

Clinical impact and mechanisms of hepatitis B virus infection concurrent with non-alcoholic fatty liver disease

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Abstract

Chronic hepatitis B (CHB) virus infection is an important threat to global health despite the administration of vaccines and the use of antiviral treatments. In recent years, as the prevalence of obesity and metabolic syndrome has increased, non-alcoholic fatty liver disease (NAFLD) in patients with CHB has become more common. Both diseases can lead to liver fibrosis and even hepatocellular carcinoma, but the risk of dual etiology, outcome, and CHB combined with NAFLD is not fully elucidated. In this review, we assess the overlapping prevalence of NAFLD and CHB, summarize recent studies of clinical and basic research related to potential interactions, and evaluate the progressive changes of treatments for CHB patients with NAFLD. This review increases the understanding of the relationship and mechanisms of interaction between steatosis and hepatitis B virus infection, and it provides new strategies for the future clinical management and treatment of CHB combined with NAFLD.

Keywords: Chronic hepatitis B; Non-alcoholic fatty liver disease; Steatosis; Mechanism

Introduction

Chronic hepatitis B (CHB) virus infection is a well-known threat to global health despite the availability of vaccines and antiviral treatments, and nearly 292 million people (3.9% of the world's population) are living with hepatitis B virus (HBV) infection (defined as being hepatitis B surface antigen [HBsAg]-positive).^[1,2] HBV infection results in >820,000 deaths worldwide each year from cirrhosis and hepatocellular carcinoma (HCC).^[3]

Non-alcoholic fatty liver disease (NAFLD) has emerged as another common chronic liver disease that affects a quarter of the global population.^[4] NAFLD is related to obesity and it includes a series of pathologies such as steatosis, non-alcoholic steatohepatitis (NASH), fibrosis, and cirrhosis. NAFLD also increases the risk of HCC.^[5] Global prevalence of NAFLD is 29.8% (95% confidence interval [CI] 28.6–31.1) with the highest prevalence in South America and the lowest in Africa.^[6] Our previous meta-analysis included 237 studies (13,044,518 participants) in Asia, and it showed that the overall prevalence of NAFLD was 29.62% (95% CI 28.13–31.15), which

was similar to the prevalence trends reported in the western world. The data above indicated that NAFLD is a global disease, regardless of the diagnostic method used.^[7] In addition, NAFLD is strongly associated with obesity, insulin resistance, metabolic syndrome, and cardiovascular disease.^[8]

As the prevalence of NAFLD increases, the coexistence of NAFLD and HBV infection is also increasing. The overall prevalence of NAFLD in CHB patients is approximately 14% to 70%.^[9,10] The existing global epidemiological data on CHB infection combined with NAFLD are mostly from single-center, retrospective cohort studies. The precise prevalence of NAFLD in CHB is widely variable due to different research cohorts that have included different populations and different diagnostic methods for NAFLD, such as ultrasound, transient elastography, or liver biopsy. A recent meta-analysis showed that the pooled prevalence of hepatic steatosis (HS) in CHB was 32.8% (95% CI 28.9–37.0), and it was especially higher in men and obese patients.^[11] Because both HBV infection and NAFLD can cause chronic liver injury, exacerbate liver damage, increase the risk of cirrhosis and HCC, and

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seriously endanger liver health, the relationship between these two diseases has attracted increasing attention.^[12,13] However, some studies have shown that the incidence of NAFLD is positively associated with a reduction in HBV serum markers in patients with CHB. We systematically reviewed the research progress related to HBV and NAFLD, trying to unravel the potential relationship between the two diseases, then help to explain the paradox above.

Interaction of CHB and NAFLD

How does NAFLD affect CHB disease progression?

The disappearance of HBsAg and undetectable HBV-DNA in the serum of patients with CHB are important hallmarks of a functional cure.^[14] HBsAg seroclearance rarely occurs in the natural history of CHB infection, and prior cohort studies have shown that the annual HBsAg seroclearance rate can range from 0.12% to 2.38%.^[15] In addition, even with interferons and antiviral therapy, the incidence of HBsAg serological clearance is still very low. Most of the recent studies have concluded that HS is positively correlated with HBsAg serum clearance and HBV-DNA suppression.

A study by Chu *et al*^[16] in 54 HBsAg carriers with HBsAg seroclearance found that compared with 108 carriers with HBsAg persistence, moderate-to-severe HS may contribute to HBsAg seroclearance in HBsAg carriers. In addition, their further research of 155 patients indicated that HBsAg carriers with a fatty liver on ultrasound and HS could accelerate HBsAg seroclearance by approximately 5 years.^[17] Similarly, our retrospective cohort study of 6786 adult patients with CHB found that compared to patients with non-fatty liver CHB, patients with fatty liver CHB had a higher HBsAg seroclearance rate.^[18] We previously found that CHB patients with NAFLD were significantly more likely to be hepatitis B e antigen (HBeAg) negative (74.3% *vs.* 62.8%) compared with CHB patients without NAFLD.^[19] Tai *et al*^[20] demonstrated a significantly higher incidence of HBsAg clearance in HBeAg-seronegative CHB patients with HS than in those without. They employed Cox regression analysis to further identify HS as an independent predictor of spontaneous HBsAg seroclearance in CHB patients, including HBeAg-seropositive and -seronegative (hazard ratio [HR]=1.222).^[21] Extreme obesity and central obesity were associated with a low prevalence of high HBV viral load in HBeAg-seropositive patients, especially in men. However, hypertriglyceridemia was associated with a low prevalence of high viral load in HBeAg-negative patients, both women and men.^[22] Increased HS was independently associated with decreased serum HBV DNA levels, suggesting a potential negative impact on viral replication.^[23] Moreover, Wang *et al*^[24] performed a study on 3212 patients with HBV infection and showed that the severity of HS in patients was negatively correlated with the expression of intrahepatic HBsAg and hepatitis B core antigen (HBcAg). It is important to note that the positive correlation between HS and HBeAg/HBsAg serologic clearance or HBV-DNA suppression should be viewed dialectically, and we cannot assume that

