



## Review

<https://doi.org/10.1631/jzus.B2200174>

# Tertiary lymphoid structures as unique constructions associated with the organization, education, and function of tumor-infiltrating immunocytes

Jing CHEN<sup>1,2</sup>, Jian CHEN<sup>1</sup>✉, Lie WANG<sup>2,3,4</sup>✉

<sup>1</sup>Department of Gastrointestinal Surgery, the Second Affiliated Hospital, Zhejiang University School of Medicine, Hangzhou 310009, China

<sup>2</sup>Institute of Immunology and Bone Marrow Transplantation Center, the First Affiliated Hospital, Zhejiang University School of Medicine, Hangzhou 310003, China

<sup>3</sup>Liangzhu Laboratory, Zhejiang University Medical Center, Hangzhou 311121, China

<sup>4</sup>Cancer Center, Zhejiang University, Hangzhou 310058, China

**Abstract:** Tertiary lymphoid structures (TLSs) are formations at sites with persistent inflammatory stimulation, including tumors. These ectopic lymphoid organs mainly consist of chemo-attracting B cells, T cells, and supporting dendritic cells (DCs). Mature TLSs exhibit functional organization for the optimal development and collaboration of adaptive immune response, delivering an augmented effect on the tumor microenvironment (TME). The description of the positive correlation between TLSs and tumor prognosis is reliable only under a certain condition involving the localization and maturation of TLSs. Emerging evidence suggests that underlying mechanisms of the anti-tumor effect of TLSs pave the way for novel immunotherapies. Several approaches have been developed to take advantage of intratumoral TLSs, either by combining it with therapeutic agents or by inducing the neogenesis of TLSs.

**Key words:** Tertiary lymphoid structure (TLS); Tumor-infiltrating immunocyte; Tumor microenvironment (TME)

## 1 Introduction

Tumor-infiltrating immune cells are key factors that hold the balance of the tumor microenvironment (TME), greatly affecting tumor progression and clinical outcomes (Giraldo et al., 2019; Lei et al., 2020). Numerous attempts have been made to manage tumors by modifying the immune states of the TME, either through immune checkpoint blockade (ICB) on dysfunctional T cells (Hodi et al., 2010; Pardoll, 2012) or through the transfusion of ex vivo engineered T cells (Tran et al., 2016; Gee et al., 2018; Depil et al., 2020). However, emerging evidence indicates that effector cells never fight alone during immune response.

Based on their presence in the TME, the localization, organization, and other cell types of the immune cell lineage can all greatly alter the tumor-associated immune response. It is worth pointing out that the compartmentalized organization defined by the physical separation of immune/non-immune clusters could independently determine the outcome of tumor sufferers. Ectopic aggregated but orderly organized immune cells are termed as tertiary lymphoid structures (TLSs) (Aloisi and Pujol-Borrell, 2006; Pitzalis et al., 2014), which are representative constructions of the mentioned context. The conspicuous performance of TLSs in tumors has been noticed (Sautès-Fridman et al., 2019), but the way they contribute to the function of tumor-infiltrating immunocytes by proper organization and education remains poorly understood. Aiming to shape the existing ideas about novel strategies of tumor management, in this paper, we briefly review the composition, organization, and prognostic value of TLSs, as well as the potential mechanism through which they educate tumor-infiltrating immunocytes.

✉ Lie WANG, wanglie@zju.edu.cn

Jian CHEN, zrchenjian@zju.edu.cn

✉ Lie WANG, <https://orcid.org/0000-0001-5094-012X>

Jian CHEN, <https://orcid.org/0000-0002-2542-9669>

Received Mar. 28, 2022; Revision accepted July 15, 2022;  
Crosschecked Aug. 25, 2022

© Zhejiang University Press 2022

## 2 Inconsistent immune performance in tumor-associated TLSs

The immune cells within the TME have been proved to be key factors in carcinogenesis, tumor progression, and cancer prognosis (Giraldo et al., 2019; Hinshaw and Shevde, 2019; Lei et al., 2020). The existing findings indicate that tumor-antagonizing or tumor-promoting functions are not only restricted to the immune cell types. It has been revealed that the same cell populations might perform opposite functions in certain conditions; however, the reasons for the unpredictability of these tumor-infiltrating immune cells have not been elucidated.

Among the various kinds of immune cells, B cells are considered to be the most controversial type. These cells are now recognized as key players in the core immune network, whose quantity could increase at a late tumor stage and result in a dual effect on recurrence and tumor progression (Bindea et al., 2013). A comprehensive analysis based on 69 studies covering 19 cancers stated that half of the cases possessed a positive association between tumor-infiltrating B cells (including plasma cells (PCs)) and the clinical outcome, whereas the remainder showed negative (9.3%) or neutral (40.7%) effect (Wouters and Nelson, 2018). Since the biases induced by cancer type, pathologic factors, or technical approaches have been removed, the inherent characteristics of the tumor-infiltrating B cells deserve increased attention. The transcriptome and proteomic analysis of ICB-treated melanoma and renal cell carcinoma cohorts revealed unique functional states of B cells in responders, which appeared more memory-like (Amaria et al., 2018; Helmink et al., 2020).

A similar paradox was also presented by follicular helper T (T<sub>fh</sub>) cells, which robustly predicted breast cancer survival or responsiveness to chemotherapy (Gu-Trantien et al., 2013) but was also reported to be negatively associated with hepatocellular carcinoma (HCC) survival (Shalapour et al., 2017).

Additional findings showed that patients with both T and B cells infiltrated into tumors were more likely to possess favorable prognosis than those with only T or only B cells. The co-occurrence of cluster of differentiation 8-positive (CD8<sup>+</sup>) T cells and CD20<sup>+</sup> B cells within tumors was found independently associated with improved survival (Cabrita et al., 2020).

Moreover, tumor-infiltrating CD8<sup>+</sup> T cells, the well accepted positive prognostic factor, only exhibited their cytotoxic capacity and prognostic value if B cells were also augmented within the tumor (Kroeger et al., 2016; Wouters and Nelson, 2018). Tumors rich in B cells were more likely to be infiltrated with increased levels of naive and memory T cells that were supposed to do well in anti-tumor response (Cabrita et al., 2020). It has been recently acknowledged that immune cells alone are incapable of proper function, as it is their coordinated action that is critical for both the physiological and pathological processes. The complex network of interactions bridges the gaps among diverse immune cells, as well as non-immune cells (Garner and de Visser, 2020).

The varied effect of multiple kinds of immunocytes triggered by their co-localization and cooperation reaches the concept of TLSs. These are well-organized aggregates of immune cells in non-lymphoid tissues that are subjected to long-lasting inflammatory response, including chronic infection, autoimmune disease, as well as tumors (Sautès-Fridman et al., 2019). TLSs are composed of an inner CD20<sup>+</sup> B cell-zone mantled with CD3<sup>+</sup> T cell-zone, anchored with dendritic cells (DCs) and vessel structure, similar to the lymphoid follicles in secondary lymphoid organs (SLOs) (Aloisi and Pujol-Borrell, 2006; Drayton et al., 2006).

TLSs provide an appropriate niche for communication among diverse immune cells. The unencapsulated structure of TLSs enables sufficient interactions between immune cells and the surrounding microenvironment (Braun et al., 2011). It is reasonable to speculate that the contradictory property of certain immune cells might be attributed to the formation and organization of TLSs to some extent. Supporting evidence shows that only when organized into TLSs, did B cells predict better prognosis and higher response rate to immunotherapy (Castino et al., 2016; Petitprez et al., 2020). B cells outside TLSs are likely to inhibit anti-tumor immunity and contribute to tumor growth (Germain et al., 2014; Cabrita et al., 2020). Similarly, T cells located inside or outside TLSs, or those sporadically infiltrated in tumors without TLSs, showed a distinct transcription signature and molecular phenotype, with the latter group of T cells exhibiting dysfunctional preference (Cabrita et al., 2020). Meanwhile, CXCL13<sup>+</sup>CD4<sup>+</sup> T<sub>fh</sub> cells, one of the major components of TLSs, always indicate extensive immune

infiltration within the tumor, intense response to chemotherapy, and longer survival (Gu-Trantien et al., 2013). CXCL13<sup>+</sup>CD4<sup>+</sup> and CXCL13<sup>+</sup>CD8<sup>+</sup> T cells could hardly enhance the ICB effect without forming TLSs (He et al., 2022). In brief, TLSs facilitate coordinated antitumor response by involving the combined actions of mainly T cells, B cells, and DCs (Germain et al., 2014; Goc et al., 2014; Carmi et al., 2015; Kroeger et al., 2016).

