

Exosomes Derived from Hypoxic Glioma Cells Reduce the Sensitivity of Glioma Cells to Temozolomide Through Carrying miR-106a-5p

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Background: Hypoxia is a frequent feature of solid tumors which significantly affects the efficacy of treatments such as chemotherapy. In addition, exosomes from hypoxic cancer cells could contribute to the chemoresistance of tumor cells through carrying miRNAs. It has been shown that miR-106-5p level was upregulated in glioma. However, whether exosomes derived from hypoxic glioma cells could affect temozolomide (TMZ) resistance in glioma through carrying miR-106a-5p remains unexplored.

Methods: Exosomes were isolated from glioma cells under normoxia or hypoxia condition. EdU staining and flow cytometry assays were used to assess the cell proliferation and cell apoptosis. The relation between miR-106a-5p and PTEN was investigated by dual luciferase assay.

Results: MiR-106a-5p was enriched in exosomes derived from hypoxic glioma cells compared to exosomes from cells under normoxia condition. Additionally, hypoxic glioma cells were able to transfer exosomes to glioma cells, resulting in a significant increase of miR-106a-5p level in cells. TMZ remarkably suppressed glioma cell proliferation and triggered cell apoptosis. However, hypoxic glioma cell-derived exosomes markedly promoted the proliferation and suppressed the apoptosis in TMZ-treated glioma cells, and miR-106a-5p inhibitor was able to abolish these phenomena. Meanwhile, PTEN was verified to be a direct target of miR-106a-5p. Furthermore, TMZ elevated PTEN and Bax level and reduced p-Akt level in glioma cells, whereas these changes were reversed by hypoxia glioma cell-derived exosomes. Furthermore, hypoxia glioma cell-derived exosomes reduced the sensitivity of glioma cells to TMZ in vivo via downregulating PTEN.

Conclusion: Collectively, exosomal miR-106a-5p derived from hypoxia glioma cells could reduce the sensitivity of glioma cells to TMZ through downregulating PTEN. Thus, our study might provide new strategies for improving the clinical efficacy of TMZ on glioma.

Keywords: cancer, hypoxia, miR-106a-5p, chemosensitivity

Introduction

Glioma is a primary malignant tumor in brains, which occurs with a median survival period. Most patients with glioma die in 2 years of diagnosis.^{1,2} The major therapies for glioma include TMZ chemotherapy and radiation, and the combination of TMZ and radiation are more successful to radiation alone.^{3,4} TMZ often promotes apoptosis by inducing the DNA damage.^{5,6} The prognosis of many patients is not ideal as the glioma cells could acquire chemoresistance.^{7,8} It has been revealed that low oxygen tension (hypoxia) established in growing tumor masses could induce the chemoresistance of tumor cells.^{9,10} The hypoxic microenvironment can induce chemoresistance and decrease the chemosensitivity during the tumor progression.^{10–12} Thus, it is essential to study the mechanisms of chemo-responsiveness in glioma cells for discovering novel therapeutic methods.

MicroRNAs (miRNAs) have been found to be associated with chemoresistance in multiple types of cancers, including glioma.^{13–15} Li et al reported that miR-519a overexpression sensitized glioblastoma cells to TMZ treatment through targeting STAT3/Bcl2 signaling.¹⁴ MiR-182b could reduce chemoresistance to TMZ in glioma stem cells.¹⁶ Previous

study revealed that miR-106a-5p was upregulated in glioma.¹⁷ Additionally, miR-106a-5p could facilitate 5-Fluorouracil resistance in colorectal cancer by targeting TGF β R2.¹⁸ However, the relationship between miR-106a-5p and TMZ resistance in glioma cells remain largely unclear.

Exosomes could participate in occurrence of glioma, which could induce the resistance during chemotherapy.^{19,20} It has been revealed that exosomal miRNAs dysregulation could affect the chemoresistance and chemosensitivity in cancers.^{21,22} Studies showed that exosomes derived from hypoxic tumor cells could contribute to TMZ resistance of tumor cells by transferring miRNAs.^{20,23} However, whether exosomes derived from hypoxic glioma cells could affect the response of glioma cells to TMZ through carrying miR-106a-5p remains unclear.

Based on the above backgrounds, we aimed to investigate the function of hypoxic exosomes from glioma cells in TMZ resistance during the progression of glioma. The study would supply a theoretical basis for discovering novel methods against glioma.

Materials and Methods

Cell Culture and Treatment

Glioma cells (U87MG, SHG44 and U251MG) and human brain glial cells (HEB) were bought from Shanghai cell bank of Chinese Academy of Sciences (Shanghai, China). Cells were cultured in DMEM medium (Thermo Fischer Scientific) with 10% fetal bovine serum (FBS), 1% penicillin and streptomycin at 37°C, 5% CO₂.

