



# Antibody Therapies for Progressive Multiple Sclerosis and for Promoting Repair

Joachim Havla<sup>1,2,3</sup> · Reinhard Hohlfeld<sup>1,2,4</sup>

Accepted: 4 March 2022 / Published online: 14 March 2022  
© The Author(s) 2022

## Abstract

Progressive multiple sclerosis (PMS) is clinically distinct from relapsing–remitting MS (RRMS). In PMS, clinical disability progression occurs independently of relapse activity. Furthermore, there is increasing evidence that the pathological mechanisms of PMS and RRMS are different. Current therapeutic options for the treatment of PMS remain inadequate, although ocrelizumab, a B-cell-depleting antibody, is now available as the first approved therapeutic option for primary progressive MS. Recent advances in understanding the pathophysiology of PMS provide hope for new innovative therapeutic options: these include antibody therapies with anti-inflammatory, neuroprotective, and/or remyelination-fostering effects. In this review, we summarize the relevant trial data relating to antibody therapy and consider future antibody options for treating PMS.

**Keywords** Progressive multiple sclerosis · Antibody therapy · CD20 · PMS · Ocrelizumab · Rituximab

## Introduction

Multiple sclerosis (MS) is the most frequent chronic inflammatory demyelinating disease of the central nervous system (CNS). Progressive multiple sclerosis (PMS) is characterized by a relentless increase of disability that is not associated with clinical relapses as they occur in the relapsing remitting form of the disease (RRMS). There is increasing evidence that the pathological mechanisms of PMS and RRMS are different. Whereas relapses are thought to be caused by acute focal inflammation, relapse-independent progression is the clinical consequence of more diffuse inflammatory and neurodegenerative processes [1, 2]. This is supported by MRI evidence of a decrease in the number of new lesions and increasing

atrophy in patients with PMS (PwPMS), as well as by clinical evidence of an increase in disability without focal inflammation [3]. According to the modified Lublin criteria [3], the previously held distinction between primary progressive and secondary progressive MS is no longer necessary; both forms are included in the category of progressive MS (PMS). This is supported by evidence that primary and secondary progressive MS lack distinguishing histopathological features [4].

Whereas the inflammation dominating the early phase of RRMS is probably still driven by peripheral immune processes, it is thought that in PMS there is a “compartmentalization” of inflammation within the CNS compartment with a predominance of chronic-active, spreading and inactive lesions [1]. In addition, cortical and gray matter lesions, as well as meningeal lymphoid B-cell aggregates may act as drivers of disability progression [4]. Furthermore, increasing evidence points to microglial activation as a driver of progression. This seems to involve increased production of reactive oxygen species or nitric oxides [2]. This oxidative stress affects axonal mitochondria, leading to alterations of mitochondrial DNA and neuroaxonal energy deficiency [2]. Oxidative stress also affects the remyelination capacity of oligodendrocytes [5].

✉ Joachim Havla  
joachim.havla@med.lmu.de

<sup>1</sup> Institute of Clinical Neuroimmunology, University Hospital, LMU Munich, Munich, Germany

<sup>2</sup> Biomedical Center (BMC), Faculty of Medicine, LMU Munich, Martinsried, Germany

<sup>3</sup> Data Integration for Future Medicine (DIFUTURE) Consortium, LMU Munich, Munich, Germany

<sup>4</sup> Munich Cluster of Systems Neurology (SyNergy), Munich, Germany

## General Aspects of Antibody-Based Therapies in PMS

### Anti-inflammatory Versus Neuroprotective Therapeutic Strategy

In view of the current concepts of the pathogenesis of PMS (see the “Introduction” section), it makes sense to consider two different treatment approaches: one aiming to curb inflammation and a second aiming to foster neuroprotection, remyelination, and repair. Because there is evidence that inflammation and neurodegeneration occur concomitantly from onset, theoretically one would like to combine these strategies from the beginning. Currently there is a lack of effective, *sensu-stricto* neuroprotective therapies. In this regard, it is worthwhile to distinguish between directly neuroprotective therapies and indirectly neuroprotective effects of anti-inflammatory interventions. By reducing pathogenic inflammation, anti-inflammatory agents help to preserve myelin and axons, thereby indirectly exerting protection.

### Time Window of Opportunity

Our understanding of the pathogenesis of PMS would seem to support a time-window-adjusted treatment strategy [6]. Indeed, phase III studies have consistently demonstrated that the time window of opportunity for anti-inflammatory medications appears to be the early phase of PMS. In addition, it was shown that younger age, shorter duration of PMS, and more pronounced clinical and MRI activity at baseline were more likely to be associated with a positive outcome [7, 8]. Therefore, anti-inflammatory therapies should be used at a disease stage dominated by an inflammatory pathomechanism. Consistent with these considerations, the currently available therapies seem to be effective mainly in the active phases of PMS, defined by superimposed relapses and focal MRI activity [7, 8]. Ideally and theoretically, anti-inflammatory therapies should act not only in the periphery but also directly in the central nervous system (CNS).

In contrast, one might predict that neuroprotective, reparative, and remyelinating therapies, should they become available in the future, may possess a larger time window of opportunity. One important potential therapeutic goal would be the activation and differentiation of oligodendrocyte progenitor cells (OPCs), increasing their potential to differentiate into myelinating oligodendrocytes [9]. Additional therapeutic aims include neuroprotection, e.g., by inhibiting apoptosis and oxidative stress, and protection of mitochondria [6].

### Risk–Benefit Ratio

The use of monoclonal antibodies in the treatment of MS has made it possible to target selected molecules thought

to play a key role in the pathogenesis. In RRMS antibody, therapy is associated with a high level of efficacy regarding disease activity. Several antibodies such as natalizumab, ocrelizumab, ofatumumab, and alemtuzumab are considered highly effective options for treating RRMS/RMS allowing for effective control of disease activity [10]. Ocrelizumab and to some extent also rituximab were shown to have a positive impact also on disease progression. Here, it seems that the subgroup of PMS patients with ongoing inflammatory activity benefit more than patients without disease activity [11]. Apart from efficacy, the risks of treatment with monoclonal antibodies deserve special attention [12, 13]. Especially during long-term therapy, the whole immune system will be affected by the specific intervention, regardless of the fact that monoclonal antibodies are focused on a specific molecular target. The long-term consequences of therapeutic modulation of the immune system are principally unpredictable, as illustrated by daclizumab, a monoclonal antibody directed against the interleukin-2-receptor. This antibody was withdrawn from the market and lost approval when serious adverse events became apparent [14–16].