NAFLD is beneficial in CHB patients. Despite HBV seromarkers being reduced in patients with CHB and NAFLD, the clinical outcomes of these patients may be worse.

While there exists growing evidence that NAFLD may provide some benefit in reducing disease progression in CHB patients, there are some studies that have suggested otherwise. Jin *et al*^[25] retrospectively analyzed 720 CHB patients in 3 years and found that HS was not significantly associated with HBsAg seroclearance, fibrosis progression, or HCC development. However, their study may be influenced by its retrospective nature with a relatively short follow-up duration.

There have also been some studies that verified the negative association between NAFLD and HBV in cellular and animal models. Zhang *et al*^[26] established mice with chronic HBV genotype B infection using a microinjection of oocytes, and HS was induced by feeding them high-fat diets. The research found that the model mice suffering from both CHB and NAFLD had reduced HBV viral replication.^[26] Similar results were obtained by Hu *et al*^[27] in an HBV-immunocompetent mouse model where NASH inhibited HBV-DNA replication and reduced the expression levels of HBeAg and HBsAg. Another study verified this result from both cell and mouse models, and it proposed that saturated fatty acids (SFAs) can inhibit HBV replication in CHB combined with NAFLD by activating the toll-like receptor 4 (TLR4) signaling pathway.^[28]

The possible mechanistic role of NAFLD in HBV infection remains unclear, and there are several speculations. As mentioned above, TLR4 is a cell surface receptor that is crucial for the activation of innate immune responses. HBV infection restricts interferon production and interferon-stimulated gene activation by the inhibition of the myeloid differentiation primary response 88 (MyD88) signaling pathway, leading to innate immunodeficiency and chronic HBV infection.^[29,30] Our previous study indicated that TLR4 induced MyD88/nuclear factor-kappa B downstream activation in B cells triggered by HBV.^[31] Zhang *et al*^[28] found that SFAs served as a potential ligand for TLR4 and activated the TLR4 signaling pathway, and this might be involved in accelerating the mechanism of inhibiting HBV replication in CHB with NAFLD. Some research has shown that metabolic factors in NAFLD may influence HBV replication. For example, Shlomain *et al*^[32] found that fasting stimulated HBV DNA replication *in vivo*, and this was primarily due to an increase in the peroxisome proliferator-activated receptor- γ coactivator (PGC)-1 α activity. PGC-1 α is a transcription factor that plays an important role in gluconeogenesis,^[32] and simultaneously, PGC-1 α is lowly expressed in NAFLD.^[33] Therefore, the metabolic alterations induced by NAFLD may inhibit HBV replication by suppressing the expression of PGC-1 α . However, apoptosis in HBV-infected cells has been reported to inhibit viral replication in these cells.^[34] In addition, increased Fas-mediated apoptosis has been observed in liver samples from patients with NAFLD.^[35,36] It is hypothesized that NAFLD may accelerate HBsAg and

HBV-DNA clearance by increasing apoptosis in HBV-infected cells. We summarize the possible pathways and mechanisms as Figure 1.

Do CHB patients have a decreased risk of NAFLD onset?

It is now well established that hepatitis C virus (HCV) infection affects changes in insulin resistance and lipid metabolism, leading to HS and more severe inflammation

in patients with chronic hepatitis C (CHC). HS is also a common histological feature of CHC. Unlike HCV, studies of HS and CHB are limited. There is currently no direct evidence that HBV increases the risk of steatosis. A meta-analysis revealed HS in HBV seems to be as frequent as in the general population, and lower than in HCV-infected patients. In addition, there was a strong negative correlation between viral load and HS, which may indicate a protective effect of the virus on HS.^[10]