Hypoxia Assay

SHG44 cells (5 x 10³ cells for 96-well plate; 2x10⁵ cells for 6-well plate) were plated and cultured for 24 h in the condition of 37°C and 5% CO₂. Subsequently, cells were transferred into hypoxic chamber (Stemcell™ Technologies) with 1% O₂ for 72 h.

Cell Transfection

SHG44 cells were transfected with mimics-ctrl (Invitrogen), miR-106a-5p mimics (Invitrogen) or miR-106a-5p inhibitor (Invitrogen) for 48 h by using lipofectamine 3000 (Invitrogen) in line with the protocol of manufacturer.

Exosome Extraction and Identification

Briefly, cell growth medium was replaced with medium (without FBS) when the confluence reached 80%. Next, the supernatant was centrifuged (10 min at 300 × g, 15 min at 2000 × g and 30 min at 10,000 × g). Ultracentrifugation (70 min at 120,000 × g twice) was then applied to collect the supernatant for exosome isolation, and transmission electron microscopy (TEM), nanoparticle tracking analysis (NTA) and Western blot were performed to identify the relevant exosomes.²⁴ For TEM analysis, the morphology of exosomes was examined at 80 kV under a TEM (HT7700, HITACHI, Japan). For NTA, the size of exosomes was determined with a ZetaView analyzer (Particle Metrix, Germany).

Exosome Labeling and Uptake

SHG44 cells (2 x 10⁵ cells) were plated onto a 6 well plate overnight. Next, PKH26-labeled exosomes (50 μg/mL) were co-incubated with SHG44 cells for 24 h before counterstaining with fluorescent phalloidin dye, which stains the cytoskeleton. Images were then captured using a confocal laser scanning microscope (ZEISS LSM880, Germany). Magnification, x600. Cell nuclei was stained with DAPI.

CCK-8 Assay

SHG44 cells (5 x 10³ cells) were seeded onto a 96-well plate overnight. After that, cells were exposed to TMZ (25 μM),^{25,26} TMZ + normoxic glioma cell-derived exosomes (N-exo, 50 μg/mL), TMZ + hypoxic glioma cell-derived exosomes (H-exo, 50 μg/mL) or TMZ + H-exo + miR-106a-5p inhibitor (inhibitor) for 48 h and then evaluated using CCK-8 (10 μL). The OD value was measured using a microplate reader (Thermo MULTISKAN MK3, Thermo Fischer Scientific, USA) at 450 nm.

Immunofluorescence

SHG44 cells (2×10^5 cells) were seeded onto a 6 well plate overnight. Next, the EdU-positive cells were evaluated by the Cell-Light EdU Apollo567 In Vitro Kit (RIBOBIO). Images were then visualized under a fluorescence microscope (OLYMPUS IX51, Japan). Magnification, x200.

Apoptosis

SHG44 cells (2×10^5 cells) were seeded onto a 6-well plate overnight. Next, cells were centrifuged at $956 \times g$ for 5 min and resuspended in 500 μ L binding buffer. After that, cells were treated with Annexin V-FITC and PI (5 μ L) and then subjected to a flow cytometer (BD AriaIII, BD Biosciences, USA). FlowJo software was applied for data quantification and analysis.

RT-qPCR

Total RNA was extracted using TRIzol[®] and reverse transcribed to cDNA using a PrimeScript kit. RT-qPCR was applied using a SYBR green (ELK Biotechnology) reaction mix, under the conditions: 2 min at 94°C, followed by 35 cycles (94°C for 30s and 55°C for 45s). The primers used were as follows: miR-106a-5p forward, 5'-TGGGTGCTTACAGTGCAGGTAG-3' and reverse, 5'-CTCAACTGGTGTCTGGAGTC-3'; U6 forward, 5'-CTCGCTTCGGCAGCACAT-3' and reverse 5'-AACGCTTCACGAATTTGCGT-3'; PTEN forward, 5'-ATTCCCAGTCAGAGGCGCTAT-3' and reverse, 5'-GAACTTGTCTTCCCCTCGTGT-3'; HIF-1 α forward, 5'-CAGAATGAAGTGTACCCTAACTAGCC-3' and reverse, 5'-ACAAATCAGCACCAAGCAGGT-3'; β -actin forward, 5'-GTCCACC GCAAATGCTTCTA-3' and reverse, 5'-TGCTGTCACCTTCACCGTTC-3'. Relative quantification was completed using the $2^{-\Delta\Delta t}$ method and U6 and β -actin were used as the house keeping gene.

