



Published in final edited form as:

*Annu Rev Neurosci.* 2022 July 08; 45: 109–129. doi:10.1146/annurev-neuro-110920-011929.

## Mesoaccumbal Dopamine Heterogeneity: What Do Dopamine Firing and Release Have to Do with It?

Johannes W. de Jong<sup>\*</sup>,

Kurt M. Fraser<sup>\*</sup>,

Stephan Lammel

Department of Molecular and Cell Biology and Helen Wills Neuroscience Institute, University of California, Berkeley, California, USA

### Abstract

Ventral tegmental area (VTA) dopamine (DA) neurons are often thought to uniformly encode reward prediction errors. Conversely, DA release in the nucleus accumbens (NAc), the prominent projection target of these neurons, has been implicated in reinforcement learning, motivation, aversion, and incentive salience. This contrast between heterogeneous functions of DA release versus a homogeneous role for DA neuron activity raises numerous questions regarding how VTA DA activity translates into NAc DA release. Further complicating this issue is increasing evidence that distinct VTA DA projections into defined NAc subregions mediate diverse behavioral functions. Here, we evaluate evidence for heterogeneity within the mesoaccumbal DA system and argue that frameworks of DA function must incorporate the precise topographic organization of VTA DA neurons to clarify their contribution to health and disease.

### Keywords

dopamine; striatum; motivation; ventral tegmental area; reward; aversion

## 1. INTRODUCTION

Midbrain dopamine (DA) neurons have received great attention because of their important roles in a broad range of motivated behaviors and their dysfunction in several neuropsychiatric disorders, including Parkinson's disease, major depressive disorder, substance use disorder, and schizophrenia (Bjorklund & Dunnett 2007, Bromberg-Martin et al. 2010, Nestler & Carlezon 2006, Robinson & Berridge 2008, Schultz 2007, Wise 2004, Wise & Robble 2020). The striatum is the major projection target of midbrain DA neurons (Albanese & Minciocchi 1983, Ikemoto 2007, Lammel et al. 2008, Poulin et al. 2018, Roeper 2013). It is often subdivided into the nucleus accumbens (NAc), which is located in

lammel@berkeley.edu .

<sup>\*</sup>These authors contributed equally to this article

### DISCLOSURE STATEMENT

The authors are not aware of any affiliations, memberships, funding, or financial holdings that might be perceived as affecting the objectivity of this review.

the ventral striatum, and the dorsal striatum based upon projections arising from the ventral tegmental area (VTA) and substantia nigra pars compacta (SNc) DA neurons, respectively.

Rapid technological advances in system neuroscience have provided researchers with unprecedented opportunities to study the midbrain DA system. The incorporation of optogenetic approaches and use of transgenic animals have allowed for reliable access to DA neurons, with laser-light delivery during electrophysiological recordings eliciting rapid neural responses and enabling identification of genetically defined DA neurons (i.e., opto-tagging) (Cohen et al. 2012). Two-photon calcium imaging through an implanted lens has been used to record the activity of hundreds of DA neurons in the VTA during complex behavioral tasks (Engelhard et al. 2019, Hamid et al. 2021). Additionally, genetically encoded G protein-coupled DA sensors have been developed to successfully monitor the dynamics of DA release with high spatiotemporal resolution in freely behaving animals (Patriarchi et al. 2018, Sun et al. 2018). Together, these approaches allowed the field to overcome some of its previous challenges, for example, the reliable identification of VTA DA neurons in vivo (Margolis et al. 2006, Ungless & Grace 2012), and advances have been made in long-standing debates about the role of DA neurons in aversion (Schultz 2019, Verharen et al. 2020). While previous notions about the function and connectivity of VTA DA neurons are slowly beginning to change, the field faces a number of new challenges. While the nature of some of these challenges is methodological, such as the development of strategies to relate data from calcium imaging and electrophysiological experiments in DA neurons (Fleming et al. 2021, Sabatini & Tian 2020), associating VTA DA firing patterns and accumbal DA release with a precise behavioral function has been more difficult than previously thought. A recent study suggested that the slow, gradual release of DA (ramping) in the NAc is controlled locally, independent from the firing of VTA DA cell bodies, and encodes information about the value of the anticipated reward (Mohebi et al. 2019), suggesting that DA dynamics mediated by different mechanisms in different locations (i.e., at the level of cell bodies versus terminals) may underly different behavioral functions. In contrast, another recent study developed an experimental paradigm using virtual reality to disambiguate reward prediction error (RPE) encoding from value, suggesting that ramping DA signals are consistent with RPE rather than value and can be observed both at the level of individual VTA DA neurons and in axon terminals (Kim et al. 2020). Thus, an important question that currently hampers our understanding of DA function is how neural activity of VTA DA neurons translates into DA release in the NAc during behavior.

Despite an increasing appreciation of DA heterogeneity, in both the SNc and the VTA, the use of such advanced approaches has surprisingly led to renewed propositions of a uniform or single function common to all DA neurons. In this review, we explore and compare studies that have recorded activity of VTA DA neurons at the level of individual cells versus DA release at the axon terminal level in the context of behaviors associated with reward learning, motivation, and aversion to better understand whether DA activity and release share common functions. Collectively, we argue that many exciting developments have emphasized numerous unique features within the DA system, and we hope to encourage investigators to incorporate this heterogeneity into computational and behavioral frameworks for DA function in health and disease.

## 2. TOPOGRAPHIC ORGANIZATION OF A HETEROGENEOUS MIDBRAIN DOPAMINE SYSTEM

Like every tissue in the body, VTA and striatum are composed of multiple cell types that vary in their gene and protein expression, morphology, and connectivity. In addition to DA neurons, the VTA contains GABAergic and glutamatergic neurons, and some DA neurons may also co-release GABA or glutamate (Morales & Margolis 2017). The striatum is composed of GABAergic medium spiny neurons, which can be largely subdivided into cells that either express DA D1 receptors or D2 receptors, as well as cholinergic interneurons (Kreitzer 2009). The pronounced cellular heterogeneity, particularly in the VTA, and the fact that some DA neurons potentially co-release glutamate and GABA, require methodological approaches that precisely identify the cellular identity of DA neurons when studying their functional role in diverse behavioral settings. In the past, DA neurons were often identified based on indirect measures, resulting in controversies about the reliability of electrophysiological criteria used to identify DA neurons *in vivo* (Margolis et al. 2006, Ungless & Grace 2012). Current opto-tagging approaches allow researchers to reliably identify DA neurons through the use of transgenic animals, restricting expression of optogenetic proteins, such as channelrhodopsin, to cells expressing genes that are associated with the DA phenotype (e.g., TH or DAT), but *in vivo* electrophysiological experiments are typically conducted from DA neurons without knowing their specific projection target.

This makes sense to some degree, given that reliable recordings from projection-defined DA neurons are not yet feasible. The primary retrograde-traveling AAV that is commercially available (Tervo et al. 2016) transfects only a small number of projection-defined VTA DA cells, and other common retrograde viruses such as CAV2-Cre would require combinatorial genetic strategies to limit expression in a projection-specific manner (e.g., use of Cre and Flp systems). Fractionating VTA neurons on the basis of unique cell types from classical tyrosine hydroxylase and DAT markers may reveal circuit- and projection-specific enrichment and a path to exploit intersectional genetic strategies to limit manipulations and cell type-specific recordings (Poulin et al. 2020). However, discrepancies of exact projection targets in the striatum labeled by medial shell-exclusive cholecystokinin-positive VTA neurons versus core-exclusive corticotropin releasing factor receptor 1-positive VTA neurons have been demonstrated, which may limit the utility of identifying projection-defined cells on the basis of the presence of these genes (Heymann et al. 2020, Poulin et al. 2018). A lasting issue remains in that all genetic tools are developed primarily in mice, with a lack of similar tool kits for rats [e.g., DAT-Flp mice (Kramer et al. 2021)], which may ultimately produce bias in the exact behavioral approaches investigators use and, in turn, more disparities in conclusions.

A major limitation of most previous *in vivo* electrophysiological studies is that recordings from DA neurons were often biased toward the lateral VTA without considering DA cells in more medial VTA subregions (e.g., Cohen et al. 2012; Eshel et al. 2015, 2016) (Figure 1). Because it has become clear that the topographic organization and connectivity of DA neurons is an important feature for their diverse functions (Collins & Saunders 2020, Poulin et al. 2020, Verharen et al. 2020), recording DA cells in only one VTA subregion alone

cannot be used to annotate the full spectrum of DA diversity. Consistent with this, recent studies using in vivo calcium imaging or electrophysiological approaches have observed pronounced functional heterogeneity of DA neurons located in more medial VTA subregions (Engelhard et al. 2019, Kim et al. 2020), whereas DA neurons in the lateral VTA seem to be more homogeneous (Eshel et al. 2016). From an anatomical perspective, it is important to mention that medial and lateral VTA DA neurons have nonoverlapping projections to distinct subregions in the NAc (Beier et al. 2015, Lammel et al. 2008). These distinct projections suggest unique functions between VTA DA neurons preventing generalization, which we discuss further below.

Traditionally, the NAc has been subdivided into shell and core substructures. The NAc shell [also often called NAc medial shell (NAcMed)] is shaped like a crescent and located medial and ventral to the core. The differences between shell and core have been defined by various histochemical (e.g., calbindin, calretinin, substance P), electrophysiological, connectivity, and functional criteria (Castro & Bruchas 2019, Ikemoto 2007, Zahm & Brog 1992). An additional ventral striatal subregion, termed nucleus accumbens lateral shell (NAcLat) (see the sidebar titled Nucleus Accumbens Lateral Shell), which is located laterally to the NAc core, has been described in several previous studies (Beier et al. 2015, Breton et al. 2019, de Jong et al. 2019, Farassat et al. 2019, Ikemoto 2007, Lammel et al. 2008, Liu et al. 2020, Yuan et al. 2019) but generally is less studied compared to the NAc core or NAc medial shell. Nonetheless, the findings that the NAcLat is part of a major disinhibitory feedback loop (Yang et al. 2018) and the most prominent projection target of DA neurons that are located in the lateral VTA (Breton et al. 2019, Lammel et al. 2008) suggest that this often overlooked structure is an integral part of the mesolimbic DA system that needs to be taken into consideration when studying the functional role of DA release in the context of DA neuron activity.

### 3. THE PHYSIOLOGY OF DOPAMINE CELL FIRING AND DOPAMINE RELEASE

A foundational study using in vivo electrophysiology recordings delineated two different firing modes of putative DA neurons: tonic (0.2–10 Hz) and burst (>10 Hz) firing patterns (Grace & Bunney 1984). In acute brain slice recordings, VTA DA neurons exhibit pacemaker or irregular firing patterns (Krabbe et al. 2015, Paladini & Roeper 2014). Conversely, burst firing, which depends on NMDA receptor activation and network activity (Zweifel et al. 2009), is absent in ex vivo slice preparation but can be induced in a subset of neurons by inhibition of T-type calcium channels (Wolfart & Roeper 2002). Both excitatory and inhibitory afferents to midbrain DA neurons have been extensively surveyed using pseudotyped rabies tracing (Beier et al. 2015, Faget et al. 2016, Lerner et al. 2015, Menegas et al. 2015, Watabe-Uchida et al. 2012). These studies have highlighted that DA neurons with distinct projection targets receive inputs from a variety of overlapping brain regions.

