



Improved cardiovascular health by supplementation with selenium and coenzyme Q10: applying structural equation modelling (SEM) to clinical outcomes and biomarkers to explore underlying mechanisms in a prospective randomized double-blind placebo-controlled intervention project in Sweden

Urban Alehagen¹ · Peter Johansson² · Erland Svensson³ · Jan Aaseth^{4,5} · Jan Alexander⁶

Received: 4 October 2021 / Accepted: 17 March 2022 / Published online: 6 April 2022
© The Author(s) 2022

Abstract

Purpose Selenium and coenzyme Q10 have synergistic antioxidant functions. In a four-year supplemental trial in elderly Swedes with a low selenium status, we found improved cardiac function, less cardiac wall tension and reduced cardiovascular mortality up to 12 years of follow-up. Here we briefly review the main results, including those from studies on biomarkers related to cardiovascular risk that were subsequently conducted. In an effort, to explain underlying mechanisms, we conducted a structured analysis of the inter-relationship between biomarkers.

Methods Selenium yeast (200 µg/day) and coenzyme Q10 (200 mg/day), or placebo was given to 443 elderly community-living persons, for 48 months. Structural Equation Modelling (SEM) was used to investigate the statistical inter-relationships between biomarkers related to inflammation, oxidative stress, insulin-like growth factor 1, expression of microRNA, fibrosis, and endothelial dysfunction and their impact on the clinical effects. The main study was registered at Clinicaltrials.gov at 30th of September 2011, and has the identifier NCT01443780.

Results In addition to positive clinical effects, the intervention with selenium and coenzyme Q10 was also associated with favourable effects on biomarkers of cardiovascular risk. Using these results in the SEM model, we showed that the weights of the first-order factors inflammation and oxidative stress were high, together forming a second-order factor inflammation/oxidative stress influencing the factors, fibrosis ($\beta=0.74$; $p<0.001$) and myocardium ($\beta=0.65$; $p<0.001$). According to the model, the intervention impacted fibrosis and myocardium through these factors, resulting in improved cardiac function and reduced CV mortality.

Conclusion Selenium reduced inflammation and oxidative stress. According to the SEM analysis, these effects reduced fibrosis and improved myocardial function pointing to the importance of supplementation in those low on selenium and coenzyme Q10.

Keywords Selenium · Coenzyme Q10 · Elderly · Mechanisms · Mortality · SEM

✉ Urban Alehagen
urban.alehagen@liu.se

¹ Division of Cardiovascular Medicine, Department of Health, Medicine and Caring Sciences, Linköping University, 581 85 Linköping, Sweden

² Department of Health, Medicine and Caring Sciences, Linköping University, 601 74 Norrköping, Sweden

³ Swedish Defence Research Agency, Stockholm, Sweden

⁴ Research Department, Innlandet Hospital Trust, 2381 Brumunddal, Norway

⁵ Faculty of Health and Social Sciences, Inland Norway University of Applied Sciences, 2418 Elverum, Norway

⁶ Norwegian Institute of Public Health, 0403 Oslo, Norway

Abbreviations

ACEI	ACE inhibitors
ARB	Angiotensin receptor blockers
CV	Cardiovascular
EF	Ejection fraction
ECG	Electrocardiogram
Hs-CRP	High-sensitivity analysis of C-reactive protein
IHD	Ischaemic heart disease
NYHA class	New York heart association functional class
PAI-1	Plasminogen activator inhibitor 1
SD	Standard deviation
SEM	Structural equation modelling
vWf	Von Willebrand factor

Background

Selenium and coenzyme Q10 were supplemented in a four-year intervention study, to an elderly population ($n = 443$) with a low selenium status and because of old age, a low level of coenzyme Q10. A significantly reduced cardiovascular mortality was observed after five, 10 and 12 years [1–3]. Here we present an overview of the intervention trial including clinical outcomes and secondary analyses conducted mainly on different biomarkers. Second, we present a model for the interconnection between biomarkers and clinical outcome obtained by means of Structural Equation Modelling (SEM) analyses. With the application of SEM analysis, which is a combination of factor and multiple regression analyses, it is possible from observed or manifest variables to find underlying or latent factors explaining the relations between the observed variables, as well as to estimate the effects of the latent variables on dependant variables. Latent variables or factors provide important information, e.g. on pathophysiological processes, which cannot be fully described by single biomarkers alone.

Previously, in an attempt to explore possible underlying mechanisms, we have undertaken a series of secondary analyses, examined variables, such as changes in biomarkers for myocardial wall tension, inflammation, oxidative stress and fibrosis, as well as impact of baseline selenium status that all could be related to the outcome presented in the main study. A limitation, however, is that these studies by themselves do not present a broader perspective of how supplementation with selenium and coenzyme Q10 can be associated to changes in basic physiological mechanisms and how alterations in these mechanisms can be associated with each other. In the present paper, we first briefly summarize our previous results and then conduct a structural equation modelling analysis leading to an overall description of possible mechanisms and observed clinical effects.

Selenium and coenzyme Q10 in humans

Selenium is a trace element that is essential for a normal cellular function, and several of the selenoproteins play crucial roles in the cellular red/ox regulation and the protection against oxidative stress. To obtain an optimal expression of one of the important selenoproteins in plasma; selenoproteins P, which transports selenium from the liver to peripheral tissues, a daily intake of 100–150 $\mu\text{g/day}$ of selenium is required [4]. Of specific interest among the 25 selenoproteins is that the thioredoxin reductases (TXRND), a group of selenoproteins that belongs to the pyridine nucleotide–disulfide oxidoreductase family, which is central to the cellular redox regulation, appear to require at least as high serum selenium levels as other human selenoenzymes to be optimized (i.e. fully expressed), apparently about 120 $\mu\text{g/L}$ when measured in erythrocytes [5].

However, the content of selenium soil varies in different parts of the world, influencing the daily intake. In Europe, the concentrations of selenium in plasma are generally low, usually well below 80–90 $\mu\text{g/L}$ [6–9], whereas in North America, the levels are generally above 120 $\mu\text{g/L}$ [10, 11]. In Finland, the average intake level of selenium in the 1970s was 25 $\mu\text{g/day}$ with a corresponding plasma selenium concentration of 50 $\mu\text{g/L}$ [12]. After the decision to add selenium to the fertilizers in 1986, the average daily intake has risen to about 80 $\mu\text{g/day}$ [12].

The French EVA study demonstrated an association between low plasma concentration of selenium and increased mortality [13]. However, in a meta-analysis including 12 different trials (including the American SELECT and NPC trials), Rees et al. could not find an association between supplementation with selenium and decreased cardiovascular mortality [14]. It is, however, important to note that both the SELECT and NPC studies included US participants with mean plasma selenium concentrations higher than in all European countries (135 $\mu\text{g/L}$, and 113 $\mu\text{g/L}$ respectively) [15], and that the vast majority of the participants in the meta-analysis reported by Rees et al. were US inhabitants from the SELECT and NPC studies.

In Sweden, there is situation similar to that which existed in Finland before selenium was added to fertilizers, with low selenium content in the soil resulting in a low daily intake as reported by Harris et al. demonstrating an intake of less than 25 $\mu\text{g/day}$ [16], and by Gao et al. who found the intake to be well below the recommended dietary intake of selenium [17]. However, as many studies were based on dietary surveys, or were performed several decades ago, we wanted to obtain new data based on analyses in blood samples from a representative elderly

community-living population. A total of 668 elderly participants were included from a rural municipality population in the Southeast of Sweden [18]. The study population showed a low mean serum selenium concentration, 67.1 µg/L. This corresponds to an average daily intake of approximately 35 µg/day, thus well below the amounts (110–150 µg/day) for an optimal expression of selenoproteins [19]. The consequence of low selenium status in terms of cardiovascular (CV) mortality was evaluated in our Swedish study population during a follow-up period of 6.85 years [20]. A significantly higher CV mortality was observed among participants in the lowest quartile (29 out of 107) compared with those in the highest quartile (16 out of 111; χ^2 : 5.35; $p = 0.02$). In a multivariate model where adjustments were made for male gender, smoking, ischaemic heart disease (IHD), diabetes, chronic obstructive pulmonary disease, and impaired systolic cardiac function ($EF < 40\%$), the increased CV mortality risk remained significant in the lowest selenium concentration quartile (HR: 1.56; 95%CI 1.03–2.36; $p = 0.04$), compared with those who had a higher selenium intake. Thus, it seems that a low daily intake of selenium has important implications for human health, which is also reported in the literature [21, 22].