### 3 Composition and organization of TLSs in cancer

As introduced previously, intratumoral TLSs maintain an active immune response (Hiraoka et al., 2015). Moreover, the cooperation among the affiliates is well-organized. Although still partly unclear, the current understanding of the generation of TLSs was mostly referred to SLOs considering the anatomical resemblance (Drayton et al., 2006). This process is initiated with the colonization of lymphoid tissue inducer (LTi) cells that interact with lymphoid tissue organizer (LTo) cells by lymphotoxin signaling (de Togni et al., 1994; Mebius et al., 1997; Dejardin et al., 2002; Yoshida et al., 2002; Veiga-Fernandes et al., 2007). The abovementioned interactions promote the expression of adhesion molecules (e.g., vascular cell adhesion molecule 1 (VCAM1), intercellular adhesion molecule 1 (ICAM1), mucosal vascular addressin cell adhesion molecule 1 (MAdCAM1), and peripheral lymph node addressin (PNAd)) and the production of chemokines (e.g., chemokine (C-C motif) ligand 19 (CCL19), CCL21, and chemokine (C-X-C motif) ligand 13 (CXCL13)), which further leads to vascularization and immune cell recruitment (Gunn et al., 1998; Ngo et al., 1999; Suzuki et al., 1999; Ansel et al., 2000; Luther et al., 2003). However, it has been described that the induction of TLSs was not restricted to lymphotoxin signaling, and several immune cell populations, such as group 3 innate lymphoid cells (ILC3), B cells, T helper 17 (Th17) cells, effector CD8<sup>+</sup> T cells, and M1 macrophages, were found to be surrogate LTi cells (Lochner et al., 2011; Peters et al., 2011; Guedj et al., 2014; Carrega et al., 2015; Peske et al., 2015). Similarly, certain stromal and immune cells were thought to act as potential surrogate LTo cells (Bénézech et al., 2010; Barone et al., 2016; Rodriguez et al., 2021).

Even with certain distinctions during the initiation of TLSs and SLOs, the main chemokines involved in the downstream process are shared between these two structures. Among the significant chemokines, CXCL13 performs as a crucial element due to its full participation throughout the TLS organization. As a typical marker of TLSs, the expression of CXCL13 can also be utilized for prognosis prediction. Patients with CXCL13 deletion suffer a significantly higher risk of relapse (Bindea et al., 2013). CXCL13, also defined as B lymphocyte chemoattractant, has been proved to be sufficient to mediate B cell recruitment, together with the downstream process for lymphoid neogenesis, including but not limited to vessel differentiation, chemokine induction, T/B segregation, and germinal center (GC) formation (Luther et al., 2000). Moreover, CXCL13 has also been reported as a unique marker for tumor antigen-specific CD4<sup>+</sup> and CD8<sup>+</sup> T cells, as well as good prognosis (Groeneveld et al., 2021; He et al., 2022).

One of the main sources of CXCL13 points to Tfh cells, indispensable in forming extrafollicular and GC responses, where they are crucial for humoral immunity (Gunn et al., 1998; Allen et al., 2007; Kerfoot et al., 2011). CXCL13-producing Tfh cells have also been defined as a positive prognostic factor in several tumors, more or less profiting from its promotion for TLS organization (Gu-Trantien et al., 2013; Li et al., 2021). In fact, CXCL13-producing antigen-specific Tfh cells, which are necessary and sufficient for TLS formation, effectively dampen colorectal cancer (CRC) development with assistance of B cells (Overacre-Delgoffe et al., 2021). In addition, certain CD8<sup>+</sup> T subsets and stromal cells also turned out to be the origin of CXCL13, accompanied by antigen presenting capacity (van de Pavert et al., 2009; Workel et al., 2019; Wu et al., 2020; He et al., 2022). Interestingly, these CXCL13-producing CD8<sup>+</sup> T cells are always recognized as exhausted CD8<sup>+</sup> T cells (CD8<sup>+</sup> Tex) owing to the highly expressed inhibitory immune checkpoint molecules (such as programmed cell death protein-1 (PD-1), cytotoxic T lymphocyte-associated protein-4 (CTLA-4), T-cell immunoglobulin domain and mucin domain-containing molecule 3 (TIM-3), lymphocyte activation gene-3 (LAG-3), and T cell immunoglobulin and immunoreceptor tyrosine-based inhibitory motif (ITIM) domain (TIGIT)) (Thommen et al., 2018; Dai et al., 2021; Jin et al., 2021). Unlike the previously

accepted poor prognostic value of CD8<sup>+</sup> T<sub>H</sub>17 in tumors, the accumulation of CD8<sup>+</sup> T<sub>H</sub>17 favors the clinical outcome in CRC, melanoma, and breast cancer, when accompanied with T<sub>H</sub>1 (Niogret et al., 2021). In addition, other chemokines such as CXCL12, CCL19, CCL21, and tumor necrosis factor (TNF) also contribute to lymphoid tissue organogenesis by supporting the infiltration of B cells, T cells, and DCs (Luther et al., 2002; Furtado et al., 2014). CCL21 was also shown to be beneficial for the formation of more and larger TLSs (Luther et al., 2002).

Alongside B and T cells that dominate in TLSs, DCs also matter in TLS formation and function. Follicular dendritic cells (FDCs) localized within GC are critical for the selection of memory B cells, while mature dendritic cell-lysosomal-associated membrane proteins (DC-LAMPs; also present as a special marker for tumor-associated TLSs) that are predominantly distributed in the T cell zone act as the major type of antigen-presenting cells (APCs) to facilitate the generation of tumor-reactive T cells (Wykes et al., 1998; di Pucchio et al., 2008; Schmitt et al., 2009; Goc et al., 2014). DC is generally required for early T<sub>H</sub>17 differentiation, indicating its indispensable role in the initial step of TLS formation (Choi et al., 2011; Goenka et al., 2011; Kato et al., 2015; Krishnaswamy et al., 2017). What is more, the formation and maintenance of TLSs are greatly dependent on antigen recognition. As long as the antigens are thoroughly eliminated, TLSs will generally disintegrate (Geurtsvankessel et al., 2009; Halle et al., 2009). The enrichment of mature DC in TLSs also displayed a potential promotive effect on T cell infiltration, activation, and cytotoxic orientation; moreover, the transmission of CD4<sup>+</sup> T cell-derived help signals consequently contributed to long-term survival (Schoenberger et al., 1998; Yang et al., 2006; Goc et al., 2014; Eickhoff et al., 2015; Hor et al., 2015; Lee et al., 2017; Weinstein et al., 2017).

#### 4 Prognostic value of TLSs in cancer

Accumulating evidence suggests the magnified anti-tumor immune response in TLSs. Meanwhile, a favorable prognostic and predictive impact of TLSs has been well presented in many kinds of solid tumor (Sautès-Fridman et al., 2019). Also, the prognostic

value of TLSs was found to be independent of tumor stage as well as other survival factors (Hiraoka et al., 2015; Posch et al., 2018). Studies focusing on tumor ICB therapy have preliminarily revealed the correlation between TLSs and the response to ICB. For multiple tumors including melanoma, CRC, non-small cell lung cancer (NSCLC), and so on, patients with intratumoral TLSs, sometimes featured with enriched B cells or PD-1<sup>hi</sup> dysfunctional CD8<sup>+</sup> T cells, could better respond to ICB therapy (Buisseret et al., 2017; Johansson-Percival et al., 2017; Cabrita et al., 2020; Helmink et al., 2020; Petitprez et al., 2020).

