tel: +82-31-787-7693

Fax: +82-31-787-4015

Email: mdmicrobe@gmail.com

*These authors contributed equally to the work.

Copyright © 2022 by The Korean Society of Infectious Diseases, Korean Society for Antimicrobial Therapy, and The Korean Society for AIDS

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<https://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

ABSTRACT

The STANDARD™ M10 severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) assay (M10 assay) (SD Biosensor Inc., Suwon, Korea) is a rapid, fully-automated, cartridge-type molecular diagnostic assay that detects SARS-CoV-2 RNA using primers and probes for each target gene (ORF1ab gene, E gene). This study evaluated its performance by assessing its concordance with the approved SARS-CoV-2 real-time PCR assay. Tests were performed on 80 nasopharyngeal samples. The sensitivity and specificity of the M10 assay were 100%. The M10 assay effectively diagnosed SARS-CoV-2 infection, and it was comparable to the approved SARS-CoV-2 real-time PCR assay. It is a viable point-of-care test due to its short turnaround time.

Keywords: COVID-19; SARS-CoV-2 RT-PCR testing; COVID-19 nucleic acid testing

The current diagnostic modalities for coronavirus disease 2019 (COVID-19) include the reverse transcription-polymerase chain reaction (RT-PCR) assay, virus culture, antigen test, and antibody test. A real-time RT-PCR test, using upper respiratory tract specimens, is recommended as a confirmatory test [1]. However, the real-time RT-PCR assay is limited due to its long turnaround time. Moreover, it requires skilled personnel and specialized equipment. Meanwhile, the COVID-19 antigen test is inexpensive with a short turnaround time. However, the antigen test is also limited due to its low sensitivity, which varies depending on the viral load of the specimen [2-4]. Thus, a simple, rapid, and sensitive clinical test to detect COVID-19 should be developed.

The STANDARD™ M10 severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) assay (M10 assay) (SD Biosensor Inc., Suwon, Korea) automates and integrates sample preparation, nucleic acid extraction, and amplification, and detects target sequences using molecular diagnostic assays. The M10 assay utilizes a cartridge, containing primers and probes for each target gene (ORF1ab gene, E gene) to detect SARS-CoV-2 RNA. Since all processes are fully automated and performed in one cartridge, cross-contamination is minimized, and the test is less affected by test proficiency. In addition, the test results can be obtained within 1 hour. Thus, it can be used as a point-of-care test. In the United States, the cartridge-type Gene Xpert Xpress SARS-CoV-2 commercial kit (Cepheid, Sunnyvale, CA, USA) has been approved

ORCID iDs

Sin Young Ham 
<http://orcid.org/0000-0002-3243-540X>
 Hyeonju Jeong 
<http://orcid.org/0000-0002-2072-5469>
 Jongtak Jung 
<https://orcid.org/0000-0003-3497-0796>
 Eu Suk Kim 
<https://orcid.org/0000-0001-7132-0157>
 Kyoung Un Park 
<https://orcid.org/0000-0002-2402-7633>
 Hong Bin Kim 
<https://orcid.org/0000-0001-6262-372X>
 Jeong Su Park 
<https://orcid.org/0000-0001-5149-1362>
 Kyoung-Ho Song 
<https://orcid.org/0000-0002-4517-3840>

Funding

This work was supported by a research fund from the manufacturer (SD Biosensor Inc., Suwon, Korea).

Ethics Statement

This clinical trial was approved by the IRB of Seoul National University Bundang Hospital (IRB number: B-2004/607-105). The IRB approved a request to waive the informed consent.

Conflict of Interest

KUP is associate editor and HBK is editorial board of Infect Chemother; however, they did not involve in the peer reviewer selection, evaluation, and decision process of this article. Otherwise, no potential conflicts of interest relevant to this article was reported.

Author Contributions

Conceptualization: JSP, KHS. Data curation: SYH, JSP, KUP. Formal analysis: SYH, KHS, ESK, KUP, HBK. Investigation: SYH, HJ, JJ. Methodology: JSP, KHS. Supervision: JSP, KHS. Writing - Original draft: SYH, JSP, KHS. Writing - review & editing: JSP, KHS, ESK, KUP, HBK.

for diagnostic use [5, 6]. This study aimed to evaluate the performance and clinical viability of the M10 assay in comparison to the approved STANDARD™ M nCoV Real-Time Detection kit (SD Biosensor Inc.) [7].

