

Review Article

Corresponding Author

Yusuke Nishimura

https://orcid.org/0000-0001-9689-0586

Department of Neurosurgery, Nagoya University School of Medicine, 65 Tsurumai-cho, Showa-ku, Nagoya 466-8560, Japan Email: yusuken0411@med.nagoya-u.ac.jp

Received: February 28, 2022 Accepted: March 9, 2022



This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (https://creativecommons.org/licenses/by-nc/4.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Copyright © 2022 by the Korean Spinal Neurosurgery Society

Recent Molecular and Genetic Findings in Intramedullary Spinal Cord Tumors

Yoshitaka Nagashima¹, Yusuke Nishimura¹, Kaoru Eguchi², Junya Yamaguchi¹, Shoichi Haimoto³, Fumiharu Ohka¹, Masakazu Takayasu⁴, Ryuta Saito¹

- ¹Department of Neurosurgery, Nagoya University School of Medicine, Nagoya, Japan
- ²Department of Neurosurgery, National Hospital Organization Nagoya Medical Center, Nagoya, Japan
- ³Department of Neurosurgery, Aichi Cancer Center Hospital, Nagoya, Japan
- ⁴Department of Neurosurgery, Inazawa Municipal Hospital, Aichi, Japan

The study of genetic alterations and molecular biology in central nervous system (CNS) tumors has improved the accuracy of estimations of patient prognosis and tumor categorization. Therefore, the updated 2021 World Health Organization (WHO) classification includes various diagnostic genes, molecules, and pathways for diagnosis, as well as histological findings. These findings are expected both to have diagnostic applications and to facilitate new targeted therapies that target tumor-specific genetic changes and molecular biology. Intramedullary spinal cord tumors (IMSCTs) are rare CNS tumors that are difficult to treat because they occur in eloquent areas. Although the genetic underpinnings of IMSCTs remain unclear compared to their intracranial counterparts, the genetic characteristics of these tumors are gradually being revealed. Here, we describe the major changes in the new 2021 WHO classification and review the major types of IMSCTs, with an emphasis on their clinical features and genetic alterations.

Keywords: Intramedullary spinal cord tumor, Astrocytoma, Ependymoma, Hemangioblastoma, Genomics, Genetics

INTRODUCTION

Central nervous system (CNS) tumors, which arise from the brain and spinal cord, are classified into over 120 entities. World Health Organization (WHO) Classification of Tumors of the Central Nervous System has been revised on several occasions since its first publication in 1979. In the prior version of the WHO classification, CNS tumors were classified based on their histological findings with a diagnosis term and WHO grade (I–IV) embedded. However, recent comprehensive genomic studies revealed that an integrated diagnosis based on pathological and molecular findings more accurately predicted the prognosis than a diagnosis based on pathological findings alone. An integrated diagnosis with molecular information was first presented in the 2016 WHO classification, ^{2,3} and the latest version was published in 2021. ⁴

Although intramedullary spinal cord tumors (IMSCTs) are included in the category of CNS tumors, they are much less common than intracranial tumors and their molecular and genetic studies considerably lag behind those of intracranial tumors. IMSCTs account for 5%–10% of all spinal cord tumors, with ependymomas and astrocytomas comprising 80%–90% of IMSCTs. Fernangioblastomas are the third most common IMSCTs, after ependymomas and astrocytomas. Recent advances in molecular and genetic research have revealed that some infratentorial tumors, including IMSCTs, have different genetic characteristics from supratentorial tumors. In this article, we discuss the major changes in the 2021 WHO classification of CNS tumors and present a narrative review of the literature on recent molecular and genetic analyses that characterize the major types of IMSCTs.

Informed consent was obtained from all subjects involved in

the study. This study was approved by the Ethics Committee or Institutional Review Board of Nagoya University (2012-0067-19).

MAJOR CHANGES IN THE 2021 WHO CLASSIFICATION OF CENTRAL NERVOUS SYSTEM TUMORS

CNS tumors were conventionally classified based on histological findings. Molecular parameters were incorporated into the classification of CNS tumors (i.e., an integrated diagnosis) in the 2016 WHO classification of CNS tumors and its update in 2021.^{2,3} The process of an integrated diagnosis for CNS tumors is still developing in the 2021 WHO classification of CNS tumors, where various key diagnostic genes, molecules, pathways are applied to define entities, and the nomenclature was made more consistent and simpler by only including the location, age, or genetic modifiers of clinical utility. Modifier terms like "anaplastic" are not used because "grading within types" is applied. Thus, terms such as "diffuse astrocytoma, IDH-mutant," "anaplastic astrocytoma, IDH-mutant," and "glioblastoma, IDHmutant" in the 2016 WHO classification of CNS tumors are now simply classified as "astrocytoma, IDH-mutant grade 2, 3, and 4." Adult-type diffuse gliomas, which were divided into 15 entities in 2016, have been classified into only 3 types in 2021. To standardize the 2021 WHO classification of CNS tumors with other non-CNS tumor classifications, the term "type" and "subtype" have been adopted instead of "entity" and "variant," respectively. For example, meningioma is considered a single type in the 2021 WHO classification of CNS tumors with 15 subtypes.