Given the surprisingly relatively uniform distribution of inputs onto VTA DA neurons with distinct projection targets, it is important to emphasize that rabies tracing does not quantify synaptic strength and the resultant functional connectivity of these inputs. Moreover, it can

be difficult to predict the computational or functional role of a defined direct input onto VTA DA neurons. For instance, a landmark study leveraged rabies tracing to record from opto-tagged monosynaptic inputs onto VTA DA neurons found that the information required to encode RPEs is distributed with high redundancy among several VTA DA afferents, and no single aspect of the RPE signal was localized in any particular afferent (Tian et al. 2016). This does not preclude that there exist afferents to VTA DA neurons that exert specific functions via biased connectivity in a projection-defined manner (Cardozo Pinto & Lammel 2018, de Jong et al. 2019, Juarez & Han 2016, Lammel et al. 2012, Morales & Margolis 2017, Nieh et al. 2016, Qi et al. 2014, Steidl et al. 2017). For example, glutamatergic afferents from the laterodorsal tegmentum and dorsal raphe nucleus are biased toward NAcLat-projecting VTA DA neurons, and these subcircuits are predominately involved in reward-related behaviors (de Jong et al. 2019, Lammel et al. 2012, Qi et al. 2014, Steidl et al. 2017). Conversely, glutamatergic afferents from the lateral hypothalamus primarily innervate medial VTA DA neurons, encode aversive stimuli, and promote DA release in the ventral NAcMed while indirectly inhibiting other VTA DA neurons (de Jong et al. 2019, Nieh et al. 2016). Thus, the questions of how DA neurons integrate information from different afferents and how it affects DA cell firing remain open.

Another important matter that remains unaddressed is to elucidate the neurobiological mechanisms that underlie tonic and phasic DA firing as well as the switch between these two firing states. However, there is a paucity of studies that have examined projection-defined DA neurons or DA neurons in different VTA subregions in this context. Nonetheless, two recent studies have made important progress. First, Farassat et al. (2019) combined juxtacellular recordings with retrograde tracing and found that NAcMed- and NAcLat-projecting VTA DA neurons have distinct baseline firing rate and burst characteristics. Second, Otomo et al. (2020) established *in vivo* patch clamp recordings from DA neurons and performed the first *in vivo* intracellular recordings from DA neurons since the seminal work of Grace & Bunney (1984) in the 1980s. This work delineated two qualitatively distinct types of burst firing characterized by distinct membrane potential dynamics and opposing shifts in action potential threshold. However, medial VTA DA neurons were underrepresented in the Otomo et al. (2020) study, and the projection target of all recorded neurons was unknown. Furthermore, although these studies provide important insights into the nature of tonic and phasic firing patterns, future research will need to resolve the mechanisms that mediate the switch between these firing states.

Finally, the relationship between somatic action potential firing and DA release at the axon terminal requires further investigations. Striatal DA release is evoked by action potentials that originate in the soma, but DA release sites are rapidly depleted, causing depression of release following repeated firing (Liu et al. 2018, 2021). This requires DA reserves to be replenished by reuptake mediated by the dopamine transporter (DAT). Importantly, DAT transporter expression varies between DA neurons with different projection targets (Lammel et al. 2008, Sesack et al. 1998), suggesting a similar pattern of neuronal firing might result in different extracellular DA concentrations at distinct DA release sites. The rapid depletion of DA release sites makes it unlikely that DA release precisely tracks cell body firing, and DA axon terminals do not form traditional synapses that would support such a 1:1 relationship of spiking to release (Liu et al. 2018). Increased synchronization

that occurs during phasic DA firing has been proposed to cause widespread diffusion of DA and activation of postsynaptic DA receptors by saturating DAT reuptake (Liu et al. 2021). Putting the emphasis on synchronized firing poses the possibility that ramping responses, which are predominantly observed when examining striatal DA release (Howe et al. 2013, Mohebi et al. 2019, Roitman et al. 2004, Wassum et al. 2012), are not exclusively the result of ramping increases in somatic firing but instead are caused by increased synchronization of tonic firing neurons. Related to this, recent work in opto-tagged VTA DA neurons provided evidence for multiplexing in that a subset of DA neurons encoded environmental cues through phasic bursts and goal-directed actions through modulation of tonic firing (Kremer et al. 2020). Further highlighting the nonlinear relationship between soma activity and axonal DA release is the observation that while individual phasic bursts cause rapid desensitization of the DA response, a slow (over multiple seconds) facilitation of striatal DA release occurs following repeated bursts of high-frequency (60 Hz) electrical stimulation of DA neurons, which is presumed to be mediated by increased refilling of the vesicular pool (Montague et al. 2004). Additionally, in vitro experiments have shown that local control by cholinergic interneurons via activation of nicotinic acetylcholine receptors on DA axon terminals can evoke action potential-independent DA release (Threlfell et al. 2012). We summarize these potential mechanisms that may contribute to ramping of DA release in the striatum in Figure 2, and discuss this topic further in the following section.

Taken together, the possibility of reliably identifying DA neurons during in vivo electrophysiological recordings and the availability of newly developed tools to measure DA release with high temporal and spatial precision have opened new avenues to study the relationship between somatic spiking and axonal DA release. However, because of the distinct input-output relationships and anatomical, molecular, and physiological properties of DA neurons, future studies need to take the topographic organization and connectivity of the DA system into consideration. Moreover, the complex dynamics of DA release and reuptake (Liu et al. 2021) and methodological limitations (Sabatini 2019) will have to be taken into account, though advances have been made to correlate DA activity observed through calcium influx to spiking activity (Fleming et al. 2021). Together, these efforts will no doubt shed light on the precise relationship between DA firing and release in different DA pathways.

#### **4. DOPAMINE CELL FIRING AND RELEASE IN REWARD LEARNING AND MOTIVATION**

Given that DA neurons are highly heterogeneous, it is somewhat surprising that leading perspectives on DA cell activity and DA release tend to propose a homogeneous role for these neurons in encoding RPEs (Cohen et al. 2012; Dabney et al. 2020; Eshel et al. 2015, 2016; Gardner et al. 2018; Gershman & Uchida 2019; Kim et al. 2020; Schultz 2019). This idea stems from the foundational studies of Schultz and colleagues (Hollerman & Schultz 1998; Schultz et al. 1993, 1997) performing single-unit recordings in the primate midbrain that have since been replicated with optogenetically assisted tagging of DA neurons in rodents (Bayer & Glimcher 2005; Cohen et al. 2012; Eshel et al. 2015, 2016). These studies have been further corroborated by the ability to record calcium activity in DA

neuron axon terminals and DA release with fluorescent biosensors and have reinforced a long-standing debate regarding DA's actions in the striatum as mediating reward learning via RPE encoding as opposed to a motivational account (Kim et al. 2020, Mohebi et al. 2019, Patriarchi et al. 2018, Sun et al. 2018, Tsutsui-Kimura et al. 2020). While we cannot cover all aspects of this debate (for recent reviews, see Berke 2018, Coddington & Dudman 2019, Collins & Saunders 2020, Dayan & Berridge 2014, Gardner et al. 2018, Gershman & Uchida 2019, Keiflin & Janak 2015, Morales & Margolis 2017, Watabe-Uchida et al. 2017), we wish to offer here the perspective that the circuit architecture of the heterogeneous DA system refutes such a universal role for DA in exclusively mediating reinforcement learning.

DA neurons of the midbrain are embedded in a spiraling network, with inputs from their primary targets in the striatum such that a mediolateral and dorsoventral gradient exists in the projections of DA neurons and, in turn, the projections back onto these neurons from the striatum. This architecture appears to be homologous in rodents and primates (Everitt & Robbins 2013, Haber & Knutson 2010, Haber et al. 2000, Joel & Weiner 2000, Yang et al. 2018), although we recently found that in mice, projections from the NAcMed directly inhibit VTA DA neurons, while projections from the NAcLat provide canonical disinhibition of lateral VTA DA neurons via a GABA neuron intermediary (Yang et al. 2018). This surprising difference in the architecture of the striato-midbrain loop has functional consequences as DA release in the NAcMed and NAcLat has distinct response functions, with NAcMed DA release more closely mirroring behavioral output (e.g., licking for reward) and NAcLat DA release having a phasic cue and reward response (de Jong et al. 2019, Yuan et al. 2019). DA release in this manner should be heterogeneous within a region, for example, a distinction between slow fluctuations in basal DA tone following behavioral output and the fast and rapid rises in DA in response to environmental stimuli (Floresco et al. 2003, Grace & Bunney 1984, Hamid et al. 2016, Hyland et al. 2002, Niv et al. 2007, Schultz 2007). That even on a rapid timescale DA release in distinct NAc subregions appears more tonic or more phasic is in opposition to proposals for unified or universal theories of DA function that have arisen from studies of DA release primarily in the NAc core (Berke 2018, Kim et al. 2020, Lowet et al. 2020, Mohebi et al. 2019). We considered whether this heterogeneity has been overlooked by analyzing the sites of recording for studies making use of optogenetic tagging of DA neurons and in papers recording DA release in the ventral striatum. Overall, it is apparent that there has been a bias to record DA neuron activity in the lateral VTA and DA release in the NAc core in recent investigations of DA circuit function (Figure 1; Supplemental Table 1). But the innervation of the NAc core and NAcMed predominantly by VTA DA neurons originating from the medial VTA has created an apparent mismatch between cell body activity recordings in the lateral VTA and measurements of DA release in the NAc medial shell or NAc core. This bias may arise in part due to the difficulties in recording electrophysiological activity along the midline of rodents, given pronounced blood vessels that impede surgical access. Difficulties in surgical access to medial VTA regions due to blood vessels close to the midline may also exist in nonhuman primates. Additionally, medial VTA DA neurons have unique cellular and electrophysiological properties that may make them resistant to identification with existing optogenetic tagging methods (Farassat et al. 2019; Lammel et al. 2008, 2011; Yang et al. 2018).

Despite an ever increasing appreciation of functional diversity within VTA DA neurons, generally it is conceivable that, in some tasks, DA release in certain striatal regions and DA neuron activity in certain VTA subregions might be reduced to a RPE signal. Given the spiraling architecture of midbrain DA neurons, it appears fair to infer a shared computation among these neurons, but it remains less clear why such an extensive system in the brain would be devoted to uniformly processing and projecting the same computation, RPEs, to all of its downstream targets. Moreover, these correlational metrics of DA activity may look like RPEs but are typically assessed absent functional considerations—that is, whether RPE signals are phenomenological or actually serve to update behavior on a trial-to-trial basis (Dayan & Berridge 2014). The original demonstrations of DA neurons as RPEs arose from the Rescorla-Wagner model of Pavlovian conditioning, which would suggest that RPEs, and in turn DA function, are causal for Pavlovian learning. However, DA function is not required for learning in many Pavlovian situations that have been used to demonstrate the existence of RPE signaling in DA neurons (Day et al. 2007, Flagel et al. 2011). In non-human primates, selective inhibition of DA release in the NAc impaired motivation to earn reward but had no impact on reward learning, suggesting a limited role for DA release in the NAc—or in a yet unspecified subregion of the NAc—to exclusively mediate learning (Vancraeynest et al. 2020).