Western Blot

Total protein was obtained from each experimental sample using RIPA buffer and quantified using the BCA kit. Equal volumes of total protein (10 μ g/lane) were separated by SDS-PAGE (10%) and transferred onto PVDF membranes. These membranes were incubated with primary antibodies targeting CD63 (1:1000; Abcam), CD81 (1:1000; Abcam), PTEN (1:1000; Abcam), Akt (1:1000; Abcam), p-Akt (1:3000; Proteintech), p53 (1:1000; Abcam), Bax (1:1000; Abcam), calnexin (1:1000; Abcam) and β -actin (1:10,000; Abcam). After that, membranes were incubated with the relevant secondary antibodies (HRP-conjugated, 1:10,000; Abcam). Blots were visualized with ECL reagent and IPP 6.0 was used to complete the densitometry analysis.

Dual Luciferase Assay

The 3'-UTR of PTEN, which contains the miR-106a-5p binding site, were purchased from GenePharma before being cloned into the pGL6-miR vectors to establish pGL6-miR PTEN 3'-UTR wild-type (WT) or mutated (MT) plasmids. These plasmids were transfected into SHG44 cells together with miR-106a-5p mimics or mimics-ctrl using Lipofectamine 2000. Next, the luciferase activity was evaluated by the Dual Luciferase Reporter Assay System (Beyotime).

Animal Study

BALB/c female nude mice aged 6–8 weeks were housed in a room with a dedicated SPF facility. SHG44 cells (5×10^6 cells) were subcutaneously injected into the left flank of each mouse. When the tumor reached about 200 mm³, mice were divided into 3 groups (n = 4 per group) randomly: control, TMZ and TMZ + H-exo group. PBS or H-exo was injected intratumorally twice a week for 3 weeks. In addition, mice in the TMZ group were injected with 25 mg/kg TMZ intraperitoneally every day. The tumor volume was calculated once a week as per this equation: length \times width². Finally, the mice were sacrificed, and the tumors were weighed. All experiments were applied in accordance with the NIH Guide and Ethics Committee of The First Affiliated Hospital of Soochow University approved this research.

Immunohistochemical (IHC) Staining

The samples were deparaffinized, rehydrated, and heated in a microwave. Then, sections were washed, incubated in H₂O₂ (3%) for 25 min and washed with PBS. Afterwards, goat serum was applied to block and incubate samples for 30 min. Later on, a primary antibody (anti-PTEN) was used to stain the samples overnight. Then, the secondary antibody (HRP-labeled) was applied to stain the samples for 30 min. All antibodies were obtained from Abcam. The result was observed by a microscope (OLYMPUS CX31, Japan). Magnification, x200.

TUNEL Staining

Paraffin sections were washed and permeabilized. Subsequently, TUNEL mixtures (50 μ L) were used to incubate the sections for at 37°C with no light for 1.5 h. Next, cell nuclei were stained with DAPI. Finally, the result was observed by using a fluorescence microscope (Nikon Eclipse Ci-L, Japan). Magnification, x200.

Statistical Analysis

Mean \pm SD was applied to express all data. One-way analysis of variance (ANOVA) followed by Tukey's test or Student's *t*-test was used for analyzing the differences between experimental groups and $P < 0.05$ suggested a significant change.

Results

MiR-106a-5p Was Upregulated in Hypoxic Glioma Cells

To study the role of miR-106a-5p in glioma, RT-qPCR was performed. As indicated in Figure 1A, miR-106a-5p level was significantly higher in glioma cells, compared with HEB cells. In addition, miR-106a-5p level in SHG44 cells was much higher than that in other glioma cells (Figure 1A). Thus, SHG44 cells were selected of use in subsequent experiments. Meanwhile, the level of miR-106a-5p and HIF-1 α were significantly upregulated in glioma cells under hypoxic condition compared to cells under normoxia condition (Figure 1B and 1C). Taken together, miR-106a-5p was upregulated in hypoxic glioma cells.