The WHO CNS tumor classification has adopted an original grading system in line with the non-CNS tumor classification to facilitate grading across different entities. ¹² In the prior WHO classification of CNS tumors, a CNS was given one diagnosis name and automatically assigned to one WHO grade. In the 2021 classification, CNS tumors are graded within types and the grading is written using Arabic numerals, not Roman. For example, astrocytoma is assigned to grade 2, 3, or 4 based on its histological and genetic findings in 2021. Nonetheless, the 2021 WHO classification has generally retained the ranges of grades used for tumor types in prior editions (grade I: curable if surgically removed, grade IV: highly malignant, leading to death without effective therapy). Notably, although grading was based on histological findings in the prior version of the WHO classification of CNS tumors, molecular findings are applied as biomark-

ers to assign grades within the tumor type in the 2021 WHO classification of CNS tumors. The WHO grading of CNS tumors is no longer a histological system. For example, *CDKN2A/B* in IDH-mutant astrocytoma and the *TERT* promoter mutation, *EGFR* amplification, and +7/-10 copy number changes in IDH-wildtype astrocytoma are enough information to assign grade 4 even if the tumor is histologically low-grade.¹³

THE CLINICAL FEATURES AND GENETIC FINDINGS OF INTRAMEDULLARY SPINAL CORD TUMORS

1. Spinal Astrocytomas

Astrocytomas are the second most common IMSCTs observed in adults, but the most common in children. Several recent studies have demonstrated a clear prognostic difference between high-grade (WHO grade 3, 4) and low-grade spinal astrocytomas (mainly pilocytic astrocytoma WHO grade 1 and diffuse astrocytoma WHO grade 2). Therefore, we discuss spinal high-grade and low-grade astrocytomas separately, with descriptions of the important genetic mutations in each category.

1) High-grade astrocytomas

Gross total resection (GTR) is almost impossible for most high-grade astrocytomas, and subtotal resection or biopsy followed by radiation and chemotherapy is performed as the standard treatment in practical settings. Even with such treatment, the prognosis of these tumors is unfavorable (Fig. 1). There is a survival advantage with an excision extent of 78% or higher in intracranial glioblastoma, comparable to total excision. ¹⁴ Even in spinal high-grade astrocytomas, the extent of excision might be correlated with the prognosis, although the exact threshold of the extent of excision that improves the prognosis has yet to be clarified. ^{15,16} The benefits of adjuvant radiation and chemotherapy are controversial, although they are often performed for patients with residual tumors. ^{9,16-18}

(1) H3 K27M

The H3 K27M mutation of the *H3F3A* gene is the most important and well-established genetic mutation in high-grade astrocytomas (Table 1). Histones are major proteins that provide structural support for chromosomes. The histone tail (the N-terminal of the histone protein) plays an important role in the transcriptional regulation of DNA. Histone tail amino acids undergo various chemical modifications such as acetylation

and methylation. The major histone is H3.1 in humans, and the histone subtypes referred to as histone variants, such as H3.2 and H3.3, are functionally encoded by a different gene. The K27M

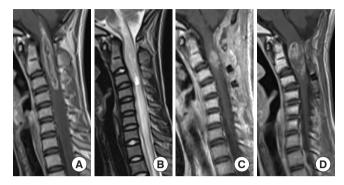


Fig. 1. Diffuse midline glioma, H3K27-altered. (A) A 15-year-old male patient with neck pain and limb weakness for 2 months presented with an intramedullary tumor with ring enhancement on gadolinium-enhanced T1-weighted magnetic resonance imaging (MRI) from the medulla oblongata to the C4 level on MRI. (B) T2-weighted MRI showed extensive edematous changes. (C) The tumor was surgically removed, although MRI one month later showed residual tumor. A histopathological examination showed anaplastic astrocytoma; however, the H3 K27M mutation was found, and the final diagnosis was diffuse midline glioma, H3K27-altered. After surgery, the patient underwent radiotherapy and chemotherapy. (D) Fifteen months after the first surgery, MRI showed tumor regrowth, and the patient died 21 months after surgery.

mutation changes lysine 27 to methionine in the N-terminus of the histone tail of the *H3F3A* gene, which mainly encodes the histone variant H3.3. This mutation is frequently detected in pediatric glioblastomas, diffuse intrinsic pontine gliomas (DIP-Gs), thalamic gliomas, and spinal cord astrocytomas¹⁹⁻²¹; however, it is not found in other CNS tumors or normal nerve tissues. This indicates that H3 K27M is a driver gene abnormality in brain stem gliomas and could be a powerful diagnostic marker of spinal diffuse astrocytomas.²²

In the 2016 WHO classification of CNS tumors, diffuse midline gliomas with the H3 K27M mutation were categorized as having a poor prognosis following the results of molecular diagnosis. Diffuse midline gliomas, characterized by a specific site mutation of H3 K27M, usually occur in the midline of the CNS, and this entity was renamed "diffuse midline glioma, H3 K27-altered" to reflect the fact that other changes (e.g., EZHIP protein overexpression) can define this entity in addition to the previously recognized H3 K27 mutations in the fifth edition of the WHO Brain Tumor Classification (2021).⁴

According to recent studies of comprehensive molecular profiling of CNS tumors, approximately 50%–60% of high-grade spinal astrocytoma cases have the H3F3A K27M (H3 K27M) mutation, as do DIPGs and thalamic gliomas. $^{6,19,21-25}$

H3 K27M-mutant spinal cord gliomas are highly malignant tumors according to the WHO classification; however, their clinical manifestations, imaging characteristics, chemotherapy,

