



Review

Diverse Roles of Ceramide in the Progression and Pathogenesis of Alzheimer's Disease

Md Riad Chowdhury^{1,2}, Hee Kyung Jin^{1,3,*} and Jae-sung Bae^{1,2,*}¹ KNU Alzheimer's Disease Research Institute, Kyungpook National University, Daegu 41566, Korea² Department of Physiology, School of Medicine, Kyungpook National University, Daegu 41944, Korea³ Department of Laboratory Animal Medicine, College of Veterinary Medicine, Kyungpook National University, Daegu 41566, Korea

* Correspondence: hkjin@knu.ac.kr (H.K.J.); jsbae@knu.ac.kr (J.-s.B.); Tel.: +82-53-950-5966 (H.K.J.); +82-53-420-4815 (J.-s.B.); Fax: +82-53-950-5955 (H.K.J.); +82-53-424-3349 (J.-s.B.)

Abstract: Alzheimer's disease (AD) is the most common neurodegenerative disorder, and is associated with several pathophysiological features, including cellular dysfunction, failure of neurotransmission, cognitive impairment, cell death, and other clinical consequences. Advanced research on the pathogenesis of AD has elucidated a mechanistic framework and revealed many therapeutic possibilities. Among the mechanisms, sphingolipids are mentioned as distinctive mediators to be associated with the pathology of AD. Reportedly, alteration in the metabolism of sphingolipids and their metabolites result in the dysfunction of mitochondria, autophagy, amyloid beta regulation, and neuronal homeostasis, which exacerbates AD progression. Considering the importance of sphingolipids, in this review, we discuss the role of ceramide, a bioactive sphingolipid metabolite, in the progression and pathogenesis of AD. Herein, we describe the ceramide synthesis pathway and its involvement in the dysregulation of homeostasis, which eventually leads to AD. Furthermore, this review references different therapeutics proposed to modulate the ceramide pathway to maintain ceramide levels and prevent the disease progression.

Keywords: Alzheimer's disease; ceramide; amyloid beta; autophagy; mitochondrial dysfunction; senescence



Citation: Chowdhury, M.R.; Jin, H.K.; Bae, J.-s. Diverse Roles of Ceramide in the Progression and Pathogenesis of Alzheimer's Disease. *Biomedicines* **2022**, *10*, 1956. <https://doi.org/10.3390/biomedicines10081956>

Academic Editors: Sónia Catarina Correia, Cristina Carvalho and Susana Cardoso

Received: 20 June 2022

Accepted: 2 August 2022

Published: 12 August 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Alzheimer's disease (AD) is defined as progressive cognitive impairment associated with the formation of senile plaques in the cerebral cortex and subcortical gray matter, which includes amyloid beta (A β) and neurofibrillary tangles [1]. The World Health Organization has declared AD a "global public health priority" because of the lack of well-established treatments. To date, researchers have proposed various theories and hypotheses regarding the causes and targets of AD. Primarily, dementia is stated as the cause of AD in people over 60 years of age, while approximately 50–75% of patients with dementia develop AD [2]. Furthermore, diabetes, hypertension, and cardiovascular disease have also been identified as risk factors in the progression of AD [3]. Currently, therapeutic interventions are being attempted based on mechanisms that have been proposed in AD pathogenesis. Some of these include antioxidant therapy, NSAIDs, cholinergic replacement therapy, hormone replacement therapy, memantine, and A β vaccines [4].

Sphingolipids are abundantly distributed in nerve cell membranes and myelin sheaths of nerve fibers [5]. The bioactive metabolites of sphingolipids, including ceramide, sphingomyelin, sphingosine, and sphingosine-1-phosphate (S1P) are synthesized by a variety of enzymatic biosynthesis pathways [6]. Generally, sphingolipids play a significant role in biological membranes, and their metabolites are involved in the regulation of various cell functions. Several reports have suggested that the metabolism of sphingolipids plays a major role in the pathogenesis of AD, and hence might be considered a potential treatment

target [5,7,8]. Additionally, small changes in sphingolipid metabolism may cause remarkable effects in age-related neurodegenerative diseases. In this regard, some studies have demonstrated that the level of ceramides is significantly increased in the brain tissue of patients with AD compared to controls, whereas sphingomyelin and sphingosine-1-phosphate are decreased [5–7]. In addition, abnormal expression of enzymes in the sphingolipid synthesis pathway have also been described previously [9]. Moreover, plasma sphingolipids are considered as potential biomarkers of neurodegenerative diseases [10]. In particular, one study revealed that the levels of sphingomyelin and ceramide were altered in the plasma of patients with AD as compared to the control group [11]. The study was further supported by another report, in which an increase in sphingomyelin levels was detected in the cerebrospinal fluid of patients with AD [12].

Different bioactive molecules of sphingolipids exhibit several regulatory functions in cellular and tissue homeostasis, particularly senescence, cell cycle, migration, proliferation, autophagy, inflammation, and immune responses [6]. While these mediators are abundant in the central nervous system, any dysfunction of sphingolipids may contribute to neurodegenerative disorders. Especially, ceramide is one of the simplest sphingolipids which is widely distributed in animal tissues, while other sphingolipids are derivatives of ceramide [13]. Therefore, in this review, we aimed to discuss the impact of ceramide, a bioactive metabolite of sphingolipids, on the progression and pathogenesis of AD. Moreover, this review mentioned a list of therapeutics that are suggested to mediate ceramide synthesis pathway and prevent the disease progression.

2. Pathogenesis of Alzheimer's Disease

Although the pathogenesis of AD is not clear, it is known that the disease involves several factors, including neurotransmitters, immunity, hereditary and environmental factors [14]. Currently, researchers have proposed different theories regarding AD, namely, the amyloid beta theory, tau theory, oxidative stress, mitochondrial dysfunction, inflammation, autophagy dysfunction, and nerve and blood vessel theory [2,14]. Among these, the concept of A β -containing senile plaque formation has previously been described by many researchers. A β is not deposited in the brain under normal conditions, as it is generally cleared by the homeostatic clearance mechanism. However, an imbalance in the rate of A β production and clearance results in the formation of amyloid plaques. A previous study revealed that A β deposition in the brain is associated with cognitive impairment in the elderly [15]. Therefore, preventing A β sedimentation could be a potential method to ameliorate AD. In contrast, the development of neurofibrillary tangles by hyperphosphorylated tau protein is a fundamental neuropathological marker of AD [16]. Abundantly distributed in neurons, the tau protein plays a role in maintaining the stability of cytoskeletal microtubules and axonal transport [17]. Nevertheless, due to chemical dysfunction in AD, tau tends to separate from microtubules and attach to other tau molecules to form tangles, blocking the transport of neurons [18]. Consequently, obstruction of synaptic communication between neurons promotes the progression and pathogenesis of AD. With regards to the synapses, previous report confirmed that during AD the synaptic plasticity changes and causes reduced synapses in the brain [14]. Collectively, the deposition of A β , aggregation of tau protein, and synaptic loss cause neuronal injury followed by neuronal death and contribute to cognitive impairment.

Chemokines are a group of secreted proteins mostly known as a regulator of cell migration, especially leukocytes. The chemokines are associated with AD and considered as a key factor in the pathology of AD because of their involvement in the regulation of inflammatory or glial cells. A previous report suggested that the level of chemokines in the brain, cerebrospinal fluid, and serum constantly fluctuate in patients with AD [14]. Due to their role in regulating proinflammatory and anti-inflammatory properties, chemokines cause neuroinflammation in the AD brain, subsequently leading to neuronal death. Microglia, a type of glial cell regulated by chemokines function as an immune defense in the central nervous system. However, previous study has reported that microglia are

unable to clear waste and toxins from the brain, and their accumulation can cause chronic neuroinflammation [18]. In addition, brain cells are dependent on mitochondrial oxidative phosphorylation for their energy. However, gene set enrichment analysis has shown significant downregulation of mitochondrial oxidative phosphorylation and interruption of the mitochondrial import pathways in AD [19]. As such, mitochondrial dysfunction may also be correlated with the pathogenesis of AD. Besides, autophagy is an important cell survival mechanism that facilitates bioenergetic homeostasis. Researchers have further discussed the involvement of autophagy dysfunction in the pathogenesis of AD, as increased autophagy causes the accumulation of A β in the brain [20].

