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Review

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Protein structure-based *in-silico* approaches to drug discovery: Guide to COVID-19 therapeutics

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ABSTRACT

With more than 5 million fatalities and close to 300 million reported cases, COVID-19 is the first documented pandemic due to a coronavirus that continues to be a major health challenge. Despite being rapid, uncontrollable, and highly infectious in its spread, it also created incentives for technology development and redefined public health needs and research agendas to fast-track innovations to be translated. Breakthroughs in computational biology peaked during the pandemic with renewed attention to making all cutting-edge technology deliver agents to combat the disease. The demand to develop effective treatments yielded surprising collaborations from previously segregated fields of science and technology. The long-standing pharmaceutical industry's aversion to repurposing existing drugs due to a lack of exponential financial gain was overrun by the health crisis and pressures created by front-line researchers and providers. Effective vaccine development even at an unprecedented pace took more than a year to develop and commence trials. Now the emergence of variants and waning protections during the booster shots is resulting in breakthrough infections that continue to strain health care systems. As of now, every protein of SARS-CoV-2 has been structurally characterized and related host pathways have been extensively mapped out. The research community has addressed the druggability of a multitude of possible targets. This has been made possible due to existing technology for virtual computerassisted drug development as well as new tools and technologies such as artificial intelligence to deliver new leads. Here in this article, we are discussing advances in the drug discovery field related to target-based drug discovery and exploring the implications of known target-specific agents on COVID-19 therapeutic management. The current scenario calls for more personalized medicine efforts and stratifying patient populations early on for their need for different combinations of prognosis-specific therapeutics. We intend to highlight target hotspots and their potential agents, with the ultimate goal of using rational design of new therapeutics to not only end this pandemic but also uncover a generalizable platform for use in future pandemics.

1. Introduction

Since the beginning of the COVID-19 pandemic, which is caused by SARS-CoV-2, there has been an impending question 'what can be the standard course of therapy, and which agents need to be trialed. The first year of the pandemic followed Murphy's Law [\(Bloch, 2003](#page-17-0)) with the ensuing chaos causing severe mortality rates due to a lack of population immunity and the use of ineffective interventions. The rapid global

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spread of the disease overwhelmed medical care systems due to exponential regional surges. As of July 4th[,] 2022, the pandemic has claimed 6.35 million lives worldwide and caused over 0.5 billion cases of infection ("[WHO Coronavirus Disease \(COVID-19\) Dashboard,](#page-24-0)" n.d.). The USA has been the worst hit with more than a million deaths out of 87.5 million cases ([Dong et al., 2020](#page-19-0); [Ruhm, 2022\)](#page-23-0). The surge in cases is often at an intensity that its severity is made worse by a shortage of medical resources. This has stymied trials conducted for several agents ([Robinson et al., 2022\)](#page-23-0). Many promising initial reports of therapeutic approaches became proven failures, and yet they often were needlessly *trialed* repeatedly by different groups. Hampering effective therapeutic development, the rush to trials often fell short in the number of patients recruited. This under empowerment and the varying degree of symptom sets leads to prognosis and therapeutic response variability which makes it difficult to stratify patient populations. This was further exacerbated by the changing pathophysiology caused by newer variants, which combined with the evolving self-medication landscape, resulted in inconsistent trial data for some agents and ultimately unreliable outcome results [\(Watson, 2022](#page-24-0)). Prohibitive costs of newer drugs, as well as antibody therapies, have generated worldwide interest in trying a variety of agents to reduce the severity of COVID-19 infection. For instance, preliminary evidence suggested that hydroxychloroquine (HQ) therapy can reduce viral load [\(Gautret et al., 2020](#page-19-0)). However, a recent meta-analysis of multiple trial data has now concluded that although HQ therapy is safe at the trial doses used, it remains ineffective in reducing mortality and severity of disease (T. [Gupta et al., 2022](#page-20-0)). Conversely, other trials have shown more promising results, such as the use of Oseltamivir (Theraflu), which statistically demonstrated to reduce mortality in COVID-19 patients ([Zendehdel et al., 2022\)](#page-24-0). Additionally, various comorbidities like old age, diabetes, obesity, hypertension, and the immunocompromised state contribute to COVID-19 mortality, their associations are still not enough to stratify patients and take universal prophylactic measures ([Gentile and Schiano Moriello, 2022](#page-19-0)) and as a result, new therapeutic interventions remain in high demand.

Computational structural biology is a interdisciplinary field performed on computer or via computer simulation that encompasses the theory and application of approaches to model, predict, and explain biological function at the molecular level, well-known as *in silico* experiment. Proteins are flexible molecules that undergo conformational changes (such as folding and unfolding or domain motions) as part of their interactions with other biopolymers as partners or drug molecules. Conformational changes of the proteins might reflect a closed, open, or intermediate states and this dynamical aspect plays a critical role in drug discovery. Nowadays, molecular dynamics makes it possible to simulate these conformational changes with a timescale ranging from nanoseconds to microseconds of time. Molecular dynamics simulations is a computer (*in silico*) technique that makes it possible to predict how a system will evolve over time and, consequently, to predict the movement of the molecules in the system. *In silico* methods (molecular modeling, molecular docking or screening, molecular dynamics, etc) could be used to efficiently identify and design drug candidates, to study their interactions with their targets. The Nobel Prize in Chemistry 2013 has been awarded to Martin Karplus, Michael Levitt and Arieh Warshel for development of multiscale models of complex chemical systems as computational techniques for structural biology [\(https://](https://www.nobelprize.org) [www.nobelprize.org\)](https://www.nobelprize.org).

In silico drug discovery has proved to be instrumental in suggesting numerous agents and many of the predicted agents have been used to manage COVID-19. It has been a long-standing principle that the fixed 3D structure of protein dictated by amino acid composition is the basis for assigning function. There have been exceptions to this principle in multiple instances when proteins have multiple structures owing to disordered regions [\(Anjum et al., 2022](#page-17-0); Prateek [Kumar et al., 2022a](#page-21-0); J. [Zhang et al., 2022\)](#page-25-0). This is more evident in RNA viral proteomes due to a higher rate of mutations and a protein often has more than one function. For instance, PLpro is a protease and a deubiquitinase, all of which are

important for viral envelope formation, and their functional activities are associated with inflammasome formation in infected cells ([Lewis](#page-21-0) [et al., 2022;](#page-21-0) J. [Zhang et al., 2022](#page-25-0)). Such redundancy, size limitations, and genetic instabilities call for highly flexible proteins which are generally seen in the experimentally solved crystal structure, their variabilities in viral proteins in the form of multiple 'states' and confirmations [\(Fornasier et al., 2022](#page-19-0); [Siragusa et al., 2022](#page-23-0)). As starting crystal structure is the bottleneck of any virtual screening effort, this variability led to numerous 'false' hits that had no agreement between binding prediction and biological activity [\(Martin et al., 2020\)](#page-22-0). Like all the other fields, the field of computational biology methods also had multiple breakthroughs which now have more applications than just COVID-19 drug discovery research. Additionally, we now have AI predictions for the shape of nearly every known protein, which can be structurally complementary to drug discovery ([Callaway, 2022\)](#page-18-0). Many laboratories have been pioneering novel technologies in the machine learning, AI, and conformational dynamics space [\(Caulfield and Medina-Franco,](#page-18-0) [2011;](#page-18-0) [Coban et al., 2020,](#page-18-0) [2021a](#page-18-0), [2021b;](#page-18-0) [Hines et al., 2019a,](#page-20-0) [2019b](#page-20-0); [Kayode et al., 2016](#page-21-0); [Puschmann et al., 2017](#page-22-0); [Savytskyi et al., 2013\)](#page-23-0).

Recently, the anti-cancer drug Pralatrexate was discovered to have *in vitro* EC50 values of 0.008 μM. While being a strong immunosuppressant it's usability in COVID-19 is highly debatable the pipeline that delivered this compound comprised of deep learning models and force field dynamics simulations [\(Zhang et al., 2020\)](#page-25-0). With newer and faster methods made available there are multiple methods producing a similar pipeline ([Rapicavoli et al., 2022](#page-22-0); [Zhang et al., 2022\)](#page-25-0). Free energy perturbation calculations enabled Zhang et al., in 2022 to improve main protease Triarylpyridinone inhibitors to have EC50 values as low as 0.080 μM ([Ramos et al. 1987](#page-22-0)).

In this review, we try to boil down protein-inhibitor relationships that have been exploited as anti-COVID-19 therapeutics or have a high validated potential for the same. Such information should be used to steer the computational learning approaches through AI to understand why these work and others don't despite having positive classic predicted interactions. Additionally, we provide a comprehensive analysis of existing, approved, and experimental therapeutics with their mechanism of action against either the viral or host protein targets.

2. Drugging COVID-19: what constitutes a "good" drug?

There have been some controversial agents that have undergone trials against SARS-CoV-2 due to some *in vitro* reports or proposed mechanisms of action [\(Ivanova et al., 2022](#page-20-0)). Many of these agents did not have a consistent effect and had surprising side effects such as QT prolongations (abnormal heart rhythms and sudden cardiac arrest) e.g Chloroquine and Hydroxychloroquine [\(Table 4\)](#page-15-0) ([Deng et al., 2022](#page-18-0)). While others were not fully effective at tolerable doses e.g. Ivermectin ([Hariyanto et al., 2022\)](#page-20-0), some were mildly effective even though they had no interaction with SARS-CoV-2 targets, e.g. oseltamivir ([Zendehdel](#page-24-0) [et al., 2022\)](#page-24-0). Some were highly dangerous, especially with the misinformation inspired panicked patient self-medications e.g. Chlorine Dioxide ([Chejfec-Ciociano et al., 2022\)](#page-18-0). Since the beginning of the pandemic, Ibuprofen was contraindicated as it is known to increase ACE2 receptor expression in the cells exacerbating viral infectiousness. However, there was widespread use of nebulized ibuprofen (NaIHS) as a wonder cure and reported to be highly effective, had negative correlations and so-called positive effects were probably due to concomitant aggressive corticosteroid therapy [\(Calonico et al., 2022\)](#page-18-0). As a result, there is a need to understand both classical drug targets and other modalities that may be therapeutic.

3. Techniques for elucidation of drug-target interaction and efficacy

One of the foundations of drug design is to utilize a molecular model of druggable targets. Today's drug discovery labs can draw from a multitude of techniques for determining experimental structures, yet the different techniques have their strengths and weaknesses. For example, membrane proteins are notoriously difficult to crystallize, so the gold standard x-ray crystallography is generally not successful. Typically, cryo-EM is utilized for large proteins/complexes, such as membrane proteins. The caveat here is that cryo-EM is in general a lower resolution technique and may bias conformations because of the airwater interface. A relatively new structural technique is x-ray freeelectron laser (XFEL), coupled with lipid-cubic phase crystallization ([Ono et al., 2022\)](#page-22-0). Essentially, this consists of growing small crystals in a lipidic environment that is more amenable for membrane proteins, which are then injected at random orientations and illuminated with extremely brilliant x-ray photons to generate diffraction patterns. This has been successfully applied to a variety of membrane proteins recently, though not as yet any COVID-19-related target; however, this technique has potential application in the field as shown with other viruses [\(Townsend et al., 2021\)](#page-24-0). Proteases and kinase inhibitors have traditionally held roles as drugs of choice for inhibiting virion production ([Bain et al., 2003;](#page-17-0) [Mahdi et al., 2020;](#page-21-0) [Pearlman, 2012](#page-22-0); "[Protein](#page-22-0) [Kinase Inhibitors,](#page-22-0)" 2012; [Zhou et al., 2015\)](#page-25-0), however, in recent times the shift to virus centric proteins has made progress [\(Chakraborty et al.,](#page-18-0) [2021; Dai et al., 2020;](#page-18-0) [Narayanan et al., 2022](#page-22-0); [Prajapat et al., 2020;](#page-22-0) [Y.-X.](#page-25-0) [Zheng et al., 2021\)](#page-25-0). Added to these new targets has been the implementation of new computational tools to more quickly address the urgency of the need ([Callaway, 2022; Coban et al., 2021b\)](#page-18-0).