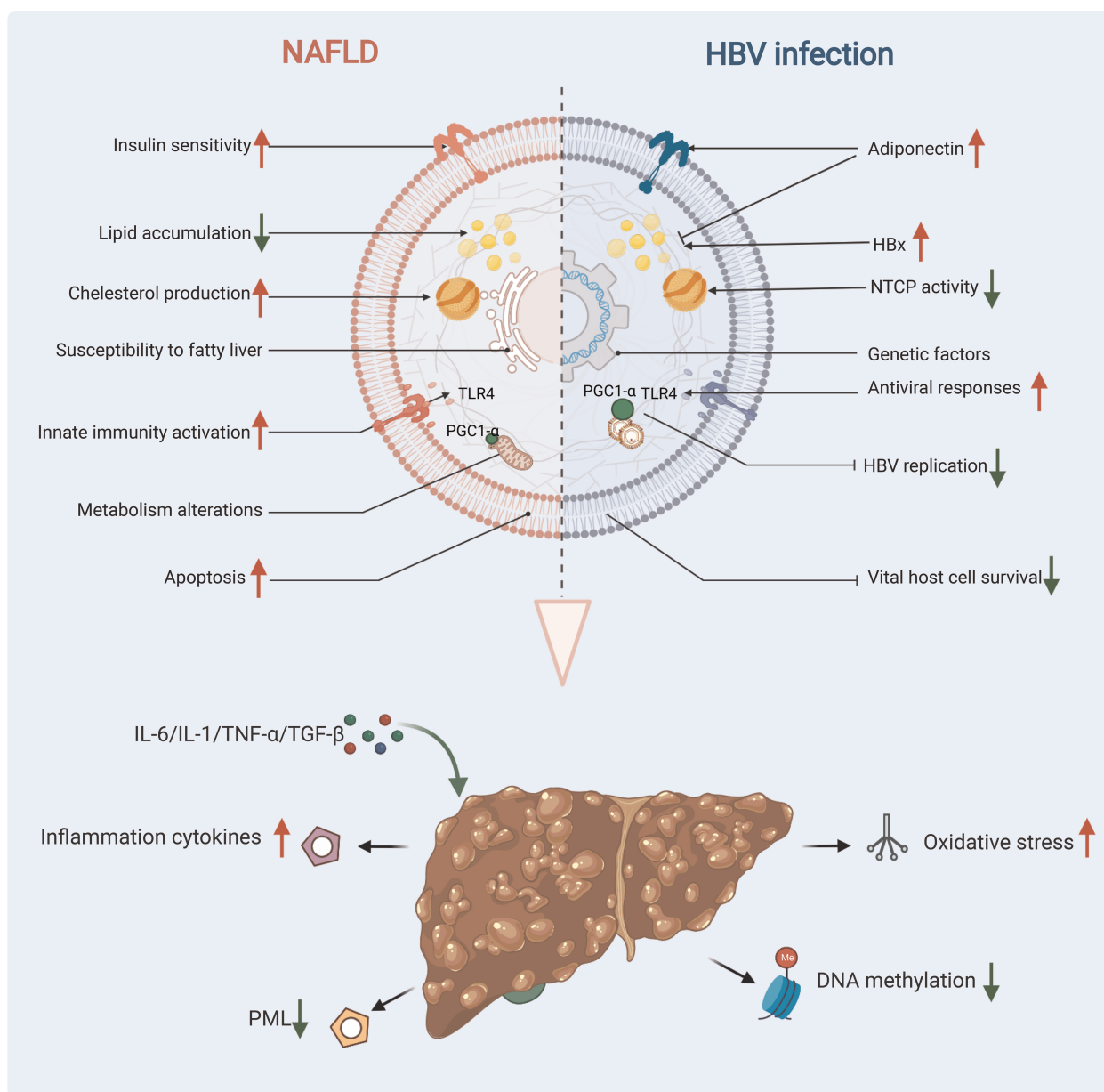


Figure 1: Potential mechanisms of interaction between NAFLD and CHB and clinical outcomes. Metabolic alterations in NAFLD patients may enhance antiviral responses through activation of innate immunity, such as by activating the TLR4 signaling pathway or suppressing the expression of PGC-1 α . NAFLD could also accelerate HBsAg and HBV-DNA clearance by increasing apoptosis in HBV-infected cells. Hepatic steatosis in CHB patients is associated with host metabolic factors, a reduced risk of hyperlipidemia has been observed in patients with CHB. In addition, the higher serum adiponectin level could account for the lower prevalence of steatosis in HBV-infected subjects. Conversely, the overexpression of HBx and the genetic susceptibility to fatty liver in CHB patients induce hepatic lipid accumulation. There are many potential mechanisms for clinical outcomes in NAFLD combined with CHB. Inflammatory cytokines (eg, IL-1, IL-6, TNF- α , TGF β) released from damaged hepatocytes in NASH can lead to activation and proliferation of hepatic stellate cells. NAFLD-related fat deposition and oxidative stress may create a pro-fibrotic and pro-carcinogenic milieu within the liver. In addition, the PML deficiency-mediated abnormal lipid metabolism induced by HBsAg and the lower levels of global DNA methylation in patients with concurrent NAFLD and CHB could both accelerate the development of cirrhosis and HCC. CHB: Chronic hepatitis B; HBsAg: Hepatitis B surface antigen; HBV: hepatitis B virus; HBx: Hepatitis B protein X; NAFLD: Non-alcoholic fatty liver disease; NASH: Non-alcoholic steatohepatitis; PGC-1 α : Peroxisome proliferator-activated receptor- γ coactivator-1 α ; PML: Promyelocyticleukemia protein; TLR4: Toll-like receptor 4; NTCP: Sodium taurocholate cotransporting polypeptide; IL-1: Interleukin-1; IL-6: Interleukin-6; TNF- α : Tumor necrosis factor- α ; TGF β : Transforming growth factor β .

Table 1: Publications on the potential role of CHB in NAFLD onset.