Another key regulator of brain function is lipid that has been increasingly involved in AD. It was previously observed that in AD patients, heterogeneous changes in lipid metabolism occurs in different regions of the brain [21]. Especially, lipid composition in the neuron is able to regulate the activity of membrane-bound proteins including BACE1, APP and presenilin thereby adjust the amyloid beta levels [21]. Accordingly, Grimm et al. reported that the level of cholesterol and sphingomyelin in the membrane can regulate the γ -secretase activity [22]. Cholesterol also accumulated in nerve terminals and A β plaques in human AD brain and APP transgenic mice [23]. Furthermore, production and accumulation of A β is stimulated due the elevation in cellular cholesterol level [24]. On the other hand, A β is reported to be modulating the phospholipase activity and exert its cytotoxic effects by disturbing the cell membranes [21]. Another lipid named ganglioside GM1 can regulate the pathogenic potential of A β by controlling susceptibility to aggregate [25]. Therefore, altered lipid metabolism could be an important contributor in the pathogenesis of AD.

In short, the association of amyloid beta toxicity, tau protein aggregation, synaptic damage, mitochondrial dysfunction, oxidative damage, neuroinflammation, and autophagy dysfunction complicates the AD pathogenesis.

3. Ceramide Synthesis Pathways

3.1. *De Novo Synthesis*

De novo synthesis of ceramide occurs at the cytosolic leaflet of the endoplasmic reticulum. The reaction is primarily initiated by the condensation of cytosolic serine and palmitoyl CoA by serine palmitoyltransferase to produce 3-ketodihydrosphingosine [26]. In turn, 3-ketodihydrosphingosine is reduced to dihydrosphingosine by 3-ketodihydrosphingosine reductase, followed by the production of dihydroceramide by dihydroceramide synthase [27]. Subsequently, dihydroceramide is catalyzed by dihydroceramide desaturase to form ceramide, which is transported to the Golgi apparatus (Figure 1) [28].

3.2. *Salvage Pathway*

Sphingomyelin and glucosylceramide are produced by the degradation of sphingolipids and glycosphingolipids in acidic subcellular compartments, lysosomes, and/or late endosomes [29]. Subsequently, sphingomyelin is cleaved to produce ceramide through the action of acid sphingomyelinase (ASM) [30]. In contrast, glucosylceramide is converted into ceramide by the acid β -glucosidase 1 enzyme [29]. Furthermore, lysosomal ceramides produced by these processes are deacetylated by acid ceramidase to produce sphingosine, which can eventually form sphingosine-1-phosphate or ceramide [31]. Ceramide also acts as a substrate to produce sphingosine, a product of sphingolipid catabolism [32]. Thus, this pathway of sphingosine recycling is termed the “salvage pathway,” and is responsible for regulating the formation of ceramide.

3.3. *Sphingomyelin Hydrolysis*

Sphingomyelin (SM) is an abundant sphingolipid in the plasma membrane and can be degraded into ceramide and phosphocholine. There are three types of SMase that catalyzes sphingomyelin namely, acid sphingomyelinase, neutral sphingomyelinase and alkaline while SM hydrolysis involves catalysis by neutral sphingomyelinase [27]. Additionally, a

previous report suggested that in some cells, ionizing radiation activates sphingomyelinase in the cell membrane to produce ceramide [33].

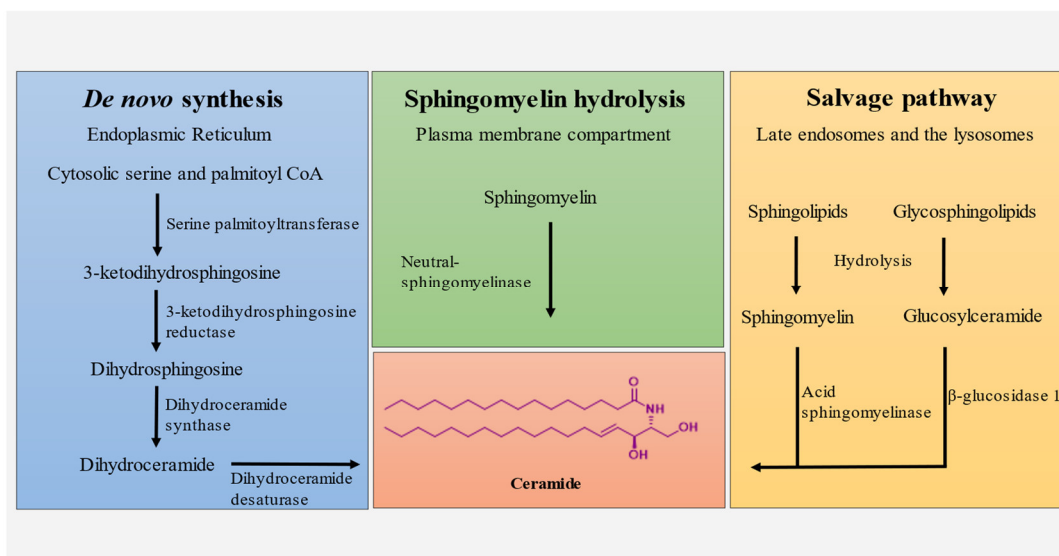


Figure 1. Ceramide synthesis pathways; de novo synthesis, sphingomyelin hydrolysis and salvage pathway.

4. The Role of Ceramide in AD Cell Lines

4.1. Amyloid Beta Plaques

Amyloid beta peptide ($A\beta$) is known to originate from the transmembrane protein amyloid beta precursor protein (APP). APP is cleaved by a beta-secretase enzyme called beta-site APP cleaving enzyme 1 (BACE1), releasing the C99 fragment of APP, which eventually results in the production of the $A\beta$ peptide. Subsequently, these peptides are secreted into the extracellular region of the brain and transported by the cerebrospinal fluid. $A\beta$ exhibits several beneficial roles in human physiology, including tumor suppression, regulation of synaptic function, and recovery from BBB (blood brain barrier) leakage and brain injury [34]. However, abnormal accumulation of $A\beta$ peptide in the brain has been reported to be the main pathogenic cause of disruption of the neuronal cell function which triggers Alzheimer's disease [34]. Moreover, it has been hypothesized that during AD, the level of secreted $A\beta$ peptide is gradually elevated in the extracellular space and forms insoluble amyloid fibrils that are resistant to degradation [35]. Interestingly, a previous report suggested that the level of ceramides in the brains of patients with AD is increased three-fold compared to age-matched controls [36]. In addition, many studies have revealed the progressive elevation of ceramide levels throughout aging in both cultured cells and the whole brain samples [37]. Thus, ceramide might be capable of regulating the production of $A\beta$ and increasing the risk of AD. In particular, a study of C6-ceramide reported that this cell-permeable analog upregulated the generation of $A\beta$ in Chinese hamster ovary cells [38]. Furthermore, the authors demonstrated that C6-ceramide promotes the β -cleavage of APP and regulates the molecular stability of BACE1, thereby increasing the rate of $A\beta$ biosynthesis [38] (Figure 2). Accordingly, the action of ceramide on BACE1 might be a strong pathway through which $A\beta$ generation increases and forms $A\beta$ plaques during AD.

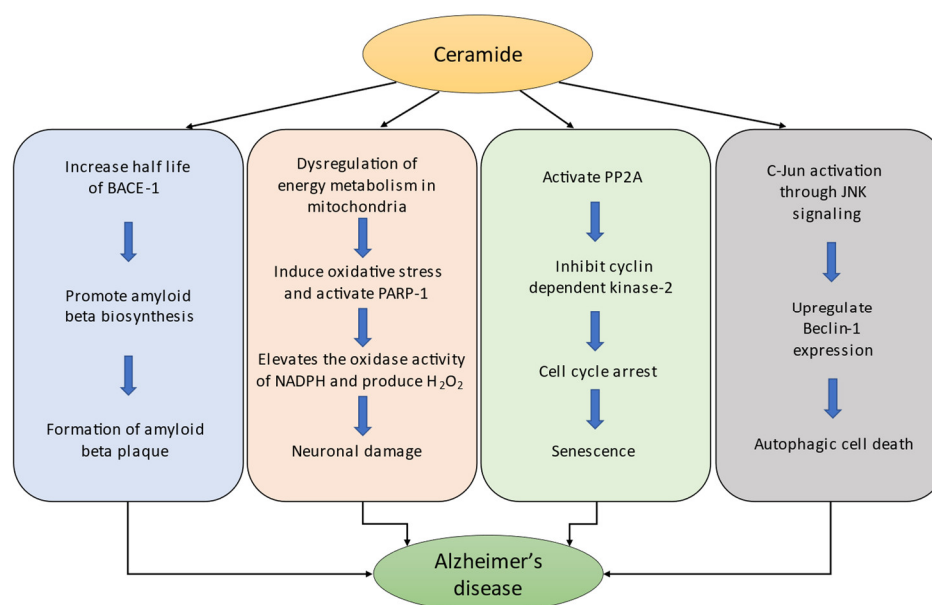


Figure 2. Mechanistic role of ceramide in the pathogenesis of AD.