4. Enter the era of the machine: learning to use algorithmguided drug design

The complex multivariate approaches to drug modeling on a molecular structure are well suited to the application of machine learning (ML) techniques. Generative chemistry is at the forefront of new medicinal chemistry design workflows, where the implementation of layered data with context to various data sources allows us to integrate complex datasets into the framework of a deep learning or machinebased intelligence that can find associations otherwise not possible. Both ML and artificial intelligence (AI) are being applied to many areas of biological research. With respect to COVID-19, ML has been used to help screen drug targets, druggable sites on the targets, drugs, and drugtarget interactions ([El-Behery et al., 2021\)](#page-19-0). This has led to the repurposing of drugs that are already FDA-approved for COVID-19 therapy, the discovery of novel molecules as potential drugs, and the identification of cryptic binding pockets introduced by virus/host protein-protein interaction [\(Dang and Song, 2022](#page-18-0)). In addition, ML has been used to mine bioinformatics data and analyze biological pathways to identify novel pathways that can lead to a greater understanding of the disease mechanism, as well as detect additional points of intervention ([Auwul](#page-17-0) [et al., 2021](#page-17-0)). AI has assisted in the analysis of samples to help make rapid diagnoses with a less expensive assay that is highly sensitive, selective, and accurate [\(Jaroenram et al., 2022](#page-20-0); [Lai et al., 2022](#page-21-0)). The method works by employing two pH-dependent dyes and a reverse transcription loop-mediated isothermal amplification (RT-LAMP) assay; the colorimetric readout data was used to train an algorithm for classification i.e. diagnosis of positive or negative infection status. Other uses of ML related to COVID-19 are the large-scale screening for anti-COVID-19 biomolecules in foods ([Laponogov et al., 2021](#page-21-0)). The study used a similar approach to standard drug screening but started with a database of food-based bioactive molecules; they identified 52 molecules predicted to disrupt the COVID-19-host interactome. Engaging in multiple treatment paradigms is beneficial in that it increases the likelihood of therapeutic benefit to the patient, decreases the chance of the virus developing resistance, and can reduce dosing to limit adverse side effects. Interruption of COVID-19 progression with multi-drug therapy looking for synergetic effect with computational biology for high-throughput screening has been successful ([Coban et al., 2021b](#page-18-0)), which has the capability of using mixed algorithms to examine the

impact of structural changes. As a result, the application of ML and AI techniques is expected to yield rapid progress in the discovery of new candidates for antiviral use.

5. In *silico* **deduced target-specific leads that reached clinical trials**

Favipiravir is a purine analog that is a potent RNA-dependent RNA polymerase (RdRp) inhibitor initially selected on basis of similarities with known target EBOLA RdRp ([da Silva et al., 2022;](#page-18-0) [Mashayekhi--](#page-22-0)[Sardoo and Hosseinjani, 2022\)](#page-22-0). Favipiravir showed a 62.8% viral clearance in 4 days compared to untreated ([Ivashchenko et al., 2021](#page-20-0)). While favipiravir has little effect on nonhospitalized patients, its use among hospitalized patients has led to faster viral clearance and better radiological imaging endpoints in multiple trials [\(Hung et al., 2022](#page-20-0)). With upcoming reports of long-term lung damage in both hospitalized and nonhospitalized patients [\(C. Wang et al., 2022;](#page-24-0) J. [Yu et al., 2022](#page-24-0)), there is a need for a retrospective follow-up trial needed to assess favipiravir's long-term benefits. Icatibant is a known bradykinin type 2 receptor antagonist that was computationally predicted to target the SARS-CoV-2 main protease [\(Liu and Wang, 2020\)](#page-21-0). However, the clinical trial (NCT04978051) results were inconclusive [\(Malchair et al.,](#page-21-0) [2022\)](#page-21-0) and there is no target-specific inhibition data available. Lopinavir & Ritonavir are other predicted inhibitors of 3CLpro ([Reina and Iglesias,](#page-23-0) [2022\)](#page-23-0), however, numerous clinical trials have failed to establish their clinical usefulness as anti-COVID-19 medications ([Cao et al., 2020](#page-18-0); [Sheahan et al., 2020\)](#page-23-0). PF-07321332 (nirmatrelvir) a rationally improved second-generation frontrunning drug from Pfizer is in the Phase3 clinical trial, It targets 3CLpro and thereby inhibits viral replication [\(Vandyck and Deval, 2021\)](#page-24-0). Ciclosporin/Cyclosporine immunomodulatory drug is a calcineurin inhibitor that was discovered through computational host interactome modeling for the SARS virus (SAR-S-CoV) ([Pfefferle et al., 2011](#page-22-0)) and was predicted to have a positive effect on COVID-19 through immunosuppression ([Ellinger et al., 2021](#page-19-0)). Further, it was found to have antiviral activity *in vitro* (Dittmar et al., [2021\)](#page-19-0). Later HR (hazard ratio) improvement value of 2.15 was observed in a combination trial with a low dose of steroid ([Galvez-Romero et al.,](#page-19-0) [2021\)](#page-19-0) and was an efficacious treatment option in the COQUIMA cohort ([Schuurmans and Hage, 2021](#page-23-0)) and multiple variants [\(Fenizia et al.,](#page-19-0) [2022\)](#page-19-0). Another 3CLpro inhibitor, found through *in silico* screenings was Cepharanthine (CEP), a small phyto-alkaloid obtained from the *Stephania cepharantha*. CEP had IC₅₀ of 1.90 μm [\(Hijikata et al., 2022\)](#page-20-0) against the Wuhan strain (wild type) and consistent activity against three other VOCs (Prabhakaran [Kumar et al., 2022](#page-21-0)). It's a promising anti-COVID-19 candidate in animal testing offering significant protection from lung fibrosis in bleomycin (BLM)-challenged rats [\(Li et al.,](#page-21-0) [2022\)](#page-21-0).

6. Cytotherapy

Cellular therapies have been proven to protect immunosuppressed patients (*>*20% mortality rate) by providing anti-viral cellular immunity and immune modulation for vulnerable patient populations ([Far](#page-19-0)[hangnia et al., 2022](#page-19-0); [Verma et al., 2022](#page-24-0)). Different trials with SARS-CoV-2 specific T-cell trials (allogeneic CSTs familial or HLA matched), Natural killer (NK) cell (e.g. FT516 cells), Tregs (T regulatory cell), and Mesenchymal Stem Cell Infusion or Stem Cell Products have shown therapeutic potential comparable to available antiviral therapies ([Conway et al., 2022\)](#page-18-0). With a longer lifespan of T-cells, there is longer-lasting protection than humoral immunity.

7. Biological activities of SARS-CoV-2 components as potential therapeutic targets

A wide variety of targets are addressable for attenuating the infection progression of SARS-CoV-2 as depicted in [Fig. 1.](#page-4-0) As previously mentioned, therapeutics active on some of these targets are now in clinical trials. Yet many more Non-Structural Protein (NSP) targets have been identified and are in various stages of development ([Table 1\)](#page-5-0). In this review, we will address both classical drug targets (enzymatic vs non-enzymatic) and new modalities for possible use as COVID therapies.

8. NSP enzymatic targets

8.1. Main Peptidase

The 3CLpro/Mpro gene is the Main Peptidase of SARS coronavirus and is responsible for \sim 11 cleavage sites in viral propeptide. As a result, it is an essential target for both viral replicase as well as structural assembly for completing the viral cycle ([Gupta et al., 2021b\)](#page-20-0). This 306 amino acid long protease has a catalytic core with C145 and H41 and is highly conserved among variants to preserve essential function [\(Gupta](#page-20-0) [et al., 2021a\)](#page-20-0) but also has multiple conformation states making drug targeting difficult [\(Savytskyi and Kornelyuk, 2022\)](#page-23-0). The most recent PF-07321332 (**nirmatrelvir**) is a Pfizer anti-SARS-CoV-2 compound targeting 3CLpro [\(Reina and Iglesias, 2022](#page-23-0)). In combination with ritonavir, a xenobiotic degradation reducing agent for PF-07321332 ([Lamb,](#page-21-0) [2022\)](#page-21-0), the drug combination has shown a strong efficacy across multiple SARS-CoV-2 variants ([Ullrich et al., 2022](#page-24-0)). Additional research on combinations with other antiviral agents targeting different components (e.g. Monupiravir/remdesivir for RdRp) is ongoing ([Table 1](#page-5-0)). Earlier, *in silico* predictions discovered a 3CLpro inhibitor, Atazanavir, that was later shown to block viral replication [\(Fintelman-Rodrigues et al., 2020\)](#page-19-0)

and showed positive outcomes in various trials ([Kalantari et al., 2021](#page-21-0)). However, due to many side effects such as hepatotoxicity, Atazanavir failed to be a drug of choice in the long run ([Mazaherpour et al., 2021](#page-22-0)). Daclatasvir is a well-accepted HCV therapeutic and its combination with sofosbuvir is well tolerated and efficacious [\(Merat, 2020\)](#page-22-0). While both Daclatasvir and sofosbuvir had anti-SARS-CoV-2 activity, the combination showed inconsistent results in different trials but had an overall positive effect ([Chan et al., 2021, p. 2\)](#page-18-0). Another anti-HCV protease inhibitor Danoprevir showed some efficacy in initial trials ([H. Chen et al.,](#page-18-0) [2020\)](#page-18-0) but was abandoned in Phase 4 trials (NCT04345276).

8.2. Papain-like proteinases

Papain-like viral protease (Plpro) is named NSP 3 and is a versatile enzyme that processes the viral polypeptide into functional proteins similar to 3CLpro but has Catalytic triad C111, H272, and D286 which is also highly conserved ([Fu et al., 2021\)](#page-19-0). While activating it also protects viral peptides being attacked by host proteasome machinery and de-ubiqutinylase Lys-linked polyUb chains ([Lewis et al., 2022](#page-21-0)). Although a potential therapeutic target, drugs blocking Plpro have yet to be identified.

8.3. RNA-dependent RNA polymerase

Viral RNA-dependent RNA polymerase (RdRp) is identified in the SARS-CoV-2 genome as the NSP 12, It's part of a large replicase complex carrying out RNA replication. This protein class has been a highly

Fig. 1. Schematic depiction of different SARS-CoV-2 proteome (ORF map) coded targets(3D ribbons or cartoons) involved in different steps of viral replication (labeled blue) and various example inhibitors (labeled red). The infection cycle starts when the SARS-CoV-2 Spike protein binds to a Human receptor followed by either viral-host cell fusion (1a) or endocytosis (1b). Fusion directly allows the viral RNA to enter the host cell (2), The large viral script is known to encode 29 viral proteins (3), A viral-specific translation yields two replicase polyproteins, pp1a and pp1ab, and many small ORFs(4). The two major polyproteins are processed by two proteases, PLpro and 3CLpro(5), generating 16 NSPs. ExoN possesses a viral exoribonuclease activity (9). Viral Helicase plays a critical role in viral replication by unwinding dsRNA formed during replication as well as tertiary structures of genomic RNA. (7). The enzyme 2'-O-MT methylates the viral 2' end which is important for selective translation and protection from host RNA degradation (8). RdRP along with different NSPs is involved in viral-host cell replication through catalyzing template synthesis of polynucleotides in the 5'-3' direction (7). NendoU is an Mn²⁺ dependent hexamer (dimer of trimer) enzyme responsible for protein interference with the innate immune system. For viral assembly of structural proteins (S, E, and M) in the endoplasmic reticulum, along with the N protein is combined with the (+) gRNA to become a compact helical nucleoprotein complex(10). They assemble to form a virus particle in the endoplasmic reticulum-Golgi apparatus compartment and are then excreted from the cell through budding mediated by the fusion of smooth-walled vesicles to the plasma membrane (11–12).

Table 1

In vitro validated anti-SARS-CoV-2 agents reported with a known target.

Table 1 (*continued*)

exploited target in several RNA viruses and the resulting inhibitors have served as a rich pool for many repurposable antivirals [\(Abolhassani](#page-17-0) [et al., 2021](#page-17-0)). While all the natural variants in SARS-CoV-2 are highly susceptible to remdesivir ([Pitts et al., 2022\)](#page-22-0), studies have shown the possibility of mutational resistance which is contraindicated for mono-therapy ([Stevens et al., 2022](#page-23-0)). Azvudine is a 4'-Modified Nucleoside and a potent anti-HIV drug candidate ([Chang, 2022\)](#page-18-0). Early trials showed Azvudine as a promising anti-COVID-19 agent with evident shortening of nucleic acid negative conversion [\(Ren et al., 2020\)](#page-23-0), but it has only been regionally approved as an anti-HIV therapeutic in China and has not been trialed elsewhere. AT527 (RO-7496998) *a.k.a.* bemnifosbuvir is an oral purine nucleotide prodrug that has potent *in vitro* antiviral activity SARS-CoV-2 ([Shannon et al., 2022](#page-23-0)) and has also shown a shortening of disease tenure in early trials [\(Good et al., 2021](#page-20-0)). Clevudine a pyrimidine analog is an anti-HBV drug that underwent a trial in the Korean republic but was grossly ineffective [\(Song et al., 2021](#page-23-0)). Sofosbuvir (PSI-7977), an approved anti-HCV phosphoramidite prodrug ([Messina et al., 2022\)](#page-22-0), is a treatment that has been shown to reduce mortality and improve associated clinical outcomes in patients with COVID-19 ([Hsu et al., 2022](#page-20-0)). Molnupiravir is a prodrug and it is hydrolyzed by esterases to form intermediate ribonucleoside N-hydroxycytidine (NHC) which is further phosphorylated intracellularly yielding active agent NHC triphosphate (NHC-TP) ([Cox et al., 2021](#page-18-0); [Wahl et al., 2021](#page-24-0)). It is a well-tolerated and highly effective anti-COVID-19 treatment owing to its high bioavailability ([Table .2\)](#page-8-0) ([Caraco et al., 2022](#page-18-0); [Jayk Bernal et al., 2022](#page-20-0); [Whitley, 2022](#page-24-0)).