Table 1. Summary of intramedullary spinal cord tumors

Tumor type	Gene mutation	Incidence	Clinical implications
High-grade astrocytomas	H3 K27M	About half of spinal high-grade astrocytomas were described to harbor the H3 K27M mutation.	H3 K27M mutation is associated with high malignancy, regardless of the histopathological findings.
Low-grade astrocytomas	BRAF	Although not restricted to spinal cord lesions, more than 75% of PA harbor the <i>KIAA1549-BRAF</i> mutation, and about 6% harbor the <i>BRAF</i> V600E mutation. The frequency of the mutation in DA is unknown.	KIAA1549-BRAF mutation is correlated with better prognosis. BRAF V600E is associated with more aggressive behavior in pediatric low-grade astrocytomas.
	IDH	Although common in intracranial counterparts, it seems to be quite rare in spinal cord lesions.	The relationship with the prognosis in spinal astrocytomas is controversial because of its rarity.
Spinal ependymomas	NF2	$N\!F\!2$ mutation is not apparent in intracranial ependy momas.	Driver mutation for spinal cord ependymomas.
	MYCN amplification	Most likely present in a quite small number of patients in spinal ependymoma.	It has characteristic clinical features and correlates with poor prognosis.
Spinal hemangioblas- tomas	VHL	Most cases of VHL disease are associated with HB and some patients with sporadic HB carry the $\it VHL$ mutation.	Benign vascular lesion.

PA, pilocytic astrocytoma; DA, diffuse astrocytoma; HB, hemangioma.

and appropriate surgical treatment have not yet been well-elucidated due to their rarity. At present, there are few reports about the diagnosis and treatment of H3 K27M-mutant spinal cord gliomas. 24,26-28 Although H3 K27M-mutant diffuse midline gliomas have currently been classified as WHO grade IV, recent studies on high-grade gliomas of the spinal cord have not revealed a clear prognostic difference between the prognosis of H3 K27 wild-type cases and H3 K27M mutant cases.^{6,24,28} However, high enhancer of zeste homolog 2 (EZH2) expression and H3 K27me3 loss may be associated with a poor prognosis. 22,29 The interaction between the H3 K27M mutation and polycomb repressive complex 2 is promoted by EZH2. This interaction results in an overall reduction of H3 K27me3, as observed in tumors other than spinal cord gliomas. Ishi et al.²² found that the combination of H3 K27me3 status and EZH2 expression had prognostic value for WHO grade 2-4 diffuse spinal cord gliomas. Maeda et al.³⁰ reported that mutant allele-specific imbalance was associated with significantly higher Ki-67 index and poorer survival, and related to downregulation of H3 K27me3 modification.

2) Low-grade astrocytomas

The majority of spinal cord low-grade astrocytomas are grade 1 pilocytic astrocytomas and grade 2 astrocytomas. These tumors have been reported to have a better prognosis than high-

grade astrocytomas.^{6,31} Surgical resection is the mainstay of treatment for patients with low-grade spinal cord astrocytoma with the intention of maximizing resection and avoiding long-term neurological dysfunction. GTR may be possible in cases with a clear tumor-parenchyma interface (Fig. 2). It is difficult to achieve GTR with more invasive tumors, such as grade 2 astrocytoma, although a better prognosis is expected than with high-grade astrocytoma because of their slower growth pattern (Fig. 3).

(1) BRAF

Two major mutations have been noted in *BRAF*: a fusion oncogene between *BRAF* and *KIAA1549* (*KIAA1549-BRAF*), and the substitution of valine to glutamate at position 600 (*BRAF* V600E). These genetic mutations have been reported to activate the mitogen-activated protein kinase (MAPK) pathway, which is associated with tumorigenesis. Pilocytic astrocytomas harboring the *BRAF* V600E mutation account for about 5%–15% of all cases, and the *KIAA1549-BRAF* fusion gene is more likely to be detected, especially in infratentorial lesions (Table 1). In a study including 10 cases of grade 1 spinal cord pilocytic astrocytomas, there were 3 cases with *BRAF-KIAA1549* translocation and 5 cases with *BRAF* copy number gain. Another study that included 26 grade 1 spinal cord pilocytic astrocytomas revealed that 10 patients harbored the *KIAA1549-BRAF* mutation and 1 patient harbored the *BRAF* V600E mutation.

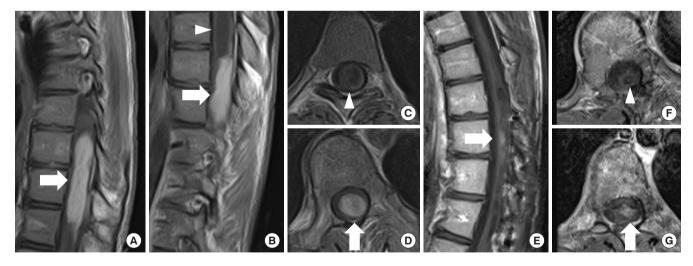


Fig. 2. Pilocytic astrocytoma, World Health Organization grade 1. (A–D) An 18-year-old man scheduled for surgery for scoliosis revealed a mass lesion at T8–12 with syrinx. The tumor was shown in contrast by gadolinium-enhanced T1-weighted magnetic resonance imaging (MRI) (arrow: tumor, arrowhead: syrinx). During the waiting period for surgery, the patient suffered from sudden onset of paralysis of the lower limbs and urinary retention, and emergency surgery was performed. The tumor had a relatively distinct surgical plane that separated it from the surrounding spinal cord parenchyma, and almost the entire tumor could be removed. Thirty-one months later, MRI showed that a tiny contrast-enhancing lesion was still present (E, G) and the syrinx resolution (F). At 40 months postoperatively, the patient's neurological symptoms were stable.