4.2. Mitochondrial Dysfunction

In neurons, the normal functional roles of mitochondria include regulating calcium homeostasis, neurotransmission, membrane excitability management, and plasticity. In addition, myriad neuronal cell functions, including calcium buffering and ATP production, are also dependent on mitochondria [39]. Thus, defective mitochondrial function is anticipated to cause cellular dysfunction and contribute to disease progression. Similarly, prolonged mitochondrial damage induces dysregulation of energy metabolism, which can cause reduced energy (ATP) production, calcium buffering, and increased reactive oxygen species (ROS) [40]. Previously, ceramide is reported to modulate mitochondrial function and oxidative phosphorylation wherein, certain ceramides are involved with reduced mitochondrial respiratory chain (MRC) activity, elevated ROS production, oxidative stress and decreased mitochondrial membrane potential [41]. Many studies have suggested that ceramides act locally on the mitochondria rather than in any other cell organelle [41–43]. Consequently, ceramide may directly contribute to the generation of free radicals in the mitochondria, leading to mitochondrial dysfunction and apoptosis [42]. This was further confirmed by another study which reported that mitochondrial dysregulation by ceramide may induce oxidative stress and activate the apoptotic indicator poly (ADP-ribose) polymerase-1 (PARP-1) [41]. Moreover, ceramide-induced oxidative stress elevates the oxidase activity of NADPH and produces H_2O_2 which causes neuronal damage [44] (Figure 2). In contrast, the central nervous system is reported to be affected most frequently by mitochondrial damage, which eventually leads to cognitive decline and dementia [43]. A recent investigation showed that disrupted mitochondrial function impairs neuronal stem cell self-renewal and causes defects in neurogenesis, differentiation, and neuronal survival [45]. Therefore, the irregular action of ceramide on mitochondria might be a possible mechanism of mitochondrial dysregulation, which eventually contributes to the progression and pathogenesis of AD.

4.3. Senescence

Senescence is an irreversible cellular process in which cells lose the ability to proliferate. Many factors are involved in the initiation of cellular senescence, including organelle stress, DNA damage, oncogene activation, and telomere dysfunction [46]. In addition, many studies have suggested that senescence is regulated by different sphingolipids [37,47,48]. Particularly, one study reported an increase in the level of ceramides in old senescent WI-38 human fibroblasts as compared to young fibroblasts [49]. Their investigation showed

that ceramide significantly inhibited the growth of WI-38 human diploid fibroblasts. Similar cell growth arrest and differentiation were observed in senescent cells by another study [48]. Ceramide has also been reported to activate PP2A (protein phosphatase 2A-like protein phosphatase), a family of enzymes that regulate cellular processes and signal transduction pathways [50]. Study have suggested that C6-ceramide directly activates PP2A, which causes subsequent inhibition of cyclin-dependent kinase-2 and leads to cell cycle arrest [51] (Figure 2). Notably, ceramide might also affect tau hyperphosphorylation through the modulation of PP2A activity and contribute to the progression of AD [42]. In addition, a hydrolase enzyme, senescence-associated beta-galactosidase (SA- β -gal), allows the identification of cell senescence in mammalian tissue. One report demonstrated that C6-ceramide-induced increased expression of SA- β -gal in WI-38 cells in a time- and dose-dependent manner [37]. Collectively, these results support the significant contribution of ceramide to cellular senescence through different mechanisms.

Senescence has been associated with several pathologies, including cancer, type-2 diabetes, fibrosis, atherosclerosis, and Alzheimer's disease [47]. Besides, different studies have described the role of senescence in the pathology of AD. Features of specific cells during AD exhibit features indicative of cells that undergoes senescence. For example, telomeric alterations and hyperproliferation-mediated DNA damage in cellular senescence have been observed in AD as well [52]. Particularly, one study demonstrated the correlation between AD and DNA double strand breaks where the authors have observed an increased γ -H2A.X (Ser139) positive neurons and GFAP positive astrocytes in the hippocampus of human AD postmortem brains compared to non-AD brains [53]. These results indicate the accumulation of DNA double strand breaks during AD condition which activates DDR (DNA damage response) and lead to senescence [53]. A previous study suggested that the removal of senescent cells in aged mice elevates their lifespan and decreases the likelihood of age-related diseases, including AD [54]. Similarly, another study compared the transcriptome in young and old mice hippocampi and reported that senescence markers were accumulated in microglia [55]. Moreover, the elimination of senescent microglia significantly decreased neuroinflammation and restrains cognitive decline [55]. This evidence supports the possibility that cellular senescence by ceramide increases the progression of AD.

4.4. Autophagy Dysfunction

Autophagy is an intracellular bulk degradation process that targets the cytosolic content and organelles in mammalian cells. Based on differences in functions and mechanisms, three types of autophagy have been described: microautophagy, macroautophagy, and chaperone-mediated autophagy [56]. The biogenesis of autophagy involves several steps, including phagophore membrane isolation, autophagosome maturation, prolongation of phagophore, and cytoplasmic content engulfment [57]. Although autophagy induction is largely triggered by cellular starvation, ceramide has also been reported to be involved in autophagy activation [58]. Several studies have confirmed that ceramide induces lethal autophagy. Scarlatti et al. demonstrated that C2 ceramide increased the expression of Beclin-1 and induced lethal autophagy [59]. Similarly, Qian et al. showed that upregulation of Beclin-1 contributed to lethal autophagy, which could be prevented by the administration of the autophagy inhibitor 3-MA [60]. In another study, the mechanism of lethal autophagy was investigated, and it was found that the c-Jun protein was activated through JNK signaling by ceramide and upregulated the expression of Beclin-1 (Figure 2) [61]. Furthermore, ceramide-induced autophagic cell death was intercepted through the inhibition of JNK activity by the administration of SP600125 [61].

Autophagic dysfunction has been reported to contribute to neurodegenerative disorders by triggering neuronal cell death [62]. A previous report suggested that autophagy dysfunction significantly contributes to the pathophysiology of AD [63]. One study by Nixon et al. found a significant accumulation of autophagosomes in the frontal cortex of AD patients in comparison to controls [64]. In addition, Yu et al. demonstrated that

APP residing in autophagic vacuoles can produce A β , which may act as a source of A β production in AD [65]. Interestingly, the production and release of A β , as well as the formation of plaques, are dependent on the induction of autophagy [66]. To further understand the role of autophagy in A β secretion, Nilsson et al. generated autophagy-deficient APP transgenic mice, and observed that the secretion of A β decreased by 90% [67]. Moreover, autophagy-lysosome dysfunction is reported to stimulate tau formation and tau species accumulation which further contributes to the pathology of AD [68]. These reports confirm a strong role of ceramide in autophagy dysfunction to exacerbate the pathogenesis of AD.

5. Role of Ceramide in Plasma on AD

Ceramides in the blood circulation are transported by lipoproteins. Dysregulation in the plasma concentration of ceramide is reported in many disorders. Likewise, ceramide dysregulation has previously been reported to be detected in the circulation of patients with AD in several studies. Accordingly, it has been hypothesized that plasma ceramide may be correlated with AD, and thus it might act as a potential target to prevent the disease progression. Particularly, one report suggested that elevated plasma ceramide levels are associated with the increased risk of cognitive impairment and AD among cognitively normal individual [69]. Besides, ceramides are also associated with platelet activation and endothelial dysfunction through the detachment of NO signaling pathway [70]. It was previously reported that platelets are the major source of both APP and A β in human blood [71]. Therefore, it is possible that the activation of platelet by plasma ceramide might contribute to the pathogenesis of AD by elevating the level of A β . Various reports have proposed that the pathogenesis of systemic inflammation and insulin resistance is associated with plasma ceramide [69,72,73]. In addition, plasma ceramides are linked with proinflammatory cytokines in patients with type-2 diabetes, obesity, and cardiovascular disease [73]. For example, the production and release of proinflammatory cytokines are stimulated by LDL-ceramide following accumulation of ceramide linked to JNK and NF- κ B signaling [74]. Consequently, elevated levels of LDL-ceramide exert proinflammatory effects on macrophages and promotes inflammation in obesity [74]. These cytokines cross the blood-brain barrier and activate inflammatory pathways in the hypothalamus to cause dysregulation in brain homeostasis [75]. Several reports have suggested that patients with type-2 diabetes develop cognitive impairment, wherein insulin resistance is initiated in the brains of patients with AD [73,76,77].