8.4. Helicase

NSP 13 is an ATP-dependent Helicase with a 5'to 3′ polarity acting on either double-stranded RNA/DNA [\(Shu et al., 2020](#page-23-0)). Among all SARS-CoV-2 NSPs, Helicase is the most conserved among different beta coronavirus species ([Jang et al., 2020\)](#page-20-0). There are reports of helicase forming a complex with RdRp/replicase complex suggesting a role in proofreading during RNA replication ([J. Chen et al., 2020\)](#page-18-0). Also, there are isolated reports of helicase affecting infected cell interferon (IFN) signaling to neighboring healthy cells by altering JAK1 phosphorylation of SAT1 ([Fung et al., 2022\)](#page-19-0). While there were multiple helicase inhibitors discovered against SARS and MERS there were not many interesting leads for SARS-CoV-2 despite high sequence similarity ([Cimolai, 2020\)](#page-18-0) except amantadine or memantine that have been shown in isolated reports to be effective in COVID-19 with neurological symptoms ([Rejdak and Grieb, 2020](#page-23-0)). Ranitidine bismuth citrate also targets helicases and was initially shown to be highly effective in protecting Syrian hamster COVID-19 animal models [\(Yuan et al., 2020](#page-24-0)). While SARS-CoV-2 helicase is highly susceptible to bismuth salts, which

are accepted to be the primary mechanism ([Shu et al., 2020](#page-23-0)), zinc chelation ([Zamai, 2021, p. 20](#page-24-0)) and allosteric main protease inhibition ([Tao et al., 2021](#page-23-0)) additional mechanisms in play. A pilot study showed that 50% of patients receiving Bismuth Subsalicylate (BSS) became RT-PCR negative, however, authors state issues with dosage and bioavailability.

8.5. NendoU

NSP 15 is a uridylate-specific endoribonuclease (NendoU) that exists as a homo hexamer [\(Tran et al., 2022](#page-24-0)). While NendoU is highly conserved among most of the nidoviruses, especially vertebrates infecting coronaviruses, its knockouts are known to replicate at par with wild types ([Grellet et al., 2022\)](#page-20-0). The role of NendoU is to protect viral RNA from host intracellular defenses [\(Boodhoo et al., 2022](#page-18-0)). A few of the known corticosteroids can inhibit SARS/MERS *in vitro* and were also reported to have potent activity against SARS-CoV-2 with IC_{50} s niclosamide (0.28 μM), ciclesonide (4.33 μM), and tilorone (4 μM) (Ko et al., [2021\)](#page-21-0). Ciclesonide has been shown to lose antiviral activity on MERS-Nendou mutants ([Matsuyama et al., 2020](#page-22-0)). While Ciclesonide has been part of many therapeutic combinations, there have been a few focused monotherapy randomized trials with inhaled formulations that have resulted in lower hospitalizations and reduced respiratory symptoms in treated patients [\(Clemency et al., 2022;](#page-18-0) [Ezer et al., 2021](#page-19-0)). Ciclesonide is of particular interest for long-haul patient management for preventing severe lung damage [\(Ruggiero et al., 2022\)](#page-23-0). Exebryl-1 a known β-amyloid anti-aggregation molecule (Alzheimer's therapy) was shown to have consistent antiviral activity between 10 and 66 μM, in various cell lines and was discovered through high throughput screens ([Choi et al., 2021](#page-18-0)). Exebryl-1 has been shown to disturb hexamerization of NendoU critical for its activity [\(Tran et al., 2022](#page-24-0)). So far there are no trials with Exebryl-1 against COVID-19, but negative drug interactions with COVID-19 medications with Alzheimer's disease does suggest a utility for this repurposable agent [\(Balli et al., 2020\)](#page-17-0).

8.6. Other targets

ADP ribose phosphatase (NSP3) is another interesting target playing a role in cellular immune evasion by SARS-CoV-2 by resisting ADPribosylation of host proteins induced by IFN [\(Russo et al., 2021](#page-23-0)). Exoribonuclease (ExoN, NSP14) is a 5'-to-3' exonuclease and has been the focus of many computational drug screening pipelines ([Castillo-Garit](#page-18-0) [et al., 2021](#page-18-0); [Gupta et al., 2021b](#page-20-0)). ExoN is inhibited by S-adenosylhomocysteine [\(Riccio et al., 2022](#page-23-0)) which is a marker for severe COVID-19 ([Ponti et al., 2021](#page-22-0)) and its abundance may have been protecting liver cholangiocytes expressing ACE-2. NSP16 is another critical target which

Table 2

Descriptions of anti-viral agents from clinical trials.

(*continued on next page*)

infiltrates in some participants with

Table 2 (*continued*)

is an Mn^{2+} dependant putative 2'-O-methyl transferase that forms a heterodimer with NSP10 ([Minasov et al., 2021\)](#page-22-0).

9. Non-enzymatic targets

9.1. 3a Ion channel

ORF3a encodes an accessory protein that forms $K+$ channels that trigger NLRP3 activation resulting in the maturation of IL-1β and cleavage/activation of Gasdermin via NFκB ([Kern et al., n.d.](#page-21-0); J. [Zhang](#page-25-0) [et al., 2022\)](#page-25-0). ORF3a is susceptible to amantadine [\(Toft-Bertelsen et al.,](#page-24-0) [2021\)](#page-24-0) which has been shown to improve patient conditions suffering from COVID-19-Related Diffuse Leukoencephalopathy ([Lam et al.,](#page-21-0) [2022\)](#page-21-0). In a larger trial with co-morbidities in Parkinson's and multiple sclerosis patients already receiving amantadine, there was significant prevention of COVID-19 infection [\(Kamel et al., 2021\)](#page-21-0). A larger trial is in progress and its results are awaited [\(Rejdak and Grieb, 2020](#page-23-0)). [Tomar](#page-24-0) [et al., 2021](#page-24-0) reported many more FDA-approved drugs with significant *in vitro* activity against heterologously expressed 3a Ion channel; Plerixafor, Kasugamycin, Capreomycin, Pentamidine, Spectinomycin, Flumatinib, Darapladib, Floxuridine, and Fludarabine ([Tomar et al., 2021](#page-24-0), [2021\)](#page-24-0).

9.2. Non-structural protein 1

NSP-1 is the host shutoff factor that halts the translational machinery of SARS-CoV-2 infected cells by binding with the mRNA channel within the ribosome [\(Simeoni et al., 2021](#page-23-0)). The main c-terminal domain playing a role in the ribosome binding can be blocked by Mitoxantrone hydrochloride (Novantrone) (Prateek [Kumar et al., 2022b\)](#page-21-0). Notably, Mitoxantrone HCL also blocks viral entry through perturbing spike-heparan sulfate interactions (Q. [Zhang et al., 2022](#page-25-0)).

9.3. Other SARS-CoV-2 targets

NTD-N-protein or N terminal domain of Nucleocapsid protein is responsible for binding and thereby assembling the RNA genome of SARS-CoV-2 [\(Ye et al., 2020\)](#page-24-0). Recently multiple *in vitro* anti-*SARS-CoV-2* molecules were discovered as interacting with the NTD-N-protein through isothermal titration calorimetry with EC₅₀s: Telmisartan (1.02 μM), Bictegravir (8.11 μM), Bisdemethoxycurcumin (1.64 μM), and MCC-555 (4.26 μM) ([Dhaka et al., 2022](#page-19-0)). Additional targets have been proposed and investigated as drug targets *in silico*. NSP2 is involved in host signaling interferences, NSP3 mediates a bipartite shift of host translational machinery to translate viral RNA only, NSP4 plays a role in the replicase complex assembly, and NSP18 is critical for replication ([Yan et al., 2022\)](#page-24-0).

10. Structural protein targets

10.1. Envelope protein

The E protein is a transmembrane cation-selective viroporin with Ca^{2+} and/or K⁺ selectivity ([Hong et al., 2020](#page-20-0); [Mandala et al., 2020](#page-22-0)). Similar to previous reports with SARS/MERS, SARS-CoV-2, the E protein also forms an inflammasome by TLR2 or NRLP5 activation through NF-kB due to K^+ influx [\(Yalcinkaya et al., 2021;](#page-24-0) [M. Zheng et al., 2021](#page-25-0)). β-boswellic acid and glycyrrhizic acid natural product combinations have been shown to shorten the recovery time ([Gomaa et al., 2022\)](#page-20-0), and in a suggestion of a possible mechanism, they have shown positive binding with the E protein *in vitro* ([Fatima et al., 2022\)](#page-19-0)*.* There are a few phytochemicals i.e. proanthocyanidins (PAC), wortmannin, and veliparib reported to block E protein *in vitro* [\(Y. Wang et al., 2022](#page-24-0)).

10.2. Spike glycoprotein

Spike protein, (S1, S2, S3) is the largest protein coded by the SARS-CoV-2 genome. It has various domains including transmembrane, S1 & S2 domains. S1 binds to different receptors (ACE2, CD147, B0AT1, and NRP1) and interacts with heparan sulfate and the S2 domain is a viral fusion domain. S1 domain has open and closed states to maintain the receptor-binding domain (RBD) specificity ([Gupta et al., 2021c; Jackson](#page-20-0) [et al., 2022\)](#page-20-0). The fusion inhibitors are discussed in detail in later RBD-ACE-2 interaction inhibition. S2 activation requires cleavage of spike protein mediated by furin and TMPRSS2 (Y. [Gupta et al., 2022](#page-20-0)). Itraconazole and Estradiol Benzoate were found to be interacting with the S2 domain of spike protein and had *in vitro* activities of IC50 0.45 (μM) and 1.02 (μM) respectively [\(Yang et al., 2021](#page-24-0)). Itraconazole synergistically improved the remdesivir efficacy *in vitro* [\(Schloer et al.,](#page-23-0) [2021\)](#page-23-0). Pan-CoV fusion inhibitor EK1 (fusion domain S2) is efficacious against all variants suggesting high target conservancy despite the high degree of amino acid mutations in SARS-CoV-2 variants ([Lan et al.,](#page-21-0) [2021\)](#page-21-0). Further, a designer peptide mimicking the HR2 sub-domain of the S2 fusion domain (VVNIQKEIDRLNEVAKNLNESLID) was designed *in silico* and validated both by MD simulations and *in vitro* testing ([Kandeel et al., 2021;](#page-21-0) [Manna et al., 2020\)](#page-22-0).

10.3. Homo sapiens (Host) COVID-19 therapeutic targets

In addition to targeting SARS-CoV-2 proteins, another therapeutic approach is to target host proteins that enable viral infection, replication, and spread [\(Fig. 2\)](#page-10-0). The interventions range from interfering with the host receptors for SARS-CoV-2 (e.g. ACE2), to blocking the proteolytic processing needed for viral particle internalization (e.g. Cathepsin L).

10.4. Viral receptors targets of human host

Fig. 2. Host proteome targets involved in COVID-19 hyperimmune and their inhibitors. Cartoon representation of molecular components involved in hyperimmune reaction leading to the severe clinical presentation (ARDS) among COVID-19 patients. The lung fibrosis observed in COVID-19 patients and resulting hypoxia is the main reason for mortality in severe cases along with immunosuppressed conditions and concomitant infections. Classic pathways are hijacked in COVID-19-associated lung fibrosis by various proteins coded by SARS-CoV-2. In COVID-19 patients due to inflammation mediators such as IL-6 and cytokine storm or increased release from damaged/dying cells, there is a loss of lung surface area to fibrosis. There is an aggravation of the infection cycle due to hypoxia-induced ACE2, TMPRSS2 overexpression, and furin cell surface localization. Multiple immune suppressants and modulators have been effective in reducing the severity and mortality as seen in large trials. However, the mechanism for which is still not well established. There are many other agents known to modulate many members of this cascade, especially the NLRP3 pathway responsible for characteristic COVID-19 storm but not yet exploited due to a rather recent elucidation.