In one recent compelling report, despite pairing cues directly with DA activation producing learning, individual DA projections to the striatum resulted in drastically different behavioral outcomes in Pavlovian conditioning. Only projections to the NAc core supported classical conditioning as evidenced by a cue-directed approach; projections to the dorsal striatum resulted in cue-generated rotations; and NAcMed projections, arising from the medial VTA, did not establish any cue-conditioned behaviors. Despite these fractionated behavioral functions in Pavlovian conditioning, all of these projections supported equivalent levels of optogenetic intracranial self-stimulation (Saunders et al. 2018). This is surprising because it suggests that even though DA release may uniformly support operant reinforcement, the contribution of the same exact DA signal has drastically different effects in Pavlovian conditioning in differing striatal subregions. Why distinct and disparate functions emerge from a proposed universal RPE signal in even the simplest of Pavlovian tasks remains unclear, as does how these functions could be transferred according to the spiraling architecture of the striato-midbrain system (Russek et al. 2017). Theories of DA function assume RPE has equivalent functions in Pavlovian and operant conditioning, yet it appears that this may not be the case. Although DA release and activity may conform to RPE encoding in these separate striatal regions, predicting the functional consequence of circuit-specific DA release appears nontrivial.

These disparities can potentially be addressed by taking into consideration local control and modulation of DA release within specific subregions of the striatum. Along with the ventromedial to dorsolateral spiraling of midbrain and striatal connectivity, glutamatergic projections from cortex, thalamus, and amygdala exhibit specialized projections throughout the striatum (Berendse et al. 1992, Groenewegen 1988, Heilbronner et al. 2016, Kelley 2004, Mogenson et al. 1980, Voorn et al. 2004, Wright et al. 1996). These projections have been shown to modulate DA release either via direct excitation of DA terminals or through cholinergic interneuron intermediaries (Adrover et al. 2020; Jones et al. 1989, 2010; Kosillo



et al. 2016; Parsons et al. 2007; Pinto et al. 2003; Threlfell et al. 2012). However, there appear to be discrepancies on how different thalamic inputs impact DA release within the striatum, and differences may exist in how such local control arises throughout the striatum (Liu et al. 2021, Parsons et al. 2007, Sulzer et al. 2016, Threlfell et al. 2012). This local control of DA release has largely been proposed as an answer to the generation of ramping DA release (Mohebi et al. 2019), which has been observed in awake behaving animals during motivated behavior, yet a mechanism for ramp generation in vivo remains to be demonstrated (Berke 2018, Collins & Saunders 2020, Hamid et al. 2016, Howe et al. 2013, Kim et al. 2020, Lloyd & Dayan 2015, Roitman et al. 2004, Wassum et al. 2012). A critical and outstanding question is how the topographically distinct composition of amygdala, cortical, and thalamic inputs to different striatal subcompartments affects local control of DA release in vivo, if they contribute at all, especially as many of the demonstrations for such cholinergic interneuron-mediated forms of release have been conducted ex vivo in the dorsal striatum (Adrover et al. 2020, Kosillo et al. 2016, Threlfell et al. 2012).

Exploiting technical advances to fluorescently record the activity of defined striatal inputs or cholinergic interneurons alongside DA release will be essential to resolving the mechanisms underlying afferent control of DA release within the striatum in awake and behaving animals. These approaches can also resolve whether ramping release is an exclusive property of DA release manifested at the axon terminal or whether there is a potential for ramping activity in DA cell bodies, as has been recently observed in a select few DA neurons (Kim et al. 2020). Despite the observation of ramping in a small population of VTA DA neurons, their ramping could be described as RPE signaling (Kim et al. 2020), whereas other researchers have argued that ramping is constrained to striatal DA release where its function is instead to convey value and drive motivated behaviors (Mohebi et al. 2019). It is interesting to note that the authors also identified a small population of ramping VTA DA neurons whose activity could not be captured by RPE models and that the activity of these neurons is most reminiscent of observations of ramping obtained with fast-scan cyclic voltammetry (FSCV) (Howe et al. 2013, Kim et al. 2020). Given the extensive axonal branching of DA neuron axons in the striatum, directly identifying DA release in the striatum as arising from a simultaneously recorded VTA DA neuron is challenging (Aransay et al. 2015, Liu et al. 2021, Matsuda et al. 2009). We are not aware of antidromic approaches being successfully applied to record VTA DA neurons in rodents in such a circuit-specific manner during behavior, which could provide a means to optically record striatal DA release coincident with the spiking activity of these neurons within the VTA (Gadagkar et al. 2016, Guyenet & Aghajanian 1978). These technically challenging approaches may be necessary to resolve the long-standing discrepancy between DA neuron activity and release, and even provide insight into striatal compartment-specific specialization of these mechanisms.

One emerging topic of interest is what DA activity and release convey in the content of what is learned regarding relationships between cues, actions, and outcomes (Gardner et al. 2018, Langdon et al. 2018). DA neurons are proposed to conform to conveying model-free signals, updating the values of actions and states absent any representation but an abstract scalar value term (Bayer & Glimcher 2005, Hollerman & Schultz 1998, Schultz et al. 1997, Sutton & Barto 2018, Waelti et al. 2001). In contrast, in many instances, VTA DA neurons exhibit activity that suggests these neurons have access to richer representations that are

evoked by cues such as the delay to reward, links between previously neutral sounds or lights, and precise sensory qualities of the upcoming reward (Sadacca et al. 2016; Stalnaker et al. 2019; Takahashi & Schoenbaum 2016; Takahashi et al. 2011, 2016, 2017). The ability of DA neurons to respond in this fashion suggests that they are more actively involved in generating very precise representations in contrast to most computational models of the function of these neurons (Dayan & Berridge 2014). Indeed, manipulations of VTA DA neurons alter behavior in tasks designed specifically to require the use of these model-based representations (Keiflin et al. 2019; Sharpe et al. 2017, 2020). For instance, learning induced by optogenetic activation of VTA DA neurons is sensitive to manipulations of the value of the predicted reward (Keiflin et al. 2019). Moreover, it appears that it is VTA, but not SNc, DA neurons that are critical for these model-based signals (Keiflin et al. 2019). This specialization in the function of distinct DA projections in model-based versus model-free learning makes difficult the assumption that DA signals are merely relayed from mesolimbic to striatonigral DA systems with time (Belin-Rauscent et al. 2012; Everitt & Robbins 2013; Willuhn et al. 2012, 2014). Indeed, increasing attention is being given to the diversity of computational metrics encoded by individual VTA DA neurons (Engelhard et al. 2019, Kremer et al. 2020). Despite this, we have little information about how rich, model-based information is transmitted throughout the striatum, as the primary evidence for such sensitivity has largely been demonstrated in the NAc core when the physiological state of the animal has been shifted (Aitken et al. 2016, Cone et al. 2016, Hsu et al. 2020). Moreover, it is unclear how model-based information itself is conveyed to VTA DA neurons to even allow such a complex computation (Langdon et al. 2018). It will be critical to disentangle the circuit-specific routing of model-based and model-free signals throughout the striatum to better refine our understanding of these neurons in motivated behavior. Generating and testing these hypotheses will require increasingly sophisticated behavioral approaches in tandem with circuit-specific recording of VTA DA neurons and DA release in defined striatal subregions.

## 5. DOPAMINE CELL FIRING AND RELEASE IN AVERSION

The processing of aversive stimuli by DA neurons has historically been a source of controversy. Foundational studies described a homogeneous population of DA neurons that were uniformly inhibited by aversive events or the omission of predicted reward (Hollerman & Schultz 1998; Kim et al. 2010, 2012; Ljungberg et al. 1991, 1992; Schultz et al. 1993; Ungless et al. 2004). This pattern of response supported the notion that these neurons participate in RPE encoding. However, recordings of putative DA neurons in the ventromedial VTA found that these neurons were excited by the presentation of a cue that predicted mild electric shock (Guarraci & Kapp 1999). Further, antidromic identification of prefrontal cortex (PFC)-projecting putative DA neurons, which are primarily found in the medial VTA, also found that most of these neurons are excited by a noxious tail pinch (Mantz et al. 1989). Despite these findings, heterogeneity in response to aversive stimuli was controversial, as these studies primarily made use of electrophysiological characteristics to identify DA neurons, which made certainty regarding their neurochemical identity difficult (Margolis et al. 2006, Ungless & Grace 2012) and left open the possibility that, even if a small number of electrophysiologically identified DA neurons responded to aversive

stimuli (Bromberg-Martin et al. 2010, Matsumoto & Hikosaka 2009, Mirenowicz & Schultz 1996), these may in fact not be dopaminergic. To clarify this discrepancy, Brischoux et al. (2009) juxtacellularly labeled VTA neurons that were recorded while electric shocks were delivered to allow the confirmation of these neurons' neurochemical identity with immunohistochemistry for tyrosine hydroxylase, the enzyme responsible for DA synthesis. Surprisingly, aversion-excited DA neurons were primarily observed in the medial VTA, whereas aversion-inhibited DA neurons were found in the lateral VTA (Brischoux et al. 2009). Given that medial VTA DA neurons primarily project to the PFC or NAcMed, this finding suggested circuit-specific specialization for the processing of aversion within the DA system (Brischoux et al. 2009, Lammel et al. 2008).

The suggestion that subsets of VTA DA neurons selectively process aversion was supported by the observation that recording DA release using microdialysis primarily detected increases in DA turnover in the NAc and PFC following restraint or electric shock (Abercrombie et al. 1989, Jackson & Moghaddam 2004, Young 2004). However, the DA response in the NAc to these stressors appears to depend on controllability, as it is diminished after repeated exposure unless the animal is given an opportunity to perform an action to terminate the aversive stimulus (Cabib & Puglisi-Allegra 1994, Imperato et al. 1993). However, the minute-long timescale of microdialysis made it difficult to link increases in DA release to the aversive experience itself or potential relief arising from its termination (Mirenowicz & Schultz 1996, Salinas-Hernández et al. 2018). Recording DA release on a seconds-long timescale with FSCV indicated that, while aversive foot shocks evoke DA release throughout the striatum, DA release in the NAcMed was selectively increased in response to shock-predictive cues (Badrinarayan et al. 2012, Budygin et al. 2012). To clarify this discrepancy between the proposed specialization of VTA DA neurons in encoding aversion versus an apparent general increase in DA in the striatum to aversive foot shock, we systematically mapped DA release throughout the NAc during aversive Pavlovian conditioning (de Jong et al. 2019) (Figure 3). We found that aversive foot shocks, as well as cues that predict them, selectively evoked DA release in the ventral NAcMed, while NAcLat DA release was decreased by aversive foot shock (de Jong et al. 2019). The selective increase in DA release in the ventral NAcMed suggests that this region can be divided into distinct dorsal and ventral compartments, as dorsal NAcMed DA release is decreased by noxious stimuli such as a bitter-tasting quinine solution (Roitman et al. 2008, Yuan et al. 2019) and increased by the termination or omission of an aversive stimulus (Budygin et al. 2012, de Jong et al. 2019). This dorsal reward and ventral aversion processing gradient in the NAcMed is also supported by the finding that direct activation of dorsal dynorphin-producing NAcMed neurons induces place preference, while ventral dynorphinergic NAcMed activation elicits place aversion (Al-Hasani et al. 2015).