Moreover, a report have demonstrated the relationship between ceramide and depression, where the ceramide pathway is considered as a target for antidepressants [78]. Xing et al. showed that plasma ceramide levels were related to depression in moderate-to-severe AD [79]. Another similar study reported that patients with anxiety and major depressive disorder progress hippocampal atrophy which is supported by a report where an increase in the level of proinflammatory cytokines tumor necrosis factor- α , interleukin-6 and interleukin-1 is seen in MDD patients [80]. Collectively, ceramide is responsible for initiating depression while major depressive disorder increases the risk of developing AD and other forms of dementia.

Overall, ceramide has been correlated with neurodegenerative diseases, and alterations in plasma ceramide levels have been identified in the cerebrospinal fluid of patients with AD [73]. In a recent report, Mielke et al. showed that an increased level of serum Cer16:0 and Cer24:0 is associated with a higher risk of developing AD [10]. In addition, clinical studies and laboratory and animal investigations have shown that an imbalance in plasma sphingomyelin and ceramide levels causes the progression of AD through amyloid beta formation and subsequent neurodegeneration [10].

6. Therapeutics Targeting Ceramide Biosynthesis

Currently, the level of ceramide can be modulated using various therapeutic drugs. When designing these agents, researchers have targeted different enzymes that contribute to the ceramide synthesis pathway. In terms of de novo synthesis of ceramide, myriocin

and fumifungin, which are structurally similar to sphingofungins, have been reported to inhibit the enzyme serine palmitoyltransferase [81] (Table 1). Mandala et al. investigated the mechanism of action of the antifungal agent lipoxamycin and revealed that lipoxamycin and hydroxylipoxamycin could potentially inhibit serine palmitoyltransferase [82]. Another compound, fumonisin B1, which is also structurally similar to that of sphingosine, has been reported to inhibit ceramide synthase in the de novo synthesis pathway [5]. In addition, dihydroceramide desaturase 1 (DES1) plays a major role in the final step of de novo ceramide biosynthesis. Triola et al. reported for the first time an effective synthesized DES1 inhibitor, GT11 (C8-cyclopropenylceramide), which is a ceramide derivative [83]. A different sphingolipid analog, XM462 (5-thiadihydroceramide), was also reported as a DES1 inhibitor to prevent ceramide biosynthesis [84]. In addition, some non-sphingolipid analogs, including fenretinide, SKI II, celecoxib, resveratrol, curcumin, and Δ 9-tetrahydrocannabinol (THC), have been reported to exhibit inhibitory activity against DES1 [85].

Table 1. List of drugs targeting different ceramide biosynthesis pathways.

Pathway	Targets	Drugs	Reference
De novo synthesis	Serine palmitoyltransferase	Myriocin	[81]
		Fumifungin	[86]
		Lipoxamycin	[82]
		Hydroxylipoxamycin	
	Ceramide synthase	Fumonisin B1	[5,87]
	Dihydroceramide desaturase 1	GT11 (C8-cyclopropenylceramide)	[83]
XM462 (5-thiadihydroceramide)		[84]	
Sphingomyelin hydrolysis	Neutral sphingomyelinase	Fenretinide	[85]
		SKI II	
		Celecoxib	
		Resveratrol	
		Curcumin	
		Δ 9-tetrahydrocannabinol (THC)	
Salvage pathway	Acid sphingomyelinase	Scyphostatin	[88]
		Spiroexpoxide	[89]
		Manumycin A	[90]
		Alutenusin	[91]
		Ubiquinol	[92]
		KARI 201	[93]
		ARC39	[94]
		AD2765	[95]
		SMA-7	[96]
		α -mangostin	[97]
Cowanol and Cowanin	[85]		
L- α phosphatidyl-D-myoinositol-3,5-biphosphate	[98]		
Phosphatidyl-myoinositol-3,4,5-triphosphate	[99]		
Fluoxetine			
Dextromethorphan			
Maprotilin			
Orphenadrine			
Nortriptyline			
Triflupromazine	[100]		
Sertraline			

Neutral sphingomyelinase (N-sphingomyelinase) is another enzyme, which is required for the hydrolysis of sphingomyelin to synthesize ceramide. Accordingly, N-sphingomyelinase inhibition is an effective approach to regulate ceramide levels. A previous report suggested that an isolated compound, scyphostatin, has a structural resemblance to ceramide and may inhibit N-sphingomyelinase [88]. Moreover, some other N-sphingomyelinase inhibitor analogs have been developed, namely spiroexpoxide, [89] manumycin A, [90] alutenusin, [91] and ubiquinol [92].

In addition, the hydrolysis of sphingomyelin is catalyzed by ASM (acid sphingomyelinase), indicating another target for the regulation of ceramide biosynthesis. To date, different types of ASM inhibitors have been reported, including natural, non-natural, physiological, and functional inhibitors. Besides, several research groups have reported some direct inhibitors of ASM including KARI 201 [93], ARC39 [94], AD2765 [95], and SMA-7 [96]. In addition, α -mangostin [97], cowanol, and cowanin [85] are natural ASM inhibitors, while various physiological inhibitors, including L- α -phosphatidyl-D-myoinositol-3,5-biphosphate [98] and phosphatidyl-myoinositol-3,4,5-triphosphate [99] also inhibits ASM. Indirect inhibitors of ASM, which act as functional inhibitors of acid sphingomyelinase (FIASMAS) are also available. FIASMAS consist of compounds such as fluoxetine, dextromethorphan, maprotilin, orphenadrine, nortriptyline, flupromazine, and sertraline [100]. These agents have been reported to inhibit ASM in the salvage pathway of ceramide synthesis, thereby reducing the production of ceramide as a therapeutic agent for neurological disorders.

Among these drugs many have undergone different phases of clinical trials including Fenretinide, Resveratrol, Ubiquinol, Fluoxetine, Nortriptyline and Sertraline for their investigation against recurrent neuroblastoma, cognitive change, sepsis, major depressive disorder, depression, Parkinson disease and anxiety respectively.

7. Conclusions and Future Perspective

As the population continues to age, the prevalence of AD is also increasing. To date, researchers have proposed several theories to explain the pathogenesis of AD. Based on these mechanisms various therapeutics are developed and currently available to prevent the disease progression. Among the mechanisms of AD pathology, most of the previous reports have demonstrated an association of sphingolipid and its bioactive metabolites in the progression and pathogenesis of AD. In this review, we emphasize the role of ceramide, an important bioactive lipid in the sphingolipid family, in AD pathology. Ceramide is associated with different pathological states, including neurodegeneration, diabetes, obesity, and inflammation. As described above, ceramide levels in the cell and plasma significantly control several characteristics of pathological changes in AD, including A β plaques, mitochondrial dysfunction, senescence, autophagy dysfunction, platelet activation and endothelial dysfunction. Moreover, unlike other sphingolipids, ceramide directly participates in initiating the abovementioned pathological changes to cause AD. Hence, ceramide biosynthesis pathways and the participating enzymes could be potential targets to regulate ceramide levels and prevent disease progression. Several studies have suggested the use of different types of drugs to inhibit specific enzymes in the ceramide synthesis pathway. Consequently, the modulation of ceramide synthesis pathway significantly controlled the level of ceramide and decreased AD progression. Nevertheless, owing to the lack of knowledge regarding their safety and efficacy, the search for potential targets and remedies remains a major point of attention.

Author Contributions: Conceptualization, J.-s.B.; writing—original draft preparation, M.R.C.; writing—review and editing, H.K.J. and J.-s.B. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by the National Research Foundation of Korea (NRF) grant funded by the Korea government (MSIT) (2018M3C7A1056513, 2020R1A2C3006875, 2020R1A2C3006734).