References	Year of publication	Number of patients	Explored association between CHB and NAFLD onset	OR, RR, or HR
Wong <i>et al</i> ^[38]	2012	1030	HBsAg seropositivity and fatty liver	0.42 (0.20–0.88)
Cheng <i>et al</i> ^[44]	2013	33,439	HBsAg seropositivity and fatty liver	0.70 (0.64–0.76)
Huang <i>et al</i> ^[40]	2020	14,452	HBsAg seropositivity and NAFLD	0.72 (0.61–0.85)
Wang <i>et al</i> ^[41]	2018	1882	HBsAg seropositivity and NAFLD	0.57 (0.34–0.98)
Lin <i>et al</i> ^[42]	2021	4734	HBsAg seropositivity and NAFLD	0.83 (0.78–0.89)
Lv <i>et al</i> ^[43]	2021	16,451	HBsAg seropositivity and hypercholesterolemia	0.62 (0.58–0.66)
Zhong <i>et al</i> ^[39]	2018	2988	HBsAg seropositivity and NAFLD	0.64 (0.42–0.95)
Yun <i>et al</i> ^[47]	2009	86	HBsAg seropositivity and hypercholesterolemia	1.19 (1.17–2.83)
Joo <i>et al</i> ^[37]	2017	83,339	HBsAg seropositivity and NAFLD	0.83 (0.73–0.94)
Zhu <i>et al</i> ^[45]	2019	4429	HBV DNA levels and NAFLD	0.37 (0.14–0.98)

CHB: Chronic hepatitis B; HBsAg: Hepatitis B surface antigen; HBV: Hepatitis B virus; HR: Hazard ratio; NAFLD: Non-alcoholic fatty liver disease; RR: Relative risk; OR: Odds ratio.

The results of recent studies have indicated a negative association with a possible risk for steatosis in CHB [Table 1]. In a large-scale cohort study of 83,339 participants, the incidence of steatosis was significantly lower in CHB patients than in the controls. Even after adjusting for possible confounders and metabolic factors, there was still a significant negative correlation between HBV infection and NAFLD.^[37] A cross-sectional study using proton-magnetic resonance spectroscopy, a highly reliable steatosis assay, to determine fatty liver status found that chronic HBV infection was significantly associated with a lower risk of fatty liver.^[38] Another large case-control study in China confirmed this conclusion.^[39] The results of four additional studies indicated a negative correlation between steatosis and CHB.^[40–43]

To date, various studies seem to indicate that HBV may have a protective effect on the development of the fatty liver. In a large-scale study in Taiwan (China) by Cheng *et al*^[44] that recruited a total of 33,439 subjects, they found an inverse relationship between HBV infection and NAFLD in subjects older than 50 years and no significant association between HBV infection and fatty liver in subjects younger than 50 years. Zhu *et al*^[45] found no association between viral factors and the NAFLD incidence rate in 2393 CHB patients who had no history of alcohol intake. Moreover, steatosis was observed in 62% of patients according to the research of Viganò *et al*^[46], and it was independently associated with body mass index (BMI) and hyperglycemia, but not with virologic factors such as HBeAg status or HBV-DNA load. The authors speculated that metabolic alterations are the leading cause of steatosis in CHB.^[46] In addition, three case-control studies have shown that HBeAg status and HBV DNA are not related to liver steatosis in patients with CHB.^[47–49] Enomoto *et al*^[50] obtained a similar result in the histology of patients with high HBV-DNA levels. Wang *et al*^[41] found that HBV infection is not associated with fatty liver disease occurrence, and they verified the results *in vitro*. Conversely, it was revealed that HBV may play a role in the development of NAFLD by increasing the production of mitochondrial reactive oxygen species *in vitro*.^[41]

Hu *et al*^[27] found that in an HBV-immunocompetent mouse model, NASH inhibited HBV replication, while

virus replication did not alter lipid metabolism. Similarly, Zhang *et al*^[26] found that in HBV genotype B transgenic mice with combined NAFLD, HBV viral factors were reduced compared to the controls, without reducing the metabolic and histologic features. Both mouse models had their shortcomings. HBV transgenic mice are immune-tolerant and cannot be spontaneously cleared as their genomes are integrated into the mouse genome. HBV-immunocompetent mice with a hydrodynamic injection can effectively transfer exogenous genes into liver cells; however, viremia peaks after 6 days and rapidly declines thereafter. Although these animal models differ from HBV infection in humans, the studies still play important roles in the study of HBV combined with NAFLD.

Some studies have pointed out that HS in CHB patients is associated with host metabolic factors. Hyperlipidemia is an important risk factor for NAFLD; however, a reduced risk of hyperlipidemia has been observed in patients with CHB. A cross-sectional study of 7695 Taiwanese adults demonstrated a lower risk of hypercholesterolemia, hypertriglyceridemia, and high low-density lipoprotein cholesterol levels in HBV-infected individuals.^[51] In addition, two large community-based cohort studies found a negative association between chronic HBV infection and serum lipid levels, such as triglycerides (TG) and cholesterol.^[52,53] In another retrospective cohort of 122 patients with CHB, a negative association between serum HBV DNA levels and TG levels was also found.^[54]

Furthermore, HBV infection may alter hepatic cholesterol metabolism. Some studies have shown that the pre-S1 domain of the HBV envelope can hinder sodium taurocholate cotransporting polypeptide (NTCP) mediated bile acid uptake and lead to compensatory cholesterol production and uptake. In addition, adiponectin improves hepatic insulin sensitivity and decreases lipid accumulation in macrophages.^[55,56] CHB patients have been reported to have higher serum adiponectin levels, and this could account for the low prevalence of steatosis in HBV-infected subjects.^[57] In addition, Hajjou *et al*^[58] identified a significant upregulation of lipid biosynthesis gene expression in the liver of HBV transgenic mice,

including adenosine 5'-triphosphate citrate lyase, fatty acid synthase, sterol regulatory element-binding factor 2, and retinol-binding protein 1. Yilmaz *et al*^[59] found that in patients infected with HBV, HS was primarily associated with metabolic factors, such as obesity, insulin resistance, and dyslipidemia, rather than viral factors.

