# Multianalyte Prognostic Signature Including Circulating Tumor DNA and Circulating Tumor Cells in Patients With Advanced Pancreatic Adenocarcinoma

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**PURPOSE** Pancreatic ductal adenocarcinoma (PDAC) is associated with a poor prognosis. Multianalyte signatures, including liquid biopsy and traditional clinical variables, have shown promise for improving prognostication in other solid tumors but have not yet been rigorously assessed for PDAC.

**MATERIALS AND METHODS** We performed a prospective cohort study of patients with newly diagnosed locally advanced pancreatic cancer (LAPC) or metastatic PDAC (mPDAC) who were planned to undergo systemic therapy. We collected peripheral blood before systemic therapy and assessed circulating tumor cells (CTCs), cell-free DNA concentration (cfDNA), and circulating tumor *KRAS* (ctKRAS)–variant allele fraction (VAF). Association of variables with overall survival (OS) was assessed in univariate and multivariate survival analysis, and comparisons were made between models containing liquid biopsy variables combined with traditional clinical prognostic variables versus models containing traditional clinical prognostic variables alone.

**RESULTS** One hundred four patients, 40 with LAPC and 64 with mPDAC, were enrolled. CTCs, cfDNA concentration, and ctKRAS VAF were all significantly higher in patients with mPDAC than patients with LAPC. ctKRAS VAF (cube root; 0.05 unit increments; hazard ratio, 1.11; 95% CI, 1.03 to 1.21; P = .01), and CTCs  $\geq$  1/mL (hazard ratio, 2.22; 95% CI, 1.34 to 3.69; P = .002) were significantly associated with worse OS in multivariate analysis while cfDNA concentration was not. A model selected by backward selection containing traditional clinical variables plus liquid biopsy variables had better discrimination of OS compared with a model containing traditional clinical variables alone (optimism-corrected Harrell's C-statistic 0.725 v 0.681).

**CONCLUSION** A multianalyte prognostic signature containing CTCs, ctKRAS, and cfDNA concentration outperformed a model containing traditional clinical variables alone suggesting that CTCs, ctKRAS, and cfDNA provide prognostic information complementary to traditional clinical variables in advanced PDAC.

JCO Precis Oncol 6:e2200060. © 2022 by American Society of Clinical Oncology

# **INTRODUCTION**

Pancreatic ductal adenocarcinoma (PDAC) is the fourth leading cause of cancer death in the United States with only modest improvements in overall survival (OS) in recent decades.<sup>1,2</sup> Clinical characteristics most consistently associated with OS include performance status, disease status (locally advanced pancreatic cancer [LAPC] *v* metastatic PDAC [mPDAC]), sex, age, liver metastasis, albumin, cancer antigen (CA)19-9, number of metastatic sites, pain, bilirubin, and lactate dehydrogenase,<sup>3</sup> although these characteristics are imperfect. There is a great need to identify other noninvasive strategies to more accurately risk stratify patients with PDAC for clinical research, clinical care, and patient counseling. Tissue biomarkers are challenging to obtain as biopsies are often hypocellular,

and the majority of patients will not be candidates for surgical resection. Conversely, blood samples are easily obtainable over the course of a patient's therapy.

Cell-free DNA (cfDNA) is shed by cells into the blood while circulating tumor DNA (ctDNA) represents DNA specifically shed by tumor.<sup>4</sup> In PDAC, ctDNA is most commonly assessed by the presence or absence of *KRAS* mutations using polymerase chain reaction or next-generation sequencing as *KRAS* has been shown to be mutated in > 90% of PDAC cases.<sup>5</sup> Rates of circulating tumor *KRAS* mutation (ctKRAS) detection in the peripheral blood range from 21% to 60% in patients with localized PDAC and 42%-72% in patients with mPDAC.<sup>6</sup> Detection of ctKRAS at baseline is consistently associated with worse OS in patients with PDAC of all stages.<sup>7-14</sup> Although this suggests that

# ASSOCIATED Content

# Data Supplement

Author affiliations and support information (if applicable) appear at the end of this article.

Accepted on June 15, 2022 and published at ascopubs.org/journal/ po on August 8, 2022: D0I https://doi.org/10. 1200/P0.22.00060



# CONTEXT

## **Key Objective**

Pancreatic adenocarcinoma has poor prognosis. Multianalyte signatures using liquid biopsy variables have shown promise for improving prognostication in patients with other solid tumors but have not been rigorously evaluated to date in patients with pancreatic adenocarcinoma.

#### **Knowledge Generated**

Circulating tumor cells and circulating tumor *KRAS*-variant allele fraction were significantly associated with overall survival in univariable analysis and in multivariable analysis with adjustment for important traditional prognostic variables. Prognostic models containing liquid biopsy variables and traditional clinical variables outperformed models containing traditional clinical variables alone, suggesting that liquid biopsy variables provide complementary information.

### Relevance

Multiple liquid biopsy analytes, including circulating tumor *KRAS* variant allele fraction and circulating tumor cells, may be applied in future clinical research to improve prognostic and predictive models in patients with pancreatic adenocarcinoma.

ctKRAS alone may be a sufficient prognostic indicator in PDAC, it would be of great interest to know if the addition of other liquid biopsy analytes, easily obtainable from the same tube of blood, could improve prognostication compared with ctKRAS alone.

Circulating tumor cells (CTCs) are another class of liquid biopsy analytes. In patients with PDAC, rates of CTC detection using various platforms-including antibodymediated enrichment (ie, CellSearch), methods using size, physical properties, and immunohistochemistry (ie, isolation by size of epithelial tumor cells, or ISET), and microfluidics approaches<sup>15-17</sup>—have varied from 5% to 89%.<sup>18</sup> The presence of CTCs is associated with significantly worse OS in patients with PDAC.<sup>18,19</sup> A more recent nonenrichment CTC detection method works by depositing all nucleated cells from a tube of blood onto glass microscope slides and identifies CTCs and other rare cells through immunofluorescence and digital pathology. Although this technology has been clinically qualified in prostate, breast, and lung cancer, this technology has not been evaluated in PDAC.<sup>20,21</sup>

There is evidence in multiple cancers that multianalyte approaches can provide complementary information compared with that of individual analytes alone. In breast cancer, a multianalyte liquid biopsy signature including CTC mRNA, CTC genomic DNA, extracellular vesicle mRNA, and cfDNA improved prognostication of OS compared with individual analytes, and clustering analysis suggested that liquid biopsy analytes were complementary and offered significant nonoverlapping information.<sup>22</sup> In PDAC, a multianalyte panel of tumor-associated extracellular vesicle microRNA and mRNA, cfDNA, ctDNA KRAS mutations, and CA19-9 showed promise for improving diagnosis of PDAC and detecting occult metastases, although these variables were not assessed for prognostic value.<sup>24</sup> A study examining baseline ctDNA and exosome DNA in PDAC found that elevation of both markers was a

stronger predictor of worse OS compared with their elevation individually.<sup>25</sup>

Assessing any multianalyte panel in combination with clinical prognostic variables compared with clinical prognostic variables alone is an important step to establish the potential benefit in clinical practice. To our knowledge, the prognostic value of a multianalyte liquid biopsy in patients with advanced PDAC has not been assessed. We hypothesized that a multianalyte prognostic model incorporating liquid biopsy variables with traditional clinical variables would improve discrimination of prognosis in patients with LAPC and mPDAC compared with a model containing traditional clinical variables alone. Additionally, we aimed to evaluate for an independent association of cfDNA, ctKRAS, and CTCs with OS after adjusting for a more robust panel of clinical covariates than has previously been performed.

#### **MATERIALS AND METHODS**

#### Patient Enrollment

Patients with LAPC or mPDAC were screened at presentation to the medical oncology clinic at the University of Pennsylvania after diagnosis. Patients were excluded for having a concurrent cancer or for receiving prior systemic therapy for advanced PDAC. This study was reviewed by the Institutional Review Board at the University of Pennsylvania. Patients provided written informed consent to participate. Systemic treatment decisions were made by clinicians and patients as part of routine clinical care and were unrelated to the present study. Sample size was a convenience sample on the basis of budget for CTC analysis.