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Jackson, G.A. Dementia: Alzheimer's Disease. *Nurs. Times* **2002**, *98*, 32–34. [[CrossRef](#)] [[PubMed](#)]
2. Srivastava, S.; Ahmad, R.; Khare, S.K. Alzheimer's disease and its treatment by different approaches: A review. *Eur. J. Med. Chem.* **2021**, *216*, 113320. [[CrossRef](#)]
3. Galle, S.A.; Geraedts, I.; Deijen, J.; Milders, M.; Drent, M. The Interrelationship between Insulin-Like Growth Factor 1, Apolipoprotein E E4, Lifestyle Factors, and The Aging Body and Brain. *J. Prev. Alzheimer's Dis.* **2020**, *7*, 265–273. [[CrossRef](#)] [[PubMed](#)]

4. Shah, R.S.; Lee, H.-G.; Xiongwei, Z.; Perry, G.; Smith, M.A.; Castellani, R.J. Current approaches in the treatment of Alzheimer's disease. *Biomed. Pharmacother.* **2008**, *62*, 199–207. [[CrossRef](#)] [[PubMed](#)]
5. Brodowicz, J.; Przegaliński, E.; Müller, C.P.; Filip, M. Ceramide and Its Related Neurochemical Networks as Targets for Some Brain Disorder Therapies. *Neurotox. Res.* **2017**, *33*, 474–484. [[CrossRef](#)]
6. Park, M.H.; Jin, H.K.; Bae, J.-S. Potential therapeutic target for aging and age-related neurodegenerative diseases: The role of acid sphingomyelinase. *Exp. Mol. Med.* **2020**, *52*, 380–389. [[CrossRef](#)] [[PubMed](#)]
7. Chakrabarti, S.S.; Bir, A.; Poddar, J.; Sinha, M.; Ganguly, A.; Chakrabarti, S. Ceramide and Sphingosine-1-Phosphate in Cell Death Pathways: Relevance to the Pathogenesis of Alzheimer's Disease. *Curr. Alzheimer Res.* **2016**, *13*, 1232–1248. [[CrossRef](#)] [[PubMed](#)]
8. Kitatani, K.; Idkowiak-Baldys, J.; Hannun, Y.A. The sphingolipid salvage pathway in ceramide metabolism and signaling. *Cell. Signal.* **2008**, *20*, 1010–1018. [[CrossRef](#)]
9. Crivelli, S.M.; Giovagnoni, C.; Visseren, L.; Scheithauer, A.-L.; de Wit, N.; Hoedt, S.D.; Losen, M.; Mulder, M.T.; Walter, J.; de Vries, H.E.; et al. Sphingolipids in Alzheimer's disease, how can we target them? *Adv. Drug Deliv. Rev.* **2020**, *159*, 214–231. [[CrossRef](#)]
10. Mielke, M.M.; Bandaru, V.V.R.; Haughey, N.J.; Xia, J.; Fried, L.P.; Yasar, S.; Albert, M.; Varma, V.; Harris, G.; Schneider, E.B.; et al. Serum ceramides increase the risk of Alzheimer disease: The Women's Health and Aging Study II. *Neurology* **2012**, *79*, 633–641. [[CrossRef](#)]
11. Han, X.; Rozen, S.G.; Boyle, S.H.; Hellegers, C.; Cheng, H.; Burke, J.; Welsh-Bohmer, K.A.; Doraiswamy, P.M.; Kaddurah-Daouk, R. Metabolomics in Early Alzheimer's Disease: Identification of Altered Plasma Sphingolipidome Using Shotgun Lipidomics. *PLoS ONE* **2011**, *6*, e21643. [[CrossRef](#)] [[PubMed](#)]
12. Kosicek, M.; Zetterberg, H.; Andreassen, N.; Peter-Katalinic, J.; Hecimovic, S. Elevated Cerebrospinal Fluid Sphingomyelin Levels in Prodromal Alzheimer's Disease. *Neurosci. Lett.* **2012**, *516*, 302–305. [[CrossRef](#)] [[PubMed](#)]
13. Sphingolipid Biochemistry Britannica. Available online: <https://www.britannica.com/science/sphingolipid> (accessed on 12 July 2022).
14. Chen, Y.-G. Research Progress in the Pathogenesis of Alzheimer's Disease. *Chin. Med. J.* **2018**, *131*, 1618–1624. [[CrossRef](#)] [[PubMed](#)]
15. Shimada, H.; Kitamura, S.; Shinotoh, H.; Endo, H.; Niwa, F.; Hirano, S.; Kimura, Y.; Zhang, M.; Kuwabara, S.; Suhara, T.; et al. Association between A β and tau accumulations and their influence on clinical features in aging and Alzheimer's disease spectrum brains: A [11 C]PBB3-PET study. *Alzheimer's Dement. Diagn. Assess. Dis. Monit.* **2016**, *6*, 11–20. [[CrossRef](#)]
16. Kempf, S.J.; Metaxas, A. Neurofibrillary tangles in Alzheimer's disease: Elucidation of the molecular mechanism by immunohistochemistry and tau protein phospho-proteomics. *Neural Regen. Res.* **2016**, *11*, 1579–1581. [[CrossRef](#)]
17. Poorkaj, P.; Kas, A.; D'Souza, I.; Zhou, Y.; Pham, Q.; Stone, M.; Olson, M.V.; Schellenberg, G.D. A genomic sequence analysis of the mouse and human microtubule-associated protein tau. *Mamm. Genome* **2001**, *12*, 700–712. [[CrossRef](#)] [[PubMed](#)]
18. What Happens to the Brain in Alzheimer's Disease? National Institute on Aging. Available online: <https://www.nia.nih.gov/health/what-happens-brain-alzheimers-disease> (accessed on 10 April 2022).
19. Wang, W.; Zhao, F.; Ma, X.; Perry, G.; Zhu, X. Mitochondria dysfunction in the pathogenesis of Alzheimer's disease: Recent advances. *Mol. Neurodegener.* **2020**, *15*, 30. [[CrossRef](#)] [[PubMed](#)]
20. Lee, S.; Sato, Y.; Nixon, R.A. Primary lysosomal dysfunction causes cargo-specific deficits of axonal transport leading to Alzheimer-like neuritic dystrophy. *Autophagy* **2011**, *7*, 1562–1563. [[CrossRef](#)] [[PubMed](#)]
21. Chan, R.B.; Oliveira, T.G.; Cortes, E.P.; Honig, L.S.; Duff, K.E.; Small, S.A.; Wenk, M.R.; Shui, G.; Di Paolo, G. Comparative Lipidomic Analysis of Mouse and Human Brain with Alzheimer Disease. *J. Biol. Chem.* **2012**, *287*, 2678–2688. [[CrossRef](#)]
22. Grimm, M.; Grimm, H.S.; Pätzold, A.J.; Zinser, E.G.; Halonen, R.; Duering, M.; Tschäpe, J.-A.; De Strooper, B.; Müller, U.; Shen, J.; et al. Regulation of cholesterol and sphingomyelin metabolism by amyloid- β and presenilin. *Nat. Cell Biol.* **2005**, *7*, 1118–1123. [[CrossRef](#)] [[PubMed](#)]
23. Mori, T.; Paris, D.; Town, T.; Rojiani, A.M.; Sparks, D.L.; Delledonne, A.; Crawford, F.; Abdullah, L.I.; Humphrey, J.A.; Dickson, D.W.; et al. Cholesterol Accumulates in Senile plaques of Alzheimer Disease patients and in Transgenic APPswMice. *J. Neuropathol. Exp. Neurol.* **2001**, *60*, 778–785. [[CrossRef](#)] [[PubMed](#)]
24. Fassbender, K.; Simons, M.; Bergmann, C.; Stroick, M.; Lütjohann, D.; Keller, P.; Runz, H.; Kühl, S.; Bertsch, T.; von Bergmann, K.; et al. Simvastatin Strongly Reduces Levels of Alzheimer's Disease β -Amyloid Peptides A β 42 and A β 40 In Vitro and In Vivo. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 5856–5861. [[CrossRef](#)]
25. Matsuzaki, K.; Kato, K.; Yanagisawa, K. A β polymerization through interaction with membrane gangliosides. *Biochim. Biophys. Acta (BBA) Mol. Cell Biol. Lipids* **2010**, *1801*, 868–877. [[CrossRef](#)] [[PubMed](#)]
26. Mandon, E.; Ehses, I.; Rother, J.; van Echten, G.; Sandhoff, K. Subcellular localization and membrane topology of serine palmitoyltransferase, 3-dehydrosphinganine reductase, and sphinganine N-acyltransferase in mouse liver. *J. Biol. Chem.* **1992**, *267*, 11144–11148. [[CrossRef](#)]
27. Rao, R.P.; Vaidyanathan, N.; Rengasamy, M.; Oommen, A.M.; Somaiya, N.; Jagannath, M.R. Sphingolipid Metabolic Pathway: An Overview of Major Roles Played in Human Diseases. *J. Lipids* **2013**, *2013*, 1–12.
28. Sugimoto, Y.; Sakoh, H.; Yamada, K. IPC Synthase as a Useful Target for Antifungal Drugs. *Curr. Drug Targets Infect. Disord.* **2004**, *4*, 311–322. [[CrossRef](#)] [[PubMed](#)]
29. Kitatani, K.; Sheldon, K.; Rajagopalan, V.; Anelli, V.V.; Jenkins, R.W.; Sun, Y.; Grabowski, G.A.; Obeid, L.; Hannun, Y.A. Involvement of Acid β -Glucosidase 1 in the Salvage Pathway of Ceramide Formation. *J. Biol. Chem.* **2009**, *284*, 12972–12978. [[CrossRef](#)]

