

RESEARCH ARTICLE

Polyethylenimine-grafted mesoporous silica nanocarriers markedly enhance the bactericidal effect of curcumin against *Staphylococcus aureus* biofilm

Ayşenur Pamukçu¹  | Nursu Erdoğan¹ | Didem Şen Karaman^{2,3}

¹Department of Biomedical Technologies, Graduate School of Natural and Applied Sciences, Izmir Katip Çelebi University, Izmir, Turkey

²Department of Biomedical Engineering, Faculty of Engineering and Architecture, Izmir Katip Çelebi University, Izmir, Turkey

³Pharmaceutical Sciences Laboratory, Faculty of Science and Engineering, Åbo Akademi University, Finland

Correspondence

Didem Şen Karaman, Department of Biomedical Engineering, Faculty of Engineering and Architecture, Izmir Katip Çelebi University, Izmir, Turkey.

Email: didem.sen.karaman@ikcu.edu.tr

Funding information

Council of Higher Education 100/2000 doctoral scholarship; The Turkish Scientific and Technological Research Council 2211-A BİDEB doctoral scholarship; The Turkish Scientific and Technological Research Council, Grant/Award Number: 319S024

Abstract

The recalcitrant nature of biofilms makes biofilm-associated infections difficult to treat in modern medicine. Biofilms have a high vulnerability to antibiotics and a limited repertoire of antibiotics could act on matured biofilms. This issue has resulted in a gradual paradigm shift in drug discovery and therapy, with anti-biofilm compounds being sought alongside new drug carriers. A potential solution to biofilm-associated infections is to employ antibiofilm treatments, which can attack biofilms from many fronts. Nanocarriers are promising in this regard because they can be entrapped within biofilm matrix, target biofilm matrix, and provide local drug delivery to inhibit biofilm formation. In this study, curcumin as an herbal extract was loaded onto hyperbranched polyethylenimine-grafted mesoporous silica nanoparticles (F-MSN-PEI/Cur) and antibiofilm investigations were performed. The F-MSN-PEI/Cur design has the potential to repurpose curcumin as an antibiofilm agent by increasing its solubility and lowering the required doses for the destruction of matured biofilms as well as suppressing biofilm development. Using imaging and spectroscopic techniques, we assessed the interaction of F-MSN-PEI/Cur with *Staphylococcus aureus* bacterial cells and determined the impact of F-MSN-PEI/Cur on eradicating matured biofilms and suppressing biofilm development. The F-MSN-PEI/Cur design is highly cytocompatible, as observed by the cytotoxicity screening investigations on L929 mouse fibroblast cell line. Our findings show that F-MSN-PEI/Cur design reduces the bacterial cell viability, inhibits biofilm formation, and induces biofilm eradication, which is attributed to F-MSN-PEI/Cur design having the potential to repurpose the antibiofilm activity of curcumin-herbal extract.

KEYWORDS

antibacterial, antibiofilm, curcumin, mesoporous silica, polyethyleneimine, *Staphylococcus aureus*

1 | INTRODUCTION

Infectious diseases caused by bacteria have been life threatening due to increased resistance to antibiotics and the intricacy of the infection.

Disease progression and the response of the host to the infection reveal two different clinical conditions, acute versus chronic infection.¹ Chronic infections are highly associated with aggregated/attached clusters of planktonic cells, termed the biofilm mode of

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2022 The Authors. *Journal of Biomedical Materials Research Part B: Applied Biomaterials* published by Wiley Periodicals LLC.

growth, with a delayed healing process.² A biofilm is a structured bacterial community embedded in a three-dimensional matrix that attaches to a solid surface or aggregates into clusters. The self-produced, so-called extracellular polymeric substances (EPS) matrix is responsible for the protection and structural stability of bacteria cells which also results in tolerance to antimicrobial treatments and environmental stress. Antibiotic tolerance in biofilms may be due to (i) EPS components preventing antibiotic diffusion through biofilms, (ii) metabolic activity and growth rate gradients within the biofilm restricting antibiotic uptake by bacteria cells, and (iii) antibiotic degradation by enzymes secreted by biofilm bacteria.³ Therefore, biofilms persist with antibiotic therapy up to 1000 times more than required for planktonic bacteria treatments.⁴ The emergence of biofilm-associated infections has increased the demand for novel approaches for delivering bactericidal drugs through biofilms. One intriguing strategy is to use nanoparticles (NPs) to deliver not only antibiotics but also natural bactericidal agents to handle the issue of biofilm-associated infections, which account for more than 60% of chronic infections in people.³

With the support of gained knowledge from research in oncology-related nanomedicine, NPs have become attractive for the treatment of infectious diseases. Among existing NP, mesoporous silica nanoparticles (MSN) can provide multifunctionality (i.e., inherently therapeutic, drug delivery, targeted delivery, precision in dosing) due to their modular design options.⁵ The recent advancements in NPs aided antibacterial treatments and already well-established research on MSN-aided oncology treatments^{6,7} have driven our interest in achieving MSN bactericidal delivery system for the treatment of biofilm-associated infections due to the unique conceptual similarities between biofilms and tumor microenvironments.⁵ In comparison to the currently used antibacterial NPs for bacterial growth inhibition, modular MSN-based bactericidal carriers may offer distinct advantages, including entrapment and penetration within the biofilm matrix via attachment to the EPS outer surface followed by migration into the biofilm. Furthermore, silica chemistry can be exploited to provide surface modification on MSN surfaces once for the benefit of high cargo loading, penetration through the biofilm matrix and better interactions with bacterial cells. In this regard, the allocation of primary, secondary, and tertiary amines on MSN surfaces via polyethylenimine (PEI) surface grafting is a legitimate strategy to deliver permeabilizing effects and disintegrate the bacterial cell membranes.^{8,9} As a result, MSN-PEI design can function as antibacterial agent carriers while also improving therapeutic efficacy by avoiding antimicrobial agent recognition and deactivation in the biofilm matrix, providing targeted delivery to cells, and increasing local concentration and antibacterial activity of bactericidal agents against bacterial cells. MSN-PEI drug carriers have the potential to improve the efficacy of bactericidal agents by repurposing existing medications and minimizing the development of drug resistance, primarily by lowering the drug concentration required for the treatment and clearance of biofilm infections.⁸

Curcumin, the primary component of turmeric, has been shown to possess a variety of pharmacological properties and to exert antibacterial activity when combined with other antibacterial

agents.^{9–13} Although curcumin's bactericidal property is imposed in combination with other compounds, its use alone is hampered by its low water solubility, stability in physiological environment, and bio-availability.¹⁴ It is classified in Class IV according to Biopharmaceutical Classification System (BCS) and its usage results in low absorption, inadequate distribution, and rapid elimination from living systems.^{15–17} It was indicated once the curcumin is orally administered (up to 12 g/day) less than 1% of it culminated in human circulation system.¹⁸ Moreover, curcumin has been noted as having very low water solubility of only 0.6 µg/ml and is susceptible to degradation particularly under alkaline conditions.¹⁹ The use of curcumin is limited due to low water solubility under acidic or neutral conditions, high decomposition rate in alkaline media and photodegradation in organic solvent.²⁰ More in detail, the degradation of curcumin in series of pH conditions ranging from 3 to 10 has been shown and the result revealed that decomposition was pH-dependent and occurred faster at neutral-basic conditions.²¹ In vitro studies showed that more than 90% of curcumin tends to degrade in 30 min under physiological pH conditions (0.1 M phosphate buffer solution, 37 °C, pH 7.2).²² Surfactants, polymer mixtures, and the formation of inclusion complexes encompass the most used and successful approaches employed in the literature.²³ Obvious approach to improve the biopharmaceutical properties of curcumin is to overcome water solubility limitations of curcumin by employing nanocarriers.²⁴ In light of our previous study wherein we have shown the loading of curcumin into MSN possessing different surface chemistries could improve the interaction of curcumin without degradation due to the presence of surface-grown PEI on MSN by providing strong interaction between PEI and curcumin.²⁵ As such, the use of polyethylenimine surface grafted MSN (MSN-PEI) as a carrier for curcumin is anticipated to be a promising strategy for repurposing curcumin as an antibiofilm agent by improving the water solubility and without providing combinatory antibiotic treatments.

In this study, we interpreted the potency of hyperbranched polyethylenimine (PEI) modified MSN as the curcumin carrier for inhibiting biofilm formation and eradicating matured biofilms. The interaction of F-MSN-PEI/Cur and planktonic bacterial cells was evaluated to resolve the rationale behind the antibiofilm efficacy of curcumin-loaded fluorescently labeled MSN-PEI (F-MSN-PEI/Cur). Our findings show that nanocarriers design, PEI surface grafted MSN significantly increase the bactericidal effect of curcumin while also providing operability in eradicating matured *S. aureus* biofilms and inhibiting the formation of *S. aureus* biofilm.

2 | MATERIALS AND METHODS

2.1 | Materials

Tetraethyl orthosilicate (TEOS, reagent grade, 98%, Sigma-Aldrich) and aminopropyl triethoxysilane (APTES, Sigma-Aldrich) were used as silica sources in the synthesis of MSN. Absolute ethanol (99% purity, IsoLAB), cetyltrimethylammonium bromide (CTAB, Merck), sodium hydroxide (Merck) and ammonium nitrate (Carlo Erba) were used as

received for MSN synthesis. Milli-Q water (18.2 M Ω cm) was used throughout the study. Fluorescein isothiocyanate (FITC) from Sigma-Aldrich was used for the fluorescent labeling of MSN. Curcumin was purchased from Sigma-Aldrich. Tryptic Soy Broth (Merck), PBS buffer saline (Sigma-Aldrich), Dulbecco's Modified Eagle Medium (Lonza Bio-Whittaker) (with 25 mM HEPES buffer and 4.5 g/L glucose) with an addition of 10% heat-inactivated fetal bovine serum (FBS) (Gibco, Thermo Scientific), 2 mM L-glutamine (Sigma-Aldrich) and 10 μ l/ml of penicillin–streptomycin (Sigma-Aldrich) was used in *in vitro* studies. The *S. aureus* strain DSM 20321 was provided by Dr. Tuba Müderris from the Department of Medical Microbiology, Izmir Katip Celebi University.