## **CTC** Isolation and Identification

Peripheral blood (approximately 10 mL per sample) was collected at the time of patient enrollment and shipped to Epic Sciences (San Diego, CA) for processing. Red blood

cells were lysed, and nucleated cells ( $3 \times 10^6$  per slide) were deposited onto 12 microscope slides. Two replicate slides per sample were thawed and stained with antibodies for pan-cytokeratins (CK), CD45, and 4',6-diamidino-2-phenylindole (DAPI). Slides then underwent a rapid fluorescent scanning method and CTCs identified in silico using proprietary digital pathology algorithms.<sup>26-28</sup> The resulting images were displayed in a web-based report and confirmed by two trained technicians. Next, CTC enumeration results were compiled and reported as CTCs/mL of blood. The category of interest for the present study was CK+/CD45– cells with an intact nucleus (DAPI+) plus CK+ clusters, counted as one event, defined as two or more adjacent CD45– cells with shared cytoplasmic boundaries, at least one of which was CK+.<sup>27</sup>

# cfDNA Isolation, Quantification, and Detection of ctKRAS Mutation

Isolation and quantification of cfDNA and ctKRAS detection by preamplification droplet digital polymerase chain reaction were performed as previously described (Data Supplement).<sup>24</sup> Samples below the assay level of detection (0.01% variant allele fraction [VAF])<sup>24</sup> were treated as zero values.

# Assessment of Traditional Clinical Variables

Baseline clinical variables (Table 1) were obtained by chart review. Laboratory variables were considered to be baseline values if collected up to 28 days before enrollment and before first systemic therapy.

# **Statistical Methods**

Descriptive statistics. Descriptive statistics were computed as mean and standard deviation for continuous variables and as a proportion of patients for binary or categorical variables. The proportion of missing observations for each variable was reported. To compare values of liquid biopsy variables between LAPC and mPDAC groups, two-tailed t tests or Wilcoxon rank-sum tests were performed (depending on normality of distribution of continuous variables). Pearson's chi-square test or Fisher's exact test was performed to compare categorical variables between LAPC and mPDAC. Fisher's exact test was used to examine the association of CTCs (binary; cut point 1/mL) with ctKRAS (binary; cut point VAF 0.05; rationale described below) across the overall cohort, and linear regression modeling was used to assess the association of CTCs/mL (continuous) with ctKRAS VAF (continuous). Statistical testing for all analyses used an  $\alpha$  of .05.

**Univariate survival analysis.** Univariate and multivariate survival analysis was performed in R (version 3.6.3) using the *rms* and *survival* packages.<sup>29-31</sup> OS was defined as the time from study enrollment to death as assessed by clinical records and online obituaries, with censoring at last clinical contact or data cutoff of January 14, 2021. The functional

form and transformations of variables used in Cox proportional hazards models are described in the Data Supplement.

Univariate Cox proportional hazards modeling was performed to examine the association of liquid biopsy variables and traditional clinical variables with OS in the overall cohort and within the LAPC and mPDAC cohorts individually. Next, treating ctKRAS VAF and cfDNA as binary variables was explored by examining the spline transformation of each variable in the log relative hazard and selecting a natural cut point. This binary treatment of liquid biopsy variables was used to plot Kaplan-Meier survival curves by levels of the liquid biopsy variable, followed by log-rank testing. Using the binary cutoffs chosen above, an exploratory analysis was performed to examine OS for patients who were dual positive for CTCs and ctKRAS, single positive, or dual negative using the Kaplan-Meier method, followed by log-rank testing.

*Multivariate Cox proportional hazards model selection.* Candidate variables for the multivariable prognostic models included variables with a P < .15 in univariate analysis. Our goals were (1) to assess the relationship of liquid biopsy variables with OS after adjusting for clinical covariates and (2) to compare models containing traditional clinical plus liquid biopsy variables to models containing traditional clinical clinical containing traditional clinical variables alone for model fit and discrimination.

We accomplished these goals with two model selection strategies. First, we included all candidate variables (both liquid biopsy and traditional clinical variables) with a P < .15 in univariate analysis (model A) and compared this with a model that contained only traditional clinical variables that passed the P < .15 threshold in univariate analysis (model B). Second, we performed backward selection by minimizing Akaike's information criterion using the stepAIC function under the *MASS* package in R.<sup>32</sup> We performed this backward selection procedure using all candidate variables with P < .15 in univariate analysis (model C) and ran a second model using this backward selection procedure using traditional clinical variables with P < .15 in univariate analysis (model C) and ran a second model using this backward selection procedure using traditional clinical variables with P < .15 in univariate analysis alone (model D).

**Evaluation of multivariate Cox proportional hazards models.** Evaluation of model assumptions and model fit is described in the Data Supplement. Model discrimination was assessed using Harrel's C-statistic.<sup>33</sup> Internal validation with bootstrapping to calculate optimism-corrected Harrel's C-statistics and optimism-corrected calibration curves are further described in the Data Supplement. Sensitivity analysis examining the impact of missing data on the effect estimates in each model is described and presented in the Data Supplement.

# RESULTS

# Patient Characteristics and Summary Statistics

Forty patients with LAPC and 64 patients with mPDAC were enrolled (104 patients in total; Table 1) between August 14,

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# TABLE 1. Patient Characteristics in the Locally Advanced and Metastatic Cohorts