30. Quintern, L.; Schuchman, E.; Levran, O.; Suchi, M.; Ferlinz, K.; Reinke, H.; Sandhoff, K.; Desnick, R. Isolation of cDNA clones encoding human acid sphingomyelinase: Occurrence of alternatively processed transcripts. *EMBO J.* **1989**, *8*, 2469–2473. [[CrossRef](#)] [[PubMed](#)]
31. Smith, E.R.; Merrill, A.H. Differential Roles of de Novo Sphingolipid Biosynthesis and Turnover in the “Burst” of Free Sphingosine and Sphinganine, and Their 1-Phosphates and N-Acyl-Derivatives, That Occurs upon Changing the Medium of Cells in Culture (*). *J. Biol. Chem.* **1995**, *270*, 18749–18758. [[CrossRef](#)] [[PubMed](#)]
32. Moro, K.; Nagahashi, M.; Gabriel, E.; Takabe, K.; Wakai, T. Clinical application of ceramide in cancer treatment. *Breast Cancer* **2019**, *26*, 407–415. [[CrossRef](#)] [[PubMed](#)]
33. Kolter, T.; Sandhoff, K. Principles of Lysosomal Membrane Digestion: Stimulation of Sphingolipid Degradation by Sphingolipid Activator Proteins and Anionic Lysosomal Lipids. *Annu. Rev. Cell Dev. Biol.* **2005**, *21*, 81–103. [[CrossRef](#)] [[PubMed](#)]
34. Brothers, H.M.; Gosztyla, M.; Robinson, S.R. The Physiological Roles of Amyloid- β Peptide Hint at New Ways to Treat Alzheimer’s Disease. *Front. Aging Neurosci.* **2018**, *10*, 118. [[CrossRef](#)] [[PubMed](#)]
35. Takahashi, R.H.; Nagao, T.; Gouras, G.K. Plaque formation and the intraneuronal accumulation of β -amyloid in Alzheimer’s disease. *Pathol. Int.* **2017**, *67*, 185–193. [[CrossRef](#)] [[PubMed](#)]
36. Han, X.; Holtzman, D.M.; McKeel, D.W., Jr.; Kelley, J.; Morris, J.C. Substantial sulfatide deficiency and ceramide elevation in very early Alzheimer’s disease: Potential role in disease pathogenesis. *J. Neurochem.* **2002**, *82*, 809–818. [[CrossRef](#)] [[PubMed](#)]
37. Mouton, R.E.; Venable, M.E. Ceramide induces expression of the senescence histochemical marker, β -galactosidase, in human fibroblasts. *Mech. Ageing Dev.* **2000**, *113*, 169–181. [[CrossRef](#)]
38. Puglielli, L.; Ellis, B.C.; Saunders, A.J.; Kovacs, D.M. Ceramide Stabilizes β -Site Amyloid Precursor Protein-cleaving Enzyme 1 and Promotes Amyloid β -Peptide Biogenesis. *J. Biol. Chem.* **2003**, *278*, 19777–19783. [[CrossRef](#)] [[PubMed](#)]
39. Liu, F.; Lu, J.; Manaenko, A.; Tang, J.; Hu, Q. Mitochondria in Ischemic Stroke: New Insight and Implications. *Aging Dis.* **2018**, *9*, 924–937. [[CrossRef](#)]
40. Norat, P.; Soldozy, S.; Sokolowski, J.D.; Gorick, C.M.; Kumar, J.S.; Chae, Y.; Yağmurlu, K.; Prada, F.; Walker, M.; Levitt, M.R.; et al. Mitochondrial dysfunction in neurological disorders: Exploring mitochondrial transplantation. *NPJ Regen. Med.* **2020**, *5*, 1–9. [[CrossRef](#)]
41. Kogot-Levin, A.; Saada, A. Ceramide and the mitochondrial respiratory chain. *Biochimie* **2014**, *100*, 88–94. [[CrossRef](#)] [[PubMed](#)]
42. Jazvinšćak Jembrek, M.; Hof, P.R.; Šimić, G. Ceramides in Alzheimer’s Disease: Key Mediators of Neuronal Apoptosis Induced by Oxidative Stress and A β Accumulation. *Oxidative Med. Cell. Longev.* **2015**, *2015*, 346783. [[CrossRef](#)] [[PubMed](#)]
43. Finsterer, J. Cognitive dysfunction in mitochondrial disorders. *Acta Neurol. Scand.* **2012**, *126*, 1–11. [[CrossRef](#)] [[PubMed](#)]
44. Adams, J. Probable Causes of Alzheimer’s Disease. *Science* **2021**, *3*, 16. [[CrossRef](#)]
45. Khacho, M.; Clark, A.; Svoboda, D.S.; Maclaurin, J.G.; Lagace, D.C.; Park, D.; Slack, R.S. Mitochondrial dysfunction underlies cognitive defects as a result of neural stem cell depletion and impaired neurogenesis. *Hum. Mol. Genet.* **2017**, *26*, 3327–3341. [[CrossRef](#)]
46. Di Micco, R.; Krizhanovsky, V.; Baker, D.; d’Adda di Fagagna, F. Cellular Senescence in Ageing: From Mechanisms to Therapeutic Opportunities. *Nat. Rev. Mol. Cell Biol.* **2021**, *22*, 75–95. [[CrossRef](#)]
47. Trayssac, M.; Hannun, Y.A.; Obeid, L.M. Role of sphingolipids in senescence: Implication in aging and age-related diseases. *J. Clin. Investig.* **2018**, *128*, 2702–2712. [[CrossRef](#)]
48. Peacocke, M.; Campisi, J. Cellular senescence: A reflection of normal growth control, differentiation, or aging? *J. Cell. Biochem.* **1991**, *45*, 147–155. [[CrossRef](#)]
49. Venable, M.E.; Lee, J.Y.; Smyth, M.J.; Bielawska, A.; Obeid, L.M. Role of Ceramide in Cellular Senescence. *J. Biol. Chem.* **1995**, *270*, 30701–30708. [[CrossRef](#)]
50. Chalfant, C.E.; Kishikawa, K.; Mumby, M.C.; Kamibayashi, C.; Bielawska, A.; Hannun, Y.A. Long Chain Ceramides Activate Protein Phosphatase-1 and Protein Phosphatase-2A. *J. Biol. Chem.* **1999**, *274*, 20313–20317. [[CrossRef](#)]
51. Lee, J.; Bielawska, A.; Obeid, L. Regulation of Cyclin-Dependent Kinase 2 Activity by Ceramide. *Exp. Cell Res.* **2000**, *261*, 303–311. [[CrossRef](#)]
52. Mahady, L.J.; He, B.; Malek-Ahmadi, M.; Mufson, E.J. Telomeric alterations in the default mode network during the progression of Alzheimer’s disease: Selective vulnerability of the precuneus. *Neuropathol. Appl. Neurobiol.* **2020**, *47*, 428–440. [[CrossRef](#)]
53. Thadathil, N.; Delotterie, D.F.; Xiao, J.; Hori, R.; McDonald, M.P.; Khan, M.M. DNA Double-Strand Break Accumulation in Alzheimer’s Disease: Evidence from Experimental Models and Postmortem Human Brains. *Mol. Neurobiol.* **2020**, *58*, 118–131. [[CrossRef](#)] [[PubMed](#)]
54. Baker, D.J.; Childs, B.G.; Durik, M.; Wijers, M.E.; Sieben, C.J.; Zhong, J.; Saltness, R.A.; Jeganathan, K.B.; Verzosa, G.C.; Pezeshki, A.; et al. Naturally occurring p16^{Ink4a}-positive cells shorten healthy lifespan. *Nature* **2016**, *530*, 184–189. [[CrossRef](#)] [[PubMed](#)]
55. Ogrodnik, M.; Evans, S.A.; Fielder, E.; Vettorelli, S.; Kruger, P.; Salmonowicz, H.; Weigand, B.M.; Patel, A.D.; Pirtskhalava, T.; Inman, C.L.; et al. Whole-body senescent cell clearance alleviates age-related brain inflammation and cognitive impairment in mice. *Aging Cell* **2021**, *20*, e13296. [[CrossRef](#)]
56. Jiang, W.; Ogretmen, B. Autophagy paradox and ceramide. *Biochim. Biophys. Acta (BBA) Mol. Cell Biol. Lipids* **2013**, *1841*, 783–792. [[CrossRef](#)] [[PubMed](#)]
57. Lee, K.M.; Hwang, S.K.; Lee, J.A. Neuronal Autophagy and Neurodevelopmental Disorders. *Exp. Neurobiol.* **2013**, *22*, 133. [[CrossRef](#)] [[PubMed](#)]