While advances have been made in defining DA subsystems that are excited or inhibited by aversive stimuli, further resolving the functions of VTA DA neuron activity and DA release in aversively motivated behaviors will require exploiting circuit-specific approaches to record and manipulate neural activity within these specialized projection-defined systems. Existing evidence suggests that VTA DA neurons are excited by and facilitate learning about aversive stimuli, which runs counter to RPE theories, but other recent studies suggest VTA DA neurons are inhibited to facilitate learning about aversive stimuli (Cai et al. 2020, Jo

et al. 2018). Perhaps taking advantage of more ethologically relevant behavioral paradigms can clarify the apparent discrepant contribution of VTA DA neurons to aversively motivated behavior (Dennis et al. 2021). Nonetheless, it appears that taking into account the precise striatal projection target of VTA DA neurons refines their contributions to aversion, and it will be critical for future studies to exploit this projection-defined heterogeneity.

## 6. CONCLUSIONS

Within the past decade there have been numerous technological advances that have facilitated a greater appreciation of the heterogeneity of the DA system. Even within the mesolimbic DA system separate subpopulations of VTA DA neurons projecting to distinct NAc subregions are specialized for the production and transmittance of defined behaviorally relevant signals. However, this poses a challenge to theories of DA function that posit homogeneous roles for mesolimbic DA neurons in encoding RPEs. VTA DA neurons also appear to exhibit a wide array of computations that run counter to the original proposition of these neurons exclusively encoding model-free variables, and it remains to be demonstrated whether these neurons integrate inputs to generate rich model-based predictions or whether this information is merely inherited from upstream brain regions. Perhaps an overemphasis on investigating DA neurons in the lateral VTA and DA release in the NAc core has resulted in preliminary universal and sometimes controversial theories of mesolimbic DA function that still require proof for their functions throughout the striatum. Advancing methodologies to record from circuit- and cell type–defined VTA DA neurons in tandem with sophisticated and elegant behavioral approaches will be essential to move the field forward and resolve these outstanding issues. Recognition of this circuit-specific specialization of DA function will be necessary to advance our understanding of the role of these neurons in health and disease.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## ACKNOWLEDGMENTS

We sincerely apologize to all of our colleagues whose work we were unable to discuss due to space and citation limitations. S.L. is a Weill Neurohub Investigator, John P. Stock Faculty Fellow, and Rita Allen Scholar. This work was supported by National Institutes of Health grants (R01-DA042889 and R01-MH123246 to S.L., F32-MH127792 to K.M.F.), the One Mind Foundation (to S.L.), a NARSAD Young Investigator Award (27936 to J.W.d.J.), and the Wayne and Gladys Valley Foundation (to S.L.).

## Glossary

### DA

dopamine

### NAc

nucleus accumbens

### VTA

ventral tegmental area

**SNC**

substantia nigra pars compacta

**Opto-tagging**

expressing optogenetic proteins in a cell type–specific manner to confirm the identity of in vivo electrophysiologically recorded cells

**Ramping**

gradual increases in extracellular dopamine that increase toward a peak as reward is earned or actions are performed

**Reward prediction error (RPE)**

a computation that can be described as the mismatch between how much reward was expected and how much reward was received

**TH**

tyrosine hydroxylase

**DAT**

dopamine transporter

**NAcMed**

nucleus accumbens medial shell

**NAcLat**

nucleus accumbens lateral shell

**Reinforcement learning**

a process by which agents refine their behavior by iteratively acting upon the environment and receiving feedback via prediction errors

**Pavlovian conditioning**

also called classical conditioning, a procedure by which stimuli are associated with the delivery of outcomes absent any action requirement

**FSCV**

fast-scan cyclic voltammetry

**Model-free**

a computation or representation that is purely value based and abstract

**Model-based**

a computation or representation that is highly complex and relies on a set of relationships between stimuli, actions, and outcomes

**PFC**

prefrontal cortex

## LITERATURE CITED

- Abercrombie ED, Keefe KA, DiFrischia DS, Zigmond MJ. 1989. Differential effect of stress on in vivo dopamine release in striatum, nucleus accumbens, and medial frontal cortex. *J. Neurochem.* 52(5):1655–58 [PubMed: 2709017]
- Adrover MF, Shin JH, Quiroz C, Ferré S, Lemos JC, Alvarez VA. 2020. Prefrontal cortex-driven dopamine signals in the striatum show unique spatial and pharmacological properties. *J. Neurosci.* 40(39):7510–22 [PubMed: 32859717]
- Aitken TJ, Greenfield VY, Wassum KM. 2016. Nucleus accumbens core dopamine signaling tracks the need-based motivational value of food-paired cues. *J. Neurochem.* 136(5):1026–36 [PubMed: 26715366]
- Albanese A, Minciacchi D. 1983. Organization of the ascending projections from the ventral tegmental area: a multiple fluorescent retrograde tracer study in the rat. *J. Comp. Neurol.* 216(4):406–20 [PubMed: 6308073]
- Al-Hasani R, McCall JG, Shin G, Gomez AM, Schmitz GP, et al. 2015. Distinct subpopulations of nucleus accumbens dynorphin neurons drive aversion and reward. *Neuron* 87(5):1063–77 [PubMed: 26335648]
- Aransay A, Rodríguez-López C, García-Amado M, Clascá F, Prensa L. 2015. Long-range projection neurons of the mouse ventral tegmental area: a single-cell axon tracing analysis. *Front. Neuroanat.* 9:59 [PubMed: 26042000]
- Badrinarayan A, Wescott SA, Vander Weele CM, Saunders BT, Couturier BE, et al. 2012. Aversive stimuli differentially modulate real-time dopamine transmission dynamics within the nucleus accumbens core and shell. *J. Neurosci.* 32(45):15779–90 [PubMed: 23136417]
- Bayer HM, Glimcher PW. 2005. Midbrain dopamine neurons encode a quantitative reward prediction error signal. *Neuron* 47(1):129–41 [PubMed: 15996553]
- Beier KT, Steinberg EE, DeLoach KE, Xie S, Miyamichi K, et al. 2015. Circuit architecture of VTA dopamine neurons revealed by systematic input-output mapping. *Cell.* 162(3):622–34 [PubMed: 26232228]
- Belin-Rauscent A, Everitt BJ, Belin D. 2012. Intrastratial shifts mediate the transition from drug-seeking actions to habits. *Biol. Psychiatry* 72(5):343–45 [PubMed: 22872011]
- Berendse HW, Graaf YG-D, Groenewegen HJ. 1992. Topographical organization and relationship with ventral striatal compartments of prefrontal corticostriatal projections in the rat. *J. Comp. Neurol.* 316(3):314–47 [PubMed: 1577988]
- Berke JD. 2018. What does dopamine mean? *Nat. Neurosci.* 21(6):787–93 [PubMed: 29760524]
- Bjorklund A, Dunnett SB. 2007. Dopamine neuron systems in the brain: an update. *Trends Neurosci.* 30:194–202 [PubMed: 17408759]
- Breton JM, Charbit AR, Snyder BJ, Fong PTK, Dias EV, et al. 2019. Relative contributions and mapping of ventral tegmental area dopamine and GABA neurons by projection target in the rat. *J. Comp. Neurol.* 527(5):916–41 [PubMed: 30393861]
- Brischoux F, Chakraborty S, Brierley DI, Ungless MA. 2009. Phasic excitation of dopamine neurons in ventral VTA by noxious stimuli. *PNAS* 106(12):4894–99 [PubMed: 19261850]
- Bromberg-Martin ES, Matsumoto M, Hikosaka O. 2010. Dopamine in motivational control: rewarding, aversive, and alerting. *Neuron* 68(5):815–34 [PubMed: 21144997]
- Budygin EA, Park J, Bass CE, Grinevich VP, Bonin KD, Wightman RM. 2012. Aversive stimulus differentially triggers subsecond dopamine release in reward regions. *Neuroscience* 201:331–37 [PubMed: 22108611]
- Cabib S, Puglisi-Allegra S. 1994. Opposite responses of mesolimbic dopamine system to controllable and uncontrollable aversive experiences. *J. Neurosci.* 14(5 Part 2):3333–40 [PubMed: 8182476]
- Cai LX, Pizano K, Gundersen GW, Hayes CL, Fleming WT, et al. 2020. Distinct signals in medial and lateral VTA dopamine neurons modulate fear extinction at different times. *eLife* 9:e54936 [PubMed: 32519951]
- Cardozo Pinto DF, Lammel S. 2018. Viral vector strategies for investigating midbrain dopamine circuits underlying motivated behaviors. *Pharmacol. Biochem. Behav.* 174:23–32 [PubMed: 28257849]