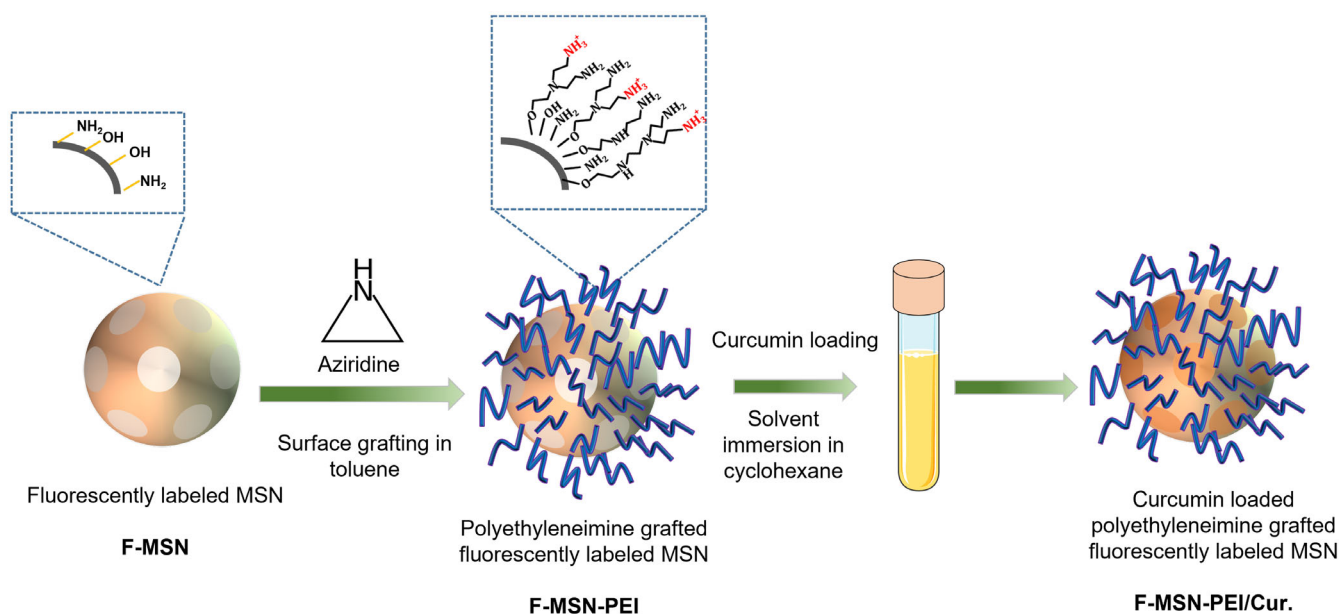
2.2 | Preparation of curcumin-loaded fluorescently labeled mesoporous silica nanoparticles

Fluorescein isothiocyanate (FITC)-labeled MCM-41-type MSN synthesis was performed employing the synthesis protocol in our previous study.²⁶ In brief, a basic aqueous solution was prepared with the addition of absolute ethanol (20% vol/vol) as a co-solvent in the presence of the structure-directing agents cetyltrimethylammonium bromide (CTAB), tetraethyl orthosilicate (TEOS), and aminopropyl ethoxy silane (APTES) as silica sources. The reaction was carried out at 33°C overnight. The molar composition of the synthesis solution was 1 TEOS: 0.01 APTES: 1.23×10^{-2} CTAB: 0.31 NaOH: 71.8 EtOH: 1063.8 H₂O. Before reaction, APTES modification with FITC was performed by reacting FITC with APTES in absolute ethanol at a molar ratio of 1:3 and stirring for 2 h to provide successful incorporation within the silica framework. After the overnight reaction, the structure-directing agent (CTAB) was removed by solvent extraction using an ethanolic NH₄NO₃ solution (20 g/L) and the obtained NPs,

namely F-MSN, were vacuum dried to be used for hyperbranched PEI surface modification. The surface modification of MSN with hyperbranched PEI by surface-initiated polymerization was carried out according to the literature protocols,²⁷ and the schematic presentation for the PEI surface grafting on F-MSN and curcumin loading was presented in Scheme 1. To initiate PEI polymerization from the MSN surfaces, aziridine was used as a monomer with toluene as the solvent, in which the MSN substrate was suspended in the presence of catalytic amounts of acetic acid. The suspension was refluxed under atmospheric pressure overnight at RT, filtered, washed with toluene, and dried under vacuum at 313 K. Henceforth, the obtained fluorescently labeled NPs will be abbreviated as F-MSN-PEI. After obtaining F-MSN-PEI, dried samples were taken for curcumin loading. Curcumin was loaded into F-MSN-PEI by the solvent immersion method to obtain a final design of F-MSN-PEI/curcumin, named as F-MSN-PEI/Cur.²⁵ In brief, sonication was used to disperse 20 mg of F-MSN-PEI in 10 ml of cyclohexane. Then, curcumin with an initial concentration of 50% wt/wt was added to the F-MSN-PEI suspension and then sonicated and allowed to vortex for 24 h at room temperature. The next day, the curcumin-loaded F-MSN-PEI (F-MSN-PEI/Cur) was centrifuged and dried in an oven.

2.3 | Physicochemical characterizations of F-MSN-PEI/Cur

The dried, extracted and PEI surface-grafted MSN as the curcumin carrier has been subjected to physicochemical characterization to explore the features of the F-MSN-PEI/Cur and the carrier F-MSN-PEI design. The size distribution of resuspended dry F-MSN-PEI in HEPES buffer at pH 7.2 was investigated by subsequent dynamic lights scattering (DLS) measurements (ZetaSizer NanoZS 90, Malvern).



SCHEME 1 Schematic presentation for the reaction steps of F-MSN-PEI/Cur.

The success of one-step hyperbranching polymerization of a surface-grown PEI was confirmed by ζ potential measurements (ZetaSizer NanoZS 90, Malvern) in HEPES buffer solution. The morphology of F-MSN-PEI was investigated by scanning electron microscope (SEM; Carl Zeiss 300VP). The F-MSN-PEI samples were dried under vacuum and then transferred onto a sample holder coated with gold (QUORUM Q150 RES).

Curcumin loading degree was determined by UV-VIS spectrophotometer. For this purpose, F-MSN-PEI/Cur samples were dispersed in ethanol at the concentration of 0.25 mg/ml by ultra-sonication and kept in the sonication bath for 30 min, left for complete elution on rotating wheel overnight.^{25,28} After eluting the adsorbed curcumin from F-MSN-PEI/Cur samples in ethanol, the NPs were centrifuged and supernatants were taken for spectrophotometric investigation. The absorbance value at the highest peak point of the 425 nm wavelength was recorded and the calculations were made by employing the curcumin standard curve in ethanol at various concentrations (1–50 $\mu\text{g/ml}$) with the same instrument and experimental parameters. Net surface charge and hydrodynamic size of F-MSN-PEI and F-MSN-PEI/Cur were determined, respectively, with ζ potential and DLS analyses using NanoZS 90 Zetasizer (Malvern). For measurements, the samples were dispersed in HEPES buffer (25 mM, pH 7.2) at a concentration of 0.25 mg/ml.

After size and morphology investigations, F-MSN-PEI/Cur powders and curcumin were taken for curcumin solubility investigations to affirm the benefit of F-MSN-PEI to improve the bioavailability of bactericidal curcumin. This particular investigation was employed at high curcumin content, which was foreseen as the needed dosing in investigating the bactericidal effect of curcumin for biofilm inhibition and eradication. The curcumin solubility investigations were performed by following the literature protocols. The solubility of curcumin was performed in buffer solution at pH 7.2 by adding 0.1% (wt/vol) Tween 80 to restrain errors due to poor solubility which is a frequently used strategy in literature studies while working with low soluble molecules.^{29–33} Tween 80 (Sigma-Aldrich) was used as a surfactant and a non-ionic emulsifier to prevent aggregation due to poor solubility of curcumin at physiological pH.³⁴ In detail, F-MSN-PEI/Cur powders were suspended as 320, 640 $\mu\text{g/ml}$ in aqueous release media (HEPES, pH 7.2, 25 mM containing 0.1% (wt/vol) Tween 80) possessing curcumin content of 57.6 and 115.2 $\mu\text{g/ml}$ and was taken into Float-A-Lyzer G2 dialysis device (MWCO:3.5–5 kDa) to investigate the improved solubility of curcumin by F-MSN-PEI carrier. The counterpart investigations were performed also for curcumin powder, which has equivalent curcumin content in the carriers. Briefly, 320 and 640 $\mu\text{g/ml}$ F-MSN-PEI/Cur samples were immersed in aqueous media containing with a 0.1% (wt/vol) Tween 80. The curcumin solubility from the F-MSN-PEI/Cur was investigated by employing a UV-visible spectrophotometer. After 24 h, F-MSN-PEI/Cur suspension from the Float-A-Lyzer G2 dialysis device was centrifuged for 10 min at 10,000 rpm and supernatants were measured by Synergy HTX Multimode Plate Reader for solubility examination. The solubility of curcumin was calculated with a standard curve comprising various curcumin concentrations in the release medium. The same

investigations were performed also for free curcumin in immersed media where the dialysis bag was immersed for 24 h. The solubility was determined in three replicates and measured three times. The results were statistically analyzed with two-way ANOVA, followed by Sidak's multiple comparison test using GraphPad Prism (version 8.4.2). In order to solidify F-MSN-PEI nanocarriers-aided curcumin solubility enhancement whether it is due to the obtained differences in the solid state of the curcumin crystals on nanocarriers, differential scanning calorimetry (DSC) investigations were also performed for curcumin powder, F-MSN-PEI and F-MSN-PEI/Cur powders. All samples were scanned by a scanning rate of 5 K/min. An empty aluminum crucible was used as a reference sample and the temperature scale of the calorimeter. A quantity of 2–6 mg (total weight) was used for the DSC measurements.

2.4 | In vitro cytotoxicity investigations of F-MSN-PEI/Cur

Cytocompatibility assessment of F-MSN-PEI/Cur was investigated in the L929 mouse fibroblast cell line. Fibroblast cells were cultured in a basal medium comprised of Dulbecco's Modified Eagle Medium (DMEM), 10% FBS, and 1% penicillin-streptomycin. Furthermore, the cells were seeded onto a standard cell culture plate at a concentration of 10^4 cells/well and incubated overnight at 37°C in 5% CO₂ environment to allow cell attachment. The following day, F-MSN-PEI/Cur was dispersed in filtered HEPES buffer and diluted with the basal medium at a concentration of 10, 25, 50, 100, and 200 $\mu\text{g/ml}$. Fibroblast cells were treated with diluted F-MSN-PEI/Cur and, subsequently incubated at 37°C with 5% CO₂ for 24, 48, and 72 h. After incubation, treated cells were rinsed gently with sterile phosphate buffer saline (PBS) to remove NPs. Cell viability was determined by the 3-[4,5-dimethylthiazol-2-yl]-2,5 diphenyl tetrazolium bromide (MTT) assay. Serum-free DMEM containing 10% (vol/vol) MTT stock solution (5 mg/ml) was added to treated cells and then incubated at 37°C. After incubation, formazan crystals were dissolved using dimethyl sulfoxide (DMSO) and absorbance values were measured at 570 nm by Synergy HTX Multimode Plate Reader.

2.5 | In vitro antibacterial and antibiofilm investigations of F-MSN-PEI/Cur

2.5.1 | Determination of bacterial growth inhibition by F-MSN-PEI/Cur

Staphylococcus aureus (DSM 20321) have been employed as a gram-positive model organism as a resemble of bacterial strains causing infection.^{35,36} To investigate the impact of F-MSN-PEI/Cur on the planktonic growth form of the bacterial cell, the growth profiles of F-MSN-PEI/Cur-treated bacterial cells were investigated. For this purpose, the optical density values of the F-MSN-PEI/Cur treated bacteria during 18 h were compared to the untreated bacteria suspensions.

In detail, *S. aureus* culture was grown overnight by inoculation of tryptic soy broth (TSB) from a single colony at 37°C and 150 rpm under aerobic conditions. The following day, the overnight grown cultures were diluted at 1:50 in TSB for 2 h to reach the exponential growth phase. Subsequently, the *S. aureus* culture concentration was adjusted to 10⁸ CFU/ml by optical density measurement at 600 nm (OD 600) using Synergy HTX Multimode Plate Reader. The bacteria suspension was then diluted in TSB to a final concentration of 2 × 10⁵ CFU/ml. A 100 µl of F-MSN-PEI/Cur (the final concentrations of 25, 50, 100, 200, 400, 800, and 1600 µg/ml) were pipetted into the wells of 96-well microtiter plates. Subsequently, 100 µl of 2 × 10⁵ CFU/ml *S. aureus* suspension was added to each well. The plate was incubated at 37°C for 18 h and OD 600 measurement was carried out with 30 min intervals. Non-treated bacteria were used as negative control while F-MSN-PEI/Cur suspensions in TSB without bacteria were used as the blank of the actual samples. By this way, we eliminated the optical density interference arising from components of F-MSN-PEI/Cur, that is, curcumin and FITC and also arisen due to the colloidal status of the particle suspensions. Three replicates were conducted for each group. Subsequently, the bacterial viability assessment was conducted for the same set of investigations to depict the bactericidal efficacy of F-MSN-PEI/Cur on the planktonic growth form of the bacterial cell. However, in this particular investigation, the bacterial cell suspensions were treated with an increasing concentration of F-MSN-PEI/Cur at 37°C for 5 h to investigate the bacterial cell viability. After incubation, resazurin solution (200 µM, resazurin sodium salt, Sigma-Aldrich) was added to each well at a final concentration of 10% (vol/vol). The plate was then incubated at 37°C in a dark environment. Fluorescence measurement was carried out at excitation/emission wavelengths of 530/590 nm using CLARIOstar[®] multimode reader. Non-treated bacteria suspensions were employed as negative controls, while F-MSN-PEI/Cur suspensions without bacteria were used as blank to eliminate F-MSN-PEI/Cur induced backgrounds.