Nasopharyngeal swabs for evaluation were obtained from Seoul National University Bundang Hospital (SNUBH) from February 1, 2020, to May 14, 2020. To calculate the sample size, we used Buderer's formula [8]. The sample size calculated with a prevalence of 50% (n/2 positive samples + n/2 negative samples), $\alpha = 0.05$ and marginal error = 0.1 was 70. Taking into consideration the possibility of inappropriate samples for tests, a total of 80 nasopharyngeal swabs were collected. Forty positive samples were collected from patients hospitalized for COVID-19 infection, and all patients were symptomatic. Samples were collected at 14.725 ± 8.45 (mean ± SD) days from the onset of symptoms. Each patient was confirmed to be COVID-19-positive by SARS-CoV-2 RT-PCR performed on the day of sample acquisition. Forty negative samples were obtained from patients who visited the emergency room in SNUBH due to suspicion of COVID-19 infection, but were confirmed as negative by SARS-CoV-2 RT-PCR. The collected samples were given unique identification codes and were randomly assigned to be deidentified.

The STANDARD™ M nCoV Real-Time Detection kit (M nCoV assay, SD Biosensor Inc.) and M10 assay were performed on 80 collected samples. Both tests were performed according to the manufacturer's instructions. All collected samples were stored at -70 °C prior to testing. Testing was conducted from December 9, 2020 to December 10, 2020.

The sensitivity and specificity of M10 assay were evaluated to assess its diagnostic effectiveness. For the binomial confidence interval, the Clopper-Pearson method was used [9].

Both assays were performed on 80 samples, and the sensitivity and specificity of M10 assay were 100%, respectively (Table 1). The overall test results are presented in Supplementary Table 1. In one sample (case number 59), only the E gene was detected in both tests. A nasopharyngeal swab collected on the same day from the same patient showed a positive result when tested by The PowerChek™ 2019-nCoV Real-time PCR Kit (Kogene Biotech, Seoul, Korea); hence, this test result was considered as a positive result.

In this study, the M10 assay exhibited a 100% overall agreement with the approved STANDARD™ M nCoV Real-Time assay for detecting SARS-CoV-2. In addition, the M10 assay has an advantage in that it has a short turnaround time of less than 1 hour. Although molecular diagnostic assays on similar platforms are commercially available [6, 10], the M10 assay is superior in that it amplifies two target genes at once, the manual hands-on time is short, and it does not require additional equipment installation. However, its limitations

Table 1. Sensitivity and specificity of STANDARD™ M10 SARS-CoV-2 assay compared with STANDARD™ M nCoV real-time detection kit

	STANDARD™ M nCoV real-time detection kit		
	Positive	Negative	Total
STANDARD™ M10 SARS-CoV-2			
Positive	40	0	40
Negative	0	40	40
Total	40	40	80
Sensitivity (95% CI)	40/40, 100% (91.19 - 100.00%)		
Specificity (95% CI)	40/40, 100% (91.19 - 100.00%)		

SARS-CoV-2, severe acute respiratory syndrome coronavirus-2; CI, confidence interval.

include a low sample throughput and high cost. Thus, the M10 assay can be used in places where skilled personnel or real-time PCR equipment is limited. Moreover, it is a viable point-of-care test for emergency cases, requiring rapid screening.

The M10 assay noted an excellent specificity (100%) and sensitivity (100%). However, this study has a few limitations. First, the number of samples included was small. Second, in one of 40 positive samples, only the E gene was amplified in the test. Clinically, it corresponds to “indeterminate” result and requires retesting, but a retest was not performed due to insufficient sample volume.

In summary, the M10 assay effectively diagnosed SARS-CoV-2 infection, and its performance was comparable to the approved SARS-CoV-2 real-time PCR assay. It is a viable point-of-care test due to its short turnaround time.