## Anti-inflammatory Approaches for Treating PMS

### T Cells Versus B cells

From the early days of immunomodulatory therapy of MS with interferon-beta, it has been known that PMS is a more difficult therapeutic target than RRMS. Yet, even in those early days, there was evidence that PMS is not entirely resistant to immunomodulatory therapy if treatment is initiated sufficiently early [17, 18]. One of the most potent options for the treatment of active RRMS is natalizumab. Natalizumab acts by inhibiting alpha-4-integrin-mediated migration of lymphocytes, notably T-lymphocytes, from blood into the CNS. The therapeutic efficacy of alpha-4-integrin inhibition was first shown in a T-cell-mediated animal model, experimental autoimmune encephalomyelitis (EAE) [19]. It was only logical to test natalizumab also in patients with PMS. The ASCEND trial program investigated whether natalizumab was able to slow relapse-independent progression in patients with secondary progressive MS. In this placebo-controlled, randomized, double-blind study, 888 patients with SPMS participated. Of these, 439 were in the natalizumab cohort and 449 in the placebo group. The observation period of 2 years was followed by an open-label extension (OLE). Disappointingly, the ASCEND study was terminated after publication of blinded study results. Natalizumab did not reduce progression on the the primary multicomponent

disability endpoint but only on its upper-limb component [20]. Specifically, there was no effect on walking deterioration as measured by the EDSS as well as the timed 25-foot walk (T25FW) as part of the multicomponent primary endpoint. However, for the third component of the primary endpoints, the 9-Hole Peg Test (9HPT), there was a significantly lower relative risk (minus 44%) of the natalizumab group developing upper extremity disability (15% vs. 23%;  $p=0.001$ ). This effect was independent of the detection of active inflammatory lesions on magnetic resonance imaging. Clinical relevance of this finding is possible, particularly in regard to upper extremity function preservation and independence [20]. Interestingly, these results are consistent with the IMPACT trial (interferon beta 1a in secondary progressive MS) [21]. Again, efficacy was seen only on the upper extremities with less relapse-independent progression but lack of efficacy on gait disturbance with shortening of walking distance [21]. Overall, a potential therapeutic effect of natalizumab purely on disability progression and independent of disease activity remains unclear. Based on the partly positive aspects, it can be assumed that the treatment duration may have been too short and that future studies should plan for a longer treatment duration [20] (Table 1).

In addition to T cells, B cells have increasingly come into focus as important elements in the immunopathogenesis of MS, not least due to therapeutic developments. Thus, B cells were established as bidirectional interaction partners of T cells, both in the periphery and in the CNS [22–24]. This concept is supported by the high efficacy of B-cell depletion with anti-CD20 monoclonal antibodies [23–32]. CD20 is a surface molecule expressed by B cells at different stages of maturation. CD20 positivity is observed across different stages ranging from pre-B cells in bone marrow to short-lived plasmablasts. However, long-lived antibody-producing plasma cells are CD20 negative [32]. Notably, CD20 is not exclusive to B cells but is also present on a subset of T cells that could play a role in the pathogenesis of MS [33]. Several anti-CD20 therapeutic monoclonal antibodies were developed to achieve B-cell depletion. The antibodies differ in their structural features (chimeric, humanized, fully human antibodies), relative potency to elicit antibody-dependent cellular and complement-mediated cytotoxicity, and pharmacokinetics [34]. They also differ in the route of administration (intravenously or subcutaneously), infusion times, and the need for premedication [34].

What exactly is the role of B cells and antibodies in MS pathogenesis? Obviously B cells are the source of antibodies, including CSF-specific oligoclonal bands and immunoglobulins deposited in MS lesions [35]. It is not clear, however, if intrathecally produced antibodies which include antibodies directed against cellular debris are pathogenic

[36]. It is therefore likely that the therapeutic efficacy of CD20-mediated B cell depletion is brought about by inhibition of B-cell functions other than antibody production, including antigen presentation to T cells, and production of various cytokines and chemokines [37]. The importance of these intercellular interactions appears to be particularly high in active early stages of MS. In later or progressive stages, plasma cell infiltrates, which are insensitive to anti-CD20-mediated depletion, might play an increasingly important role [38]. Nevertheless, clinical efficacy of CD20 antibody therapy was shown in both relapsing and progressive phases of MS. It should be noted that anti-CD20 antibodies differ in their molecular and pharmacological properties. For example, rituximab acts mostly by complement-dependent B-cell depletion, whereas ocrelizumab acts mostly via antibody-dependent cellular cytotoxicity [39]. These distinctive properties not only impact the mechanisms and efficacy of monoclonal CD20 antibodies, but also the required dosing [40]. Although current MS dosing regimens result in near complete depletion of circulating B cells, dose-dependent differential kinetics of B cell reconstitution are evident. One possible interpretation could therefore be that near-complete peripheral B-cell depletion may be accompanied by varying degrees of depletion in tissues, immune cell niches, or secluded compartments such as the CNS [41].

However, why and to what extent the described differential mechanisms of monoclonal CD20 antibodies, which are administered outside the CNS, may contribute to their beneficial effects on MS progression remains unresolved. It is also not clear whether the efficacy of anti-CD20-mediated depletion in the CNS correlates with the therapeutic effect on disease progression. In progressive MS, there is an increasing “compartmentalization” of inflammation within the CNS, with accumulation of clonally expanded B cells in meningeal B-cell follicle-like structures. They provide a niche in the CNS which supports and maintains pathogenic B-cell function [42, 43] and could be instrumental in driving chronic progression [42]. B-cell-associated pathogenetic features include cortical lesions as well as diffuse microglial activation [43].

Depletion of peripheral B cells is accompanied by a marked decrease in B cells not only in the cerebrospinal fluid (CSF) but also in the perivascular spaces of the brain [44]. Relevant crossing of CD20 antibodies across the intact blood–brain barrier is unlikely [45]. Therefore, it seems that monoclonal CD20 antibodies can only be effective if systemic depletion of peripheral B cells occurs during a phase of the disease course when peripheral immune cells are actively recruited to the CNS [46]. Thus, an obvious therapeutic approach could be the intrathecal administration of monoclonal anti-CD20 antibodies. The efficacy of such therapeutic strategies is currently investigated with ongoing animal research and clinical studies [46].