2.5.2 | Determination of biofilm inhibition and eradication degree after F-MSN-PEI/Cur treatments

Inhibiting the biofilm formation by F-MSN-PEI/Cur treatments

The inhibition of biofilm formation by F-MSN-PEI/Cur and equivalent free curcumin content was performed by treating the bacteria cells with increasing concentrations of F-MSN-PEI/Cur suspensions and equivalent quantities of free curcumin solutions during the of biofilm formation. In detail, overnight culture of *S. aureus* was set to 10⁸ CFU/ml by OD 600 measurement followed by dilution in TSB supplemented with 0.2% (vol/vol) glucose to obtain a concentration of 2 × 10⁵ CFU/ml. Afterwards, 100 µl of F-MSN-PEI/Cur (the final concentrations 400, 800, and 1600 µg/ml) and in parallel 100 µl of curcumin solutions (the final concentrations 72, 144, and 288 µg/ml as equivalent amount in F-MSN-PEI/Cur treatments) were pipetted into the wells of 96-well microtiter plates. Then, 100 µl of 2 × 10⁵ CFU/ml *S. aureus* suspension was pipetted into the wells to obtain final *S. aureus*

bacterial concentration 10⁵ CFU/ml. Suspensions in the wells were gently mixed by pipetting to provide a homogeneous dispersion of the F-MSN-PEI/Cur. For the negative control, 10⁵ CFU/ml *S. aureus* suspension was also incubated. The plates were incubated for 24 h at 37°C and 150 rpm. The quantification of the biofilm viability and biomass were performed after the treatments.

Eradicating the matured biofilm by F-MSN-PEI/Cur treatments

The eradication of mature *S. aureus* biofilms was also assessed with the same strategy. However, in this case, matured *S. aureus* biofilms were obtained as described above with only broth media. After mature biofilms were obtained on the following day, the medium in the wells was removed and the wells were washed with PBS three times to remove planktonic or loosely attached bacteria. Then, the ascending dose of F-MSN-PEI/Cur suspensions and free curcumin solutions were applied in parallel to assess biofilm eradication degree after F-MSN-PEI/Cur treatments, as well as the influence of PEI grafted MSN as a curcumin carrier. The mature biofilms were then treated with F-MSN-PEI/Cur suspensions of 400, 800, and 1600 µg/ml and in parallel equivalent free curcumin concentrations of 72, 144, and 288 µg/ml in HEPES buffer. Plates were incubated for 24 h at 37°C and 150 rpm to determine biofilm availability and total biomass of the treated biofilms. 24 h grown *S. aureus* biofilms started with a concentration of 1 × 10⁵ CFU/ml was used as a negative control. The quantification of biofilm viability and biomass were performed after the treatments.

Biofilm inhibition and eradication degree analysis after F-MSN-PEI/Cur treatments

(i) Quantifying the biofilm viability and total biomass

To evaluate the degree of biofilm inhibition and eradication upon exposure to F-MSN-PEI/Cur suspensions and curcumin solutions, biofilm viability, and biomass were quantified using resazurin and safranin assays, respectively. To this end, after the treatments in Section 2.5.2 the supernatants in the wells were gently discarded, and biofilms were washed with PBS thrice to remove loosely attached bacteria. For biofilm viability analysis, 200 µl of 20 µM resazurin solution diluted in PBS was added to each well. The plate was incubated at room temperature and 150 rpm until a color change was observed in the negative control group. Fluorescence measurement was carried out at excitation/emission wavelength of 530/590 nm using CLARIOstar[®] multimode reader. For quantification of biomass, safranin dye was employed due to its known ability to bind to negative charges and therefore target many different molecules of bacteria and EPS.^{37,38} After the treatments in Section 2.5.2 the supernatants in the wells were gently removed, and the biofilms were washed with PBS thrice. Thereafter, 200 µl of 0.1% (vol/vol) safranin solution was added to each well, and the plates were incubated at room temperature for 30 min. After incubation, safranin solutions were discarded, wells were washed with PBS twice to remove excess safranin, and the plates were air-dried at room temperature. A 200 µl of 30% (vol/vol) glacial acetic acid was then pipetted into each well to dissolve safranin

TABLE 1 Interpretation of biofilm production.

Average OD	Biofilm production
$OD \leq OD_c$	Non-biofilm producer
$OD_c < OD \leq 2 \times OD_c$	Weak biofilm producer
$2 \times OD_c < OD \leq 4 \times OD_c$	Moderate biofilm producer
$4 \times OD_c < OD$	Strong biofilm producer

staining on biofilms. The biofilm mass was determined by measuring safranin absorbance at 525 nm using a Synergy HTX Multimode Plate Reader.

(ii) Estimating biofilm production ability of *S. aureus* during biofilm inhibition studies

The ability of the *S. aureus* cells for biofilm production after exposure to F-MSN-PEI/Cur suspensions was evaluated by employing the criteria described by Hassan et al. as demonstrated in Table 1.³⁹ Biofilm production capacity of F-MSN-PEI/Cur treated bacterial cells was interpreted by using the OD_{525} values obtained from the safranin assay. For comparison, the optical density cutoff value (OD_c) was calculated as mean (μ) OD of uninoculated TSB + $3 \times SD$ (σ) of uninoculated TSB.

(iii) Investigating F-MSN-PEI/Cur entrapment within *S. aureus* biofilm

The degree of the F-MSN-PEI/Cur entrapment within the *S. aureus* biofilm was evaluated by the fluorescent intensity obtained due to FITC labeled F-MSN-PEI/Cur in order to ensure the diffusing ability of the F-MSN-PEI/Cur throughout to matured biofilm matrix.⁴⁰ For this purpose, the matured *S. aureus* biofilms were prepared as described in Section 2.5.2. Then, F-MSN-PEI/Cur suspensions in HEPES buffer were added to wells containing *S. aureus* biofilms at three different concentrations (400, 800, and 1600 $\mu\text{g/ml}$). After 24 h., the suspensions on the biofilms containing non-entrapped F-MSN-PEI/Cur in the wells were transferred to a 96-well plate and the fluorescence emission was determined using the Synergy HTX Multimode Plate Reader at excitation/emission wavelengths of 485/528 nm. The experimental design used F-MSN-PEI/Cur suspensions in the absence of *S. aureus* biofilms at same treatment concentrations as those used in the presence of *S. aureus* biofilms as the starting point for fluorescent intensity measurements. To calculate the entrapped F-MSN-PEI/Cur concentration within the *S. aureus* biofilm, the difference between the initial F-MSN-PEI/Cur concentration and the non-entrapped F-MSN-PEI/Cur concentration was calculated and used to calculate the F-MSN-PEI/Cur entrapment percentage. Afterwards, the concentration-dependent entrapment efficiency of F-MSN-PEI/Cur was then calculated using the following equation as described by Devlin et al.⁴⁰

$$F-MSN-PEI/Cur \text{ entrapment\%} = \frac{F_c - F_e}{F_c} \times 100$$

where F_c is the fluorescent intensity of the F-MSN-PEI/Cur suspensions, F_e is the fluorescent intensity of the F-MSN-PEI/Cur suspension entrapped within the *S. aureus* biofilms during 24 h. Therefore, the entrapped percentile of the added F-MSN-PEI/Cur suspensions within the *S. aureus* biofilms was evaluated.

(iv) Electron microscopy imaging of *S. aureus* biofilm after exposure to F-MSN-PEI/Cur treatment

The biofilms of *S. aureus* after exposure to the highest dose of F-MSN-PEI/Cur (1600 $\mu\text{g/ml}$) were investigated by scanning electron microscopy (SEM) for both inhibition and eradication studies to predict the architectural changes of the bacteria cells and biofilm matrix due to the F-MSN-PEI/Cur treatment. In detail, *S. aureus* biofilm was cultured in 24-well plates containing sterile glass coupons with a starting concentration of 6×10^5 CFU/ml with simultaneous addition of F-MSN-PEI/Cur for inhibition or addition of F-MSN-PEI/Cur to mature biofilm for eradication. After 24 h incubation, biofilms were washed with $1 \times$ PBS thrice to remove unattached bacteria and NPs. The *S. aureus* biofilms were then fixed in 2.5% (vol/vol) glutaraldehyde for 1 h., washed gently with PBS three times and air-dried at room temperature, respectively. After fixation, the biofilm samples were gradually dehydrated with increasing concentrations of ethanol (25, 50, 70, 90, 100% (vol/vol)) for 20 min each. *S. aureus* biofilms were sputter coated with gold followed by visualization by Carl Zeiss 300 VP SEM instrument.

2.6 | Data interpretation and statistical analysis

All experiments were carried out in triplicates and data are represented as mean \pm SD. Statistical analyses were performed using GraphPad Prism software (version 8.4.2) with the level of significance (α) set to 0.05.

During all studies comprising biofilm inhibition, eradication and entrapment, biofilm, biofilm + curcumin and biofilm + F-MSN-PEI/Cur were also prepared as blanks for resazurin and safranin assays to eliminate biofilm, FITC, and curcumin-induced interferences as the background. Nevertheless, some peaks for safranin were overlapped with curcumin peaks in absorbance spectra even after subtracting the blanks. This problem has been overcome by decomposition of the safranin and curcumin peaks using peak analyzer tool in OriginPro software (version 2019b) to process the area only under the safranin peaks for total biomass degree calculations.

The robustness of O.D. reading, safranin and resazurin assays were evaluated by calculating Z' factor, screening window coefficient, with the following equation:

$$Z' = \frac{1 - (3 \times \sigma_{\max} + 3 \times \sigma_{\min})}{|\mu_{\max} - \mu_{\min}|}$$

where μ_{\max} and μ_{\min} are the maximum and minimum mean of assay signals, and σ_{\max} and σ_{\min} are the SDs of the maximum and minimum

assay signals. The test method is interpreted as appropriate if the Z' factor is between 0.5 and 1, acceptable in some cases if it is between 0 and 0.5, and not acceptable if the Z' factor value is <0.

3 | RESULTS

3.1 | Characterization of the F-MSN-PEI/Cur

The synthesis protocol yielded spherically shaped monodisperse F-MSN-PEI and F-MSN-PEI/Cur in the size range of 150–250 nm (Figure 1A,B). 250 µg/ml F-MSN-PEI and F-MSN-PEI/Cur NP suspensions were dispersed in HEPES buffer (pH 7.2, 25 mM) for hydrodynamic size and net surface charge (ζ -potential) measurements, respectively (Figure 1C). The hydrodynamic size and ζ -potential data are presented in Table 2. The F-MSN-PEI samples were fully dispersible in HEPES buffer after all processing steps, including template removal, surface modification of F-MSN with polyethylenimine

(F-MSN-PEI) and further curcumin loading (F-MSN-PEI/Cur). The success of surface modification was ensured by the ζ -potential measurements as presented in Table 2. The re-dispersibility of F-MSN-PEI/Cur was investigated. The results obtained (Table 2) revealed that the loaded drug content does not cause an adverse effect on the re-dispersibility of F-MSN-PEI even the hydrodynamic radius and the polydispersity index (PDI) values increased, in the acceptable range of PDI 0.1–0.4, which accounted for monodispersed size distribution. Furthermore, after curcumin loading, the ζ -potential values could be produced with net positive values ($+21 \pm 4.8$ mV).