However, of importance, as reported by Xia et al., coenzyme Q10 needs the presence of selenium for reduction of the oxidized form, ubiquinone, into the active form, ubiquinol [23]. Coenzyme Q10 is essential for all living cells, and it is one of the important antioxidants in lipid structures — membranes and lipoproteins of the body and of particular importance for crucial ATP-generating steps within the mitochondrial respiratory chain. In the extramitochondrial space, the selenoenzyme TXNDR1 is a main reductive enzyme restoring ubiquinol from ubiquinone [23]. The endogenous production of coenzyme Q10 decreases with increasing age, and the production in the myocardium is reduced to about half at the age of 80 years [24], why supplementation to elderly, especially for those living in areas with low selenium contents in the soil seems relevant [24, 25]. Both the synthesis of coenzyme Q10 and selenoproteins, in this case TXNDR is dependent on a functional mevalonate pathway [25].

These considerations suggest that optimal levels in the organism of *both* selenium and coenzyme Q10 are needed for good human health.

Based on the rationale outlined above, a group of elderly Swedish individuals was recruited into a prospective randomized double-blind placebo-controlled trial. The study population included consisted of 443 individuals in the age 70–88 years (Table 1). They received the selenized yeast and coenzyme Q10 (SelenoPrecise 200 µg B.I.D, Pharma NordApS, Vejle, Denmark and Bio-Quinon 100 mg B.I.D, Pharma NordApS, Vejle, Denmark) or placebo as a dietary

supplement for 48 months and were re-examined every six months [1].

Summary of effects on primary and secondary outcomes

Glycation products

Fructosamine is the glycation product of proteins, mainly albumin, in the body. The concentration of fructosamine monitors the blood–glucose level during the last two to four weeks. However, fructosamine appears to be associated with other pathogenic mechanisms in the body. Even in those without diabetes, a higher concentration of fructosamine indicates an increased CV risk [26]. Also, there is a strong association between concentration of fructosamine and biomarkers of inflammation and increases in patients with rheumatoid arthritis, but without diabetes [27]. Similarly, in the non-diabetic part of the study population, we found a strong association between the fructosamine concentration and intervention with selenium and coenzyme Q10, as a significantly decreased fructosamine concentration was observed in the active treatment group (Fig. 1) [28].

The effect on fructosamine could be seen in both genders, and both in the older, and in the younger part of the study population. This indicates that an anti-inflammatory effect i.e. of selenium also may protect against protein glycation, which has previously been discussed in the literature [29].

Insulin-like growth factor 1 (IGF-1)

Insulin-like growth factor 1 (IGF-1) has a multitude of actions, including anti-inflammatory and anti-apoptotic actions on myocardial cells [30, 31]. The concentration of IGF-1 decreases with increasing age [32]. A lower concentration of IGF-1 is also associated with increased CV risk [33, 34]. In our study, we observed higher levels of both IGF-1, and the age-adjusted IGF-1 SD score as a result of the intervention [35].

These increased IGF-1 concentrations accompanying the intervention could mediate positive intracellular effects including less inflammatory activity and probably also an anti-apoptotic effect.

Inflammation

Several selenoproteins have important functions in the inflammatory response accompanying various diseases [36]. To evaluate the effects of supplementation with selenium and coenzyme Q10 on inflammation, we used sP-selectin and CRP using high sensitivity assays. We could report a highly significant difference after 48 months of

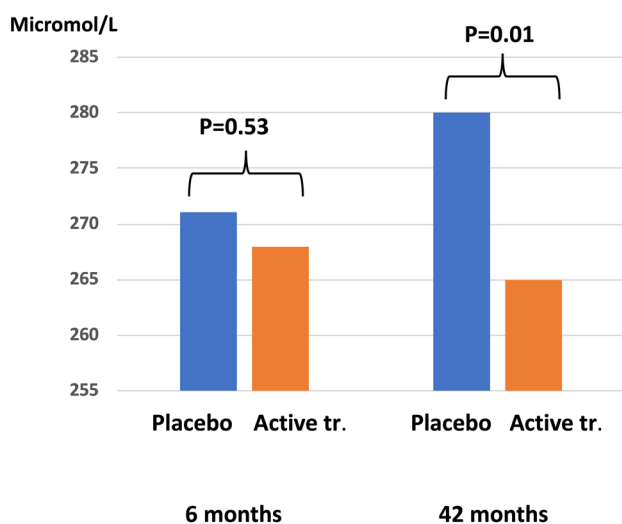
Table 1 Baseline characteristics of the study population receiving dietary supplementation of selenium and coenzyme Q10 combined or placebo during 4 years

	Active treatment group <i>N</i> =221	Placebo group <i>N</i> =222	<i>P</i>
Males/Females <i>n</i>	115/106	110/112	
Age years, mean (SD)	77.0 (3.6)	77.3 (3.4)	0.36
<i>History</i>			
Diabetes, <i>n</i> (%)	47 (21.3)	48 (21.6)	0.93
Hypertension, <i>n</i> (%)	158 (71.5)	168 (75.7)	0.32
IHD, <i>n</i> (%)	47 (21.3)	53 (23.9)	0.51
NYHA class I, <i>n</i> (%)	118 (53.4)	108 (48.6)	0.32
NYHA class II, <i>n</i> (%)	61 (27.6)	64 (28.8)	0.77
NYHA class III, <i>n</i> (%)	41 (18.6)	47 (21.2)	0.49
NYHA class IV, <i>n</i> (%)	0	0	
NYHA unclassified, <i>n</i>	0	3	
<i>Medications</i>			
ACEI, <i>n</i> (%)	35 (15.8)	55 (24.8)	0.02
ARB, <i>n</i> (%)	10 (4.5)	13 (5.9)	0.53
Beta blockers, <i>n</i> (%)	81 (36.7)	73 (32.9)	0.40
Digitalis, <i>n</i> (%)	11 (5.0)	11 (5.0)	0.99
Diuretics, <i>n</i> (%)	70 (31.7)	88 (39.6)	0.08
Statins, <i>n</i> (%)	45 (20.4)	51 (23.0)	0.50
<i>Examinations</i>			
EF <40%, <i>n</i> (%)	16 (7.2)	17 (7.7)	0.87
Atrial fibrillation, <i>n</i> (%)	24 (10.9)	25 (11.3)	0.89
NT-proBNP, ng/L, mean (SD)	547 (1578)	517 (877)	0.81

Note: *ACEI* ACE inhibitors, *ARB* Angiotensin receptor blockers, *EF* Ejection fraction, *IHD* Ischemic heart disease, *NT-proBNP* N-terminal fragment of proBNP, *NYHA* New York Heart Association functional class, *SD* Standard Deviation

Note: Values are means \pm SDs or frequency (percent)

Note: Student's unpaired two-sided t test was used for continuous variables and the chi-square test was used for analysis of one discrete variable

**Fig. 1** Plasma concentration of fructosamine in those with active treatment compared with placebo after six and 42 months

both biomarkers, with a lower concentration in those supplemented with selenium and coenzyme Q10 [37]. The effect appears to be partly mediated by p38 mitogen-activated protein kinases or c-JNK signalling pathways [38, 39]. Therefore, it seems reasonable that the supplementation of selenium and coenzyme Q10 has effects also on inflammation.

To further validate these results, five additional biomarkers of inflammation (osteopontin, osteoprotegerin, sTNF receptor 1, sTNF receptor 2 and the tumour necrosis factor-like weak inducer of apoptosis called TWEAK) were examined [40].

Significant differences were observed in four out of five of the biomarkers, with signs of less inflammatory activity in those on active supplementation with selenium and coenzyme Q10, indicating less inflammatory activity in those supplemented with selenium and coenzyme Q10, as compared to those on placebo.

Oxidative stress

Several selenoproteins and coenzyme Q10 are both important endogenous antioxidants [41, 42]. The reduced form of coenzyme Q10, ubiquinol, protects against lipid peroxidation, and together with selenium, it can reduce the inflammatory response, as discussed above [43, 44].

With increasing age, the level of oxidative stress is increased [45–47]. Copeptin, a surrogate biomarker for vasopressin, and the biomarker adrenomedullin have both been shown to act as markers of the level of oxidative stress in tissues [48, 49]. In those on supplementation with selenium and coenzyme Q10, the levels of the two biomarkers decreased in comparison with those on placebo [50] (Fig. 2). This indicated a reduced intracellular oxidative stress due to the intervention.

Endothelial function

Endothelial function and selenium concentration are associated, and this might have clinical implications [51, 52]. Therefore, two biomarkers of endothelial function were investigated, the von Willebrand factor, and plasminogen activator inhibitor.

The von Willebrand factor is a plasma glycoprotein that is primarily active in the coagulation process, through a process in the vascular endothelium [53], but also an indicator of endothelial dysfunction [54].

Plasminogen activator inhibitor (PAI-1) is secreted from the endothelium and hepatocytes [55, 56], has well-known

antifibrinolytic effects. The factors are also indicators of the endothelial function.

