

# True seals achieved global distribution by breaking Bergmann's rule

James P. Rule,<sup>1,2,3</sup>  Felix G. Marx,<sup>4,5</sup>  Alistair R. Evans,<sup>2,6</sup>  Erich M.G. Fitzgerald,<sup>2,6,7,8</sup>  
 and Justin W. Adams<sup>1,2</sup> 

<sup>1</sup>Department of Anatomy and Developmental Biology, Biomedicine Discovery Institute, Monash University, Clayton, VIC 3800, Australia

<sup>2</sup>Geosciences Museums Victoria, Melbourne, VIC 3001, Australia

<sup>3</sup>E-mail: jrule.palaeo@gmail.com

<sup>4</sup>Museum of New Zealand Te Papa Tongarewa, Wellington 6011, New Zealand

<sup>5</sup>Department of Geology, University of Otago, Dunedin 9016, New Zealand

<sup>6</sup>School of Biological Sciences, Monash University, Melbourne, VIC 3800, Australia

<sup>7</sup>National Museum of Natural History, Smithsonian Institution, Washington, D.C. 20013

<sup>8</sup>Department of Life Sciences, Natural History Museum, London SW7 5BD, United Kingdom

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True seals (phocids) have achieved a global distribution by crossing the equator multiple times in their evolutionary history. This is remarkable, as warm tropical waters are regarded as a barrier to marine mammal dispersal and—following Bergmann's rule—may have limited crossings to small-bodied species only. Here, we show that ancestral phocids were medium sized and did not obviously follow Bergmann's rule. Instead, they ranged across a broad spectrum of environmental temperatures, without undergoing shifts in temperature- or size-related evolutionary rates following dispersals across the equator. We conclude that the tropics have not constrained phocid biogeography.

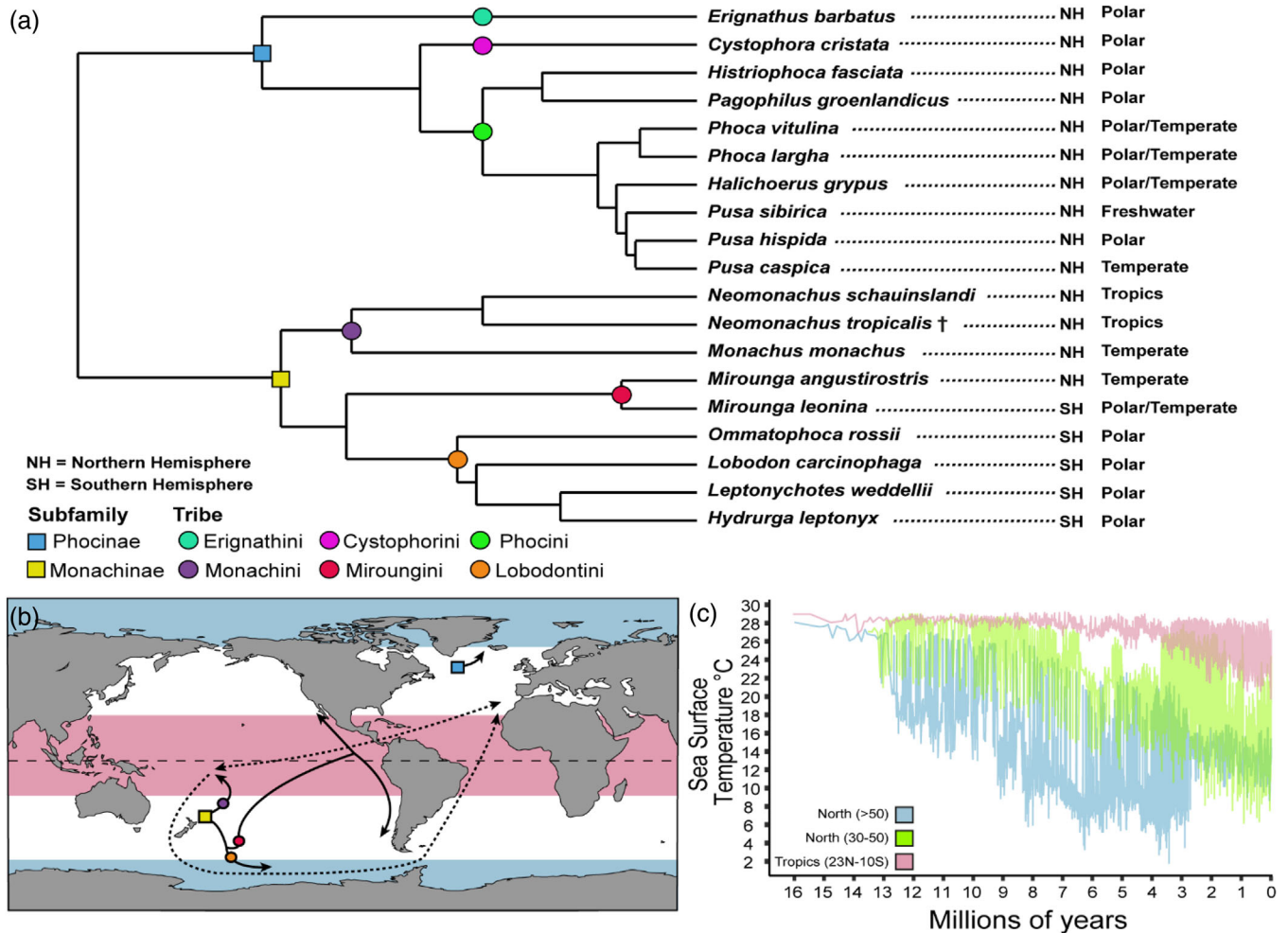
**KEY WORDS:** Antitropical distribution, Bergmann's rule, biogeography, body size, Phocidae, sea surface temperature.

The global distribution of true seals reflects their success as secondarily aquatic tetrapods. Since returning to the water, they have evolved a wide range of body sizes (Churchill et al. 2015), adapted to thermoregulation in aquatic environments (Liwanag et al. 2012), and dispersed around the globe (Berta et al. 2015). Their current range includes both polar regions and the tropics (Berta et al. 2018; Fig. 1), resulting in conflicting hypotheses as to whether they originated in a cold (Fulton and Strobeck 2010; Davies 1958a,b) or warm environment (Repenning et al. 1979; Deméré et al. 2003; Fyler et al. 2005; Mason et al. 2020).

Modern marine mammals are mostly cold adapted, with their relatively large size and blubber insulation putting them at risk of overheating in warm environments (Davies 1958a, 1963; Holt et al. 2020). Consequently, they are thought to follow Bergmann's rule (Bergmann 1847), which postulates an inverse relationship

between body size and environmental temperature (Sepúlveda et al. 2013; Torres-Romero et al. 2016; Adamczak et al. 2020). If so, warm tropical waters should be home to relatively small body forms only, whereas larger species might be expected to show antitropical distributions (Holt et al. 2020).

True seals arose in the Northern Hemisphere (Fulton and Strobeck 2010; Fig. 1). Their earliest (Miocene—Pliocene) Southern Hemisphere representatives are notably small (Rule et al. 2021a). This perhaps indicates restricted cross-equatorial dispersal consistent with Bergmann's rule, as the latitudinal thermal gradient was present throughout the Neogene (Fig. 1c). Yet phocids appear to have crossed the tropics several times during their evolutionary history (Rule et al. 2020a; Fig. 1), in stark contrast to the single major southern dispersal of their otariid cousins, the fur seals and sea lions (Yonezawa et al. 2009; Churchill



**Figure 1.** Biogeography of crown Phocidae. (a) Phylogeny of extant true seals (Rule et al. 2021b) with geographic distributions. (b) Dispersals for crown-phocids (modified from Rule et al. 2020a), with the tropics in light red and polar environments in light blue. (c) Variation of Sea Surface Temperature by latitude through time (Herbert et al. 2016), demonstrating consistently high temperatures in the tropics, and broader Sea Surface Temperatures across latitudes closer to the present.

et al. 2014). Here, we test phocid dispersal capabilities by testing whether (i) equatorial crossings were accompanied by notable shifts in body size and/or environmental temperature, and (ii) Bergmann's rule truly applies to them.

## Materials and Methods

We based our study on the phylogeny of Rule et al. (2020a, 2021b), excluding tips with no phenotypic and/or environmental data. Analyses were carried out in RStudio version 1.2.1335 (R Version 3.6.0) using the packages “ape,” “phytools,” “geiger,” “ratematrix,” “nlme,” and “RRphylo” (Paradis et al. 2004; Revell 2012; Pennell et al. 2014; Caetano and Harmon 2017; Pinheiro et al. 2017; Castiglione et al. 2018). Taxa resolved as ancestors in the original tree were assigned artificial branch lengths of 0.01 million years to enable analyses to run.

Maximum and minimum total body lengths for each specimen were taken from the literature (Stirling 1971; King 1983; Modig 1996; Andersen et al. 1999; Bininda-Emonds and Gittleman 2000; Samaranch and Gonzalez 2000; Lindenfors et al. 2002; Laws et al. 2003; Rogers 2009; Churchill et al. 2015; Valenzuela-Toro et al. 2016; Dewaele et al. 2017; Rule et al. 2020b, 2021a; Tables A3, A4) or, where unavailable (for 12 out of 17 extinct taxa), maximum total body length was estimated following Rule et al. (2020b; Tables A1, A2). All length data were then  $\log_{10}$  transformed prior to analysis. For extant species, we used the minimum, median, and maximum sea surface temperature (SST) of their entire geographic range (IUCN Red List) as a proxy for environmental temperature (Appendix). For extinct taxa, median SST estimates aligning with the tip age and geographic region of the fossils in question were taken from the literature (Dowsett and Wiggs 1992; Barrick et al. 1993; Warne 2005; Amiot et al. 2008; Dowsett et al. 2012; Herbert et al. 2016).

**Table 1.** Summary statistics on datasets analyses. TBL =  $\log_{10}$  total body length, SST = sea surface temperature.

Dataset	N	Mean	Median	Minimum	Maximum	Standard Deviation
Maximum TBL extant + extinct	36	2.31	2.29	1.83	2.75	0.17
Maximum TBL minus ancestors	26	2.37	2.37	2.11	2.75	0.15
Maximum TBL extant only	19	2.37	2.37	2.11	2.75	0.17
Minimum TBL extant only	19	2.32	2.33	2.52	2.52	0.13
Median SST extant + extinct	32	13.60	14.40	4.00	27.00	7.01
Median SST minus ancestors	26	12.32	12.00	4.00	27.00	7.02
Minimum SST extant only	19	3.47	0.00	0.00	24.00	7.63
Median SST extant only	19	10.37	7.50	4.00	27.00	7.12
Maximum SST extant only	19	17.37	15.00	8.00	30.00	8.01

We used median SST (Tables A5, A6) to enable direct comparisons between extant and extinct phocids, and to account for migratory movements meant to avoid seasonal extremes.

We examined the evolution of maximum total body length and median sea surface temperature (Table 1) for (i) extant species only, (ii) extant + extinct species minus ancestors (to rule out the possibility of ultrashort branch lengths producing artificial shifts), and (iii) the complete phylogeny via “RRphylo” (Castiglione et al. 2018, 2019a,b). For extant species only, we also analyzed minimum total body length, and minimum and maximum SST, to test the effects of extremes on the evolutionary analyses. Ancestral states of both traits ( $\log_{10}$  total body length and SST) were estimated for the extant and complete datasets only. We tested for evolutionary rate shifts using the *auto-recognize* feature of the *search.shift* function (Castiglione et al. 2018). Finally, we used the function *overfitRR* to measure the uncertainty associated with our RRphylo results.