Hepatitis B protein X (HBx), one of the four HBV proteins, has an important role in HBV infection. Previous studies have confirmed that the overexpression of HBx induces hepatic lipid accumulation in HepG2-HBx stable cells and in HBx-transgenic mice. Therefore, HBx is a risk factor for steatosis.^[41] Kang *et al*^[60] showed that hepatic lipid accumulation may be caused by the inhibition of apolipoprotein B secretion by HBx protein, while Kim *et al*^[61] suggested that increased HBx expression mediated by sterol regulatory element-binding protein 1 and peroxisome proliferator-activated receptor γ (PPAR- γ) also increases hepatic lipid accumulation. Subsequently, HBx has been reported to upregulate fatty acid-binding protein 1 to promote hepatic lipid accumulation in the development of steatosis in HBV-induced cells.^[62] In addition, in HBx-transfected hepatocytes, HBx stimulated the expression and transcriptional activation of CCAAT/enhancer-binding protein α (C/EBP α) and PPAR- γ , both regulators of C/EBP α adipocyte differentiation.^[61]

The evidence suggested the possibility of genetic susceptibility to fatty liver in CHB. Single-nucleotide polymorphisms (SNPs) of patatin-like phospholipase domain-containing protein 3 (PNPLA3), and transmembrane 6 superfamily member 2 (TM6SF2) are two important genetic determinants of NAFLD.^[63,64] In cohort studies of biopsy-proven CHB patients with or without HS, several SNPs of PNPLA3 were independently associated with steatosis and inflammation. In addition, when compared to the TT genotype, the C allele at PNPLA3 rs1010023 conferred a

higher risk of HS in CHB patients (OR = 1.768, 95% CI: 1.027–3.105; $P = 0.045$).^[65] The T allele of rs58542926 in TM6SF2 has been associated with altered lipids and HS in CHB patients.^[66] The genetic and metabolic changes related to HBV infection at the cellular level could help explain the clinical outcomes, but phenotypic differences related to NAFLD at the individual level require further study. We summarize the possible pathways and mechanisms as Figure 1.

Progression and Outcomes of CHB with NAFLD

In NAFLD patients, fibrosis is the feature most closely associated with long-term adverse events compared to other histological features. NASH, a severe form of NAFLD, has a rapid progression in fibrosis, and it is the major cause of liver fibrosis, cirrhosis, and HCC in advanced NAFLD.^[67] In developed countries, NAFLD could account for >30% of HCC cases.^[5] As we know, CHB infection is an important cause of HCC formation, and previously, >70% of the HCC incidence was attributed to chronic viral hepatitis.^[68] Both diseases are capable of independently increasing the risk of HCC development. Today, patients with CHB have an increased incidence of combined HS, but there is no conclusive evidence linking HS and liver fibrosis, cirrhosis, and HCC in patients with CHB infection. Table 2 is a summary of the recent studies.

In Hong Kong (China), Cheng *et al*^[69] prospectively recruited 1466 patients with CHB from 2006 to 2008. They found that metabolic syndrome increased the risk of cardiovascular events, such as acute coronary syndrome, and cerebral vascular events, but not cirrhotic complications, HCC, and/or liver-related mortality.^[69] Similarly, our previous retrospective cohort study reported 6786 adult Asian participants with CHB from 1991 to 2016.

Table 2: Publications on the progression and outcomes of CHB with NAFLD.

References	Year of publication	Number of patients	HS and fibrosis (surrogate/method)	HBsAg seroclearance	Fibrosis progression	Cirrhosis progression	HCC development
Li <i>et al</i> ^[18]	2021	6786	Ultrasonography or CT	More in FL	Not reported	Less in FL	Less in FL
Chang <i>et al</i> ^[25]	2021	720	Transient elastography	No difference	No difference	Not reported	No difference
Cheng <i>et al</i> ^[69]	2016	1466	Transient elastography	Not reported	No difference	No difference	No difference
Lee <i>et al</i> ^[13]	2019	321	Liver biopsy	Not reported	Not reported	Not reported	No difference
Lim <i>et al</i> ^[73]	2020	289	Liver biopsy	Not reported	Not reported	Not reported	No difference
Chen <i>et al</i> ^[70]	2017	162	Liver biopsy	Not reported	More in HS	No difference	Not reported
Mandana <i>et al</i> ^[74]	2021	420	Liver biopsy	Not reported	More in steatohepatitis	No difference	Not reported
Wong <i>et al</i> ^[78]	2020	614	Transient elastography	Not reported	More in HS	Not reported	No difference
Chu <i>et al</i> ^[17]	2013	155	Ultrasonography	More in HS	Not reported	More in HS	Not reported
Mak <i>et al</i> ^[74]	2020	330	Transient elastography	More in HS	More in HS	Not reported	Not reported
Charatcharoenwithaya <i>et al</i> ^[76]	2017	256	Liver biopsy	Not reported	More in steatohepatitis	Not reported	Not reported
Choi <i>et al</i> ^[9]	2020	1089	Liver biopsy	Not reported	More in NASH	Not reported	More in NASH
van Kleef <i>et al</i> ^[79]	2021	1076	Liver biopsy	Not reported	Not reported	Not reported	More in MAFLD
Peleg <i>et al</i> ^[81]	2019	524	Liver biopsy and ultrasonographic	Not reported	More in liver steatosis	Not reported	More in liver steatosis
Kim <i>et al</i> ^[80]	2021	48,335	FLI and HSI	Not reported	Not reported	Not reported	More in HS
Chan <i>et al</i> ^[12]	2017	270	Liver biopsy	Not reported	Not reported	More in FL	More in FL
Karacaer <i>et al</i> ^[72]	2016	254	Ultrasonography	Not reported	More in HS	Not reported	Not reported