The degree of curcumin loading on F-MSN-PEI/Cur was determined to be 18% (wt/wt). The solubility of curcumin from the F-MSN-PEI carrier was evaluated to verify F-MSN-PEI is effective in enhancing curcumin solubility together with the commonly employed strategy of immersion into surfactant (i.e., non-ionic Tween 80) added media.²³ To understand the effect of F-MSN-PEI on curcumin solubility, solubilized curcumin with the aid of F-MSN-PEI nanocarriers at different concentrations of 320 and 640 µg/ml was investigated, and

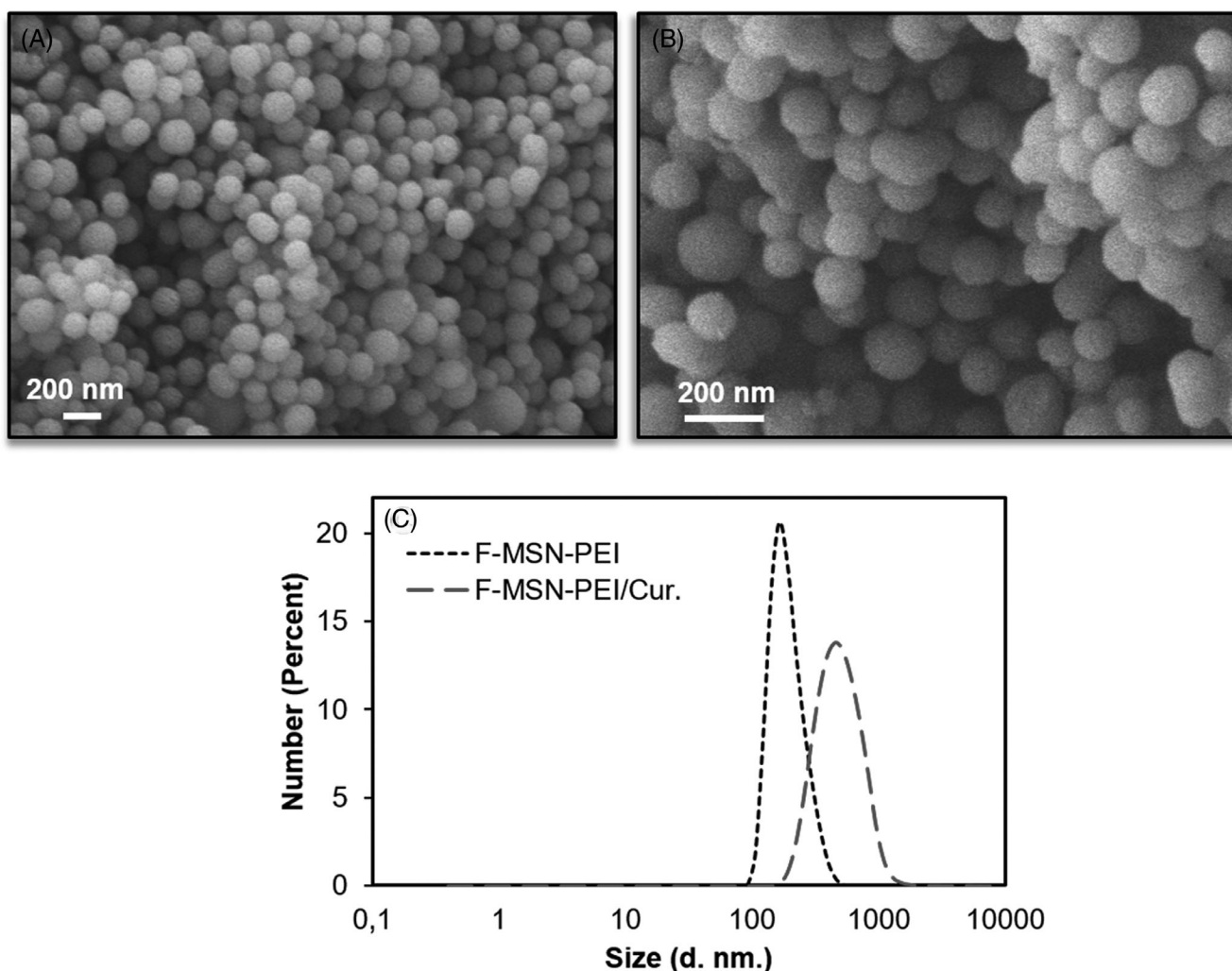


FIGURE 1 (A) SEM micrograph of F-MSN-PEI, (B) SEM micrograph of F-MSN-PEI/Cur, (C) Hydrodynamic size distribution of F-MSN-PEI and F-MSN-PEI/Cur.

TABLE 2 ζ potential, hydrodynamic radius and PDI results of F-MSN and F-MSN-PEI/Cur. The results are expressed as mean \pm SD ($n \geq 3$).

Characterization	F-MSN-PEI	F-MSN-PEI/Cur
ζ Potential (mV)	44.3 \pm 6.5	21.2 \pm 4.1
Hydrodynamic radius (nm)	236.1 \pm 3.2	689.3 \pm 106
Polydispersity index (PDI)	0.061 \pm 0.02	0.316 \pm 0.04

the free curcumin solutions were subjected to same spectrophotometric analysis. The solubility investigations were performed for F-MSN-PEI/Cur which contains 57.6 and 115.2 $\mu\text{g/ml}$ curcumin and the counterpart free curcumin solutions at the concentration of 57.6 and 115.2 $\mu\text{g/ml}$. In this particular investigation, we have employed higher concentrations than the solubility limits, which was foreseen as the needed dosing in investigating the bactericidal effect of curcumin in the circumstance of biofilm inhibition and eradications. The observations showed that free curcumin solubility could be improved up to 58.6 \pm 1.3 $\mu\text{g/ml}$ and further 75.6 \pm 0.4 $\mu\text{g/ml}$ in 0.1% (wt/vol) Tween 80 (non-ionic surfactant) aqueous solution once the starting concentration of curcumin increased.^{41,42} In the literature investigations, researchers have claimed the solubility of curcumin as 0.6 $\mu\text{g/ml}$ in water¹⁹ whereas in our investigations once the 0.1% (wt/vol) Tween 80 was added to HEPES buffer solution (25 mM, pH 7.2) the curcumin was solubilized 100-fold higher than the literature finding. Our observations shown in Figure 2 indicated that when curcumin loaded onto F-MSN-PEI and immersed in the 0.1% (wt/vol) Tween 80 containing HEPES buffer the solubility of curcumin improved up to 110.6 \pm 5.2 $\mu\text{g/ml}$ which is almost 183-fold higher 0.6 $\mu\text{g/ml}$ where the solubility was investigated in water.¹⁹ This result could be ascribed as 0.1% (wt/vol) Tween 80 could aid the solubilization of curcumin however the incorporation of curcumin into F-MSN-PEI is more effective for improving the curcumin solubility compared to only 0.1% (wt/vol) Tween80 addition into immersion medium.

Furthermore, in order to solidify F-MSN-PEI nanocarriers-aided curcumin solubility enhancement regardless of the presence of the solubility enhancer (Tween 80, non-ionic surfactant presence) in the immersion media, DSC investigations were performed for the F-MSN-PEI/Cur and curcumin powders (Figure S2). Shifts of exothermic and endothermic peaks are usually associated with interactions between drug and carrier.⁴³ However, in our investigations we have not observed a distinct shift in the melting point of curcumin, which could be indicated as the drug molecules could have been adsorbed via weak interactions by diffusing into pores and as aggregated in the PEI polymeric network of F-MSN-PEI carrier which may still lead the accommodation of both amorphous or disordered crystalline state. In Figure S2, a broader peak was observed for F-MSN-PEI/Cur which could be accepted as the indicative of a disturbance in the crystalline habit of the curcumin once in the F-MSN-PEI/Cur formulation which can be ascribed to a physical interaction between the excipients during processing. Almost 5-fold ΔH reduction was also observed as

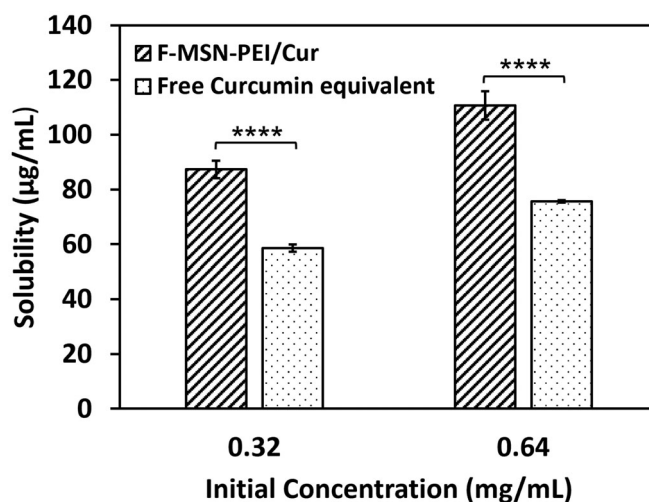


FIGURE 2 Solubility of curcumin in the F-MSN-PEI/Cur design in comparison to free equivalent curcumin content at different concentrations. The results are expressed as mean \pm SD ($n \geq 3$). Statistical analysis was performed between F-MSN-PEI/Cur and equivalent curcumin at each concentration by applying two-way ANOVA followed by Sidak's multiple comparison test. * $p < .03$, ** $p < .002$, *** $p < .0002$, **** $p < .0001$

given in Figure S2 for F-MSN-PEI/Cur compared to curcumin powder. This could be attributed to the disorganization of curcumin crystals after loading into F-MSN-PEI nanocarriers.⁴⁴ The slightly lower T_m could be ascribed for this, while the $\Delta T_{1/2}$ remains unchanged.

3.2 | In vitro cytotoxicity of F-MSN-PEI/Cur

The cytocompatibility of F-MSN-PEI/Cur was evaluated and F-MSN-PEI/Cur was found to be cytocompatible at concentrations up to 100 $\mu\text{g/ml}$ (Figure 3) over 24, 48, and 72 h incubation periods. When the dosing of F-MSN-PEI/Cur is increased higher than the 10 $\mu\text{g/ml}$ actively proliferating cells were observed as shown in Figure 3 for 24 h treatments. However, the Figure 3 results reveal when the incubation time is increased the proliferative effect is not dominating as shorter term incubating.

3.3 | In vitro antibacterial and antibiofilm investigations of F-MSN-PEI/Cur

3.3.1 | Bacterial growth inhibition and bactericidal effect of F-MSN-PEI/Cur

The antibacterial activity of F-MSN-PEI/Cur were verified by incubating *S. aureus* suspensions with F-MSN-PEI/Cur followed by optical measurement (OD 600) of bacteria suspension. The optical density values of the F-MSN-PEI/Cur treated bacteria cell suspensions and untreated bacteria suspensions are presented in Figure 4. *S. aureus*

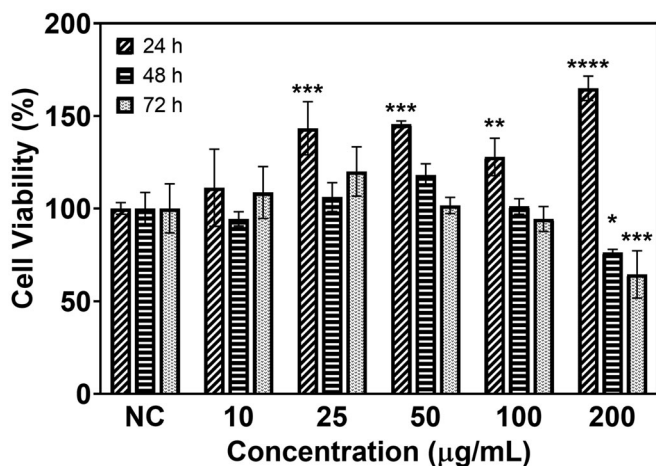


FIGURE 3 In vitro cytocompatibility of F-MSN-PEI/Cur with L929 cells. Error bars represent SD ($n \geq 3$). The results were statistically analyzed with the GraphPad Prism software (V 8.4.2) using two-way ANOVA followed by the Dunnett multiple comparisons test. The degree of significance was $*p < .03$, $**p < .002$, $***p < .0002$, $****p < .0001$.