We ran two sets of analyses to test for a possible relationship between  $\log_{10}$  total body length and sea surface temperature (i.e., Bergmann’s rule). First, we regressed maximum  $\log_{10}$  total body length against median SST via a linear regression and phylogenetic generalized least squares. Second, we tested for correlated evolution via a Bayesian Markov chain Monte Carlo (MCMC) analysis of evolutionary rate matrices, as implemented in the package “ratematrix” (Caetano and Harmon 2017). We ran two chains for 1,000,000 generations, sampling every 1000 generations and discarding the first 25% as burn-in. We checked the ac-

ceptance ratio for the two chain logs, and tested for convergence between them. When convergence was achieved, we merged the two MCMC chains and plotted the rate matrix to test for an evolutionary correlation between the two traits.

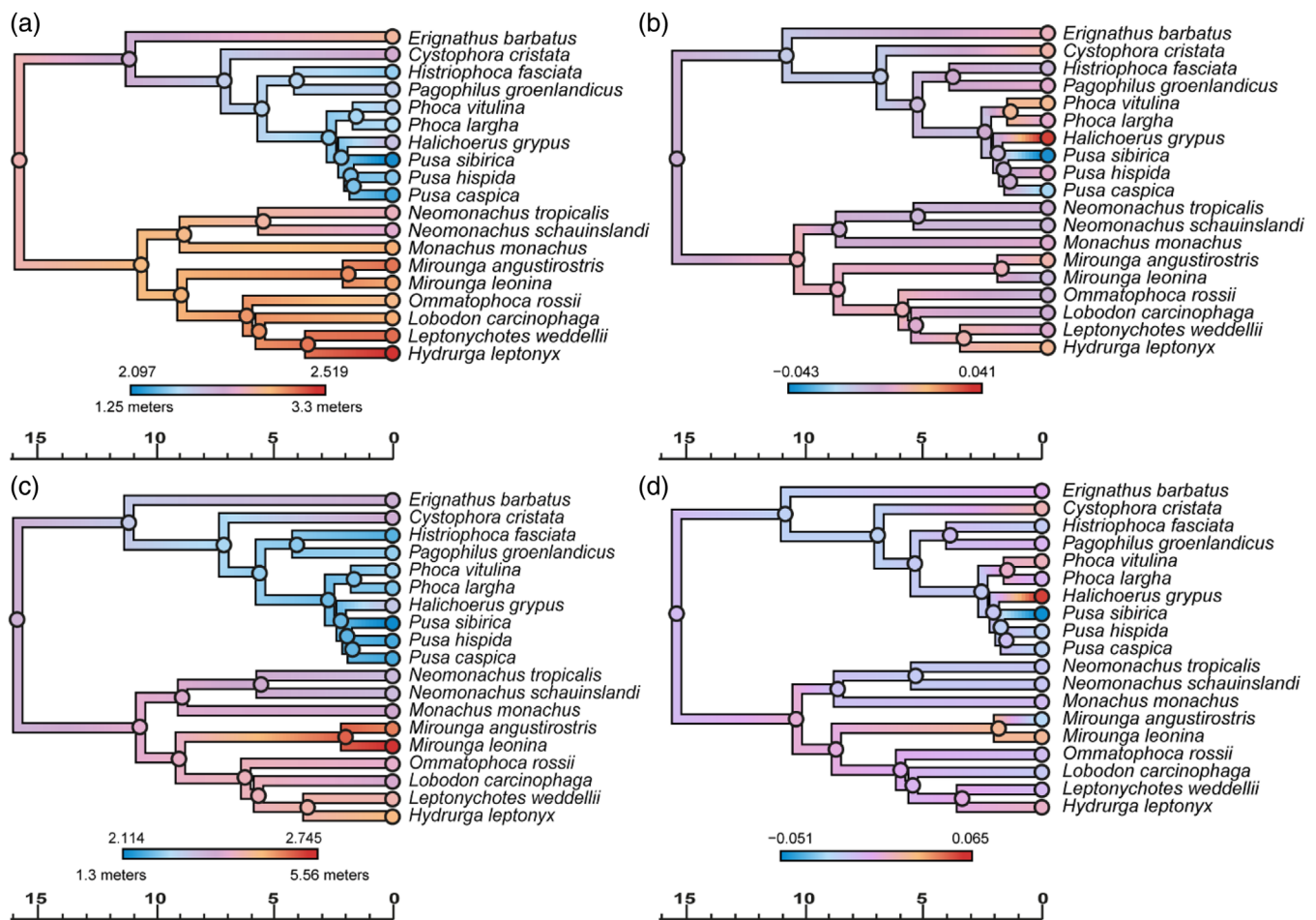
## Results

For the evolutionary rate shift analyses, including ancestors in the phylogeny did not result in additional evolutionary shifts (Tables 3, 4), and the results of the extant + extinct evolutionary rate analyses were better supported than the extant only analyses (Tables 2, A7; Fig. A1). Extant-only ancestral state estimations suggest that archaic phocines were far smaller (1.97–2.12 m) than monachines (2.56–2.93 m), with their last common ancestor being 2.32–2.50 m (Fig. 2). Phocids as a whole appear adapted to cold water (<12°C), with only monk seals being tolerant of warmer environments (Fig. 3). Taking into account fossil taxa reveals a more even pattern, with ancestral phocids, phocines, and monachines all showing a similar range of body lengths (1.64–2.25 m) and environmental temperatures (~19°C) (Fig. 4).

Both the extant (Fig. 2) and the extant + extinct (Fig. 4) datasets show little variation in evolutionary rates for  $\log_{10}$  total body length. Nevertheless, significant decreases in the rate of body size evolution characterize Antarctic seals (lobodontins) + elephant seals (ARD [Actual Rate Difference]: -0.008), and monk seals (ARD: -0.006) in the extant-only datasets, and stem phocids in the extant + extinct dataset (ARDs: -0.019 and

**Table 2.** OverfitRR results for the 95% confidence intervals of the root value obtained by the RRphylo analysis. TBL = log<sub>10</sub> total body length.

Dataset	Root Value	2.5% CI	97.5% CI
Extant + extinct maximum TBL	2.35	2.35	2.36
Minus ancestors maximum TBL	2.36	2.36	2.36
Extant only minimum TBL	2.37	2.16	2.38
Extant only maximum TBL	2.4	2.2	2.42
Extant + extinct median SST	19.21	16.79	19.28
Minus ancestors median SST	14.18	14.62	14.81
Extant only Minimum SST	1.74	0.18	0.4
Extant only Median SST	7.15	5.21	25.8
Extant only maximum SST	12.58	10.12	27.62

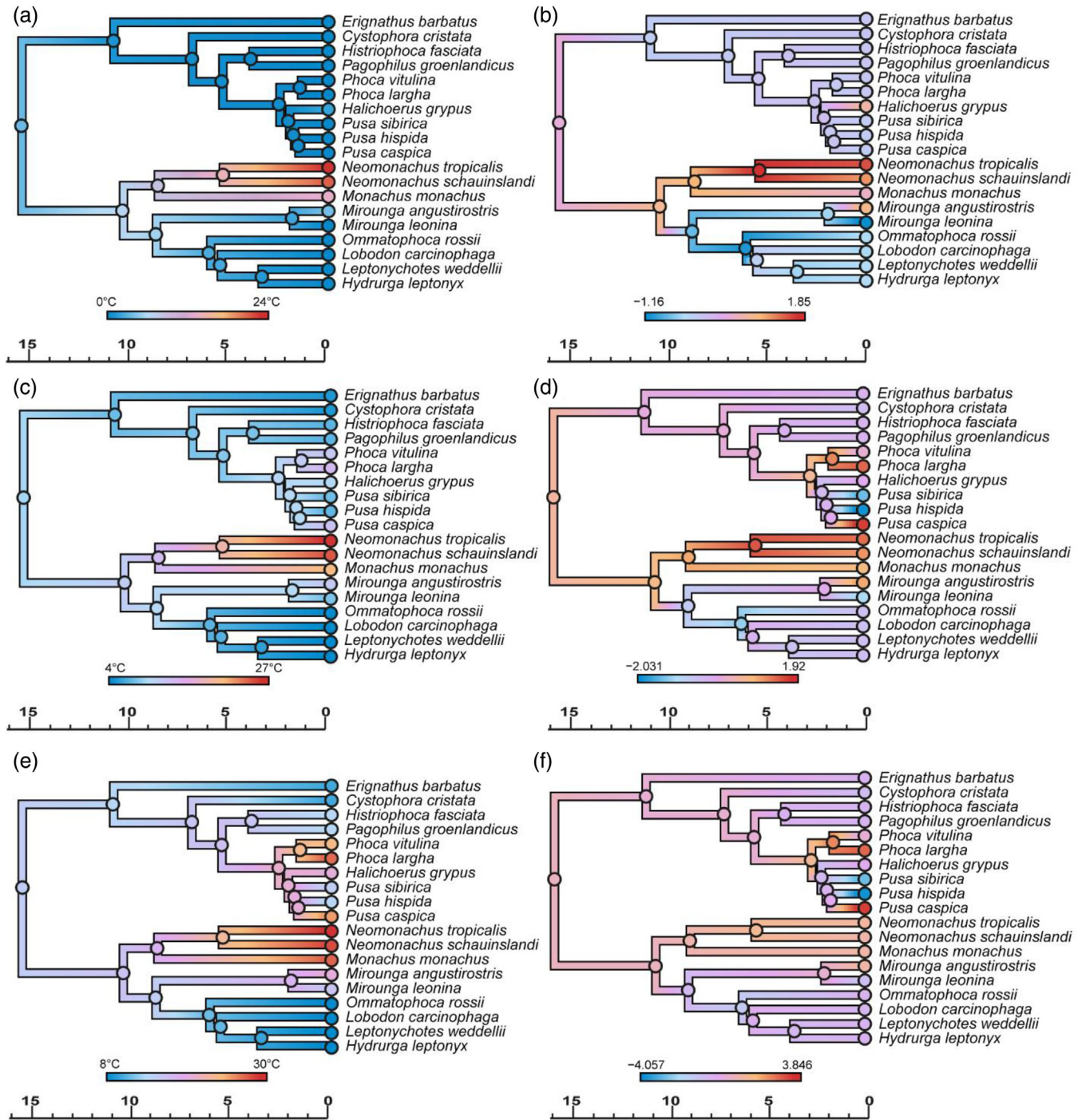
**Figure 2.** Evolution of body size in extant true seals estimated by RRphylo analysis, using phylogeny from Rule et al. (2021b). (a) Ancestral state estimation and (b) evolutionary rates for Log<sub>10</sub> minimum total body length. (c) Ancestral state estimation and (d) evolutionary rates for Log<sub>10</sub> maximum total body length. Timescales in millions of years.

–0.022; Table 3). There was more variation in evolutionary rates for sea surface temperature within the monachine clade than the phocine clade (Figs. 3–4). The extant data furthermore suggest rate decreases associated with a shift toward colder minimum SSTs for Phocinae and colder maximum SSTs for Monachinae;

however, these significant evolutionary rate decreases disappear when fossils are included (Table 4).