CHB: Chronic hepatitis B; CT: Computed tomography; FL: Fatty liver; FLI: Fatty liver index; HBsAg: Hepatitis B surface antigen; HS: Hepatic steatosis; HSI: Hepatic steatosis index; MAFLD: metabolic associated fatty liver disease; NAFLD: Non-alcoholic fatty liver disease; NASH: Non-alcoholic steatohepatitis.

During the mean follow-up of 131.88 ± 77.28 months, 564 patients developed cirrhosis, 281 patients developed HCC, and 478 patients had HBsAg seroclearance. Importantly, we found that a combined fatty liver did not increase the occurrence of liver cirrhosis and HCC in the CHB patients.^[19] It has also been shown that significant or advanced liver fibrosis is associated with the grade of necroinflammation and host factors but not with HS in CHB patients.^[70,71] In addition, several other studies have suggested that HS is not a risk factor for the probability of fibrosis progression or HCC development in CHB patients.^[13,25,71-73]

However, most studies have confirmed that obesity, diabetes, and MetS are independent risk factors for fibrosis, cirrhosis, and HCC in CHB patients, suggesting a synergistic effect of metabolic factors and CHB on HCC pathogenesis. A recent cohort study from North America included 420 CHB cases who underwent liver biopsies. The study found that the combination was associated with advanced fibrosis and higher biochemical measures of hepatic inflammation over time.^[74] Similar results were obtained in two Hong Kong studies.^[12,75] In addition, Charatcharoenwitthaya *et al*^[76] demonstrated that steatohepatitis was related to the severity of liver fibrosis, and Mak *et al*^[77] showed that the rate of liver fibrosis progression in patients with persistent severe steatosis was higher than in those without steatosis in CHB. Moreover, Chen *et al*^[70] found that the 5-year cumulative incidence of cirrhosis was 4.17% in patients without HS and 5.19% in patients with HS, and advanced fibrosis may be a risk factor for progression to cirrhosis in NAFLD. Furthermore, HS may promote the progression of HCC.^[70] In particular, Chan *et al*^[12] found that the incidence of HCC was significantly higher in patients with steatosis >5% than in those with steatosis <5%, and that combined fatty liver was an independent risk factor for the development of HBV-related HCC, with an associated 7.3-fold increased risk.

A longitudinal cohort study from Wong *et al*^[78] found that the presence of HS was independently associated with advanced fibrosis. However, HS was not associated with any adverse outcomes in CHB patients. These researchers indicated that HS does not predict adverse outcomes in CHB patients, unlike advanced fibrosis.^[78] Conversely, a large combined tertiary center cohort that included 1089 patients who underwent liver biopsies between 1985 and 2016 at one of the two tertiary centers showed that combined severe HS was closely associated with the development of liver fibrosis in CHB patients. The combination of NASH also increases the risk of liver-related events, including decompensated cirrhosis, HCC, and even death, in CHB patients.^[9] Similar results were obtained in a recent study by van Kleef *et al*^[79]. Additionally, a study by Kim *et al*^[80] reported that CHB patients with liver steatosis had a higher risk of all-cause mortality and cancer development compared to patients without liver steatosis. Another retrospective study included 524 treatment-naïve patients with CHB with a mean follow-up of 6 years. It found that CHB patients with liver steatosis had an increased risk of all-cause mortality and cancer development compared to

CHB patients without liver steatosis, regardless of their baseline HBV viral load.^[81] All of these results indicated that in addition to viral suppression, screening for and managing metabolic abnormalities is important to prevent disease progression in HBV.

The mechanism of NAFLD in CHB-related fibrosis, cirrhosis, and HCC remains unclear, possible pathways and mechanisms are shown in Figure 1. It is speculated that NAFLD-mediated inflammatory injury and fibrogenesis in HBV infection are responsible.^[74] It is well known that danger-associated molecular patterns released from damaged hepatocytes in NASH as well as inflammatory cytokines can lead to activation and proliferation of hepatic stellate cells, which can exacerbate fibrosis or eventually cirrhosis.^[82,83] NAFLD-related accumulated fat deposition, oxidative stress, and a low-grade inflammatory response may create a pro-fibrotic and pro-carcinogenic milieu within the liver and further accelerate the development of CHB-related cirrhosis and HCC. In addition, a recent study found that the knockdown of the promyelocytic leukemia protein (PML; a tumor suppressor involved in genome maintenance and fatty acid oxidation) in HBsAg-transgenic mice predisposed them to obesity and drove early-steatosis-specific liver tumorigenesis. This demonstrated that PML deficiency-mediated abnormal lipid metabolism induced by HBsAg appears to be a hallmark of chronic HBV-associated pathogenesis leading to steatosis-associated hepatocarcinogenesis.^[84]

