BTZ-Derived Benzisothiazolinones with In Vitro Activity against Mycobacterium tuberculosis

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elucidation show little or no antimycobacterial activity in vitro. The structures of the four compounds were investigated by X-ray crystallography and NMR spectroscopy. Molecular covalent docking of the new

compounds to Mycobacerium tuberculosis decaprenylphosphoryl-β-d-ribose 2'-epimerase (DprE1) suggests that the active BITs exert antimycobacterial activity through inhibition of DprE1 like BTZs.

BTZ-SO

KEYWORDS: benzisothiazolinone, BTZ043, crystal structure, antimycobacterial activity, DprE1, tuberculosis

uberculosis (TB), the disease usually caused by Mycobacterium tuberculosis, globally continues to be the leading cause of death from a bacterial infectious agent.¹ According to the World Health Organization, a total of 1.5 million people died from and an estimated 9.9 million people fell ill with TB in 2020.² In particular, the increasing emergence of drug-resistant TB is a threat to public health.^{3,} Whereas about 85% of patients with drug-susceptible active TB can be successfully treated with a six-month regime of four first-line drugs,² treatment of drug-resistant TB in general takes longer, requires more expensive drugs that result in more side effects, and has lower success rates.^{5,6} Drug discovery efforts are thus vital to combat drug-resistance and to reduce the duration, cost, and side effects of TB therapy through maintaining a copious anti-TB drug supply.⁷⁻⁹

8-Nitro-1,3-benzothiazin-4-ones (BTZs) have been identified as potent antitubercular agents.¹⁰ The two most advanced compounds of this class, BTZ043 (Scheme 1) and macozinone (PBTZ169),¹¹ have progressed to clinical studies.⁷ BTZs and several other electron-deficient nitroaromatic agents target the mycobacterial enzyme decaprenylphosphoryl-β-d-ribose 2'epimerase (DprE1), $^{12-14}$ which is crucial for the biosynthesis of arabinogalactan, a major structural component of the mycobacterial cell wall. Upon binding to the active site of DprE1, the BTZ nitro group is reduced to a reactive nitroso

group, which then forms a covalent semimercaptal with the thiol group of cysteine 387.15 BTZs are thus mechanism-based inhibitors, which irreversibly inactivate the enzyme. For this mechanism of action, an electron-deficient warhead is essential. Besides the nitro group it was recently shown that 8-cyano BTZs also have antimycobacterial activity.¹⁶ Efforts to modify the substituent at position 6 and the side chain at position 2 of the BTZ scaffold to improve physicochemical properties have been made.^{17–20} Alteration of the BTZ core by means of scaffold simplification²¹ and metabolically relevant chemical transformations of the scaffold have also been reported.^{22,23}

In 2015, Tiwari et al. described the purported sulfoxide ("BTZ-SO") and sulfone ("BTZ-SO2") of BTZ043 in this journal,²⁴ putatively obtained in very low yields by reaction of BTZ043 with the *m*-chloroperoxybenzoic acid (Scheme 1). The chemical structures of "BTZ-SO" and "BTZ-SO,", as

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Scheme 1. Reaction of BTZs 1a and 1b (BTZ043) with *m*-Chloroperoxybenzoic Acid (*m*CPBA): Putative Chemical Structures "BTZ-SO" and "BTZ-SO₂" Assigned by Tiwari et al.²⁴ and Corresponding Revised Chemical Structures Reported by Eckhardt et al.²⁵



Scheme 2. Syntheses of N-Acyl BITs 4a and 4b and O-Acyl Benzisothiazolols 5a and 5b^a



^{*a*}Reagents and conditions: (i) CuI, 1,10-phenanthroline, sulfur, K_2CO_3 , DMF, 110 °C, overnight; (ii) piperidine-1-carbonyl chloride (a) or (S)-2-methyl-1,4-dioxa-8-azaspiro[4.5]decane-8-carbonyl chloride (b), pyridine, CH₂Cl₂, rt, 24 h.

depicted in Scheme 1, were assigned on the basis of ¹H NMR spectroscopy and ESI mass spectrometry.