The biological advantages of TLSs manifest as an increased opportunity for tumor-specific immune cells to encounter the matched antigens. On the other hand, the enhanced immune response benefits from the collaboration among diverse immune cells. It is well known that the fate of T<sub>H</sub>17 cells, also defined as tumor-specific CD4<sup>+</sup> T cells in TME, can be regulated by B cells and B cell-recognized neoantigens. Afterwards, the primed T<sub>H</sub>17 cells enhance the activation and cytotoxic function of CD8<sup>+</sup> T cells by producing interleukin-21 (IL-21) for an anti-tumor effect (Cui et al., 2021; Niogret et al., 2021). In turn, the recruitment of T<sub>H</sub>17 cells is partly due to CXCL13 expression by intratumoral CD8<sup>+</sup> T cells (Niogret et al., 2021). Furthermore, the sufficient antigen-driven interaction between CD4<sup>+</sup> T cells and DC produces CCL3/4 to directly attract CD8<sup>+</sup> T cells and increase the possibility of optimal stimulation of antigen-specific CD8<sup>+</sup> T cells (Castellino et al., 2006).

The exact mechanisms for boosting the immune reactivity of TLSs still remain elusive. The prevailing ideas can be roughly summarized into three main groups. (1) Some effector cells are preferred to be recruited into TLSs, such as ILC3 (Carrega et al., 2015), natural killer T (NKT) cells (Bénézech et al., 2015), and effective PCs (Lochner et al., 2011), which participate in the organization of TLSs. (2) Local priming derived from TLSs orients naive populations to have a particular status, forming the evidence that naive T cells can be attracted into TLSs and mature into CTLs (Lee et al., 2006; Garner and de Visser, 2020), and the resident stem-like CD8<sup>+</sup> T cells are likely to enter dysfunctional terminal state that can sustain antigen stimulation for the maintenance of TLSs (Jansen et al., 2019). Meanwhile, B cells within TLSs are more likely switched into memory B cells,

and prefer to mature toward IgG-producing PCs that boost ICB response by labeling tumor cells with IgG (Helmink et al., 2020; Kinker et al., 2021; Meylan et al., 2022), and also promoting CD8<sup>+</sup> T cells infiltration (Castino et al., 2016). Without TLSs, however, B cells would function suboptimally and more likely acquire suppressive function (Siliņa et al., 2018; Cillo et al., 2020; Ruffin et al., 2021). It could be interpreted that TLSs-resident B cells are well embraced by T cells and DCs, and protected from immunosuppressive stimuli by tumor cells (Germain et al., 2014; Siliņa et al., 2018; Cabrita et al., 2020). (3) Existing effector populations could be reeducated to gain novel characteristics. Th17 cells, one type of surrogate LTi cells, could be induced into Tfh-like population with upregulated CXCL13 expression for TLS induction (Lochner et al., 2011; Peters et al., 2011). At the same time, Tfh cells could possess cytotoxic ability as functional Th1-oriented cells under the education of TLSs (Goc et al., 2014; Noël et al., 2021). Even Tfh cells that locate in distinct sites diverge along their differentiation trajectory, with Tfh outside GC proliferating continuously while GC-resident Tfh stopping proliferation soon after differentiation but performing better in vesicle organization and exocytosis (Yeh et al., 2022). The previously defined “dysfunctional” CD8<sup>+</sup> Tex cells have been found as a highly proliferative and dynamically differentiating subset in the TME (Li et al., 2019), indicating their unique role in tumor-associated immune response. CD8<sup>+</sup> Tex cells always locate within tumor sites thanks to tumor antigen-specific priming, and these cells can be educated by stroma-derived transforming growth factor- $\beta$  (TGF- $\beta$ ) to secrete Tfh/B cell-recruiting CXCL13 to assist with the formation of TLSs exactly inside tumors (Workel et al., 2019). On the other hand, the “exhausted” phenotype of CD8<sup>+</sup> T cells caused by continuous stimulation could counteract with the help of CD4<sup>+</sup> T cells. It has been reported that antigen-activated CD4<sup>+</sup> T cells could improve the clonal expansion of effector CD8<sup>+</sup> T cells and strengthen tumor-reactive CD8<sup>+</sup> T cells by downregulating coinhibitory receptors, increasing migration-related chemokine receptors, and boosting the generation of the cytotoxic effect (Bennett et al., 1997; Ridge et al., 1998; Janssen et al., 2003; Shedlock and Shen, 2003; Ahrends et al., 2017).

As a matter of fact, the composition and function of TLSs vary along with tumor progression, and their

presence will not necessarily subvert the immunosuppressive state of the TME, exemplified as highly inflamed tumors (Sautès-Fridman et al., 2019). There are many factors that determine the phenotype of TLSs, including but not limited to their quantity, location, composition, and degree of maturation. Visually, the morphology of TLSs can differ significantly according to their location, even within the same tumor sample (Munoz-Erazo et al., 2020), but whether they differ in biological function remains unclear. What is already known is that the location of TLSs is a determinant of their anti-tumor ability. The favorable prognostic value of intratumoral TLSs has been confirmed in HCC, while the adverse effect of tumor-adjacent TLSs has also been established. The tumor margin area is prone to be in an inflamed condition, which potentially alters TLS to become a refuge for tumor stem cells that might cause late recurrence (Calderaro et al., 2019).

It is reasonable to presume that not only the density and location, but also the composition and degree of maturation of TLSs could significantly impact their function. Evidence shows that the integration of TLS density and TLS maturation can better predict tumor prognosis than each parameter separately (Posch et al., 2018). To our knowledge, there are mainly three stages during TLSs maturation. At the initial stage, immature TLSs are segregated aggregates of recruited T and B cells. In the intermediate maturation stage, follicular DCs penetrate into the niches. Finally, GCs are organized within the structure and distinguish mature TLSs. The maturation of TLSs not only affects the activation, proliferation, and affinity maturation of B cells, but also determines the therapeutic response and prognostic value of TLSs (Berek et al., 1991; Germain et al., 2014; Siliņa et al., 2018). In other words, only TLSs with GCs are active and functional.

## 5 Clinical application and therapeutic induction of TLSs

In view of the given anti-tumor effect of mature TLSs, these fascinating structures could turn into ideal therapeutic targets for a variety of solid tumors.

Since it has been well documented that the presence of mature TLSs always predicts better prognosis in the vast majority of solid tumors (Sautès-Fridman

et al., 2019), the strategy for these patients is to make full use of the anti-tumor effect of TLSs. Large clinical cohorts have revealed the positive correlation between TLSs and canonical tumor therapy. Better response to neoadjuvant therapy, which leads to increased pathological complete response, was found to be significantly associated with higher TILs in breast cancer, regardless of the molecular subtypes (Song et al., 2017; Denkert et al., 2018). The application of ICB in sarcoma, melanoma, and renal cell carcinoma was reinforced by the presence of tumor-associated TLSs (Cabrita et al., 2020; Helmink et al., 2020; Petitprez et al., 2020).