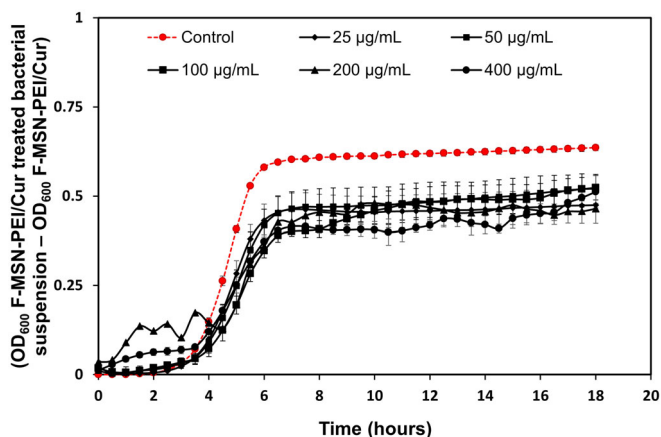


FIGURE 4 Growth profile of *S. aureus* cells upon exposure to ascending concentrations of F-MSN-PEI/Cur suspensions during 18 h to investigate the changes in growth profile in the presence of F-MSN-PEI/Cur. Error bars represent SD ($n \geq 3$).

bacterial cell growth profiles tends to change due to the F-MSN-PEI/Cur treatment. In order to clarify impact of the optical density changes due to the colloidal interference of the F-MSN-PEI/Cur we have followed the optical density of F-MSN-PEI/Cur suspensions in TSB (Figure S1). Obtained results showed drastic fluctuations and changes in optical density values when F-MSN-PEI suspensions higher than 400 $\mu\text{g}/\text{ml}$. The observed trend in Figure S1 could be due to the flocculation/sedimentation of the F-MSN-PEI/Cur content in the growth medium which may lead to the optical density fluctuations. In order to emphasize the elimination of NPs originated optical density disturbance the y-axis of Figure 4 has been presented by subtracting the optical density values of F-MSN-PEI/Cur suspensions as the background from F-MSN-PEI/Cur treated bacterial suspensions. The lowest optical density values were obtained by 400 $\mu\text{g}/\text{ml}$ treatment of

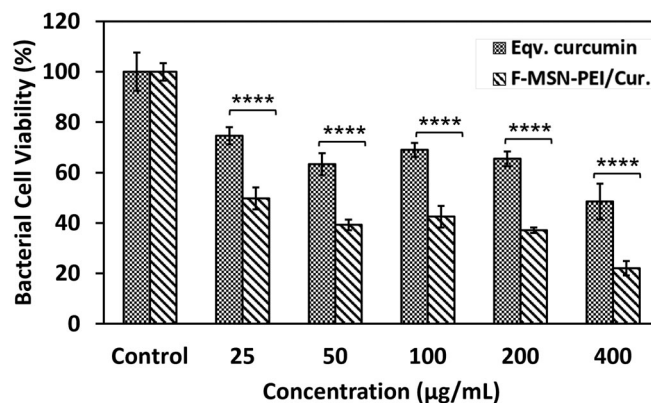


FIGURE 5 Bacterial cell viability investigation after 3 h treatment of bacterial suspensions of *S. aureus* with ascending F-MSN-PEI/Cur formulation. Statistical analysis was performed between F-MSN-PEI/Cur and equivalent curcumin at each concentration by applying two-way ANOVA followed by Sidak's multiple comparison test. $*p < .03$, $**p < .002$, $***p < .0002$, $****p < .0001$

the bacteria suspension (Figure 4) and the trend changes in the exponential growth phase were observed with the high concentration treatments (100, 200, and 400 $\mu\text{g}/\text{ml}$) compared to 25 and 50 $\mu\text{g}/\text{ml}$ F-MSN-PEI treatments.

F-MSN-PEI/Cur and equivalent free curcumin amount loaded into F-MSN-PEI/Cur were then studied for its short-term bactericidal action. The metabolic activity of the bacteria was studied using a resazurin assay after incubation of bacterial suspensions with increasing F-MSN-PEI/Cur and free curcumin dosing for 3 h (the beginning of the exponential growth phase of control bacteria). As the dose of F-MSN-PEI/Cur was increased, the metabolic activity of bacterial cells decreased, which is often related with reduced bacterial cell survival as shown in Figure 5. F-MSN-PEI/Cur dosing at 25 $\mu\text{g}/\text{ml}$ inhibited bacterial cell viability by almost 50%, while higher dosings of 200 and 400 $\mu\text{g}/\text{ml}$ reduced cell viability down to $38.03 \pm 1.11\%$ and $21.34 \pm 2.87\%$, respectively. The treatment with curcumin, on the other hand, did not result in the same degree of cell viability reduction as seen in the Figure 5. However, the substantial interference by F-MSN-PEI/Cur in testing media was observed for the fluorescence reading of resazurin in these short-term investigations. Therefore, we did not execute the investigations with the higher concentrations of 800 and 1600 $\mu\text{g}/\text{ml}$ in this particular investigation.

3.3.2 | Biofilm inhibiting and eradicating efficiency of F-MSN-PEI/Cur

The impact of F-MSN-PEI/Cur and free curcumin treatments on inhibiting the *S. aureus* biofilm formation was evaluated. After 24 h treatments, the biofilms were quantified by analyzing total biomass change and the bacterial cell metabolic activity in biofilm matrix, compared to the untreated groups. The experiments were carried out with the 400–800 to 1600 $\mu\text{g}/\text{ml}$ of F-MSN-PEI/Cur dosing and free curcumin equivalent to those in F-MSN-PEI/Cur designs. Figure 6 shows

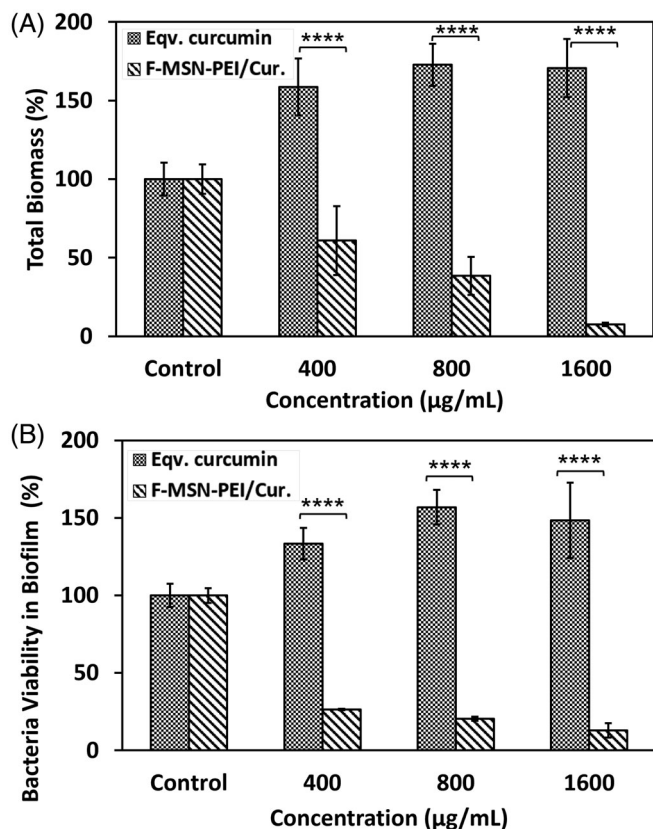


FIGURE 6 Investigation of the biofilm inhibiting potency of F-MSN-PEI/Cur and equivalent curcumin treatment by (A) safranin assay for total biomass and (B) resazurin assay for bacterial cell viability in biofilm. Statistical analysis was performed between treatment groups by applying two-way ANOVA followed by Sidak's multiple comparison test, ns: 0.1234, * $p < .03$, ** $p < .002$, *** $p < .0002$, **** $p < .0001$.

that almost 60% of the total biomass could be reduced when the bacteria cells were treated with 400 µg/ml of F-MSN-PEI/Cur dosing, whereas the highest dosing of 1600 µg/ml could reduce 90% of the total biomass compared to the negative control group. However, when we treated biofilm-forming bacteria with curcumin, the total biomass and cell viability increment in biofilm matrix was observed in Figure 6 as a comparison to the effect of the F-MSN-PEI/Cur design. The obtained results indicated that 72, 144, and 288 µg/ml curcumin dosing without nanocarriers yielded almost 1.5-fold more biomass compared to negative control groups. The observations indicated statistically significant differences between free curcumin and F-MSN-PEI/Cur groups.

The efficacy of ascending concentrations of F-MSN-PEI/Cur and the free curcumin dosing on matured *S. aureus* biofilms are presented in Figure 7. The total biomass in Figure 7A showed that F-MSN-PEI/Cur treatments reduce the biomass significantly compared to free curcumin groups. The biofilm eradication potency of F-MSN-PEI/Cur is higher than free curcumin. However, equivalent dose of free curcumin treatment in eradication investigation worked more effectively compared to biofilm inhibition investigations. The reduction in the metabolic activity of the bacterial cells in the biofilm matrix was presented

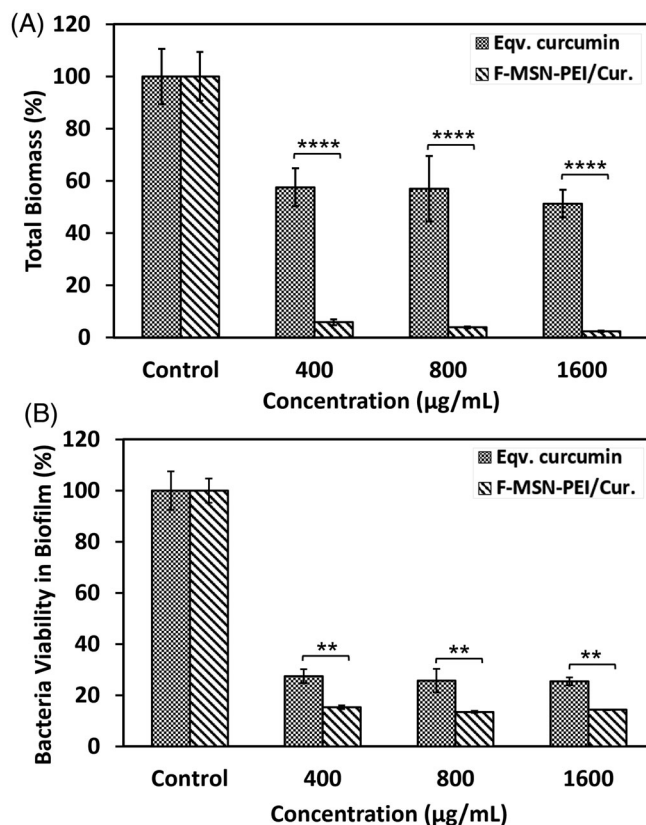


FIGURE 7 Investigation of biofilm eradication after F-MSN-PEI/Cur and equivalent curcumin treatment by (A) safranin assay for total biomass, (B) resazurin assay for bacteria cell viability. Statistical analysis was performed between treatment groups by applying two-way ANOVA followed by Sidak's multiple comparison test, ns: 0.1234, * $p < .03$, ** $p < .002$, *** $p < .0002$, **** $p < .0001$.

in Figure 7B. The obtained results showed statistically significant differences between free curcumin and F-MSN-PEI/Cur treated biofilm viability regardless of the increased dosing.

The Z' factor, which predict the robustness of an assay, was calculated for the optical density reading assays used for bacterial growth profiles, safranin assay for biomass determination and resazurin assay for cell viability to affirm the separation between the distributions of positive and negative controls. The calculations resulted in the Z' factor between 0.1 and 0.6 for safranin assays, which is indicated as a moderate assay. However, the Z factor was calculated as 0.94 for optical density investigations and 0.84 for the resazurin assay, indicating the robustness and reproducibility of the assays.⁴⁵

The biofilm forming capacity of the treated bacterial cells with ascending dosing of F-MSN-PEI/Cur were found that the treatment conditions completely eliminate the biofilm forming ability of *S. aureus* biofilms as presented in Table 3.³⁹

SEM images of *S. aureus* biofilms after exposure to the highest dose of F-MSN-PEI/Cur (1600 µg/ml) for both inhibition and eradication investigations and negative control group were presented in Figure 8 to predict the architectural changes of the biofilm matrix after treatments. Reduction in the biomass for both cases was

obtained compared to untreated condition. In addition, the debris of the biofilm matrix could be also visualized in Figure 8C whereas proper bacterium structures without any destruction could be visualized in Figure 8B.