SUPPLEMENTARY MATERIAL

Supplementary Table 1

Test results of STANDARD™ M nCoV Real-Time Detection kit and STANDARD™ M10 SARS-CoV-2 assay

[Click here to view](#)

REFERENCES

1. Hong KH, Lee SW, Kim TS, Huh HJ, Lee J, Kim SY, Park JS, Kim GJ, Sung H, Roh KH, Kim JS, Kim HS, Lee ST, Seong MW, Ryoo N, Lee H, Kwon KC, Yoo CK. Guidelines for laboratory diagnosis of coronavirus disease 2019 (COVID-19) in Korea. *Ann Lab Med* 2020;40:351-60.
[PUBMED](#) | [CROSSREF](#)
2. Oh SM, Jeong H, Chang E, Choe PG, Kang CK, Park WB, Kim TS, Kwon WY, Oh MD, Kim NJ. Clinical application of the standard Q COVID-19 Ag test for the detection of SARS-CoV-2 infection. *J Korean Med Sci* 2021;36:e101.
[PUBMED](#) | [CROSSREF](#)
3. Turcato G, Zaboli A, Pfeifer N, Ciccariello L, Sibilio S, Tezza G, Ausserhofer D. Clinical application of a rapid antigen test for the detection of SARS-CoV-2 infection in symptomatic and asymptomatic patients evaluated in the emergency department: a preliminary report. *J Infect* 2021;82:e14-6.
[PUBMED](#) | [CROSSREF](#)
4. Pray IW, Ford L, Cole D, Lee C, Bigouette JP, Abedi GR, Bushman D, Delahoy MJ, Currie D, Cherney B, Kirby M, Fajardo G, Caudill M, Langolf K, Kahrs J, Kelly P, Pitts C, Lim A, Aulik N, Tamin A, Harcourt JL, Queen K, Zhang J, Whitaker B, Browne H, Medrzycki M, Shewmaker P, Folster J, Bankamp B, Bowen MD, Thornburg NJ, Goffard K, Limbago B, Bateman A, Tate JE, Gieryn D, Kirking HL, Westergaard R; CDC COVID-19 Surge Laboratory Group. Performance of an antigen-based test for asymptomatic and symptomatic SARS-CoV-2 testing at two university campuses - Wisconsin, September-October 2020. *MMWR Morb Mortal Wkly Rep* 2021;69:1642-7.
[PUBMED](#) | [CROSSREF](#)
5. Stevens B, Hogan CA, Sahoo MK, Huang C, Garamani N, Zehnder J, Kurzer J, Pinsky BA. Comparison of a point-of-care assay and a high-complexity assay for detection of SARS-CoV-2 RNA. *J Appl Lab Med* 2020;5:1307-12.
[PUBMED](#) | [CROSSREF](#)
6. Goldenberger D, Leuzinger K, Sogaard KK, Gosert R, Roloff T, Naegele K, Cuénod A, Mari A, Seth-Smith H, Rentsch K, Hinić V, Hirsch HH, Egli A. Brief validation of the novel GeneXpert Xpress SARS-CoV-2 PCR assay. *J Virol Methods* 2020;284:113925.
[PUBMED](#) | [CROSSREF](#)

7. Hur KH, Park K, Lim Y, Jeong YS, Sung H, Kim MN. Evaluation of four commercial kits for SARS-CoV-2 real-time reverse-transcription polymerase chain reaction approved by emergency-use-authorization in Korea. *Front Med (Lausanne)* 2020;7:521.
[PUBMED](#) | [CROSSREF](#)
8. Buderer NM. Statistical methodology: I. Incorporating the prevalence of disease into the sample size calculation for sensitivity and specificity. *Acad Emerg Med* 1996;3:895-900.
[PUBMED](#) | [CROSSREF](#)
9. Clopper CJ, Pearson ES. The use of confidence or fiducial limits illustrated in the case of the binomial. *Biometrika* 1934;26:404-13.
[CROSSREF](#)
10. Basu A, Zinger T, Inglima K, Woo KM, Atie O, Yurasits L, See B, Aguero-Rosenfeld ME. Performance of abbot ID now COVID-19 rapid nucleic acid amplification test using nasopharyngeal swabs transported in viral transport media and dry nasal swabs in a New York City academic institution. *J Clin Microbiol* 2020;58:e01136-20.
[PUBMED](#) | [CROSSREF](#)