10.5. Host receptors

ACE2 is the most abundant and highest affinity receptor of SARS-CoV-2 spike protein and is the first step in viral entry into the host cell. There are multiple reports that ACE2 polymorphisms and Spike protein modulate viral infectivity [\(Suryamohan et al., 2021](#page-23-0)). Various known ACE2 inhibitors, as well as expression modulators, have been proposed to be viable anti-COVID-19 therapeutics. There is another novel approach of molecular mimicry where B38-CAP an ACE2 homolog carboxypeptidase of bacterial origin protected patients from lung injury without apparent viral neutralization, but through a mechanism of RAS inactivation and decreased Acute Respiratory Distress Syndrome (ARDS) ([Yamaguchi et al., 2021\)](#page-24-0). This is coherent with the previous reports of lung damage protection with recombinant soluble ACE2 in animal models [\(Imai et al., 2005, p. 20](#page-20-0)). Also, soluble recombinant human ACE2 has a high SARS-CoV-2 neutralizing potential as shown *in vitro* [\(Monteil](#page-22-0) [et al., 2020](#page-22-0)). Giapreza, the angiotensin II substrate of ACE2, had variable outcomes from different studies. The conclusive multicentric trial concluded a decrease in blood pressure and improved fraction of inspired oxygen (FiO2) levels but there was no apparent benefit in terms of mortality among severe ARDS patients. ACE2 agonists have also shown a decrease in Spike-ACE2 interaction as their binding site is closer to the interface compare to antagonists e.g. Losartan/Valsartan that bind in the catalytic core and have no positive effect as reported in multiple trials [\(Geriak et al., 2021](#page-19-0); [Puskarich et al., 2021](#page-22-0), [2022](#page-22-0)). A small randomized trial with 51 patients receiving C21 and an ACE2 agonist

showed a significant reduction in the requirement of mechanical ventilation ([Tornling et al., 2021\)](#page-24-0). Methylene Blue is a nonspecific ACE2-Spike interaction inhibitor and has been used to inactivate residual viruses in convalescent plasma ([Table 4](#page-15-0)) ([Alemany et al., 2022](#page-17-0)). Ceftazidime is an injectable broad-spectrum beta-lactam antibiotic that is a third-generation cephalosporin. Ceftazidime was found to effectively block ACE-2 spike interactions *in vitro* ([C. Lin et al., 2021](#page-21-0))*.* It was trialed on 136 patients in a study and showed a significant reduction in recovery (PCR negativity) [\(Eid et al., 2021\)](#page-19-0). On the contrary, Ramipril is highly contraindicated in COVID-19 patients as it is known to highly up-regulate ACE2 and increase SARS-CoV-2 virion loads ([Theodor](#page-24-0)[akopoulou et al., 2022](#page-24-0)).

Neuropilin-1 (NRP1) is another host surface receptor mediating SARS-CoV-2 entry [\(Cantuti-Castelvetri et al., 2020](#page-18-0); [Kyrou et al., 2021\)](#page-21-0) and has been associated with neurological morbidities seen in COVID-19 ([Davies et al., 2020\)](#page-18-0). Apart from protein receptor binding spike protein also interacts with cell surface heparan sulfate and is the basis for antiviral activity of heparin [\(Gupta et al., 2021c](#page-20-0)) and sulfated polysaccharides [\(Kwon et al., 2020](#page-21-0)) abundant in many natural products. There is a high interest in using sulfated polysaccharides as anti-COVID-19 also due to the reduction in coagulopathy seen in COVID-19 patients [\(B. Tu et al., 2022\)](#page-24-0). There is still a possibility of SARS-CoV-2 variants evolving or already evolved to use different receptors like other coronaviruses ([Nassar et al., 2021\)](#page-22-0).

l**e 3**
ss targeting different viral/host proteins with *in vitro v*alidatio

Molecular Aspects of Medicine 91 (2023) 101151

Molecular Aspects of Medicine 91 (2023) 101151

Table 3 (*continued*)

11. Spike processing enzymatic targets

11.1. Cathepsin L

Cathepsin L (CTSL) is a transmembrane peptidase/serine subfamily member 2/4 and plays an important role in spike activation in endosomes. The widespread now-dominant mutation in the SARS-CoV-2 Spike glycoprotein D614G is predicted to confer a site loss for CTSL ([Gobeil et al., 2020;](#page-20-0) Y. [Gupta et al., 2022](#page-20-0)). Amantadine acts as a lysosomotropic agent by disturbing Cathepsin L's functional environment ([Smieszek et al., 2020\)](#page-23-0). A few reports are showing decreased leukopathy ([Lam et al., 2022\)](#page-21-0) and the slowdown of neurodegeneration presentations of COVID-19 by amantadine [\(Rejdak and Grieb, 2020](#page-23-0)).

11.2. Furin

Furin is a Ca^{2+-} dependent endopeptidase that processes many secretory proteins as well as protein digestion ([Than et al., 2005](#page-24-0)). During hypoxia, furin can translocate to the cell surface and is thought to be responsible for the rapid worsening of hypoxia patients in COVID-19 by increased spike processing at the cell surface resulting in direct fusion ([Arsenault et al., 2012;](#page-17-0) Y. [Gupta et al., 2022](#page-20-0)). Both Furin is essential for SARS-CoV-2 invasion ([Bestle et al., 2020](#page-17-0)) and known furin inhibitors MI-1851 and E-64d have both shown *in vitro* efficacy against SARS-CoV-2 [\(Table 3](#page-11-0)).

11.3. TMPRSS2

Transmembrane serine protease 2 (TMPRSS2) is a cell surface activator of spike protein essential to exposing and activating the viral fusion domain ([Bestle et al., 2020;](#page-17-0) [Hoffmann et al., 2020\)](#page-20-0). Nafamostat (CKD-314/Nafabelltan) a TMPRSS2 inhibitor was found to instigate a significantly higher recovery rate among treated patients and was well tolerated [\(Table 4\)](#page-15-0) [\(Zhuravel et al., 2021\)](#page-25-0). Another TMPRSS2 inhibitor

Camostat mesylate (FOY-305) in contrast didn't show any positive effect in a phase III trial [\(Kinoshita et al., 2022\)](#page-21-0). One speculation for inconclusive outcome with Camostat is the drug might need a better dosage formulation for effective treatment ([Kosinsky et al., 2022\)](#page-21-0). There are additional inhibitors of TMPRSS2 with promising results *in vitro* e.g. Aprotinin, MI-1900, MI-432, E-64d, PCI-27483, and Otamixaban.

11.4. Targets associated with host immune response

The TLR 2/6/9 agonist PUL-042 is a phase III investigational compound that can induce epithelial resistance to SARS-CoV-2 in animal models ([Evans et al., 2020\)](#page-19-0). Famotidine is a selective histamine H2-receptor (H2R) antagonist ([Malone et al., 2021\)](#page-22-0) that also inhibits 3CLpro of SARS-CoV-2 [\(Loffredo et al., 2021\)](#page-21-0). Famotidine had a positive effect with a reduced risk of clinical deterioration leading to intubation or death when tested in a small retrospective cohort [\(Freedberg et al.,](#page-19-0) [2020\)](#page-19-0). Currently, famotidine is part of multiple combinations in various trials. There are hypothetical reports of targeting different immune components such as Basigin CD_antigen: CD147, 5F7, Collagenase stimulatory factor, Leukocyte activation antigen M6, Extracellular matrix metalloproteinase inducer, Tumor cell-derived collagenase stimulatory factor, GCSF-Receptor Signaling Complex CSF3, IL-1β, leukocytic pyrogen, leukocytic endogenous mediator, and mononuclear cell factor, yet discussing all of these is beyond the scope of the current review. Major confounding comorbidity arising in a portion of the SARS-CoV-2 infected populations is the activation of a cytokine storm leading to the development of ARDS. To block the cytokine storm from activating in COVID-19 patients, various antibody cocktails blocking these factors have been used in ongoing trials [\(Elahi et al., 2022;](#page-19-0) [Harrison, 2020](#page-20-0); [Harrison et al., 2021](#page-20-0)). Many recombinant proteins e.g. Recombinant TNF (INB03) and Recombinant human interferon α 1 β (Novaferon) have also been tried (Drozdzal [et al., 2021\)](#page-19-0). Other targets include Peginterferon Lambda-1a, and Chemokine Receptor Type 2 (CCR2) ([Hu et al.,](#page-20-0) [2021\)](#page-20-0). The Interleukin-1 receptor-associated Kinase 4 (IRAK4) Inhibitor

Table 4

PF-06650833 is predicted to restore immunological balance [\(Gupta and](#page-20-0) [Chun, 2021\)](#page-20-0) and is under trial ([Franchin, 2021](#page-19-0)). Sigma-1 receptor (sigma non-opioid intracellular receptor 1) is an important factor associated with the mortality of COVID-19 patients ([Lehrer and Rheinstein,](#page-21-0) [2021\)](#page-21-0) several inhibitors have been predicted to be anti-COVID-19 e.g. Haloperidol, PD-144418, clemastine, Cloperastine, and progesterone. Naringenin, targeting the endo-lysosomal Two-Pore Channels (TPCs) has been shown as having anti-SARS-CoV-2 activity ([Clementi et al.,](#page-18-0) [2021\)](#page-18-0).

12. Mechanistic targets

12.1. Dihydroorotate dehydrogenase

Dihydroorotate dehydrogenase (mitochondrial DHODH), is a Dihydroorotate oxidase involved in pyrimidine synthesis within cells. DHODH inhibition has been shown to decrease viral replication/turnover rates [\(Kaur et al., 2021](#page-21-0)) as well as increase the incorporation of nucleoside analog antivirals such as N4-hydroxycytidine (NHC) which is an activated metabolite of Molnupiravir ([Stegmann et al., 2021](#page-23-0)).

Brequinar (DUP 785, NSC 368390) in combination with nucleoside analog Dipyridamole has shown high *in vitro* efficacy ([Demarest et al.,](#page-18-0) [2022;](#page-18-0) [Xiong et al., 2020\)](#page-24-0) and is in Phase II trials. There are many more DHODH inhibitors showing high anti-SARS-CoV-2 activities e.g. PTC299 ([Luban et al., 2021\)](#page-21-0), Teriflunomide ([Maghzi et al., 2020\)](#page-21-0), and Leflunomide ([Hu et al., 2020](#page-20-0)). Leflunomide also showed faster PCR negativity in COVID-19 patients in a small trial [\(Hu et al., 2020\)](#page-20-0).

12.2. Cathepsin B

Cathepsin B (APP secretase/Cathepsin B1) is an important enzyme overexpressed in hyperimmune inflammatory disorders and hence can be a target for ARDS mitigation ([Ding et al., 2022](#page-19-0)).

12.3. Caspase

COVID-19 inflammasome causes cell death through caspase pathways, specifically caspase 8 ([Li et al., 2020](#page-21-0)). Belnacasan and Emricasan are Caspase inhibitors that showed inhibition of inflammasome *in vitro* ([Jeong et al., 2022](#page-20-0))*.*

12.4. Calpain

Calpain inhibitor BLD-2660 is an anti-fibrotic and part of many ongoing trials shown to mitigate lung fibrosis in combinations with antivirals [\(Djordje et al., 2021\)](#page-19-0).

12.5. Ferroportin

Multiple reports point to SARS-CoV-2 mediated lung injury being mediated by ferroptosis with a portion of spike protein mimicking hepcidin hormone (Y. [Gupta et al., 2022](#page-20-0)). Vitamin D is known to induce ferroportin overexpression which effluxes out the excess iron thereby preventing ferroptosis to reduce lung injury [\(Moran-Lev et al., 2018](#page-22-0)). Low levels of vitamin D were associated with higher COVID-19 mortality and it has been part of various combinations as an inexpensive therapeutic supplement for COVID-19 patients ([Z. Wang et al., 2022\)](#page-24-0).

12.6. Eukaryotic elongation factor 1A2 (eEF1A2)

Nitazoxanide is a thiazolide chemical compound that induces $eIF2\alpha$ (eukaryotic translation initiation factor-2) overexpression and PKR (double-stranded-RNA-activated protein kinase) phosphorylation, which has been used clinically to control Japanese encephalitis virus replication ([Elazor et al., 2008;](#page-19-0) [Shi et al., 2014](#page-23-0)). Nitazoxanide has been part of various combinations for SARS-CoV-2 infections and has shown depression in disease trajectory if started early on (Mendieta Zerón et al., [2021; Miorin et al., n.d.](#page-22-0); [Rocco et al., 2021\)](#page-23-0). Paradoxically, Plitidepsin (dehydrodidemnin B/Aplidin) is a marine-derived cyclic depsipeptide inhibiting eEF1A2 that is authorized in a few countries for treating refractory multiple myeloma. Preclinical and randomized phase-I trials showed Plitidepsin to be well tolerated and block the SARS-CoV-2 virus at the nanomolar range ([Varona et al., 2022\)](#page-24-0). Both eEF1A2 inhibition and overexpression seem to be detrimental to SARS-CoV-2 pathogenesis.

12.7. Inosine-5′ *-monophosphate dehydrogenase (IMPDH)*

Merimepodib (MMPD) is a IMPDH inhibitor that showed 2.5-log decrease in viral titers (p-value $= 0.0004$) with 4hr pretreatment ([Bukreyeva et al., 2020](#page-18-0)). When used in combination with Remdesivir, there was a rapid undetected level of achievement of viral load *in vitro*; a trial with the same combination is ongoing [\(Wimmer and Keestra,](#page-24-0) [2022\)](#page-24-0).