Clarized mature         Marine (m = 16.4)	Characteristic	0vorall (N = 104)	Locally Advanced $(n - 40)$	Motostatia (n - 64)	P	
sake, No. (%)           Female         42 (40.4)         16 (40.0)         26 (40.6)           Male         62 (59.6)         24 (60.0)         38 (59.4)           Age, yards           57 (69.1)           White         89 (65.6)         32 (80.0)         57 (69.1)           Back         9 (87.)         7 (17.5)         2 (3.1)           Assan or others         6 (5.8)         1 (2.5)         5 (7.8)           BMI           46.0 (0.0)         1 (1.6)           Datetes mellitus, No. (%)         1 (1.0)         0 (0)         1 (1.6)           Datetes mellitus, No. (%)         1 (1.0)         1 (2.5)         0 (0)           Mean (SD)         3.8100 (0.4520)         3.9100 (0.3300)         3.7550 (0.5050)           OCOG PS, No. (%)         1 (1.0)         1 (2.5)         0 (0)           ECOG O-1         84 (80.8)         35 (87.5)         49 (76.6)           ECOG S 2         14 (13.5)         3 (7.5)         11 (17.2)           Messing, No. (%)         1 (1.0)         0 (0)         1 (1.6)           Levr metalsas, No. (%)         1 (1.0)         0 (0)         1 (1.6)           Levr metalsas, No. (%)         1 (1.0)         0 (0)		Overall (N = 104)	Auvaliceu (li = 40)	Wetastatic (II = O4)	r	
Termane         42 (40.4)         16 (40.0)         26 (60.0)           Male         62 (99.6)         24 (60.0)         38 (99.4)           Age, years	Sex, No. (%)	40 (40 4)	16 (40.0)	00 (40 0)		
Mile         62 (39.6)         24 (60.0)         38 (39.4)           Mean (SD)         65.5000 (9.5400)         65.1000 (9.0100)         65.7000 (9.9100)           Race, No. (%)         While         89 (95.6)         32 (90.0)         57 (89.1)           Black         9 (8.7)         7 (17.5)         2 (3.1)         Asian or others         6 (5.8)         1 (2.5)         5 (7.8)           BM         Mean (SD)         27.1000 (5.7200)         26.3000 (6.0500)         27.6000 (6.0900)           Means, No. (%)         1 (1.0)         0 (0)         1 (1.6)         1 (1.6)           Dabates melitus, No. (%)         1 (1.0)         1 (2.5)         0 (0)         1 (2.5)           Albumin, g(d)         1 (1.0)         1 (2.5)         0 (0)         1 (2.5)         0 (0)           ECOG 0 1         84 (90.8)         35 (87.5)         49 (75.6)         1 (2.5)         0 (0)           ECOG 5 2         14 (13.5)         3 (7.5)         11 (17.2)         Mean (min, max)         1,110 (3.00, 468,000)         23 (3.00, 7,830)         2,710 (5.00, 468,000)           Mesing, No. (%)         1 (1.0)         0 (0)         1 (1.6)         1 (1.6)         1 (1.6)           Mesing, No. (%)         1 (1.0)         0 (0)         5 (79.7)         1	Female	42 (40.4)	16 (40.0)	26 (40.6)		
Age, gets           Mean (SD)         65.5000 (9.5400)         65.1000 (9.0100)         65.7000 (9.9100)           Race, No. (%)         9 (8.7)         7 (17.5)         2 (3.1)           Black         9 (8.7)         7 (17.5)         2 (3.1)           Asim or others         6 (5.8)         1 (2.5)         5 (7.8)           BMI         9 (8.7)         7 (17.5)         2 (3.1)           Mean (SD)         27.1000 (5.7200)         26.3000 (5.0500)         27.6000 (6.0900)           Missing, No. (%)         1 (1.0)         0 (0)         1 (1.6)           Dabetes melitus, No. (%)         1 (1.0)         1 (2.5)         0 (0)           Mean (SD)         3.8100 (0.4520)         3.9100 (0.3300)         3.7500 (0.5050)           Missing, No. (%)         1 (1.0)         1 (2.5)         0 (0)           ECOG 6.1         84 (40.8)         35 (87.5)         49 (76.6)           ECOG 5.2         14 (13.5)         3 (7.5)         11 (17.2)           Missing, No. (%)         1 (1.0)         0 (0)         1 (1.6)           Live metastasis, No. (%)         1 (1.0)         0 (0,0)         1 (79.7)           Initial treatment, No. (%)         1 (1.0)         0 (0,0)         51 (79.7)           Gemottabui		62 (59.6)	24 (60.0)	38 (59.4)		
Mean (SD)         65.300 (9.940)         65.000 (9.0100)         65.000 (9.9100)           White         89 (85.6)         32 (80.0)         57 (89.1)           Black         9 (8.7)         7 (17.5)         2 (3.1)           Asian or others         6 (5.8)         1 (2.5)         5 (7.8)           BMI         Mean (SD)         27.1000 (5.7200)         26.3000 (5.6500)         27.6000 (6.0900)           Missing, No. (%)         1 (1.0)         0 (0)         1 (1.6)           Diabetes melitus, No. (%)         1 (1.0)         0 (0)         24 (37.5)           Aburnin, gdt.	Age, years		CE 1000 (0.0100)	CE 7000 (0.0100)		
Race, No. (%)         White         89 (85.6)         32 (80.0)         57 (89.1)           Black         9 (8.7)         7 (17.5)         2 (3.1)           Asian or others         6 (5.8)         1 (2.5)         5 (7.8)           BMI	Mean (SD)	65.5000 (9.5400)	65.1000 (9.0100)	65./000 (9.9100)		
Write         69 (65,6)         32 (80,0)         57 (17,5)         2 (3,1)           Asian or others         6 (5.8)         1 (2,5)         5 (7,8)           BM         27,1000 (5,7200)         26,3000 (5,0500)         27,6000 (6,0900)           Mean (SD)         27,1000 (5,7200)         26,3000 (5,0500)         27,6000 (6,0900)           Missing, No. (%)         1 (1,0)         0 (0)         1 (1,6)           Diabetes melitus, No. (%)         3,8100 (0,4520)         3,9100 (0,3300)         3,7500 (0,5050)           Mean (SD)         3,8100 (0,4520)         3,9100 (0,3300)         3,7500 (0,5050)           Missing, No. (%)         1 (1,0)         1 (2,5)         0 (0)           ECOG 0,1         84 (80,8)         35 (87,5)         49 (76,6)           ECOG 0,1         84 (80,8)         35 (87,5)         49 (76,6)           ECOG 0,1         84 (80,8)         35 (87,5)         41 (1,7,2)           Missing, No. (%)         1 (1,0)         0 (0)         1 (1,1)           Missing, No. (%)         1 (1,0)         0 (0)         1 (1,6)           Liver metastasis, No. (%)         1 (1,0)         0 (0,0)         51 (79,7)           Initial treatment, No. (%)         9 (8,7)         1 (2,5)         8 (12,5)	Race, No. (%)	00 (05 0)	20 (00 0)	57 (00.1)		
Biack         9 (8,7)         7 (1,7.5)         2 (3,1)           Asian or others         6 (5,8)         1 (2.5)         5 (7.8)           BMI         Mean (SD)         27.1000 (5.7200)         26.3000 (5.0500)         27.6000 (6.0900)           Mean (SD)         27.1000 (5.7200)         26.3000 (5.0500)         27.6000 (6.0900)           Diabetes malitus, No. (%)         1 (1.0)         0 (0)         24 (37.5)           Alburnin, g/dL	White	89 (85.6)	32 (80.0)	57 (89.1)		
Asin or others         6 (5.8)         1 (2.5)         5 (7.8)           BMI <td>Black</td> <td>9 (8.7)</td> <td>/ (17.5)</td> <td>2 (3.1)</td> <td></td>	Black	9 (8.7)	/ (17.5)	2 (3.1)		
BMI Mean (SD) 27.1000 (5.7200) 26.0000 (5.0500) 27.6000 (6.0900) Missing, No. (%) 1 (1.0) 0 (0) 1 (1.6) Diabetes mellitus, No. (%) Yes 42 (40.4) 18 (45.0) 24 (37.5) Albumin, g/dl. Mean (SD) 3.8100 (0.4520) 3.9100 (0.3300) 3.7500 (0.5050) Missing, No. (%) 1 (1.0) 1 (2.5) 0 (0) ECOG PS, No. (%) ECOG 0-1 84 (80.8) 35 (87.5) 49 (76.6) ECOG 0-2 14 (13.5) 3 (7.5) 11 (17.2) Missing 6 (58.8) 2 (5.0) 4 (6.2) CA19-9, U/mL Median (min, max) 1,110 (3.00, 468,000) 232 (3.00, 7,830) 2.710 (5.00, 468,000) Missing, No. (%) 1 (1.0) 0 (0) 11 (.6) Liver metastasis, No. (%) Tver metastasis, No. (%) Genoticabline plus nab-pacitaxel 56 (53.8) 20 (50.0) 36 (56.2) None/genotiabline/others 9 (8.7) 1 (2.5) 8 (12.5) POLFRINOX 39 (37.5) 19 (47.5) 20 (31.2) CTCs, No. (%) <1 CTC/mL 60 (57.7) 29 (72.5) 31 (48.4) ≥ 1 CTC/mL 70 (20.28) 8 (20.00) 22 (34.4) Missing No. (%) (14 (13.5) 3 (7.5) 11 (17.2) CTCs, No. (%) (20.00) 0.6380 (1.5400) 12.5000 (38.7000) Missing, No. (%) (20.0200) 0.6380 (1.5400) 12.5000 (38.7000) Missing, No. (%) (20.0200) 0.6380 (1.5400) 12.5000 (38.7000) Missing, No. (%) (20.0200) 0.00817 (0.0372) 0.00812 (0.1120) Missing No. (%) (25.0) 4 (6.2) CtRAS variant allele frequency <0000000 0.00817 (0.0372) 0.00812 (0.1120) Missing, No. (%) (6 (58) 2 (50.0) 4 (6.2)	Asian or others	6 (5.8)	1 (2.5)	5 (7.8)		
Mean (SD)         27.1000 (s.7200)         26.3000 (s.0300)         27.6000 (s.0300)           Missing, No. (%)         1 (1.0)         0 (0)         1 (1.6)           Diabetes mellitus, No. (%)         24 (37.5)         Alburnin, g/dl.           Mean (SD)         3.8100 (0.4520)         3.9100 (0.3300)         3.7500 (0.5050)           Missing, No. (%)         1 (1.0)         1 (2.5)         0 (0)           ECOG 0-1         84 (80.8)         35 (87.5)         49 (76.6)           ECOG 0-1         84 (80.8)         3 (7.5)         11 (1.7.2)           Missing         6 (5.8)         2 (5.0)         4 (6.2)           Missing, No. (%)         1 (1.0)         0 (0.0)         51 (79.7)           Initial treatment, No. (%)         20 (50.0)         36 (56.2)         Non eigemediable/dures         9 (8.7)         1 (2.5)         8 (12.5)	BWI					