**Table 1** Monoclonal antibodies for progressive multiple sclerosis (table according to [6, 101, 103])

Monoclonal antibody	Study	Details	Primary endpoint	Key results	Reference
<b>Phase III</b>					
Natalizumab	Phase III SPMS (ASCEND)	MC, R, DB, PC, SD 2 years, 887 patients, MA(y) 47, MDoP(y) 5, eEDSS 3.0–6.5	Time to 3 month composite CDP	Negative (no treatment effect on multicomponent outcome, EDSS or the T25FW; but reduced 9HPT progression)	[20]
Rituximab	Phase II/III PPMS (OLYMPUS)	MC, R, DB, PC, 2 years, 439 patients, MA(y) 49, MDoP(y) 9, eEDSS 2.0–6.5	Time to CDP	Negative (no significant benefit in CDP for RIX vs. placebo)	[104]
Ocrelizumab	Phase III PPMS (ORATORIO)	MC, R, DB, PC, minimum 120 weeks, event driven study, 732 patients, MA(y) 45, MDoP(y) 6, eEDSS 3.0–6.5	Time to 12-week confirmed CDP	Positive (the relative risk of 12-week confirmed disability progression was significantly decreased by 24% ( $p=0.03$ ))	[29, 53]
	Phase III PPMS (O Hand)	MC, R, DB, PC, 27 months, aim 1000 patients, eEDSS $\geq 3.0-8.0$	Time to upper limb disability progression confirmed For at least 12 weeks	Recruiting	Analysis ongoing
	Phase III PPMS/SPMS (Consonance)	MC, open-label, single-arm, aim 900 patients, 4 years, eEDSS $\leq 6.5$	Proportion of patients with no evidence of progression (NEP) on 6 month EDSS CDP	Recruiting	Analysis ongoing
<b>Phase II</b>					
Opicinumab	Phase II ON (RENEW)	MC, R, DB, PC, 32 weeks	Remyelination at 24 weeks, measured as recovery using full-field visual evoked potential (FF-VEP)	Negative (remyelination did not differ significantly between the opicinumab and placebo groups in the ITT population at week 24)	[105]
Temelimab	Phase II RMS (ProTEct-MS)	MC, R, DB, PC, 1 year, eEDSS 2.5–5.5	Mean overall response score (ORS): EDSS, T25FW, 9HPT-DH, 9HPT-NDH	Ongoing	Pre-study [106]
Elezanumab	Phase II PPMS/SPMS	MC, R, DB, PC, 123 patients, 1 year	Composite score of EDSS, T25FW, 9HPT	Completed	Analysis ongoing

SC single center, MC multicenter, R randomized, DB double blinded, eEDSS entry EDSS, PC placebo controlled, SD study duration, MA(y) mean age years, MDoP(y) mean duration of progression years

## Rituximab

Rituximab is a chimeric monoclonal CD20 antibody that is used off-label in many places for the treatment of MS. This antibody has not been approved by the FDA (US Food and Drug Administration) or European Medicines Agency (EMA) for the treatment of MS [47–51]. However, the efficacy of rituximab has been studied in several trials in MS. In the 96-week phase II/III OLYMPUS study, a randomized controlled trial in primary progressive multiple sclerosis (PPMS), rituximab did not significantly improve confirmed disability progression (CDP,  $p=0.14$ ) or reduce brain atrophy rate ( $p=0.62$ ) (Table 1). However, there was significantly less T2 hyperintense lesion volume increase at week 96 ( $p<0.001$ ) compared with placebo [28].

Subgroup analyses showed that rituximab could delay time to CDP in younger PPMS patients (age < 51 years) or in patients with Gd-enhancing lesions at baseline. Thus, a positive prediction regarding future treatment response could be derived from subgroup analyses [52] and from very consistent ORATORIO experiences. The ORATORIO study evaluated the safety and efficacy of ocrelizumab in PPMS. ORATORIO is an international, multicenter, double-blind, randomized, placebo-controlled phase III study [29, 53].

To achieve depletion of CNS-resident B cells, intrathecal administration of rituximab has been explored as a mode of application. In a trial of intrathecal rituximab in PMS patients with MRI evidence of leptomeningeal contrast enhancement, there was a profound reduction of peripheral B cells and transient reduction of B cells in the CSF. However, the number of contrast-enhancing leptomeningeal sites did not change following treatment [54].

## Ocrelizumab

The humanized anti-CD20 monoclonal antibody ocrelizumab was approved at a dose of 600 mg i.v. twice yearly for the treatment of PPMS with evidence of disease activity in March 2017 (FDA) and January 2018 (EMA). The antibody targets CD20-expressing lymphocytes, mostly B cells but also a smaller subset of T cells [32, 33].

The EMA has licensed ocrelizumab based on the pivotal data for the treatment of adult patients with early PPMS, characterized by disease duration and degree of disability, as well as imaging features typical of inflammatory activity [55, 56]. The approval was based on the ORATORIO trial and on the study population investigated (Table 1). ORATORIO enrolled 732 PPMS patients and treated them with either ocrelizumab or placebo every 6 months for at least 120 weeks [26]. The study population represented early-stage PPMS patients, i.e., 18 to 55 years of age (inclusive), with an EDSS of 3.0 to 6.5 at the time of screening, and a duration of disease since the onset of first MS symptoms

of less than 10 years (for patients with an EDSS of  $\leq 5.0$  at screening) or less than 15 years (for patients with an EDSS of  $> 5.0$  at screening). The implication for clinical practice is that evidence of inflammatory activity, defined by Gd-uptaking T1 lesions and/or active [new or enlarging] T2 lesions), should be obtained by MRI in all patients who are considered candidates for treatment with ocrelizumab. Patients older than 55 years were not studied in the clinical trials [57]. In the pivotal study, ocrelizumab met both the primary endpoint (reduction in the risk of disability progression confirmed at 12 weeks) and the secondary endpoints.

The proportion of patients with CDP in the EDSS score at 12 weeks was reduced by 24% compared with placebo (significant reduction in 12-week CDP ( $p=0.03$ ) and its confirmation in 24-week CDP ( $p=0.04$ )) [26]. CDP at 24 weeks was defined as follows:

(1) Increase in EDSS score, (2)  $\geq 20\%$  increase in time to complete the 9-Hole Peg Test [9HPT], (3)  $\geq 20\%$  increase in time to complete the timed 25-foot walk [T25FW], and composite progression, defined as the first confirmed occurrence of any of these three (1–3) individual measures. In addition, time to need for a wheelchair (EDSS  $\geq 7$ ) was considered [29].

Subanalyses of hand function (9HPT) and walking ability (T25FW) confirmed the superiority of ocrelizumab with significantly less worsening in these motor function subscores [26]. Ocrelizumab significantly reduced the risk of disability progression, less T2 lesion volume for ocrelizumab-treated patients (minus 92% vs. placebo;  $p<0.001$ ), and brain atrophy compared with placebo ( $p=0.02$ ). However, this efficacy was driven by the subgroup of younger study participants (< 40 years) with disease activity, e.g., disease activity on MRI ( $\geq 1$  Gd-enhancing lesions) [26, 58]. As mentioned before this is consistent with experiences from the OLYMPUS trial (rituximab in PPMS) [28].

After the double-blind phase of the pivotal study, study participants could enter the optional open-label extension phase (OLE). Patients previously treated with ocrelizumab remained on treatment (initial ocrelizumab cohort), and patients from the placebo group were switched to the ocrelizumab treatment arm (initial placebo cohort).