With regard to the other group of the patients who lack mature TLSs within the tumor sites, different attempts aiming to trigger the induction of TLSs have been made. Considering that lymphatic vessels are essential for immune cell trafficking, it is reasonable that the major attempts to promote TLS formation began with the induction of high endothelial venules (HEVs) to promote immune infiltration, which has proved feasible (Jiang, 2020). This was achieved by applying lymphotoxin (Lee et al., 2006; Mounzer et al., 2010), vascular targeted LIGHT (an alias for TNF superfamily member 14 (TNFSF14)) (Johansson-Percival et al., 2017), or a combination of antiangiogenesis with programmed death-ligand 1 (PD-L1) blockade (Allen et al., 2017). HEVs surrounding or passing through TLSs function as gateways for lymphocyte traffic into tumors, as well as scaffolds for lymphoid structure formation (Martinet et al., 2011; Asrir et al., 2022). The existing research shows that a high density of HEVs inside tumors independently assures longer survival and lower risk of relapse (Martinet et al., 2011; Bindea et al., 2013). Besides, the tumor destruction achieved by Treg depletion was partly attributed to the formation of induced HEVs (Joshi et al., 2015; Colbeck et al., 2017). The above evidence elevates the accessibility and feasibility of targeting HEVs to induce TLS formation for tumor treatment. However, most of these attempts were limited to animal studies; hence additional clinical studies are in urgent need. It is encouraging that progress has been achieved in other clinical trials. Certain patients might benefit from canonical therapy, exemplified by hepatoblastoma patients carrying an adenomatous polyposis coli (APC) germline mutation, who could acquire better prognosis by cisplatin-induced TLSs (Morcrette et al., 2019). In addition, the application of human

papillomavirus (HPV) vaccine and allogeneic pancreatic ductal adenocarcinoma vaccine successfully increased the infiltration of effector T cells, which further organized as TLSs to improve survival (Lutz et al., 2014; Maldonado et al., 2014).

It should be noted that, since the potent immune reactivity of TLSs provides their ability to antagonize tumor cells, concomitant autoimmunity should not be overlooked. Limited studies have revealed a risk of lymphocyte leakage for TLS formation caused by ICB therapy. The enrichment of cytotoxic cells within ectopic TLSs led to muscle fiber degeneration, identified as PD-1 myopathy (Matsubara et al., 2019). As a management of autoimmune toxicity caused by immunotherapy, the application of corticosteroids could markedly impair the formation and function of TLSs (Siliņa et al., 2018; van Dijk et al., 2020), leaving us with an elaborate risk-benefit ratio that needs careful evaluation.

## 6 Conclusions and prospects

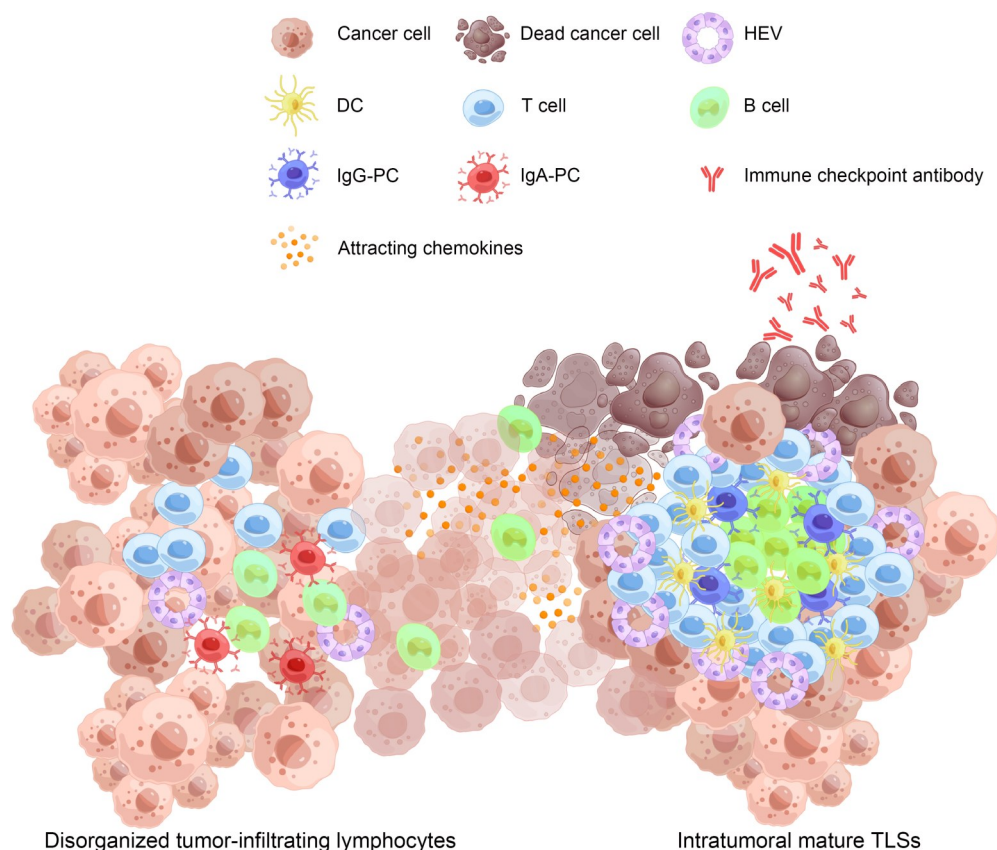
It is beyond doubt that TLSs possess promising prognostic and predictive values in cancers. They provide a chance for various immune cells to be well orchestrated to maximize the immune response (Fig. 1). Meanwhile, the effect of TLSs acting on tumor-associated immune response is also plausible, which is susceptible to their localization, stage of maturation, and other factors. The flexibility of TLSs also offers us opportunities to manipulate their formation by adjusting certain conditions to achieve the desired therapeutic goal. To this end, a lot of research has been put forward and made preliminary achievements. Follow-up efforts are still needed for a more detailed understanding and a more effective use of tumor-associated TLSs for anticancer therapy.

### Acknowledgments

This work was supported by the Zhejiang Provincial Key Project of Research and Development (No. 2019C03043), the National Natural Science Foundation of China (Nos. 32030035, 31870874, 32000623, and 32100693), and the Zhejiang Provincial Natural Science Foundation (No. LZ21C080001) of China.

### Author contributions

Jing CHEN was responsible for reference searching and manuscript writing, and Lie WANG and Jian CHEN for



**Fig. 1** Composition and potential function of TLSs in cancer. The above schematic represents immune cells infiltrated within the tumor site in two main forms. (1) Disorganized distribution within the tumor: scattered immune cells induced by tumor antigens are oriented to tumor-promoting phenotypes, represented as dysfunctional T cells and IgA-PC. (2) Assembled as TLSs within the tumor: TLSs are organized with an inner B cell-zone, mantled by the T cell-zone, with DC spreading over the structures. Surrounding HEVs act as gateways for chemo-attracted immunocytes entering the site. TLSs augment the collaboration of the aggregated immunocytes, leading to tumor-antagonizing status mainly mediated by effector T cells and IgG-PC. The presence of TLSs greatly boosts ICB therapy and efficiently disrupts tumor cells. TLS: tertiary lymphoid structure; HEV: high endothelial venule; DC: dendritic cell; IgG-PC: IgG-producing plasma cell; IgA-PC: IgA-producing plasma cell; ICB: immune checkpoint blockade.

supervision and manuscript revision. All authors have approved the final manuscript.

### Compliance with ethics guidelines

Jing CHEN, Jian CHEN, and Lie WANG declare that they have no conflict of interest.

This review does not contain any studies with human or animal subjects performed by any of the authors.