Unexpected upfield shifts of the two aromatic ¹H NMR signals, as compared with the parent BTZ043, were attributed to the influence of the lone pair at the sulfur atom and loss of aromaticity due to puckering of the thiazinone ring, as observed in AM1 computed molecular structures of the anticipated "BTZ-SO" and "BTZ-SO₂". Recently, X-ray crystallography revealed that the supposed BTZ sulfones resulting from treatment of BTZs **1a** and **1b** with *m*-chloroperoxybenzoic acid are in fact the corresponding benzisothiazolinone (BIT) 1-oxides (Scheme 1), which represent constitutional isomers.²⁵ Crystallographic evidence for the unintended BIT 1-oxide formation, retrosynthesis of a BIT and revisiting the ¹H NMR data eventually led to the revision of the chemical structures as depicted in Scheme 1 by

Eckhardt et al. in 2020. In vitro activity data included in the previous reports revealed that the BITs exhibit comparable or better in vitro activity against M. tuberculosis and Mycobacterium aurum than the corresponding BTZs 1a and 1b.²⁵ This prompted us to further investigate these compounds. In this contribution, we report on the synthesis, structural characterization, and in vitro and in silico antimycobacterial evaluation of two BITs and their O-acyl benzisothiazol-3-ol constitutional isomers. The chemistry used to prepare the compounds studied is outlined in Scheme 2. N-Acyl BITs 4 and O-acyl benzisothiazolols 5 were synthesized in two steps from 2chloro-5-trifluoromethyl-3-nitrobenzamide²⁶ (2). In the first step, BIT 3 was synthesized from 2 by a copper-mediated sulfur-nitrogen coupling reaction.²⁷ In the second step, 3 was reacted with piperidine-1-carbonyl chloride (for 4a and 5a) or (S)-2-methyl-1,4-dioxa-8-azaspiro[4.5]decane-8-carbonyl



Figure 1. Crystal structure of $4a \cdot 0.25 H_2O$. (a) Asymmetric unit. Carbon-bound hydrogen atoms are omitted for clarity. (b) Structure overlay (benzene rings) of the three crystallographically unique molecules (pink, blue, gray). (c) Displacement ellipsoid plot (50% probability) of molecule 1. Rotational disorder of the trifluoromethyl group and positional disorder of a water hydrogen atom are not shown for clarity. Color scheme: C, gray; H, white; N, blue; O, red; S, yellow.

chloride^{23,28} (for 4b and 5b)^{29,30} to obtain 4 and 5. Using pyridine as a base instead of triethylamine significantly improved the yield in the latter step. Compounds 4a and 5a as well as 4b and 5b represent constitutional isomers, which were separated by flash chromatography in both cases. The benzisothiazol-3-ol system in 5a and 5b represents a tautomeric form of the BIT system in 4a and 4b. Concomitant *O*-acylation benzisothiazol-3-ol of the tautomer, which is expected to be less stable than the BIT form, affords isolable *O*-acyl derivatives 5. The ratio of 4 to 5 was around 1:2 in both cases. Higher electron density and better steric accessibility of the oxygen atom of the isothiazol-3-ol form compared with the



Figure 3. Asymmetric unit of the crystal structure of **4b**. Displacement ellipsoid plots are drawn at the 50% probability level. Rotational disorder of the trifluoromethyl groups is omitted for clarity. Color scheme: C, gray; H, white; N, blue; O, red; S, yellow.



Figure 4. Section of the crystal structure of **5a**, showing two molecules related by a crystallographic inversion center. Displacement ellipsoids are drawn at the 50% probability level. The crystal structure was refined with aspherical scattering factors using NoSpherA2.³⁸ Color scheme: C, gray; H, white; N, blue; O, red; S, yellow.



Figure 2. Chemical diagrams and structure overlay (benzene rings) of DFT-optimized molecular structures of 4a and 4b (orange) and the BTO constitutional isomers $6a^{32}$ and 6b (green).³³ Hydrogen atoms are omitted for clarity.



Figure 5. Asymmetric unit of the crystal structure of **5b**. Displacement ellipsoids are drawn at the 50% probability level. Color scheme: C, gray; H, white; N, blue; O, red; S, yellow.

isothiazol-3-one nitrogen atom likely explain the observed regioselectivity.

The structures of the reaction products **4a** and **4b** as well as isomeric **5a** and **5b** were unambiguously assigned by X-ray crystallography. Compound **4a** crystallized as a 0.25 hydrate with three crystallographically unique molecules.³¹ As shown in Figure 1a, the three distinct molecules all adopt a *Z* conformation about the C_{acyl} -N2_{BIT} bond. The appended piperidine ring adopts a chair conformation in each case (Figure 1b).