Five hundred twenty-seven patients in the ORATORIO study program (97%) entered the OLE phase, and of these patients, 86% were analyzed. At 6.5 years, the proportion of patients with disability progression was lower in the initial OCR cohort vs. initial placebo cohort. EDSS progression was seen in 13.1% fewer patients in the initial OCR cohort ( $p=0.0018$ ), 12.5% fewer had relevant worsening in the 9HPT ( $p=0.0035$ ), and 7.5% fewer had relevant worsening in the T25FW ( $p=0.058$ ). Compound progression was seen by 10.1% fewer in the initial OCR cohort ( $p=0.0023$ ). Paraclinically, T2 lesion volume (0.45% vs. 13%,  $p<0.0001$ ) and T1 hypointense lesion volume (36.68%

vs. 60.93%,  $p < 0.0001$ ) were significantly reduced. Over the entire period, the serious adverse event rate was 12.6 per 100 patient-years; the most common serious adverse event was infection at 4 per 100 patient-years. No new safety signals occurred over 6.5 years compared with the double-blind phase of ORATORIO [29].

## Future Prospects of B-Cell Targeting in PMS

### Inebilizumab

Direct depletion of CD19-positive cells may represent another B-cell targeting therapeutic strategy. CD19 is a member of the Ig superfamily and is involved in signal transduction following B cell receptor activation, in the regulation of B cell activation and humoral immunity [59]. As a therapeutic approach, CD19 is of interest because it is expressed on a greater range of B-cell lineage members, including pro-B cells and plasmablasts [59], than is CD20.

Inebilizumab is a glycosylated, afucosylated anti-CD19 antibody. The results of a 24-week phase I randomized controlled trial in patients with RRMS compared with placebo are available [60]. An investigation in PMS is currently pending.

## Neuroprotective and Remyelination-Fostering Strategies

### Therapeutic Targeting of Microglia and Oligodendrocytes

Apart from oligodendrocytes, microglia has emerged as a prime potential target for treating PMS. Histopathologically, PMS is associated with inactive, but also with chronically active, “smoldering” lesions, which are surrounded by a rim of microglial activation [61]. Moreover, diffuse microglial activation can occur independent of focal lesions [62, 63]. Positron emission tomography (PET) could represent one possibility of monitoring microglial activation in the normal appearing white matter (NAWM). Here, the upregulation of the mitochondrial translocator protein TSPO provides a marker that can be visualized with modern tracers [64–66].

The exact pathophysiological function of microglia in progression remains speculative. Expression of proinflammatory cytokines [67] and also the contribution of microglia to mitochondrial damage may be relevant [68]. It should be noted, however, that microglia may also contribute to repair and remyelination via mechanisms of phagocytosis and production of anti-inflammatory cytokines [69, 70] promoting the recruitment of oligodendrocyte precursor cells (OPC) to the lesion site [71, 72].

In view of these complexities, a differentiated treatment strategy is required for promoting neuroprotection and remyelination. The overall goal is to promote remyelination capacity, as well as to reduce proinflammatory microglial activity [73]. Presently, however, no singular specific target is known that fulfills both requirements. Another consideration, which is relevant for antibody-based therapies, is that neuroprotective and remyelination-fostering therapies would have to reach the CNS in sufficiently high concentrations, which may be more easily achieved with small molecule drugs.

### Opicinumab

The transmembrane cell surface glycoprotein LINGO-1 has a significant role in controlling oligodendrocyte precursor proteins (OPCs) and neurons [74–77]. LINGO-1 is known to be upregulated in MS lesions. It has been shown in vitro and in animal studies that blockade of LINGO-1 can lead to increased axonal myelination as well as improvement in clinical scores [78, 79]. This was the basis for investigating the monoclonal antibody opicinumab, a fully humanized anti-LINGO1-antibody, in several clinical trials [80–85].

In the RENEW trial, a randomized, placebo-controlled, multicenter phase II study (33 treated, 36 placebo) in patients with a first unilateral acute optic neuritis episode, no benefits regarding remyelination were shown in an intention-to-treat analysis [86] (Table 1). Visual outcomes, specifically visual evoked potentials (VEP), optical coherence tomography (OCT), and MRI, were examined [83]. However, a post hoc analysis indicated that older patients in particular may benefit from therapy with opicinumab [82].

The SYNERGY trial also failed to show a significant beneficial treatment advantage of the opicinumab group over the comparison cohort. This was a randomized, double-blind, placebo-controlled, dose-ranging phase II study ( $N = 419$ ) to investigate the clinical efficacy, safety, and pharmacokinetics of opicinumab in which patients were treated with intramuscular interferon beta-1a in combination with either placebo or a variable dose of opicinumab (3/10/30/100 mg/kg) [81]. The primary endpoint was the percentage of participants with confirmed clinical improvement over 72 weeks of treatment. The study was negative overall, although trends in some subscores were apparent [81]. There was no dose-linear improvement in disability for the opicinumab treatment arm. However, compared to the RENEW trial, RRMS patients with younger age, shorter disease duration, and less brain atrophy showed some benefit. Overall, the remyelination potential of opicinumab and the reasons for the negative results of the study remain unclear at this time. Based on the study design, the study could have been underpowered, the selection of patients might not have been optimal, and poor penetration of antibodies through the blood–brain

barrier could have contributed to the results. Furthermore, the expression of LINGO-1 may be variable, depending on disease activity [87].

## Outlook (I): Tackle the Immune Cell Niche

In addition to the therapeutic development of additional and well-established antibodies, with so-called small molecules, a new option of treating PwPMS is emerging. One possible therapeutic approach is the inhibition of Bruton's tyrosine kinase (BTK) [88]. BTK regulates the activation, proliferation, and differentiation of B cells into plasma cells [89, 90]. BTK is a cytoplasmic kinase expressed on some cells of the hematopoietic cell lineage including B cells and myeloid cells. BTK is not expressed on T or NK cells [91]. As "small molecules," BTK have the advantage that they can cross the blood–brain barrier more easily than antibodies, thereby allowing for more effective targeting of the CNS-resident (compartmentalized) B-cell population [89, 90].

Different BTK inhibitors are currently being explored in clinical trials (see [89] for a comprehensive review). Ideally, a BTK inhibitor not only should easily cross the BBB, but it also should bind BTK in a highly selective but reversible manner. One example is evobrutinib. Evobrutinib was able to meet the primary endpoint (number of T1 gadolinium-enhancing lesions) in a 24-week phase II clinical trial comparing oral evobrutinib at various doses with placebo or dimethyl fumarate in patients with RRMS or active SPMS, but showed no efficacy on disability progression [92]. As presented by Gheen et al. [93], fenebrutinib is currently being studied in a phase III trial in PPMS (examination of fenebrutinib, a highly selective BTKi, on disease progression of multiple sclerosis). Another BTK inhibitor, tolebrutinib, was studied at different doses in a 16-week phase IIb study in RRMS. Also here, efficacy on disease activity was noted. However, based on this study, no conclusions can be drawn regarding potential efficacy in PMS. A study in PPMS (PERSEUS) and non-active SPMS (HERCULES) is currently recruiting (ClinicalTrials.gov Identifier: NCT04458051; ClinicalTrials.gov Identifier: NCT04411641). Whether BTK inhibitors have neuroprotective in addition to their anti-inflammatory properties remains to be elucidated [94].