DNA methylation is one of the most common epigenetic modifications, and a loss of global DNA methylation in sequences that are normally methylated can lead to chromosomal abnormalities and ultimately disease development.^[85,86] A recent study found that patients with concurrent NAFLD and CHB exhibited lower levels of global DNA methylation than patients with NAFLD alone. The concurrence of CHB and liver fibrosis in NAFLD patients was associated with a decrease in the global DNA methylation levels.^[87] NAFLD-related cumulative fat deposition, oxidative stress, and inflammatory responses may produce a pro-fibrotic and pro-cancer environment in the liver and further accelerate the development of cirrhosis and HCC. However, the disease progression is influenced by various confounding factors, and the interaction between HBV and metabolic homeostasis in the progression of liver disease requires further study.

Effect of Antiviral Treatment on NAFLD

Antiviral therapy is commonly used in preventing disease progression and the incidence of HCC in CHB patients. However, the data regarding the implication of HS on antiviral therapy for CHB have been conflicting. A retrospective single-centered cohort study in 2020 found that the presence of NAFLD and metabolic features associated with NAFLD did not negatively affect the complete virologic suppression or biochemical response to CHB treatment outcomes during a follow-up of up to 60 months. In addition, they recommended that CHB patients with NAFLD should not require any special considerations while choosing CHB treatment.^[19] Similar

results were observed in three other studies in CHB patients treated with entecavir (ETV) or tenofovir disoproxil fumarate (TDF).^[88-91] In addition, Chen *et al*^[92] found that HS had no significant impact on the HBeAg seroclearance in HBeAg-positive CHB patients with nucleos(t)ide analog (NA) treatment. Moreover, a retrospective multicenter cohort study that included a total of 145 cases with chronic HBV infection from three different medical centers indicated that NAFLD did not affect the virological response at 6 and 12 weeks of ETV or TDF treatment. In addition, the presence of steatohepatitis was found to predict a more favorable response to treatment at 6 months in the TDF arm than in patients without.^[93]

However, it has also been demonstrated that the efficacy of antiviral therapy is worse in CHB patients with combined HS. Liang *et al*^[94] found that HS can reduce the efficacy of PEG-IFN α -2a in the treatment of CHB patients, and its mechanism may be related to different HBeAg expression patterns in liver tissue. Jin *et al*^[95] found that HS was a significant independent factor associated with ETV treatment failure in a prospective study of 267 Chinese patients with CHB infection. Concurrently, they indicated this was due to lipid accumulation and the reduced activity of cytochromes in HS on drug metabolism.^[95] In addition, a meta-analysis that included eight prospective cohort studies showed that the biochemical and virological responses at both 48 and 96 weeks were significantly lower in patients with CHB with HS as compared to that in patients with only CHB. HS lowers the efficacy of antiviral treatment in patients with CHB, especially when HS is diagnosed on ultrasound findings and treated with NAs.^[96] Recently, a study reported that the NA-treated group increased body fat mass and visceral fat area in CHB patients.^[97]

Some studies have indicated that NA therapy might influence HS, and NA therapy may contribute to body fat accumulation in CHB patients.^[98] An *in vitro* study reported that TDF modulated lipid metabolism by upregulating hepatic CD36 via activating PPAR- α .^[99] The high expression of hepatic CD36 improved HS and insulin resistance by reducing hepatic lipids, and this may explain the findings. In a study of patients with CHB, switching to tenofovir alafenamide improved metabolic dysfunction, reduced the serum alanine aminotransferase (ALT) levels, and promoted ALT normalization in patients, despite significant increases in body weight and BMI.^[100] Although the impact of antiviral therapy on HS in these patients remains debatable, the onset or progression of NAFLD during antiviral therapy should still be monitored in case of underlying negative impacts.

In contrast, there are fewer studies that have examined the effect of fatty liver treatment on CHB progression. A retrospective cohort study in Korea showed that of 13,063 patients with CHB, 980 (7.5%) were statin users and 12,083 (92.5%) were statin-naïve. The annual liver cancer mortality was significantly lower among statin users than non-users. A multivariable analysis after adjusting for demographic and metabolic factors showed consistent results.^[101] This study had some limitations. It did not

include detailed data on the serum HBV DNA levels or cirrhosis status. In addition, this study did not consider the effect of antiviral therapy on liver cancer mortality. In addition, the study population consisted exclusively of Korean populations, which limits the generalizability of the findings. We also require more studies regarding the effects of NAFLD treatments, such as medications and bariatric surgery, on the progression of HBV infection and antiviral efficacy.

Potential Cell and Animal Models for Researching CHB and NAFLD

In vitro cell models supporting HBV replication and infection are important tools for basic studies of the HBV life cycle, and these models play essential roles in the identification of novel anti-HBV targets and efficacy evaluations of drug candidates while helping to better investigate the mechanisms of HBV combined with NAFLD.