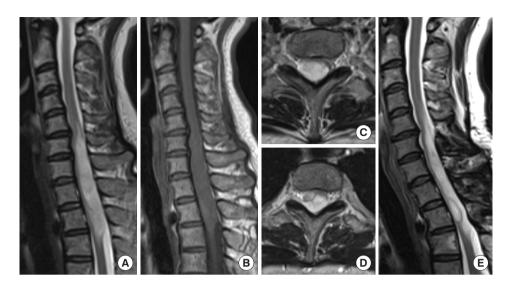


Fig. 3. Diffuse astrocytoma, World Health Organization grade 2, *IDH1* R132H mutant. A 42-year-old woman experienced numbness in both lower extremities for about 3 years and numbness in the right upper extremity and muscle weakness in the right lower extremity for 2 months. Magnetic resonance imaging (MRI) revealed an intramedullary spinal tumor at the C5–T3 levels. The tumor diffusely expanded with high signal intensity on T2-weighted MRI (A, C, D) and without any enhancements on gadolinium-enhanced T1-weighted MRI (B). It was difficult to identify a distinct surgical plane for the tumor, and partial removal was achieved. The tumor was diagnosed as diffuse astrocytoma, *IDH1* R132H mutant. Forty-two months later, MRI showed no evidence of tumor growth (E), and her symptoms were stable.

BRAF mutations have also been reported in grade 2 spinal cord astrocytomas. In a study including 10 cases of grade 2 spinal cord diffuse astrocytomas, 1 case had BRAF-KIAA1549 translocation and 2 cases had BRAF amplification.³⁸ Another study that included 17 grade 1 spinal cord pilocytic astrocytomas revealed that 2 patients harbored the BRAF V600E mutation.³¹ Low-grade gliomas with BRAF mutations can be classified into new tumor types according to the 2021 WHO classification. Reflecting the practical and conceptual importance of separating pediatric gliomas from other diffuse gliomas, 2 additional groups were added: pediatric diffuse low-grade gliomas and pediatric diffuse high-grade gliomas. "Diffuse low-grade glioma, MAPK pathway-altered" in the group of pediatric diffuse low-grade gliomas is defined as a pediatric glioma with broad histologic features, including astrocytic, oligodendroglial, or mixed morphology that shows activation of the MAPK pathway, such as BRAF mutations.4

The influence of *BRAF* mutations on the prognosis remains controversial. Some studies have shown that the *KIAA1549-BRAF* fusion is associated with improved prognosis in pediatric low-grade astrocytomas, ^{36,39,40} while another report revealed that the absence of the *KIAA1549-BRAF* fusion did not contribute significantly to the prognosis of spinal cord grade 1 pilocytic astrocytomas. ³¹ The *BRAF* V600E point mutation is thought to

be associated with more aggressive behavior in pediatric lowgrade astrocytomas.⁴⁰

(2) IDH

Isocitrate dehydrogenase (IDH) mutations were first identified in 2008 in intracranial glioblastoma⁴¹ and in >80% of WHO grade 2 and 3 cases. 42,43 Because IDH mutations are associated with the prognosis of intracranial gliomas, 43,44 IDH mutations are considered clinically significant, and the 2021 WHO classification classifies the common diffuse gliomas of adults into 3 types: "astrocytoma, IDH-mutant"; "oligodendroglioma, IDHmutant and 1p/19q-codeleted"; and "glioblastoma, IDH-wildtype."4 However, IDH mutations are extremely rare in spinal gliomas and their incidence in spinal cord gliomas is not well understood32,45 (Table 1). According to the results of immunohistochemistry and Sanger sequencing for 120 midline gliomas, including 35 spinal gliomas, 61 patients tested positive for the H3 K27M mutation, while only 2 cases exhibited the IDH1 R132H mutation.46 In another study examining the molecular characteristics of 83 spinal gliomas, there were no IDH1 mutations, although H3 K27M mutations were found in 35 cases.⁶ Furthermore, the results of the genetic analysis of spinal cord gliomas by next-generation sequencing (NGS) have been recently reported. NGS for 61 intramedullary astrocytomas including 17

grade 2 diffuse astrocytomas, revealed 2 cases of IDH mutations.³¹ In another study of NGS for 26 spinal astrocytomas, there were 2 IDH mutation cases.⁷ Thus, spinal cord gliomas are less likely to harbor the IDH mutation than intracranial gliomas.

In addition, the majority of IDH gene mutations in intracranial gliomas are IDH1 R132H43; however, the IDH mutation variants found in spinal cord gliomas may be different from those found in intracranial gliomas. A recent retrospective study further supports this hypothesis. In this study of IDH1 R132H mutant gliomas and noncanonical IDH-mutant (not IDH1 R132H) gliomas, none of the 166 IDH1 R132H mutant gliomas included an infratentorial region, while nine of 155 (5.5%) noncanonical IDH-mutant gliomas were infratentorial regions.⁴⁷ Intracranial gliomas and infratentorial gliomas including the spinal cord seem to have different genetic underpinnings. Even though IDH-mutant spinal gliomas are very rare, several spinal gliomas with variants other than IDH1 R132H have been reported, and IDH1 R132H mutations are probably not so prevalent in spinal gliomas. Konovalov et al. 48 reported 5 cases of spinal cord astrocytomas with IDH mutations: 2 had IDH1 R132H mutations, while 1 had an IDH1 R132G mutation. In addition, the remaining 2 cases had translocations at positions 82 (Arg → Lys, R82K) and 76 (Ile → Thr, I76T) of the IDH1 gene, which had never been described in intracranial gliomas in the past. In another study, Takai et al.49 reported one case of spinal astrocytoma with an IDH1 R132S mutation, and we previously reported 2 cases with IDH1 R132C and IDH1 R132H mutations, respectively. 45 There is also a report of spinal cord glioma with IDH2 R172.31 Even though IDH-mutant spinal gliomas are very rare, several spinal gliomas with variants other than IDH1 R132H have been reported, and IDH1 R132H mutations are probably not very prevalent in spinal gliomas. Therefore, Immunohistochemistry using an anti-IDH1 R132H antibody, which is commonly used for intracranial gliomas, is not sufficient for spinal gliomas, and genetic testing is desirable.