Missing, No. (%)         1 (1.0)         0 (0)         1 (1.5)           Diabetes mellitus, No. (%)         7         42 (40.4)         18 (45.0)         24 (37.5)           Alburnin, g/dL         Mean (SD)         3.8100 (0.4520)         3.9100 (0.3300)         3.7500 (0.5050)           Missing, No. (%)         1 (1.0)         1 (2.5)         0 (0)           ECOG O-1         84 (80.8)         35 (87.5)         49 (76.6)           ECOG O-1         84 (80.8)         35 (87.5)         11 (17.2)           Missing         6 (5.8)         2 (5.0)         4 (6.2)           CA19-9, U/mL	Mean (SD)	27.1000 (5.7200)	26.3000 (5.0500)	27.6000 (6.0900)		
Dabetes mellitus, No. (%)           Yes         42 (40.4)         18 (45.0)         24 (37.5)           Albumin, gdL         Mean (SD)         3.8100 (0.4520)         3.9100 (0.3300)         3.7500 (0.5050)           Missing, No. (%)         1 (1.0)         1 (2.5)         0 (0)           ECOG PS, No. (%)         ECOG 0-1         84 (80.8)         35 (87.5)         49 (76.6)           ECOG 0-1         84 (80.8)         35 (87.5)         49 (76.6)         CA           ECOG 0-1         84 (80.8)         35 (87.5)         49 (76.6)         CA           ECOG 0-1         84 (80.8)         35 (87.5)         49 (76.6)         CA           CA19-9, U/mL         CA         CA         20.00         4 (6.2)         CA           Median (min, max)         1,110 (3.00, 468,000)         232 (3.00, 7,830)         2,710 (5.00, 468,000)         Missing, No. (%)         1 (1.6)           Liver metastasis, No. (%)         1 (1.0)         0 (00         1 (1.6)         Missing         1 (2.5)         8 (12.5)           Gemcitabine plus nab-pacitaxel         56 (53.8)         20 (50.0)         36 (56.2)         One/gemcitabine/others         9 (8.7)         1 (2.5)         8 (12.5)         CA           Gemcitabine plus nab-pacitaxel         56 (53.8)	Missing, No. (%)	1 (1.0)	0 (0)	1 (1.6)		
Yes         42 (40,4)         18 (45,0)         24 (37,5)           Albumin, g/dL	Diabetes mellitus, No. (%)					
Albumin, g/dL         Mean (SD)       3.8100 (0.4520)       3.9100 (0.3300)       3.7500 (0.5050)         Missing, No. (%)       1 (1.0)       1 (2.5)       0 (0)         ECOG PS, No. (%)       ECOG > 2       14 (13.5)       3 (7.5)       11 (17.2)         Missing       6 (5.8)       2 (5.0)       4 (6.2)       CA19-9, U/mL         Median (min, max)       1,110 [3.00, 468,000]       232 [3.00, 7,830]       2,710 [5.00, 468,000]       CA19-9, U/mL         Iter metastasis, No. (%)       1 (1.0)       0 (0)       1 (1.6)       Urver metastasis, No. (%)         Liver metastasis, No. (%)       1 (1.0)       0 (0.0)       51 (79.7)       Intial freatment, No. (%)         Genericabine plus nab-pacitizzel       56 (53.8)       20 (50.0)       36 (56.2)         Nonegemetitabine/others       9 (8.7)       1 (2.5)       8 (12.5)         FOLFIRINOX       39 (37.5)       19 (47.5)       20 (50.0)         Sing       14 (13.5)       3 (7.5)       11 (17.2)         Mesing       14 (13.5)       3 (7.5)       11 (17.2)         CTo, No. (%)       7.6500 (30.2000)       0.6380 (1.5400)       22 (34.4)         Missing       14 (13.5)       3 (7.5)       11 (17.2)         CTo, CTox/mL	Yes	42 (40.4)	18 (45.0)	24 (37.5)		
Mean (SD)         3.8100 (0.4520)         3.9100 (0.3300)         3.7500 (0.5050)           Missing, No. (%)         1 (1.0)         1 (2.5)         0 (0)           ECOG PS, No. (%)         ECOG 0.1         84 (80.8)         35 (87.5)         49 (76.6)           ECOG 0.1         84 (80.8)         35 (87.5)         49 (76.6)           ECOG 0.1         84 (80.8)         2 (5.0)         4 (6.2)           CAI9-9, U/mL	Albumin, g/dL					
Missing, No. (%)         1 (1.0)         1 (2.5)         0 (0)           ECOG 0-1         84 (80.8)         35 (87.5)         49 (76.6)           ECOG 0-1         84 (80.8)         35 (87.5)         11 (17.2)           Missing         6 (5.8)         2 (5.0)         4 (6.2)           CA19-9, U/mL          0 (0)         1 (1.6)           Median (min, max)         1,110 (3.00, 468,000)         232 (3.00, 7,830)         2,710 (5.00, 468,000)           Missing, No. (%)         1 (1.0)         0 (0)         1 (1.6)           Liver metastasis, No. (%)         1 (1.0)         0 (0)         51 (79.7)           Yes         51 (149.0)         0 (0.0)         51 (79.7)           Initial treatment, No. (%)          20 (50.0)         36 (56.2)           None/gemcitabine/others         9 (8.7)         1 (2.5)         8 (12.5)           FOLFIRINOX         39 (37.5)         19 (47.5)         20 (31.2)           CTcs, No. (%)         30 (28.8)         8 (20.0)         22 (34.4)           Missing         14 (13.5)         3 (7.5)         11 (17.2)           CTcs, CTcs/mL         .006         .006300 (1.5400)         12.5000 (38.7000)           Missing, No. (%)         14 (13.5)         3 (7.	Mean (SD)	3.8100 (0.4520)	3.9100 (0.3300)	3.7500 (0.5050)		
ECOG 0-1         84 (80.8)         35 (87.5)         49 (76.6)           ECOG 0-1         84 (80.8)         35 (87.5)         11 (17.2)           Missing         6 (5.8)         2 (5.0)         4 (6.2)           CA19-9, U/mL	Missing, No. (%)	1 (1.0)	1 (2.5)	0 (0)		
ECOG 0-184 (80.8)35 (87.5)49 (76.6)ECOG $\geq$ 214 (13.5)3 (7.5)11 (17.2)Missing6 (5.8)2 (5.0)4 (6.2)CA19-9, U/mLMedian (min, max)1,110 (3.00, 468,000)232 (3.00, 7,830)2,710 [5.00, 468,000]Median (min, max)1,110 (1.0)0 (0)1 (1.6)Liver metastasis, No. (%)1 (1.0)0 (0.0)51 (79.7)Initial treatment, No. (%)55 (53.8)20 (50.0)36 (56.2)Gemcitabine plus nab-paciltaxel56 (53.8)20 (50.0)36 (56.2)None/gemcitabine/others9 (8.7)1 (2.5)8 (12.5)FOLFIRINOX39 (37.5)19 (47.5)20 (31.2)CTCs, No. (%)	ECOG PS, No. (%)					
ECOG ≥ 214 (13.5)3 (7.5)11 (17.2)Missing6 (5.8)2 (5.0)4 (6.2)CA199, U/mL	ECOG 0-1	84 (80.8)	35 (87.5)	49 (76.6)		
Missing         6 (5.8)         2 (5.0)         4 (6.2)           CA19-9, U/mL	$ECOG \ge 2$	14 (13.5)	3 (7.5)	11 (17.2)		
CA19-9, U/mL           Median (min, max)         1,110 [3.00, 468,000]         232 [3.00, 7,830]         2,710 [5.00, 468,000]           Missing, No. (%)         1 (1.0)         0 (0)         1 (1.6)           Liver metastasis, No. (%)         0 (0.0)         51 (79.7)           Yes         51 (49.0)         0 (0.0)         51 (79.7)           Initial treatment, No. (%)         20 (50.0)         36 (56.2)           Gemcitabine plus nab-pacilitaxel         56 (53.8)         20 (50.0)         36 (56.2)           None/gemcitabine/others         9 (8.7)         1 (2.5)         8 (12.5)           FOLFIRINOX         39 (37.5)         19 (47.5)         20 (31.2)           CTCs, No. (%)         60 (57.7)         29 (72.5)         31 (48.4)           ≥ 1 CTC/mL         60 (57.7)         29 (72.5)         31 (48.4)           ≥ 1 CTC/mL         30 (28.8)         8 (20.0)         22 (34.4)           Missing         14 (13.5)         3 (7.5)         11 (17.2)           Mean (SD)         7.6500 (30.2000)         0.6380 (1.5400)         12.5000 (38.7000)           Missing, No. (%)         14 (13.5)         3 (7.5)         11 (17.2)           Not detected         36 (63.5)         16 (40.0)         50 (78.1) <t< td=""><td>Missing</td><td>6 (5.8)</td><td>2 (5.0)</td><td>4 (6.2)</td><td></td></t<>	Missing	6 (5.8)	2 (5.0)	4 (6.2)		
Median (min, max)         1,110 [3.00, 468,000]         232 [3.00, 7,830]         2,710 [5.00, 468,000]           Missing, No. (%)         1 (1.0)         0 (0)         1 (1.6)           Liver metastasis, No. (%)         0 (0.0)         51 (79.7)           Yes         51 (49.0)         0 (0.0)         51 (79.7)           Initial treatment, No. (%)              Gemcitabine plus nab-pacitaxel         56 (53.8)         20 (50.0)         36 (56.2)           None/gemcitabine/others         9 (8.7)         1 (2.5)         8 (12.5)           FOLFIRINOX         39 (37.5)         19 (47.5)         20 (31.2)           CTCs, No. (%)          30 (28.8)         8 (20.0)         22 (34.4)           Missing         14 (13.5)         3 (7.5)         11 (17.2)           CTCs, CTCs/mL	CA19-9, U/mL					
Missing, No. (%)         1 (1.0)         0 (0)         1 (1.6)           Liver metastasis, No. (%)         Yes         51 (49.0)         0 (0.0)         51 (79.7)           Initial treatment, No. (%)         Gemcitabine plus nab-paclitaxel         56 (53.8)         20 (50.0)         36 (56.2)           None/gemcitabine/others         9 (8.7)         1 (2.5)         8 (12.5)           FOLFIRINOX         39 (37.5)         19 (47.5)         20 (31.2)           CTCs, No. (%)         30 (28.8)         8 (20.0)         22 (34.4)           ≥ 1 CTC/mL         60 (57.7)         29 (72.5)         31 (48.4)           ≥ 1 CTC/mL         30 (28.8)         8 (20.0)         22 (34.4)           Missing         14 (13.5)         3 (7.5)         11 (17.2)           CTCs, CTCs/mL	Median (min, max)	1,110 [3.00, 468,000]	232 [3.00, 7,830]	2,710 [5.00, 468,000]		