Neither the linear regression ( $P = 0.23$ ) nor the PGLS ( $P = 0.75$ ) show any relationship between body length and environmental temperature (Fig. 5; Tables A8, A9). For the evolutionary





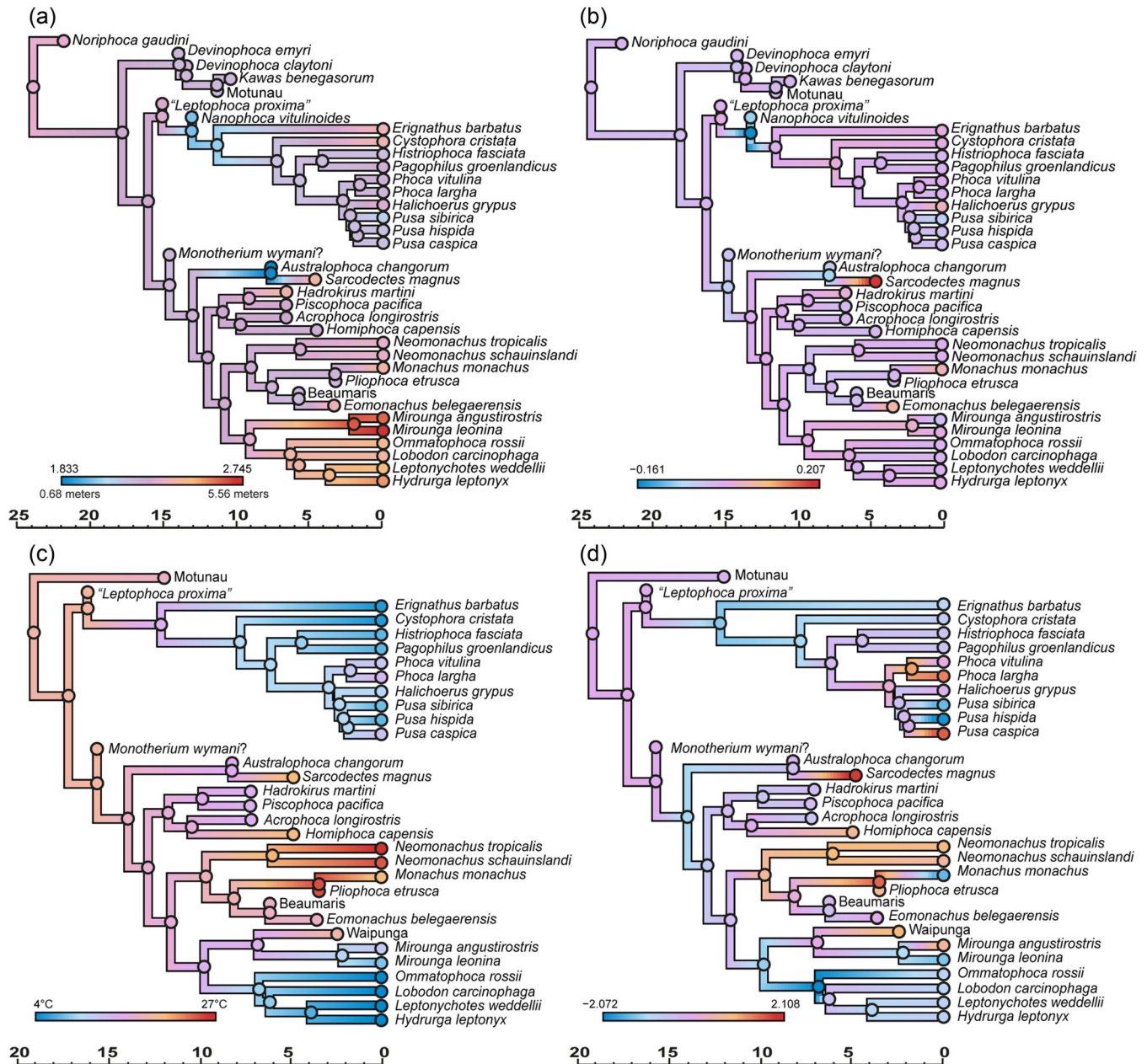
**Figure 3.** Evolution of sea surface temperature (SST) in extant true seals estimated by RRphylo analysis, using phylogeny from Rule et al. (2021b). Ancestral state estimation (a) and evolutionary rates (b) for minimum SST. Ancestral state estimation (c) and evolutionary rates (d) for median SST. Ancestral state estimation (e) and evolutionary rates (f) for maximum SST. Timescales in millions of years.

rate matrix analysis, convergence was achieved between the two MCMC chains after 1,000,000 generations (Figs. A2–A6; Tables A10, A11). The posterior distribution of the evolutionary rate matrices (Figs. 6, A7) of the merged MCMC chains shows no evolutionary correlation between  $\log_{10}$  total body length and sea surface temperature.

## Discussion

### EVOLUTION OF BODY SIZE AND SST IN TRUE SEALS

Previous studies disagreed on whether early phocids were small (Churchill et al. 2015) or large (Wyss 1994). In isolation, our extant phylogeny supports intermediate ancestral sizes (Fig. 2), which is also supported when fossil taxa are taken into account

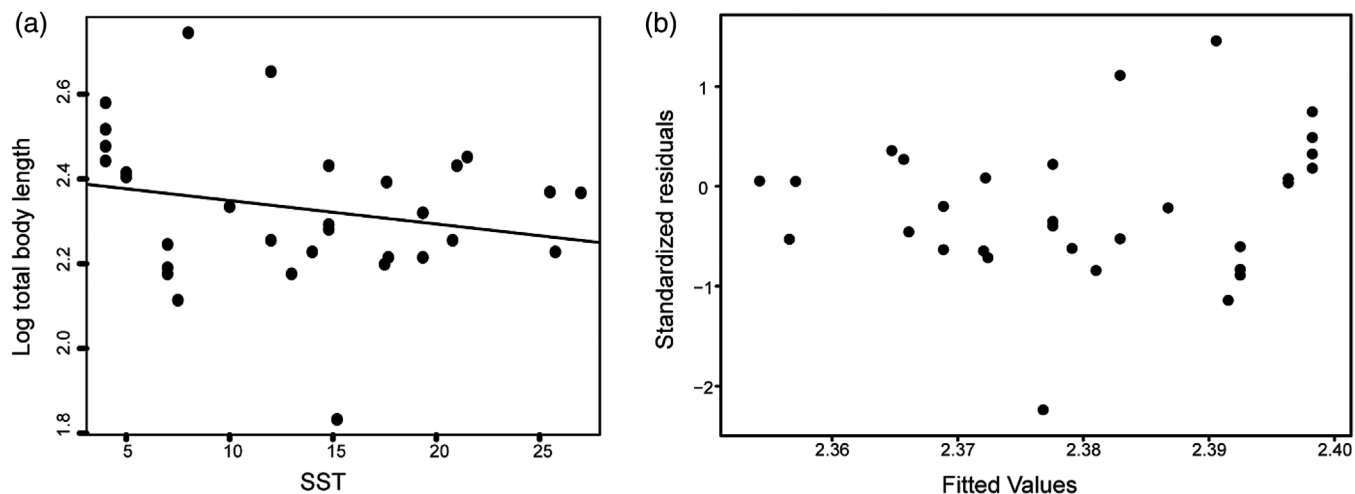


**Figure 4.** Evolution of body size and SST in extant and extinct true seals estimated by RRphylo analysis, using phylogeny from Rule et al. (2021b). Ancestral state estimation (a) and evolutionary rates (b) for  $\log_{10}$  total body length. Ancestral state estimation (c) and evolutionary rates (d) for median SST. Timescales in millions of years.

(Fig. 4). Both phocines and monachines waxed and waned in size over time (Wyss 1994; Churchill et al. 2015; Valenzuela-Toro et al. 2016; Dewaele et al. 2017; Rule et al. 2020b, 2021a), and between them gave rise to both the smallest (0.68 m) and the largest (>5 m) seals known to date. Our results suggest that these extremes represent derived conditions.

Extant-only ancestral state estimations of SST (Fig. 3) support a cold-water origin of true seals (Davies 1958a,b; Fulton and Strobeck 2010), as opposed to a more temperate range when fossils are included (Fig. 4). The latter suggests separate origins

for the cold-water adaptations of phocines and Antarctic seals (Repenning et al. 1979; Deméré et al. 2003; Fyler et al. 2005; Mason et al. 2020), with pagophily likely arising in response to Plio-Pleistocene cooling. Likewise, the tropical affinities of monk seals appear to be a derived condition. Overall, the modern contrast between polar and tropical phocids appears relictual, and largely reflects local extinctions of phocids at mid-latitudes during the late Neogene (Avery and Klein 2011; Valenzuela-Toro et al. 2013; Pimiento et al. 2017; Dewaele et al. 2018; Rule et al. 2019).



**Figure 5.** Regression analyses for log total body length and median SST in extant and extinct true seals. (a) Linear regression of log<sub>10</sub> total body length versus median SST (adjusted  $R^2 = 0.017$ ,  $P$ -value = 0.226). (b) Phylogenetic generalized least squares regression for log<sub>10</sub> total body length versus median SST ( $P$ -value = 0.753).

**Table 3.** Results of the *search.shift* analysis of log<sub>10</sub> total body length (TBL) in RRphylo. Only significant results are shown; for full results, see Supporting Information. ARD = Average Rate Difference.

Extinct + Extant maximum TBL			
Clade	ARD	$P$ -value	
<i>D. emyri</i> + <i>D. claytoni</i> + <i>Kawas</i> + Motunau	-0.019	<0.01	
<i>D. claytoni</i> + <i>Kawas</i> + Motunau	-0.022	<0.01	
Minus ancestors maximum TBL			
Clade	ARD	$P$ -value	
Monachini	-0.011	<0.01	
Extant only maximum TBL			
Clade	ARD	$P$ -value	
Lobodontini + Miroungini	-0.008	<0.03	
Extant only minimum TBL			
Clade	ARD	$P$ -value	
Monachinae	-0.006	<0.01	

Trait evolution is best assessed based on phylogenies including both extant and extinct taxa (Quental and Marshall 2010). This is supported by our findings, with extant + extinct datasets producing different, and more robust, results than those comprising living species only. Previous studies focusing on extant phocids likely underestimated their past ecological diversity (Davies 1958a,b; Fyler et al. 2005; Fulton and Strobeck 2010; Mason et al. 2020), which in turn may have prevented a more widespread extinction of the group during the late Neogene (Knope et al. 2020).

**Table 4.** Results of the *search.shift* analysis of sea surface temperature (SST) in RRphylo. Only significant results are shown; for full results, see Supporting Information. ARD = Average Rate Difference.

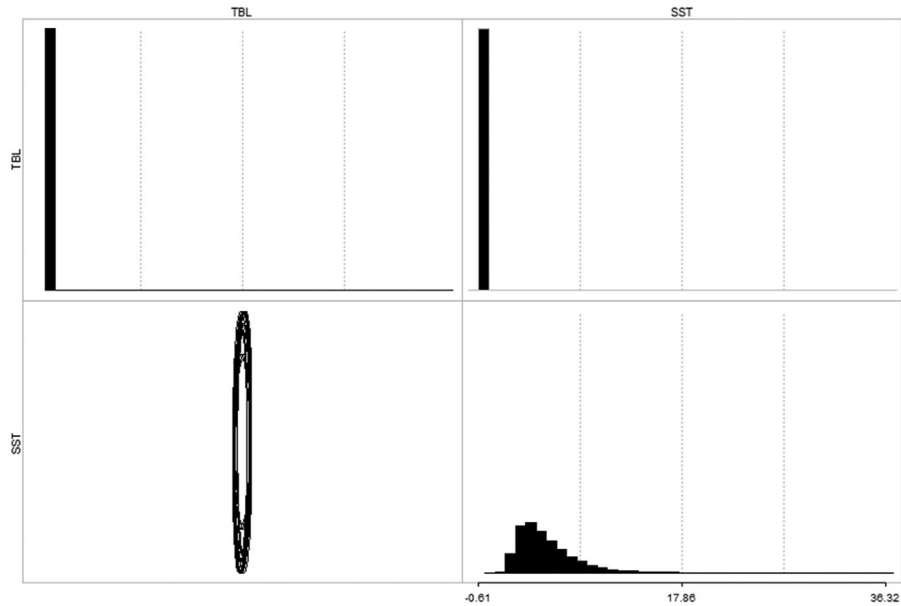
Minus ancestors median SST			
Clade	ARD	$P$ -value	
<i>Homiphoca</i> + <i>Hadrokiurus</i> + <i>Piscophoca</i> + <i>Acrophoca</i>	-0.369	<0.02	
Extant only maximum SST			
Clade	ARD	$P$ -value	
Lobodontini	-0.658	<0.01	
Lobodontini + Mirounga	-0.482	<0.03	
Extant only minimum SST			
Clade	ARD	$P$ -value	
<i>Cystophora</i> + Phocini	-0.594	<0.01	
Phocini	-0.522	<0.01	
<i>Halichoerus</i> + <i>Phoca</i> + <i>Pusa</i>	-0.387	<0.01	
<i>Pusa</i>	-0.370	0.03	

**THERMAL BARRIERS TO DISPERSAL**

True seals repeatedly crossed the tropics in the course of their evolution (Rule et al. 2020a), even though they are thought to hinder marine mammal dispersal (Davies 1963; Holt et al. 2020). A warm-water equatorial barrier could explain the small size of the oldest true seals from the Southern Hemisphere (Rule et al. 2020b, 2021a), but surprisingly is not evident in our evolutionary rate shift analysis. Overall, equatorial crossings for true seals are thus not obviously constrained by body size.