The prognostic impact of IDH mutations in spinal cord gliomas is controversial. ^{7,31,45,49,50} Because IDH mutations are rare in spinal gliomas, the current knowledge of IDH mutations and the associated prognosis is inadequate. However, the prognosis of IDH-mutant spinal gliomas does not seem to be different depending on the IDH variant. A study of IDH variants and patients' prognosis found no significant difference in prognosis between *IDH1* R132H mutant glioma and noncanonical IDH-mutant glioma. ⁴⁷ The clinical significance of IDH mutations in spine gliomas needs to be confirmed in a larger cohort.



Fig. 4. Spinal ependymoma. A 44-year-old woman who had been treated for multiple intracranial meningiomas developed numbness in her right upper extremity. Magnetic resonance imaging (MRI) revealed an intramedullary tumor at the C5–6 level with an enlarged spinal cord. (A) T2-weighted MRI showed a hyperintense tumor with a syrinx and a partially hypointense lesion reflecting hemosiderin. (B) The tumor was shown in contrast by gadolinium-enhanced T1-weighted MRI. Tumor removal was performed via a posterior approach, and gross tumor resection was achieved because the surgical plane of the tumor was clear. (C, D) T2-weighted MRI 67 months after surgical removal showed low intensity that reflected hemosiderin, whereas there was no evident tumor recurrence and no enhancement on gadolinium-enhanced T1-weighted MRI.

2. Spinal Ependymomas

Spinal ependymomas are the most common IMSCTs. 9,51,52 Almost all ependymomas are benign tumors with clear tumor borders, and long-term survival can be expected by targeting GTR (Fig. 4). 8,53 Therefore, the long-term functional prognosis should also be considered in the treatment. 54-57

1) NF2

Neurofibromin (*NF2*) gene mutations are driver mutations for spinal cord ependymomas and appear to be the most prevalent genetic mutations in spinal ependymomas⁵⁸ (Table 1). Pajtler et al.¹⁰ analyzed about 500 ependymomas, including 47 spinal lesions and revealed that most spinal ependymomas had a loss of the 22q locus, which harbors the *NF2* gene, although *NF2* mutations were not seen in intracranial ependymomas. In another study, 47% (9 of 19) of spinal ependymomas had *NF2* mutations.^{9,59} *NF2* is a tumor suppressor gene, and aberrations of the *NF2* gene make cells less responsive to contact inhibition, thereby promoting tumorigenesis.⁵⁸

2) MYCN amplification

"Spinal ependymoma, *MYCN*-amplified" is a new category in the 2021 WHO classification.⁴ Although *MYCN*-amplified ependymomas are very rare, they are associated with aggressive

behavior and unfavorable outcomes^{60,61} (Table 1). Ghasemi et al.⁶² investigated 13 *MYCN*-amplified ependymomas, of which 10 were WHO grade 3 and 3 were WHO grade 2 on histopathological examination. Compared to other subtypes of ependymomas, these groups had worse median progression-free survival (17 months) and median overall survival (87 months). These tumors were also characterized by a favored location of the cervical and thoracic spine, and were predominantly intradural and extramedullary.⁶³ The presence of diffuse leptomeningeal spread and dissemination has also been revealed as a distinctive feature of these tumors. Further study is required to develop new strategies to improve the prognosis of patients with *MYCN*-amplified spinal ependymoma.

3. Spinal Hemangioblastomas

Spinal cord hemangioblastomas are benign vascular lesions and constitute the third most common IMSCTs.⁸ Surgical resection is recommended in cases of symptomatic lesions or lesions that appear to be growing on repeat imaging studies. Because hemangioblastoma generally shows a well-defined tumor border that allows GTR, radiotherapy has a limited role.^{8,64} Although these are vascular-rich tumors, intraoperative hemorrhage is generally not a problem due to the availability of techniques such as temporary intraoperative arterial occlusion and preoperative embolization (Fig. 5).^{8,65} Although hemangioblas-



Fig. 5. Hemangioblastoma. A 44-year-old man presented with numbness in the right upper extremity and right paralysis. A radiological examination revealed a tumor at the C5 level. (A) T2-weighted MRI showed a well-defined tumor with a syrinx and flow void. (B) The tumor was strongly shown in contrast with gadolinium-enhanced T1-weighted magnetic resonance imaging (MRI). (C) Dynamic computed tomography angiography also showed a strongly contrasted tumor with dilated feeder and drainer. Preoperative angiography and feeder occlusion were performed, and the tumor was totally resected microscopically via a posterior approach. Immediately after surgery, the patient's symptoms improved. (D) Thirty-six months later, MRI showed no recurrence of the tumor and the syrinx had collapsed.

toma regrowth is very rare once the tumor is completely removed, patients with von Hippel-Lindau (VHL) disease may show multiple hemangioblastomas with new lesions repeatedly arising.