### References

- Ahrends T, Spanjaard A, Pilzecker B, et al., 2017. CD4<sup>+</sup> T cell help confers a cytotoxic T cell effector program including coinhibitory receptor downregulation and increased tissue invasiveness. *Immunity*, 47(5):848-861.e5. <https://doi.org/10.1016/j.immuni.2017.10.009>
- Allen CDC, Okada T, Tang HL, et al., 2007. Imaging of germinal center selection events during affinity maturation. *Science*, 315(5811):528-531. <https://doi.org/10.1126/science.1136736>
- Allen E, Jabouille A, Rivera LB, et al., 2017. Combined anti-angiogenic and anti-PD-L1 therapy stimulates tumor immunity through HEV formation. *Sci Transl Med*, 9(385):eaak9679. <https://doi.org/10.1126/scitranslmed.aak9679>
- Aloisi F, Pujol-Borrell R, 2006. Lymphoid neogenesis in chronic inflammatory diseases. *Nat Rev Immunol*, 6(3):205-217. <https://doi.org/10.1038/nri1786>
- Amaria RN, Reddy SM, Tawbi HA, et al., 2018. Neoadjuvant immune checkpoint blockade in high-risk resectable melanoma. *Nat Med*, 24(11):1649-1654. <https://doi.org/10.1038/s41591-018-0197-1>
- Ansel KM, Ngo VN, Hyman PL, et al., 2000. A chemokine-driven positive feedback loop organizes lymphoid follicles. *Nature*, 406(6793):309-314. <https://doi.org/10.1038/35018581>
- Asrir A, Tardiveau C, Coudert J, et al., 2022. Tumor-associated high endothelial venules mediate lymphocyte entry into

- tumors and predict response to PD-1 plus CTLA-4 combination immunotherapy. *Cancer Cell*, 40(3):318-334.e9. <https://doi.org/10.1016/j.ccell.2022.01.002>
- Barone F, Gardner DH, Nayar S, et al., 2016. Stromal fibroblasts in tertiary lymphoid structures: a novel target in chronic inflammation. *Front Immunol*, 7:477. <https://doi.org/10.3389/fimmu.2016.00477>
- Bénézech C, White A, Mader E, et al., 2010. Ontogeny of stromal organizer cells during lymph node development. *J Immunol*, 184(8):4521-4530. <https://doi.org/10.4049/jimmunol.0903113>
- Bénézech C, Luu NT, Walker JA, et al., 2015. Inflammation-induced formation of fat-associated lymphoid clusters. *Nat Immunol*, 16(8):819-828. <https://doi.org/10.1038/ni.3215>
- Bennett SRM, Carbone FR, Karamalis F, et al., 1997. Induction of a CD8<sup>+</sup> cytotoxic T lymphocyte response by cross-priming requires cognate CD4<sup>+</sup> T cell help. *J Exp Med*, 186(1):65-70. <https://doi.org/10.1084/jem.186.1.65>
- Berek C, Berger A, Apel M, 1991. Maturation of the immune response in germinal centers. *Cell*, 67(6):1121-1129. [https://doi.org/10.1016/0092-8674\(91\)90289-b](https://doi.org/10.1016/0092-8674(91)90289-b)
- Bindea G, Mlecnik B, Tosolini M, et al., 2013. Spatiotemporal dynamics of intratumoral immune cells reveal the immune landscape in human cancer. *Immunity*, 39(4):782-795. <https://doi.org/10.1016/j.immuni.2013.10.003>
- Braun A, Worbs T, Moschovakis GL, et al., 2011. Afferent lymph-derived T cells and DCs use different chemokine receptor CCR7-dependent routes for entry into the lymph node and intranodal migration. *Nat Immunol*, 12(9):879-887. <https://doi.org/10.1038/ni.2085>
- Buisseret L, Garaud S, de Wind A, et al., 2017. Tumor-infiltrating lymphocyte composition, organization and PD-1/PD-L1 expression are linked in breast cancer. *Oncot Immunology*, 6(1):e1257452. <https://doi.org/10.1080/2162402X.2016.1257452>
- Cabrita R, Lauss M, Sanna A, et al., 2020. Tertiary lymphoid structures improve immunotherapy and survival in melanoma. *Nature*, 577(7791):561-565. <https://doi.org/10.1038/s41586-019-1914-8>
- Calderaro J, Petitprez F, Becht E, et al., 2019. Intra-tumoral tertiary lymphoid structures are associated with a low risk of early recurrence of hepatocellular carcinoma. *J Hepatol*, 70(1):58-65. <https://doi.org/10.1016/j.jhep.2018.09.003>
- Carmi Y, Spitzer MH, Linde IL, et al., 2015. Allogeneic IgG combined with dendritic cell stimuli induce antitumor T-cell immunity. *Nature*, 521(7550):99-104. <https://doi.org/10.1038/nature14424>
- Carrega P, Loiacono F, di Carlo E, et al., 2015. NCR<sup>+</sup>ILC3 concentrate in human lung cancer and associate with intratumoral lymphoid structures. *Nat Commun*, 6:8280. <https://doi.org/10.1038/ncomms9280>
- Castellino F, Huang AY, Altan-Bonnet G, et al., 2006. Chemokines enhance immunity by guiding naive CD8<sup>+</sup> T cells to sites of CD4<sup>+</sup> T cell-dendritic cell interaction. *Nature*, 440(7086):890-895. <https://doi.org/10.1038/nature04651>
- Castino GF, Cortese N, Capretti G, et al., 2016. Spatial distribution of B cells predicts prognosis in human pancreatic adenocarcinoma. *Oncot Immunology*, 5(4):e1085147. <https://doi.org/10.1080/2162402X.2015.1085147>
- Choi YS, Kageyama R, Eto D, et al., 2011. ICOS receptor instructs T follicular helper cell versus effector cell differentiation via induction of the transcriptional repressor Bcl6. *Immunity*, 34(6):932-946. <https://doi.org/10.1016/j.immuni.2011.03.023>
- Cillo AR, Kurten CHL, Tabib T, et al., 2020. Immune landscape of viral- and carcinogen-driven head and neck cancer. *Immunity*, 52(1):183-199.e9. <https://doi.org/10.1016/j.immuni.2019.11.014>
- Colbeck EJ, Jones E, Hindley JP, et al., 2017. Treg depletion licenses T cell-driven HEV neogenesis and promotes tumor destruction. *Cancer Immunol Res*, 5(11):1005-1015. <https://doi.org/10.1158/2326-6066.CIR-17-0131>
- Cui C, Wang JW, Fagerberg E, et al., 2021. Neoantigen-driven B cell and CD4 T follicular helper cell collaboration promotes anti-tumor CD8 T cell responses. *Cell*, 184(25):6101-6118.e13. <https://doi.org/10.1016/j.cell.2021.11.007>
- Dai SY, Zeng H, Liu ZP, et al., 2021. Intratumoral CXCL13<sup>+</sup> CD8<sup>+</sup> T cell infiltration determines poor clinical outcomes and immunoevasive contexture in patients with clear cell renal cell carcinoma. *J Immunother Cancer*, 9(2):e001823. <https://doi.org/10.1136/jitc-2020-001823>
- Dejardin E, Droin NM, Delhase M, et al., 2002. The lymphotoxin-β receptor induces different patterns of gene expression via two NF-κB pathways. *Immunity*, 17(4):525-535. [https://doi.org/10.1016/s1074-7613\(02\)00423-5](https://doi.org/10.1016/s1074-7613(02)00423-5)
- Denkert C, von Minckwitz G, Darb-Esfahani S, et al., 2018. Tumour-infiltrating lymphocytes and prognosis in different subtypes of breast cancer: a pooled analysis of 3771 patients treated with neoadjuvant therapy. *Lancet Oncol*, 19(1):40-50. [https://doi.org/10.1016/S1470-2045\(17\)30904-X](https://doi.org/10.1016/S1470-2045(17)30904-X)
- Depil S, Duchateau P, Grupp SA, et al., 2020. 'Off-the-shelf' allogeneic CAR T cells: development and challenges. *Nat Rev Drug Discov*, 19(3):185-199. <https://doi.org/10.1038/s41573-019-0051-2>
- de Togni P, Goellner J, Ruddle NH, et al., 1994. Abnormal development of peripheral lymphoid organs in mice deficient in lymphotoxin. *Science*, 264(5159):703-707. <https://doi.org/10.1126/science.8171322>
- di Pucchio T, Chatterjee B, Smed-Sörensen A, et al., 2008. Direct proteasome-independent cross-presentation of viral antigen by plasmacytoid dendritic cells on major histocompatibility complex class I. *Nat Immunol*, 9(5):551-557. <https://doi.org/10.1038/ni.1602>
- Drayton DL, Liao S, Mounzer RH, et al., 2006. Lymphoid organ development: from ontogeny to neogenesis. *Nat Immunol*, 7(4):344-353. <https://doi.org/10.1038/ni1330>
- Eickhoff S, Brewitz A, Gerner MY, et al., 2015. Robust antiviral immunity requires multiple distinct T cell-dendritic cell interactions. *Cell*, 162(6):1322-1337. <https://doi.org/10.1016/j.cell.2015.08.004>
- Furtado GC, Pacer ME, Bongers G, et al., 2014. TNFα-dependent development of lymphoid tissue in the absence of RORγt<sup>+</sup> lymphoid tissue inducer cells. *Mucosal Immunol*, 7(3):602-614.