No shifts in SST were detected once extinct seals were taken into account (Table 4). Therefore, phocids appear tolerant of a





**Figure 6.** Posterior distribution of the evolutionary rate matrices for the merged MCMC chains. Histograms show the posterior distribution of evolutionary rate variance values for  $\log_{10}$  total body length (TBL, top left) and sea surface temperature (SST, bottom right); and pairwise evolutionary covariance values between  $\log_{10}$  total body length and sea surface temperature (top right). Ellipses (bottom left) are 50 bivariate distributions randomly sampled from the posterior distribution. The vertical orientation of the ellipses demonstrates that there is no evolutionary correlation between  $\log_{10}$  total body length and sea surface temperature. The elongated shape of the ellipses demonstrates that  $\log_{10}$  total body length has faster evolutionary rates than sea surface temperature.

broad range of environmental temperatures, which plausibly enabled them to invade both the tropics and polar environments with relative ease.

### BERGMANN'S RULE

Bergmann's rule is thought to restrict the body size of marine mammals at lower latitudes (Torres-Romero et al. 2016; Adamczak et al. 2020), which may limit cross-equatorial dispersals. The rule applies to fur seals and sea lions (Sepúlveda et al. 2013) but seemingly not phocids, with our regressions and evolutionary rate matrix analysis showing no relationship between total body length and SST (Figs. 5, 6; Tables A8, A9). Body size evolution in true seals was thus not obviously driven by temperature, and instead may reflect nutrient availability and/or feeding ecology (Dewaele et al. 2017, 2018; Rule et al. 2021a).

### GLOBAL DISPERSAL OF TRUE SEALS

Unlike fur seals, sea lions, and walruses—all of which remained restricted to the North Pacific for much of their evolution—true seals have long enjoyed a global distribution (Berta et al. 2018; Velez-Juarbe and Valenzuela-Toro 2019). Broad temperature tolerances may help to explain this pattern, with true seals being able to invade new environments relatively easily. By contrast, fur seals and sea lions only crossed into the Southern Hemisphere following Pliocene cooling and an attendant increase in productivity along the equator (Churchill et al. 2014). The

same cooling event produced sea-level fluctuations that impacted coastal habitats (with lowered sea levels eliminating shallow coastal waters) and likely disrupted the global distribution of phocids by driving their replacement with otariids at southern temperate latitudes (Boessenecker 2013; Valenzuela-Toro et al. 2013; Govender 2015; Pimiento et al. 2017; Rule et al. 2019). This idea is again consistent with our results, which suggest that—contrary to earlier suggestions (Ray 1976)—changing climates likely did not exceed the temperature tolerances of phocids as such. This suggests that phocids will be affected by physical oceanic and ecological changes from future climatic change, rather than directly by changes in temperature.

### AUTHOR CONTRIBUTIONS

JPR, FGM, ARE, and JWA conceived and designed the study. JPR collected the data. JPR and FGM analyzed the data. JPR drafted the initial version of the manuscript. All authors contributed to and approved the final versions of the manuscript.

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[Correction added on 13 MAY 2022, after first online publication: CAUL funding statement has been added.]

## CONFLICT OF INTEREST

The authors declare no conflict of interest.

## DATA ARCHIVING

Additional datasets and results are available as Supporting Information. The R-script (<https://doi.org/10.6084/m9.figshare.13515458>), input files (<https://doi.org/10.6084/m9.figshare.13515461>), and output data (<https://doi.org/10.6084/m9.figshare.13515470>) are available on the Figshare repository. Fossils used in this study are deposited in the following permanent and accessible institutions: Museum of New Zealand Te Papa Tongarewa, Canterbury Museum, Museo Paleontológico 'Egidio Feruglio', Museums Victoria, Muséum national d'Histoire naturelle, and Smithsonian Institution National Museum of Natural History

## LITERATURE CITED

- Adamczak, S.K., Pabst, D.A., McLellan, W.A. & Thorne, L.H. (2020) Do bigger bodies require bigger radiators? Insights into thermal ecology from closely related marine mammal species and implications for biogeographic rules. *J. Biogeogr.*, 47, 1193–1206.
- Amiot, R., Göhlich, U.B., Lecuyer, C., De Muizon, C., Cappelletta, H., Fourrel, F., Hérain, M.-A. & Martineau, F. (2008) Oxygen isotope compositions of phosphate from Middle Miocene–Early Pliocene marine vertebrates of Peru. *Palaeogeogr. Palaeoclimatol. Palaeoecol.*, 264, 85–92.
- Amson, E. & Muizon, C.D. (2014) A new durophagous phocid (Mammalia: Carnivora) from the late Neogene of Peru and considerations on monachine seals phylogeny. *Journal of Systematic Palaeontology*, 12, 523–548.
- Andersen, M., Hjelset, A., Gjertz, I., Lydersen, C. & Gulliksen, B. (1999) Growth, age at sexual maturity and condition in bearded seals (*Erigonathus barbatus*) from Svalbard, Norway. *Polar Biol.*, 21, 179–185.
- Avery, G. & Klein, R.G. (2011) Review of fossil phocid and otariid seals from the southern and western coasts of South Africa. *Trans. R. Soc. S. Afr.*, 66, 14–24.
- Barrick, R.E., Fischer, A.G. & Bohaska, D.J. (1993) Paleotemperatures versus sea level: oxygen isotope signal from fish bone phosphate of the Miocene Calvert Cliffs, Maryland. *Paleoceanography*, 8, 845–858.
- Bergmann, C. (1847) Ueber die Verhältnisse der Warmeökonomie der Thiere zu ihrer Grosse. *Gott. Stud.*, 1, 595–708.
- Berta, A., Churchill, M. & Boessenecker, R.W. (2018) The origin and evolutionary biology of pinnipeds: seals, sea lions, and walruses. *Ann. Rev. Earth Planet. Sci.*, 46, 203–228.
- Berta, A., Sumich, J.L. & Kovacs, K.M. (2015) Marine mammals: evolutionary biology. 3rd ed. Academic Press, Cambridge, MA.
- Bininda-Emonds, O.R. & Gittleman, J.L. (2000) Are pinnipeds functionally different from fissiped carnivores? The importance of phylogenetic comparative analyses. *Evolution*, 54, 1011–1023.
- Boessenecker, R.W. (2013) A new marine vertebrate assemblage from the Late Neogene Purisima Formation in Central California, part II: pinnipeds and cetaceans. *Geodiversitas*, 35, 815–940.
- Caetano, D.S. & Harmon, L.J. (2017) ratematrix: an R package for studying evolutionary integration among several traits on phylogenetic trees. *Methods in Ecology and Evolution*, 8, 1920–1927.
- Castiglione, S., Serio, C., Mondanaro, A., Di Febbraro, M., Profico, A., Girardi, G. & Raia, P. (2019a) Simultaneous detection of macroevolutionary patterns in phenotypic means and rate of change with and within phylogenetic trees including extinct species. *PLoS one*, 14, e0210101.
- Castiglione, S., Serio, C., Tamagnini, D., Melchionna, M., Mondanaro, A., Di Febbraro, M., Profico, A., Piras, P., Barattolo, F. & Raia, P. (2019b) A new, fast method to search for morphological convergence with shape data. *PLoS One*, 14, e0226949.
- Castiglione, S., Tesone, G., Piccolo, M., Melchionna, M., Mondanaro, A., Serio, C., Di Febbraro, M. & Raia, P. (2018) A new method for testing evolutionary rate variation and shifts in phenotypic evolution. *Methods in Ecology and Evolution*, 9, 974–983.
- Churchill, M., Boessenecker, R.W. & Clementz, M.T. (2014) Colonization of the Southern Hemisphere by fur seals and sea lions (Carnivora: Otariidae) revealed by combined evidence phylogenetic and Bayesian biogeographical analysis. *Zoological Journal of the Linnean Society*, 172, 200–225.
- Churchill, M., Clementz, M.T. & Kohno, N. (2015) Cope's rule and the evolution of body size in Pinnipedimorpha (Mammalia: Carnivora). *Evolution*, 69, 201–215.
- Davies, J. (1958a) The Pinnipedia: an essay in zoogeography. *Geographical Review*, 48, 474–493.
- Davies, J. (1958b) Pleistocene geography and the distribution of northern pinnipeds. *Ecology*, 39, 97–113.
- Davies, J. (1963) The antitropical factor in cetacean speciation. *Evolution*, 17, 107–116.
- Deméré, T.A., Berta, A. & Adam, P.J. (2003) Pinnipedimorph evolutionary biogeography. *Bull. Am. Mus. Nat. Hist.*, 279, 32–76.
- Dewaele, L., Amson, E., Lambert, O. & Louwye, S. (2017) Reappraisal of the extinct seal “*Phoca*” *vitulinoides* from the Neogene of the North Sea Basin, with bearing on its geological age, phylogenetic affinities, and locomotion. *PeerJ*, 5, e3316.
- Dewaele, L., Lambert, O. & Louwye, S. (2018) A late surviving Pliocene seal from high latitudes of the North Atlantic realm: the latest monachine seal on the southern margin of the North Sea. *PeerJ*, 6, e5734.
- Dowsett, H.J. & Wiggs, L.B. (1992) Planktonic foraminiferal assemblage of the Yorktown Formation, Virginia, USA. *Micro-paleontology*, 38, 75–86.
- Dowsett, H.J., Robinson, M.M., Haywood, A.M., Hill, D.J., Dolan, A.M., Stoll, D.K., Chan, W.-L., Abe-Ouchi, A., Chandler, M.A. & Rosenbloom, N.A. (2012) Assessing confidence in Pliocene sea surface temperatures to evaluate predictive models. *Nature Climate Change*, 2, 365–371.
- Fulton, T.L. & Strobeck, C. (2010) Multiple fossil calibrations, nuclear loci and mitochondrial genomes provide new insight into biogeography and divergence timing for true seals (Phocidae, Pinnipedia). *Journal of Biogeography*, 37, 814–829.
- Fyler, C.A., Reeder, T.W., Berta, A., Antonelis, G., Aguilar, A. & Androukaki, E. (2005) Historical biogeography and phylogeny of monachine seals (Pinnipedia : Phocidae) based on mitochondrial and nuclear DNA data. *Journal of Biogeography*, 32, 1267–1279.
- Govender, R. (2015) Preliminary phylogenetics and biogeographic history of the Pliocene seal, *Homiphoca capensis* from Langebaanweg,