Liver metastasis, No. (%)           Yes         51 (49.0)         0 (0.0)         51 (79.7)           Initial treatment, No. (%)         20 (50.0)         36 (56.2)           Gemcitabine plus nab-paclitaxel         56 (53.8)         20 (50.0)         36 (56.2)           None/gemcitabine/others         9 (8.7)         1 (2.5)         8 (12.5)           FOLFIRINOX         39 (37.5)         19 (47.5)         20 (31.2)           CTCs, No. (%)         39 (37.5)         19 (47.5)         20 (31.2)            60 (57.7)         29 (72.5)         31 (48.4)           ≥ 1 CTC/mL         60 (57.7)         29 (72.5)         31 (48.4)           ≥ 1 CTC/mL         30 (28.8)         8 (20.0)         22 (34.4)           Missing         14 (13.5)         3 (7.5)         11 (17.2)           CTCs, CTCs/mL	Missing, No. (%)	1 (1.0)	0 (0)	1 (1.6)		
Yes         51 (49.0)         0 (0.0)         51 (79.7)           Initial treatment, No. (%)         Gemcitabine plus nab-paclitaxel         56 (53.8)         20 (50.0)         36 (56.2)           None/gemcitabine/others         9 (8.7)         1 (2.5)         8 (12.5)           FOLFIRINOX         39 (37.5)         19 (47.5)         20 (31.2)           CTCs, No. (%)         20 (50.0)         22 (34.4)            30 (28.8)         8 (20.0)         22 (34.4)           Missing         14 (13.5)         3 (7.5)         11 (17.2)           CTCs, CTCs/mL	Liver metastasis, No. (%)					
Initial treatment, No. (%)           Gemcitabine plus nab-pacilitaxel         56 (53.8)         20 (50.0)         36 (56.2)           None/gemcitabine/others         9 (8.7)         1 (2.5)         8 (12.5)           FOLFIRINOX         39 (37.5)         19 (47.5)         20 (31.2)           CTCs, No. (%)         20 (50.0)         31 (48.4)           ≥ 1 CTC/mL         60 (57.7)         29 (72.5)         31 (48.4)           ≥ 1 CTC/mL         30 (28.8)         8 (20.0)         22 (34.4)           Missing         14 (13.5)         3 (7.5)         11 (17.2)           CTCs, CTCs/mL	Yes	51 (49.0)	0 (0.0)	51 (79.7)		
Gemcitabine plus nab-pacitaxel         56 (53.8)         20 (50.0)         36 (56.2)           None/gemcitabine/others         9 (8.7)         1 (2.5)         8 (12.5)           FOLFIRINOX         39 (37.5)         19 (47.5)         20 (31.2)           CTCs, No. (%)         29 (72.5)         31 (48.4)           ≥ 1 CTC/mL         60 (57.7)         29 (72.5)         31 (48.4)           ≥ 1 CTC/mL         30 (28.8)         8 (20.0)         22 (34.4)           Missing         14 (13.5)         3 (7.5)         11 (17.2)           CTCs, CTCs/mL	Initial treatment, No. (%)					
None/gemcitabine/others         9 (8.7)         1 (2.5)         8 (12.5)           FOLFIRINOX         39 (37.5)         19 (47.5)         20 (31.2)           CTCs, No. (%)         20 (31.2)         .049           < 1 CTC/mL	Gemcitabine plus nab-paclitaxel	56 (53.8)	20 (50.0)	36 (56.2)		
FOLFIRINOX         39 (37.5)         19 (47.5)         20 (31.2)           CTCs, No. (%)         .049           < 1 CTC/mL	None/gemcitabine/others	9 (8.7)	1 (2.5)	8 (12.5)		
CTCs, No. (%)       .049         < 1 CTC/mL	FOLFIRINOX	39 (37.5)	19 (47.5)	20 (31.2)		
< 1 CTC/mL         60 (57.7)         29 (72.5)         31 (48.4)           ≥ 1 CTC/mL         30 (28.8)         8 (20.0)         22 (34.4)           Missing         14 (13.5)         3 (7.5)         11 (17.2)           CTCs, CTCs/mL         .006         .007         .007           Mean (SD)         7.6500 (30.2000)         0.6380 (1.5400)         12.5000 (38.7000)         .007           Missing, No. (%)         14 (13.5)         3 (7.5)         11 (17.2)         .007           Missing, No. (%)         14 (13.5)         3 (7.5)         11 (17.2)         .007           Missing, No. (%)         14 (13.5)         3 (7.5)         11 (17.2)         .007           Missing, No. (%)         6 (5.8)         22 (55.0)         10 (15.6)         .001           Not detected         32 (30.8)         22 (55.0)         10 (15.6)         .001           Detected         66 (63.5)         16 (40.0)         50 (78.1)         .001           Missing         6 (5.8)         2 (5.0)         4 (6.2)         .001           Missing, No. (%)         6 (5.8)         2 (5.0)         4 (6.2)	CTCs, No. (%)				.049	
≥ 1 CTC/mL         30 (28.8)         8 (20.0)         22 (34.4)           Missing         14 (13.5)         3 (7.5)         11 (17.2)           CTCs, CTCs/mL         .006           Mean (SD)         7.6500 (30.2000)         0.6380 (1.5400)         12.5000 (38.7000)           Missing, No. (%)         14 (13.5)         3 (7.5)         11 (17.2)           CtKRAS mutation, No. (%)         14 (13.5)         3 (7.5)         11 (17.2)           Ot detected         32 (30.8)         22 (55.0)         10 (15.6)           Not detected         32 (30.8)         22 (55.0)         10 (15.6)           Detected         66 (63.5)         16 (40.0)         50 (78.1)           Missing         6 (5.8)         2 (5.0)         4 (6.2)           Missing, No. (%)         0.0529 (0.0970)         0.00817 (0.0372)         0.0812 (0.1120)           Missing, No. (%)         6 (5.8)         2 (5.0)         4 (6.2)	< 1 CTC/mL	60 (57.7)	29 (72.5)	31 (48.4)		
Missing         14 (13.5)         3 (7.5)         11 (17.2)           CTCs, CTCs/mL         .006           Mean (SD)         7.6500 (30.2000)         0.6380 (1.5400)         12.5000 (38.7000)           Missing, No. (%)         14 (13.5)         3 (7.5)         11 (17.2)           CtKRAS mutation, No. (%)         14 (13.5)         3 (7.5)         11 (17.2)           Not detected         32 (30.8)         3 (7.5)         11 (17.2)           Not detected         32 (30.8)         22 (55.0)         10 (15.6)           Detected         32 (30.8)         22 (55.0)         10 (15.6)           Missing         66 (63.5)         16 (40.0)         50 (78.1)           Missing         6 (5.8)         2 (5.0)         4 (6.2)           ctKRAS variant allele frequency         <<.001	$\geq$ 1 CTC/mL	30 (28.8)	8 (20.0)	22 (34.4)		
CTCs, CTCs/mL       .006         Mean (SD)       7.6500 (30.2000)       0.6380 (1.5400)       12.5000 (38.7000)         Missing, No. (%)       14 (13.5)       3 (7.5)       11 (17.2)         ctKRAS mutation, No. (%)       22 (55.0)       10 (15.6)         Not detected       32 (30.8)       22 (55.0)       10 (15.6)         Detected       66 (63.5)       16 (40.0)       50 (78.1)         Missing       6 (5.8)       2 (5.0)       4 (6.2)         ctKRAS variant allele frequency       < .001	Missing	14 (13.5)	3 (7.5)	11 (17.2)		
Mean (SD)         7.6500 (30.2000)         0.6380 (1.5400)         12.5000 (38.7000)           Missing, No. (%)         14 (13.5)         3 (7.5)         11 (17.2)           ctKRAS mutation, No. (%)           <	CTCs, CTCs/mL				.006	
Missing, No. (%)         14 (13.5)         3 (7.5)         11 (17.2)           ctKRAS mutation, No. (%)          <	Mean (SD)	7.6500 (30.2000)	0.6380 (1.5400)	12.5000 (38.7000)		
ctKRAS mutation, No. (%)         < .001           Not detected         32 (30.8)         22 (55.0)         10 (15.6)           Detected         66 (63.5)         16 (40.0)         50 (78.1)           Missing         6 (5.8)         2 (5.0)         4 (6.2)           ctKRAS variant allele frequency         < .001	Missing, No. (%)	14 (13.5)	3 (7.5)	11 (17.2)		
Not detected         32 (30.8)         22 (55.0)         10 (15.6)           Detected         66 (63.5)         16 (40.0)         50 (78.1)           Missing         6 (5.8)         2 (5.0)         4 (6.2)           ctKRAS variant allele frequency         <	ctKRAS mutation, No. (%)				< .001	
Detected         66 (63.5)         16 (40.0)         50 (78.1)           Missing         6 (5.8)         2 (5.0)         4 (6.2)           ctKRAS variant allele frequency         < .001         < .001           Mean (SD)         0.0529 (0.0970)         0.00817 (0.0372)         0.0812 (0.1120)           Missing, No. (%)         6 (5.8)         2 (5.0)         4 (6.2)	Not detected	32 (30.8)	22 (55.0)	10 (15.6)		
Missing         6 (5.8)         2 (5.0)         4 (6.2)           ctKRAS variant allele frequency         < .001	Detected	66 (63.5)	16 (40.0)	50 (78.1)		
ctKRAS variant allele frequency         < .001           Mean (SD)         0.0529 (0.0970)         0.00817 (0.0372)         0.0812 (0.1120)           Missing, No. (%)         6 (5.8)         2 (5.0)         4 (6.2)	Missing	6 (5.8)	2 (5.0)	4 (6.2)		
Mean (SD)         0.0529 (0.0970)         0.00817 (0.0372)         0.0812 (0.1120)           Missing, No. (%)         6 (5.8)         2 (5.0)         4 (6.2)	ctKRAS variant allele frequency				< .001	
Missing, No. (%) 6 (5.8) 2 (5.0) 4 (6.2)	Mean (SD)	0.0529 (0.0970)	0.00817 (0.0372)	0.0812 (0.1120)		
	Missing, No. (%)	6 (5.8)	2 (5.0)	4 (6.2)		