- <https://doi.org/10.1038/mi.2013.79>
- Garner H, de Visser KE, 2020. Immune crosstalk in cancer progression and metastatic spread: a complex conversation. *Nat Rev Immunol*, 20(8):483-497. <https://doi.org/10.1038/s41577-019-0271-z>
- Gee MH, Han A, Lofgren SM, et al., 2018. Antigen identification for orphan T cell receptors expressed on tumor-infiltrating lymphocytes. *Cell*, 172(3):549-563.e16. <https://doi.org/10.1016/j.cell.2017.11.043>
- Germain C, Gnjatic S, Tamzalit F, et al., 2014. Presence of B cells in tertiary lymphoid structures is associated with a protective immunity in patients with lung cancer. *Am J Respir Crit Care Med*, 189(7):832-844. <https://doi.org/10.1164/rccm.201309-1611OC>
- Geurtsvankessel CH, Willart MAM, Bergen IM, et al., 2009. Dendritic cells are crucial for maintenance of tertiary lymphoid structures in the lung of influenza virus-infected mice. *J Exp Med*, 206(11):2339-2349. <https://doi.org/10.1084/jem.20090410>
- Giraldo NA, Sanchez-Salas R, Peske JD, et al., 2019. The clinical role of the TME in solid cancer. *Br J Cancer*, 120(1):45-53. <https://doi.org/10.1038/s41416-018-0327-z>
- Goc J, Germain C, Vo-Bourgais TKD, et al., 2014. Dendritic cells in tumor-associated tertiary lymphoid structures signal a Th1 cytotoxic immune contexture and license the positive prognostic value of infiltrating CD8<sup>+</sup> T cells. *Cancer Res*, 74(3):705-715. <https://doi.org/10.1158/0008-5472.CAN-13-1342>
- Goenka R, Barnett LG, Silver JS, et al., 2011. Cutting edge: dendritic cell-restricted antigen presentation initiates the follicular helper T cell program but cannot complete ultimate effector differentiation. *J Immunol*, 187(3):1091-1095. <https://doi.org/10.4049/jimmunol.1100853>
- Groeneveld CS, Fontugne J, Cabel L, et al., 2021. Tertiary lymphoid structures marker *CXCL13* is associated with better survival for patients with advanced-stage bladder cancer treated with immunotherapy. *Eur J Cancer*, 148:181-189. <https://doi.org/10.1016/j.ejca.2021.01.036>
- Gu-Trantien C, Loi S, Garaud S, et al., 2013. CD4<sup>+</sup> follicular helper T cell infiltration predicts breast cancer survival. *J Clin Invest*, 123(7):2873-2892. <https://doi.org/10.1172/JCI67428>
- Guedj K, Khallou-Laschet J, Clement M, et al., 2014. M1 macrophages act as LTβR-independent lymphoid tissue inducer cells during atherosclerosis-related lymphoid neogenesis. *Cardiovasc Res*, 101(3):434-443. <https://doi.org/10.1093/cvr/cvt263>
- Gunn MD, Ngo VN, Ansel KM, et al., 1998. A B-cell-homing chemokine made in lymphoid follicles activates Burkitt's lymphoma receptor-1. *Nature*, 391(6669):799-803. <https://doi.org/10.1038/35876>
- Halle S, Dujardin HC, Bakocevic N, et al., 2009. Induced bronchus-associated lymphoid tissue serves as a general priming site for T cells and is maintained by dendritic cells. *J Exp Med*, 206(12):2593-2601. <https://doi.org/10.1084/jem.20091472>
- He JJ, Xiong XX, Yang H, et al., 2022. Defined tumor antigen-specific T cells potentiate personalized TCR-T cell therapy and prediction of immunotherapy response. *Cell Res*, 32(6):530-542. <https://doi.org/10.1038/s41422-022-00627-9>
- Helmink BA, Reddy SM, Gao JJ, et al., 2020. B cells and tertiary lymphoid structures promote immunotherapy response. *Nature*, 577(7791):549-555. <https://doi.org/10.1038/s41586-019-1922-8>
- Hinshaw DC, Shevde LA, 2019. The tumor microenvironment innately modulates cancer progression. *Cancer Res*, 79(18):4557-4566. <https://doi.org/10.1158/0008-5472.CAN-18-3962>
- Hiraoka N, Ino Y, Yamazaki-Itoh R, et al., 2015. Intratumoral tertiary lymphoid organ is a favourable prognosticator in patients with pancreatic cancer. *Br J Cancer*, 112(11):1782-1790. <https://doi.org/10.1038/bjc.2015.145>
- Hodi FS, O'Day SJ, McDermott DF, et al., 2010. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med*, 363(8):711-723. <https://doi.org/10.1056/NEJMoa1003466>
- Hor JL, Whitney PG, Zaid A, et al., 2015. Spatiotemporally distinct interactions with dendritic cell subsets facilitates CD4<sup>+</sup> and CD8<sup>+</sup> T cell activation to localized viral infection. *Immunity*, 43(3):554-565. <https://doi.org/10.1016/j.immuni.2015.07.020>
- Jansen CS, Prokhnevskaya N, Master VA, et al., 2019. An intratumoral niche maintains and differentiates stem-like CD8 T cells. *Nature*, 576(7787):465-470. <https://doi.org/10.1038/s41586-019-1836-5>
- Janssen EM, Lemmens EE, Wolfe T, et al., 2003. CD4<sup>+</sup> T cells are required for secondary expansion and memory in CD8<sup>+</sup> T lymphocytes. *Nature*, 421(6925):852-856. <https://doi.org/10.1038/nature01441>
- Jiang XG, 2020. Lymphatic vasculature in tumor metastasis and immunobiology. *J Zhejiang Univ-Sci B (Biomed & Biotechnol)*, 21(1):3-11. <https://doi.org/10.1631/jzus.B1800633>
- Jin KF, Cao YF, Gu Y, et al., 2021. Poor clinical outcomes and immunoevasive contexture in CXCL13<sup>+</sup>CD8<sup>+</sup> T cells enriched gastric cancer patients. *Oncol Immunology*, 10(1):1915560. <https://doi.org/10.1080/2162402X.2021.1915560>
- Johansson-Percival A, He B, Li ZJ, et al., 2017. De novo induction of intratumoral lymphoid structures and vessel normalization enhances immunotherapy in resistant tumors. *Nat Immunol*, 18(11):1207-1217. <https://doi.org/10.1038/ni.3836>
- Joshi NS, Akama-Garren EH, Lu YS, et al., 2015. Regulatory T cells in tumor-associated tertiary lymphoid structures suppress anti-tumor T cell responses. *Immunity*, 43(3):579-590. <https://doi.org/10.1016/j.immuni.2015.08.006>
- Kato Y, Zaid A, Davey GM, et al., 2015. Targeting antigen to Clec9A primes follicular Th cell memory responses capable of robust recall. *J Immunol*, 195(3):1006-1014. <https://doi.org/10.4049/jimmunol.1500767>
- Kerfoot SM, Yaari G, Patel JR, et al., 2011. Germinal center B cell and T follicular helper cell development initiates in the interfollicular zone. *Immunity*, 34(6):947-960. <https://doi.org/10.1016/j.immuni.2011.03.024>
- Kinker GS, Vitiello GAF, Ferreira WAS, et al., 2021. B cell orchestration of anti-tumor immune responses: a matter of cell localization and communication. *Front Cell Dev Biol*, 9:678127. <https://doi.org/10.3389/fcell.2021.678127>
- Krishnaswamy JK, Gowthaman U, Zhang BY, et al., 2017.