13. Target independent drugs

13.1. NSAIDs

Indomethacin is an NSAID that inhibits prostaglandin E synthase 2 (PGES-2) [\(Lucas, 2016\)](#page-21-0). Its mechanism of action is still an enigma, while its primary target is IL6 suppression through PGES-2 inhibition, it is also proposed to block multiple factors for severe COVID-19 e.g. suppressing ACE2, TMPRSS2, cytokines, and inflammation in general [\(Alkotaji and](#page-17-0) [Al-Zidan, 2021\)](#page-17-0). Indomethacin has shown 100% protection from the development of hypoxia/desaturation with $SpO2 \leq 93$ compared to 16–22% in the untreated pool of patients ([Ravichandran et al., 2022](#page-23-0)).

14. Newer approaches to drugging targets

A variety of novel targets are being investigated with non-standard drug targeting. Ensovibep (MP0420) is a DARPins derivative that is an emerging class of novel therapeutics. This molecule's three distinct DARPin domains are designed to simultaneously target the receptor binding ridge on each RBD of the spike trimer [\(Chonira et al., 2022](#page-18-0)). MP0420 had an IC50 of an average of 2.3 ng/ml except for the mutation F486V, it was twice as effective as neutralizing antibodies; REGN10933 and REGN10987, and had a better efficacy against variants of concern ([Reichen et al., n.d.\)](#page-23-0).

A novel therapeutic paradigm is a proteolysis-targeting chimera (PROTAC), an application of targeted protein degradation, which has successfully been applied toward COVID-19 targets ([Shaheer et al.,](#page-23-0) [2021\)](#page-23-0). Essentially, PROTACs have a region that binds the viral target and the same region that binds a ubiquitin ligase, thereby positioning it to traffic the target for degradation. Since the virus must enter the cell, it is thereafter susceptible to PROTACs. Viral proteins are also exogenous, making them good targets from a standpoint of specificity. Furthermore, fragments generated from degradation can result in novel antigens that stimulate the host immune response. MPRO in particular has been selected as a viable candidate for PROTACs [\(Shaheer et al., 2021](#page-23-0)). Other potential targets include viral envelope proteins, PLpro, and RNA-dependent RNA polymerase (RdRp). PROTACs use a ligand as the basis for targeted protein degradation, novel therapeutics can be based on existing drugs or those in development, for the appropriate intracellular targets. For example, indomethacin has gained attention after drug repurposing studies identified its antiviral capabilities ([Shekhar](#page-23-0) [et al., 2022](#page-23-0); [Zeng et al., 2020\)](#page-25-0). A recent study investigated the effectiveness of indomethacin-based PROTACs in pan anti-coronavirus therapy [\(Desantis et al., 2021](#page-18-0)). Their findings indicated the indomethacin-PROTAC was more potent at inhibiting coronavirus, as well as was able to be effective against multiple strains of coronavirus ([Table 4](#page-15-0)).

A major limitation of PROTACs is that they are only useable for intracellular targets, or at least ones with an intracellular component; this limitation precludes a vast range of potential targets of high importance. A very recent technique called molecular degraders of extracellular proteins through the asialoglycoprotein receptor (MoDE-As) addresses the glaring weakness of targeted protein degradation. MoDE-As can target extracellular proteins for degradation [\(Caianiello](#page-18-0) [et al., 2021](#page-18-0)). This is accomplished via the formation of a ternary complex between a target protein, the ligand, and hepatocyte ASGPRs; this complex is then endocytosed, trafficked to the lysosome and the target protein is degraded by the host machinery. While MoDE-As has not yet been applied to COVID-19 therapy, it is a viable technique to intervene with viral protein targets before they enter the cells. Furthermore, there is evidence that the SARS-CoV-2 spike protein interacts with the ASGPR in hepatocytes through a lesser-known mechanism of entry [\(Collins and](#page-18-0) [Steer, 2021;](#page-18-0) [Gu et al., 2022](#page-20-0)).

15. Conclusion

COVID-19 disease can be safely called a virus-induced hyper-immune disorder. There are thus numerous factors still being discovered from the host point of view which can be mitigated by various therapeutics to reduce the severe clinical presentations (W. [Zhang et al.,](#page-25-0) [2022\)](#page-25-0). Also with new roles assigned to various viral components essential in pathogenesis and severe disease progression, numerous virus-coded proteins have been proposed as drug targets albeit only a few have bioactive inhibitors ([Martin et al., 2020\)](#page-22-0). Although there are numerous agents with known *in vitro* activity, there is an urgent need to form suitable combinations based on the synergy of the agents, a stratified patient population taking into consideration important pathways leading to either ARDS or Long-haul disorders. Also, various *trialed* agents with borderline protection or a population-specific activity can be used to fortify newly discovered strong antivirals like Nirmatrelvir or Molnopiravir. As there is no single pathway in this COVID-19 sequela, there is an urgent need for utilizing personalized medicine combinations composed of the most tolerated and active agent combinations.

Intriguingly, when viewing from a drug discovery perspective, there is a learning phase we must endeavor to better understand the druggability of identified viral targets with known and potential inhibitors to continue developing new antivirals to be better prepared for the emergence of drug resistance to current candidates and therapeutics [\(Gandhi](#page-19-0) [et al., 2022](#page-19-0)), especially when it's now known as immunocompromised patients are the source of new resistant variant emergence ([Chen et al.,](#page-18-0) [2021;](#page-18-0) [Gandhi et al., 2022](#page-19-0); [Leung et al., 2022](#page-21-0)).

Within this realm of rapidly advancing, technology is a convergent race between computational and experimental methods, which furthers the acceleration of drug discovery[\(Dara et al., 2022;](#page-18-0) [Hinton, 2007](#page-20-0); Jiménez-Luna et al., 2021; [Lima et al., 2016;](#page-21-0) [Patel et al., 2020](#page-22-0); [Sher](#page-23-0)[rington and Kirkpatrick, 1975;](#page-23-0) [Talevi et al., 2020](#page-23-0)). We are using ML increasingly in multiple areas of science and even in other areas (e.g. social science), whilst we are making stronger strides in computational design techniques. ML is now commonplace in digital pathology, search engines, recognition (voice, facial, pattern), market and financial predictions, astronomy, cryptography, agriculture, and more. The use of AL, ML, and deep learning techniques is to better find and rapidly identify data from multiple sources, extract valuable insights, visualize the data meaningfully, and give context. Within drug discovery, there is an ongoing explosion of the use of ML with molecular modeling for protein structure prediction and drug-protein interaction analyses. For example, the pioneering of Boltzmann machines using decision trees and then adaptive rules for protein structures was a crucial development that allowed the generation of predetermined global variables on molecular structures to dictate conformational searches in directions under the reinforced learning pattern dictated [\(Caulfield and Devkota, 2012](#page-18-0); [Caulfield, 2011](#page-18-0); [Caulfield et al., 2011](#page-18-0); [Coban et al., 2021b;](#page-18-0) [Kayode et al.,](#page-21-0) [2016;](#page-21-0) [von Roemeling et al., 2018\)](#page-24-0). The particularly useful application of this allowed such things as cryo-EM fitting and rapid space searches ([Caulfield and Devkota, 2012](#page-18-0); [Caulfield et al., 2011\)](#page-18-0) using entropy as the controller.

Particularly of note is the emergence of AI and ML to the forefront of protein structural modeling, conformational dynamics exploratory mission of many labs to find key druggable states, and the determination of the human genomic variance as a contributing factor to the way viruses capitalize on variation. Virus exploitation of human genetic variance is also being tackled by computationalists to better understand how genetics plays a role in virus proliferation, which will allow better tools to predict potential virus offshoots in the future. One can imagine a day when there will be a virtual medicine cabinet pre-stocked with the needed antivirals specific to the patient's genetic predispositions and particular cell pathways. In such a scenario, we will have AI-based medicine that has the genetic profile, molecular structures for the targets needed, rapidly available custom chemistry, and rapid safetyprofiling needed for the new chemical entities to be used in humans

on-demand with acceptable safety tolerances. While this particular view of AI and ML is not anytime soon, the palatability of this particular star trek viewpoint is very realizable and within our horizon.

References

- Abidi, S.H., Almansour, N.M., Amerzhanov, D., Allemailem, K.S., Rafaqat, W., Ibrahim, M.A., la Fleur, P., Lukac, M., Ali, S., 2021. Repurposing potential of posaconazole and grazoprevir as inhibitors of SARS-CoV-2 helicase. Sci. Rep. 11, 1–11.<https://doi.org/10.1038/s41598-021-89724-0>.
- [Abolhassani, H., Bashiri, G., Montazeri, M., Kouchakzadeh, H., Shojaosadati, S.A.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref2) [Siadat, S.E.R., 2021. Ongoing clinical trials and the potential therapeutics for](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref2) [COVID-19 treatment. In: COVID-19. Springer, pp. 27](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref2)–89.
- Ahmed, M., Farag, A., Boys, I.N., Wang, P., L. Eitson, J., Ohlson, M.B., Fan, W., McDougal, M.B., Menendez-Montez, I., Uyen Nhi Nguyen, N.U.N.N., Mar, K., Ortiz, F., Young Kim, S., Williams, N., lemoff, A.L., DeBerardinis, R., Schoggins, J, W., Sadek, H., 2021. Identification of atovaquone and mebendazole as repurposed drugs with antiviral activity against SARS-CoV-2. Chemistry. [https://doi.org/](https://doi.org/10.26434/chemrxiv-2021-b3fv1-v7) [10.26434/chemrxiv-2021-b3fv1-v7](https://doi.org/10.26434/chemrxiv-2021-b3fv1-v7).) (preprint), Version 6.
- Alemany, A., Millat-Martinez, P., Corbacho-Monné, M., Malchair, P., Ouchi, D., Ruiz-Comellas, A., Ramírez-Morros, A., Codina, J.R., Simon, R.A., Videla, S., others, 2022. High-titre methylene blue-treated convalescent plasma as an early treatment for outpatients with COVID-19: a randomised, placebo-controlled trial. Lancet Respir. Med. 10, 278–288. [https://doi.org/10.1016/S2213-2600\(21\)00545-2.](https://doi.org/10.1016/S2213-2600(21)00545-2)
- Alkotaji, M., Al-Zidan, R.N., 2021. Indomethacin: can it counteract bradykinin effects in COVID-19 patients? Curr. Pharm. Rep. 7, 102–106. [https://doi.org/10.1007/](https://doi.org/10.1007/s40495-021-00257-6) [s40495-021-00257-6](https://doi.org/10.1007/s40495-021-00257-6).
- Amani, Bahman, Khanijahani, A., Amani, Behnam, Hashemi, P., 2021. Lopinavir/ ritonavir for COVID-19: a systematic review and meta-analysis. J. Pharm. Pharmaceut. Sci. 24, 246–257. [https://doi.org/10.18433/jpps31668.](https://doi.org/10.18433/jpps31668)
- Amani, Behnam, Amani, Bahman, Zareei, S., Zareei, M., 2021. Efficacy and safety of arbidol (umifenovir) in patients with COVID-19: a systematic review and metaanalysis. Immun. Inflamm. Dis. 9, 1197–1208. <https://doi.org/10.1002/iid3.502>.
- Andreana, I., Bincoletto, V., Milla, P., Dosio, F., Stella, B., Arpicco, S., 2022. Nanotechnological approaches for pentamidine delivery. Drug Deliv. and Transl. Res. 12, 1911–1927. [https://doi.org/10.1007/s13346-022-01127-4.](https://doi.org/10.1007/s13346-022-01127-4)
- Angamo, M.T., Mohammed, M.A., Peterson, G.M., 2022. Efficacy and safety of remdesivir in hospitalised COVID-19 patients: a systematic review and metaanalysis. Infection 50, 27–41. <https://doi.org/10.1007/s15010-021-01671-0>.
- Anjum, F., Mohammad, T., Asrani, P., Shafie, A., Singh, S., Yadav, D.K., Uversky, V.N., Hassan, M.I., 2022. Identification of intrinsically disorder regions in non-structural proteins of SARS-CoV-2: new insights into drug and vaccine resistance. Mol. Cell. Biochem. 477, 1607–1619.<https://doi.org/10.1007/s11010-022-04393-5>.
- Arsenault, D., Lucien, F., Dubois, C.M., 2012. Hypoxia enhances cancer cell invasion through relocalization of the proprotein convertase furin from the trans-golgi network to the cell surface. J. Cell. Physiol. 227, 789–800. [https://doi.org/10.1002/](https://doi.org/10.1002/jcp.22792) [jcp.22792.](https://doi.org/10.1002/jcp.22792)
- Auwul, M.R., Rahman, M.R., Gov, E., Shahjaman, M., Moni, M.A., 2021. Bioinformatics and machine learning approach identifies potential drug targets and pathways in COVID-19. Briefings Bioinf. 22, bbab120 <https://doi.org/10.1093/bib/bbab120>.
- Bai, Y., Ye, F., Feng, Y., Liao, H., Song, H., Qi, J., Gao, G.F., Tan, W., Fu, L., Shi, Y., 2021. Structural basis for the inhibition of the SARS-CoV-2 main protease by the anti-HCV drug narlaprevir. Signal Transduct. Targeted Ther. 6, 1–3. [https://doi.org/10.1038/](https://doi.org/10.1038/s41392-021-00468-9) [s41392-021-00468-9](https://doi.org/10.1038/s41392-021-00468-9).
- Bain, J., McLauchlan, H., Elliott, M., Cohen, P., 2003. The specificities of protein kinase inhibitors: an update. Biochem. J. 371, 199–204. [https://doi.org/10.1042/](https://doi.org/10.1042/BJ20021535) [BJ20021535](https://doi.org/10.1042/BJ20021535).
- Baker, J.D., Uhrich, R.L., Kraemer, G.C., Love, J.E., Kraemer, B.C., 2021. A drug repurposing screen identifies hepatitis C antivirals as inhibitors of the SARS-CoV2 main protease. PLoS One 16, e0245962. [https://doi.org/10.1371/journal.](https://doi.org/10.1371/journal.pone.0245962) [pone.0245962](https://doi.org/10.1371/journal.pone.0245962).
- Balli, F., Kara, E., Demirkan, S.K., 2020. The another side of COVID-19 in Alzheimer's disease patients: drug-drug interactions. Int. J. Clin. Pract. 74 (10), e13596 [https://](https://doi.org/10.1111/ijcp.13596) doi.org/10.1111/ijcp.13596. In press.
- [Barratt-Due, A., Olsen, I.C., Henriksen, K.N., K\aasine, T., Lund-Johansen, F., Hoel, H.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref17) [Holten, A.R., Tveita, A., Mathiessen, A., Haugli, M., others, 2021. Evaluation of](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref17) [Remdesivir and Hydroxychloroquine on Viral Clearance in COVID-19 Patients:](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref17) [Results from the NOR-Solidarity Randomised Trial.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref17)
- Beckerman, R., Gori, A., Jeyakumar, S., Malin, J.J., Paredes, R., Póvoa, P., Smith, N.J. Teixeira-Pinto, A., 2022. Remdesivir for the treatment of patients hospitalized with COVID-19 receiving supplemental oxygen: a targeted literature review and metaanalysis. Sci. Rep. 12, 9622. [https://doi.org/10.1038/s41598-022-13680-6.](https://doi.org/10.1038/s41598-022-13680-6)
- Beigel, J.H., Tomashek, K.M., Dodd, L.E., Mehta, A.K., Zingman, B.S., Kalil, A.C., Hohmann, E., Chu, H.Y., Luetkemeyer, A., Kline, S., others, 2020. Remdesivir for the treatment of covid-19. N. Engl. J. Med. <https://doi.org/10.1056/NEJMoa2007764>
- Bestle, D., Heindl, M.R., Limburg, H., Pilgram, O., Moulton, H., Stein, D.A., Hardes, K., Eickmann, M., Dolnik, O., Rohde, C., others, 2020. TMPRSS2 and furin are both essential for proteolytic activation of SARS-CoV-2 in human airway cells. Life science alliance 3. <https://doi.org/10.26508/lsa.202000786>.