MiR-106a-5p Could Be Transferred from Hypoxic Glioma Cells to Glioma Cells via Exosomes

Hypoxic cancer cell-derived exosomes are believed to play several crucial roles in chemoresistance during the tumor progression.^{27,28} Given this, we then isolated exosomes from normoxic (H-exo) or hypoxic (N-exo) glioma cells. Our

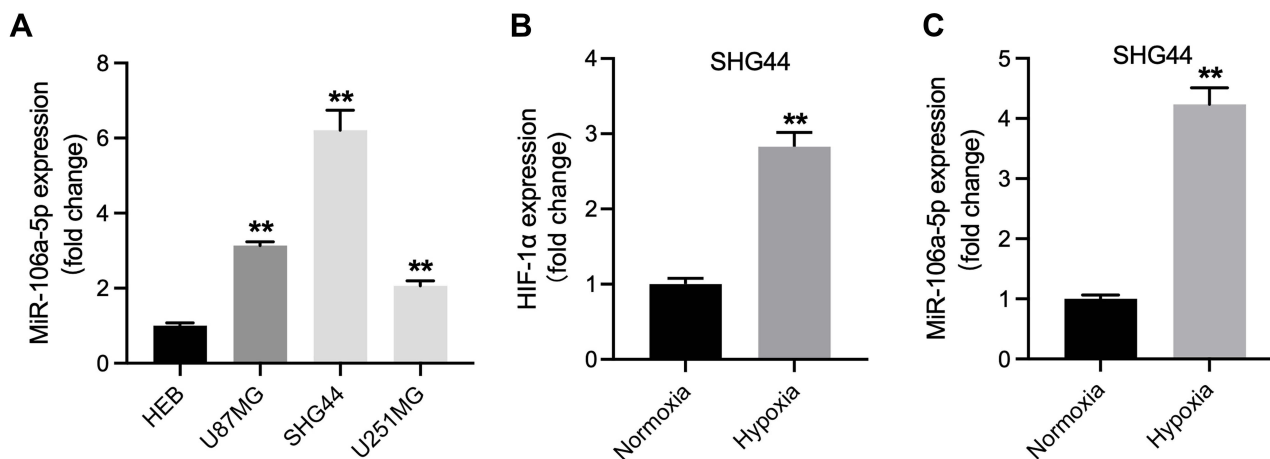


Figure 1 MiR-106a-5p was upregulated in hypoxic glioma cells. (A) The level of miR-106a-5p in HEB, U87MG, SHG44 and U251MG cells was detected by RT-qPCR. (B) The level of HIF-1 α in hypoxic or normoxia glioma cells was assessed by RT-qPCR. (C) The level of miR-106a-5p in hypoxic or normoxia glioma cells was assessed by RT-qPCR. ** $P < 0.01$ compared to HEB or normoxia.