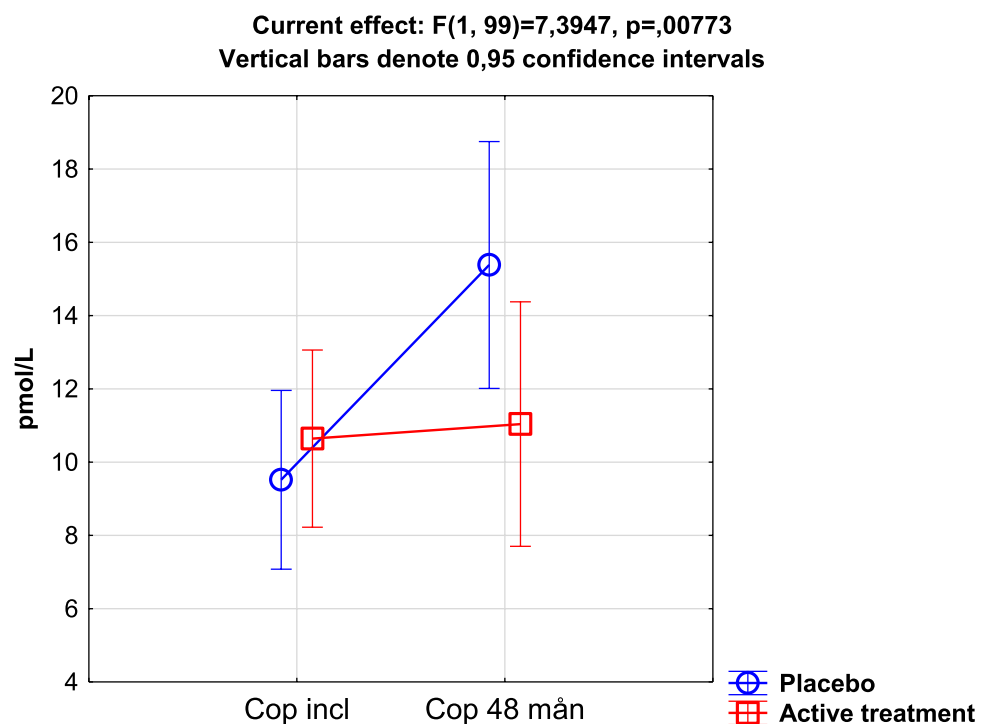
Our studies disclosed a highly significant difference between active treatment and placebo after 36 months with significantly higher concentrations of the two biomarkers in the placebo group, compared with the active treatment group [57], indicating that the intervention results in less endothelial dysfunction. This could indicate less injury of the vascular endothelium, an effect that later could be manifested as established CV disease, and where the end-result could result in a CV death.

Finally, D-dimer, is a fragment of degraded fibrin, and as an indicator of fibrinolysis [58], it is used as a biomarker for venous thromboembolism [59], or pulmonary embolism [60]. However, there is also an association between concentration of D-dimer and mortality risk, even in the absence of thromboembolism, possibly because D-dimer is also a biomarker of inflammation [61]. In addition, there may be an association between D-dimer and endothelial dysfunction as well [62]. In our study, a significantly lower concentration of D-dimer could be noted in the supplementation group, compared with those on placebo [63].

Fibrosis

In the process of ageing and development of diseases in the CV system, fibrosis is one of the characteristics observed [64]. Eight different biomarkers for fibrosis (endostatin, tissue metalloproteinase 1 (TIMP1), galectin 3, and growth differentiation factor 15 (GDF-15), cathepsin S, matrix

Fig. 2 Plasma concentration of copeptin in the active treatment group, compared with placebo at inclusion and after 48 months



metalloproteinases 1 and 9 (MMP-1, MMP-9) and Suppression of tumorigenicity 2 (ST2)) have been evaluated. From these evaluations, significant reductions of concentrations in seven of the eight biomarkers were observed. This could imply that the intervention influenced the fibrous process in the CV system with less fibrosis as a result.

microRNA

The knowledge of microRNAs is fairly recent, and, today, more than 2000 different microRNAs have been characterized [65, 66]. The microRNAs have important functions in the regulation of the protein-encoding mammal genes, and one single microRNA can regulate the production of many different proteins. We found that the combined supplementation resulted in a highly significant difference in expression in 90 out of 145 microRNAs and with up to a fourfold change, when comparing those to placebo [67]. Among these, several microRNAs were related to CV disease, myocardial repair and myocardial function. Substantial changes in microRNA expression observed may act to modulate the synthesis not only of the selenoproteins, but possibly also of other proteins.

Cardiac function

Heart failure is characterized by increased wall tension that leads to increased synthesis of the natriuretic peptide BNP and its N-terminal fragment of proBNP (NT-proBNP). The most important effect of the release of BNP is increased diuresis, and vascular dilatation. However, besides heart failure, increased age is also associated with increased synthesis of the peptide.

Supplementation with selenium and CoQ10 resulted in a decreased NT-proBNP concentration, when compared with the placebo group, indicating reduced myocardial wall tension [1]. The decrease in NT-proBNP is also in accord with a significantly improved systolic cardiac function in the active treatment group as measured by change in ejection fraction (EF) as seen in echocardiography [1].

Cardiovascular mortality and quality of life

The intervention resulted in a significantly reduced CV mortality (5.9% in the active supplementation group versus 12.6% in the placebo group; $p=0.015$), by applying a follow-up time of five years, and a corresponding CV mortality risk reduction in those supplemented with selenium and coenzyme Q10, was found.

By applying a longer follow-up period, 10 years, still a significant reduction of CV mortality remained (46 out of 221 in the active treatment group compared with 86 out of 222 in the placebo group; $p<0.0001$), and with a

corresponding risk reduction, apparent also after adjustment for several well-known variables influencing CV risk [2]. In addition, the positive long-term results have been validated in a follow-up evaluation after 12 years with a HR of 0.59 (95%CI 0.42–0.81; $p=0.001$) of CV mortality risk when using a multivariate model [3]. It should be emphasized that the active intervention was terminated already after four years.

Furthermore, in the project, the health-related quality of life has been evaluated by use of the generic health-related quality of life questionnaire SF-36. The result was a better quality of life reported both in the physical and in the emotional dimensions compared to those on placebo supplementation, which support the results on positive effects combined supplementation discussed above [68].

Aim of present analysis

The aims of the present secondary analysis were to use clinical and biomarker results summarized above to further explore underlying mechanisms behind the reduced CV mortality and improved cardiac function in the elderly Swedish population that was supplemented with selenium and coenzyme Q10, by the use of the structural equation modelling (SEM) technique, and to elucidate cause–effect relations of the supplementation.

Methods of the present secondary analysis

The previously presented main study and the summarized sub-studies are based upon the classical experiment-control group design (a “randomized double-blind placebo-controlled trial”), whereas the present secondary analysis applying a SEM-analyses approach, represents an alternative analytical technique for causal inferences. The SEM analyses also represent a cross-validation of the findings from the main study. Our research group has used SEM modelling in previous publications [69, 70].

Statistical method: structural equation modelling analysis

A stepwise approach in the analysis was adopted.

In a first step, Confirmative Factor Analysis (CFA) was used to reduce the complexity of the 13 measured biomarkers into a lower number of latent, underlying variables or factors. The choice of variables to be measured is central in the evaluation process. However, from the previously published sub-analyses, we could see that obviously some mechanisms were influenced to a greater extent than others. We could see

that inflammation, oxidative stress, and fibrosis were examples of such mechanisms. Primarily, we analysed all variables from these evaluations; however, as some of the variables had a low communality, they could be excluded without any loss of information. Lastly, we knew from a clinical point of view that a factor indicating cardiac function was important, and thus the NT-proBNP concentration was also included.

In a second step, Structural Equation Modelling (SEM) ad modum LISREL was used to explore associations or structural relations/effects between the underlying factors. The SEM approach allows researchers to determine if there are direct as well as indirect relationships between independent and dependent variables [71].

In the third step, the effect of selenium concentration *at baseline* on the model structure was analysed. Finally, linear structural relationships and factor structures were combined into one comprehensive model. Goodness-of-fit of the SEM model was examined with Chi-Square, the Root Mean Square Error of Approximation (RMSEA), and the Comparative Fit Index (CFI). A non-significant Chi-Square, a RMSEA <0.06 and a CFI ≥ 0.95 indicate a good model fit [71].

In the *first step* of the present analysis, 13 different biomarkers (i.e. copeptin, MR-proADM, TNFr1, TNFr2, osteoprotegerin, osteopontin, endostatin, galectin, TIMP1, cathepsin S, GDF-15, MMP-1, and NT-proBNP) were factored into latent variables or factors. The factor and modelling analyses were based on data from the *initial* measurements and thus reflect the natural variation of the biomarkers in the total group at baseline.

Then, the factors identified were used in reliability analyses and comparisons of groups between the first and second occasion 48 months later.

Finally, the relative predictive effects of selenium status and NT-proBNP values (as independent variables) on the subjects age (as dependent variable) were analysed in a regression model. The value was called “CV age” and represents the optimal linear predictive power of selenium and NT-proBNP.