58. Young, M.M.; Kester, M.; Wang, H.G. Sphingolipids: Regulators of Crosstalk between Apoptosis and Autophagy. *J. Lipid Res.* **2013**, *54*, 5–19. [[CrossRef](#)]
59. Scarlatti, F.; Bauvy, C.; Ventruti, A.; Sala, G.; Cluzaud, F.; Vandewalle, A.; Ghidoni, R.; Codogno, P. Ceramide-Mediated Macroautophagy Involves Inhibition of Protein Kinase B and up-Regulation of Beclin 1. *J. Biol. Chem.* **2004**, *279*, 18384–18391. [[CrossRef](#)]
60. Qian, W.; Liu, J.; Jin, J.; Ni, W.; Xu, W. Arsenic trioxide induces not only apoptosis but also autophagic cell death in leukemia cell lines via up-regulation of Beclin-1. *Leuk. Res.* **2007**, *31*, 329–339. [[CrossRef](#)]
61. Li, D.; Wang, L.; Deng, R.; Tang, J.; Shen, Y.; Guo, J.-F.; Wang, Y.; Xia, L.-P.; Feng, G.-K.; Liu, Q.-Q.; et al. The Pivotal Role of C-Jun NH2-Terminal Kinase-Mediated Beclin 1 Expression during Anticancer Agents-Induced Autophagy in Cancer Cells. *Oncogene* **2009**, *28*, 886–898. [[CrossRef](#)]
62. Koike, M.; Shibata, M.; Tadakoshi, M.; Gotoh, K.; Komatsu, M.; Waguri, S.; Kawahara, N.; Kuida, K.; Nagata, S.; Kominami, E.; et al. Inhibition of Autophagy Prevents Hippocampal Pyramidal Neuron Death after Hypoxic-Ischemic Injury. *Am. J. Pathol.* **2008**, *172*, 454–469. [[CrossRef](#)]
63. Cheng, J.; North, B.J.; Zhang, T.; Dai, X.; Tao, K.; Guo, J.; Wei, W. The emerging roles of protein homeostasis-governing pathways in Alzheimer's disease. *Aging Cell* **2018**, *17*, e12801. [[CrossRef](#)] [[PubMed](#)]
64. Nixon, R.A.; Wegiel, J.; Kumar, A.; Yu, W.H.; Peterhoff, C.; Cataldo, A.; Cuervo, A.M. Extensive Involvement of Autophagy in Alzheimer Disease: An Immuno-Electron Microscopy Study. *J. Neuropathol. Exp. Neurol.* **2005**, *64*, 113–122. [[CrossRef](#)] [[PubMed](#)]
65. Yu, J.; Pan, W.; Shi, R.; Yang, T.; Li, Y.; Yu, G.; Bai, Y.; Schuchman, E.H.; He, X.; Zhang, G. Ceramide is Upregulated and Associated with Mortality in Patients with Chronic Heart Failure. *Can. J. Cardiol.* **2015**, *31*, 357–363. [[CrossRef](#)] [[PubMed](#)]
66. Kurtishi, A.; Rosen, B.; Patil, K.S.; Alves, G.W.; Møller, S.G. Cellular Proteostasis in Neurodegeneration. *Mol. Neurobiol.* **2018**, *56*, 3676–3689. [[CrossRef](#)] [[PubMed](#)]
67. Nilsson, P.; Saido, T.C. Dual roles for autophagy: Degradation and secretion of Alzheimer's disease A β peptide. *BioEssays* **2014**, *36*, 570–578. [[CrossRef](#)]
68. Li, Q.; Liu, Y.; Sun, M. Autophagy and Alzheimer's Disease. *Cell. Mol. Neurobiol.* **2016**, *37*, 377–388. [[CrossRef](#)] [[PubMed](#)]
69. Mielke, M.M.; Haughey, N.J.; Han, D.; An, Y.; Bandaru, V.V.R.; Lyketsos, C.G.; Ferrucci, L.; Resnick, S.M. The Association between Plasma Ceramides and Sphingomyelins and Risk of Alzheimer's Disease Differs by Sex and APOE in the Baltimore Longitudinal Study of Aging. *J. Alzheimer's Dis.* **2017**, *60*, 819–828. [[CrossRef](#)]
70. Meeusen, J.W.; Donato, L.J.; Bryant, S.C.; Baudhuin, L.M.; Berger, P.B.; Jaffe, A.S. Plasma Ceramides: A Novel Predictor of Major Adverse Cardiovascular Events After Coronary Angiography. *Arterioscler. Thromb. Vasc. Biol.* **2018**, *38*, 1933–1939. [[CrossRef](#)] [[PubMed](#)]
71. Sevush, S.; Jy, W.; Horstman, L.L.; Mao, W.-W.; Kolodny, L.; Ahn, Y.S. Platelet Activation in Alzheimer Disease. *Arch. Neurol.* **1998**, *55*, 530–536. [[CrossRef](#)]
72. Kim, M.; Nevado-Holgado, A.; Whiley, L.; Snowden, S.G.; Soininen, H.; Kloszewska, I.; Mecocci, P.; Tsolaki, M.; Vellas, B.; Thambisetty, M.; et al. Association between Plasma Ceramides and Phosphatidylcholines and Hippocampal Brain Volume in Late Onset Alzheimer's Disease. *J. Alzheimer's Dis.* **2017**, *60*, 809–817. [[CrossRef](#)]
73. De Mello, V.D.F.; Lankinen, M.; Schwab, U.; Kolehmainen, M.; Lehto, S.; Seppänen-Laakso, T.; Oresic, M.; Pulkkinen, L.; Uusitupa, M.; Erkkilä, A.T. Link between plasma ceramides, inflammation and insulin resistance: Association with serum IL-6 concentration in patients with coronary heart disease. *Diabetologia* **2009**, *52*, 2612–2615. [[CrossRef](#)] [[PubMed](#)]
74. Boon, J.; Hoy, A.J.; Stark, R.; Brown, R.D.; Meex, R.C.; Henstridge, D.C.; Schenk, S.; Meikle, P.J.; Horowitz, J.F.; Kingwell, B.A.; et al. Ceramides Contained in LDL Are Elevated in Type 2 Diabetes and Promote Inflammation and Skeletal Muscle Insulin Resistance. *Diabetes* **2013**, *62*, 401–410. [[CrossRef](#)]
75. De Souza, C.T.; Araujo, E.P.; Bordin, S.; Ashimine, R.; Zollner, R.L.; Boschero, A.C.; Saad, M.J.A.; Velloso, L.A. Consumption of a Fat-Rich Diet Activates a Proinflammatory Response and Induces Insulin Resistance in the Hypothalamus. *Endocrinology* **2005**, *146*, 4192–4199. [[CrossRef](#)]
76. Cukierman, T.; Gerstein, H.C.; Williamson, J.D. Cognitive decline and dementia in diabetes—Systematic overview of prospective observational studies. *Diabetologia* **2005**, *48*, 2460–2469. [[CrossRef](#)] [[PubMed](#)]
77. Talbot, K.; Wang, H.Y.; Kazi, H.; Han, L.Y.; Bakshi, K.P.; Stucky, A.; Fuino, R.L.; Kawaguchi, K.R.; Samoyedny, A.J.; Wilson, R.S.; et al. Demonstrated Brain Insulin Resistance in Alzheimer's Disease Patients is Associated with IGF-1 Resistance, IRS-1 Dysregulation, and Cognitive Decline. *J. Clin. Investig.* **2012**, *122*, 1316–1338. [[CrossRef](#)] [[PubMed](#)]
78. Gulbins, E.; Walter, S.; Becker, K.A.; Halmer, R.; Liu, Y.; Reichel, M.; Edwards, M.J.; Müller, C.P.; Fassbender, K.; Kornhuber, J. A central role for the acid sphingomyelinase/ceramide system in neurogenesis and major depression. *J. Neurochem.* **2015**, *134*, 183–192. [[CrossRef](#)] [[PubMed](#)]
79. Xing, Y.; Tang, Y.; Zhao, L.; Wang, Q.; Qin, W.; Zhang, J.-L.; Jia, J. Plasma Ceramides and Neuropsychiatric Symptoms of Alzheimer's Disease. *J. Alzheimer's Dis.* **2016**, *52*, 1029–1035. [[CrossRef](#)] [[PubMed](#)]
80. Dowlati, Y.; Herrmann, N.; Swardfager, W.; Liu, H.; Sham, L.; Reim, E.K.; Lanctôt, K.L. A Meta-Analysis of Cytokines in Major Depression. *Biol. Psychiatry* **2010**, *67*, 446–457. [[CrossRef](#)] [[PubMed](#)]
81. Wadsworth, J.M.; Clarke, D.J.; McMahon, S.A.; Lowther, J.P.; Beattie, A.E.; Langridge-Smith, P.R.R.; Broughton, H.B.; Dunn, T.M.; Naismith, J.H.; Campopiano, D.J. The Chemical Basis of Serine Palmitoyltransferase Inhibition by Myriocin. *J. Am. Chem. Soc.* **2013**, *135*, 14276–14285. [[CrossRef](#)]