[Bloch, A., 2003. Murphy](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref21)'s Law. Penguin.

Blum, V.F., Cimerman, S., Hunter, J.R., Tierno, P., Lacerda, A., Soeiro, A., Cardoso, F., Bellei, N.C., Maricato, J., Mantovani, N., Vassao, M., Dias, D., Galinskas, J., Janini, L. M.R., Santos-Oliveira, J.R., Da-Cruz, A.M., Diaz, R.S., 2021. Nitazoxanide superiority to placebo to treat moderate COVID-19 – a Pilot prove of concept randomized

double-blind clinical trial. eClinicalMedicine 37, 100981. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.eclinm.2021.100981) [eclinm.2021.100981.](https://doi.org/10.1016/j.eclinm.2021.100981)

- Bojkova, D., Bechtel, M., McLaughlin, K.-M., McGreig, J.E., Klann, K., Bellinghausen, C., Rohde, G., Jonigk, D., Braubach, P., Ciesek, S., Münch, C., Wass, M.N., Michaelis, M., Cinatl, J., 2020. Aprotinin inhibits SARS-CoV-2 replication. Cells 9, 2377. [https://](https://doi.org/10.3390/cells9112377) [doi.org/10.3390/cells9112377.](https://doi.org/10.3390/cells9112377)
- Boodhoo, N., Matsuyama-Kato, A., Shojadoost, B., Behboudi, S., Sharif, S., 2022. The severe acute respiratory syndrome coronavirus 2 non-structural proteins 1 and 15 proteins mediate antiviral immune evasion. Current Research in Virological Science 3, 100021.<https://doi.org/10.1016/j.crviro.2022.100021>.
- Brewitz, L., Kamps, J.J.A.G., Lukacik, P., Strain-Damerell, C., Zhao, Y., Tumber, A., Malla, T.R., Orville, A.M., Walsh, M.A., Schofield, C.J., 2022. Mass spectrometric assays reveal discrepancies in inhibition profiles for the SARS-CoV-2 papain-like protease. ChemMedChem 17. [https://doi.org/10.1002/cmdc.202200016.](https://doi.org/10.1002/cmdc.202200016) [Bukreyeva, N., Mantlo, E.K., Sattler, R.A., Huang, C., Paessler, S., Zeldis, J., 2020. The](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref26)
- [IMPDH inhibitor merimepodib suppresses SARS-CoV-2 replication in vitro. bioRxiv.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref26) Caianiello, D.F., Zhang, M., Ray, J.D., Howell, R.A., Swartzel, J.C., Branham, E.M.J., Chirkin, E., Sabbasani, V.R., Gong, A.Z., McDonald, D.M., Muthusamy, V.,
- Spiegel, D.A., 2021. Bifunctional small molecules that mediate the degradation of extracellular proteins. Nat. Chem. Biol. 17, 947–953. [https://doi.org/10.1038/](https://doi.org/10.1038/s41589-021-00851-1) [s41589-021-00851-1](https://doi.org/10.1038/s41589-021-00851-1).
- Cairns, D.M., Dulko, D., Griffiths, J.K., Golan, Y., Cohen, T., Trinquart, L., Price, L.L., Beaulac, K.R., Selker, H.P., 2022. Efficacy of niclosamide vs placebo in SARS-CoV-2 respiratory viral clearance, viral shedding, and duration of symptoms among patients with mild to moderate COVID-19: a phase 2 randomized clinical trial. JAMA Netw. Open 5, e2144942. [https://doi.org/10.1001/jamanetworkopen.2021.44942.](https://doi.org/10.1001/jamanetworkopen.2021.44942)
- [Callaway, E., 2022. 'The entire protein universe](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref29)': AI predicts shape of nearly every [known protein. nature news article.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref29)
- [Calonico, S., Di Tella, R., Del Valle, J.C.L., 2022. Causal Inference during a Pandemic:](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref30) [Evidence on the Effectiveness of Nebulized Ibuprofen as an Unproven Treatment for](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref30) [COVID-19 in Argentina. National Bureau of Economic Research.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref30)
- Cantuti-Castelvetri, L., Ojha, R., Pedro, L.D., Djannatian, M., Franz, J., Kuivanen, S., van der Meer, F., Kallio, K., Kaya, T., Anastasina, M., others, 2020. Neuropilin-1 facilitates SARS-CoV-2 cell entry and infectivity. Science 370, 856–860. [https://doi.](https://doi.org/10.1126/science.abd2985) [org/10.1126/science.abd2985](https://doi.org/10.1126/science.abd2985).
- Cao, B., Wang, Y., Wen, D., Liu, W., Wang, J., Fan, G., Ruan, L., Song, B., Cai, Y., Wei, M., others, 2020. A trial of lopinavir–ritonavir in adults hospitalized with severe Covid-19. N. Engl. J. Med. <https://doi.org/10.1056/NEJMoa2001282>.
- Caraco, Y., Crofoot, G.E., Moncada, P.A., Galustyan, A.N., Musungaie, D.B., Payne, B., Kovalchuk, E., Gonzalez, A., Brown, M.L., Williams-Diaz, A., others, 2022. Phase 2/3 trial of molnupiravir for treatment of Covid-19 in nonhospitalized adults. NEJM Evidence 1, EVIDoa2100043. <https://doi.org/10.1056/EVIDoa2100043>.
- Carpinteiro, A., Edwards, M.J., Hoffmann, M., Kochs, G., Gripp, B., Weigang, S., Adams, C., Carpinteiro, E., Gulbins, A., Keitsch, S., Sehl, C., Soddemann, M., Wilker, B., Kamler, M., Bertsch, T., Lang, K.S., Patel, S., Wilson, G.C., Walter, S., Hengel, H., Pöhlmann, S., Lang, P.A., Kornhuber, J., Becker, K.A., Ahmad, S.A., Fassbender, K., Gulbins, E., 2020. Pharmacological inhibition of acid sphingomyelinase prevents uptake of SARS-CoV-2 by epithelial cells. Cell Rep. Med. 1, 100142 [https://doi.org/10.1016/j.xcrm.2020.100142.](https://doi.org/10.1016/j.xcrm.2020.100142)
- Castillo-Garit, J.A., Cañizares-Carmenate, Y., Pérez-Giménez, F., 2021. Biosynthetic enzymes of the SARS-CoV-2 as potential targets for the discovery of new antiviral drugs. Nereis. Interdisciplinary Ibero-American Journal of Methods, Modelling and Simulation 17–23. [https://doi.org/10.46583/nereis_2021.13.844.](https://doi.org/10.46583/nereis_2021.13.844)
- Caulfield, T., Devkota, B., 2012. Motion of transfer RNA from the A/T state into the Asite using docking and simulations. Proteins 80, 2489–2500. [https://doi.org/](https://doi.org/10.1002/prot.24131) [10.1002/prot.24131](https://doi.org/10.1002/prot.24131).
- Caulfield, T., Medina-Franco, J.L., 2011. Molecular dynamics simulations of human DNA methyltransferase 3B with selective inhibitor nanaomycin A. J. Struct. Biol. 176, 185–191. <https://doi.org/10.1016/j.jsb.2011.07.015>.
- Caulfield, T.R., 2011. Inter-ring rotation of apolipoprotein A-I protein monomers for the double-belt model using biased molecular dynamics. J. Mol. Graph. Model. 29, 1006–1014. [https://doi.org/10.1016/j.jmgm.2011.04.005.](https://doi.org/10.1016/j.jmgm.2011.04.005)
- Caulfield, T.R., Devkota, B., Rollins, G.C., 2011. Examinations of tRNA range of motion using simulations of cryo-EM microscopy and X-ray data. J. Biophys. 2011, 219515 <https://doi.org/10.1155/2011/219515>.
- Chakraborty, C., Bhattacharya, M., Mallick, B., Sharma, A.R., Lee, S.-S., Agoramoorthy, G., 2021. SARS-CoV-2 protein drug targets landscape: a potential pharmacological insight view for the new drug development. Expet Rev. Clin.
- Pharmacol. 14, 225–238. [https://doi.org/10.1080/17512433.2021.1874348.](https://doi.org/10.1080/17512433.2021.1874348) Chan, H.-T., Chao, C.-M., Lai, C.-C., 2021. Sofosbuvir/daclatasvir in the treatment of COVID-19 infection: a meta-analysis. J. Infect. 82, e34–e35. [https://doi.org/](https://doi.org/10.1016/j.jinf.2020.12.021) [10.1016/j.jinf.2020.12.021.](https://doi.org/10.1016/j.jinf.2020.12.021)
- Chang, J., 2022. 4′ -Modified nucleosides for antiviral drug discovery: achievements and perspectives. Accounts Chem. Res. 55, 565–578. [https://doi.org/10.1021/acs.](https://doi.org/10.1021/acs.accounts.1c00697) [accounts.1c00697](https://doi.org/10.1021/acs.accounts.1c00697).
- Chejfec-Ciociano, J.M., Martínez-Herrera, J.P., Parra-Guerra, A.D., Chejfec, R., Barbosa-Camacho, F.J., Ibarrola-Peña, J.C., Cervantes-Guevara, G., Cervantes-Cardona, G.A., Fuentes-Orozco, C., Cervantes-Pérez, E., others, 2022. Misinformation about and interest in chlorine Dioxide during the COVID-19 pandemic in Mexico identified using google trends data: infodemiology study. JMIR Infodemiology 2, e29894. /doi.org/10.2196/29894.
- Chen, H., Zhang, Z., Wang, L., Huang, Z., Gong, F., Li, X., Chen, Y., Wu, J.J., 2020. First clinical study using HCV protease inhibitor danoprevir to treat COVID-19 patients. Medicine 99. https://doi.org/10.1097/MD.000000000002335
- Chen, J., Malone, B., Llewellyn, E., Grasso, M., Shelton, P.M., Olinares, P.D.B., Maruthi, K., Eng, E.T., Vatandaslar, H., Chait, B.T., others, 2020. Structural basis for

helicase-polymerase coupling in the SARS-CoV-2 replication-transcription complex. Cell 182, 1560–1573. <https://doi.org/10.1016/j.cell.2020.07.033>.