The F-MSN-PEI/Cur entrapment efficiency into the *S. aureus* biofilm was investigated to rationalize the correlation between the entrapment of F-MSN-PEI/Cur within the biofilm matrix and the biofilm eradication. After incubation of the matured biofilms with F-MSN-PEI/Cur suspensions, the fluorescence intensity of the supernatant containing F-MSN-PEI/Cur that have not yet entrapped into the biofilms and the control wells comprising the same concentration of

F-MSN-PEI/Cur suspensions as starting concentration were measured. The obtained results are presented in Figure 9 demonstrating the entrapment efficiency as $34.11 \pm 3.11\%$, $44.78 \pm 2.76\%$, and $51.12 \pm 3.99\%$ for 400, 800, and 1600 $\mu\text{g/ml}$ respectively. A significant increment in F-MSN-PEI/Cur entrapment could be observed when the dosing increased from 400 to 800 and 1600 $\mu\text{g/ml}$, however, the saturation of biofilm matrix with the F-MSN-PEI/Cur after the 800 $\mu\text{g/ml}$ dosing was observed which could be related with the stacking of the F-MSN-PEI/Cur carrier more in the mid layer and upper layers of the biofilm matrix and not dose depended penetration through the biofilm matrix.

TABLE 3 Interpretation of biofilm formation ability of F-MSN-PEI/Cur treated bacterial cells.

Treatments	OD _c	OD	Interpretation
0 $\mu\text{g/ml}$	0.14	0.30	Moderate biofilm producer
400 $\mu\text{g/ml}$	0.74	0.17	Non-biofilm producer
800 $\mu\text{g/ml}$	0.43	0.11	Non-biofilm producer
1600 $\mu\text{g/ml}$	0.07	0.002	Non-biofilm producer

4 | DISCUSSION

S. aureus is emerging as a problematic opportunistic pathogen that causes osteomyelitis, endocarditis, indwelling device infections, and wound infections.⁴⁶ Due to the limited repertoire of antibiotics available today, there is a high demand for developing a new solution to overcome the failures of antibiotic treatments in *S. aureus* originated

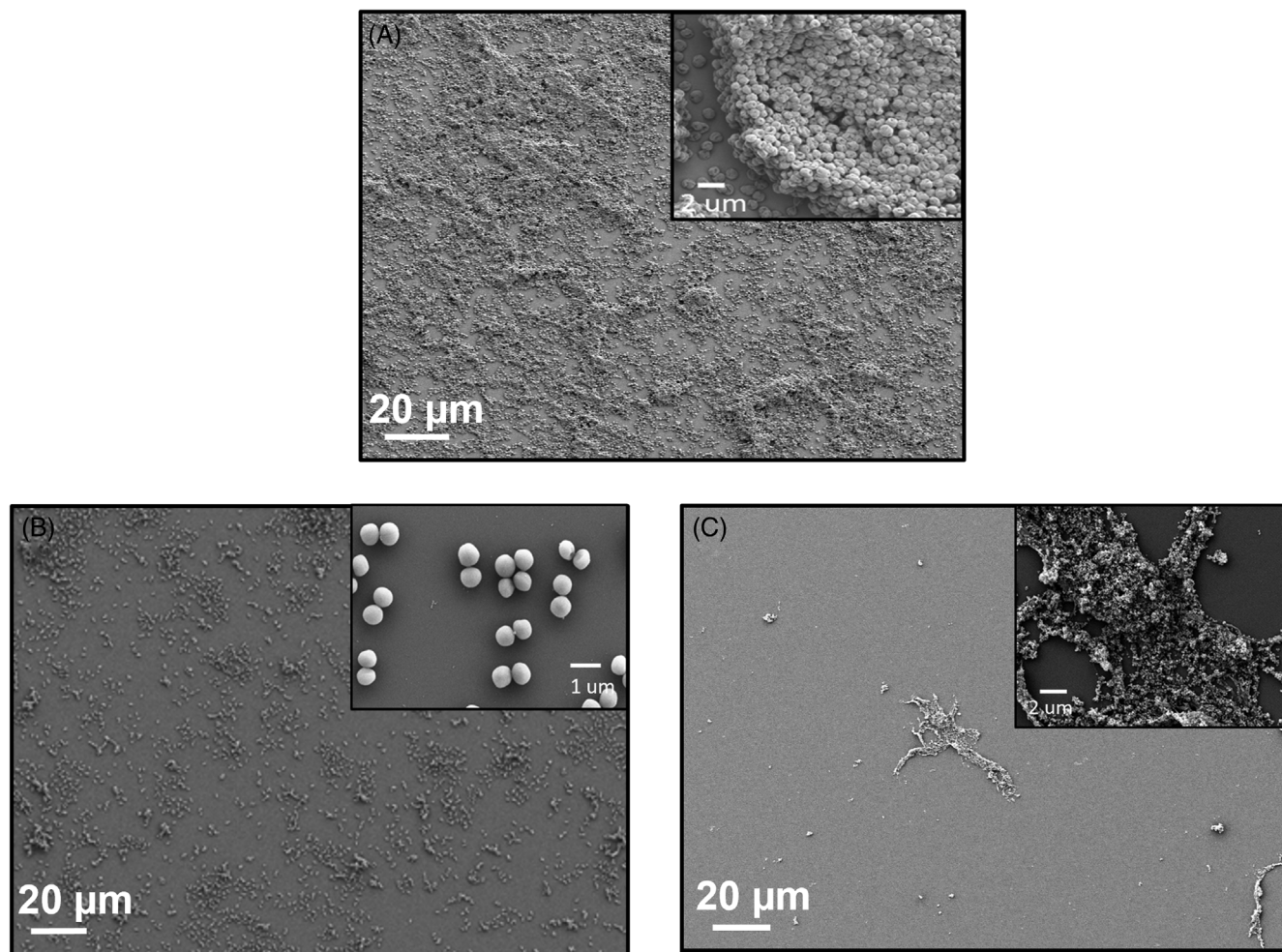


FIGURE 8 SEM micrograph of (A) untreated *S. aureus* biofilm, (B) F-MSN-PEI/Cur treated *S. aureus* bacteria during biofilm formation (C) F-MSN-PEI/Cur treated matured *S. aureus* biofilm.

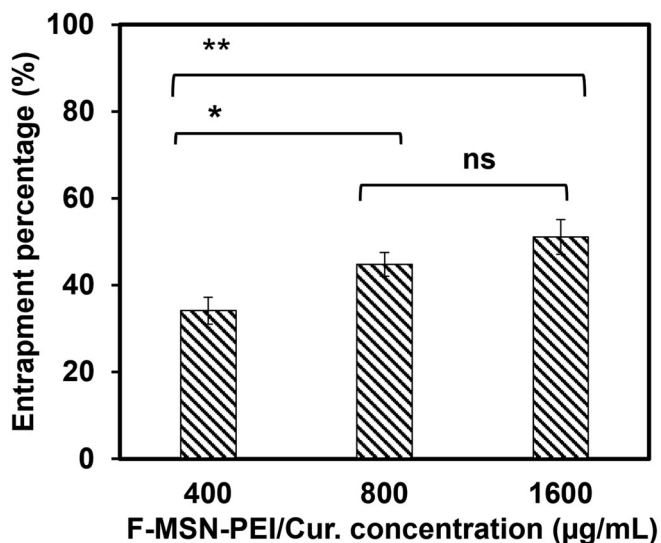


FIGURE 9 Entrapment efficiency of the F-MSN-PEI/Cur formulation into *S. aureus* biofilm. The results for entrapment efficiency were statistically analyzed in Graphpad Prism software (V 8.4.2) by ordinary one-way ANOVA followed by Tukey's multiple comparisons test. The degree of significance for both graphs were ns:0.1234, * $p < .03$, ** $p < .002$, *** $p < .0002$, **** $p < .0001$.

infections. The unique properties of NPs play an important role in inducing the antibacterial activity of antibiotics. Among the existing NPs, MSN provides a superior effect for cellular interactions. The use of MSN has been reported to be an effective antibiotic delivery system for antimicrobial agents. In the present study, the PEI surface grafted MSN as the carrier of herbal extract, curcumin, and the inhibition of planktonic and biofilm forms of *S. aureus* by F-MSN-PEI/Cur have been presented. F-MSN-PEI/Cur treatments both for the prevention of biofilm and eradication of matured biofilms was observed and affirmed by the reduction in biomass and cell viability in the biofilm matrix, which ultimately means the destruction of the biofilm-forming capability of the cells and the biofilm microenvironments. These results could be derived from the increased local bioavailability of curcumin by the entrapment of the F-MSN-PEI/Cur within the biofilm matrix which is due to the penetration capability of F-MSN-PEI carrier through the matured biofilm matrix. The main mechanism of action in inhibiting *S. aureus* growth by curcumin was shown to be perturbation of bacterial membrane integrity.¹³ In literature studies, it was demonstrated that damaged bacterial membrane is correlated to impaired cell metabolism and cell death.⁴⁷ Cell membrane damage in *S. aureus* after exposure to curcumin was also corroborated with electron microscopy images revealing distorted cell shape and cell debris. In addition, it was shown that curcumin induces damage in bacterial DNA as well by causing DNA fragmentation which is an indicative of apoptotic process.⁴⁸ Even though the modes of curcumin-mediated *S. aureus* cell killing were mainly investigated in planktonic form of bacteria in literature studies, the same mechanisms are possibly responsible for the anti-biofilm action of curcumin. Increasing the bioavailability of curcumin by loading the curcumin on F-MSN-PEI carrier

is therefore of great concern to obtain the desired anti-biofilm effects since higher doses of the active ingredients is required against biofilms compared to planktonic bacteria.

In addition, the F-MSN-PEI carrier could aid in improving the biocompatibility of curcumin as a bactericidal agent. It was revealed that curcumin show distinct toxicity that is concentration dependent.^{49,50} The incorporation of curcumin into F-MSN-PEI carriers did not cause any cytotoxicity and pungency, which could be due to both the combinatory effect of silica nanocarriers and improved solubility in aqueous physiological environments. Our results showed that curcumin loaded F-MSN-PEI supports fibroblast cell viability significantly during 24 h incubation. In the literature, different forms of silica have effect on cell behavior and the obtained results indicated that positively charged silica particles are taken up rapidly by cells and showed intracellular silicic acid release which resulted in positive effect on cell proliferation and migration.^{51,52} Moreover, curcumin is known as highly effective for wound healing process. For instance, Dai et al. reported that human fibroblast HS-27 cells incubated with cell culture media mixed extract from curcumin incorporated nanofibrous mats led to more proliferation than without curcumin containing mats and provide increased fibroblast migration hence, a higher wound closure rate inclined in vitro. In parallel, Kang et al. indicated that curcumin promoted the fibroblast proliferation until 12.5 µM/L.⁵⁰ Similarly, in vivo wound healing results demonstrated that curcumin containing mats cause faster wound closure at 7 and 15 days⁵³ and another in vivo study proved that curcumin showed thicker granulation tissue and greater re-epithelization at 7 and 12 days of post operation.⁵⁴ Additionally, the outer surface modification component of F-MSN-PEI design, PEI is well known for its cytotoxicity. Despite the toxicity consideration PEI functionalization is utilized commonly especially for gene delivery due to capability to employ reactive groups, increase stability, tune surface charge. It was indicated that PEI toxicity could be attributed to be molecular weight dependent.⁵⁵ However, Desai and coworkers demonstrated that PEI adsorbed silica NPs in different forms showed less toxicity than PEI alone even at high concentrations. Also, toxicity did not correspond to the particles concentrations but PEI/SiO₂ ratio. The study proved that incorporation of silica and PEI could aid to suppress the toxicity of PEI.⁵⁶ All in all, the in vitro cytotoxicity investigations did not lead reduction in cell viability especially for 24 h incubation which is also matching with the treatment period in the bactericidal investigations.