- Migratory CD11b<sup>+</sup> conventional dendritic cells induce T follicular helper cell-dependent antibody responses. *Sci Immunol*, 2(18):eaam9169.  
<https://doi.org/10.1126/sciimmunol.aam9169>
- Kroeger DR, Milne K, Nelson BH, 2016. Tumor-infiltrating plasma cells are associated with tertiary lymphoid structures, cytolytic T-cell responses, and superior prognosis in ovarian cancer. *Clin Cancer Res*, 22(12):3005-3015.  
<https://doi.org/10.1158/1078-0432.CCR-15-2762>
- Lee JM, Lee MH, Garon E, et al., 2017. Phase I trial of intratumoral injection of *CCL21* gene-modified dendritic cells in lung cancer elicits tumor-specific immune responses and CD8<sup>+</sup> T-cell infiltration. *Clin Cancer Res*, 23(16):4556-4568.  
<https://doi.org/10.1158/1078-0432.CCR-16-2821>
- Lee Y, Chin RK, Christiansen P, et al., 2006. Recruitment and activation of naive T cells in the islets by lymphotoxin  $\beta$  receptor-dependent tertiary lymphoid structure. *Immunity*, 25(3):499-509.  
<https://doi.org/10.1016/j.immuni.2006.06.016>
- Lei X, Lei Y, Li JK, et al., 2020. Immune cells within the tumor microenvironment: biological functions and roles in cancer immunotherapy. *Cancer Lett*, 470:126-133.  
<https://doi.org/10.1016/j.canlet.2019.11.009>
- Li HJ, van der Leun AM, Yofe I, et al., 2019. Dysfunctional CD8 T cells form a proliferative, dynamically regulated compartment within human melanoma. *Cell*, 176(4):775-789.e18.  
<https://doi.org/10.1016/j.cell.2018.11.043>
- Li JP, Wu CY, Chen MY, et al., 2021. PD-1<sup>+</sup>CXCR5<sup>+</sup>CD4<sup>+</sup>Th-CXCL13 cell subset drives B cells into tertiary lymphoid structures of nasopharyngeal carcinoma. *J Immunother Cancer*, 9(7):e002101.  
<https://doi.org/10.1136/jitc-2020-002101>
- Lochner M, Ohnmacht C, Presley L, et al., 2011. Microbiota-induced tertiary lymphoid tissues aggravate inflammatory disease in the absence of ROR $\gamma$  T and LTi cells. *J Exp Med*, 208(1):125-134.  
<https://doi.org/10.1084/jem.20100052>
- Luther SA, Lopez T, Bai W, et al., 2000. BLC expression in pancreatic islets causes B cell recruitment and lymphotoxin-dependent lymphoid neogenesis. *Immunity*, 12(5):471-481.  
[https://doi.org/10.1016/s1074-7613\(00\)80199-5](https://doi.org/10.1016/s1074-7613(00)80199-5)
- Luther SA, Bidgol A, Hargreaves DC, et al., 2002. Differing activities of homeostatic chemokines CCL19, CCL21, and CXCL12 in lymphocyte and dendritic cell recruitment and lymphoid neogenesis. *J Immunol*, 169(1):424-433.  
<https://doi.org/10.4049/jimmunol.169.1.424>
- Luther SA, Ansel KM, Cyster JG, 2003. Overlapping roles of CXCL13, interleukin 7 receptor  $\alpha$ , and CCR7 ligands in lymph node development. *J Exp Med*, 197(9):1191-1198.  
<https://doi.org/10.1084/jem.20021294>
- Lutz ER, Wu AA, Bigelow E, et al., 2014. Immunotherapy converts nonimmunogenic pancreatic tumors into immunogenic foci of immune regulation. *Cancer Immunol Res*, 2(7):616-631.  
<https://doi.org/10.1158/2326-6066.CIR-14-0027>
- Maldonado L, Teague JE, Morrow MP, et al., 2014. Intramuscular therapeutic vaccination targeting HPV16 induces T cell responses that localize in mucosal lesions. *Sci Transl Med*, 6(221):221ra213.  
<https://doi.org/10.1126/scitranslmed.3007323>
- Martinet L, Garrido I, Filleron T, et al., 2011. Human solid tumors contain high endothelial venules: association with T- and B-lymphocyte infiltration and favorable prognosis in breast cancer. *Cancer Res*, 71(17):5678-5687.  
<https://doi.org/10.1158/0008-5472.CAN-11-0431>
- Matsubara S, Seki M, Suzuki S, et al., 2019. Tertiary lymphoid organs in the inflammatory myopathy associated with PD-1 inhibitors. *J Immunother Cancer*, 7(1):256.  
<https://doi.org/10.1186/s40425-019-0736-4>
- Mebius RE, Rennert P, Weissman IL, 1997. Developing lymph nodes collect CD4<sup>+</sup>CD3<sup>-</sup>LT $\beta$ <sup>+</sup> cells that can differentiate to APC, NK cells, and follicular cells but not T or B cells. *Immunity*, 7(4):493-504.  
[https://doi.org/10.1016/s1074-7613\(00\)80371-4](https://doi.org/10.1016/s1074-7613(00)80371-4)
- Meylan M, Petitprez F, Becht E, et al., 2022. Tertiary lymphoid structures generate and propagate anti-tumor antibody-producing plasma cells in renal cell cancer. *Immunity*, 55(3):527-541.e5.  
<https://doi.org/10.1016/j.immuni.2022.02.001>
- Morcrette G, Hirsch TZ, Badour E, et al., 2019. APC germline hepatoblastomas demonstrate cisplatin-induced intra-tumor tertiary lymphoid structures. *Oncol Immunology*, 8(6):e1583547.  
<https://doi.org/10.1080/2162402X.2019.1583547>
- Mounzer RH, Svendsen OS, Baluk P, et al., 2010. Lymphotoxin- $\alpha$  contributes to lymphangiogenesis. *Blood*, 116(12):2173-2182.  
<https://doi.org/10.1182/blood-2009-12-256065>
- Munoz-Erazo L, Rhodes JL, Marion VC, et al., 2020. Tertiary lymphoid structures in cancer—considerations for patient prognosis. *Cell Mol Immunol*, 17(6):570-575.  
<https://doi.org/10.1038/s41423-020-0457-0>
- Ngo VN, Korner H, Gunn MD, et al., 1999. Lymphotoxin  $\alpha/\beta$  and tumor necrosis factor are required for stromal cell expression of homing chemokines in B and T cell areas of the spleen. *J Exp Med*, 189(2):403-412.  
<https://doi.org/10.1084/jem.189.2.403>
- Niogret J, Berger H, Rebe C, et al., 2021. Follicular helper-T cells restore CD8<sup>+</sup>-dependent antitumor immunity and anti-PD-L1/PD-1 efficacy. *J Immunother Cancer*, 9(6):e002157.  
<https://doi.org/10.1136/jitc-2020-002157>
- Noël G, Fontsa ML, Garaud S, et al., 2021. Functional Th1-oriented T follicular helper cells that infiltrate human breast cancer promote effective adaptive immunity. *J Clin Invest*, 131(19):e139905.  
<https://doi.org/10.1172/JCI139905>
- Overacre-Delgoffe AE, Bumgarner HJ, Cillo AR, et al., 2021. Microbiota-specific T follicular helper cells drive tertiary lymphoid structures and anti-tumor immunity against colorectal cancer. *Immunity*, 54(12):2812-2824.e4.  
<https://doi.org/10.1016/j.immuni.2021.11.003>
- Pardoll DM, 2012. The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer*, 12(4):252-264.  
<https://doi.org/10.1038/nrc3239>
- Peske JD, Thompson ED, Gemta L, et al., 2015. Effector lymphocyte-induced lymph node-like vasculature enables naive T-cell entry into tumours and enhanced anti-tumour immunity. *Nat Commun*, 6:7114.  
<https://doi.org/10.1038/ncomms8114>
- Peters A, Pitcher LA, Sullivan JM, et al., 2011. Th17 cells induce ectopic lymphoid follicles in central nervous system tissue inflammation. *Immunity*, 35(6):986-996.  
<https://doi.org/10.1016/j.immuni.2011.10.015>