## Outlook (II): New Approaches Regarding Study Designs, Strategies, and Objectives

It remains to be evaluated whether the traditional outcome measures such as disability progression measured with the EDSS remain the most suitable tool for studies in PMS. It is expected that future studies will need to focus on more

specific biological or preclinical outcome measures [95]. These could include novel cell-specific markers of inflammatory and degenerative mechanisms as well as a broader selection of already existing preclinical markers. Besides electrophysiological investigations such as somatosensory (SEP), motor (MEP) and visual evoked potentials (VEP), modern magnetic resonance imaging (MRI) parameters, and specific positron emission tomography (PET), examinations could play a role in future study designs [95]. Last but not least, optical coherence tomography (OCT) could be potentially suitable as a marker of retinal neuroaxonal degeneration [96]. In this regard, the International Progressive MS Alliance (<https://www.progressivemsalliance.org/>) has recently published a proposal of a core data set that could be used as a standard for future trial designs in PwPMS [95]. This includes the established clinical scores (EDSS, upper limb dexterity), preclinical measures (Neurofilament light [NfL] protein, brain atrophy, VEP and/or OCT), immune treatment response markers (sCD21, SCD27, sCD14, CXCL13, BAFF), and non-hypothesis driven measures like peripheral transcriptomics [95].

Other important aspects include the selection of the appropriate study population. For example, PwPMS should be selected taking into account disease dynamics during the disease course, but also the sex distribution within the study population, which should correspond to the sex distribution in the targeted MS population [95]. The study design is also critical for the success of a therapeutic development. It is questionable whether the complex interaction between inflammatory and neurodegenerative processes can be adequately addressed with a monotherapy. Of course, cumulative risks of polytherapy, tolerability, and costs to healthcare systems must be kept in mind for any development. However, combination therapy including immunosuppressants, but also remyelination or neuroprotective treatment strategies should be considered in the future. Regarding the design of PMS treatment trials, modern study designs make use of adaptive, enrichment, futility, or crossover design elements [95].

Even with improved study designs, the heterogeneity of MS populations continues to contribute an unavoidable element of unpredictability. Viewed in terms of pathophysiology, therapeutic targets should include proinflammatory microglia, active astrocytes, mitochondrial dysfunction, and oxidative stress. At the same time, the ability to remyelinate should be supported [97]. Related to the remyelination capacity of PwMS, it is known that there are two groups of patients, those considered good and those classified as poor/insufficient remyelimators [98]. Influencing factors could be age, duration of disease, presence of oligodendrocyte progenitor cells (OPCs), genetic factors, and environmental factors [99]. Interestingly, gender also appears to play a role: although more women than men are

affected by MS overall, the disease course in men appears overall more aggressive [100].

In conclusion, it seems promising to investigate strategies to promote myelin repair from endogenous as well as exogenous cell sources. In addition, the different factors influencing remyelination should be therapeutically addressed to support the spontaneous repair of demyelinated axons [97].

## Conclusions

The currently existing therapeutic options for the treatment of PMS remain inadequate. However, recent advances in understanding the pathophysiology of PMS offer hope for new innovative therapeutic options. Effective therapy must address the compartmentalized inflammation in the CNS. Furthermore, the classical requirement for immunotherapy in RRMS, namely early initiation of an effective therapy, is equally relevant for treating PMS. However, the data available so far on neuroprotection and remyelination in PMS do not yet allow definition of unequivocal target priorities. What seems clear, however, is that the principle of early therapy applies not only to RRMS but also to PMS. Neuroprotection is most effective when there are still myelinated axons to protect, and remyelination is most effective, when there are still demyelinated axons to remyelinate [101].

In particular, PMS is considered a very demanding challenge in the care of MS patients [101, 102]. Future therapy strategies should therefore always be multimodal. Besides possible antibody therapy options for anti-inflammation, neuroprotection, or remyelination, symptomatic therapy should also play a crucial role. Measuring the success of therapy by practical criteria is of key importance. One example is NEPAD, an acronym standing for “no evidence of progression or active disease,” i.e., “Lublin not active and without progression” [29, 57]. Last but not least, special attention must always be paid to benefit/risk analysis. Safety of treatment and strategies for dealing with adverse effects are at least as important as efficacy.

**Supplementary Information** The online version contains supplementary material available at <https://doi.org/10.1007/s13311-022-01214-x>.

**Required Author Forms** Disclosure forms provided by the authors are available with the online version of this article.

**Disclosures** RH has received grant support and honoraria from Novartis, DSanofi/Genzyme, Biogen, Teva, Merck, Janssen/Johnson-Johnson, and Roche. JH reports personal fees, research grants and non-financial support from Merck, Novartis, Roche, Biogen, Alexion, Celgene, and Janssen and non-financial support of the Guthy-Jackson Charitable Foundation, all outside the submitted work.

**Funding** Open Access funding enabled and organized by Projekt DEAL. The authors' scientific work has been supported by the German

Research Council, Bavarian and national association of the German MS society (DMSG), Dr. Leopold And Carmen Ellinger Foundation, and the association “Verein zur Therapieforschung für MS Kranke e.V.” J.H. is (partially) funded by the German Federal Ministry of Education and Research [Grant Numbers 01ZZ1603[A-D] and 01ZZ1804[A-H] (DIFUTURE)].