(Continued on following page)

TABLE 1. Patient Characteristics in the Locally Advanced and Metastatic Cohorts (Continued)

		Locally			
Characteristic	Overall (N = 104)	Advanced $(n = 40)$	Metastatic (n = 64)	Р	
cfDNA concentration, ng/mL				.007	
Mean (SD)	19.7000 (41.6000)	8.1000 (5.2800)	27.0000 (51.9000)		
Missing, No. (%)	6 (5.8)	2 (5.0)	4 (6.2)		

Abbreviations: BMI, body mass index; CA, cancer antigen; cfDNA, cell-free DNA; CTC, circulating tumor cell; ctKRAS, circulating tumor *KRAS*; ECOG PS, Eastern Cooperative Oncology Group performance status; FOLFIRINOX, fluorouracil, leucovorin, irinotecan, and oxaliplatin; SD, standard deviation.

2015, and June 22, 2018. Ninety-nine patients (95.2%) died during follow-up. Of patients who were censored, the median follow-up was 40.1 months (range 9.3-44.7 months). The median OS was 11.9 months (95% CI, 9.2 to 15.5) in the overall population, 16.2 months (95% CI, 9.2 to 19.3) in the LAPC cohort, and 8.5 months (95% CI, 7.0 to 12.9 months) in mPDAC. cfDNA, ctKRAS VAF, and CTCs were all significantly higher in patients with mPDAC compared with LAPC (Fig 1).

### **Univariate Survival Analysis**

Univariate association of liquid biopsy and traditional clinical variables with OS is presented in Table 2. In addition, we examined the univariate association of liquid biopsy variables with OS within the LAPC and mPDAC subgroups. There was increased hazard of death for each 0.05 unit increase in cube root of ctKRAS VAF that was not statistically significant in the LAPC cohort (hazard ratio [HR], 1.12; 95% CI, 0.99 to 1.27; P = .08) but was



**FIG 1.** Box plots demonstrating distribution of (A) cfDNA concentration (ng/mL); (B) ctKRAS mutation variant allele frequency (ctKRAS VAF); and (C) CTCs (CTCs/mL of blood) in locally advanced and metastatic cohorts. (A) *P* values displayed represent the results of a two-tailed, two-sample *t*-test and (B) and (C) Wilcoxon rank-sum tests as variables were non-normally distributed. Kaplan-Meier survival curves stratified by (D) cfDNA concentration  $\geq$  15 ng/mL or < 15 ng/mL; (E) ctKRAS VAF  $\geq$  0.05 or < 0.05; and (F) CTCs  $\geq$  1/mL or < 1/mL. Binary cutoffs were chosen as discussed in the Statistical Methods section. *P* values represent the result of a log-rank test comparing the survival curves across reported strata. Numbers of patients at risk at each time point for each group are presented below Kaplan-Meier curves. cfDNA, cell-free DNA; CTC, circulating tumor cell; ctKRAS, circulating tumor *KRAS*; OS, overall survival; VAF, variant allele fraction.