Table 1. In Vitro Activity (MIC₉₀ in μ M) of 4a, 4b, 5a, and 5b and the Two Reference BTZs 1a and 1b against *M. smegmatis* mc² 155, *M. aurum* DSM 43999 and *M. tuberculosis* H37Rv

	M. smegmatis mc ² 155 ^a	M. aurum DSM 43999 ^b	M. tuberculosis H37Rv ^b
1a	6.25	25-12.5	>4.0
1b (BTZ043)	0.025	0.008	<0.06
4a	0.20	0.10	1.0
4b	0.32	0.25-0.13	0.5
5a	>100	>100	>32
5b	>100	>100	>32
^{<i>a</i>} 7H9 medium with OADC.	n supplemented witl	n ADS. ^{<i>b</i>} 7H9 medi	um supplemented

The solvate water forms a hydrogen bond to the piperidinoyl oxygen atom of molecule 1 (Figure 1c). Figure 2 depicts overlay diagrams of DFT-calculated molecular structures of 4a and 4b and the corresponding constitutionally isomeric benzothiazole *N*-oxides (BTOs, 6), potent antimycobacterial agents described previously in the literature (vide infra),^{32,33} which reveals that the molecules are isosteric.

Compound 4b crystallizes with two molecules in the asymmetric unit (Figure 3).³⁴ Like in 4a, the conformation



Figure 6. Aliphatic region of the 13 C NMR spectra of 4a, 5a, 4b, and 5b in CDCl₃ at room temperature (the full spectra are shown in the Supporting Information).

Table 2. Physicochemical Properties of 4a, 4b, 5a, and 5b, the Two Reference BTZs 1a and 1b, and the precursor 3

	solubility $(\mu M)^a$	$c \log P^b$
la	85	3.39
1b (BTZ043)	74	2.45
3	444	1.23
4a	92	4.14
4b	103	3.47
5a	60	3.75
5b	39	2.81

^{*a*}Determined by nephelometry in PBS (pH 7.4) with 2% (v/v) DMSO.⁴⁴ b Calculated with *Chem3D Ultra* version 20.1.1.125.



Figure 7. Highest ranked molecular covalent docking solution of 4a to *M. tuberculosis* DprE1.

about the C_{acyl} -N2_{BIT} bond is Z in both unique molecules. The two crystallographically distinct molecules are thus diastereomeric conformers. Such was also encountered in the crystal structure of **1b** (BTZ043).³⁵

Figures 4 and 5 depict the molecular structures of 5a and 5b, respectively, in the crystal.^{36,37} The C3–N2 bond length of the benzothiazole heterocyclic system of 1.298(1) Å is significantly shorter by ca. 0.4 Å than that in 4a and 4b, consistent with C=N double bond character in 5a and 5b. The opposite enantiomeric conformer is also present in the centrosymmetric crystal structure. As for $1b^{35}$ and 4b, the crystal structure of 5b comprises two crystallographically unique diastereomeric conformers.

Compounds 4 and 5 were also structurally characterized by ¹H and ¹³C NMR spectroscopy in solution at room temperature (see the Supporting Information). Figure 6 shows the aliphatic regions of the ¹³C NMR spectra in CDCl₃, which exhibit the most marked differences for the compounds studied. The resonance signals in the aliphatic regions were assigned based on chemical shifts. With three signals, 4a shows the simplest spectrum in this region. The signals are assigned to the piperidine methylene groups. The occurrence of one set of signals for the α - and β -methylene groups indicates that rotation about the urea $C_{carbonyl} - N_{piperidine}$ bond is fast. The same was observed for BTZ 1a.² In contrast, two signals are encountered for each of the α - and β -methylene groups in the spectrum of 5a, which can the explained by restricted rotation of the piperidine ring around the C_{carbonyl}-N_{piperidine} bond. For 4b and 5b, the most upfield shifted signal is assigned to the methyl group of the methyldioxolan moiety. For 4b, two signals appear for the piperidine β -methylene pubs.acs.org/acsmedchemlett

groups, whereas the α -methylene carbon atoms are virtually isochronous, suggesting that there is no restricted rotation of the side chain around the C_{carbonyl}-N_{piperidine} bond. For BTZ 1b, variable-temperature ¹³C NMR spectroscopy revealed a coalescence temperature T_c of 301 \pm 1 K and a Gibbs free energy ΔG^{\ddagger} of 14.6 \pm 1 kcal mol⁻¹ for this process.³⁵ The ¹³C NMR spectrum of **5b** displays four discrete signals for each of the α - and β -piperidine methylene groups, which can be ascribed to diastereomeric conformers, as revealed by X-ray crystallography (Figure 5), resulting from restricted rotation of the side chain around both the carbamate $C_{carbonyl} - N_{piperidine}$ and C_{BIT} -O_{BIT} bonds. Restricted rotation around the C_{BIT} -O_{BIT} bond in 5a due to steric clashes between the carbamate group and benzisothiazolol moiety should result in stable enantiomeric conformers, which cannot be distinguished in the NMR spectrum. As shown in Figure 4, both enantiomeric conformers of 5a are present in the solid state.