- South Africa. *Transactions of the Royal Society of South Africa*, 70, 25-39.
- Herbert, T.D., Lawrence, K.T., Tzanova, A., Peterson, L.C., Caballero-Gill, R. & Kelly, C.S. (2016) Late Miocene global cooling and the rise of modern ecosystems. *Nature Geoscience*, 9, 843-847.
- Holt, B., Marx, F.G., Fritz, S.A., Lessard, J.-P. & Rahbek, C. (2020) Evolutionary diversification in the marine realm: a global case study with marine mammals. *Frontiers of Biogeography*, 12, e45184.
- King, J. (1983) *Seals of the world*. Oxford Univ. Press, Oxford, U.K.
- Koretsky, I.A. & Holec, P. P. (2002) A primitive seal (Mammalia: Phocidae) from the early middle Miocene of Central Paratethys. *Smithson. Contrib. Paleobiol.*, 93, 163-178.
- Koretsky, I.A. & Rahmat, S.J. (2015) A new species of the subfamily Devinophocinae (Carnivora, Phocidae) from the Central Paratethys. *Res. Paleontol. Stratigraph.*, 121, 31-47.
- Knobe, M.L., Bush, A.M., Frishkoff, L.O., Heim, N.A. & Payne, J.L. (2020) Ecologically diverse clades dominate the oceans via extinction resistance. *Science*, 367, 1035-1038.
- Laws, R., Baird, A. & Bryden, M. (2003) Size and growth of the crabeater seal *Lobodon carcinophagus* (Mammalia: Carnivora). *Journal of Zoology*, 259, 103-108.
- Lindfors, P., Tullberg, B.S. & Biuw, M. (2002) Phylogenetic analyses of sexual selection and sexual size dimorphism in pinnipeds. *Behavioral Ecology and Sociobiology*, 52, 188-193.
- Liwanag, H.E., Berta, A., Costa, D.P., Budge, S.M. & Williams, T.M. (2012) Morphological and thermal properties of mammalian insulation: the evolutionary transition to blubber in pinnipeds. *Biological Journal of the Linnean Society*, 107, 774-787.
- Mason, M.J., Wenger, L.M., Hammer, Ø. & Blix, A.S. (2020) Structure and function of respiratory turbinates in phocid seals. *Polar Biology*, 43, 157-173.
- Modig, A. (1996) Effects of body size and harem size on male reproductive behaviour in the southern elephant seal. *Animal Behaviour*, 51, 1295-1306.
- Paradis, E., Claude, J. & Strimmer, K. (2004) APE: analyses of phylogenetics and evolution in R language. *Bioinformatics*, 20, 289-290.
- Pennell, M.W., Eastman, J.M., Slater, G.J., Brown, J.W., Uyeda, J.C., FitzJohn, R.G., Alfaro, M.E. & Harmon, L.J. (2014) geiger v2. 0: an expanded suite of methods for fitting macroevolutionary models to phylogenetic trees. *Bioinformatics*, 30, 2216-2218.
- Pimiento, C., Griffin, J.N., Clements, C.F., Silvestro, D., Varela, S., Uhen, M.D. & Jaramillo, C. (2017) The Pliocene marine megafauna extinction and its impact on functional diversity. *Nature ecology & evolution*, 1, 1100-1106.
- Pinheiro, J., Bates, D., DebRoy, S., Sarkar, D., Heisterkamp, S., Van Willigen, B. & Maintainer, R. (2017) 'nlme': linear and nonlinear mixed effects models, version 3. R package.
- Quental, T.B. & Marshall, C.R. (2010) Diversity dynamics: molecular phylogenies need the fossil record. *Trends in ecology & evolution*, 25, 434-441.
- Ray, C.E. (1976) Geography of phocid evolution. *Systematic Biology*, 25, 391-406.
- Repenning, C.A., Ray, C.E. & Grigorescu, D. (1979) Pinniped biogeography. *Historical biogeography, plate tectonics, and the changing environment*, 357-369.
- Revell, L.J. (2012) phytools: an R package for phylogenetic comparative biology (and other things). *Methods in ecology and evolution*, 3, 217-223.
- Rogers, T.L. (2009) Leopard seal: *Hydrurga leptonyx*. Pp. 673-674 in W. F. Perrin, B. Würsig & J. G. M. Thewissen, eds. *Encyclopedia of marine mammals*. Elsevier, Amsterdam, The Netherlands.
- Rule, J.P., Hocking, D.P. & Fitzgerald, E.M.G. (2019) Pliocene monachine seal (Pinnipedia: Phocidae) from Australia constrains timing of pinniped turnover in the Southern Hemisphere. *Journal of Vertebrate Paleontology*, 39, e1734015.
- Rule, J.P., Adams, J.W., Marx, F.G., Evans, A.R., Tennyson, A.J., Scofield, R.P. & Fitzgerald, E.M. (2020a) First monk seal from the Southern Hemisphere rewrites the evolutionary history of true seals. *Proceedings of the Royal Society B*, 287, 20202318.
- Rule, J.P., Adams, J.W., Rovinsky, D.S., Hocking, D.P., Evans, A.R. & Fitzgerald, E.M. (2020b) A new large-bodied Pliocene seal with unusual cutting teeth. *Royal Society Open Science*, 7, 201591.
- Rule, J.P., Adams, J.W. & Fitzgerald, E.M. (2021a) Colonization of the ancient southern oceans by small-sized Phocidae: new evidence from Australia. *Zoological Journal of the Linnean Society*, 191, 1160-1180.
- Rule, J.P., Adams, J.W., Marx, F.G., Evans, A.R., Tennyson, A.J., Scofield, R.P. & Fitzgerald, E.M. (2021b) Correction to: first monk seal from the Southern Hemisphere rewrites the evolutionary history of true seals. *Proceedings of the Royal Society B*, 288, 20211858.
- Samaranch, R. & Gonzalez, L. (2000) Changes in morphology with age in Mediterranean monk seals (*Monachus monachus*). *Marine mammal science*, 16, 141-157.
- Sepúlveda, M., Oliva, D., Duran, L.R., Urrea, A., Pedraza, S.N., Majluf, P., Goodall, N. & Crespo, E.A. (2013) Testing Bergmann's rule and the Rosenzweig hypothesis with craniometric studies of the South American sea lion. *Oecologia*, 171, 809-817.
- Stirling, I. (1971) *Leptonychotes weddellii*. *Mammalian Species*, 19, 1-5.
- Torres-Romero, E.J., Morales-Castilla, I. & Olalla-Tárraga, M.Á. (2016) Bergmann's rule in the oceans? Temperature strongly correlates with global interspecific patterns of body size in marine mammals. *Global Ecology and Biogeography*, 25, 1206-1215.
- Valenzuela-Toro, A.M., Gutstein, C.S., Varas-Malca, R.M., Suarez, M.E. & Pyenson, N.D. (2013) Pinniped turnover in the South Pacific Ocean: new evidence from the Plio-Pleistocene of the Atacama Desert. *Chile. Journal of Vertebrate Paleontology*, 33, 216-223.
- Valenzuela-Toro, A.M., Pyenson, N.D., Gutstein, C.S. & Suarez, M.E. (2016) A new dwarf seal from the late Neogene of South America and the evolution of pinnipeds in the Southern Hemisphere. *Pap Palaeontol.*, 2, 101-115.
- Velez-Juarbe, J. & Valenzuela-Toro, A.M. (2019) Oldest record of monk seals from the North Pacific and biogeographic implications. *Biology letters*, 15, 20190108.
- Warne, M.T. (2005) The global Mio-Pliocene climatic equability and coastal ostracod faunas of southeast Australia. *Palaeogeography, palaeoclimatology, palaeoecology*, 225, 248-265.
- Wyss, A.R. (1994) The evolution of body size in phocids: some ontogenetic and phylogenetic observations. Pp. 69-75 in A. Berta & T. A. Deméré, eds. *Contributions in marine mammal paleontology honoring Frank C. Whitmore Jr.* Forgotten Books, Lond.
- Yonezawa, T., Kohno, N. & Hasegawa, M. (2009) The monophyletic origin of sea lions and fur seals (Carnivora; Otariidae) in the Southern Hemisphere. *Gene*, 441, 89-99.

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## Appendix

## Body Size Estimations

**Table A1.** Equations used for total body length estimations of extinct taxa.

Acronym	Definition	Equation	Reference
BZB	Bizygomatic width	$1.03 \times \text{Log}(\#) + 1.10$	Churchill et al. 2014
CW	Width across canines	$0.72 \times \text{Log}(\#) + 1.83$	Churchill et al. 2014
HOC	Height of occipital shield	$1.30 \times \text{Log}(\#) + 1.28$	Churchill et al. 2014
LB	Length of tympanic bulla	$0.75 \times \text{Log}(\#) + 1.91$	Churchill et al. 2014
LUPC	Length of upper postcanine toothrow	$0.96 \times \text{Log}(\#) + 1.64$	Churchill et al. 2014
LUTR	Length of upper toothrow	$1.00 \times \text{Log}(\#) + 1.50$	Churchill et al. 2014
OCB	Width across occipital condyles	$1.54 \times \text{Log}(\#) + 1.02$	Churchill et al. 2014
PL	Palate length	$0.96 \times \text{Log}(\#) + 1.32$	Churchill et al. 2014
WB	Width of tympanic bulla	$1.08 \times \text{Log}(\#) + 1.58$	Churchill et al. 2014
<i>P. sibirica</i> % estimate	Humerus 8.12% total body length in <i>Pusa sibirica</i>	$\# / 0.0812$	Dewaele et al. 2017
<i>P. vitulina</i> % estimate	Humerus 7.76% total body length in <i>Phoca vitulina</i>	$\# / 0.0776$	Dewaele et al. 2017
<i>O. rossii</i> % estimate	Humerus 5.95% total body length in <i>Ommatophoca rossii</i>	$\# / 0.0595$	Dewaele et al. 2017
<i>L. weddellii</i> % estimate	Humerus 6.5% total body length in <i>Leptonychotes weddellii</i>	$\# / 0.065$	Dewaele et al. 2017

Table A2. Estimations for total body length of extinct taxa.

Taxa	Specimen	Equation Used	Measurement (mm)	Value from Equation	Estimate (mm)	Average Estimate (mm)	Notes
<i>Noriphoca gaudini</i>	MSNUN 123	BZB	184.00	3.43	2708.71	2288.15	Measurement data taken from Dewaele et al. (2018)
		LUPC	83.00	3.48	3036.09		
		CW	61.00	3.12	1304.48		
<i>Devinophoca emyri</i>	USNM 553684	PL	122.00	3.32	2103.32		Measurement data taken from Koretsky and Rahmat (2015)
		BZB	127.70	3.27	1859.42	1638.92	
		WB	35.70	3.26	1806.68		
<i>Devinophoca claytoni</i>	Z14523	LUPC	46.90	3.24	1755.20		Measurement data taken from Koretsky and Holec (2002)
		LB	33.60	3.05	1134.37		
		BZB	124.00	3.26	1803.95	1682.74	
		WB	49.30	3.41	2560.20		
Motunau	CM Zfa333	LUPC	49.00	3.26	1830.58		Total body length estimates for taken from Dewaele et al. (2017). Due to <i>N. vitulinoides</i> being a phocine, only phocine estimates were used.
		CW	40.00	2.98	962.68		
		LB	38.50	3.10	1256.31		
		LB	37.74	3.09	1237.66	1637.47	
		WB	39.90	3.31	2037.28		
<i>Leptophoca proxima</i>	CMM-V-2021	All Subsets	CW(33.05), OCB(51.43)	3.32	2094.43	2094.43	
<i>Nanophoca vitulinoides</i>	IRSNB M2276c, IRSNB 1063-M242, IRSNB M2278, IRSNB M2271, IRSNB M2276d, IRSNB 1105-M239	<i>P. sibirica</i> % estimate	–	–	920.00	982.50	Total body length estimates for taken from Dewaele et al. (2017). Due to <i>N. vitulinoides</i> being a phocine, only phocine estimates were used.
		–	–	–	–	–	
		–	–	–	–	–	
		–	–	–	–	–	
		–	–	–	–	–	
		–	–	–	–	–	
<i>Monotherium wynani</i>	USNM 214909	<i>P. vitulina</i> % estimate	–	–	1045.00		USNM 214909 is a cast of MCZ 8741
		WB	41.40	3.33	2120.12	1638.60	

(Continued)



Table A2. (Continued).