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

## References

- Lassmann H, van Horsen J, Mahad D. Progressive multiple sclerosis: pathology and pathogenesis. *Nat Rev Neurol*. 2012;8:647–56.
- Mahad DH, Trapp BD, Lassmann H. Pathological mechanisms in progressive multiple sclerosis. *Lancet Neurol*. 2015;14.
- Lublin FD, Reingold SC, Cohen JA, Defining the clinical course of multiple sclerosis: the, et al. revisions. *Neurology*. 2013;2014:83.
- Correale J, Gaitan MI, Ysraelit MC, Fiol MP. Progressive multiple sclerosis: from pathogenic mechanisms to treatment. *Brain*. 2017;140:527–46.
- Bramow S, Frischer JM, Lassmann H, et al. Demyelination versus remyelination in progressive multiple sclerosis. *Brain*. 2010;133:2983–98.
- Sorensen PS, Fox RJ, Comi G. The window of opportunity for treatment of progressive multiple sclerosis. *Curr Opin Neurol*. 2020;33:262–70.
- Comi G. Disease-modifying treatments for progressive multiple sclerosis. *Mult Scler*. 2013;19:1428–36.
- Sellebjerg F, Bornsen L, Ammitzboll C, et al. Defining active progressive multiple sclerosis. *Mult Scler*. 2017;23:1727–35.
- Kremer D, Göttle P, Hartung HP, Küry P. Pushing forward: remyelination as the new frontier in CNS diseases. *Trends Neurosci*. 2016;39.
- Costello F, Stuve O, Weber MS, Zamvil SS, Frohman E. Combination therapies for multiple sclerosis: scientific rationale, clinical trials, and clinical practice. *Curr Opin Neurol*. 2007;20:281–5.
- Gelfand JM, Cree BAC, Hauser SL. Ocrelizumab and other CD20(+) B-cell-depleting therapies in multiple sclerosis. *Neurotherapeutics*. 2017;14.
- Klotz L, Havla J, Schwab N, et al. Risks and risk management in modern multiple sclerosis immunotherapeutic treatment. *Ther Adv Neurol Disord*. 2019;12:1756286419836571.
- Havla J, Warnke C, Derfuss T, et al. Interdisciplinary risk management in the treatment of multiple sclerosis. *Dtsch Arztebl Int*. 2016;113:879–86.
- Williams T, Chataway J. Immune-mediated encephalitis with daclizumab: the final nail. *Mult Scler*. 2018.

15. Stork L, Bruck W, von Gottberg P, et al. Severe meningo-/encephalitis after daclizumab therapy for multiple sclerosis. *Mult Scler*. 2019;25:1618–32.
16. Rauer S, Stork L, Urbach H, et al. Drug reaction with eosinophilia and systemic symptoms after daclizumab therapy. *Neurology*. 2018;91:e359–63.
17. Koch MW, Mostert J, Uitdehaag B, Cutter G. Clinical outcome measures in SPMS trials: an analysis of the IMPACT and ASCEND original trial data sets. *Mult Scler*. 2020;26:1540–9.
18. Kappos L, Weinshenker B, Pozzilli C, et al. Interferon beta-1b in secondary progressive MS: a combined analysis of the two trials. *Neurology*. 2004;63:1779–87.
19. Yednock TA, Cannon C, Fritz LC, et al. Prevention of experimental autoimmune encephalomyelitis by antibodies against alpha 4 beta 1 integrin. *Nature*. 1992;356:63–6.
20. Kapoor R, Ho PR, Campbell N, et al. Effect of natalizumab on disease progression in secondary progressive multiple sclerosis (ASCEND): a phase 3, randomised, double-blind, placebo-controlled trial with an open-label extension. *Lancet Neurol*. 2018;17.
21. Cohen JA, Cutter GR, Fischer JS, et al. Benefit of interferon beta-1a on MSFC progression in secondary progressive MS. *Neurology*. 2002;59:679–87.
22. Li R, Patterson KR, Bar-Or A. Reassessing B cell contributions in multiple sclerosis. *Nat Immunol*. 2018;19.
23. Hohlfeld R, Dornmair K, Meinl E, Wekerle H. The search for the target antigens of multiple sclerosis, part 1: autoreactive CD4+ T lymphocytes as pathogenic effectors and therapeutic targets. *Lancet Neurol*. 2016;15:198–209.
24. Hohlfeld R, Dornmair K, Meinl E, Wekerle H. The search for the target antigens of multiple sclerosis, part 2: CD8+ T cells, B cells, and antibodies in the focus of reverse-translational research. *Lancet Neurol*. 2016;15:317–31.
25. Hauser SL, Waubant E, Arnold DL, et al. B-cell depletion with rituximab in relapsing-remitting multiple sclerosis. *N Engl J Med*. 2008;358:676–88.
26. Montalban X, Hauser SL, Kappos L, et al. Ocrelizumab versus placebo in primary progressive multiple sclerosis. *N Engl J Med*. 2017;376.
27. Kappos L, Li D, Calabresi PA, et al. Ocrelizumab in relapsing-remitting multiple sclerosis: a phase 2, randomised, placebo-controlled, multicentre trial. *Lancet*. 2011;378.
28. Hawker K, O'Connor P, Freedman MS, et al. Rituximab in patients with primary progressive multiple sclerosis: results of a randomized double-blind placebo-controlled multicenter trial. *Ann Neurol*. 2009;66.
29. Wolinsky JS, Arnold DL, Brochet B, et al. Long-term follow-up from the ORATORIO trial of ocrelizumab for primary progressive multiple sclerosis: a post-hoc analysis from the ongoing open-label extension of the randomised, placebo-controlled, phase 3 trial. *Lancet Neurol*. 2020;19.
30. Hauser SL, Bar-Or A, Comi G, et al. Ocrelizumab versus interferon beta-1a in relapsing multiple sclerosis. *N Engl J Med*. 2017;376.
31. Hauser SL, Bar-Or A, Cohen JA, et al. Ofatumumab versus teriflunomide in multiple sclerosis. *N Engl J Med*. 2020;383.
32. Hohlfeld R, Meinl E. Ocrelizumab in multiple sclerosis: markers and mechanisms. *Lancet Neurol*. 2017;16:259–61.
33. Meinl E, Hohlfeld R. CD20(+) T cells as pathogenic players and therapeutic targets in MS. *Ann Neurol*. 2021;90:722–4.
34. Franks SE, Getahun A, Hogarth PM, Cambier JC. Targeting B cells in treatment of autoimmunity. *Curr Opin Immunol*. 2016;43.
35. Obermeier B, Mentele R, Malotka J, et al. Matching of oligoclonal immunoglobulin transcriptomes and proteomes of cerebrospinal fluid in multiple sclerosis. *Nat Med*. 2008;14.
36. Brandle SM, Cerina M, Weber S, et al. Cross-reactivity of a pathogenic autoantibody to a tumor antigen in GABAA receptor encephalitis. *Proc Natl Acad Sci U S A*. 2021;118.
37. Baecher-Allan C, Kaskow BJ, Weiner HL. Multiple sclerosis: mechanisms and immunotherapy. *Neuron*. 2018;97.
38. Machado-Santos J, Saji E, Troscher AR, et al. The compartmentalized inflammatory response in the multiple sclerosis brain is composed of tissue-resident CD8+ T lymphocytes and B cells. *Brain*. 2018;141.
39. Klein C, Lammens A, Schafer W, et al. Epitope interactions of monoclonal antibodies targeting CD20 and their relationship to functional properties. *MAbs*. 2013;5:22–33.
40. Bar-Or A, Grove RA, Austin DJ, et al. Subcutaneous ofatumumab in patients with relapsing-remitting multiple sclerosis: the MIRROR study. *Neurology*. 2018;90.
41. Bar-Or A, O'Brien SM, Sweeney ML, Fox EJ, Cohen JA. Clinical perspectives on the molecular and pharmacological attributes of anti-CD20 therapies for multiple sclerosis. *CNS Drugs*. 2021;35:985–97.
42. Magliozzi R, Howell O, Vora A, et al. Meningeal B-cell follicles in secondary progressive multiple sclerosis associate with early onset of disease and severe cortical pathology. *Brain*. 2007;130.
43. Kramann N, Neid K, Menken L, et al. Increased meningeal T and plasma cell infiltration is associated with early subpial cortical demyelination in common marmosets with experimental autoimmune encephalomyelitis. *Brain Pathol*. 2015;25:276–86.
44. Martin Mdel P, Cravens PD, Winger R, et al. Depletion of B lymphocytes from cerebral perivascular spaces by rituximab. *Arch Neurol*. 2009;66:1016–20.
45. Rubenstein JL, Combs D, Rosenberg J, et al. Rituximab therapy for CNS lymphomas: targeting the leptomeningeal compartment. *Blood*. 2003;101:466–8.
46. Fereidan-Esfahani M, Bruck W, Weber MS. Targeting central nervous system B cells in progression of multiple sclerosis: is intrathecal anti-CD20 a therapeutic option? *JAMA Neurol*. 2015;72:1407–8.
47. Zecca C, Bovis F, Novi G, et al. Treatment of multiple sclerosis with rituximab: a multicentric Italian-Swiss experience. *Mult Scler*. 2020;26.
48. Yamout BI, El-Ayoubi NK, Nicolas J, et al. Safety and efficacy of rituximab in multiple sclerosis: a retrospective observational study. *J Immunol Res*. 2018.
49. Spelman T, Frisell T, Piehl F, Hillert J. Comparative effectiveness of rituximab relative to IFN- $\beta$  or glatiramer acetate in relapsing-remitting MS from the Swedish MS registry. *Mult Scler*. 2018;24.
50. Scotti B, Disanto G, Sacco R, et al. Effectiveness and safety of rituximab in multiple sclerosis: an observational study from Southern Switzerland. *PLoS One*. 2018;13.
51. Salzer J, Svenningsson R, Alping P, et al. Rituximab in multiple sclerosis: a retrospective observational study on safety and efficacy. *Neurology*. 2016;87.
52. Ingle GT, Sastre-Garriga J, Miller DH, Thompson AJ. Is inflammation important in early PPMS? A longitudinal MRI study. *J Neurol Neurosurg Psychiatry*. 2005;76.
53. Montalban X, Hauser SL, Kappos L, et al. Ocrelizumab versus placebo in primary progressive multiple sclerosis. *N Engl J Med*. 2017;376.
54. Bhargava P, Wicken C, Smith MD, et al. Trial of intrathecal rituximab in progressive multiple sclerosis patients with evidence of leptomeningeal contrast enhancement. *Mult Scler Relat Disord*. 2019;30.
55. Ocrevus. Available from: <https://www.ema.europa.eu/en/medicines/human/EPAR/ocrevus>.