- Castro DC, Bruchas MR. 2019. A motivational and neuropeptidergic hub: anatomical and functional diversity within the nucleus accumbens shell. *Neuron* 102(3):529–52 [PubMed: 31071288]
- Coddington LT, Dudman JT. 2019. Learning from action: reconsidering movement signaling in midbrain dopamine neuron activity. *Neuron* 104(1):63–77 [PubMed: 31600516]
- Cohen JY, Haesler S, Vong L, Lowell BB, Uchida N. 2012. Neuron-type-specific signals for reward and punishment in the ventral tegmental area. *Nature* 482(7383):85–88 [PubMed: 22258508]
- Collins AL, Saunders BT. 2020. Heterogeneity in striatal dopamine circuits: form and function in dynamic reward seeking. *J. Neurosci. Res.* 98(6):1046–69 [PubMed: 32056298]
- Cone JJ, Fortin SM, McHenry JA, Stuber GD, McCutcheon JE, Roitman MF. 2016. Physiological state gates acquisition and expression of mesolimbic reward prediction signals. *PNAS* 113(7):1943–48 [PubMed: 26831116]
- Dabney W, Kurth-Nelson Z, Uchida N, Starkweather CK, Hassabis D, et al. 2020. A distributional code for value in dopamine-based reinforcement learning. *Nature* 577(7792):671–75 [PubMed: 31942076]
- Day JJ, Roitman MF, Wightman RM, Carelli RM. 2007. Associative learning mediates dynamic shifts in dopamine signaling in the nucleus accumbens. *Nat. Neurosci.* 10(8):1020–28 [PubMed: 17603481]
- Dayan P, Berridge KC. 2014. Model-based and model-free Pavlovian reward learning: reevaluation, revision, and revelation. *Cogn. Affect. Behav. Neurosci.* 14(2):473–92 [PubMed: 24647659]
- de Jong JW, Afjei SA, Pollak Dorocic I, Peck JR, Liu C, et al. 2019. A neural circuit mechanism for encoding aversive stimuli in the mesolimbic dopamine system. *Neuron* 101(1):133–51.e7 [PubMed: 30503173] This study demonstrates that dopamine release dynamics are specialized in distinct striatal areas to encode reward or aversion.
- Dennis EJ, El Hady A, Michael A, Clemens A, Tervo DRG, et al. 2021. Systems neuroscience of natural behaviors in rodents. *J. Neurosci.* 41(5):911–19 [PubMed: 33443081]
- Engelhard B, Finkelstein J, Cox J, Fleming W, Jang HJ, et al. 2019. Specialized coding of sensory, motor and cognitive variables in VTA dopamine neurons. *Nature* 570(7762):509–13 [PubMed: 31142844] This study reveals novel computational metrics within individual dopamine neurons by exploiting longitudinal 2-photon imaging.
- Eshel N, Bukwich M, Rao V, Hemmelder V, Tian J, Uchida N. 2015. Arithmetic and local circuitry underlying dopamine prediction errors. *Nature* 525(7568):243–46 [PubMed: 26322583]
- Eshel N, Tian J, Bukwich M, Uchida N. 2016. Dopamine neurons share common response function for reward prediction error. *Nat. Neurosci.* 19(3):479–86 [PubMed: 26854803]
- Everitt BJ, Robbins TW. 2013. From the ventral to the dorsal striatum: devolving views of their roles in drug addiction. *Neurosci. Biobehav. Rev* 37(9 Pt. A):1946–54 [PubMed: 23438892]
- Faget L, Osakada F, Duan J, Ressler R, Johnson AB, et al. 2016. Afferent inputs to neurotransmitter-defined cell types in the ventral tegmental area. *Cell Rep.* 15(12):2796–808 [PubMed: 27292633]
- Farassat N, Costa KM, Stojanovic S, Albert S, Kovacheva L, et al. 2019. In vivo functional diversity of midbrain dopamine neurons within identified axonal projections. *eLife* 8:e48408 [PubMed: 31580257] This study provides detailed mapping of electrophysiological properties of location- and projection-defined dopamine neurons in vivo.
- Flagel SB, Clark JJ, Robinson TE, Mayo L, Czuj A, et al. 2011. A selective role for dopamine in stimulus-reward learning. *Nature* 469(7328):53–57 [PubMed: 21150898]
- Fleming W, Jewell S, Engelhard B, Witten DM, Witten IB. 2021. Inferring spikes from calcium imaging in dopamine neurons. *PLOS ONE* 16(6):e0252345 [PubMed: 34086726]
- Floresco SB, West AR, Ash B, Moore H, Grace AA. 2003. Afferent modulation of dopamine neuron firing differentially regulates tonic and phasic dopamine transmission. *Nat. Neurosci.* 6(9):968–73 [PubMed: 12897785]
- Franklin KBJ, Paxinos G. 2001. Paxinos and Franklin's The Mouse Brain in Stereotaxic Coordinates. Amsterdam: Academic Press. 2nd ed.
- Gadagkar V, Puzerey PA, Chen R, Baird-Daniel E, Farhang AR, Goldberg JH. 2016. Dopamine neurons encode performance error in singing birds. *Science* 354(6317):1278–82 [PubMed: 27940871]

- Gardner MPH, Schoenbaum G, Gershman SJ. 2018. Rethinking dopamine as generalized prediction error. *Proc. Biol. Sci.* 285(1891):20181645 [PubMed: 30464063]
- Gershman SJ, Uchida N. 2019. Believing in dopamine. *Nat. Rev. Neurosci.* 20(11):703–14 [PubMed: 31570826]
- Grace AA, Bunney BS. 1984. The control of firing pattern in nigral dopamine neurons: burst firing. *J. Neurosci.* 4(11):2877–90 [PubMed: 6150071]
- Groenewegen HJ. 1988. Organization of the afferent connections of the mediodorsal thalamic nucleus in the rat, related to the mediodorsal-prefrontal topography. *Neuroscience* 24(2):379–431 [PubMed: 2452377]
- Guarraci FA, Kapp BS. 1999. An electrophysiological characterization of ventral tegmental area dopaminergic neurons during differential Pavlovian fear conditioning in the awake rabbit. *Behav. Brain Res.* 99(2):169–79 [PubMed: 10512583]
- Guyenet PG, Aghajanian GK. 1978. Antidromic identification of dopaminergic and other output neurons of the rat substantia nigra. *Brain Res.* 150(1):69–84 [PubMed: 78748]
- Haber SN, Fudge JL, McFarland NR. 2000. Striatonigrostriatal pathways in primates form an ascending spiral from the shell to the dorsolateral striatum. *J. Neurosci.* 20(6):2369–82 [PubMed: 10704511]
- Haber SN, Knutson B. 2010. The reward circuit: linking primate anatomy and human imaging. *Neuropsychopharmacology* 35(1):4–26 [PubMed: 19812543]
- Hamid AA, Pettibone JR, Mabrouk OS, Hetrick VL, Schmidt R, et al. 2016. Mesolimbic dopamine signals the value of work. *Nat. Neurosci.* 19(1):117–26 [PubMed: 26595651]
- Heilbronner SR, Rodriguez-Romaguera J, Quirk GJ, Groenewegen HJ, Haber SN. 2016. Circuit-based corticostriatal homologies between rat and primate. *Biol. Psychiatry* 80(7):509–21 [PubMed: 27450032] This anatomical study demonstrates that homologous striatal compartments in rodents and primates receive specialized glutamatergic inputs.
- Heymann G, Jo YS, Reichard KL, McFarland N, Chavkin C, et al. 2020. Synergy of distinct dopamine projection populations in behavioral reinforcement. *Neuron* 105(5):909–20.e5 [PubMed: 31879163]
- Hollerman JR, Schultz W. 1998. Dopamine neurons report an error in the temporal prediction of reward during learning. *Nat. Neurosci.* 1(4):304–9 [PubMed: 10195164]
- Howe MW, Tierney PL, Sandberg SG, Phillips PEM, Graybiel AM. 2013. Prolonged dopamine signalling in striatum signals proximity and value of distant rewards. *Nature* 500(7464):575–79 [PubMed: 23913271]
- Hsu TM, Bazzino P, Hurh SJ, Konanur VR, Roitman JD, Roitman MF. 2020. Thirst recruits phasic dopamine signaling through subfornical organ neurons. *PNAS* 117(48):30744–54 [PubMed: 33199591]
- Hyland BI, Reynolds JNJ, Hay J, Perk CG, Miller R. 2002. Firing modes of midbrain dopamine cells in the freely moving rat. *Neuroscience* 114(2):475–92 [PubMed: 12204216]
- Ikemoto S. 2007. Dopamine reward circuitry: two projection systems from the ventral midbrain to the nucleus accumbens-olfactory tubercle complex. *Brain Res. Rev.* 56(1):27–78 [PubMed: 17574681]
- Imperato A, Cabib S, Puglisi-Allegra S. 1993. Repeated stressful experiences differently affect the time-dependent responses of the mesolimbic dopamine system to the stressor. *Brain Res.* 601(1–2):333–36 [PubMed: 8431783]
- Jackson ME, Moghaddam B. 2004. Stimulus-specific plasticity of prefrontal cortex dopamine neurotransmission. *J. Neurochem.* 88(6):1327–34 [PubMed: 15009632]
- Jo YS, Heymann G, Zweifel LS. 2018. Dopamine neurons reflect the uncertainty in fear generalization. *Neuron* 100(4):916–25.e3 [PubMed: 30318411]
- Joel D, Weiner I. 2000. The connections of the dopaminergic system with the striatum in rats and primates: an analysis with respect to the functional and compartmental organization of the striatum. *Neuroscience* 96(3):451–74 [PubMed: 10717427]
- Jones JL, Day JJ, Aragona BJ, Wheeler RA, Wightman RM, Carelli RM. 2010. Basolateral amygdala modulates terminal dopamine release in the nucleus accumbens and conditioned responding. *Biol. Psychiatry* 67(8):737–44 [PubMed: 20044074]



- Jones MW, Kilpatrick IC, Phillipson OT. 1989. Regulation of dopamine function in the nucleus accumbens of the rat by the thalamic paraventricular nucleus and adjacent midline nuclei. *Exp. Brain Res.* 76(3):572–80 [PubMed: 2676574]
- Juarez B, Han M-H. 2016. Diversity of dopaminergic neural circuits in response to drug exposure. *Neuropsychopharmacology* 41(10):2424–46 [PubMed: 26934955]
- Keiflin R, Janak PH. 2015. Dopamine prediction errors in reward learning and addiction: from theory to neural circuitry. *Neuron* 88(2):247–63 [PubMed: 26494275]
- Keiflin R, Pribut HJ, Shah NB, Janak PH. 2019. Ventral tegmental dopamine neurons participate in reward identity predictions. *Curr. Biol.* 29(1):93–103.e3 [PubMed: 30581025] This study delineates the actions of ventral tegmental area dopamine neurons in forming rich, model-based representations to support learning.
- Kelley AE. 2004. Ventral striatal control of appetitive motivation: role in ingestive behavior and reward-related learning. *Neurosci. Biobehav. Rev.* 27(8):765–76 [PubMed: 15019426]
- Kim HR, Malik AN, Mikhael JG, Bech P, Tsutsui-Kimura I, et al. 2020. A unified framework for dopamine signals across timescales. *Cell* 183(6):1600–16.e25 [PubMed: 33248024] This study argues that dopamine activity and release is best described as reward prediction error.
- Kim Y, Wood J, Moghaddam B. 2012. Coordinated activity of ventral tegmental neurons adapts to appetitive and aversive learning. *PLOS ONE* 7(1):e29766 [PubMed: 22238652]
- Kim Y-B, Matthews M, Moghaddam B. 2010. Putative  $\gamma$ -aminobutyric acid neurons in the ventral tegmental area have a similar pattern of plasticity as dopamine neurons during appetitive and aversive learning. *Eur. J. Neurosci.* 32(9):1564–72 [PubMed: 21040517]
- Kosillo P, Zhang Y-F, Threlfell S, Cragg SJ. 2016. Cortical control of striatal dopamine transmission via striatal cholinergic interneurons. *Cereb. Cortex* 26(11):4160–69 [PubMed: 27566978]
- Krabbe S, Duda J, Schieman J, Poetschke C, Schneider G, et al. 2015. Increased dopamine D2 receptor activity in the striatum alters the firing pattern of dopamine neurons in the ventral tegmental area. *PNAS* 112(12):E1498–506 [PubMed: 25675529]
- Kramer DJ, Aisenberg EE, Kosillo P, Friedmann D, Stafford DA, et al. 2021. Generation of a DAT-P2A-Flpo mouse line for intersectional genetic targeting of dopamine neuron subpopulations. *Cell Rep.* 35(6):109123 [PubMed: 33979604]
- Kreitzer AC. 2009. Physiology and pharmacology of striatal neurons. *Annu. Rev. Neurosci.* 32:127–47 [PubMed: 19400717]
- Kremer Y, Flakowski J, Rohner C, Lüscher C. 2020. Context-dependent multiplexing by individual VTA dopamine neurons. *J. Neurosci.* 40(39):7489–509 [PubMed: 32859713] The authors demonstrate diverse behaviorally relevant computations within the spiking of individual dopamine neurons.
- Lammel S, Hetzel A, Häckel O, Jones I, Liss B, Roeper J. 2008. Unique properties of mesoprefrontal neurons within a dual mesocorticolimbic dopamine system. *Neuron* 57(5):760–73 [PubMed: 18341995]
- Lammel S, Ion DI, Roeper J, Malenka RC. 2011. Projection-specific modulation of dopamine neuron synapses by aversive and rewarding stimuli. *Neuron* 70(5):855–62 [PubMed: 21658580]
- Lammel S, Lim BK, Ran C, Huang KW, Betley MJ, et al. 2012. Input-specific control of reward and aversion in the ventral tegmental area. *Nature* 491(7423):212–17 [PubMed: 23064228]
- Langdon AJ, Sharpe MJ, Schoenbaum G, Niv Y. 2018. Model-based predictions for dopamine. *Curr. Opin. Neurobiol.* 49:1–7 [PubMed: 29096115]
- Lerner TN, Shilyansky C, Davidson TJ, Evans KE, Beier KT, et al. 2015. Intact-brain analyses reveal distinct information carried by SNc dopamine subcircuits. *Cell.* 162(3):635–47 [PubMed: 26232229]
- Liu C, Goel P, Kaeser PS. 2021. Spatial and temporal scales of dopamine transmission. *Nat. Rev. Neurosci.* 22(6):345–58 [PubMed: 33837376]
- Liu C, Kershberg L, Wang J, Schneeberger S, Kaeser PS. 2018. Dopamine secretion is mediated by sparse active zone-like release sites. *Cell.* 172(4):706–18.e15 [PubMed: 29398114] This is a foundational study detailing unique challenges in relating activity of dopamine neurons to subsequent dopamine release.