1) VHL

Approximately 20% to 40% of patients who develop hemangioblastomas have VHL disease, ⁶⁶ which is an inherited disorder that causes multiple tumors and cysts in various parts of the body (Table 1). Glasker et al. ⁶⁷ reported that 94% of VHL disease-associated hemangioblastomas harbor *VHL* mutations and 62% exhibit loss of heterozygosity (LOH) at the VHL locus (3p25-56). By contrast, of 13 sporadic hemangioblastomas, 23% expressed germline mutations in *VHL* and 50% had LOH of the *VHL* locus. ⁶⁷ The *VHL* gene encodes an E3 ubiquitin ligase that targets hypoxia-inducible factor-1a (HIF- α), which is known to be a regulator of vascular growth. ⁹ Mutations or deletions of the *VHL* gene cause cells to be unable to adequately degrade HIF- α , leading to vascular proliferation. Activated HIF- α and vascular endothelial growth factor were found to be correlated and increased in *VHL* mutant cells. ^{32,68}

CONCLUSION

The discovery of the genetic and molecular mechanisms of CNS tumors is beginning to impact the management of intracranial tumors, with improved predictions of prognosis and availability of targeted therapy. The 2021 WHO classification has been modified to reflect these facts. However, the genetic underpinnings of spinal cord tumors remain less well understood as those of their intracranial counterparts due to their rarity and difficulty in treatment because of their location in eloquent areas. Molecular and genetic differences exist between tumors located in the spinal cord and intracranial regions, even within the same pathological type. Therefore, further genetic studies on IMSCTs are warranted in order to develop novel therapies and improve the prognosis of patients suffering from these challenging tumors.

NOTES

Conflict of Interest: The authors have nothing to disclose.

Funding/Support: This study received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

Author Contribution: Conceptualization: YN, YN, FO, MT; Data curation: YN, KE, JY; Project administration: YN, MT; Writ-

ing - original draft: YN, KE, JY; Writing - review & editing: YN, YN, SH, FO, MT, RS.

ORCID

Yusuke Nishimura: 0000-0001-9689-0586 Fumiharu Ohka: 0000-0002-5569-5626 Ryuta Saito: 0000-0002-2484-1360

REFERENCES

- Zulch K. Histological typing of tumours of the central nervous system. Vol no. 21. Geneva (Switzerland): World Health Organization; 1979.
- Louis DN, Ohgaki H, Wiestler OD, et al. WHO classification of tumours of the central nervous system. Revised 4th ed. Lyon (France): International Agency for Research on Cancer; 2016.
- Louis DN, Perry A, Reifenberger G, et al. The 2016 World Health Organization Classification of Tumors of the Central Nervous System: a summary. Acta Neuropathol. 2016;131: 803-20
- Louis DN, Perry A, Wesseling P, et al. The 2021 WHO Classification of Tumors of the Central Nervous System: a summary. Neuro Oncol 2021;23:1231-51.
- Zuchegna C, Di Zazzo E, Moncharmont B, et al. Dual-specificity phosphatase (DUSP6) in human glioblastoma: epithelial-to-mesenchymal transition (EMT) involvement. BMC Res Notes 2020;13:374.
- Chai RC, Zhang YW, Liu YQ, et al. The molecular characteristics of spinal cord gliomas with or without H3 K27M mutation. Acta Neuropathol Commun 2020;8:40.
- Biczok A, Strubing FL, Eder JM, et al. Molecular diagnostics helps to identify distinct subgroups of spinal astrocytomas. Acta Neuropathol Commun 2021;9:119.
- 8. Tobin MK, Geraghty JR, Engelhard HH, The molecular characteristics of spinal cord gliomas with or without H3 K27M mutation Intramedullary spinal cord tumors: a review of current and future treatment strategies. Neurosurg Focus 2015;39:E14.
- Azad TD, Jiang B, Bettegowda C. Molecular foundations of primary spinal tumors-implications for surgical management. Ann Transl Med 2019;7:222.
- Pajtler KW, Witt H, Sill M, et al. Molecular classification of ependymal tumors across All CNS compartments, histopathological grades, and age groups. Cancer Cell 2015;27:728-43.