Variable	Referent	HR	95% CI	Р
Age/5ª	NA	1.11	1.00 to 1.24	.060
Male sex	Female sex	1.18	0.79 to 1.77	.430
Treatment with none/gemcitabine/others	Gemcitabine plus nab-paclitaxel	1.26	0.59 to 2.66	.550
Treatment with FOLFIRINOX	Gemcitabine plus nab-paclitaxel	0.81	0.53 to 1.24	.330
Diabetes mellitus	No diabetes	1.03	0.69 to 1.53	.900
Former smoker	Never smoker	1.20	0.77 to 1.85	.420
Current smoker	Never smoker	1.04	0.51 to 2.10	.920
Black/African American race	White Race	0.66	0.32 to 1.37	.270
Asian/other race	White Race	0.95	0.38 to 2.36	.920
BMI ≥ 35	BMI < 35	1.58	0.79 to 3.15	.200
Albumin	NA	0.33	0.20 to 0.54	< .001
ECOG PS of 2 or greater	ECOG 0-1	2.29	1.28 to 4.10	.005
Metastatic	Locally advanced	1.84	1.22 to 2.80	.004
Liver metastasis	No liver metastasis	1.80	1.21 to 2.69	.004
Log (CA19-9) <sup>b</sup>	NA	1.17	1.08 to 1.27	< .001
cfDNA concentration <sup>c</sup>	NA	1.04	1.02 to 1.05	< .001
ctKRAS <sup>d</sup>	NA	1.16	1.10 to 1.22	< .001
$CTCs \ge 1/mL$ of blood	< 1 CTC/mL	1.98	1.26 to 3.12	.003

 TABLE 2.
 Results of Univariate Cox Proportional Hazards Models for Association With Candidate Variables With Overall Survival

 Variable
 Referent
 HR
 95% CI

Abbreviations: BMI, body mass index; CA, cancer antigen; cfDNA, cell-free DNA; CTC, circulating tumor cell; ctKRAS, circulating tumor *KRAS*; ECOG PS, Eastern Cooperative Oncology Group performance status; FOLFIRINOX, fluorouracil, leucovorin, irinotecan, and oxaliplatin; HR, hazard ratio; NA, not available.

<sup>a</sup>Indicates HR for age reported for 5-year increments.

<sup>b</sup>Log transformation of CA19-9 (units/mL).

°cfDNA concentration in ng/mL capped at a maximum value of 50 ng/mL.

<sup>d</sup>Cube root transformation of ctKRAS variant allele fraction with HR reported in 0.05 unit increments.

statistically significant in the mPDAC cohort (HR, 1.15; 95% CI, 1.08 to 1.22; P < .0001). CTCs  $\geq$  1/mL was not associated with OS in LAPC (HR, 0.72; 95% CI, 0.31 to 1.69; P = .45) but was significantly associated with increased hazard of death in mPDAC (HR, 5.54; 95% CI, 2.67 to 11.47; P < .0001).Increasing cfDNA concentration was significantly associated with increased hazard of death in both LAPC (HR, 1.08; 95% CI, 1.00 to 1.17; P = .04) and mPDAC cohorts (HR, 1.03; 95% CI, 1.01 to 1.04; P = .0003). When treated as binary variables, CTCs  $\geq$  1/mL, cfDNA  $\geq$  15 ng/mL, and ctKRAS VAF  $\geq$  0.05 were significantly associated with worse OS (Fig 1).

which both analytes were detectable. Interestingly, there was no significant association between CTCs  $\geq$  1/mL and ctKRAS VAF  $\geq$  0.05 (P = .13; Table 3) or when treated as continuous variables (P = .47; Data Supplement) suggesting that these analytes may be complementary. Within the LAPC cohort, 27 of 37 patients (73%) with nonmissing data for CTCs and ctKRAS had no detectable CTCs or ctKRAS (dual negative, CTCs < 1/mL and ctKRAS VAF < 0.05), and the remaining 10 of 37 patients (27%) were deemed single positive (CTCs > 1/mL or ctKRAS VAF > 0.05). However, within the mPDAC cohort, 10 of 53 patients (19%) evaluable for both analytes were dual positive, 22 (42%) were single positive, and 21 patients (40%) were dual negative for CTCs and ctKRAS.

Given that ctKRAS and CTCs are two different measures of tumor shedding, we next examined the frequency with

TABLE 3.	Association	of	ctKRAS	VAF	≥	0.05	With	CTCs	≥	1/mL
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CTC Status	ctKRAS VAF < 0.05, No. (% total)	ctKRAS ≥ 0.05, No. (% total)	Row Total
CTC $<$ 1/mL, No. (% total)	48 (53)	12 (13)	60
CTC $\geq$ 1/mL, No. (% total)	20 (22)	10 (11)	30
Column total	68	22	90

NOTE. Fisher's exact test P = .13.

Abbreviations: CTC, circulating tumor cell; ctKRAS, circulating tumor KRAS; VAF, variant allele fraction.

mPDAC cohort, there was a significant difference in OS by log-rank testing and by univariate Cox proportional hazards modeling (Fig 2).

## **Multivariate Cox Proportional Hazards Models**

A multivariate model was selected to assess the association of liquid biopsy variables with OS after adjusting for clinical covariates, selected as identified in the Statistical Methods section. CTCs  $\geq 1/mL$ , cube root of ctKRAS VAF, and albumin were independently associated with OS in a model containing all candidate variables that had a P < .15 in univariate analysis (model A; Table 4). cfDNA concentration was not independently associated with OS in this model.



**FIG 2.** Assessment of OS by CTC and ctKRAS status by (A) the Kaplan-Meier method in patients with mPDAC along with the number of patients at risk and (B) by univariate Cox proportional hazards modeling in patients with mPDAC. Patients were categorized as dual negative for CTCs and ctKRAS (CTCs < 1/mL and ctKRAS VAF < 0.05), single positive (CTCs  $\geq$  1/mL or ctKRAS VAF  $\geq$  0.05), or dual positive (CTCs  $\geq$  1/mL and ctKRAS VAF  $\geq$  0.05). Twenty-one patients with mPDAC were dual negative, 22 were single positive, and 10 were dual positive. Among patients with mPDAC, dual positivity was associated with worse OS than dual negativity (HR, 18.84; 95% CI, 6.38 to 55.68; *P* < .001) and single positivity was associated with worse OS than dual negativity (HR, 5.98; 95% CI, 2.64 to 13.54; *P* < .001). CTC, circulating tumor cell; ctKRAS, circulating tumor *KRAS*; HR, hazard ratio; mPDAC, metastatic pancreatic ductal adenocarcinoma; OS, overall survival; VAF, variant allele fraction.

Likelihood ratio testing was performed to compare fit of each model with that of model A (Table 4). These comparisons suggest that models containing ctKRAS, CTCs, and cfDNA (models A and C) had statistically significantly improved model fit compared with models which considered traditional clinical variables only (models B and D).

Internal validation using bootstrapping showed optimismcorrected Harrell's C-statistics of 0.698, 0.673, 0.725, and 0.681 for models A-D, respectively. These results suggest that models containing ctKRAS, CTCs, and cfDNA (models A and C) have higher discrimination and are more likely to perform better in external data sets than models containing traditional prognostic variables alone (models B and D). Calibration plots showed reasonable calibration for models A-D (Data Supplement).

## DISCUSSION

In a cohort of patients with LAPC or mPDAC, we found that liquid biopsy variables, including ctKRAS VAF and CTCs were significantly associated with OS in univariate analysis. There was no significant association between CTCs and ctKRAS when assessed as continuous or binary variables. In patients with mPDAC, those classified as dual positive for CTCs  $\geq$  1/mL and ctKRAS VAF  $\geq$  0.05 had significantly worse OS compared with patients who were single positive and dual negative, respectively. After adjusting for clinical confounders in multivariate analysis, CTCs and ctKRAS VAF remained independently associated with OS. We also found that a prognostic model using liquid biopsy plus traditional clinical variables had significantly improved model fit compared with a model developed using traditional clinical variables alone. In addition, the ability of the models containing liquid biopsy variables plus traditional clinical variables (models A and C) to discriminate between patients with differential survival, as measured by an optimism-corrected Harrell's C-statistic, was higher compared with models using traditional clinical variables only (models B and D). These findings suggest that liquid biopsy variables provide information that is complementary to each other and to that of traditional clinical variables.