Taxa	Specimen	Equation Used	Measurement (mm)	Value from Equation	Estimate (mm)	Average Estimate (mm)	Notes
<i>Hadrokirus marini</i>	MNHN.F.SAS 1627	LB	34.50	3.06	1157.08		
		All Subsets	CW(52.70), OCB(56.90)	3.43	2698.73	2698.73	Measurement data taken from Amson and Muizon (2014)
<i>Sarcodectes magnus</i>	USNM 475486	LUTR	109.76	3.54	3470.92	2831.29	
		LUPC	91.32	3.52	3327.69		
	USNM 181601	WB	53.91	3.45	2819.70		
		LB	51.42	3.19	1560.81		
	USNM 534034	<i>O. rossii</i> hum/body length	180.44	–	3032.61		
		<i>L. weddellii</i> hum/body length		–	2776.00		
<i>Pliophoca etrusca</i>	MSNUP I-13993	CW	44.00	3.01	1031.06	1684.99	Measurement data taken from Berta et al. (2015)
		<i>O. rossii</i> hum/body length	125.00	–	2100.84		
		<i>L. weddellii</i> hum/body length		–	1923.08		
Beaumaris	NMV P160399	LB	35.66	3.07	1186.14	1580.86	
		WB	38.78	3.30	1975.59		
<i>Eomonachus belegaerensis</i>	NMNZ S.046692	BZB	131.12	3.28	1910.73	2467.96	
		WB	39.77	3.31	2030.11		
		LB	31.24	3.03	1074.07		
		OCB	51.05	3.65	4470.06		
		HOC	66.02	3.65	4421.59		
	NMNZ S.047276	LB	35.85	3.08	1190.88		
		WB	38.58	3.29	1964.59		
	NMNZ S.047422	OCB	52.07	3.66	4608.34		
		LB	32.67	3.05	1110.74		
		WB	38.18	3.29	1942.60		
		LUPC	63.05	3.37	2331.83		
	CM 2020.74.1	LUTR	80.20	3.40	2536.15		
		LUPC	67.56	3.40	2491.74		

**Table A3.** Average of estimates of total body length for extinct taxa taken from the literature, or calculated in this study.

Taxon	Body Length (m) Averaged Estimates	Source and Method
<i>Noriphoca gaudini</i>	2.29	Churchill et al. (2015) (CW, PL, BZB, LUPC)
<i>Devinophoca emyri</i>	1.64	Churchill et al. (2015) (BZB, WB, LUPC, LB)
<i>Devinophoca claytoni</i>	1.68	Churchill et al. (2015) (BZB, WB, LUPC, CW, LB)
†		
<i>Kawas benegasorum</i> †	1.63	Rule et al. (2020a)
CM ZFa333 Motunau †	1.64	Churchill et al. (2015) (LB, WB)
<i>Leptophoca proxima</i> †	2.09	Churchill et al. (2015) (All subsets)
<i>Nanophoca vitulinoides</i> †	0.98	Dewaele et al. (2017) (Modified for this study using Phocinae humeri percentages only)
<i>Monotherium wymani</i>	1.64	Churchill et al. (2015) (WB, LB)
†		
<i>Homiphoca capensis</i> †	1.80	Churchill et al. (2015)
<i>Piscophoca pacifica</i> †	1.96	Churchill et al. (2015)
<i>Acrophoca longirostris</i>	1.91	Churchill et al. (2015)
†		
<i>Hadrokirus martini</i> †	2.70	Churchill et al. (2015) (All subsets)
<i>Australophoca changorum</i> †	0.68	Valenzuela-Toro et al. (2016)
<i>Sarcodectes magnus</i> †	2.83	Churchill et al. (2015) and Rule et al. (2020b) (LUTR, LUPC, WB, LB) and Dewaele et al. (2017) (Monachinae humeri percentages only)
<i>Pliophoca etrusca</i> †	1.69	Churchill et al. (2015) (CW) and Dewaele et al. (2017) (Monachinae humeri percentages only)
NMV P160399 Beaumaris †	1.58	Churchill et al. (2015) (WB, LB)
<i>Eomonachus belegaerensis</i> †	2.47	Churchill et al. (2015) (BZB, WB, LB, OCB, HOC, LUPC, LUTR)

**Table A4.** Total body length of extant taxa taken from the literature.

Taxon	Maximum Body Length (m)	Maximum Male Body Length (m)	Maximum Female Body Length (m)	Reference
<i>Erigonathus barbatus</i>	2.54	2.54	2.42	Andersen et al. (1999)
<i>Cystophora cristata</i>	2.6	2.6	2.06	Bininda-Emonds and Gittleman (2000)
<i>Histiophoca fasciata</i>	1.55	1.53	1.55	Bininda-Emonds and Gittleman (2000)
<i>Pagophilus groenlandicus</i>	1.76	1.76	1.69	Bininda-Emonds and Gittleman (2000)
<i>Phoca largha</i>	1.69	1.69	1.59	Bininda-Emonds and Gittleman (2000)
<i>Phoca vitulina</i>	1.80	1.8	1.65	Lindenfors et al. (2002)
<i>Halichoerus grypus</i>	2.16	2.16	1.8	Bininda-Emonds and Gittleman (2000)
<i>Pusa capsica</i>	1.50	1.5	1.36	Bininda-Emonds and Gittleman (2000)
<i>Pusa sibirica</i>	1.30	1.3	1.25	Bininda-Emonds and Gittleman (2000)
<i>Pusa hispida</i>	1.5	1.5	1.5	King (1983)
<i>Monachus monachus</i>	2.7	2.7	2.65	Samaranch and Gonzalez (2000) (male), Bininda-Emonds and Gittleman (2000) (female)
<i>Neomonachus schauinslandi</i>	2.34	2.14	2.43	Bininda-Emonds and Gittleman (2000)
<i>Neomonachus tropicalis</i>	2.33	2.33	2.26	Bininda-Emonds and Gittleman (2000) (male)
<i>Mirounga leonina</i>	5.56	5.56	2.70	Modig (1996) (male), Bininda-Emonds and Gittleman (2000) (female)
<i>Mirounga angustirostris</i>	4.50	4.50	2.95	Bininda-Emonds and Gittleman (2000)
<i>Lobodon carcinophagus</i>	2.77	2.64	2.77	Laws et al. (2003)
<i>Ommatophoca rossii</i>	3	3.00	2.50	King (1983)
<i>Hydrurga leptonyx</i>	3.8	3.3	3.8	Rogers (2009)
<i>Leptonychotes weddellii</i>	3.29	2.97	3.29	King (1983), Stirling (1971) (female)

## Sea Surface Temperature data

**Table A5.** Sea Surface Temperature data for extant taxa from NOAA Earth System Research Laboratory database.

Taxon	Winter SST	Summer SST	Median SST
<i>Erignathus barbatus</i>	0–2	8–10	5
<i>Cystophora cristata</i>	0–2	8–10	5
<i>Histiophoca fasciata</i>	0–2	12–14	7
<i>Pagophilus groenlandicus</i>	0–2	12–14	7
<i>Phoca largha</i>	0–2	26–28	14
<i>Phoca vitulina</i>	0–2	22–24	12
<i>Halichoerus grypus</i>	2–4	18–20	10
<i>Pusa caspica</i>	0–2	24–26	13
<i>Pusa sibirica</i>	0–2	15	7.5
<i>Pusa hispida</i>	0–2	12–14	7
<i>Monachus monachus</i>	14–16	26–28	21
<i>Neomonachus schauinslandi</i>	22–24	28–29	25.5
<i>Neomonachus tropicalis</i>	24–26	29–30	27
<i>Mirounga leonina</i>	0–2	10–16	8
<i>Mirounga angustirostris</i>	4–6	18–20	12
<i>Lobodon carcinophagus</i>	0–2	6–8	4
<i>Ommatophoca rossii</i>	0–2	6–8	4
<i>Hydrurga leptonyx</i>	0–2	6–8	4
<i>Leptonychotes weddellii</i>	0–2	6–8	4



**Table A6.** Sea surface temperature (SST) estimate from the area and locality of fossil taxa.

Taxon	SST	Data	Reference
<i>Monotherium wymani</i>	14.7–24 (Median 19.35)	Skeletal Oxygen Isotope	Barrick et al. (1993)
<i>Leptophoca proxima</i>	14.7–24 (Median 19.35)	Skeletal Oxygen Isotope	Barrick et al. (1993)
<i>Australophoca changorum</i>	15.2	Skeletal Oxygen Isotope	Amiot et al. (2008)
<i>Hadrokirus martini</i>	14.8	Skeletal Oxygen Isotope	Amiot et al. (2008)
<i>Acrophoca longirostris</i>	14.8	Skeletal Oxygen Isotope	Amiot et al. (2008)
<i>Piscophoca pacifica</i>	14.8	Skeletal Oxygen Isotope	Amiot et al. (2008)
<i>Sarcodectes magnus</i>	21.5	Biostratigraphy	Dowsett and Wiggs (1992), Dowsett et al. (2012)
Beumaris	15–20 (Median 17.5)	Biostratigraphy	Warne 2005
<i>Eomonachus belegaerensis</i>	17.6	Biostratigraphy	Dowsett et al. (2012)
Motunau	17.68	Isotope (oxygen and carbon)	Herbert et al. (2016)
Waipunga	19.06	Isotope (oxygen and carbon)	Herbert et al. (2016)
<i>Pliophoca etrusca</i>	25.76	Isotope (oxygen and carbon)	Herbert et al. (2016)
<i>Homiphoca capensis</i>	20.79	Isotope (oxygen and carbon)	Herbert et al. (2016)

## RRphylo Results

**Table A7.** OverfitRR analysis of search.shift results, with 100 ancestral state estimation regression simulations. For each clade, values for p.shift+ and p.shift- are the percentage of simulations that obtained statistically significant ( $P$ -value  $< 0.05$ ) positive or negative evolutionary rate shifts. "All clades" reports results assuming all nodes evolved under a single rate.

Phylogeny	Clade	p.shift+	p.shift-
Extant + extinct maximum TBL	All clades	0.4	0
	43	0.4	0.1
	44	0.4	0.1
	45	0.4	0.1
	46	0.4	0.1
	47	0.4	0
	49	0.4	0
	57	0.4	0
	61	0.4	0
	62	0.4	0
Minus ancestor maximum TBL	All clades	0.13	0
	35	0.77	0
	44	0	0.92
Extant only minimum TBL	All clades	0.04	0
	27	0.61	0
	30	0	0.23
Extant only maximum TBL	27	0.85	0
Extant + extinct median SST	All clades	0.89	0
	52	0.97	0
	43	0.18	0.01
Minus ancestors median SST	All clades	0.57	0
	35	0.1	0.01
	39	0	0
	42	1	0
Extant only minimum SST	All clades	0.01	0.12
	22	0	0.92
	31	0.76	0
Extant only median SST	All clades	0.74	0
	28	0.53	0.01
	31	0.6	0
Extant only maximum SST	All clades	0.12	0.02
	28	0.62	0
	35	0	0.69