F-MSN-PEI drug carrier could aid in improving the bactericidal effect of curcumin. Due to the spatial confinement of drug molecules within the F-MSN-PEI mesoporous network, drugs with poor solubility can be transformed into their amorphous/disordered crystalline counterparts with higher solubility.⁵⁷ Also, it was shown that amine-grafted mesoporous silica employed a prolonged and adjustable release profile of curcumin.⁵⁸ In addition, the surface chemistry of the NPs has been shown to play a prominent role in altering the interaction between NPs and the biofilm matrix. This fact eventually determines the penetration ability, depth, and distribution of the NPs through the biofilm matrix.⁵⁹ The complex nature of the biofilm matrix consists of different macromolecules with varying charges and

hydrophobicity. Recently, researchers have shown that the penetration and localization of NPs through the biofilm matrix are highly dominated by the net surface charge of silica nanoparticles (SiNPs).⁶⁰ Among the investigated surface properties of SiNPs, positively charged SiNPs resulted in the highest degree of entrapment, and hence the accumulation, by the biofilm matrix. Another consideration for NPs entrapment through biofilms is to determine which component (protein, eDNA, or polysaccharides) comprises the majority of the biofilm EPS matrix which has a direct effect on the affinity of the NPs. Studies investigating the relationship between the type of surface functionalization and different biofilm samples show that the entrapment degree of MSN within biofilm matrix can be tuned by the altered surface chemistry on mesoporous silica matrix.⁶¹ In our study, we showed that the polyionic nature of PEI on MSN carriers together with the loading of curcumin from the herbal extract (design-MSN-PEI/Cur) could provide a superior effect for inhibiting biofilm formation compared to free curcumin. This should be related to the polyionic nature of the PEI that could destruct the integrity of the bacterial cell membrane before bacterial cells reach the biofilm formation stages involving attachment of cells to a surface followed by the assembly of the cells to form micro colonies, and the differentiation of biofilm into a mature structure.⁶² In the literature, researchers have revealed that hydrophobic surfaces are more susceptible to bacterial cell adhesion than hydrophilic surfaces. Hence, the hydrophilic nature of F-MSN-PEI could inhibit the bacterial cell adhesion.⁶³ In addition, the porous architecture of the silica matrix has been revealed to prevent the bacterial cell aggregations that take place as the initial step of biofilm formation, as also claimed in the literature studies.⁶⁴ On the other hand, free curcumin treatment of bacterial cells during biofilm formation steps has induced the biomass formation compared to control groups, which could be due to the secretion of different components and could cause relapsing of the biofilm formation in the long term. When the short-term treatment of *S. aureus* cells is taken into account, the viability of bacterial cells could already be reduced by 20% with 400 µg/ml F-MSN-PEI/Cur treatment compared to the negative control group, which could be depicted as that changes in the metabolic activity of bacterial cells could inhibit the biofilm formation potency of treated bacterial cells.

As presented in Table 3, F-MSN-PEI/Cur could play a role in inhibiting the biofilm-forming capacity of the bacterial cells by switching them to non-biofilm-forming bacterial cells. However, there was only an acceptable level of statistical difference in the reduction of biofilm viability in the matured biofilms in Figure 7B, whereas the statistical differences were significantly higher in Figure 6B when the F-MSN-PEI/Cur and equivalent free curcumin treatments were compared. This could be related to the slow degradation of curcumin during the incubation time and/or stacking depth of F-MSN-PEI through *S. aureus* biofilm matrix. In our previous laboratory investigations as a part of another study, confocal imaging was performed for the F-MSN-PEI treated biofilms, and the plot of fluorescent intensity of F-MSN-PEI (count) versus depth of *S. aureus* biofilm (µm) after 24 h was plotted. The obtained analysis results of fluorescence count for 4, 8, and 14 µm depths of the biofilms revealed that F-MSN-PEI could

penetrate through the deepest level of the biofilm matrix. However, the highest intensity, which could be depicted as the highest accumulation amount of F-MSN-PEI, could be achieved between 3 and 6 µm depths of the biofilm matrix. The accumulation of F-MSN-PEI/Cur in the middle layer could be due to the hydrophilic component that could overcome the hydrophobicity of the *S. aureus* cell membrane.⁶⁵ This could be the reason for the higher accumulation of the F-MSN-PEI with the hydrophilic nature in the encapsulating layer of bacterial cells and could destroy the integrity of the EPS matrix, which leads mainly to a decrease in total biomass compared to the untreated preformed biofilms. Taken together, we showed that the antimicrobial and antibiofilm activities of curcumin can be enhanced by loading curcumin onto F-MSN-PEI nanocarriers. The F-MSN-PEI matrix as a drug carrier provides an opportunity for revolutionizing the bactericidal activity of curcumin, the common source of turmeric, for eradication of mature biofilms and prevention of biofilm formation without any combinatory antibiotic treatment.

5 | CONCLUSION

The successful development of curcumin loaded PEI grafted mesoporous silica NPs nanocomposite, denoted F-MSN-PEI/Cur proves both antibacterial and antibiofilm activity. Short-term treatments of *S. aureus* cells in planktonic form with F-MSN-PEI/Cur suspensions could provide a significant reduction in the cell viability even at low dosing (25 µg/ml) compared to free curcumin treatments, and the highest concentration of 400 µg/ml treatments could reduce the cell viability down to 20% compared to untreated control. These facts inhibit biofilm formation by reducing the viability of bacterial cells and total biomass in the biofilm matrix. Furthermore, increasing F-MSN-PEI/Cur dosing could prevent biofilm formation and eradicate matured biofilms compared to free curcumin treatments. Therefore, it is evident that PEI grafted mesoporous silica nanocarriers have great potency for repurposing of herbal extracts (i.e., curcumin) as antibiofilm agents against biofilms without any concomitant antibiotic treatment.

ACKNOWLEDGMENTS

The Turkish Scientific and Technological Research Council (project #319S024) is acknowledged for funding support. Financial support from The Turkish Scientific and Technological Research Council 2211-A BİDEB doctoral scholarship (Ayşenur Pamukçu) and the Council of Higher Education for 100/2000 doctoral scholarship (Ayşenur Pamukçu, Nursu Erdoğan) are gratefully acknowledged.

DATA AVAILABILITY STATEMENT

Data available on request from the authors. The data that support the findings of this study are available from the corresponding author upon reasonable request.

ORCID

Ayşenur Pamukçu  <https://orcid.org/0000-0002-7650-4147>

REFERENCES

- Roberts AEL, Kragh KN, Bjarnsholt T, Diggle SP. The limitations of in vitro experimentation in understanding biofilms and chronic infection. *J Mol Biol*. 2015;427:3646-3661. doi:10.1016/j.jmb.2015.09.002
- Høiby N, Bjarnsholt T, Moser C, et al. ESCMID* guideline for the diagnosis and treatment of biofilm infections. *Clin Microbiol Infect*. 2014;21(2015):S1-S25. doi:10.1016/j.cmi.2014.10.024
- Liu Y, Li Y, Shi L. Controlled drug delivery systems in eradicating bacterial biofilm-associated infections. *J Control Release*. 2021;329:1102-1116. doi:10.1016/j.jconrel.2020.10.038
- Olson ME, Ceri H, Morck DW, Buret AG, Read RR. Biofilm bacteria: formation and comparative susceptibility to antibiotics. *Can J Vet Res Rev Can Rech Veterinaire*. 2002;66:86-92.
- Şen Karaman D, Manner S, Rosenholm JM. Mesoporous silica nanoparticles as diagnostic and therapeutic tools: how can they combat bacterial infection? *Ther Deliv*. 2018;9:241-244. doi:10.4155/tde-2017-0111
- Benoit DS, Koo H. Targeted, triggered drug delivery to tumor and biofilm microenvironments. *Nanomedicine*. 2016;11:873-879. doi:10.2217/nnm-2016-0014
- Bjarnsholt T, Whiteley M, Rumbaugh KP, Stewart PS, Jensen PØ, Frimodt-Møller N. The importance of understanding the infectious microenvironment. *Lancet Infect Dis*. 2021;22:e88-e92. doi:10.1016/S1473-3099(21)00122-5
- Gupta A, Landis RF, Rotello V. Nanoparticle-based antimicrobials: surface functionality is critical. *F1000Research*. 2016;5:364. doi:10.12688/f1000research.7595.1
- Feghali P, Ibrahim R, Nawas T. Antibacterial activity of curcumin against lebanese clinical isolates of *Staphylococcus aureus*. *MOJ Toxicol*. 2018;4:81-83. doi:10.15406/mojt.2018.04.00094
- Kumbar VM, Peram MR, Kugaji MS, et al. Effect of curcumin on growth, biofilm formation and virulence factor gene expression of *Porphyromonas gingivalis*. *Odontology*. 2021;109:18-28. doi:10.1007/s10266-020-00514-y
- Moshe M, Lellouche J, Banin E. Curcumin: a natural antibiofilm agent. *Science and Technology Against Microbial Pathogens*. World Scientific; 2011:89-93. doi:10.1142/9789814354868_0017
- Zorofchian Moghadamtousi S, Abdul Kadir H, Hassandarvish P, Tajik H, Abubakar S, Zandi K. A review on antibacterial, antiviral, and antifungal activity of curcumin. *Biomed Res Int*. 2014;2014:1-12. doi:10.1155/2014/186864
- Teow S-Y, Liew K, Ali SA, Khoo AS-B, Peh S-C. Antibacterial action of curcumin against *Staphylococcus aureus*: a brief review. *J Trop Med*. 2016;2016:1-10. doi:10.1155/2016/2853045
- Gao M, Long X, Du J, et al. Enhanced curcumin solubility and antibacterial activity by encapsulation in PLGA oily core nanocapsules. *Food Funct*. 2020;11:448-455. doi:10.1039/C9FO00901A
- Chen C, Sun W, Wang X, Wang Y, Wang P. Rational design of curcumin loaded multifunctional mesoporous silica nanoparticles to enhance the cytotoxicity for targeted and controlled drug release. *Mater Sci Eng C*. 2018;85:88-96. doi:10.1016/j.msec.2017.12.007
- Shaikh J, Ankola DD, Beniwal V, Singh D, Kumar MNVR. Nanoparticle encapsulation improves oral bioavailability of curcumin by at least 9-fold when compared to curcumin administered with piperine as absorption enhancer. *Eur J Pharm Sci*. 2009;37:223-230. doi:10.1016/j.ejps.2009.02.019
- John MK, Xie H, Bell EC, Liang D. Development and pharmacokinetic evaluation of a curcumin co-solvent formulation. *Anticancer Res*. 2013;33:4285-4291.
- Suresh K, Nangia A. Curcumin: pharmaceutical solids as a platform to improve solubility and bioavailability. *CrstEngComm*. 2018;20:3277-3296. doi:10.1039/C8CE00469B
- Kurien BT, Singh A, Matsumoto H, Scofield RH. Improving the solubility and pharmacological efficacy of curcumin by heat treatment. *Assay Drug Dev Technol*. 2007;5:567-576. doi:10.1089/adt.2007.064
- Tønnesen HH, Måsson M, Loftsson T. Studies of curcumin and curcuminoids. XXVII. Cyclodextrin complexation: solubility, chemical and photochemical stability. *Int J Pharm*. 2002;244:127-135. doi:10.1016/S0378-5173(02)00323-X
- Wang Y-J, Pan M-H, Cheng A-L, et al. Stability of curcumin in buffer solutions and characterization of its degradation products. *J Pharm Biomed Anal*. 1997;15:1867-1876. doi:10.1016/S0731-7085(96)02024-9
- Esatbeyoglu T, Huebbe P, Ernst IMA, Chin D, Wagner AE, Rimbach G. Curcumin-from molecule to biological function. *Angew Chem Int Ed*. 2012;51:5308-5332. doi:10.1002/anie.201107724
- Tabanelli R, Brogi S, Calderone V. Improving curcumin bioavailability: current strategies and future perspectives. *Pharmaceutics*. 2021;13:1715. doi:10.3390/pharmaceutics13101715
- Naksuriya O, Okonogi S, Schiffeleers RM, Hennink WE. Curcumin nanoformulations: a review of pharmaceutical properties and preclinical studies and clinical data related to cancer treatment. *Biomaterials*. 2014;35:3365-3383. doi:10.1016/j.biomaterials.2013.12.090
- Patra D, Şen Karaman D, Desai D, El Khoury E, Rosenholm JM. Preparation of curcumin loaded mesoporous silica nanoparticles: determining polarizability inside the mesopores. *Mater Res Bull*. 2016;84:267-272. doi:10.1016/j.materresbull.2016.08.012
- Özliseli E, Şen Karaman D, Chakraborti S, et al. Rational evaluation of human serum albumin coated mesoporous silica nanoparticles for xenogenic-free stem cell therapies. *Colloids Surf A Physicochem Eng Asp*. 2020;600:124945. doi:10.1016/j.colsurfa.2020.124945
- Rosenholm JM, Penninkangas A, Lindén M. Amino-functionalization of large-pore mesoscopically ordered silica by a one-step hyperbranching polymerization of a surface-grown polyethyleneimine. *Chem Commun*. 2006; 37:3909-3911. doi:10.1039/B607886A
- Martín A, García RA, Karaman DS, Rosenholm JM. Polyethyleneimine-functionalized large pore ordered silica materials for poorly water-soluble drug delivery. *J Mater Sci*. 2014;49:1437-1447. doi:10.1007/s10853-013-7828-1
- Yadav YC, Pattnaik S, Swain K. Curcumin loaded mesoporous silica nanoparticles: assessment of bioavailability and cardioprotective effect. *Drug Dev Ind Pharm*. 2019;45:1889-1895. doi:10.1080/03639045.2019.1672717
- Gong C, Wu Q, Wang Y, et al. A biodegradable hydrogel system containing curcumin encapsulated in micelles for cutaneous wound healing. *Biomaterials*. 2013;34:6377-6387. doi:10.1016/j.biomaterials.2013.05.005
- Fu S-Z, Meng X-H, Fan J, et al. Acceleration of dermal wound healing by using electrospun curcumin-loaded poly(ϵ -caprolactone)-poly(ethylene glycol)-poly(ϵ -caprolactone) fibrous mats: acceleration of dermal wound healing. *J Biomed Mater Res B Appl Biomater*. 2014;102:533-542. doi:10.1002/jbm.b.33032
- Lungare S, Hallam K, Badhan RKS. Phytochemical-loaded mesoporous silica nanoparticles for nose-to-brain olfactory drug delivery. *Int J Pharm*. 2016;513:280-293. doi:10.1016/j.ijpharm.2016.09.042
- Sareen S, Joseph L, Mathew G. Improvement in solubility of poor water-soluble drugs by solid dispersion. *Int J Pharm Investig*. 2012;2:12-17. doi:10.4103/2230-973X.96921
- Shin GH, Li J, Cho JH, Kim JT, Park HJ. Enhancement of curcumin solubility by phase change from crystalline to amorphous in cur-TPGS nanosuspension: enhancement of curcumin solubility. *J Food Sci*. 2016;81:N494-N501. doi:10.1111/1750-3841.13208
- Ghaseminezhad SM, Shojaosadati SA, Meyer RL. Ag/Fe₃O₄ nanocomposites penetrate and eradicate *S. aureus* biofilm in an in vitro chronic wound model. *Colloids Surf B Biointerfaces*. 2018;163:192-200. doi:10.1016/j.colsurfb.2017.12.035
- Dempster C, Banat I, Mrrchant C. Antimicrobial and antibiofilm potential of biosurfactants as novel combination therapy against bacterium that cause skin infections. Microbiology Society Annual Conference. Access Microbiology 2019. doi:10.1099/acmi.ac2019.po0566