Results

Data reduction and modelling analyses

Confirmative Factor Analyses (CFA) were, in the first step, performed on data from the initial measurement occasion. In the final analysis, three factors, viz. “**Inflammation**”, “**Oxidative Stress**”, and “**Fibrosis**” were extracted. The common variance between TNF receptor1, TNF receptor2, osteoprotegerin, and osteopontin was explained by the factor

“**Inflammation**”, the common variance between copeptin, and MR-proADM was explained by the factor “**Oxidative Stress**”, and the common variance between endostatin, galectin 3, TIMP 1, cathepsin S, GDF-15 and MMP-1 was explained by the factor “**Fibrosis**”. The fit of the factor structure was good with a RMSEA of 0.042, and a Critical Fit Index (CFI) of 0.96, indicating that 96 percent of the co-variances between the 13 biomarkers were explained by the three factors. The construct reliability for “**Inflammation**” was 0.63, 0.70 for “**Oxidative Stress**”, and 0.63 for “**Fibrosis**”. The index presents the proportion of true variance explained by the factor, and values higher than 0.60 indicate a high reliability.

In the *second* step, SEM ad modum LISREL was used to model or explore associations or structural relations/effects between the underlying factors “**Inflammation**”, “**Oxidative Stress**”, and “**Fibrosis**”. To the hypothesized basic model, a factor “Myocardium” representing NT-proBNP (indicating myocardial wall tension) was added. In the model also, a second-order factor “Inflammation/Oxidative Stress” combining the factors “**Inflammation**” and “**Oxidative Stress**” was hypothesized.

When scrutinizing the correlation matrix of the variables, we found that half of the biomarkers at the first occasion were slightly, but significantly, related to the age of the participant (the mean correlation was 0.25 with a range from 0.18 to 0.37). This relation may inflate the correlations of the input matrix and affect the factor relations of the SEM model. This age effect was controlled by means of partial correlation analysis, and the corrected correlation matrix was compared with the uncorrected one. From the comparison, we found that the age effect was only marginal, and 98 percent of the variable relations remained the same. Accordingly, our use of the original matrix in our analyses was valid.

Finding relations between age and several of the biomarkers was expected and has also been reported elsewhere. There was a small but significant relation (-0.13 , $p=0.001$) between selenium concentration (at the first occasion) and age. This has been previously reported in the literature, but in a much younger population [8]. To compensate for the age relation, we created a *quotient* between selenium concentration and age. This independent factor here called “Sel/age” is related 0.84 to selenium concentration, and -0.64 to age.

In the *third* step, this quotient Sel/age was included as an independent factor. By means of this factor we could analyse if and to what extent selenium influences the factors of the basic model.

The final model is presented in Fig. 3. As can be seen from the model (Fig. 3), the first-order factors “**Inflammation**” and “**Oxidative Stress**” form a second-order factor “Infl/Oxidative Stress”. The weights of the first-order factors on the second-order factor are high and about the

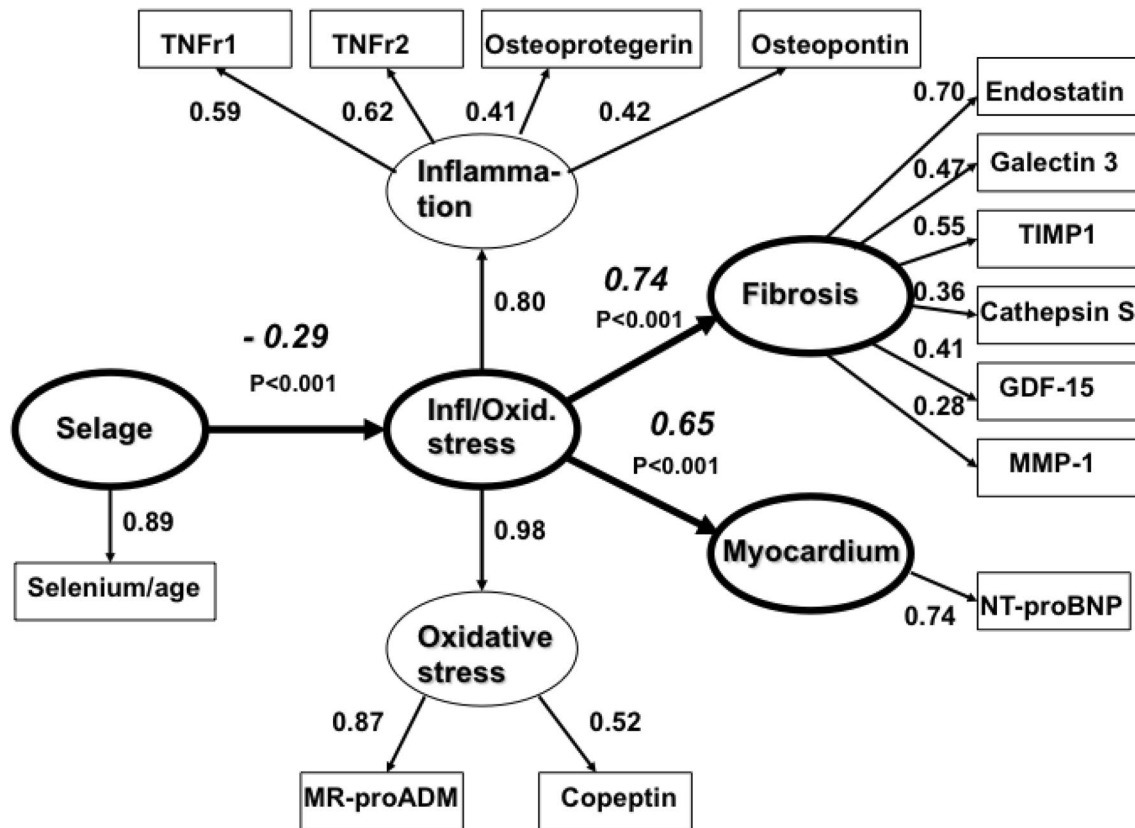


Fig. 3 Structural equation model of the relations between Selenium (Selenium/age), the second-order factor Infl/Oxid. Stress, Fibrosis and Myocardium. Ellipses depict latent variables or factors and

squares manifest or measured variables. All effects (Beta-values italicized) and factor loadings are significant ($p < .001$). Model fit: Chi-square = 56.43, $df = 57$, $p = 0.497$, RMSEA = 0.000, CFI = 0.99

same (0.98, and 0.80). As hypothesized, the factor “Infl/Oxidative Stress” has strong influences on the factors “Fibrosis” ($\beta = 0.74$; $p < 0.001$) and “Myocardium” ($\beta = 0.65$; $p < 0.001$). Finally, the model shows that the independent factor “Sel/age” significantly influences the second-order factor “Infl/Oxidative Stress” ($\beta = -0.29$; $p < 0.001$). All indirect effects (β weights) of “Sel/age” are significant: “Sel/age” → “Inflammation”, -0.23 , ($p = 0.001$), “Sel/age” → “Oxidative Stress”, -0.29 , ($p < 0.001$), “Sel/age” → “Fibrosis”, -0.21 , ($p = 0.003$), and “Sel/age” → “Myocardium”, -0.19 , ($p = 0.007$). This means that the second-order factor “Infl/Oxidative Stress”, representing the factors “Inflammation” and “Oxidative Stress”, acts as a mediating factor between the independent selenium factor “Sel/age” and the dependent factors “Fibrosis” and “Myocardium”. The fit of the presented model is almost perfect (Chi-Square = 56.43, $df = 57$, $p = 0.497$, RMSEA = 0.000, CFI = 0.99), meaning that it explains almost all of the true variance between the manifest or measured variables.

As noted above, the factors “Inflammation”, “Oxidative Stress”, and “Fibrosis” have high construct reliability. Their test–retest reliabilities are also high. The correlation between test and retest for “Inflammation” was 0.57

($p < 0.001$), for “Oxidative Stress” 0.67 ($p < 0.001$), and for “Fibrosis” 0.76 ($p < 0.001$). This means that 32, 45, and 45 percent, respectively, of the factor variance after 48 months can be explained by the factor variance at the initial test occasion.

Changes after intervention

Changes in the active treatment and the placebo groups were evaluated over the 48 months. For the factor “Inflammation”, a significant increase between inclusion and 48 months was noted in the placebo group ($T = -4.1$; $p < 0.001$), whereas in the active treatment group, a significant decrease was noted ($T = 2.6$; $p = 0.010$) (Fig. 4). Upon evaluation of the effect on the factor “Fibrosis”, it was revealed that in the placebo group, no significant difference was seen between inclusion and after 48 months ($T = 1.2$; $p = 0.23$); however, in the active treatment group, a significant decrease was noted ($T = 8.1$; $p < 0.001$). In the placebo group, the factor “Oxidative Stress”, increased significantly ($T = -2.21$; $p = 0.03$), whereas in the active treatment group, there was no significant difference between inclusion and 48 months ($T = 0.39$; $p = 0.70$).