Human hepatocytes are specific hosts for HBV. For quite some time, primary human hepatocytes (PHH) were the only research model supporting HBV infection *in vitro*.^[102] However, due to the limited source of human primary hepatocytes, the demanding *in vitro* culture techniques, and the inability to continuously expand have limited their application in HBV research.^[103] Due to the availability of PHH and donor problems, HepaRG cells have been investigated as an alternative source. The bipotential progenitor-like HepaRG cell line was isolated from an HCV-induced hepatocarcinoma and can be induced to differentiate as hepatocytes or cholangiocytes *in vitro*.^[104] Human hepatoma cell lines that replicate HBV after transfection or the stable integration of HBV genomes have been widely used for the analysis of HBV replication and viral-host interactions, and HepG2.2.15 is the most commonly used of these.^[105,106] NTCP is a multiple transmembrane bile acid transporter protein specifically expressed on the surface of hepatocyte membranes that binds specifically to the pre-S1 region of the HBV viral outer membrane protein and is a functional receptor for HBV that is absent from expression in HCC cell lines. Therefore, the commonly used HCC cell lines, such as HepG2 and Huh7 are resistant to HBV infection, but exogenous expression of NTCP in them can restore their HBV susceptibility.^[107] It has been widely used for the study of basic biological problems of HBV and also has provided tools for the development of early antiviral drugs. To investigate the mutual effects and mechanisms of CHB patients with combined fatty liver *in vitro*, palmitic acid and oleic acid are the most common free fatty acids that can induce steatosis in HBV cell models.^[28]

In addition to cellular models, animal models are crucial to imitate NAFLD and HBV infection simultaneously. HBV can only infect humans and chimpanzees, but the use of chimpanzees in HBV research is strongly restricted.^[108] To date, the tree shrew (*Tupaia belangeri*) is the only non-primate animal found to be susceptible to HBV infection. Primary *Tupaia* hepatocytes are readily available compared with PHH, and they have been used in HBV research for many years.^[109]

Humanized chimeric mouse

The first human liver chimeric mouse model was developed in immunodeficient (Rag2^{-/-}, severe combined immunodeficiency [SCID], SCID/beige) mice using the urokinase-type plasminogen activator (uPA) transgene. The transplantation of human hepatocytes into uPA-SCID mice results in a liver-humanized model with a high human hepatocyte reconstitution rate that is supportive of HBV and HCV infection.^[110,111] These models can simulate the natural history of HBV infection *in vivo* as well as the innate and adaptive immune responses against HBV.

HBV transgenic mouse model

It has long been reported in the literature that HBV transgenic mice that selectively express HBV proteins have been used to study topics related to HBV infection. In 1995, a full HBV genome transgenic mouse model was established, producing infectious HBV virions morphologically identical to human-derived virions.^[112] Although HBV is immunotolerant and cannot be spontaneously cleared due to its genomic integration into the mouse genome, this model has made an important contribution to elucidating the pathogenesis of HBV infection.

HBV transfected mouse model

Adeno-associated viral vectors containing the HBV genome efficiently transduce hepatocytes in immunocompetent mice. HBV replication can last for 3 months after intravenous injection of adenoviral vectors containing HBV.^[113] Rapid injection of large amounts of liquid containing naked DNA into mice by the tail vein, called hydrodynamic injection, which can effectively transfer exogenous genes into hepatocytes. Hydrodynamic injection was found to successfully deliver replication-competent HBV genome into the liver of immunocompetent mice, with viremia peaking after 6 days and declining rapidly thereafter.^[114]

Mouse models of NAFLD

Animal models of NAFLD are primarily based on various types of diet, such as a HFD, a high glucose, sucrose, fructose, methionine, and choline-deficient (MCD) diet, a choline-deficient L-amino acid-defined diet, a high-cholesterol diet, and a cholesterol and cholate diet. Even in the genetic animal models of NASH, diet is used as a means of a secondary trigger for disease progression.^[115] A HFD composed of 71% fat, 11% carbohydrates, and 18% proteins fed to mice for 16 weeks is known to result in the development of insulin resistance with marked panlobular steatosis, inflammation, and induce fibrosis.^[116] The model displays NAFLD features similar to human NAFLD, but the pathological outcome is not as severe.^[115] MCD is a commonly used diet that produces the most severe phenotype of NASH in the shortest amount of time. This diet has high sucrose (40%) and 10% fat, but it is deficient in methionine and choline, and it is known to quickly induce measurable hallmarks of NAFLD, such as HS in mice and rats in 2 to 4 weeks, and this progresses to inflammation and fibrosis shortly thereafter.^[117,118]

These HBV mouse models combined with well-established NAFLD mice models, such as HFD or MCD, promise to serve as applicable alternative models for exploring the interaction of HBV infection and NAFLD *in vivo*.

Conclusions

Both NAFLD and CHB are chronic liver diseases that inevitably threaten the health of patients by disrupting liver function and exacerbating end-stage cirrhosis and HCC. Although there are abundant results that have demonstrated serum clearance of hepatitis B in patients with HS, the prevalence of NAFLD may still be lower in CHB patients. There remains a knowledge gap on how this happens. The outcome of treatment with oral antivirals or interferons appears to be unaffected by HS. However, viral factors appear to influence necroinflammation and fibrosis in the liver rather than HS. As the mechanisms of interactions between steatosis and HBV infection become clearer, future studies will provide new strategies for the clinical management and treatment of CHB combined with NAFLD.

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Conflicts of interest

None.

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