56. Graf J, Albrecht P, Goebels N, Aktas O, Hartung HP [Ocrelizumab for treatment of multiple sclerosis]. *Nervenarzt*. 2020;91:722–34.
57. Montalban X, Hauser SL, Kappos L, et al. Ocrelizumab versus placebo in primary progressive multiple sclerosis. *N Engl J Med*. 2017;376:209–20.
58. Turner B, Cree BAC, Kappos L, et al. Ocrelizumab efficacy in subgroups of patients with relapsing multiple sclerosis. *J Neurol*. 2019;266.
59. Tedder TF. CD19: a promising B cell target for rheumatoid arthritis. *Nat Rev Rheumatol*. 2009;5.
60. Agius MA, Klodowska-Duda G, Maciejowski M, et al. Safety and tolerability of inebilizumab (MEDI-551), an anti-CD19 monoclonal antibody, in patients with relapsing forms of multiple sclerosis: results from a phase 1 randomised, placebo-controlled, escalating intravenous and subcutaneous dose study. *Mult Scler*. 2019;25.
61. Frischer JM, Bramow S, Dal-Bianco A, et al. The relation between inflammation and neurodegeneration in multiple sclerosis brains. *Brain*. 2009;132.
62. De Groot CJ, Bergers E, Kamphorst W, et al. Post-mortem MRI-guided sampling of multiple sclerosis brain lesions: increased yield of active demyelinating and (p)reactive lesions. *Brain*. 2001;124:1635–45.
63. Van der Poel M, Ulas T, Mizze MR, et al. Transcriptional profiling of human microglia reveals grey-white matter heterogeneity and multiple sclerosis-associated changes. *Nat Commun*. 2019;10:1139.
64. Unterrainer M, Mahler C, Vomacka L, et al. TSPO PET with [(18)F]GE-180 sensitively detects focal neuroinflammation in patients with relapsing-remitting multiple sclerosis. *Eur J Nucl Med Mol Imaging*. 2018;45:1423–31.
65. Mahler C, Schumacher AM, Unterrainer M, et al. TSPO PET imaging of natalizumab-associated progressive multifocal leukoencephalopathy. *Brain*. 2021;144:2683–95.
66. Giannetti P, Politis M, Su P, et al. Microglia activation in multiple sclerosis black holes predicts outcome in progressive patients: an in vivo [(11)C](R)-PK11195-PET pilot study. *Neurobiol Dis*. 2014;65:203–10.
67. O'Loughlin E, Madore C, Lassmann H, Butovsky O. Microglial phenotypes and functions in multiple sclerosis. *Cold Spring Harb Perspect Med*. 2018;8.
68. Lisak RP, Benjamins JA, Bealmear B, et al. Differential effects of Th1, monocyte/macrophage and Th2 cytokine mixtures on early gene expression for molecules associated with metabolism, signaling and regulation in central nervous system mixed glial cell cultures. *J Neuroinflammation*. 2009;6:4.
69. Karamita M, Barnum C, Mobius W, et al. Therapeutic inhibition of soluble brain TNF promotes remyelination by increasing myelin phagocytosis by microglia. *JCI Insight*. 2017;2.
70. Lampron A, Laroche A, Laflamme N, et al. Inefficient clearance of myelin debris by microglia impairs remyelinating processes. *J Exp Med*. 2015;212:481–95.
71. Boyd A, Zhang H, Williams A. Insufficient OPC migration into demyelinated lesions is a cause of poor remyelination in MS and mouse models. *Acta Neuropathol*. 2013;125:841–59.
72. Franklin RJ, Goldman SA. Glia disease and repair-remyelination. *Cold Spring Harb Perspect Biol*. 2015;7:a020594.
73. Djedovic N, Stanisavljevic S, Jevtic B, et al. Anti-encephalitogenic effects of ethyl pyruvate are reflected in the central nervous system and the gut. *Biomed Pharmacother*. 2017;96:78–85.
74. Mi S, Miller RH, Tang W, et al. Promotion of central nervous system remyelination by induced differentiation of oligodendrocyte precursor cells. *Ann Neurol*. 2009;65:304–15.
75. Mi S, Miller RH, Lee X, et al. LINGO-1 negatively regulates myelination by oligodendrocytes. *Nat Neurosci*. 2005;8:745–51.
76. Mi S, Lee X, Shao Z, et al. LINGO-1 is a component of the Nogo-66 receptor/p75 signaling complex. *Nat Neurosci*. 2004;7:221–8.
77. Ahmed Z, Douglas MR, John G, Berry M, Logan A. AMIGO3 is an NgR1/p75 co-receptor signalling axon growth inhibition in the acute phase of adult central nervous system injury. *PLoS One*. 2013;8:e61878.
78. Mi S, Hu B, Hahm K, et al. LINGO-1 antagonist promotes spinal cord remyelination and axonal integrity in MOG-induced experimental autoimmune encephalomyelitis. *Nat Med*. 2007;13:1228–33.
79. Rudick RA, Mi S, Sandrock AW Jr. LINGO-1 antagonists as therapy for multiple sclerosis: in vitro and in vivo evidence. *Expert Opin Biol Ther*. 2008;8:1561–70.
80. Klistorner A, Chai Y, Leocani L, et al. Assessment of opicinumab in acute optic neuritis using multifocal visual evoked potential. *CNS Drugs*. 2018;32:1159–71.
81. Cadavid D, Mellion M, Hupperts R, et al. Safety and efficacy of opicinumab in patients with relapsing multiple sclerosis (SYNERGY): a randomised, placebo-controlled, phase 2 trial. *Lancet Neurol*. 2019;18:845–56.
82. Cadavid D, Balcer L, Galetta S, et al. Predictors of response to opicinumab in acute optic neuritis. *Ann Clin Transl Neurol*. 2018;5:1154–62.
83. Cadavid D, Balcer L, Galetta S, et al. Safety and efficacy of opicinumab in acute optic neuritis (RENEW): a randomised, placebo-controlled, phase 2 trial. *Lancet Neurol*. 2017;16:189–99.
84. Petrillo J, Balcer L, Galetta S, et al. Initial impairment and recovery of vision-related functioning in participants with acute optic neuritis from the RENEW trial of opicinumab. *J Neuroophthalmol*. 2019;39:153–60.
85. Ranger A, Ray S, Szak S, et al. Anti-LINGO-1 has no detectable immunomodulatory effects in preclinical and phase 1 studies. *Neurol Neuroimmunol Neuroinflamm*. 2018;5:e417.
86. Aktas O, Albrecht P, Hartung HP. Optic neuritis as a phase 2 paradigm for neuroprotection therapies of multiple sclerosis: update on current trials and perspectives. *Curr Opin Neurol*. 2016;29.
87. Hanf KJM, Arndt JW, Liu Y, et al. Functional activity of anti-LINGO-1 antibody opicinumab requires target engagement at a secondary binding site. *MAbs*. 2020;12:1713648.
88. Margoni M, Preziosa P, Filippi M, Rocca MA. Anti-CD20 therapies for multiple sclerosis: current status and future perspectives. *J Neurol*. 2021.
89. Contentti EC, Correale J. Bruton's tyrosine kinase inhibitors: a promising emerging treatment option for multiple sclerosis. *Expert Opin Emerg Drugs*. 2020;25.
90. Torke S, Weber MS. Inhibition of Bruton's tyrosine kinase as a novel therapeutic approach in multiple sclerosis. *Expert Opin Investig Drugs*. 2020;29.
91. Hendriks RW, Yuvaraj S, Kil LP. Targeting Bruton's tyrosine kinase in B cell malignancies. *Nat Rev Cancer*. 2014;14:219–32.
92. Montalban X, Arnold DL, Weber MS, et al. Placebo-controlled trial of an oral BTK inhibitor in multiple sclerosis. *N Engl J Med*. 2019;380.
93. MSVirtual – Poster Abstracts. *Mult Scler J*. 2020;(26):118–659.
94. Reich DS, Honeycutt WD, Laganke C, et al. Efficacy and safety outcomes in patients with relapsing forms of MS treated with the CNS-penetrating BTK inhibitor SAR442168: results from the phase 2b trial. *Eur J Neurol*. 2020;27.