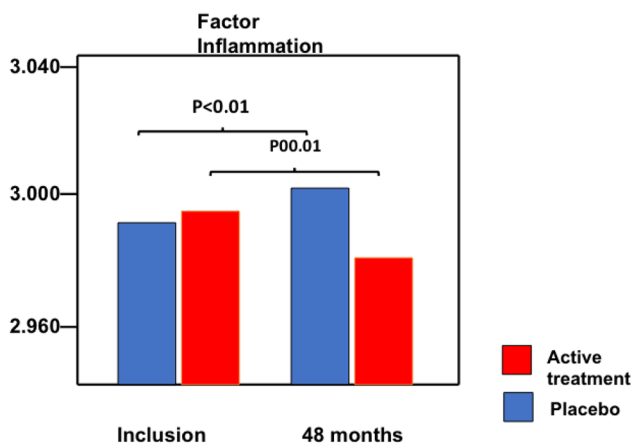


Fig. 4 The obtained changes in factor weights between the factors “Inflammation” before and after intervention with selenium and coenzyme Q₁₀

NT-proBNP is a critical marker of the heart function, and its reference range in healthy individuals increases with increasing age after about 50 years of age [72, 73]. In the present population (age 79–88), selenium supplementation was associated with reduced NT-proBNP-levels. In a multiple regression analysis, using data obtained at baseline, we examined the relative predictive effects of selenium and NT-proBNP (as independent variables) on the subjects age (as dependent variable). From the analysis, an algorithm predicting the functional age of the heart or “CV age” (“CV age” = $82.185 - 0.185 * \text{selenium} + 0.354 * \text{NT-proBNP}$) was evaluated in the cohort. Both the weight for selenium (-0.185 , $t = -2.875$, $p = 0.004$) and the weight for NT-proBNP (0.354 , $t = 5.502$, $p < -0.001$) as well as the fit of the total regression model (ANOVA: Mean Square = 161.570, F-ratio = 21.841, $p < 0.001$) were significant.

Figure 5 shows a linear regression smoothing plane optimizing the fit of data points in a three-dimensional space.

As can be seen from the figure, the highest value of NT-proBNP, in combination with the lowest value of selenium predicts an “CV age” of about 88 years (A), and the lowest value of NT-proBNP in combination with the highest value of selenium predicts a an “CV age” of about 72 years (B). In the present population of elderly individuals with a suboptimal selenium status, the correlation between real age and “CV age” is as low as 0.40 ($p < 0.001$). However, it should be taken into consideration that the range of true age of the subjects is 79–88 years. Furthermore, it is apparent that extrapolations of this relation to populations with optimal selenium intake (> about 100 $\mu\text{g}/\text{day}$), for instance, the cohort in the Framingham study, should not be done.

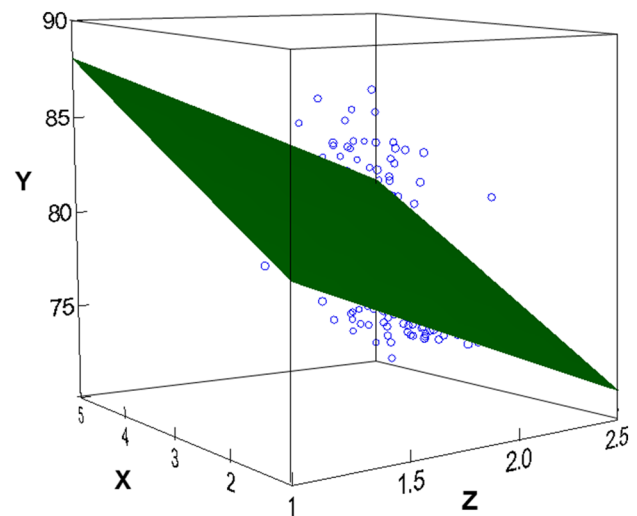


Fig. 5 A linear regression smoothing plane optimizing the fit of data points in a three-dimensional space. The figure is based on the regression equation predicting “CV age” from logarithmized values of NT-proBNP and Selenium. X = LogNT-proBNP, Y = “CV age”, and Z = logselenium

Discussion

The main findings of the randomized double-blind placebo-controlled intervention with selenium and CoQ₁₀ in elderly Swedes low in selenium and coenzyme Q₁₀ reviewed here, were a significantly increased systolic heart function, a reduced myocardial wall tension, and a reduced CV mortality. Based on these findings, a series of secondary analyses to explore some of the underlying mechanisms for the positive clinical results were conducted. Through these analyses, we found evidence for a causal relation between the intervention and changes in a series of biomarkers, including expression of circulating microRNAs.

To further examine the relationships between the impacts on different biomarkers, we undertook an overall analysis of our observations using the SEM approach, which is considered a robust technique regarding causal inferences. The presented SEM analysis provides mechanistic a cross-validation of the findings from the classical RCT design by first developing a model based on data obtained at baseline (Fig. 3), and second, using the follow-up data, we also examined the impact of the intervention in the model (Figs. 4 and 5) in studies using only the RCT approach statistically significant, but clinically insignificant differences, can be generated.

From the SEM analysis, we conclude that the second-order factor “Infl/Oxidative Stress”, representing the factors Inflammation and Oxidative Stress, acts as a mediating factor between the apparently independent selenium factor

Sel/age, and the dependent factors, fibrosis and myocardium. Identification of these independent and dependent factors is based on determinations before the intervention, that there is a well-known inter-relationship between age and inflammation is illustrated by the concept of “Inflammaging” presented by Franceschi in 2007 [74].

The multiple regression analyses indicate that NT-proBNP and selenium together, but independently of each other, may predict a functional age, “CV age”, for the present subjects. While the NT-proBNP-value may act to increase this theoretical age, increasing selenium levels reduces the “CV age”.

The results obtained support the notion that selenium status is a central determinant for several processes in the body, as seen in the present population with a relative deficiency in selenium/coenzyme Q₁₀. Further, the supplementation has verified effects on inflammation, on oxidative stress, on fibrosis and also directly on the myocardium.

The reported decreased levels of both the von Willebrand factor, and PAI-1, resulting from the intervention, indicate that endothelial dysfunction, as an early sign of vascular damage, is alleviated by the intervention.

As clinical results evaluated also include “end-stage” results in a long process (cardiovascular mortality and experienced health-related quality of life [75]), the early signs of impairment such as endothelial dysfunction are important for the prevention.

The endpoint CV mortality is the most definitive endpoint to be used when evaluating effects of intervention on cardiac health. The four years of intervention with selenium and coenzyme Q₁₀ reduced CV mortality not only after five years of follow-up but also eight years after termination of the supplementation. This long-lasting effect may indicate beneficial structural changes due to the intervention.

As the actual genesis to the CV mortality in the present study differs between heart failure, myocardial infarction, fatal arrhythmias, including cerebrovascular events, we suggest that the effect of the intervention could be found in basic vascular mechanisms, e.g. endothelial dysfunction and inflammation.

There are several studies in the literature where either selenium or coenzyme Q₁₀ was given as monotherapy. Rees et al. presented a Cochrane systematic review to elucidate possible effects of intervention with selenium on prevention of cardiovascular disease [14]. Their conclusion was that no data included in their review could support the supplementation with selenium. However, an important fact in their study was that out of the 19,715 participants, 18,415 (93.4%) came from a US-based population, a population that had high baseline selenium [10, 11] and thus, it is not surprising that supplementation with selenium did not give significant effects. Underscoring this is our finding that the tertile

lowest in plasma selenium concentration in terms of CV mortality benefited most from the active intervention [40].

There are some studies where interventions have been performed using a multi-supplement approach, where selenium and coenzyme Q₁₀ have been part of the mixture given. Improved cardiac function and perceived quality of life were reported by Witte et al. in a study of patients with heart failure using capsules containing 14 different micronutrients, including coenzyme Q₁₀ and selenium, or placebo [76].

Even if our main study had a relatively small sample size, the follow-up period was long, up to 12 years, and the secondary analyses consisted of several evaluated variables. The significant results obtained from the many different evaluations confirms that obvious effects do occur from the intervention, and it seems that the lower the selenium intake, the more powerful results of the intervention.

It should be emphasized that the baseline selenium status in the elderly population of the present Kisel study was suboptimal or inadequate, implying that extrapolation of the results to other populations with a higher selenium intake should be done with care.

To re-examine our observations, we would propose a large-scale long-term intervention study in another population with comparable baseline selenium values and use of the same type of intervention.

Conclusion

An intervention study for four years with selenium and coenzyme Q₁₀ combined performed in elderly community-living Swedish subjects showed substantial and long-lasting clinical effects that included improved cardiac function and reduced cardiovascular mortality. We have here given a short summary of some of the most important findings, including the findings on biomarkers of inflammation, oxidative stress and fibrosis. From these findings, it could be demonstrated that supplementation with selenium and coenzyme Q₁₀ could significantly reduce the level of these processes. To validate these results and to further explore possible mechanisms behind these clinical results, we conducted a secondary analysis based on previously published results on a series of biomarkers measured at baseline and during the follow-up period. By applying Structural Equation Modeling (SEM), we identified a model of possible mechanisms. From the model, it could be deduced that the lower the selenium concentration of the individual, the higher was inflammatory activity as well as biomarkers on oxidative stress. Furthermore, suboptimal selenium status was accompanied by a higher level of fibrosis and increased myocardial wall tension which in turn influences the risk of cardiovascular

mortality. The model was validated using data obtained after 48 months of intervention.