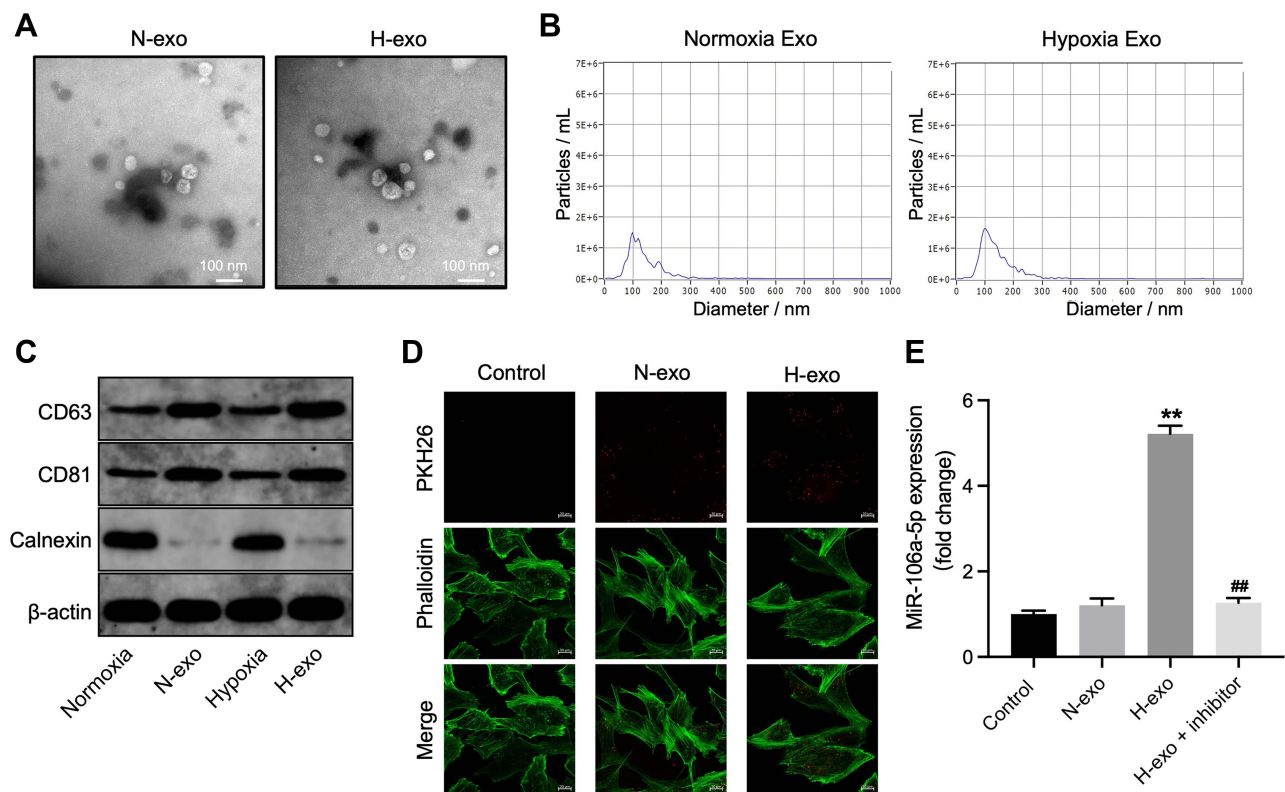


Figure 2 MiR-106a-5p could be transferred from hypoxic glioma cells to glioma cells via exosomes. (A) Exosomes were isolated from hypoxic or normoxia glioma cells (H-exo or N-exo), and were observed by TEM. (B) NTA was applied for exosome identification. (C) The expressions of CD81, CD63 and calnexin in normoxia cells, hypoxia cells, H-exo or N-exo were examined by Western blot. (D) The exosomes absorbed by glioma cells were observed by fluorescence staining. (E) Glioma cells were treated with N-exo, H-exo or H-exo + miR-106a-5p inhibitor. The level of miR-106a-5p in glioma cells was assessed by RT-qPCR. ** $P < 0.01$ compared to control. ### $P < 0.01$ compared to hypoxia Exo.

initial extraction data showed that these extracts were rounded particles (30–150 nm in diameter) (Figure 2B), and our subsequent evaluations revealed a high proportion of CD63 and CD81 expression in these extracts (Figure 2C). However, these extracts were negative for the endoplasmic reticulum calnexin protein (Figure 2C). These results showed that exosomes were isolated successfully.

Next, we investigated whether these exosomes can be transmitted into glioma cells by incubating glioma cells with PKH26-labeled H-exo or PKH26-labeled N-exo. After 24 h of incubation, PKH26 red dye was observed in glioma cells both in H-exo and N-exo groups (Figure 2D). In addition, compared to N-exo, H-exo sharply increased miR-106a-5p level in glioma cells, whereas that change was obviously reversed by miR-106a-5p inhibitor (Figure 2E). Collectively, miR-106a-5p was enriched in hypoxic exosomes derived from glioma cells and can be transferred from hypoxic glioma cells to glioma cells via exosomes.

Exosomes Derived from Hypoxic Glioma Cells Reduced TMZ Sensitivity in Glioma Cells Through Carrying miR-106a-5p

To detect the role of exosomal miR-106a-5p in TMZ sensitivity in glioma cells, CCK8, EdU staining and flow cytometry assays were performed. As shown in Figure 3A, TMZ significantly inhibited the viability of glioma cells, and exhibited about 50% growth inhibition. In addition, TMZ markedly reduced the proliferation and induced the apoptosis of glioma cells (Figure 3B and C). However, H-exo notably abolished the anti-tumor effect of TMZ in glioma cells, while these phenomena were reversed in the presence of miR-106a-5p inhibitor (Figure 3A–C). To sum up, hypoxic exosomes derived from glioma cells could reduce the sensitivity of glioma cells to TMZ through carrying miR-106a-5p.