One novel aspect of our study is that the association of CTCs detected by a nonenrichment method with OS has not been formally evaluated in PDAC. We found that the presence of CTCs at a concentration of  $\geq 1/mL$  was significantly associated with increased hazard of death in both univariate and multivariate analysis. This result is consistent with that observed in CTCs detected by other methods in patients with advanced PDAC.<sup>18,19</sup>

We explored whether our liquid biopsy variables might have different associations with OS in patients with LAPC versus mPDAC. Interestingly, for cfDNA and ctKRAS VAF, the effect size and direction of the association with OS was similar in the LAPC and mPDAC cohorts (although the result was not statistically significant for ctKRAS VAF in the LAPC cohort). However, there was no relationship between

	Model A: All Variables With <i>P</i> < .15 in Univariate Analysis		Model B: All Traditional Clinical Variables With <i>P</i> < .15 in Univariate Analysis		Model C: Stepwise B Selection of All Candida With <i>P</i> < .15 in Univaria	ackward te Variables ate Analysis	Model D: Stepwise Backward Selection of Traditional Clinical Variables Only With P < .15 in Univariate Analysis		
Variable	HR (95% CI)	Р	HR (95% CI)	Р	HR (95% CI)	Р	HR (95% CI)	Р	
ctKRAS <sup>a</sup>	1.11 (1.03 to 1.21)	.010 <sup>b</sup>			1.14 (1.08 to 1.21)	< .001 <sup>b</sup>			
$\geq$ 1 CTC/mL	2.22 (1.34 to 3.69)	.002 <sup>b</sup>			2.16 (1.33 to 3.50)	.002 <sup>b</sup>			
cfDNA <sup>c</sup>	1.02 (1.00 to 1.04)	.130			1.02 (1.00 to 1.03)	.080			
Albumin	0.40 (0.19 to 0.83)	.010 <sup>b</sup>	0.34 (0.19 to 0.61)	.003 <sup>b</sup>	0.40 (0.22 to 0.72)	.002 <sup>b</sup>	0.30 (0.18 to 0.50)	$< .001^{b}$	
Age/5	1.13 (0.99 to 1.29)	.080	1.08 (0.95 to 1.23)	.250	1.15 (1.02 to 1.29)	.020 <sup>b</sup>	1.10 (0.97 to 1.24)	.120	
Metastatic	1.96 (0.83 to 4.65)	.130	1.08 (0.49 to 2.39)	.850			1.43 (0.89 to 2.29)	.140	
Log of CA19-9	1.04 (0.94 to 1.16)	.450	1.17 (1.06 to 1.28)	.002 <sup>b</sup>			1.17 (1.07 to 1.28)	$< .001^{b}$	
Liver metastasis	0.62 (0.26 to 1.51)	.290	1.40 (0.68 to 2.88)	.360					
ECOG $\geq 2$	1.20 (0.57 to 2.54)	.630	1.25 (0.64 to 2.43)	.510					
Model summary									
P For LR test comparing reduced model with model A <sup>d</sup>	NA (reference model)		< .001 <sup>b</sup>		.420		< .001 <sup>b</sup>		
No. of patients <sup>e</sup>	84		96		89		102		
Optimism-corrected	0.698		0.673		0.725		0.681		

Harrell's C-statistic

NOTE. Age/5 indicates the HR represents a 5-year increment in age. Variable transformations indicated in the table aimed to improve interpretability of HRs.

Abbreviations: CA, cancer antigen; cfDNA, cell-free DNA; CTC, circulating tumor cell; ctKRAS, circulating tumor *KRAS*; ECOG, Eastern Cooperative Oncology Group; HR, hazard ratio; LR, likelihood ratio; NA, not available.

<sup>a</sup>Represents cube root transformation of ctKRAS variant allele fraction in 0.05 unit increments.

<sup>b</sup>Indicates *P* values significant at an  $\alpha$  of .05.

°Represents cfDNA in ng/mL capped at a maximum value of 50 ng/mL.

<sup>d</sup>*P* < .05 indicates that full model (model A) has statistically significantly higher log likelihood, or better fit to the data, than the reduced model and that the variables in the larger model should be retained when choosing between the two. LR testing was performed using a data set that included complete cases on all of the variables in the largest model (model A).

<sup>e</sup>To calculate effect sizes and optimism-corrected Harrell's C, each model was performed on a data set of cases that were complete on all variables included in each model. This explains why each model is run on a different number of patients.

CTC > 1/mL and OS in the LAPC cohort while there was a very strong relationship between  $CTC \ge 1/mL$  and worse OS in the mPDAC cohort. This result must be considered exploratory as CTC detection rate in LAPC patients was quite low in this study (20%) lending insufficient power to detect an association with survival. This lower detection rate of CTCs in LAPC compared with mPDAC is consistent with previously observed results.34 When examining the combination of CTCs and ctKRAS within the mPDAC cohort, patients with dual positivity for CTCs and ctKRAS had significantly worse OS compared with those with single positivity, who in turn had worse OS compared with those with dual negativity (Fig 2). Overall, these results suggest that although cfDNA concentration and ctKRAS VAF are likely useful prognostic biomarkers in both LAPC and mPDAC, CTC detection and the combination of CTCs and ctKRAS may be more useful when used in patients with more advanced disease.

Limitations include the study of a moderate sample size enrolled at a single institution. Second, although tumor volumetric measurements were outside the scope of our study, previous work has shown an association between ctDNA VAF and tumor volume in patients with mPDAC and that ctDNA VAF and tumor volume were significantly

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W.J.C. and J.E.T. share first authorship to this work. W.-T.H. is the second author on the work. M.H.O. and E.L.C. share senior authorship to this work.

### **PRIOR PRESENTATION**

Items related to current article were presented at the American Society of Clinical Oncology (ASCO) Annual Meeting, Chicago, IL, June 4-8, 2021.

### SUPPORT

Supported by Grant No. 1R01 CA207643-01, by the Penn Center for Pancreatic Research, the Basser Young Leadership Council, the Konner Fund, the Pearl and Philip Basser Award, and An Anonymous Foundation.

#### DATA SHARING STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors upon request, without undue reservation.

associated with OS.<sup>35</sup> Future research may address whether prognostic models including ctDNA VAF, tumor volumetric measurements, or both perform better in patients with PDAC. Third, although methods of ctDNA detection (other than ctKRAS) have also demonstrated prognostic value, a comparison of methods of ctDNA assessment was outside the scope of this study.<sup>36</sup> Fourth, although we performed robust internal validation using a bootstrapping approach to estimate the likely performance of the models in an external data set, the models have not undergone external validation and thus are not ready for clinical application. Nevertheless, liquid biopsy variables including ctKRAS, cfDNA, and CTCs have demonstrated promise for improving prognostication beyond traditional clinical variables alone in patients with LAPC and mPDAC.

Future research may include external validation of our prognostic model and assess the utility of ctKRAS, cfDNA, and CTCs in monitoring response to therapy. Additional studies would evaluate the utility of a prognostic model for selecting patients with LAPC who have received neoadjuvant therapy for curative-intent surgery and for stratifying patients with mPDAC onto clinical trials earlier in therapy.

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The following represents disclosure information provided by authors of this manuscript. All relationships are considered compensated unless otherwise noted. Relationships are self-held unless noted. I = Immediate Family Member, Inst = My Institution. Relationships may not relate to the subject matter of this manuscript. For more information about ASCO's conflict of interest policy, please refer to www.asco.org/rwc or ascopubs. org/po/author-center.

Open Payments is a public database containing information reported by companies about payments made to US-licensed physicians (Open Payments).

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No other potential conflicts of interest were reported.

## ACKNOWLEDGMENT

We would like to acknowledge, and thank, all of the patients and family members who participated in this study for their contribution to this research.

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