- Petitprez F, de Reyniès A, Keung EZ, et al., 2020. B cells are associated with survival and immunotherapy response in sarcoma. *Nature*, 577(7791):556-560.  
<https://doi.org/10.1038/s41586-019-1906-8>
- Pitzalis C, Jones GW, Bombardieri M, et al., 2014. Ectopic lymphoid-like structures in infection, cancer and autoimmunity. *Nat Rev Immunol*, 14(7):447-462.  
<https://doi.org/10.1038/nri3700>
- Posch F, Silina K, Leibl S, et al., 2018. Maturation of tertiary lymphoid structures and recurrence of stage II and III colorectal cancer. *OncoImmunology*, 7(2):e1378844.  
<https://doi.org/10.1080/2162402X.2017.1378844>
- Ridge JP, di Rosa F, Matzinger P, 1998. A conditioned dendritic cell can be a temporal bridge between a CD4<sup>+</sup> T-helper and a T-killer cell. *Nature*, 393(6684):474-478.  
<https://doi.org/10.1038/30989>
- Rodriguez AB, Peske JD, Woods AN, et al., 2021. Immune mechanisms orchestrate tertiary lymphoid structures in tumors via cancer-associated fibroblasts. *Cell Rep*, 36(3):109422.  
<https://doi.org/10.1016/j.celrep.2021.109422>
- Ruffin AT, Cillo AR, Tabib T, et al., 2021. B cell signatures and tertiary lymphoid structures contribute to outcome in head and neck squamous cell carcinoma. *Nat Commun*, 12:3349.  
<https://doi.org/10.1038/s41467-021-23355-x>
- Sautès-Fridman C, Petitprez F, Calderaro J, et al., 2019. Tertiary lymphoid structures in the era of cancer immunotherapy. *Nat Rev Cancer*, 19(6):307-325.  
<https://doi.org/10.1038/s41568-019-0144-6>
- Schmitt N, Morita R, Bourdery L, et al., 2009. Human dendritic cells induce the differentiation of interleukin-21-producing T follicular helper-like cells through interleukin-12. *Immunity*, 31(1):158-169.  
<https://doi.org/10.1016/j.immuni.2009.04.016>
- Schoenberger SP, Toes REM, van der Voort EIH, et al., 1998. T-cell help for cytotoxic T lymphocytes is mediated by CD40-CD40L interactions. *Nature*, 393(6684):480-483.  
<https://doi.org/10.1038/31002>
- Shalpour S, Lin XJ, Bastian IN, et al., 2017. Inflammation-induced IgA<sup>+</sup> cells dismantle anti-liver cancer immunity. *Nature*, 551(7680):340-345.  
<https://doi.org/10.1038/nature24302>
- Shedlock DJ, Shen H, 2003. Requirement for CD4 T cell help in generating functional CD8 T cell memory. *Science*, 300(5617):337-339.  
<https://doi.org/10.1126/science.1082305>
- Siliņa K, Soltermann A, Attar FM, et al., 2018. Germinal centers determine the prognostic relevance of tertiary lymphoid structures and are impaired by corticosteroids in lung squamous cell carcinoma. *Cancer Res*, 78(5):1308-1320.  
<https://doi.org/10.1158/0008-5472.CAN-17-1987>
- Song IH, Heo SH, Bang WS, et al., 2017. Predictive value of tertiary lymphoid structures assessed by high endothelial venule counts in the neoadjuvant setting of triple-negative breast cancer. *Cancer Res Treat*, 49(2):399-407.  
<https://doi.org/10.4143/crt.2016.215>
- Suzuki G, Sawa H, Kobayashi Y, et al., 1999. Pertussis toxin-sensitive signal controls the trafficking of thymocytes across the corticomedullary junction in the thymus. *J Immunol*, 162(10):5981-5985.
- Thommen DS, Koelzer VH, Herzig P, et al., 2018. A transcriptionally and functionally distinct PD-1<sup>+</sup> CD8<sup>+</sup> T cell pool with predictive potential in non-small-cell lung cancer treated with PD-1 blockade. *Nat Med*, 24(7):994-1004.  
<https://doi.org/10.1038/s41591-018-0057-z>
- Tran E, Robbins PF, Lu YC, et al., 2016. T-cell transfer therapy targeting mutant KRAS in cancer. *N Engl J Med*, 375(23):2255-2262.  
<https://doi.org/10.1056/NEJMoa1609279>
- van de Pavert SA, Olivier BJ, Goverse G, et al., 2009. Chemokine CXCL13 is essential for lymph node initiation and is induced by retinoic acid and neuronal stimulation. *Nat Immunol*, 10(11):1193-1199.  
<https://doi.org/10.1038/ni.1789>
- van Dijk N, Gil-Jimenez A, Silina K, et al., 2020. Preoperative ipilimumab plus nivolumab in locoregionally advanced urothelial cancer: the NABUCCO trial. *Nat Med*, 26(12):1839-1844.  
<https://doi.org/10.1038/s41591-020-1085-z>
- Veiga-Fernandes H, Coles MC, Foster KE, et al., 2007. Tyrosine kinase receptor RET is a key regulator of Peyer's Patch organogenesis. *Nature*, 446(7135):547-551.  
<https://doi.org/10.1038/nature05597>
- Weinstein AM, Chen L, Brzana EA, et al., 2017. Tbet and IL-36γ cooperate in therapeutic DC-mediated promotion of ectopic lymphoid organogenesis in the tumor microenvironment. *OncoImmunology*, 6(6):e1322238.  
<https://doi.org/10.1080/2162402X.2017.1322238>
- Workeel HH, Lubbers JM, Arnold R, et al., 2019. A transcriptionally distinct CXCL13<sup>+</sup>CD103<sup>+</sup>CD8<sup>+</sup> T-cell population is associated with B-cell recruitment and neoantigen load in human cancer. *Cancer Immunol Res*, 7(5):784-796.  
<https://doi.org/10.1158/2326-6066.CIR-18-0517>
- Wouters MCA, Nelson BH, 2018. Prognostic significance of tumor-infiltrating B cells and plasma cells in human cancer. *Clin Cancer Res*, 24(24):6125-6135.  
<https://doi.org/10.1158/1078-0432.CCR-18-1481>
- Wu SZ, Roden DL, Wang CF, et al., 2020. Stromal cell diversity associated with immune evasion in human triple-negative breast cancer. *EMBO J*, 39(19):e104063.  
<https://doi.org/10.15252/embj.2019104063>
- Wykes M, Pombo A, Jenkins C, et al., 1998. Dendritic cells interact directly with naive B lymphocytes to transfer antigen and initiate class switching in a primary T-dependent response. *J Immunol*, 161(3):1313-1319.
- Yang SC, Batra RK, Hillinger S, et al., 2006. Intrapulmonary administration of CCL21 gene-modified dendritic cells reduces tumor burden in spontaneous murine bronchoalveolar cell carcinoma. *Cancer Res*, 66(6):3205-3213.  
<https://doi.org/10.1158/0008-5472.CAN-05-3619>
- Yeh CH, Finney J, Okada T, et al., 2022. Primary germinal center-resident T follicular helper cells are a physiologically distinct subset of CXCR5<sup>hi</sup>PD-1<sup>hi</sup> T follicular helper cells. *Immunity*, 55(2):272-289.e7.  
<https://doi.org/10.1016/j.immuni.2021.12.015>
- Yoshida H, Naito A, Inoue J, et al., 2002. Different cytokines induce surface lymphotoxin-αβ on IL-7 receptor-α cells that differentially engender lymph nodes and Peyer's patches. *Immunity*, 17(6):823-833.  
[https://doi.org/10.1016/s1074-7613\(02\)00479-x](https://doi.org/10.1016/s1074-7613(02)00479-x)