37. Stiefel P, Rosenberg U, Schneider J, Mauerhofer S, Maniura-Weber K, Ren Q. Is biofilm removal properly assessed? Comparison of different quantification methods in a 96-well plate system. *Appl Microbiol Biotechnol*. 2016;100:4135-4145. doi:10.1007/s00253-016-7396-9
38. Azeredo J, Azevedo NF, Briandet R, et al. Critical review on biofilm methods. *Crit Rev Microbiol*. 2017;43:313-351. doi:10.1080/1040841X.2016.1208146
39. Hassan A, Usman J, Kaleem F, Omaid M, Khalid A, Iqbal M. Evaluation of different detection methods of biofilm formation in the clinical isolates. *Braz J Infect Dis*. 2011;15:305-311. doi:10.1016/S1413-8670(11)70197-0
40. Devlin H, Hiebner D, Barros C, et al. A high throughput method to investigate nanoparticle entrapment efficiencies in biofilms. *Colloids Surf B Biointerfaces*. 2020;193:111123. doi:10.1016/j.colsurfb.2020.111123
41. Chen LR. Dissolution behavior of a poorly water soluble compound in the presence of tween 80. *Pharm Res*. 2003;20:797-801. doi:10.1023/A:1023493821302
42. Ashfaq M, Shah S, Rasul A, et al. Enhancement of the solubility and bioavailability of pitavastatin through a self-nanoemulsifying drug delivery system (SNEDDS). *Pharmaceutics*. 2022;14:482. doi:10.3390/pharmaceutics14030482
43. Nair RS, Morris A, Billa N, Leong C-O. An evaluation of curcumin-encapsulated chitosan nanoparticles for transdermal delivery. *AAPS PharmSciTech*. 2019;20:69. doi:10.1208/s12249-018-1279-6
44. Gardikis K, Hatziantoniou S, Viras K, Demetzos C. Effect of a bioactive curcumin derivative on DPPC membrane: a DSC and Raman spectroscopy study. *Thermochim Acta*. 2006;447:1-4. doi:10.1016/j.tca.2006.03.007
45. Zhang JH, Chung TD, Oldenburg KR. A simple statistical parameter for use in evaluation and validation of high throughput screening assays. *J Biomol Screen*. 1999;4:67-73. doi:10.1177/108705719900400206
46. Conlon BP. *Staphylococcus aureus* chronic and relapsing infections: evidence of a role for persister cells: an investigation of persister cells, their formation and their role in *S. aureus* disease. *Bioessays*. 2014;36:991-996. doi:10.1002/bies.201400080
47. Tyagi P, Singh M, Kumari H, Kumari A, Mukhopadhyay K. Bactericidal activity of curcumin I is associated with damaging of bacterial membrane. *PLoS One*. 2015;10:e0121313. doi:10.1371/journal.pone.0121313
48. de Andrade Neto JB, de Farias Cabral VP, Nogueira LFB, et al. Anti-MRSA activity of curcumin in planktonic cells and biofilms and determination of possible action mechanisms. *Microb Pathog*. 2021;155:104892. doi:10.1016/j.micpath.2021.104892
49. Scharstuhl A, Mutsaers HAM, Pennings SWC, Szarek WA, Russel FGM, Wagener FADTG. Curcumin-induced fibroblast apoptosis and in vitro wound contraction are regulated by antioxidants and heme oxygenase: implications for scar formation. *J Cell Mol Med*. 2009;13:712-725. doi:10.1111/j.1582-4934.2008.00339.x
50. Kang J, Huang H, Zhu F. Effect of curcumin on growth and function of fibroblast in human hyperplastic scar. *J Integr Tradit West Med*. 2009;29:1100-1103.
51. Quignard S, Coradin T, Powell JJ, Jugdaohsingh R. Silica nanoparticles as sources of silicic acid favoring wound healing in vitro. *Colloids Surf B Biointerfaces*. 2017;155:530-537. doi:10.1016/j.colsurfb.2017.04.049
52. Akbik D, Ghadiri M, Chrzanowski W, Rohanizadeh R. Curcumin as a wound healing agent. *Life Sci*. 2014;116:1-7. doi:10.1016/j.lfs.2014.08.016
53. Dai X, Liu J, Zheng H, et al. Nano-formulated curcumin accelerates acute wound healing through Dkk-1-mediated fibroblast mobilization and MCP-1-mediated anti-inflammation. *NPG Asia Mater*. 2017;9:e368-e368. doi:10.1038/am.2017.31
54. Yen Y-H, Pu C-M, Liu C-W, et al. Curcumin accelerates cutaneous wound healing via multiple biological actions: the involvement of TNF- α , MMP-9, α -SMA, and collagen. *Int Wound J*. 2018;15:605-617. doi:10.1111/iwj.12904
55. Xia T, Kovochich M, Liong M, et al. Polyethyleneimine coating enhances the cellular uptake of mesoporous silica nanoparticles and allows safe delivery of siRNA and DNA constructs. *ACS Nano*. 2009;3:3273-3286. doi:10.1021/nn900918w
56. Desai D, Karaman DS, Prabhakar N, et al. Design considerations for mesoporous silica nanoparticulate systems in facilitating biomedical applications. *Open Mater Sci*. 2014;1:16-43. doi:10.2478/mesbi-2014-0001
57. Salonen J, Laitinen L, Kaukonen AM, et al. Mesoporous silicon microparticles for oral drug delivery: loading and release of five model drugs. *J Control Release*. 2005;108:362-374. doi:10.1016/j.jconrel.2005.08.017
58. Hartono SB, Hadisoewignyo L, Yang Y, Meka AK, Antaresti C. Amine functionalized cubic mesoporous silica nanoparticles as an oral delivery system for curcumin bioavailability enhancement. *Nanotechnol*. 2016;27:505605. doi:10.1088/0957-4484/27/50/505605
59. Li X, Yeh Y-C, Giri K, et al. Control of nanoparticle penetration into biofilms through surface design. *Chem Commun*. 2014;51:282-285. doi:10.1039/C4CC07737G
60. Hiebner DW, Barros C, Quinn L, Vitale S, Casey E. Surface functionalization-dependent localization and affinity of SiO₂ nanoparticles within the biofilm EPS matrix. *Biofilm*. 2020;2:100029. doi:10.1016/j.biofilm.2020.100029
61. Fulaz S, Devlin H, Vitale S, Quinn L, O'Gara JP, Casey E. Tailoring nanoparticle-biofilm interactions to increase the efficacy of antimicrobial agents against *Staphylococcus aureus*. *Int J Nanomedicine*. 2020;15:4779-4791. doi:10.2147/IJN.S256227
62. Roy R, Tiwari M, Donelli G, Tiwari V. Strategies for combating bacterial biofilms: a focus on anti-biofilm agents and their mechanisms of action. *Virulence*. 2018;9:522-554. doi:10.1080/21505594.2017.1313372
63. Doyle RJ. Contribution of the hydrophobic effect to microbial infection. *Microbes Infect*. 2000;2:391-400. doi:10.1016/S1286-4579(00)00328-2
64. Pezzoni M, Catalano PN, Pizarro RA, et al. Antibiofilm effect of supramolecularly templated mesoporous silica coatings. *Mater Sci Eng C*. 2017;77:1044-1049. doi:10.1016/j.msec.2017.04.022
65. Reifsteck F, Wee, B.J.Y. S. Wilkinson, hydrophobicity—hydrophilicity of staphylococci. *J Med Microbiol*. 1987;24(1987):65-73. doi:10.1099/00222615-24-1-65

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Pamukçu A, Erdoğan N, Şen Karaman D. Polyethylenimine-grafted mesoporous silica nanocarriers markedly enhance the bactericidal effect of curcumin against *Staphylococcus aureus* biofilm. *J Biomed Mater Res*. 2022;110(11):2506-2520. doi:10.1002/jbm.b.35108