- Liu Y, Jean-Richard-Dit-Bressel P, Yau JO-Y, Willing A, Prasad AA, et al. 2020. The mesolimbic dopamine activity signatures of relapse to alcohol-seeking. *J. Neurosci.* 40(33):6409–27 [PubMed: 32669355]
- Ljungberg T, Apicella P, Schultz W. 1991. Responses of monkey midbrain dopamine neurons during delayed alternation performance. *Brain Res.* 567(2):337–41 [PubMed: 1817739]
- Ljungberg T, Apicella P, Schultz W. 1992. Responses of monkey dopamine neurons during learning of behavioral reactions. *J. Neurophysiol.* 67(1):145–63 [PubMed: 1552316]
- Lloyd K, Dayan P. 2015. Tamping ramping: algorithmic, implementational, and computational explanations of phasic dopamine signals in the accumbens. *PLOS Comput. Biol.* 11(12):e1004622 [PubMed: 26699940]
- Lowet AS, Zheng Q, Matias S, Drugowitsch J, Uchida N. 2020. Distributional reinforcement learning in the brain. *Trends Neurosci.* 43(12):980–97 [PubMed: 33092893]
- Mantz J, Thierry AM, Glowinski J. 1989. Effect of noxious tail pinch on the discharge rate of mesocortical and mesolimbic dopamine neurons: selective activation of the mesocortical system. *Brain Res.* 476(2):377–81 [PubMed: 2702475]
- Margolis EB, Lock H, Hjelmstad GO, Fields HL. 2006. The ventral tegmental area revisited: Is there an electrophysiological marker for dopaminergic neurons? *J. Physiol.* 577:907–24 [PubMed: 16959856]
- Matsuda W, Furuta T, Nakamura KC, Hioki H, Fujiyama F, et al. 2009. Single nigrostriatal dopaminergic neurons form widely spread and highly dense axonal arborizations in the neostriatum. *J. Neurosci.* 29(2):444–53 [PubMed: 19144844]
- Matsumoto M, Hikosaka O. 2009. Two types of dopamine neuron distinctly convey positive and negative motivational signals. *Nature* 459(7248):837–41 [PubMed: 19448610]
- Menegas W, Bergan JF, Ogawa SK, Isogai Y, Venkataraju KU, et al. 2015. Dopamine neurons projecting to the posterior striatum form an anatomically distinct subclass. *eLife* 4:e10032 [PubMed: 26322384]
- Mirenowicz J, Schultz W. 1996. Preferential activation of midbrain dopamine neurons by appetitive rather than aversive stimuli. *Nature* 379(6564):449–51 [PubMed: 8559249]
- Mogenson GJ, Jones DL, Yim CY. 1980. From motivation to action: functional interface between the limbic system and the motor system. *Prog. Neurobiol.* 14(2–3):69–97 [PubMed: 6999537]
- Mohebi A, Pettibone JR, Hamid AA, Wong J-MT, Vinson LT, et al. 2019. Dissociable dopamine dynamics for learning and motivation. *Nature* 570(7759):65–70 [PubMed: 31118513]
- Montague PR, McClure SM, Baldwin PR, Phillips PEM, Budygin EA, et al. 2004. Dynamic gain control of dopamine delivery in freely moving animals. *J. Neurosci.* 24(7):1754–59 [PubMed: 14973252]
- Morales M, Margolis EB. 2017. Ventral tegmental area: cellular heterogeneity, connectivity and behaviour. *Nat. Rev. Neurosci.* 18(2):73–85 [PubMed: 28053327]
- Nestler EJ, Carlezon WA. 2006. The mesolimbic dopamine reward circuit in depression. *Biol. Psychiatry* 59(12):1151–59 [PubMed: 16566899]
- Nieh EH, Vander Weele CM, Matthews GA, Presbrey KN, Wichmann R, et al. 2016. Inhibitory input from the lateral hypothalamus to the ventral tegmental area disinhibits dopamine neurons and promotes behavioral activation. *Neuron* 90(6):1286–98 [PubMed: 27238864]
- Niv Y, Daw ND, Joel D, Dayan P. 2007. Tonic dopamine: opportunity costs and the control of response vigor. *Psychopharmacology* 191(3):507–20 [PubMed: 17031711]
- Otomo K, Perkins J, Kulkarni A, Stojanovic S, Roeper J, Paladini CA. 2020. In vivo patch-clamp recordings reveal distinct subthreshold signatures and threshold dynamics of midbrain dopamine neurons. *Nat. Commun.* 11(1):6286 [PubMed: 33293613]
- Paladini CA, Roeper J. 2014. Generating bursts (and pauses) in the dopamine midbrain neurons. *Neuroscience* 282:109–21 [PubMed: 25073045]
- Parsons MP, Li S, Kirouac GJ. 2007. Functional and anatomical connection between the paraventricular nucleus of the thalamus and dopamine fibers of the nucleus accumbens. *J. Comp. Neurol.* 500(6):1050–63 [PubMed: 17183538]

- Patriarchi T, Cho JR, Merten K, Howe MW, Marley A, et al. 2018. Ultrafast neuronal imaging of dopamine dynamics with designed genetically encoded sensors. *Science* 360(6396):eaat4422 [PubMed: 29853555]
- Paxinos G, Watson C. 2007. *The Rat Brain in Stereotaxic Coordinates*. San Diego, CA: Academic. 6th ed.
- Pinto A, Jankowski M, Sesack SR. 2003. Projections from the paraventricular nucleus of the thalamus to the rat prefrontal cortex and nucleus accumbens shell: ultrastructural characteristics and spatial relationships with dopamine afferents. *J. Comp. Neurol.* 459(2):142–55 [PubMed: 12640666]
- Poulin J-F, Caronia G, Hofer C, Cui Q, Helm B, et al. 2018. Mapping projections of molecularly defined dopamine neuron subtypes using intersectional genetic approaches. *Nat. Neurosci.* 21(9):1260–71 [PubMed: 30104732]
- Poulin J-F, Gaertner Z, Moreno-Ramos OA, Awatramani R. 2020. Classification of midbrain dopamine neurons using single-cell gene expression profiling approaches. *Trends Neurosci.* 43(3):155–69 [PubMed: 32101709]
- Qi J, Zhang S, Wang H-L, Wang H, de Jesus Aceves Buendia J, et al. 2014. A glutamatergic reward input from the dorsal raphe to ventral tegmental area dopamine neurons. *Nat. Commun.* 5:5390 [PubMed: 25388237]
- Robinson TE, Berridge KC. 2008. The incentive sensitization theory of addiction: some current issues. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* 363(1507):3137–46 [PubMed: 18640920]
- Roeper J 2013. Dissecting the diversity of midbrain dopamine neurons. *Trends Neurosci.* 36(6):336–42 [PubMed: 23582338]
- Roitman MF, Stuber GD, Phillips PEM, Wightman RM, Carelli RM. 2004. Dopamine operates as a subsecond modulator of food seeking. *J. Neurosci.* 24(6):1265–71 [PubMed: 14960596]
- Roitman MF, Wheeler RA, Wightman RM, Carelli RM. 2008. Real-time chemical responses in the nucleus accumbens differentiate rewarding and aversive stimuli. *Nat. Neurosci.* 11(12):1376–77 [PubMed: 18978779]
- Russek EM, Momennejad I, Botvinick MM, Gershman SJ, Daw ND. 2017. Predictive representations can link model-based reinforcement learning to model-free mechanisms. *PLOS Comput. Biol.* 13(9):e1005768 [PubMed: 28945743]
- Sabatini BL. 2019. The impact of reporter kinetics on the interpretation of data gathered with fluorescent reporters. *bioRxiv* 834895. 10.1101/834895
- Sabatini BL, Tian L. 2020. Imaging neurotransmitter and neuromodulator dynamics in vivo with genetically encoded indicators. *Neuron* 108(1):17–32 [PubMed: 33058762]
- Sadacca BF, Jones JL, Schoenbaum G. 2016. Midbrain dopamine neurons compute inferred and cached value prediction errors in a common framework. *eLife* 5:e13665 [PubMed: 26949249]
- Salinas-Hernández XI, Vogel P, Betz S, Kalisch R, Sigurdsson T, Duvarci S. 2018. Dopamine neurons drive fear extinction learning by signaling the omission of expected aversive outcomes. *eLife* 7:e38818 [PubMed: 30421719]
- Saunders BT, Richard JM, Margolis EB, Janak PH. 2018. Dopamine neurons create Pavlovian conditioned stimuli with circuit-defined motivational properties. *Nat. Neurosci.* 21(8):1072–83 [PubMed: 30038277] This elegant study delineates circuit-specific behavioral functions of dopamine neurons in Pavlovian conditioning.
- Schultz W 2007. Multiple dopamine functions at different time courses. *Annu. Rev. Neurosci.* 30:259–88 [PubMed: 17600522]
- Schultz W 2019. Recent advances in understanding the role of phasic dopamine activity. *F1000Research* 8:1680
- Schultz W, Apicella P, Ljungberg T. 1993. Responses of monkey dopamine neurons to reward and conditioned stimuli during successive steps of learning a delayed response task. *J. Neurosci.* 13(3):900–913 [PubMed: 8441015]
- Schultz W, Dayan P, Montague P. 1997. A neural substrate of prediction and reward. *Science* 275(5306):1593–99 [PubMed: 9054347]
- Sesack SR, Hawrylak VA, Matus C, Guido MA, Levey AI. 1998. Dopamine axon varicosities in the prelimbic division of the rat prefrontal cortex exhibit sparse immunoreactivity for the dopamine transporter. *J. Neurosci.* 18(7):2697–708 [PubMed: 9502827]