- Chen, L., Zody, M.C., Di Germanio, C., Martinelli, R., Mediavilla, J.R., Cunningham, M. H., Composto, K., Chow, K.F., Kordalewska, M., Corvelo, A., others, 2021. Emergence of multiple SARS-CoV-2 antibody escape variants in an immunocompromised host undergoing convalescent plasma treatment. mSphere 6. //doi.org/10.1128/mSphere.00480-21 e00480-21.
- Choi, R., Zhou, M., Shek, R., Wilson, J.W., Tillery, L., Craig, J.K., Salukhe, I.A., Hickson, S.E., Kumar, N., James, R.M., others, 2021. High-throughput screening of the ReFRAME, Pandemic Box, and COVID Box drug repurposing libraries against SARS-CoV-2 nsp15 endoribonuclease to identify small-molecule inhibitors of viral activity. PLoS One 16, e0250019. [https://doi.org/10.1371/journal.pone.0250019.](https://doi.org/10.1371/journal.pone.0250019)
- [Chonira, V.K., Kwon, Y.-D., Gorman, J., Case, J.B., Ku, Z., Simeon, R., Casner, R.G.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref48) [Harris, D.R., Olia, A.S., Stevens, T., others, 2022. Potent and Pan-Neutralization of](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref48) [SARS-CoV-2 Variants of Concern by DARPins. bioRxiv](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref48).
- Cimolai, N., 2020. Potentially repurposing adamantanes for COVID-19. J. Med. Virol. 92, 531–532. <https://doi.org/10.1002/jmv.25752>.
- Clemency, B.M., Varughese, R., Gonzalez-Rojas, Y., Morse, C.G., Phipatanakul, W., Koster, D.J., Blaiss, M.S., 2022. Efficacy of inhaled ciclesonide for outpatient treatment of adolescents and adults with symptomatic COVID-19: a randomized clinical trial. JAMA Intern. Med. 182, 42–49. [https://doi.org/10.1001/](https://doi.org/10.1001/jamainternmed.2021.6759) [jamainternmed.2021.6759.](https://doi.org/10.1001/jamainternmed.2021.6759)
- Clementi, N., Scagnolari, C., D'Amore, A., Palombi, F., Criscuolo, E., Frasca, F., Pierangeli, A., Mancini, N., Antonelli, G., Clementi, M., others, 2021. Naringenin is a powerful inhibitor of SARS-CoV-2 infection in vitro. Pharmacol. Res. 163, 105255 [https://doi.org/10.1016/j.phrs.2020.105255.](https://doi.org/10.1016/j.phrs.2020.105255)
- Coban, M.A., Blackburn, P.R., Whitelaw, M.L., Haelst, M.M. van, Atwal, P.S., Caulfield, T.R., 2020. Structural models for the dynamic effects of loss-of-function variants in the human SIM1 protein transcriptional activation domain. Biomolecules 10, E1314. [https://doi.org/10.3390/biom10091314.](https://doi.org/10.3390/biom10091314)
- Coban, M.A., Fraga, S., Caulfield, T.R., 2021a. Structural and computational perspectives of selectively targeting mutant proteins. Curr. Drug Discov. Technol. 18, 365–378. <https://doi.org/10.2174/1570163817666200311114819>.
- Coban, M.A., Morrison, J., Maharjan, S., Hernandez Medina, D.H., Li, W., Zhang, Y.S., Freeman, W.D., Radisky, E.S., Le Roch, K.G., Weisend, C.M., Ebihara, H., Caulfield, T.R., 2021b. Attacking COVID-19 progression using multi-drug therapy for synergetic target engagement. Biomolecules 11, 787. [https://doi.org/10.3390/](https://doi.org/10.3390/biom11060787) [biom11060787.](https://doi.org/10.3390/biom11060787)
- Collins, D.P., Steer, C.J., 2021. Binding of the SARS-CoV-2 spike protein to the asialoglycoprotein receptor on human primary hepatocytes and immortalized hepatocyte-like cells by confocal analysis. Hepat. Med. 13, 37–44. [https://doi.org/](https://doi.org/10.2147/HMER.S301979) [10.2147/HMER.S301979.](https://doi.org/10.2147/HMER.S301979)
- Conway, S.R., Keller, M.D., Bollard, C.M., 2022. Cellular therapies for the treatment and prevention of SARS-CoV-2 infection. Blood. [https://doi.org/10.1182/](https://doi.org/10.1182/blood.2021012249) [blood.2021012249](https://doi.org/10.1182/blood.2021012249).
- Costanzo, M., De Giglio, M.A.R., Roviello, G.N., 2020. SARS-CoV-2: recent reports on antiviral therapies based on lopinavir/ritonavir, darunavir/umifenovir, hydroxychloroquine, remdesivir, favipiravir and other drugs for the treatment of the new coronavirus. Curr. Med. Chem. [https://doi.org/10.2174/](https://doi.org/10.2174/0929867327666200416131117) [0929867327666200416131117](https://doi.org/10.2174/0929867327666200416131117).
- Cox, R.M., Wolf, J.D., Plemper, R.K., 2021. Therapeutically administered ribonucleoside analogue MK-4482/EIDD-2801 blocks SARS-CoV-2 transmission in ferrets. Nat. Microbiol. 6, 11–18. <https://doi.org/10.1038/s41564-020-00835-2>.
- [da Silva, M.F., de Araújo-Júnior, J.X., da Silva-Júnior, E.F., Heimfarth, L., Martins-](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref59)[Filho, P.R., Quintans, J. de S.S., Quintans-Júnior, L.J., 2022. Bradykinin-target](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref59) [therapies in SARS-CoV-2 infection: current evidence and perspectives. Naunyn-](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref59)Schmiedeberg'[s Arch. Pharmacol. 1](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref59)–9.
- Dai, W., Zhang, B., Jiang, X.-M., Su, H., Li, J., Zhao, Y., Xie, X., Jin, Z., Peng, J., Liu, F., Li, C., Li, Y., Bai, F., Wang, H., Cheng, X., Cen, X., Hu, S., Yang, X., Wang, J., Liu, X., Xiao, G., Jiang, H., Rao, Z., Zhang, L.-K., Xu, Y., Yang, H., Liu, H., 2020. Structurebased design of antiviral drug candidates targeting the SARS-CoV-2 main protease. Science 368, 1331–1335. [https://doi.org/10.1126/science.abb4489.](https://doi.org/10.1126/science.abb4489)
- [Dang, M., Song, J., 2022. A review of the effects of ATP and hydroxychloroquine on the](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref61) [phase separation of the SARS-CoV-2 nucleocapsid protein. Biophys. Rev. 1](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref61)–7.
- Daniel, J.A., Chau, N., Abdel-Hamid, M.K., Hu, L., von Kleist, L., Whiting, A., Krishnan, S., Maamary, P., Joseph, S.R., Simpson, F., Haucke, V., McCluskey, A., Robinson, P.J., 2015. Phenothiazine-derived antipsychotic drugs inhibit dynamin and clathrin-mediated endocytosis: antipsychotic drugs inhibit dynamin. Traffic 16, 635-654. https://doi.org/10.1111/tra.1227.
- Dara, S., Dhamercherla, S., Jadav, S.S., Babu, C.M., Ahsan, M.J., 2022. Machine learning in drug discovery: a review. Artif. Intell. Rev. 55, 1947–1999. [https://doi.org/](https://doi.org/10.1007/s10462-021-10058-4) [10.1007/s10462-021-10058-4.](https://doi.org/10.1007/s10462-021-10058-4)
- [Davies, J., Randeva, H.S., Chatha, K., Hall, M., Spandidos, D.A., Karteris, E., Kyrou, I.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref64) [2020. Neuropilin-1 as a new potential SARS-CoV-2 infection mediator implicated in](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref64) [the neurologic features and central nervous system involvement of COVID-19. Mol.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref64) [Med. Rep. 22, 4221](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref64)–4226.
- [Demarest, J.F., Kienle, M., Boytz, R., Ayres, M., Kim, E., Chung, D., Gandhi, V., Davey, R.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref65) [A., Sykes, D.B., Shohdy, N., others, 2022. Brequinar and Dipyridamole in](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref65) [combination exhibits synergistic antiviral activity against SARS-CoV-2 in vitro:](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref65) [rationale for a host-acting antiviral treatment strategy for COVID-19. bioRxiv](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref65).
- Deng, J., Zhou, F., Heybati, K., Ali, S., Zuo, Q.K., Hou, W., Dhivagaran, T., Ramaraju, H. B., Chang, O., Wong, C.Y., others, 2022. Efficacy of chloroquine and hydroxychloroquine for the treatment of hospitalized COVID-19 patients: a metaanalysis. Future Virol. 17, 95–118. <https://doi.org/10.2217/fvl-2021-0119>.
- Desantis, J., Mercorelli, B., Celegato, M., Croci, F., Bazzacco, A., Baroni, M., Siragusa, L., Cruciani, G., Loregian, A., Goracci, L., 2021. Indomethacin-based PROTACs as pan-

coronavirus antiviral agents. Eur. J. Med. Chem. 226, 113814 [https://doi.org/](https://doi.org/10.1016/j.ejmech.2021.113814) [10.1016/j.ejmech.2021.113814](https://doi.org/10.1016/j.ejmech.2021.113814).

Devi, K.P., Pourkarim, M.R., Thijssen, M., Sureda, A., Khayatkashani, M., Cismaru, C.A., Neagoe, I.B., Habtemariam, S., Razmjouei, S., Khayat Kashani, H.R., 2022. A perspective on the applications of furin inhibitors for the treatment of SARS-CoV-2. Pharmacol. Rep. 74, 425–430. [https://doi.org/10.1007/s43440-021-00344-x.](https://doi.org/10.1007/s43440-021-00344-x)

[Dhaka, P., Singh, A., Choudhary, S., Kumar, P., Sharma, G.K., Tomar, S., 2022. Discovery](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref69) [of Anti-SARS-CoV-2 Molecules Using Structure-Assisted Repurposing Approach](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref69) [Targeting N-Protein. bioRxiv](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref69).

Ding, X., Ye, N., Qiu, M., Guo, H., Li, J., Zhou, X., Yang, M., Xi, J., Liang, Y., Gong, Y., others, 2022. Cathepsin B is a potential therapeutic target for coronavirus disease 2019 patients with lung adenocarcinoma. Chem. Biol. Interact. 353, 109796 [https://](https://doi.org/10.1016/j.cbi.2022.109796) [doi.org/10.1016/j.cbi.2022.109796.](https://doi.org/10.1016/j.cbi.2022.109796)

Dittmar, M., Lee, J.S., Whig, K., Segrist, E., Li, M., Kamalia, B., Castellana, L., Ayyanathan, K., Cardenas-Diaz, F.L., Morrisey, E.E., others, 2021. Drug repurposing screens reveal cell-type-specific entry pathways and FDA-approved drugs active against SARS-Cov-2. Cell Rep. 35, 108959 [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.celrep.2021.108959) [celrep.2021.108959](https://doi.org/10.1016/j.celrep.2021.108959).

[Djordje, A., Avila, S.V., Forat, L., Xiaoxuan, F., Sanchez-Petitto, G., Siglin, J., Baddley, J.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref72) [Mannuel, H.D., Hanan, A., Hankey, K.G., others, 2021. Deep dissection of the](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref72) [antiviral immune profile of patients with COVID-19. Commun. Biol. 4](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref72).

Dong, E., Du, H., Gardner, L., 2020. An interactive web-based dashboard to track COVID-19 in real time. Lancet Infect. Dis. 20, 533–534. [https://doi.org/10.1016/S1473-](https://doi.org/10.1016/S1473-3099(20)30120-1) [3099\(20\)30120-1](https://doi.org/10.1016/S1473-3099(20)30120-1).

Drayman, N., DeMarco, J.K., Jones, K.A., Azizi, S.-A., Froggatt, H.M., Tan, K., Maltseva, N.I., Chen, S., Nicolaescu, V., Dvorkin, S., others, 2021. Masitinib is a broad coronavirus 3CL inhibitor that blocks replication of SARS-CoV-2. Science 373, 931–936. [https://doi.org/10.1126/science.abg5827.](https://doi.org/10.1126/science.abg5827)

Drożdżal, S., Rosik, J., Lechowicz, K., Machaj, F., Szostak, B., Przybyciński, J., Lorzadeh, S., Kotfis, K., Ghavami, S., Los, M.J., 2021. An update on drugs with therapeutic potential for SARS-CoV-2 (COVID-19) treatment. Drug Resist. Updates 59, 100794. <https://doi.org/10.1016/j.drup.2021.100794>.