Antimycobacterial activities of 4a, 4b, 5a, and 5b and BTZs 1a and 1b as reference compounds were determined in vitro against Mycobacterium smegmatis, M. aurum, and M. tuberculosis (Table 1). M. smegmatis and M. tuberculosis are susceptible to irreversible inhibition of DprE1 through BTZs,^{10,39} because of the presence of a cysteine residue in the active site (vide supra). M. smegmatis is a fast-growing mycobacterium, generally considered nonpathogenic, and a useful model organism for *M. tuberculosis.*⁴⁰ We found that the *N*-acyl BITs 4a and 4b with the side chain appended to the BIT nitrogen atom exert activity against the three mycobacterial strains. Interestingly, 4a is more active than BTZ 1a bearing the same piperidine side chain. A similar trend, however, is not observed for 4b and the corresponding BTZ 1b. The in vitro activity of 4b bearing the BTZ043-like side chain does not reach that of the 1b but is similar to that of 4a. The MIC values against the three mycobacterial strains are still some orders of magnitude higher for 4b than for 1b. The isomeric Oacyl benzisothiazol-3-ol derivatives 5a and 5b do not inhibit mycobacterial growth up to the concentrations tested. We propose that the carbamate group attached to the BIT oxygen atom (O3) and resulting altered hydrogen bond preferences account for the inactivity of 5a and 5b (vide infra). The BIT precursor 3, lacking a side chain, was tested in vitro against M. smegmatis and found to be inactive (MIC₉₀ > 100 μ M) as expected. The in vitro test results for 3, 5a, and 5b confirm that not only an electron-deficient nitroaromatic warhead but also the nature and spatial orientation of the side chain are critical for efficient mechanism-based mycobacterial growth inhibition. Kinetic solubilities of the compounds studied in PBS are essentially comparable with each other and with the two reference BTZs, and the c log P values are lower for the compounds bearing the BTZ043-like spiro ketal side chain than for those with an appended piperidine ring (Table 2). The c log P values of the active compounds 4a and 4b are higher than those of the corresponding inactive constitutional isomers 5a and 5b but do not exceed a value of 5, which is desirable for drug-like compounds.⁴¹

These results raise the question of how the antimycobacterial properties of **4a** and **4b** compare with those of the structurally most related compounds. As reported previously, **4a** and **4b** exhibited higher in vitro activity against *M. tuberculosis* and *M. aurum* than the corresponding BIT 1-oxides.²⁵ For the corresponding isosteric BTO constitutional isomers (vide supra), in vitro activity data against *M. tuberculosis* H37Rv can be found in the literature: 0.25 μ g



Figure 8. Highest ranked molecular docking solutions for (A) **4a**, (B) **4b**, (C) **5a**, and (D) **5b** (docked to 6HEZ.B; flexible side chains for Lys134, Trp230, Val365, Leu363, Lys418; toggled water molecules; solution number as in Table S2).

mL $^{-1}$ (0.67 $\mu M)$ for $6a^{32}$ (medium unknown) and 0.24 μM (7H12 medium) and 0.23 μ M (GAS medium) for **6b**.³³ Thus, 4a/4b and 6a/6b exhibit comparable in vitro activities against M. tuberculosis. Comparing the molecular electrostatic potential (MEP) mapped onto the van der Waals surface for 4a/4b versus 6a/6b reveals a striking similarity of the isosteric molecules (see the Supporting Information). The comparable antimycobacterial properties are consistent with these findings. The authors of 6a also reported the crystal structure of a related BTO (bearing a different side chain) in complex with M. tuberculosis DprE1 (PDB ID: 4PFA),³² which revealed a semimercaptal adduct formed between the nitroso form of the ligand and the Cys387 residue like for BTZs (vide supra). The authors of 6b proved by metabolic labeling of M. tuberculosis DprE1 and in vitro testing against M. aurum that the target for this BTO compound is DprE1, based on the assumption that M. aurum lacks a cysteine residue in the DprE1 active site⁴² U. Möllmann, M. Miller, personal communication: In ref 33, the M. aurum strain SB66 from the collection of the Leibniz Institute for Natural Product Research and Infection Biology -Hans Knöll Institute – (HKI, Jena, Germany) was used.].³³

As shown in Table 2 below, the active BIT derivatives 4a and 4b have a similar low solubility as lead compound BTZ043 ($\leq 100 \ \mu$ M), whereas the c log *P* values are slightly higher.