Thus, the model shows that in individuals with low selenium and coenzyme Q₁₀ concentration, supplementation with the two substances makes a difference in terms of inflammation, oxidative stress, fibrosis and cardiovascular risk.

Author contributions Conceived and designed the research project, UA and PJ; conducted the research, UA; analysed data and performed the statistical analyses, UA and ES; wrote the paper, UA, and JAa, and JA, and PJ and ES; had the final responsibility for the final content; UA. All authors have read and approved the final manuscript.

Funding/support Open access funding provided by Linköping University. Part of the analysis cost was supported by grants from Pharma Nord Aps, Denmark, the County Council of Östergötland, Linköping University. The funding organisations had no role in the design, management, analysis, or interpretation of the data, nor in the preparation, review or approval of the manuscript. No economic compensation was distributed.

Data availability Under Swedish Law, the authors cannot share the data used in this study and cannot conduct any further research other than what is specified in the ethical permissions application. For inquiries about the data, researchers should first contact the owner of the database, the University of Linköping. Please contact the corresponding author with requests for and assistance with data. If the university approves the request, researchers can submit an application to the Regional Ethical Review Board for the specific research question that the researcher wants to examine.

Declarations

Conflict of interest The authors declare no conflicts of interest.

Ethical approval The study was approved by the Regional Ethical Committee (Forskningsetikommittén, Hälsouniversitetet, SE-581 85 Linköping, Sweden; No. D03-176), and conforms to the ethical guidelines of the 1975 Declaration of Helsinki. (As the Medical Product Agency considered the trial as a trial of one food supplement and not a medication, it declined to review the study protocol). This study has been registered retrospectively at Clinicaltrials.gov, and has the identifier NCT01443780, as it was not mandatory to register at the time the study began.

Informed consent Informed consent was obtained from each patient regarding participation and publication of data.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

1. Alehagen U, Johansson P, Bjornstedt M, Rosen A, Dahlstrom U (2013) Cardiovascular mortality and N-terminal-proBNP reduced after combined selenium and coenzyme Q10 supplementation: a 5-year prospective randomized double-blind placebo-controlled trial among elderly Swedish citizens. *Int J Cardiol* 167:1860–1866. <https://doi.org/10.1016/j.ijcard.2012.04.156>
2. Alehagen U, Aaseth J, Johansson P (2015) Reduced cardiovascular mortality 10 years after supplementation with selenium and coenzyme q10 for four years: follow-up results of a prospective randomized double-blind placebo-controlled trial in elderly citizens. *PLoS One* 10:e0141641. <https://doi.org/10.1371/journal.pone.0141641>
3. Alehagen U, Aaseth J, Alexander J, Johansson P (2018) Still reduced cardiovascular mortality 12 years after supplementation with selenium and coenzyme Q10 for four years: a validation of previous 10-year follow-up results of a prospective randomized double-blind placebo-controlled trial in elderly. *PLoS ONE* 13:e0193120. <https://doi.org/10.1371/journal.pone.0193120>
4. Brodin O, Hackler J, Misra S, Wendt S, Sun Q, Laaf E, Stoppe C, Bjornstedt M, Schomburg L (2020) Selenoprotein P as biomarker of selenium status in clinical trials with therapeutic dosages of selenite. *Nutrients*. <https://doi.org/10.3390/nu12041067>
5. Karunasinghe N, Han DY, Zhu S, Duan H, Ko YJ, Yu JF, Triggs CM, Ferguson LR (2013) Effects of supplementation with selenium, as selenized yeast, in a healthy male population from New Zealand. *Nutr Cancer* 65:355–366. <https://doi.org/10.1080/01635581.2013.760743>
6. Van Cauwenbergh R, Robberecht H, Van Vlaslaer V, Deelstra H (2004) Comparison of the serum selenium content of healthy adults living in the Antwerp region (Belgium) with recent literature data. *J Trace Elem Med Biol* 18:99–112. <https://doi.org/10.1016/j.jtemb.2004.04.004>
7. Burri J, Haldimann M, Dudler V (2008) Selenium status of the Swiss population: assessment and change over a decade. *J Trace Elem Med Biol* 22:112–119. <https://doi.org/10.1016/j.jtemb.2007.11.002>
8. Letsiou S, Nomikos T, Panagiotakos D, Pergantis SA, Fragopoulou E, Antonopoulou S, Pitsavos C, Stefanadis C (2009) Serum total selenium status in Greek adults and its relation to age. The ATTICA study cohort. *Biol Trace Elem Res* 128:8–17. <https://doi.org/10.1007/s12011-008-8252-2>
9. Spina A, Guallar E, Rayman MP, Tigbe W, Kandala NB, Stranges S (2013) Anthropometric indices and selenium status in British adults: the U.K. National diet and nutrition survey. *Free Radic Biol Med* 65:1315–1321. <https://doi.org/10.1016/j.freeradbiomed.2013.09.025>
10. Kafai MR, Ganji V (2003) Sex, age, geographical location, smoking, and alcohol consumption influence serum selenium concentrations in the USA: third National Health and Nutrition Examination Survey, 1988–1994. *J Trace Elem Med Biol* 17:13–18. [https://doi.org/10.1016/S0946-672X\(03\)80040-8](https://doi.org/10.1016/S0946-672X(03)80040-8)
11. Bleys J, Navas-Acien A, Laclaustra M, Pastor-Barriuso R, Menke A, Ordovas J, Stranges S, Guallar E (2009) Serum selenium and peripheral arterial disease: results from the national health and nutrition examination survey, 2003–2004. *Am J Epidemiol* 169:996–1003
12. Alfthan G, Euroala M, Ekholm P, Venalainen ER, Root T, Korkalainen K, Hartikainen H, Salminen P, Hietaniemi V, Aspila P, Aro A, for the Selenium Working G, (2014) Effects of nationwide addition of selenium to fertilizers on foods, and animal and human health in Finland: From deficiency to optimal selenium status of the population. *J Trace Elem Med Biol*. <https://doi.org/10.1016/j.jtemb.2014.04.009>