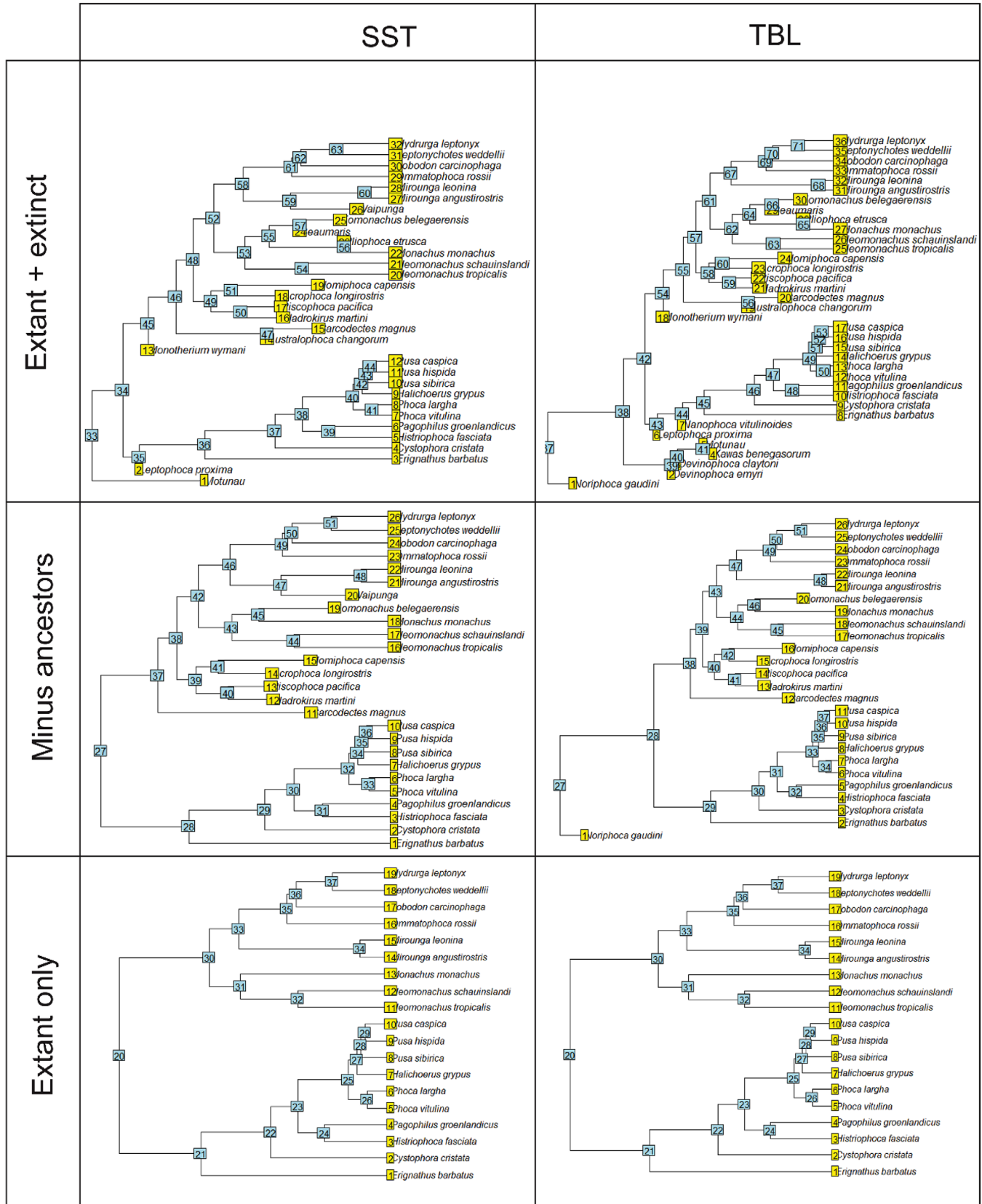


Figure A1. Clade numbers for overfitRR analysis of all datasets in Table A7.

## Regression Results

**Table A8.** Linear regression of log total body length and sea surface temperature tolerance for Phocidae. DF = degrees of freedom.

	Extant + Extinct		Extant Only				
	TBL ~ SST	Lrg TBL ~ Max SST	Lrg TBL ~ Med SST	Lrg TBL ~ Min SST	Sml TBL ~ Max SST	Sml TBL ~ Med SST	Sml TBL ~ Min SST
Min	-0.49	-0.27	-0.27	-0.26	-0.23	-0.22	-0.21
Q1	-0.10	-0.12	-0.14	-0.13	-0.09	-0.11	-0.10
Median	-0.02	-0.001	0.03	-0.03	0.33	0.03	-0.03
Q3	0.11	0.08	0.09	0.09	0.10	0.11	0.10
Max	0.39	0.36	0.36	0.38	0.17	0.20	0.21
Intercept	2.41	2.47	2.4	2.37	2.40	2.33	2.31
Slope	-0.01	-0.006	-0.003	0.002	-0.005	-0.001	0.003
Standard Error	0.01	0.005	0.006	0.006	0.004	0.004	0.004
<i>T</i> -value	-1.24	-1.13	-0.47	0.27	-1.29	-0.27	0.77
<i>P</i> -value	0.23	0.28	0.65	0.79	0.22	0.79	0.45
Residual standard error	0.17, 29 DF	0.17, 17 DF	0.18, 17 DF	0.18, 17 DF	0.13, 17 DF	0.13, 17 DF	0.13
Multiple <i>R</i> <sup>2</sup>	0.05	0.07	0.01	0.004	0.09	0.004	0.03
Adjusted <i>R</i> <sup>2</sup>	0.02	0.02	-0.05	-0.05	0.04	-0.05	-0.02
<i>F</i> -statistic	1.53, 1, and 29 DF	1.27, 1, and 17 DF	0.22, 1, and 17 DF	0.07, 1, and 17 DF	1.65, 1, and 17 DF	0.07, 1, and 17 DF	0.59, 1, and 17 DF
<i>P</i> -value	0.23	0.28	0.65	0.79	0.22	0.79	0.45



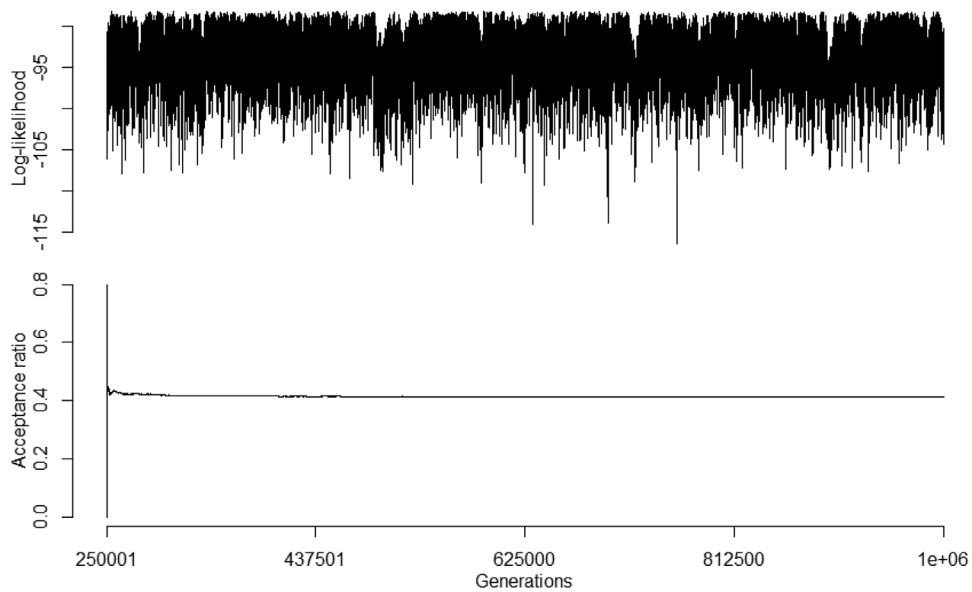
**Table A9.** Phylogenetic generalised least squares regression of log total body length and sea surface temperature tolerance for Phocidae.

	Extant + Extinct	Extant Only					
	TBL ~ SST	Lrg TBL ~ Max SST	Lrg TBL ~ Med SST	Lrg TBL ~ Min SST	Sml TBL ~ Max SST	Sml TBL ~ Med SST	Sml TBL ~ Min SST
AIC	-22.43	-25.97	-26.54	-26.71	-40.04	-40.16	-39.94
BIC	-18.13	-23.14	-23.71	-23.88	-37.21	-37.33	-37.10
Log likelihood	14.21	15.99	16.27	16.36	23.02	23.08	22.97
Min	-2.24	-1.88	-1.98	-2.06	-2.34	-2.39	-2.36
Q1	-0.63	-1.06	-1.12	-1.28	-1.22	-1.28	-1.38
Median	-0.22	-0.17	-0.13	-0.14	-0.10	0.02	-0.09
Q3	0.20	0.28	0.31	0.34	0.62	0.59	0.62
Max	1.46	2.08	2.05	1.97	1.42	1.39	1.46
Intercept	2.41	2.43	2.46	2.44	2.37	2.37	2.36
Slope	-0.002	-0.001	-0.004	-0.005	-0.001	-0.002	-0.001
Standard error	0.006	0.004	0.005	0.005	0.003	0.004	0.004
<i>T</i> -value	-0.32	-0.32	-0.79	-0.88	-0.37	-0.50	-0.21
<i>P</i> -value	0.75	0.75	0.44	0.39	0.71	0.63	0.84
Residual standard error	0.24	0.16	0.16	0.16	0.11	0.11	0.11
Degrees of freedom	31 total, 29 residual	19 total, 17 residual	19 total, 17 residual	19 total, 17 residual	19 total, 17 residual	19 total, 17 residual	19 total, 17 residual

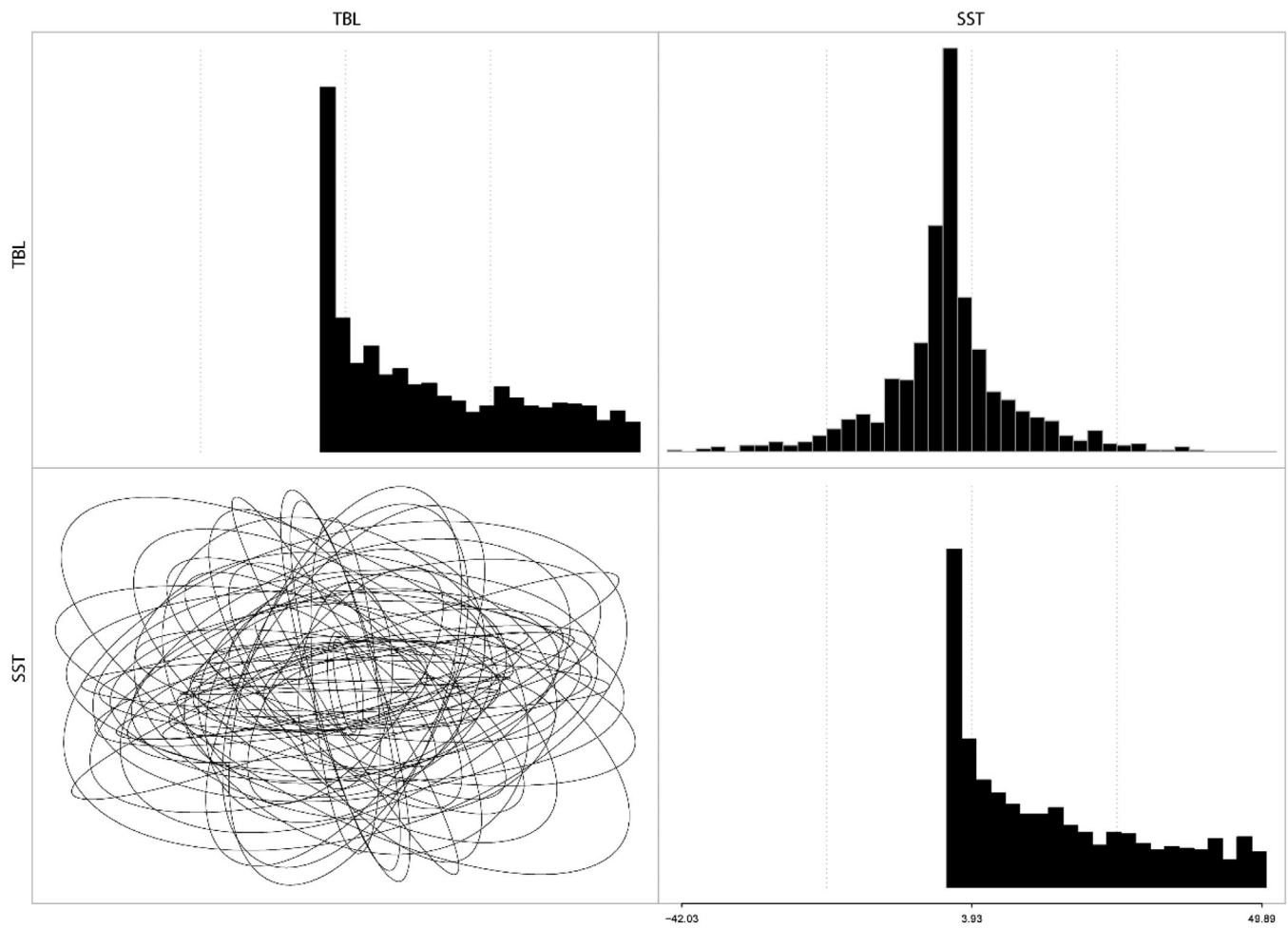
## Ratematrix Results

**Table A10.** Evolutionary rate matrix for extant + extinct taxa using the package ratematrix. TBL = total body length, SST = sea surface temperature.