82. Mandala, S.M.; Frommer, B.R.; Thornton, R.A.; Kurtz, M.B.; Young, N.M.; Cabello, M.A.; Genilloud, O.; Liesch, J.M.; Smith, J.L.; Horn, W.S. Inhibition of serine palmitoyl-transferase activity by lipoxamycin. *J. Antibiot.* **1994**, *47*, 376–379. [[CrossRef](#)] [[PubMed](#)]
83. Triola, G.; Fabria, G.; Llebaria, A. Synthesis of a Cyclopropene Analogue of Ceramide, a Potent Inhibitor of Dihydroceramide Desaturase. *Angew. Chem. Int. Ed.* **2001**, *40*, 1960–1962. [[CrossRef](#)]
84. Munoz-Olaya, J.M.; Matabosch, X.; Bedia, C.; Egidio-Gabás, M.; Casas, J.; Llebaria, A.; Delgado, A.; Fabriàs, G. Synthesis and Biological Activity of a Novel Inhibitor of Dihydroceramide Desaturase. *ChemMedChem* **2008**, *3*, 946–953. [[CrossRef](#)] [[PubMed](#)]
85. Skácel, J.; Slusher, B.S.; Tsukamoto, T. Small Molecule Inhibitors Targeting Biosynthesis of Ceramide, the Central Hub of the Sphingolipid Network. *J. Med. Chem.* **2021**, *64*, 279–297. [[CrossRef](#)] [[PubMed](#)]
86. Zweerink, M.; Edison, A.; Wells, G.; Pinto, W.; Lester, R. Characterization of a novel, potent, and specific inhibitor of serine palmitoyltransferase. *J. Biol. Chem.* **1992**, *267*, 25032–25038. [[CrossRef](#)]
87. Merrill, A.; Schmelza, E.M.; Dillehay, D.; Spiegel, S.; Shayman, J.; Schroeder, J.; Riley, R.; Voss, K.; Wang, E. Sphingolipids—The Enigmatic Lipid Class: Biochemistry, Physiology, and Pathophysiology. *Toxicol. Appl. Pharmacol.* **1997**, *142*, 208–225. [[CrossRef](#)] [[PubMed](#)]
88. Nara, F.; Tanakaa, M.; Hosoyab, T.; Suzuki-Konagaia, K.; Ogita, T. Scyphostatin, a neutral sphingomyelinase inhibitor from a discomycete, *Trichopeziza mollissima*: Taxonomy of the producing organism, fermentation, isolation, and physico-chemical properties. *J. Antibiot.* **1999**, *52*, 525–530. [[CrossRef](#)] [[PubMed](#)]
89. Arenz, C.; Giannis, A. Synthesis of the First Selective Irreversible Inhibitor of Neutral Sphingomyelinase. *Angew. Chem. Int. Ed.* **2000**, *39*, 1440–1442. [[CrossRef](#)]
90. Arenz, C.; Thutewohl, M.; Block, O.; Waldmann, H.; Altenbach, H.-J.; Giannis, A. Manumycin A and Its Analogues are Irreversible Inhibitors of Neutral Sphingomyelinase. *Chembiochem* **2001**, *2*, 141–143. [[CrossRef](#)]
91. Uchida, R.; Tomoda, H.; Dong, Y.; Omura, S. Alutenusin, a specific neutral sphingomyelinase inhibitor, produced by *Penicillium* sp. FO-7436. *J. Antibiot.* **1999**, *52*, 572–574. [[CrossRef](#)] [[PubMed](#)]
92. Martín, S.F.; Navarro, F.; Forthoffer, N.; Navas, P.; Villalba, J.M. Neutral magnesium-dependent sphingomyelinase from liver plasma membrane: Purification and inhibition by ubiquinol. *J. Bioenerg. Biomembr.* **2001**, *33*, 143–153. [[CrossRef](#)] [[PubMed](#)]
93. Park, M.H.; Park, K.H.; Choi, B.J.; Han, W.H.; Yoon, H.J.; Jung, H.Y.; Lee, J.; Song, I.-S.; Lim, D.Y.; Choi, M.-K.; et al. Discovery of a dual-action small molecule that improves neuropathological features of Alzheimer’s disease mice. *Proc. Natl. Acad. Sci. USA* **2022**, *119*, e2115082119. [[CrossRef](#)] [[PubMed](#)]
94. Yang, K.; Yu, J.; Nong, K.; Wang, Y.; Niu, A.; Chen, W.; Dong, J.; Wang, J. Discovery of Potent, Selective, and Direct Acid Sphingomyelinase Inhibitors with Antidepressant Activity. *J. Med. Chem.* **2020**, *63*, 961–974. [[CrossRef](#)] [[PubMed](#)]
95. Darroch, P.I.; Dagan, A.; Granot, T.; He, X.; Gatt, S.; Schuchman, E.H. A Lipid Analogue That Inhibits Sphingomyelin Hydrolysis and Synthesis, Increases Ceramide, and Leads to Cell Death. *J. Lipid Res.* **2005**, *46*, 2315–2324. [[CrossRef](#)]
96. Sakata, A.; Ochiai, T.; Shimeno, H.; Hikishima, S.; Yokomatsu, T.; Shibuya, S.; Toda, A.; Eyanagi, R.; Soeda, S. Acid sphingomyelinase inhibition suppresses lipopolysaccharide-mediated release of inflammatory cytokines from macrophages and protects against disease pathology in dextran sulphate sodium-induced colitis in mice. *Immunology* **2007**, *122*, 54–64. [[CrossRef](#)] [[PubMed](#)]
97. Okudaira, C.; Ikeda, Y.; Kondo, S.; Furuya, S.; Hirabayashi, Y.; Koyano, T.; Saito, Y.; Umezawa, K. Inhibition of Acidic Sphingomyelinase by Xanthone Compounds Isolated from *Garcinia Speciosa*. *J. Enzym. Inhib.* **2000**, *15*, 129–138. [[CrossRef](#)] [[PubMed](#)]
98. Kölzer, M.; Arenz, C.; Ferlinz, K.; Werth, N.; Schulze, H.; Klingenstein, R.; Sandhoff, K. Phosphatidylinositol-3,5-Bisphosphate is a Potent and Selective Inhibitor of Acid Sphingomyelinase. *Biol. Chem.* **2003**, *384*, 1293–1298. [[CrossRef](#)] [[PubMed](#)]
99. Testai, F.D.; Landek, M.A.; Goswami, R.; Ahmed, M.; Dawson, G. Acid sphingomyelinase and inhibition by phosphate ion: Role of inhibition by phosphatidyl-myoinositol 3,4,5-triphosphate in oligodendrocyte cell signaling. *J. Neurochem.* **2004**, *89*, 636–644. [[CrossRef](#)] [[PubMed](#)]
100. Kornhuber, J.; Tripal, P.; Gulbins, E.; Muehlbacher, M. Functional Inhibitors of Acid Sphingomyelinase (FIASMA). *Sphingolipids Basic Sci. Drug Dev.* **2013**, *215*, 169–186. [[CrossRef](#)]