13. Akbaraly NT, Arnaud J, Hininger-Favier I, Gourlet V, Rousset AM, Berr C (2005) Selenium and mortality in the elderly: results from the EVA study. *Clin Chem* 51:2117–2123. <https://doi.org/10.1373/clinchem.2005.055301>
14. Rees K, Hartley L, Day C, Flowers N, Clarke A, Stranges S (2013) Selenium supplementation for the primary prevention of cardiovascular disease. *Cochrane Database Syst Rev* 1:CD009671 <https://doi.org/10.1002/14651858.CD009671.pub2>
15. Hatfield DL, Gladyshev VN (2009) The outcome of selenium and vitamin e cancer prevention trial (SELECT) reveals the need for better understanding of selenium biology. *Mol Interv* 9:18–21. <https://doi.org/10.1124/mi.9.1.6>
16. Harris HR, Bergkvist L, Wolk A (2012) Selenium intake and breast cancer mortality in a cohort of Swedish women. *Breast Cancer Res Treat* 134:1269–1277. <https://doi.org/10.1007/s10549-012-2139-9>
17. Gao H, Hagg S, Sjogren P, Lambert PC, Ingelsson E, van Dam RM (2014) Serum selenium in relation to measures of glucose metabolism and incidence of Type 2 diabetes in an older Swedish population. *Diabet Med* 31:787–793. <https://doi.org/10.1111/dme.12429>
18. Alehagen U, Alexander J, Aaseth J (2016) Supplementation with selenium and coenzyme Q10 reduces cardiovascular mortality in elderly with low selenium status. A secondary analysis of a randomised clinical trial. *PLoS One* 11:e0157541. <https://doi.org/10.1371/journal.pone.0157541>
19. Hurst R, Armah CN, Dainty JR, Hart DJ, Teucher B, Goldson AJ, Broadley MR, Motley AK, Fairweather-Tait SJ (2010) Establishing optimal selenium status: results of a randomized, double-blind, placebo-controlled trial. *Am J Clin Nutr* 91:923–931. <https://doi.org/10.3945/ajcn.2009.28169>
20. Alehagen U, Johansson P, Bjornstedt M, Rosen A, Post C, Aaseth J (2016) Relatively high mortality risk in elderly Swedish subjects with low selenium status. *Eur J Clin Nutr* 70:91–96. <https://doi.org/10.1038/ejcn.2015.92>
21. Fairweather-Tait SJ, Bao Y, Broadley MR, Collings R, Ford D, Hesketh JE, Hurst R (2011) Selenium in human health and disease. *Antioxid Redox Signal* 14:1337–1383. <https://doi.org/10.1089/ars.2010.3275>
22. Rayman MP (2000) The importance of selenium to human health. *Lancet* 356:233–241
23. Xia L, Nordman T, Olsson JM, Damdimopoulos A, Bjorkhem-Bergman L, Nalvarte I, Eriksson LC, Arner ES, Spyrou G, Bjornstedt M (2003) The mammalian cytosolic selenoenzyme thioredoxin reductase reduces ubiquinone. A novel mechanism for defense against oxidative stress. *J Biol Chem* 278:2141–2146
24. Kalen A, Appelkvist EL, Dallner G (1989) Age-related changes in the lipid compositions of rat and human tissues. *Lipids* 24:579–584
25. Aaseth J, Alexander J, Alehagen U (2021) Coenzyme Q10 supplementation - In ageing and disease. *Mech Ageing Dev* 197:111521. <https://doi.org/10.1016/j.mad.2021.111521>
26. Selvin E, Rawlings AM, Lutsey PL, Maruthur N, Pankow JS, Steffes M, Coresh J (2015) Fructosamine and glycated albumin and the risk of cardiovascular outcomes and death. *Circulation* 132:269–277. <https://doi.org/10.1161/CIRCULATIONAHA.115.015415>
27. Babu NP, Bobby Z, Selvaraj N, Harish BN (2006) Increased fructosamine in non-diabetic rheumatoid arthritis patients: role of lipid peroxides and glutathione. *Clin Chem Lab Med* 44:848–852. <https://doi.org/10.1515/CCLM.2006.159>
28. Alehagen U, Aaseth J, Alexander J, Johansson P, Larsson A (2020) Supplemental selenium and coenzyme Q10 reduce glycation along with cardiovascular mortality in an elderly population with low selenium status - A four-year, prospective, randomised, double-blind placebo-controlled trial. *J Trace Elem Med Biol* 61:126541. <https://doi.org/10.1016/j.jtemb.2020.126541>
29. Malmstrom H, Wandell PE, Holzmann MJ, Arnlov J, Jungner I, Hammar N, Walldius G, Carlsson AC (2016) Low fructosamine and mortality - A long term follow-up of 215,011 non-diabetic subjects in the Swedish AMORIS study. *Nutr Metab Cardiovasc Dis* 26:1120–1128. <https://doi.org/10.1016/j.numecd.2016.08.006>
30. Li Q, Li B, Wang X, Leri A, Jana KP, Liu Y, Kajstura J, Baserga R, Anversa P (1997) Overexpression of insulin-like growth factor-1 in mice protects from myocyte death after infarction, attenuating ventricular dilation, wall stress, and cardiac hypertrophy. *J Clin Invest* 100:1991–1999. <https://doi.org/10.1172/JCI119730>
31. Poudel SB, Dixit M, Neginskaya M, Nagaraj K, Pavlov E, Werner H, Yakar S (2020) Effects of GH/IGF on the aging mitochondria. *Cells*. <https://doi.org/10.3390/cells9061384>
32. Rudman D, Kutner MH, Rogers CM, Lubin MF, Fleming GA, Bain RP (1981) Impaired growth hormone secretion in the adult population: relation to age and adiposity. *J Clin Invest* 67:1361–1369
33. Juul A, Scheike T, Davidsen M, Gyllenberg J, Jorgensen T (2002) Low serum insulin-like growth factor I is associated with increased risk of ischemic heart disease: a population-based case-control study. *Circulation* 106:939–944
34. Schutte AE, Conti E, Mels CM, Smith W, Kruger R, Botha S, Gnassi L, Volpe M, Huisman HW (2016) Attenuated IGF-1 predicts all-cause and cardiovascular mortality in a Black population: A five-year prospective study. *Eur J Prev Cardiol* 23:1690–1699. <https://doi.org/10.1177/2047487316661436>
35. Alehagen U, Johansson P, Aaseth J, Alexander J, Brismar K (2017) Increase in insulin-like growth factor 1 (IGF-1) and insulin-like growth factor binding protein 1 after supplementation with selenium and coenzyme Q10. A prospective randomized double-blind placebo-controlled trial among elderly Swedish citizens. *PLoS One* 12:e0178614. <https://doi.org/10.1371/journal.pone.0178614>
36. Mertens K, Lowes DA, Webster NR, Talib J, Hall L, Davies MJ, Beattie JH, Galley HF (2015) Low zinc and selenium concentrations in sepsis are associated with oxidative damage and inflammation. *Br J Anaesth* 114:990–999. <https://doi.org/10.1093/bja/aev073>
37. Alehagen U, Lindahl TL, Aaseth J, Svensson E, Johansson P (2015) Levels of sP-selectin and hs-CRP decrease with dietary intervention with selenium and coenzyme Q10 combined: a secondary analysis of a randomized clinical trial. *PLoS One* 10:e0137680. <https://doi.org/10.1371/journal.pone.0137680>
38. Li YB, Han JY, Jiang W, Wang J (2011) Selenium inhibits high glucose-induced cyclooxygenase-2 and P-selectin expression in vascular endothelial cells. *Mol Biol Rep* 38:2301–2306. <https://doi.org/10.1007/s11033-010-0362-1>
39. Rashtchizadeh N, Karimi P, Dehgan P, Salimi Movahed M (2015) Effects of selenium in the MAPK signaling cascade. *J Cardiovasc Thorac Res* 7:107–112. <https://doi.org/10.15171/jcvtr.2015.23>
40. Alehagen U, Alexander J, Aaseth J, Larsson A (2019) Decrease in inflammatory biomarker concentration by intervention with selenium and coenzyme Q10: a subanalysis of osteopontin, osteoprotegerin, TNF α 1, TNF α 2 and TWEAK. *J Inflamm (Lond)* 16:5. <https://doi.org/10.1186/s12950-019-0210-6>
41. Richie JP Jr, Das A, Calcagnotto AM, Sinha R, Neidig W, Liao J, Lengerich EJ, Berg A, Hartman TJ, Ciccarella A, Baker A, Kaag MG, Goodin S, DiPaola RS, El-Bayoumy K (2014) Comparative effects of two different forms of selenium on oxidative stress biomarkers in healthy men: a randomized clinical trial. *Cancer Prev Res (Phila)* 7:796–804. <https://doi.org/10.1158/1940-6207.CAPR-14-0042>