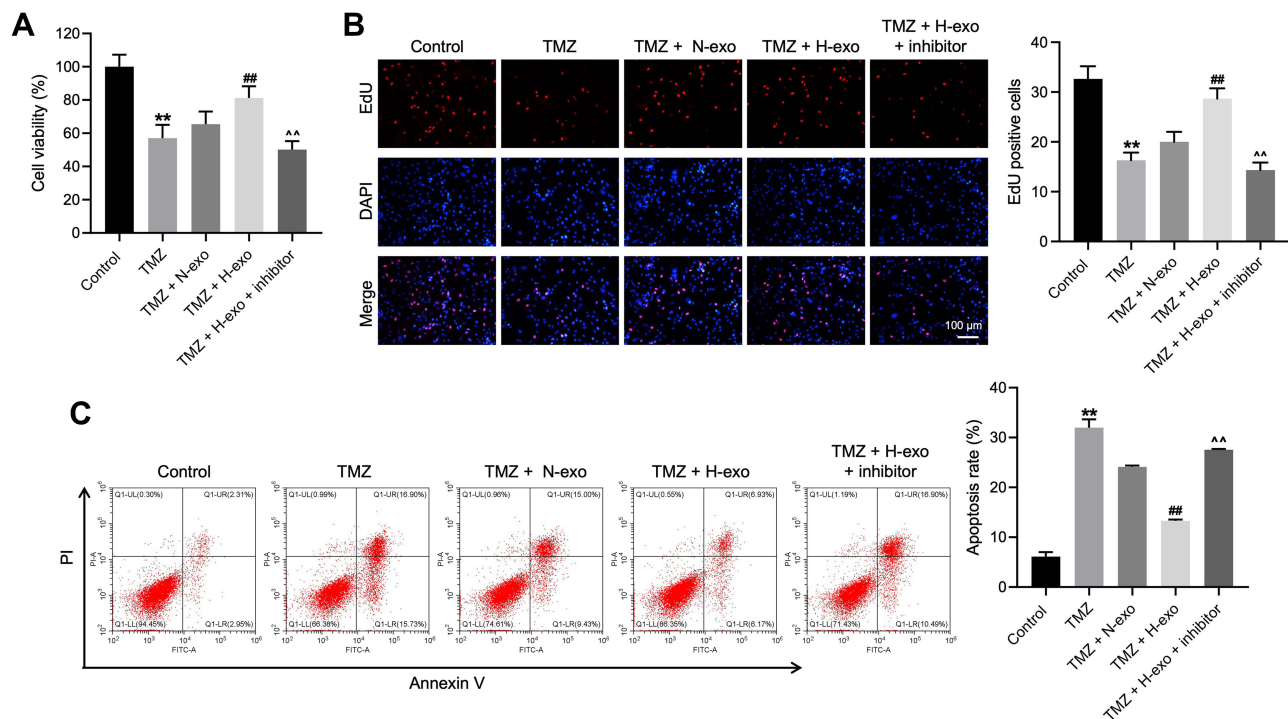


Figure 3 Exosomes derived from hypoxic glioma cells reduced TMZ sensitivity in glioma cells through carrying miR-106a-5p. Glioma cells were treated with TMZ, TMZ + N-exo, TMZ + H-exo or TMZ + H-exo + miR-106a-5p inhibitor. **(A)** The viability of glioma cells was assessed by CCK8 assay. **(B)** The proliferation of glioma cells was detected by EdU staining. **(C)** The apoptosis of glioma cells was assessed by flow cytometry. ** $P < 0.01$ compared to control. *** $P < 0.01$ compared to TMZ. ** $P < 0.01$ compared to TMZ + H-exo.

PTEN Was Identified to Be the Downstream Target of miR-106a-5p

To explore the downstream mRNA of miR-106a-5p, TargetScan was used. As revealed in Figure 4A, PTEN was predicted to be the downstream mRNA of miR-106a-5p. Additionally, the relative luciferase activity in WT-PTEN was significantly inhibited by upregulation of miR-106a-5p (Figure 4B). In addition, the level of PTEN in glioma cells was notably inhibited by miR-106a-5p mimics (Figure 4C). In summary, PTEN was a downstream target of miR-106a-5p.

Exosomes Derived from Hypoxic Glioma Cells Reduced TMZ Sensitivity in Glioma Cells Through Modulating PTEN/Akt Signaling

To further explore the mechanism by which exosomes from hypoxic glioma cells reduce TMZ sensitivity in glioma cells, Western blot was used. The data showed that TMZ significantly elevated the expressions of PTEN, Bax, p53 and decreased the expression of p-Akt in glioma cells (Figure 5A–E). However, H-exo notably reduced the expressions of PTEN, Bax, p53 and increased the expression of p-Akt in TMZ-treated glioma cells compared to TMZ treatment group, whereas these phenomena were reversed by miR-106a-5p inhibitor (Figure 5A–E). Taken together, hypoxic exosomes derived from glioma cells could reduce the sensitivity of glioma cells to TMZ through modulating PTEN/Akt signaling.