- Sharpe MJ, Batchelor HM, Mueller LE, Chang CY, Maes EJP, et al. 2020. Dopamine transients do not act as model-free prediction errors during associative learning. *Nat. Commun.* 11(1):106 [PubMed: 31913274] This study provides evidence from three behavioral preparations that dopamine neurons participate in forming model-based representations.
- Sharpe MJ, Chang CY, Liu MA, Batchelor HM, Mueller LE, et al. 2017. Dopamine transients are sufficient and necessary for acquisition of model-based associations. *Nat. Neurosci.* 20(5):735–42 [PubMed: 28368385]
- Stalnaker TA, Howard JD, Takahashi YK, Gershman SJ, Kahnt T, Schoenbaum G. 2019. Dopamine neuron ensembles signal the content of sensory prediction errors. *eLife* 8:e49315 [PubMed: 31674910]
- Steidl S, Wang H, Ordonez M, Zhang S, Morales M. 2017. Optogenetic excitation in the ventral tegmental area of glutamatergic or cholinergic inputs from the laterodorsal tegmental area drives reward. *Eur. J. Neurosci.* 45(4):559–71 [PubMed: 27740714]
- Sulzer D, Cragg SJ, Rice ME. 2016. Striatal dopamine neurotransmission: regulation of release and uptake. *Basal Ganglia* 6(3):123–48 [PubMed: 27141430]
- Sun F, Zeng J, Jing M, Zhou J, Feng J, et al. 2018. A genetically encoded fluorescent sensor enables rapid and specific detection of dopamine in flies, fish, and mice. *Cell* 174(2):481–96.e19 [PubMed: 30007419]
- Sutton RS, Barto AG. 2018. *Reinforcement Learning: An Introduction*. Cambridge, MA: MIT Press. 2nd ed.
- Takahashi YK, Batchelor HM, Liu B, Khanna A, Morales M, Schoenbaum G. 2017. Dopamine neurons respond to errors in the prediction of sensory features of expected rewards. *Neuron* 95(6):1395–405.e3 [PubMed: 28910622]
- Takahashi YK, Langdon AJ, Niv Y, Schoenbaum G. 2016. Temporal specificity of reward prediction errors signaled by putative dopamine neurons in rat VTA depends on ventral striatum. *Neuron* 91(1):182–93 [PubMed: 27292535]
- Takahashi YK, Roesch MR, Wilson RC, Toreson K, O'Donnell P, et al. 2011. Expectancy-related changes in firing of dopamine neurons depend on orbitofrontal cortex. *Nat. Neurosci.* 14(12):1590–97 [PubMed: 22037501]
- Takahashi YK, Schoenbaum G. 2016. Ventral striatal lesions disrupt dopamine neuron signaling of differences in cue value caused by changes in reward timing but not number. *Behav. Neurosci.* 130(6):593–99 [PubMed: 27854448]
- Tervo DGR, Hwang B-Y, Viswanathan S, Gaj T, Lavzin M, et al. 2016. A designer AAV variant permits efficient retrograde access to projection neurons. *Neuron* 92(2):372–82 [PubMed: 27720486]
- Threlfell S, Lalic T, Platt NJ, Jennings KA, Deisseroth K, Cragg SJ. 2012. Striatal dopamine release is triggered by synchronized activity in cholinergic interneurons. *Neuron* 75(1):58–64 [PubMed: 22794260]
- Tian J, Huang R, Cohen JY, Osakada F, Kobak D, et al. 2016. Distributed and mixed information in monosynaptic inputs to dopamine neurons. *Neuron* 91(6):1374–89 [PubMed: 27618675]
- Tsutsui-Kimura I, Matsumoto H, Akiti K, Yamada MM, Uchida N, Watabe-Uchida M. 2020. Distinct temporal difference error signals in dopamine axons in three regions of the striatum in a decision-making task. *eLife* 9:e62390 [PubMed: 33345774]
- Ungless MA, Grace AA. 2012. Are you or aren't you? Challenges associated with physiologically identifying dopamine neurons. *Trends Neurosci.* 35:422–30 [PubMed: 22459161]
- Ungless MA, Magill PJ, Bolam JP. 2004. Uniform inhibition of dopamine neurons in the ventral tegmental area by aversive stimuli. *Science* 303(5666):2040–42 [PubMed: 15044807]
- Vancraeynest P, Arsenault JT, Li X, Zhu Q, Kobayashi K, et al. 2020. Selective mesoaccumbal pathway inactivation affects motivation but not reinforcement-based learning in macaques. *Neuron* 108(3):568–81.e6 [PubMed: 32758424]
- Verharen JPH, Zhu Y, Lammel S. 2020. Aversion hot spots in the dopamine system. *Curr. Opin. Neurobiol.* 64:46–52 [PubMed: 32146296]
- Voorn P, Vanderschuren LJMJ, Groenewegen HJ, Robbins TW, Pennartz CMA. 2004. Putting a spin on the dorsal-ventral divide of the striatum. *Trends Neurosci.* 27(8):468–74 [PubMed: 15271494]

- Waelti P, Dickinson A, Schultz W. 2001. Dopamine responses comply with basic assumptions of formal learning theory. *Nature* 412(6842):43–48 [PubMed: 11452299]
- Wassum KM, Ostlund SB, Maidment NT. 2012. Phasic mesolimbic dopamine signaling precedes and predicts performance of a self-initiated action sequence task. *Biol. Psychiatry* 71(10):846–54 [PubMed: 22305286]
- Watabe-Uchida M, Eshel N, Uchida N. 2017. Neural circuitry of reward prediction error. *Annu. Rev. Neurosci.* 40:373–94 [PubMed: 28441114]
- Watabe-Uchida M, Zhu L, Ogawa SK, Vamanrao A, Uchida N. 2012. Whole-brain mapping of direct inputs to midbrain dopamine neurons. *Neuron* 74(5):858–73 [PubMed: 22681690]
- Willuhn I, Burgeno LM, Everitt BJ, Phillips PEM. 2012. Hierarchical recruitment of phasic dopamine signaling in the striatum during the progression of cocaine use. *PNAS* 109(50):20703–8 [PubMed: 23184975]
- Willuhn I, Burgeno LM, Groblewski PA, Phillips PEM. 2014. Excessive cocaine use results from decreased phasic dopamine signaling in the striatum. *Nat. Neurosci.* 17(5):704–9 [PubMed: 24705184]
- Wise RA. 2004. Dopamine, learning and motivation. *Nat. Rev. Neurosci.* 5(6):483–94 [PubMed: 15152198]
- Wise RA, Robble MA. 2020. Dopamine and addiction. *Annu. Rev. Psychol.* 71:79–106 [PubMed: 31905114]
- Wolfart J, Roeper J. 2002. Selective coupling of T-type calcium channels to SK potassium channels prevents intrinsic bursting in dopaminergic midbrain neurons. *J. Neurosci.* 22(9):3404–13 [PubMed: 11978817]
- Wright CI, Beijer AV, Groenewegen HJ. 1996. Basal amygdaloid complex afferents to the rat nucleus accumbens are compartmentally organized. *J. Neurosci.* 16(5):1877–93 [PubMed: 8774456]
- Yang H, de Jong JW, Tak Y, Peck J, Bateup HS, Lammel S. 2018. Nucleus accumbens subnuclei regulate motivated behavior via direct inhibition and disinhibition of VTA dopamine subpopulations. *Neuron* 97(2):434–49.e4 [PubMed: 29307710]
- Young AMJ. 2004. Increased extracellular dopamine in nucleus accumbens in response to unconditioned and conditioned aversive stimuli: studies using 1 min microdialysis in rats. *J. Neurosci. Methods.* 138(1–2):57–63 [PubMed: 15325112]
- Yuan L, Dou Y-N, Sun Y-G. 2019. Topography of reward and aversion encoding in the mesolimbic dopaminergic system. *J. Neurosci.* 39(33):6472–81 [PubMed: 31217328]
- Zahm DS, Brog JS. 1992. On the significance of subterritories in the “accumbens” part of the rat ventral striatum. *Neuroscience* 50(4):751–67 [PubMed: 1448200]
- Zweifel LS, Parker JG, Lobb CJ, Rainwater A, Wall VZ, et al. 2009. Disruption of NMDAR-dependent burst firing by dopamine neurons provides selective assessment of phasic dopamine-dependent behavior. *PNAS* 106(18):7281–88 [PubMed: 19342487]

**SUMMARY POINTS**

1. The midbrain dopamine (DA) system is highly heterogeneous from the molecular composition and anatomical organization of DA neurons to the function of projection-defined DA systems in behavior.
2. Whether DA release and the firing activity of DA neurons are dissociable is a topic of current debate.
3. Aversion-excited DA neurons have distinct projections and physiological characteristics, are located in the medial ventral tegmental area, and appear to belong to a unique class.
4. Defining DA neurons with respect to their striatal target refines their role in behavior.