- 11. Karsy M, Guan J, Sivakumar W, et al. The genetic basis of intradural spinal tumors and its impact on clinical treatment. Neurosurg Focus 2015;39:E3.
- 12. Louis DN, von Deimling A. Grading of diffuse astrocytic gliomas: Broders, Kernohan, Zulch, the WHO... and Shakespeare. Acta Neuropathol 2017;134:517-20.
- 13. Brat DJ, Aldape K, Colman H, et al. cIMPACT-NOW update 3: recommended diagnostic criteria for "Diffuse astrocytic glioma, IDH-wildtype, with molecular features of glioblastoma, WHO grade IV". Acta Neuropathol 2018;136:805-10.
- Sanai N, Polley MY, McDermott MW, et al. An extent of resection threshold for newly diagnosed glioblastomas. J. Neurosurg 2011;115:3-8.
- 15. McGirt MJ, Goldstein IM, Chaichana KL, et al. Extent of surgical resection of malignant astrocytomas of the spinal cord: outcome analysis of 35 patients. Neurosurgery 2008; 63:55-60; discussion 60-1.
- 16. Nagoshi N, Tsuji O, Suzuki S, et al. Clinical outcomes and a therapeutic indication of intramedullary spinal cord astrocytoma. Spinal Cord 2022;60:216-22.
- 17. Zou Y, Sun J, Zhou Y, et al. Prognostic factors and treatment of spinal astrocytomas: a multi-institutional cohort analysis. Spine (Phila Pa 1976) 2018;43:E565-73.
- 18. Konar SK, Bir SC, Maiti TK, et al. A systematic review of overall survival in pediatric primary glioblastoma multiforme of the spinal cord. J Neurosurg Pediatr 2017;19:239-48.
- 19. Schwartzentruber J, Korshunov A, Liu XY, et al. Driver mutations in histone H3.3 and chromatin remodelling genes in paediatric glioblastoma. Nature 2012;482:226-31.
- 20. Wu G, Broniscer A, McEachron TA, et al. Somatic histone H3 alterations in pediatric diffuse intrinsic pontine gliomas and non-brainstem glioblastomas. Nat Genet 2012;44:251-3.
- 21. Gessi M, Gielen GH, Dreschmann V, et al. High frequency of H3F3A (K27M) mutations characterizes pediatric and adult high-grade gliomas of the spinal cord. Acta Neuropathol 2015;130:435-7.
- 22. Ishi Y, Takamiya S, Seki T, et al. Prognostic role of H3K27M mutation, histone H3K27 methylation status, and EZH2 expression in diffuse spinal cord gliomas. Brain Tumor Pathol 2020;37:81-8.
- 23. Aihara K, Mukasa A, Gotoh K, et al. H3F3A K27M mutations in thalamic gliomas from young adult patients. Neuro Oncol 2014;16:140-6.
- 24. Yi S, Choi S, Shin DA, et al. Impact of H3.3 K27M mutation on prognosis and survival of grade IV spinal cord glioma on

- the basis of new 2016 World Health Organization Classification of the central nervous system. Neurosurgery 2019;84: 1072-81.
- 25. Alvi MA, Ida CM, Paolini MA, et al. Spinal cord high-grade infiltrating gliomas in adults: clinico-pathological and molecular evaluation. Mod Pathol 2019;32:1236-43.
- 26. Jung JS, Choi YS, Ahn SS, et al. Differentiation between spinal cord diffuse midline glioma with histone H3 K27M mutation and wild type: comparative magnetic resonance imaging. Neuroradiology 2019;61:313-22.
- 27. Cheng R, Li DP, Zhang N, et al. Spinal cord diffuse midline glioma with histone H3 K27M mutation in a pediatric patient. Front Surg 2021;8:616334.
- 28. Akinduro OO, Garcia DP, Higgins DMO, et al. A multicenter analysis of the prognostic value of histone H3 K27M mutation in adult high-grade spinal glioma. J Neurosurg Spine 2021;35:834-43.
- 29. Karlowee V, Amatya VJ, Takayasu T, et al. Immunostaining of increased expression of enhancer of zeste homolog 2 (EZH2) in diffuse midline glioma H3K27M-mutant patients with poor survival. Pathobiology 2019;86:152-61.
- 30. Maeda S, Ohka F, Okuno Y, et al. H3F3A mutant allele specific imbalance in an aggressive subtype of diffuse midline glioma, H3 K27M-mutant. Acta Neuropathol Commun 2020; 8:8.
- 31. Lebrun L, Melendez B, Blanchard O, et al. Clinical, radiological and molecular characterization of intramedullary astrocytomas. Acta Neuropathol Commun 2020;8:128.
- 32. Abd-El-Barr MM, Huang KT, Moses ZB, et al. Recent advances in intradural spinal tumors. Neuro Oncol 2018;20: 729-42.
- 33. Jones DT, Gronych J, Lichter P, et al. MAPK pathway activation in pilocytic astrocytoma. Cell Mol Life Sci 2012;69:1799-811.
- 34. Davies H, Bignell GR, Cox C, et al. Mutations of the BRAF gene in human cancer. Nature 2002;417:949-54.
- 35. Collins VP, Jones DT, Giannini C. Pilocytic astrocytoma: pathology, molecular mechanisms and markers. Acta Neuropathol 2015;129:775-88.
- 36. Grob ST, Nobre L, Campbell KR, et al. Clinical and molecular characterization of a multi-institutional cohort of pediatric spinal cord low-grade gliomas. Neurooncol Adv 2020;2: vdaa103.
- 37. Penman CL, Faulkner C, Lowis SP, et al. Current understanding of BRAF alterations in diagnosis, prognosis, and therapeutic targeting in pediatric low-grade gliomas. Front Oncol