In the present work, we observed that BITs 4a/4b and BTZs 1a/1b inhibit growth of *M. aurum* DSM 43999, which is consistent with our previously reported results for 1a and 4a.²⁵ In their report on the purported "BTZ-SO",²⁴ whose structure we revised to 4b (vide supra), Tiwari et al. described growth inhibition of *M. aurum* SB66 by "BTZ-SO", but not by 1b

(BTZ043). This also could be due to DprE1 sequence differences between the two *M. aurum* strains. We should also note that BITs are known to have various molecular targets in microorganisms.⁴³

Molecular covalent docking of the newly synthesized compounds 4a, 4b, 5a, and 5b to M. tuberculosis DprE1 was performed to gain insight into possible binding modes. The docking pose of 1b corresponds to the orientation encountered in the crystal structure in complex with the M. tuberculosis DprE1 (PDB ID: 6HEZ)¹⁵ (see the Supporting Information). The binding mode of the active compounds 4a and 4b shows a comparable orientation in the active site for all highly evaluated docking poses. Therefore, it is reasonable to assume that the secondary interactions can be formed similarly to 1b. In comparison to 4b, the position of 4a changes slightly. This can be attributed to the fact that the bridged hydrogen bond, formed to FAD, which is apparent in the crystal structure of 1b in complex with DprE1 (chain B), is observed in docked solutions of 4b but not 4a. Stabilizing effects are, however, noticeable via hydrogen bonds to Lys134 and His315 for this compound. In particular, the possible formation of a hydrogen bond between the amino group of the Lys134 side chain and O3 of 4a is striking (Figure 7).

The results for the inactive **5a** and **5b** are quite different. For these molecules, none of the in silico methods could provide a placement similar to that described above. All docking solutions for **5a** and **5b** resulted in a pose of the molecules rotated by nearly 180° . This is likely due to the fundamental change in the molecular structure with regard to the linkage of the piperidinoyl side chain to the molecular scaffold via the benzothiazole oxygen atom O3, which subsequently leads to steric hindrance in the DprE1 active site. In addition, the hydrogen bond observed in the crystal structure of 1b in complex with DprE1 can no longer be formed. All these indications rationalize the inactivity of 5a and 5b, which is confirmed by the total fitness scores of active 4a and 4b and inactive 5a and 5b (Table S1). The highest ranked solutions are shown in Figure 8.

In summary, the N-acyl BITs 4 and O-acyl benzisothiazolols 5 were synthesized and their structures investigated by X-ray crystallography and NMR spectroscopy. The N-acyl BITs 4 exhibited only slightly poorer activity against M. smegmatis mc^2 155a, M. aurum DSM 43999b and M. tuberculosis H37Rv than 1b (BTZ043) and significantly better activity than 1a, whereas 5 showed little or no antimycobacterial activity. Notably, 4a is more active by an order of magnitude than BTZ 1a bearing the same piperidine side chain. The in vitro test results for the BIT precursor 3 lacking a side chain, 5a and 5b confirm that not only an electron-deficient nitroaromatic warhead but also the nature and spatial orientation of the side group are critical for efficient mechanism-based mycobacterial growth inhibition. The inability of 5 to inhibit M. tuberculosis DprE1 was rationalized using in silico docking experiments on 4 and 5, which attributed this to the linkage of the piperidinoyl side chain to the molecular scaffold via the benzothiazole oxygen atom O3.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acsmedchemlett.2c00215.

Experimental and computational details along with characterizations of the compounds synthesized (PDF) DFT-calculated structure of 4a (XYZ)

DF1-calculated structure of 4a (A12

DFT-calculated structure of 4b (XYZ)

DFT-calculated structure of 5a (XYZ)

DFT-calculated structure of **5b** (XYZ)

DFT-calculated structure of **6a** (XYZ)

DFT-calculated structure of **6b** (XYZ)

Accession Codes

CCDC 2171377–2171380 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/structures.