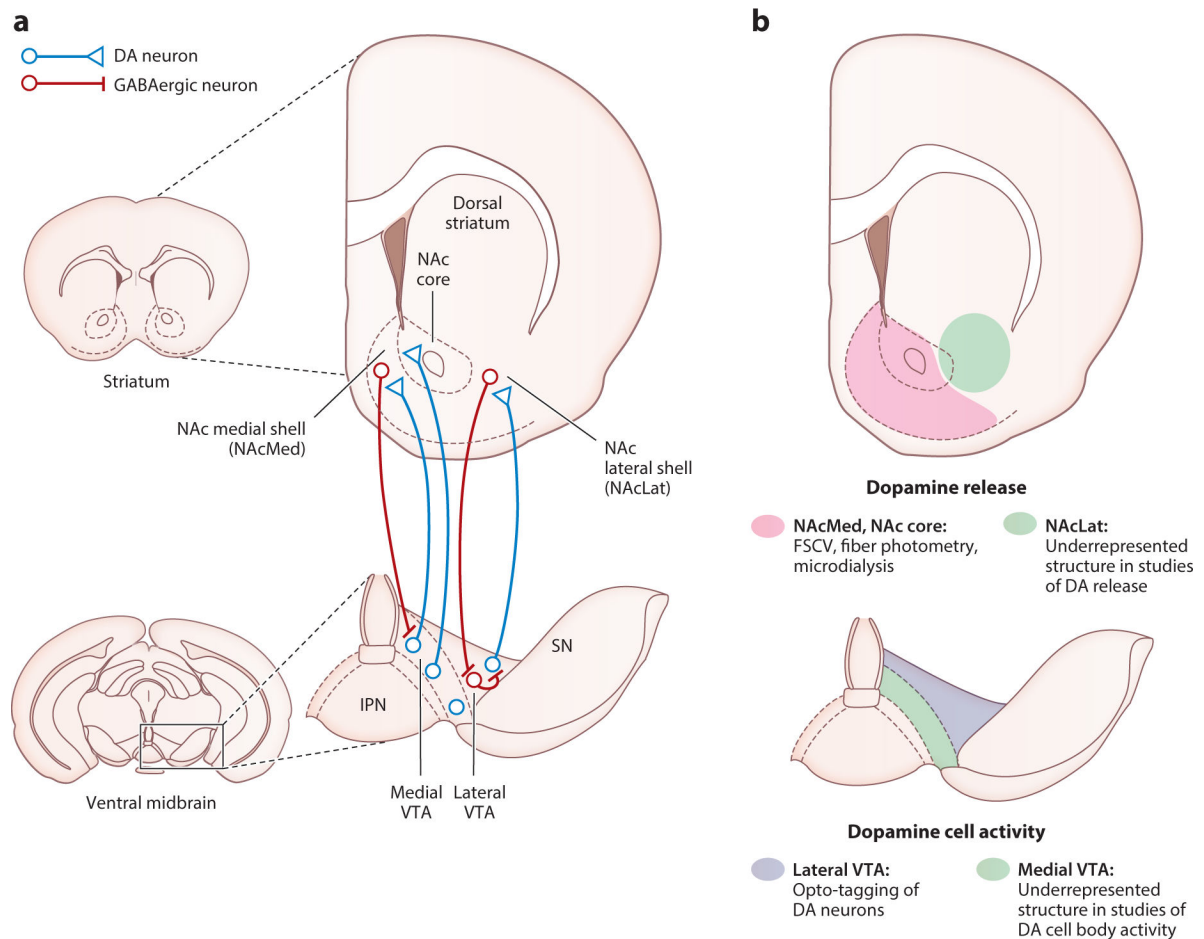
### FUTURE ISSUES

1. How can we integrate reward, motivation, and aversion encoding within different DA subsystems into a coherent framework of DA function?
2. What is the functional consequence of DA signaling in defined striatal territories, and how are these altered by experience or in disease?
3. Why is reward prediction error signaling so prominent both within the activity of DA neurons and in DA release in their target regions, and what is the significance of such a signal?
4. Should we consider DA neurons as performing individualized computations or as populations or subpopulations that collectively encode metrics critical for behavior?
5. Do DA neurons generate model-based computations, and if so, what inputs onto DA neurons support model-free versus model-based behaviors?
6. Are the mechanisms that govern local control of DA release conserved throughout the striatum?
7. Is ramping of DA release generated at the axon terminal, or does ramping arise from ramping activity of dopamine cell bodies?

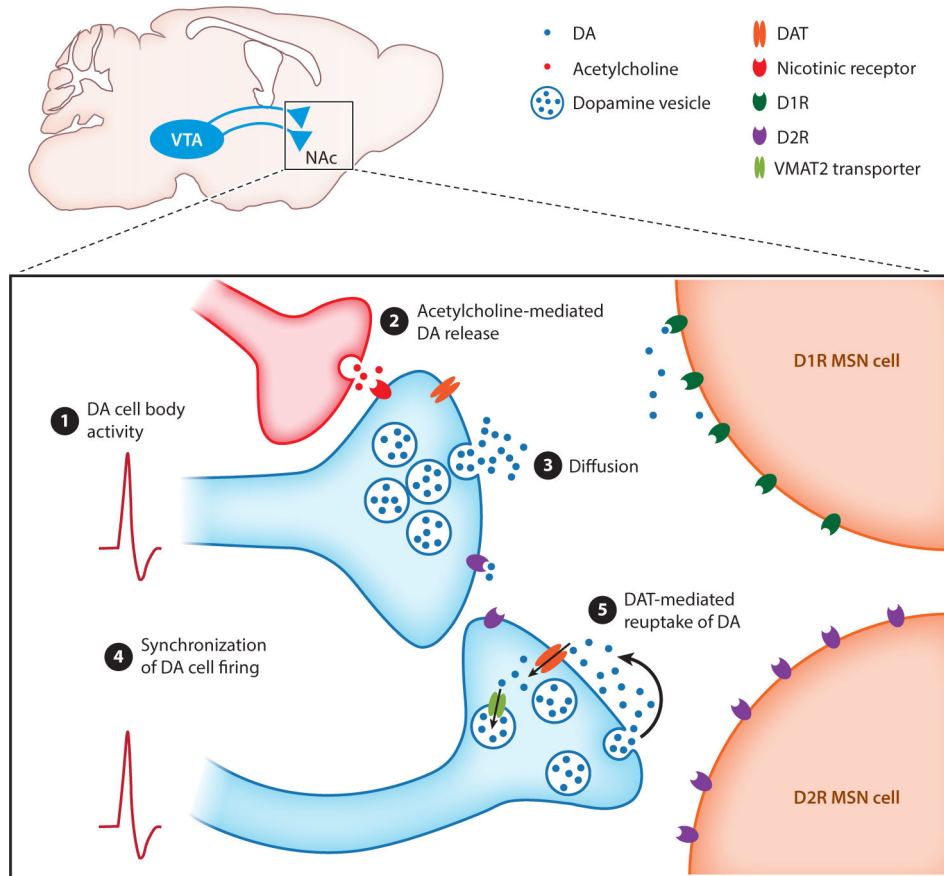
### NUCLEUS ACCUMBENS LATERAL SHELL

The terminology nucleus accumbens lateral shell (NAcLat) may be misleading since it may suggest that it is part of the nucleus accumbens (NAc) shell, though the most striking distinction appears to be that the NAcLat is the most prominent projection target from dopamine (DA) neurons in the lateral ventral tegmental area (VTA), whereas the NAc medial shell is innervated by DA neurons in the medial VTA (Breton et al. 2019, Farassat et al. 2019, Lammel et al. 2008). Further, NAc core-projecting DA neurons are predominantly located in the medial VTA, but a few studies in rats have also observed NAc core-projecting DA neurons in more lateral VTA regions (Breton et al. 2019, Saunders et al. 2018). Nevertheless, we think the terminology NAcLat is preferable as it refers to an anatomical region defined in *The Mouse Brain in Stereotaxic Coordinates* (Franklin & Paxinos 2001) and *The Rat Brain in Stereotaxic Coordinates* (Paxinos & Watson 2007). Whether this region is more appropriately labeled NAcLat, ventrolateral striatum, or lateral NAc remains a semantic question. Using nomenclature based on widely established anatomical atlases will facilitate the targeting of this brain region and in turn the reproducibility of results.

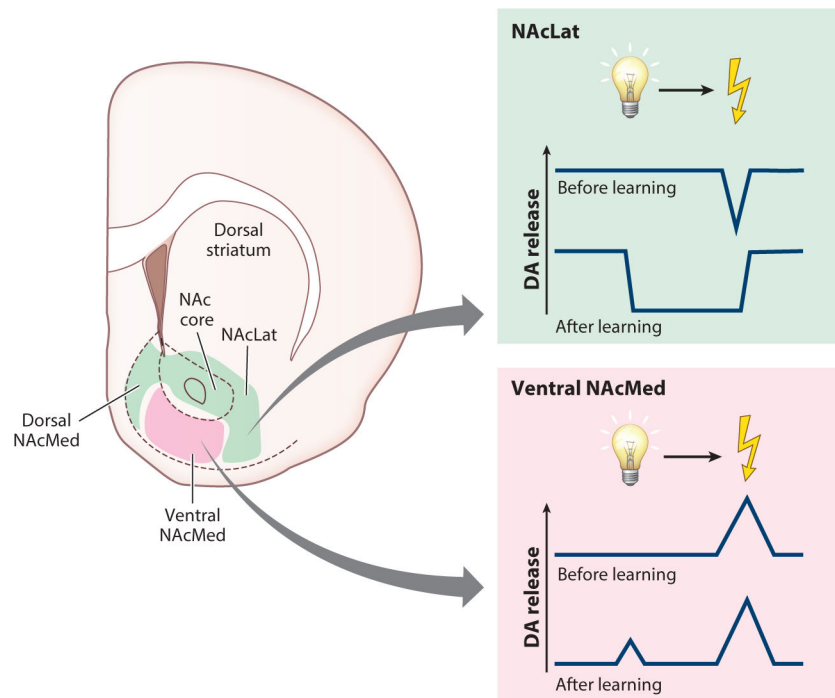


**Figure 1.**

The mesoaccumbal dopamine system. (a) Schematic showing anatomical organization of the mesoaccumbal DA system. DA neurons projecting to the NAc originate in the VTA. These neurons exhibit a topographical gradient of projections from medioventral and dorsolateral VTA to medial and lateral NAc, respectively. MSNs in the NAcMed directly inhibit medial VTA DA neurons projecting to the NAcMed, whereas MSNs in the NAcLat target GABAergic neurons in the lateral VTA that ultimately disinhibit NAcLat-projecting lateral VTA DA neurons. (b) Approaches for monitoring DA neuron activity and DA release are often biased toward specific VTA and NAc subregions, respectively. In the NAc (*top*), recordings of DA release are often biased toward the NAcMed and NAc core. In the VTA (*bottom*), recordings of DA neural activity are often biased toward the lateral VTA. Note that the preferred recording locations in the VTA and NAc are inconsistent with the topography of the mesoaccumbal DA system. Thus, we argue that studies of DA function should carefully consider the precise topographic organization of the mesoaccumbal DA system when investigating DA cell activity and release in the context of behavior. Abbreviations: DA, dopamine; FSCV, fast-scan cyclic voltammetry; IPN, interpeduncular nucleus; MSN, medium spiny neuron; NAc, nucleus accumbens; NAcLat, nucleus accumbens lateral shell; NAcMed, nucleus accumbens medial shell; SN, substantia nigra; VTA, ventral tegmental area.



**Figure 2.** Possible mechanisms for ramping of DA release in the striatum. Schematic of dopaminergic release sites (*blue*) in the striatum releasing DA in close proximity to DA D1R and D2R expressing MSN. Ramping of DA release may be generated through (①) action potential firing at the level of DA cell bodies, (②) modulation of DA release by cholinergic interneurons, (③) diffusion from numerous sparse release sites, (④) synchronization of VTA DA cell firing, and (⑤) saturation of DAT-mediated reuptake mechanisms. Abbreviations: D1R, D1 receptor; D2R, D2 receptor; DA, dopamine; DAT, dopamine transporter; MSN, medium spiny neuron; NAc, nucleus accumbens; VMAT2, vesicular monoamine transporter type 2; VTA, ventral tegmental area.



**Figure 3.** Aversion encoding in the mesoaccumbal DA system. Aversive stimuli (e.g., foot shocks) and their associated cues elicit decreases in DA release and DA terminal activity in the dorsal NAcMed, in the NAc core, and throughout the NAcLat. In contrast, these stimuli elicit increases in DA release and DA terminal activity within the ventral NAcMed. Abbreviations: DA, dopamine; NAc, nucleus accumbens; NAcLat, nucleus accumbens lateral shell; NAcMed, nucleus accumbens medial shell.