95. Dangond F, Donnelly A, Hohlfeld R, et al. Facing the urgency of therapies for progressive MS - a Progressive MS Alliance proposal. *Nat Rev Neurol*. 2021;17:185–92.
96. Petzold A, Balcer LJ, Calabresi PA, et al. Retinal layer segmentation in multiple sclerosis: a systematic review and meta-analysis. *Lancet Neurol*. 2017;16:797–812.
97. Cayre M, Falque M, Mercier O, Magalon K, Durbec P. Myelin repair: from animal models to humans. *Front Cell Neurosci*. 2021;15:604865.
98. Bodini B, Veronese M, Garcia-Lorenzo D, et al. Dynamic imaging of individual remyelination profiles in multiple sclerosis. *Ann Neurol*. 2016;79:726–38.
99. Frischer JM, Weigand SD, Guo Y, et al. Clinical and pathological insights into the dynamic nature of the white matter multiple sclerosis plaque. *Ann Neurol*. 2015;78:710–21.
100. Confavreux C, Vukusic S. Age at disability milestones in multiple sclerosis. *Brain*. 2006;129:595–605.
101. Abdelhak A, Weber MS, Tumani H. Primary progressive multiple sclerosis: putting together the Puzzle. *Front Neurol*. 2017;8:234.
102. Ontaneda D, Thompson AJ, Fox RJ, Cohen JA. Progressive multiple sclerosis: prospects for disease therapy, repair, and restoration of function. *Lancet*. 2017;389:1357–66.
103. Graf J, Aktas O, Rejdak K, Hartung HP. Monoclonal antibodies for multiple sclerosis: an update. *BioDrugs*. 2019;33:61–78.
104. Hawker K, O'Connor P, Freedman MS, et al. Rituximab in patients with primary progressive multiple sclerosis: results of a randomized double-blind placebo-controlled multicenter trial. *Ann Neurol*. 2009;66.
105. Cadavid D, Balcer L, Galetta S, et al. Safety and efficacy of opicinumab in acute optic neuritis (RENEW): a randomised, placebo-controlled, phase 2 trial. *Lancet Neurol*. 2017;16.
106. Hartung HP, Derfuss T, Cree BA, et al. Efficacy and safety of temelimab in multiple sclerosis: Results of a randomized phase 2b and extension study. *Mult Scler*. 2021;13524585211024997.

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.