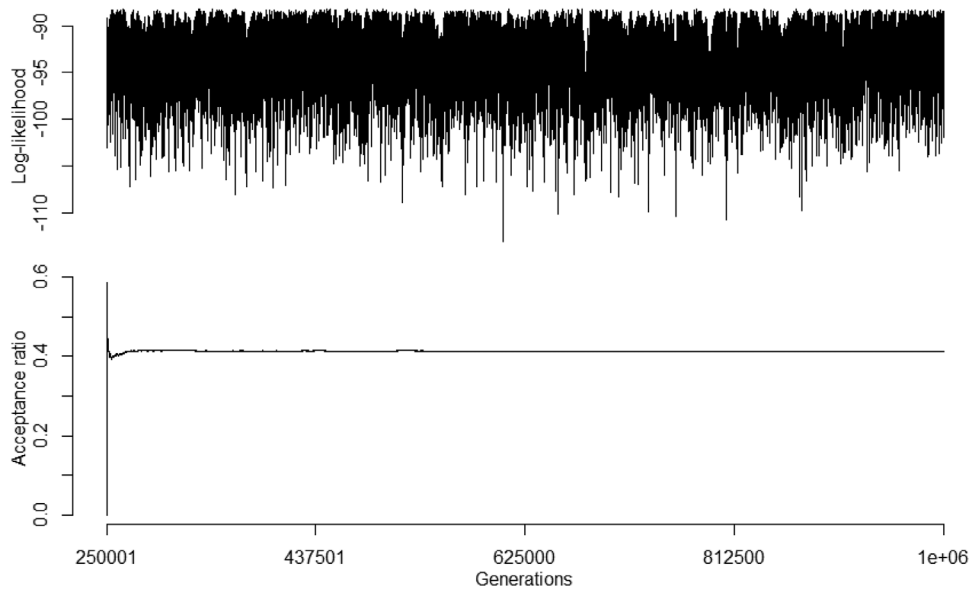
	TBL	SST
TBL	0.008157	0.019207
SST	0.019207	4.289097



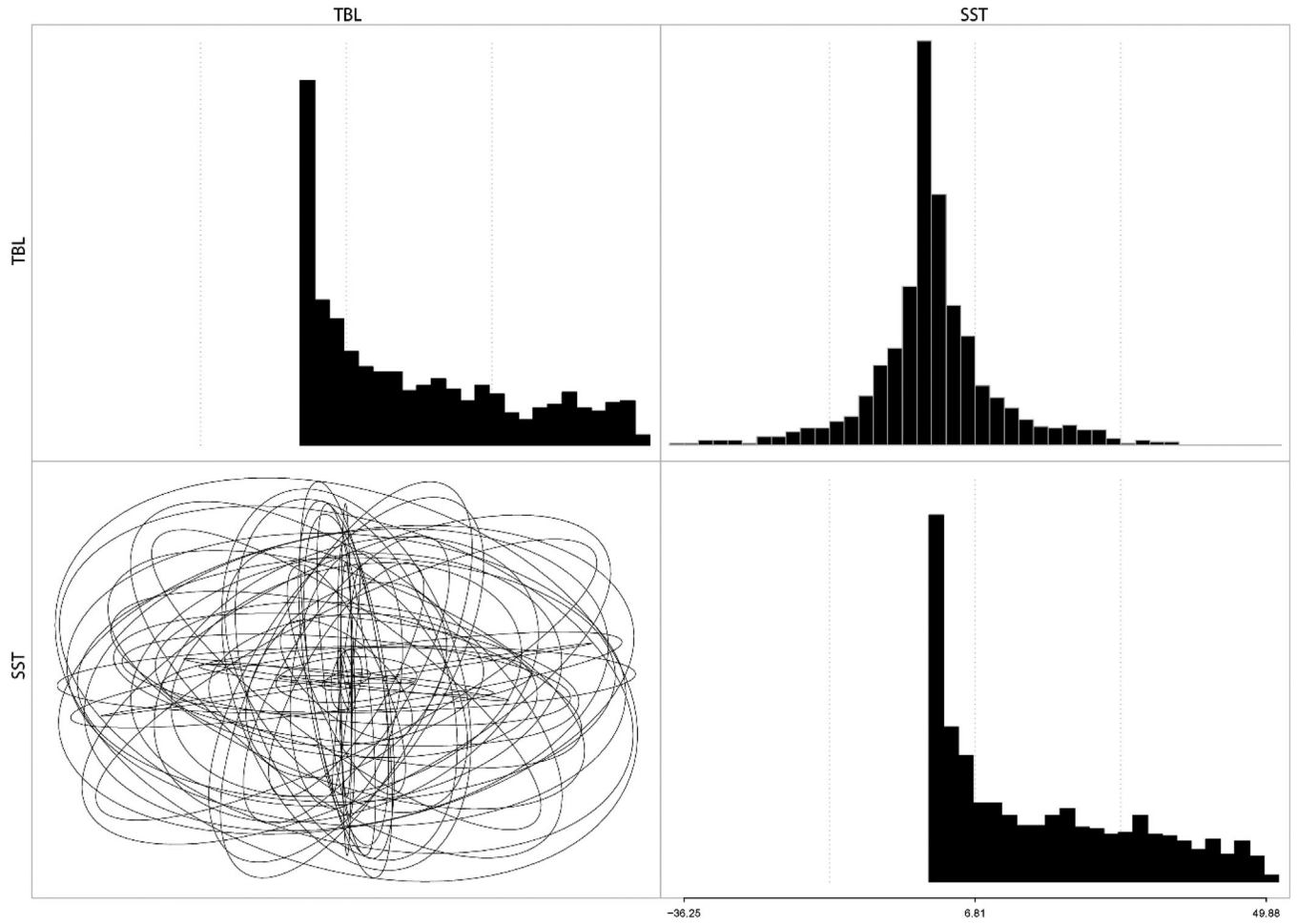
**Figure A2.** Log-likelihood trace plot and acceptance ratio of first evolutionary rate matrix MCMC chain. MCMC chain ran for 1 million generations, with the first 25% discarded as burnin and sampling every 1000 generations. Acceptance ratio for the MCMC chain was  $\sim 0.41$  (correlation = 0.62; standard deviation = 0.15; root = 0.93).



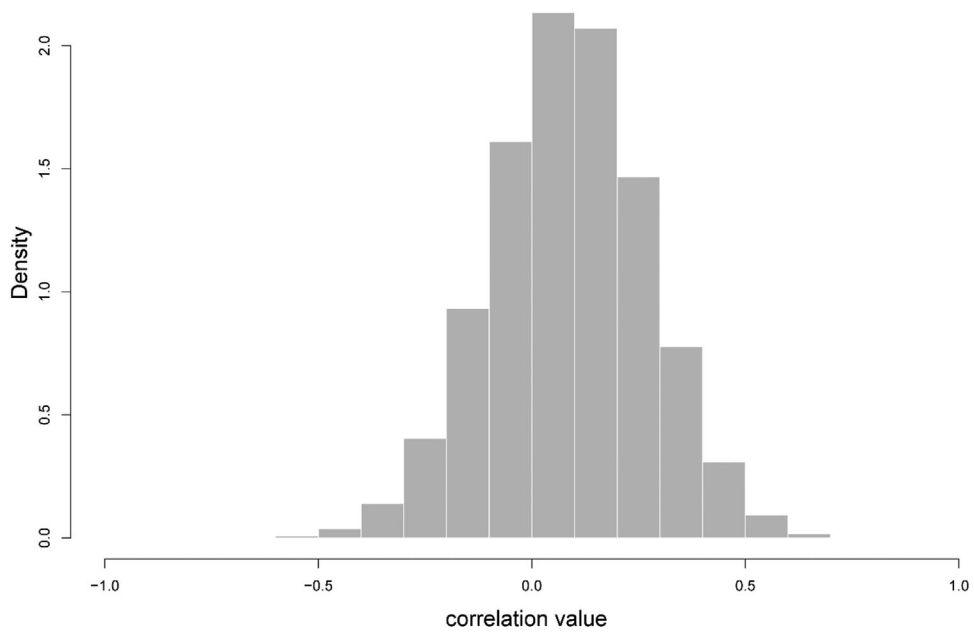
**Figure A3.** The prior distribution of the first evolutionary rate matrix MCMC chain.



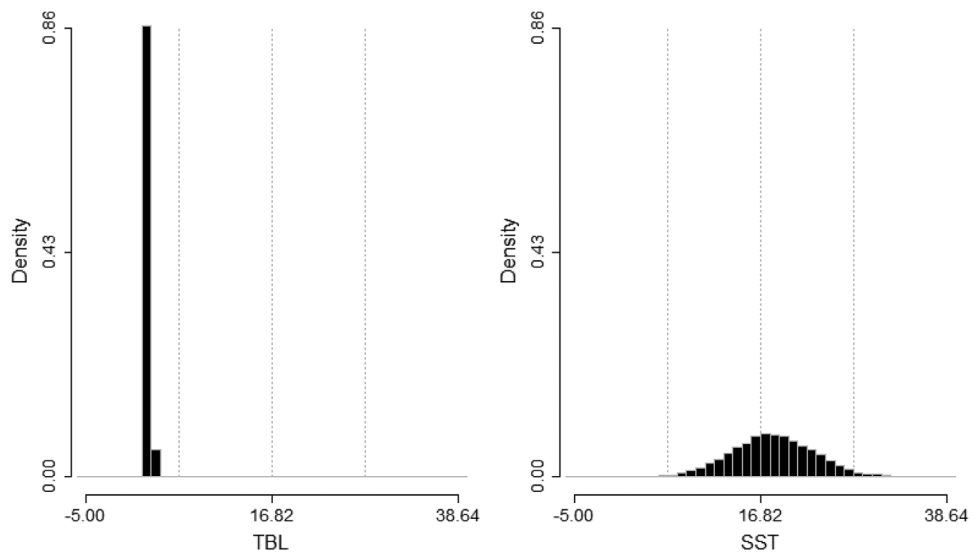
**Figure A4.** Log-likelihood trace plot and acceptance ratio of the second evolutionary rate matrix MCMC chain. MCMC chain ran for 1 million generations, with the first 25% discarded as burnin and sampling every 1000 generations. Acceptance ratio for the MCMC chain was  $\sim 0.41$  (correlation = 0.62; standard deviation = 0.15; root = 0.93).



**Figure A5.** The prior distribution of the second evolutionary rate matrix MCMC chain.



**Figure A6.** Histogram of the posterior distribution of evolutionary correlation among log total body length and sea surface temperature, extracted from the two merged MCMC chains. Minimum =  $-0.68$ ; 1st quartile =  $-0.03$ ; Median =  $0.09$ ; Mean =  $0.09$ ; 3rd quartile =  $0.21$ ; Maximum =  $0.79$ .



**Figure A7.** Posterior distribution of root values for log total body length (TBL) and sea surface temperature (SST) sampled from the merged MCMC chains.



**Table A11.** Gleman's *R* convergence check between the two ratematrix MCMC chains, with potential scale reduction factors for the root values and evolutionary rate matrices, and effective sample size.

	Point Estimate	Upper Confidence Interval	Effective Sample Size
TBL root	1.00	1.00	251.54
SST root	1.01	1.03	302.44
Matrix TBL-TBL	1.00	1.00	57,230.45
Matrix TBL-SST	1.00	1.00	44,030.33
Matrix SST-TBL	1.00	1.00	44,030.33
Matrix SST-SST	1.00	1.00	61,000.78

### *Supporting Information*

Additional supporting information may be found online in the Supporting Information section at the end of the article.

Supporting Information