Exosomes Derived from Hypoxic Glioma Cells Reduced TMZ Sensitivity in Glioma Cells in vivo

Next, we further explored the role of exosomal miR-106a-5p in TMZ sensitivity in vivo. As illustrated in Figure 6A–C, the tumor volume and weight in mice were obviously decreased by TMZ treatment, while the anti-tumor effect of TMZ was partially abolished by H-exo. In addition, TMZ-induced cell apoptosis in tumor tissues was markedly reduced by H-exo (Figure 6D). Furthermore, the level of PTEN in tumor tissues was greatly upregulated by TMZ, whereas that

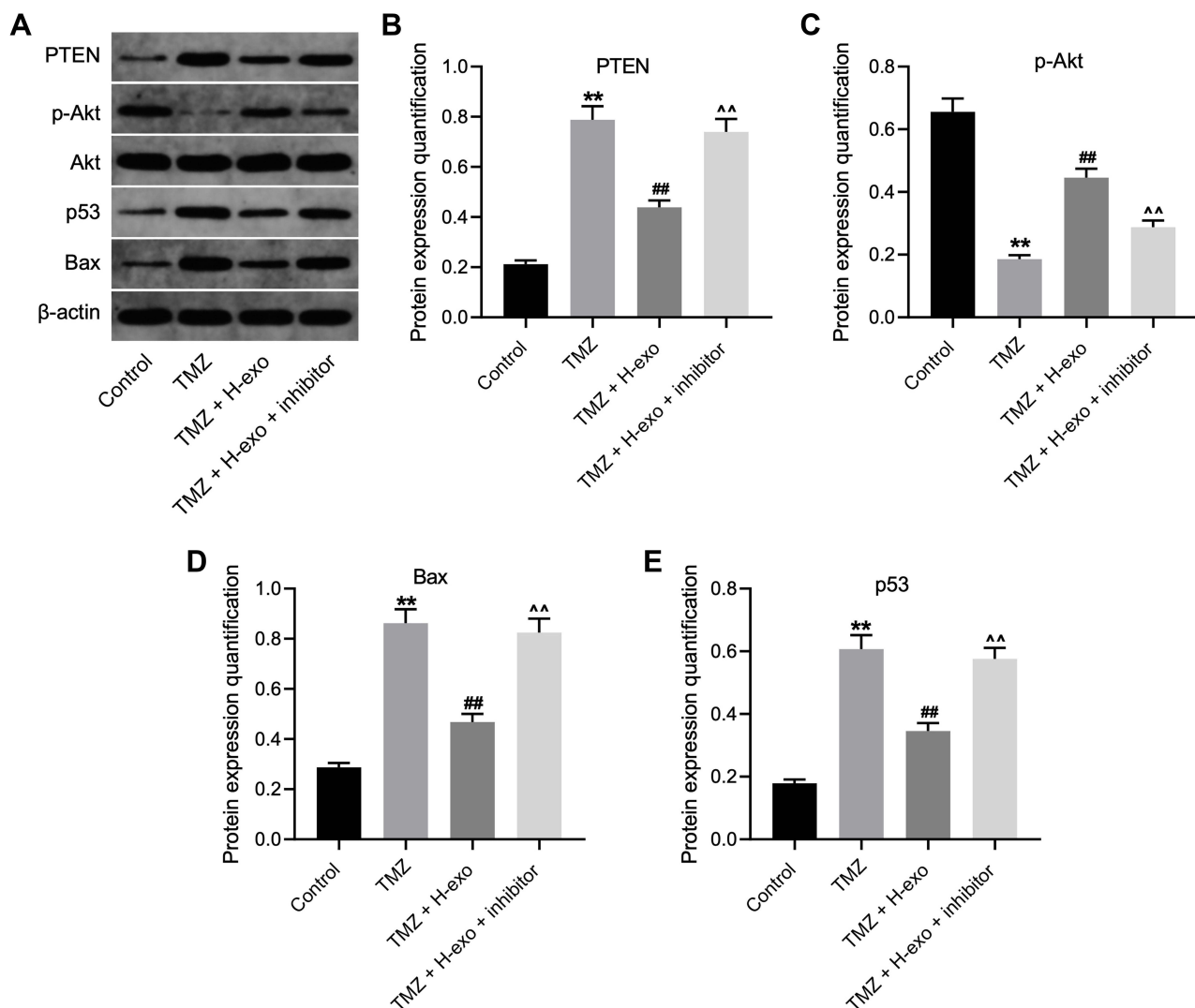


Figure 5 Exosomes derived from hypoxic glioma cells reduced TMZ sensitivity in glioma cells through inactivation of Akt signaling. **(A)** The protein levels of Akt, p-Akt, p53, Bax and PTEN in glioma cells were assessed by Western blot. **(B–E)** The relative expressions Akt, p-Akt, p53, Bax and PTEN in glioma cells were quantified by normalizing to β -actin. ** $P < 0.01$ compared to control. ## $P < 0.01$ compared to TMZ. ^^ $P < 0.01$ compared to TMZ + H-exo.

reduce the expression of PTEN and upregulate the level of p-Akt in TMZ-treated glioma cells. Meanwhile, Zhu et al found miR-106a-5p could promote the tumorigenesis of gastric cancer through targeting Smad7,⁴⁰ and our study was similar to this previous research. Smad7 was known to be an inhibitor of cancer cell invasion, and its overexpression could inhibit the malignant behavior of cancer cells.^{41,42} Hence, the similar function between PTEN and Smad7 might result in the similarity between our study and Zhu et al.