- 2015;5:54.
- 38. Shankar GM, Lelic N, Gill CM, et al. BRAF alteration status and the histone H3F3A gene K27M mutation segregate spinal cord astrocytoma histology. Acta Neuropathol 2016;131: 147-50.
- 39. Hawkins C, Walker E, Mohamed N, et al. BRAF-KIAA1549 fusion predicts better clinical outcome in pediatric low-grade astrocytoma. Clin Cancer Res 2011;17:4790-8.
- 40. Yang RR, Aibaidula A, Wang WW, et al. Pediatric low-grade gliomas can be molecularly stratified for risk. Acta Neuropathol 2018;136:641-55.
- 41. Parsons DW, Jones S, Zhang X, et al. An integrated genomic analysis of human glioblastoma multiforme. Science 2008; 321:1807-12.
- 42. Han S, Liu Y, Cai SJ, et al. IDH mutation in glioma: molecular mechanisms and potential therapeutic targets. Br J Cancer 2020;122:1580-9.
- 43. Yan H, Parsons DW, Jin G, et al. IDH1 and IDH2 mutations in gliomas. N Engl J Med 2009;360:765-73.
- 44. Ichimura K, Pearson DM, Kocialkowski S, et al. IDH1 mutations are present in the majority of common adult gliomas but rare in primary glioblastomas. Neuro Oncol 2009;11: 341-7.
- 45. Nagashima Y, Nishimura Y, Ohka F, et al. Driver genetic mutations in spinal cord gliomas direct the degree of functional impairment in tumor-associated spinal cord injury. Cells 2021;10:2525.
- 46. Wang L, Li Z, Zhang M, et al. H3 K27M-mutant diffuse midline gliomas in different anatomical locations. Hum Pathol 2018;78:89-96.
- 47. Poetsch L, Bronnimann C, Loiseau H, et al. Characteristics of IDH-mutant gliomas with non-canonical IDH mutation. J Neurooncol 2021;151:279-86.
- 48. Konovalov NA, Asyutin DS, Shayhaev EG, et al. Rare cases of IDH1 mutations in spinal cord astrocytomas. Acta Naturae 2020;12:70-3.
- 49. Takai K, Tanaka S, Sota T, et al. Spinal cord astrocytoma with isocitrate dehydrogenase 1 gene mutation. World Neurosurg 2017;108:991.e13- 6.
- 50. Sun M, Wang L, Lu D, et al. Concomitant KIAA1549-BRAF fusion and IDH mutation in pediatric spinal cord astrocytoma: a case report and literature review. Brain Tumor Pathol 2021;38:132-7.
- 51. Milano MT, Johnson MD, Sul J, et al. Primary spinal cord glioma: a Surveillance, Epidemiology, and End Results database study. J Neurooncol 2010;98:83-92.

- 52. Samartzis D, Gillis CC, Shih P, et al. Intramedullary spinal cord tumors: part I-epidemiology, pathophysiology, and diagnosis. Global Spine J 2015;5:425-35.
- 53. Constantini S, Miller DC, Allen JC, et al. Radical excision of intramedullary spinal cord tumors: surgical morbidity and long-term follow-up evaluation in 164 children and young adults. J Neurosurg 2000;93:183-93.
- 54. Ahmed R, Menezes AH, Awe OO, et al. Long-term disease and neurological outcomes in patients with pediatric intramedullary spinal cord tumors. J Neurosurg Pediatr 2014;13: 600-12.
- 55. Gonda T, Nagashima Y, Nishimura Y, et al. Postoperative cervicothoracic kyphosis following infantile intramedullary tumor resection accelerates neurological deterioration. NMC Case Rep J 2021;8:705-11.
- 56. Behmanesh B, Gessler F, Quick-Weller J, et al. Regional spinal cord atrophy is associated with poor outcome after surgery on intramedullary spinal cord ependymoma: a new aspect of delayed neurological deterioration. World Neurosurg 2017;100:250-5.
- 57. Lundar T, Due-Tonnessen BJ, Fric R, et al. Outcome after treatment of spinal ependymoma in children and adolescents: long-term follow-up of a single consecutive institutional series of 33 patients treated over eight decades. World Neurosurg. 2021;150:e228-e35.
- 58. Connolly ID, Ali R, Li Y, Gephart MH. Genetic and molecular distinctions in spinal ependymomas: a review. Clin Neurol Neurosurg 2015;139:210-5.
- 59. Bettegowda C, Agrawal N, Jiao Y, et al. Exomic sequencing of four rare central nervous system tumor types. Oncotarget 2013;4:572-83.

- 60. Shatara M, Schieffer KM, Klawinski D, et al. Clinically aggressive pediatric spinal ependymoma with novel MYC amplification demonstrates molecular and histopathologic similarity to newly described MYCN-amplified spinal ependymomas. Acta Neuropathol Commun 2021;9:192.
- 61. Raffeld M, Abdullaev Z, Pack SD, et al. High level MYCN amplification and distinct methylation signature define an aggressive subtype of spinal cord ependymoma. Acta Neuropathol Commun 2020;8:101.
- 62. Ghasemi DR, Sill M, Okonechnikov K, et al. MYCN amplification drives an aggressive form of spinal ependymoma. Acta Neuropathol 2019;138:1075-89.
- 63. Larrew T, Saway BF, Lowe SR, et al. Molecular classification and therapeutic targets in ependymoma. Cancers (Basel) 2021;13:6218.
- Chamberlain MC, Tredway TL. Adult primary intradural spinal cord tumors: a review. Curr Neurol Neurosci Rep 2011; 11:320-8.
- 65. Saliou G, Giammattei L, Ozanne A, et al. Role of preoperative embolization of intramedullary hemangioblastoma. Neurochirurgie 2017;63:372-5.
- 66. Augtein J, Green SM, Monro AM, et al. Adrenergic neurone blocking agents derived from 1,4-benzodioxan. J Med Chem 1965;8:446-56.
- 67. Glasker S, Bender BU, Apel TW, et al. Reconsideration of biallelic inactivation of the VHL tumour suppressor gene in hemangioblastomas of the central nervous system. J Neurol Neurosurg Psychiatry 2001;70:644-8.
- 68. Bader HL, Hsu T. Systemic VHL gene functions and the VHL disease. FEBS Lett 2012;586:1562-9.