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DEDICATION

Dedicated to Professor Bodo Dobner on the occasion of his 70th birthday.

ABBREVIATIONS

BTZ, 8-nitro-1,3-benzothiazin-4-one; BIT, benzisothiazolinone; BTO, benzothiazole *N*-oxides; DprE1, decaprenylphosphoryl-β-d-ribose 2'-epimerase; *m*CPBA, *m*-chloroperoxybenzoic acid; MEP, molecular electrostatic potential

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(31) Crystal data for **4a**: $C_{14}H_{12.50}F_3N_3O_{4.25}S$, $M_r = 379.83$, T = 100(2) K, $\lambda = 1.54178$ Å, triclinic, space group PI, a = 10.0915(4), b = 14.5439(5), c = 17.3813(6) Å, $\beta = 78.721(2)$, $\gamma = 80.711(2)^{\circ}$, V = 2377.82(15) Å³, Z = 6, $\rho_{calc} = 1.592$ mg m⁻³, $\mu = 2.408$ mm⁻¹, F(000) = 1167, crystal size $0.133 \times 0.073 \times 0.043$ mm³, θ range = 2.69–72.00°, reflections collected/unique = 88503/8735 ($R_{int} = 0.0834$), 750 parameters, 237 restraints, S = 1.073, R_1 [$I > 2\sigma(I)$] = 0.0420, $wR_2 = 0.1499$, $\Delta \rho_{max}/\Delta \rho_{min} = 0.43/-0.89$ e Å⁻³.

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(34) Crystal data for 4b: $C_{17}H_{16}F_3N_3O_6S$, $M_r = 447.39$, T = 100(2)K, $\lambda = 1.54178$ Å, monoclinic, space group $P2_1$, a = 13.4977(4), b = 9.0737(3), c = 15.2780(4) Å, $\beta = 90.489(2)^\circ$, V = 1871.09(10) Å³, Z = 4, $\rho_{calc} = 1.588$ mg m⁻³, $\mu = 2.212$ mm⁻¹, F(000) = 920, crystal size 0.161 × 0.062 × 0.051 mm³, θ range = 2.89–71.99°, reflections collected/unique = 58657/7075 ($R_{int} = 0.0634$), 600 parameters, 235 restraints, S = 1.099, R_1 [$I > 2\sigma(I)$] = 0.0485, $wR_2 = 0.1309$, $\Delta\rho_{max}/\Delta\rho_{min} = 0.37/-0.29$ e Å⁻³.

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(36) Crystal data for **5a**: $C_{14}H_{12}F_3N_3O_4S$, $M_r = 375.33$, T = 100(2)K, $\lambda = 0.71073$ Å, orthorhombic, space group *Pbca*, a = 16.8517(10), b = 7.6877(5), c = 23.4052(14) Å, V = 3032.2(3) Å³, Z = 8, $\rho_{calc} = 1.644$ mg m⁻³, $\mu = 0.275$ mm⁻¹, F(000) = 1538.080, crystal size 0.094 × 0.034 × 0.031 mm³, θ range = $1.74-31.75^{\circ}$, reflections collected/ unique = 96114/5114 ($R_{int} = 0.0535$), 274 parameters, S = 1.0508, R_1 [$I > 2\sigma(I)$] = 0.0241, $wR_2 = 0.0466$, $\Delta \rho_{max}/\Delta \rho_{min} = 0.46/-0.44$ e Å⁻³. (37) Crystal data for **5b**: $C_{17}H_{16}F_3N_3O_6S$, $M_r = 447.39$, T = 100(2)K, $\lambda = 0.71073$ Å, monoclinic, space group $P2_1$, a = 7.2884(7), b = 17.5581(16), c = 14.3508(13) Å, $\beta = 98.454(4)^{\circ}$, V = 1816.5(3) Å³, Z = 4, $\rho_{calc} = 1.636$ mg m⁻³, $\mu = 0.252$ mm⁻¹, F(000) = 920, crystal size 0.130 × 0.051 × 0.041 mm³, θ range = $1.44-48.67^{\circ}$, reflections collected/unique = 434272/34730 ($R_{int} = 0.0734$), 543 parameters, 1 restraint, S = 1.008, R_1 [$I > 2\sigma(I)$] = 0.0473, $wR_2 = 0.1191$, $\Delta \rho_{max}/\Delta \rho_{min} = 0.57/-0.69$ e Å⁻³.

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