Indeed, there are some shortcomings in this research. First, in this study, we explored the role of hypoxic glioma cell-derived exosomal miR-106-5p in the regulation of TMZ resistance of glioma. However, whether other miRNAs (eg, miR-1246, miR-10b-5p and miR-301a) are involved in the regulation of TMZ resistance of glioma need to be investigated in the future. Second, more downstream targets of miR-106a-5p in glioma remain unexplored. Third, more signaling pathways involved in hypoxic exosomes-mediated chemoresistance are needed to be further discovered. Thus, more investigations are essential in the coming future.

In summary, exosomes derived from hypoxic glioma cells could reduce the sensitivity of glioma cells to TMZ through carrying miR-106a-5p. Hence, our research might provide a theoretical basis for exploring new strategies against glioma.

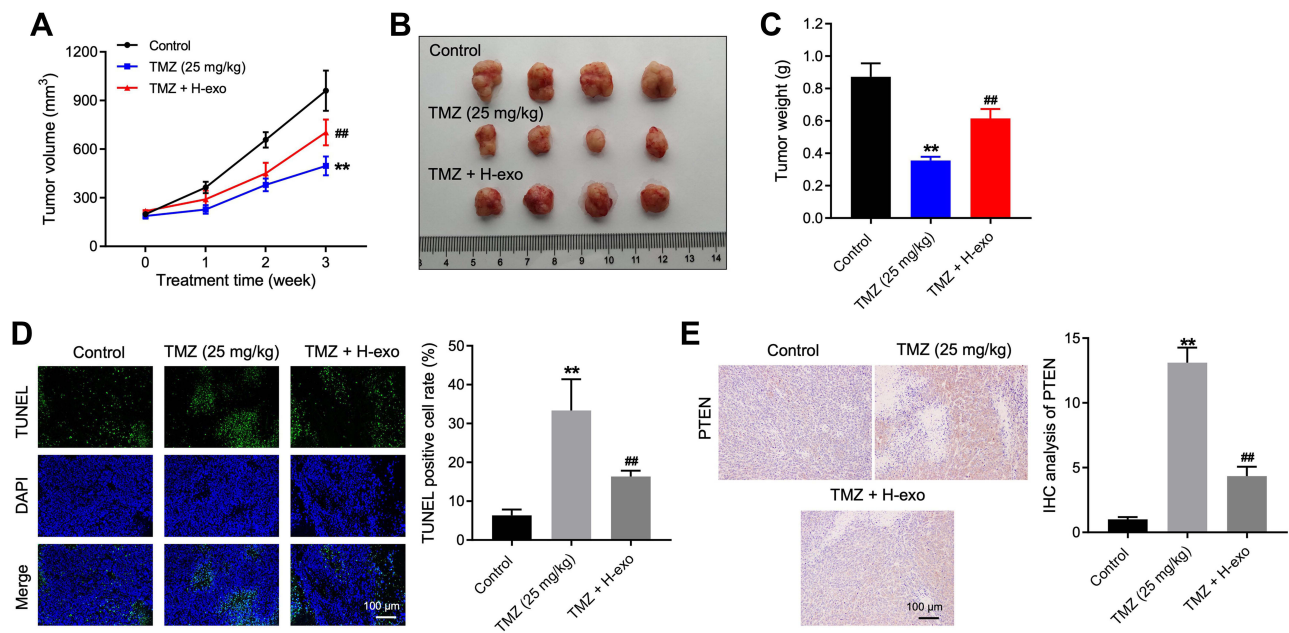


Figure 6 Exosomes derived from hypoxic glioma cells reduced TMZ sensitivity in glioma cells in vivo. **(A)** The tumor volume of mice was calculated. **(B)** The tumor tissues of mice were collected and pictured. **(C)** The tumor weight of mice was recorded. **(D)** The cell apoptosis of tumor tissues in mice was detected by TUNEL staining. **(E)** The level of PTEN in tumor tissues of mice was investigated by IHC staining. ** $P < 0.01$ compared to control. ## $P < 0.01$ compared to TMZ.

Disclosure

The authors declared no competing interests in this research.

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