42. Yen CH, Chu YJ, Lee BJ, Lin YC, Lin PT (2018) Effect of liquid ubiquinol supplementation on glucose, lipids and antioxidant capacity in type 2 diabetes patients: a double-blind, randomised, placebo-controlled trial. *Br J Nutr* 120:57–63. <https://doi.org/10.1017/S0007114518001241>
43. Bullon P, Roman-Malo L, Marin-Aguilar F, Alvarez-Suarez JM, Giampieri F, Battino M, Cordero MD (2015) Lipophilic antioxidants prevent lipopolysaccharide-induced mitochondrial dysfunction through mitochondrial biogenesis improvement. *Pharmacol Res* 91:1–8. <https://doi.org/10.1016/j.phrs.2014.10.007>
44. Ito K, Watanabe C, Nakamura A, Oikawa-Tada S, Murata M (2015) Reduced coenzyme Q10 decreases urinary 8-Oxo-7,8-Dihydro-2'-deoxyguanosine concentrations in healthy young female subjects. *J Med Food* 18:835–840. <https://doi.org/10.1089/jmf.2014.3302>
45. Guntani A, Matsumoto T, Kyuragi R, Iwasa K, Onohara T, Itoh H, Katusic ZS, Maehara Y (2011) Reduced proliferation of aged human vascular smooth muscle cells—role of oxygen-derived free radicals and BubR1 expression. *J Surg Res* 170:143–149. <https://doi.org/10.1016/j.jss.2011.03.024>
46. Moon SK, Thompson LJ, Madamanchi N, Ballinger S, Papaconstantinou J, Horaist C, Runge MS, Patterson C (2001) Aging, oxidative responses, and proliferative capacity in cultured mouse aortic smooth muscle cells. *Am J Physiol Heart Circ Physiol* 280:H2779–2788. <https://doi.org/10.1152/ajpheart.2001.280.6.H2779>
47. Mahmoudi M, Gorenne I, Mercer J, Figg N, Littlewood T, Bennett M (2008) Statins use a novel Nijmegen breakage syndrome-1-dependent pathway to accelerate DNA repair in vascular smooth muscle cells. *Circ Res* 103:717–725. <https://doi.org/10.1161/CIRCRESAHA.108.182899>
48. Shimosawa T, Shibagaki Y, Ishibashi K, Kitamura K, Kangawa K, Kato S, Ando K, Fujita T (2002) Adrenomedullin, an endogenous peptide, counteracts cardiovascular damage. *Circulation* 105:106–111
49. Hu W, Shi L, Li MY, Zhou PH, Qiu B, Yin K, Zhang HH, Gao Y, Kang R, Qin SL, Ning JZ, Wang W, Zhang LJ (2017) Adrenomedullin protects Leydig cells against lipopolysaccharide-induced oxidative stress and inflammatory reaction via MAPK/NF-kappaB signalling pathways. *Sci Rep* 7:16479. <https://doi.org/10.1038/s41598-017-16008-x>
50. Alehagen U, Aaseth J, Johansson P (2015) Less increase of copeptin and MR-proADM due to intervention with selenium and coenzyme Q10 combined: Results from a 4-year prospective randomized double-blind placebo-controlled trial among elderly Swedish citizens. *BioFactors* 41:443–452. <https://doi.org/10.1002/biof.1245>
51. Gac P, Urbanik D, Pawlas N, Poreba M, Martynowicz H, Prokopowicz A, Januszewska L, Markiewicz-Gorka I, Jaremkow A, Mazur G, Poreba R, Pawlas K (2020) Total antioxidant status reduction conditioned by a serum selenium concentration decrease as a mechanism of the ultrasonographically measured brachial artery dilatation impairment in patients with arterial hypertension. *Environ Toxicol Pharmacol* 75:103332. <https://doi.org/10.1016/j.etap.2020.103332>
52. Vural P, Kabaca G, Firat RD, Degirmencioglu S (2017) Administration of selenium decreases lipid peroxidation and increases vascular endothelial growth factor in streptozotocin induced diabetes mellitus. *Cell J* 19:452–460. <https://doi.org/10.22074/cellj.2017.4161>
53. Padilla A, Moake JL, Bernardo A, Ball C, Wang Y, Arya M, Nolasco L, Turner N, Berndt MC, Anvari B, Lopez JA, Dong JF (2004) P-selectin anchors newly released ultralarge von Willebrand factor multimers to the endothelial cell surface. *Blood* 103:2150–2156. <https://doi.org/10.1182/blood-2003-08-2956>
54. Kawecki C, Lenting PJ, Denis CV (2017) von Willebrand factor and inflammation. *J Thromb Haemost* 15:1285–1294. <https://doi.org/10.1111/jth.13696>
55. Czekay RP, Wilkins-Port CE, Higgins SP, Freytag J, Overstreet JM, Klein RM, Higgins CE, Samarakoon R, Higgins PJ (2011) PAI-1: An Integrator of cell signaling and migration. *Int J Cell Biol* 2011:562481. <https://doi.org/10.1155/2011/562481>
56. Declerck PJ, Gils A (2013) Three decades of research on plasminogen activator inhibitor-1: a multifaceted serpin. *Semin Thromb Hemost* 39:356–364. <https://doi.org/10.1055/s-0033-1334487>
57. Alehagen U, Alexander J, Aaseth J, Larsson A, Lindahl TL (2020) Significant decrease of von Willebrand factor and plasminogen activator inhibitor-1 by providing supplementation with selenium and coenzyme Q10 to an elderly population with a low selenium status. *Eur J Nutr*. <https://doi.org/10.1007/s00394-020-02193-5>
58. Adam SS, Key NS, Greenberg CS (2009) D-dimer antigen: current concepts and future prospects. *Blood* 113:2878–2887. <https://doi.org/10.1182/blood-2008-06-165845>
59. van der Graaf F, van den Borne H, van der Kolk M, de Wild PJ, Janssen GW, van Uum SH (2000) Exclusion of deep venous thrombosis with D-dimer testing—comparison of 13 D-dimer methods in 99 outpatients suspected of deep venous thrombosis using venography as reference standard. *Thromb Haemost* 83:191–198
60. Kearon C, de Wit K, Parpia S, Schulman S, Afilalo M, Hirsch A, Spencer FA, Sharma S, D'Aragon F, Deshaies JF, Le Gal G, Lazo-Langner A, Wu C, Rudd-Scott L, Bates SM, Julian JA, Investigators PES (2019) Diagnosis of pulmonary embolism with d-dimer adjusted to clinical probability. *N Engl J Med* 381:2125–2134. <https://doi.org/10.1056/NEJMoa1909159>
61. Bruinstroop E, van de Ree MA, Huisman MV (2009) The use of D-dimer in specific clinical conditions: a narrative review. *Eur J Intern Med* 20:441–446. <https://doi.org/10.1016/j.ejim.2008.12.004>
62. Hamaad A, Sosin MD, Blann AD, Lip GY, MacFadyen RJ (2009) Markers of thrombosis and hemostasis in acute coronary syndromes: relationship to increased heart rate and reduced heart-rate variability. *Clin Cardiol* 32:204–209. <https://doi.org/10.1002/clc.20321>
63. Alehagen U, Aaseth J, Lindahl TL, Larsson A, Alexander J (2021) Dietary supplementation with selenium and coenzyme Q10 prevents increase in plasma D-dimer while lowering cardiovascular mortality in an elderly Swedish population. *Nutrients*. <https://doi.org/10.3390/nu13041344>
64. Kong P, Christia P, Frangogiannis NG (2014) The pathogenesis of cardiac fibrosis. *Cell Mol Life Sci* 71:549–574. <https://doi.org/10.1007/s00018-013-1349-6>
65. Lee RC, Feinbaum RL, Ambros V (1993) The *C. elegans* heterochronic gene *lin-4* encodes small RNAs with antisense complementarity to *lin-14*. *Cell* 75:843–854
66. Wightman B, Ha I, Ruvkun G (1993) Posttranscriptional regulation of the heterochronic gene *lin-14* by *lin-4* mediates temporal pattern formation in *C. elegans*. *Cell* 75:855–862
67. Alehagen U, Johansson P, Aaseth J, Alexander J, Wagsater D (2017) Significant changes in circulating microRNA by dietary supplementation of selenium and coenzyme Q10 in healthy elderly males. A subgroup analysis of a prospective randomized double-blind placebo-controlled trial among elderly Swedish citizens. *PLoS One* 12:e0174880. <https://doi.org/10.1371/journal.pone.0174880>
68. Johansson P, Dahlstrom O, Dahlstrom U, Alehagen U (2015) Improved health-related quality of life, and more days out of hospital with supplementation with selenium and coenzyme Q10 combined. Results from a double blind, placebo-controlled

- prospective study. *J Nutr Health Aging* 19:870–877. <https://doi.org/10.1007/s12603-015-0509-9>
69. Johansson P, Riegel B, Svensson E, Brostrom A, Alehagen U, Dahlstrom U, Jaarsma T (2014) Sickness behavior in community-dwelling elderly: associations with impaired cardiac function and inflammation. *Biol Res Nurs* 16:105–113. <https://doi.org/10.1177/1099800412466170>
 70. Johansson P, Alehagen U, Svensson E, Svanborg E, Dahlstrom U, Brostrom A (2010) Determinants of global perceived health in community-dwelling elderly screened for heart failure and sleep-disordered breathing. *J Cardiovasc Nurs* 25:E16–26. <https://doi.org/10.1097/JCN.0b013e3181d6de6f>
 71. Jöreskog K, Sörbom D (1993) LISREL 8: Structural equation modeling with the simplis command language Lawrence Erlbaum Associates Inc. New Jersey
 72. Vergaro G, Januzzi JL Jr, Cohen Solal A, Aimo A, Arzilli C, Zyw L, Valleggi A, Giannoni A, Prontera C, Barison A, Poletti R, Gabutti A, Mammini C, Passino C, Emdin M (2018) NT-proBNP prognostic value is maintained in elderly and very elderly patients with chronic systolic heart failure. *Int J Cardiol* 271:324–330. <https://doi.org/10.1016/j.ijcard.2018.04.006>
 73. Choi HI, Lee MY, Oh BK, Lee SJ, Kang JG, Lee SH, Lee JY, Kim BJ, Kim BS, Kang JH, Sung KC (2021) Effects of age, sex, and obesity on n-terminal pro B-type natriuretic peptide concentrations in the general population. *Circ J* 85:647–654. <https://doi.org/10.1253/circj.CJ-20-1104>
 74. Franceschi C, Capri M, Monti D, Giunta S, Olivieri F, Sevini F, Panourgia MP, Invidia L, Celani L, Scurti M, Cevenini E, Castellani GC, Salvioli S (2007) Inflammaging and anti-inflammaging: a systemic perspective on aging and longevity emerged from studies in humans. *Mech Ageing Dev* 128:92–105. <https://doi.org/10.1016/j.mad.2006.11.016>
 75. Wilson IB, Cleary PD (1995) Linking clinical variables with health-related quality of life. A conceptual model of patient outcomes. *JAMA* 273:59–65
 76. Witte KK, Nikitin NP, Parker AC, von Haehling S, Volk HD, Anker SD, Clark AL, Cleland JG (2005) The effect of micro-nutrient supplementation on quality-of-life and left ventricular function in elderly patients with chronic heart failure. *Eur Heart J* 26:2238–2244