Eberle, R.J., Olivier, D.S., Amaral, M.S., Gering, I., Willbold, D., Arni, R.K., Coronado, M. A., 2021. The repurposed drugs suramin and quinacrine cooperatively inhibit SARS-CoV-2 3CLpro in vitro. Viruses 13, 873. [https://doi.org/10.3390/v13050873.](https://doi.org/10.3390/v13050873)

Ebrahimi Chaharom, F., Pourafkari, L., Ebrahimi Chaharom, A.A., Nader, N.D., 2022. Effects of corticosteroids on Covid-19 patients: a systematic review and metaanalysis on clinical outcomes. Pulm. Pharmacol. Therapeut. 72, 102107 [https://doi.](https://doi.org/10.1016/j.pupt.2021.102107) [org/10.1016/j.pupt.2021.102107](https://doi.org/10.1016/j.pupt.2021.102107).

Eid, R.A., Elgendy, M.O., El-Gendy, A.O., Elgendy, S.O., Belbahri, L., Sayed, A.M., Rateb, M.E., 2021. Efficacy of ceftazidime and cefepime in the management of COVID-19 patients: single center report from Egypt. Antibiotics 10, 1278. [https://](https://doi.org/10.3390/antibiotics10111278) doi.org/10.3390/antibiotics10111278.

Elahi, R., Karami, P., Heidary, A.H., Esmaeilzadeh, A., 2022. An updated overview of recent advances, challenges, and clinical considerations of IL-6 signaling blockade in severe coronavirus disease 2019 (COVID-19). Int. Immunopharm., 108536 [https://](https://doi.org/10.1016/j.intimp.2022.108536) [doi.org/10.1016/j.intimp.2022.108536.](https://doi.org/10.1016/j.intimp.2022.108536)

[Elazor, M., Liu, M., McKenna, S., Liu, P., Gehrig, E.A., Elfert, A., Puglisi, J., Rossignol, J.-](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref80) [F., Glenn, J.S., 2008. Nitazoxanide \(ntz\) is an inducer Eif2A and Pkr](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref80) [phosphorylation. In: Hepatology. JOHN WILEY](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref80) & SONS INC 111 RIVER ST, [HOBOKEN, NJ 07030 USA, 1151A-1151A](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref80).

El-Behery, H., Attia, A.-F., El-Feshawy, N., Torkey, H., 2021. Efficient machine learning model for predicting drug-target interactions with case study for Covid-19. Comput. Biol. Chem. 93, 107536 [https://doi.org/10.1016/j.compbiolchem.2021.107536.](https://doi.org/10.1016/j.compbiolchem.2021.107536)

Elebeedy, D., Elkhatib, W.F., Kandeil, A., Ghanem, A., Kutkat, O., Alnajjar, R., Saleh, M. A., Abd El Maksoud, A.I., Badawy, I., Al-Karmalawy, A.A., 2021. Anti-SARS-CoV-2 activities of tanshinone IIA, carnosic acid, rosmarinic acid, salvianolic acid, baicalein, and glycyrrhetinic acid between computational and in vitro insights. RSC Adv. 11, 29267–29286. <https://doi.org/10.1039/D1RA05268C>.

Ellinger, B., Bojkova, D., Zaliani, A., Cinatl, J., Claussen, C., Westhaus, S., Keminer, O., Reinshagen, J., Kuzikov, M., Wolf, M., others, 2021. A SARS-CoV-2 cytopathicity dataset generated by high-content screening of a large drug repurposing collection. Sci. Data 8, 1–10. [https://doi.org/10.1038/s41597-021-00848-4.](https://doi.org/10.1038/s41597-021-00848-4)

Evans, S.E., Tseng, C.-T.K., Scott, B.L., Höök, A.M., Dickey, B.F., 2020. Inducible epithelial resistance against coronavirus pneumonia in mice. Am. J. Respir. Cell Mol. Biol. 63, 540–541.<https://doi.org/10.1165/rcmb.2020-0247LE>.

[Ezer, N., Belga, S., Daneman, N., Chan, A., Smith, B.M., Daniels, S.-A., Moran, K.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref85) [Besson, C., Smyth, L.Y., Bartlett, S.J., others, 2021. Inhaled and intranasal](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref85) [ciclesonide for the treatment of covid-19 in adult outpatients: CONTAIN phase II](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref85) [randomised controlled trial. Br. Med. J. 375](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref85).

Farag, A., Wang, P., Boys, I.N., L. Eitson, J., Ohlson, M.B., Fan, W., McDougal, M.B., Ahmed, M., Schoggins, J, W., Sadek, H., 2020. Identification of atovaquone, Ouabain and mebendazole as FDA approved drugs tar-geting SARS-CoV-2 (version 4) (preprint). Chemistry. [https://doi.org/10.26434/chemrxiv.12003930.v4.](https://doi.org/10.26434/chemrxiv.12003930.v4)

Farhangnia, P., Dehrouyeh, S., Safdarian, A.R., Farahani, S.V., Gorgani, M., Rezaei, N., Akbarpour, M., Delbandi, A.-A., 2022. Recent advances in passive immunotherapies for COVID-19: the Evidence-Based approaches and clinical trials. Int. Immunopharm., 108786 <https://doi.org/10.1016/j.intimp.2022.108786>.

Fatima, S.W., Alam, S., Khare, S.K., 2022. Molecular and structural insights of -boswellic acid and glycyrrhizic acid as potent SARS-CoV-2 Envelope protein inhibitors. Phytomedicine 2, 100241. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.phyplu.2022.100241) [phyplu.2022.100241.](https://doi.org/10.1016/j.phyplu.2022.100241)

Fenizia, C., Galbiati, S., Vanetti, C., Vago, R., Clerici, M., Tacchetti, C., Daniele, T., 2022. Cyclosporine A inhibits viral infection and release as well as cytokine production in lung cells by three SARS-CoV-2 variants. Microbiol. Spectr. 10, e01504–e01521. <https://doi.org/10.1128/spectrum.01504-21>.

Fillmore, N., Bell, S., Shen, C., Nguyen, V., La, J., Dubreuil, M., Strymish, J., Brophy, M., Mehta, G., Wu, H., Lieberman, J., Do, N., Sander, C., 2021. Disulfiram associated with lower risk of Covid-19: a retrospective cohort study (preprint). Epidemiology. <https://doi.org/10.1101/2021.03.10.21253331>.

Fink, K., Nitsche, A., Neumann, M., Grossegesse, M., Eisele, K.-H., Danysz, W., 2021. Amantadine inhibits SARS-CoV-2 in vitro. Viruses 13, 539. [https://doi.org/10.3390/](https://doi.org/10.3390/v13040539) v₁₃₀₄₀₅₃₉

Fintelman-Rodrigues, N., Sacramento, C.Q., Ribeiro Lima, C., Souza da Silva, F., Ferreira, A.C., Mattos, M., de Freitas, C.S., Cardoso Soares, V., da Silva Gomes Dias, S., Temerozo, J.R., others, 2020. Atazanavir, alone or in combination with ritonavir, inhibits SARS-CoV-2 replication and proinflammatory cytokine production. Antimicrob. Agents Chemother. 64 [https://doi.org/10.1128/](https://doi.org/10.1128/AAC.00825-20) [AAC.00825-20](https://doi.org/10.1128/AAC.00825-20) e00825-20.

[Fong, C., 2020. Covid-19: Predicting Inhibition of SARS-CoV-2 in Caco-2 Cells: Viral](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref93) [Cellular Entry \(PhD Thesis\). Eigenenergy, Adelaide, Australia](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref93).

[Foo, C.S., Abdelnabi, R., Kaptein, S.J., Zhang, X., ter Horst, S., Mols, R., Delang, L.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref94) [Rocha-Pereira, J., Coelmont, L., Leyssen, P., others, 2021. Nelfinavir Markedly](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref94) [Improves Lung Pathology in SARS-CoV-2-Infected Syrian Hamsters Despite a Lack of](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref94) [an Antiviral Effect. bioRxiv](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref94).

Fornasier, E., Macchia, M.L., Giachin, G., Sosic, A., Pavan, M., Sturlese, M., Salata, C., Moro, S., Gatto, B., Bellanda, M., others, 2022. A new inactive conformation of SARS-CoV-2 main protease. Acta Crystallogr. D: Struct. Biol. 78 [https://doi.org/](https://doi.org/10.1107/S2059798322000948) [10.1107/S2059798322000948.](https://doi.org/10.1107/S2059798322000948)

[Franchin, G., 2021. IRAK 4 Inhibitor \(PF-06650833\) in Hospitalized Patients with](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref96) [COVID-19 Pneumonia and Exuberant Inflammation.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref96)

Fred, S.M., Kuivanen, S., Ugurlu, H., Casarotto, P.C., Levanov, L., Saksela, K., Vapalahti, O., Castrén, E., 2022. Antidepressant and antipsychotic drugs reduce viral infection by SARS-CoV-2 and fluoxetine shows antiviral activity against the novel variants in vitro. Front. Pharmacol. 12, 755600 https://doi.org/10.3389/ [fphar.2021.755600](https://doi.org/10.3389/fphar.2021.755600).

Freedberg, D.E., Conigliaro, J., Wang, T.C., Tracey, K.J., Callahan, M.V., Abrams, J.A., Sobieszczyk, M.E., Markowitz, D.D., Gupta, A., O'Donnell, M.R., others, 2020. Famotidine use is associated with improved clinical outcomes in hospitalized COVID-19 patients: a propensity score matched retrospective cohort study. Gastroenterology 159, 1129–1131. [https://doi.org/10.1053/j.gastro.2020.05.053.](https://doi.org/10.1053/j.gastro.2020.05.053)

Fu, Z., Huang, B., Tang, J., Liu, S., Liu, M., Ye, Y., Liu, Z., Xiong, Y., Zhu, W., Cao, D., others, 2021. The complex structure of GRL0617 and SARS-CoV-2 PLpro reveals a hot spot for antiviral drug discovery. Nat. Commun. 12, 1-12. https://doi.org/ [10.1038/s41467-020-20718-8.](https://doi.org/10.1038/s41467-020-20718-8)

Fung, S.-Y., Siu, K.-L., Lin, H., Chan, C.-P., Yeung, M.L., Jin, D.-Y., 2022. SARS-CoV-2 NSP13 helicase suppresses interferon signaling by perturbing JAK1 phosphorylation of STAT1. Cell Biosci. 12, 1–12. <https://doi.org/10.1186/s13578-022-00770-1>.

Galvez-Romero, J.L., Palmeros-Rojas, O., Real-Ramírez, F.A., Sánchez-Romero, S., Tome-Maxil, R., Ramírez-Sandoval, M.P., Olivos-Rodríguez, R., Flores-Encarnación, S.E., Cabrera-Estrada, A.A., Ávila-Morales, J., others, 2021. Cyclosporine A plus low-dose steroid treatment in COVID-19 improves clinical outcomes in patients with moderate to severe disease: a pilot study. J. Intern. Med. 289, 906-920. https://doi.org/ [10.1111/joim.13223](https://doi.org/10.1111/joim.13223).

Gandhi, S., Klein, J., Robertson, A.J., Peña-Hernández, M.A., Lin, M.J., Roychoudhury, P., Lu, P., Fournier, J., Ferguson, D., Mohamed Bakhash, S.A., others, 2022. De novo emergence of a remdesivir resistance mutation during treatment of persistent SARS-CoV-2 infection in an immunocompromised patient: a case report. Nat. Commun. 13, 1–8. [https://doi.org/10.1038/s41467-022-29104-y.](https://doi.org/10.1038/s41467-022-29104-y)

Gao, J., Zhang, L., Liu, X., Li, F., Ma, R., Zhu, Z., Zhang, J., Wu, J., Shi, Y., Pan, Y., Ge, Y., Ruan, K., 2020. Repurposing low-molecular-weight drugs against the main protease of severe acute respiratory syndrome coronavirus 2. J. Phys. Chem. Lett. 11, 7267–7272. [https://doi.org/10.1021/acs.jpclett.0c01894.](https://doi.org/10.1021/acs.jpclett.0c01894)

Garcia, G., Sharma, A., Ramaiah, A., Sen, C., Purkayastha, A., Kohn, D.B., Parcells, M.S., Beck, S., Kim, H., Bakowski, M.A., Kirkpatrick, M.G., Riva, L., Wolff, K.C., Han, B., Yuen, C., Ulmert, D., Purbey, P.K., Scumpia, P., Beutler, N., Rogers, T.F., Chatterjee, A.K., Gabriel, G., Bartenschlager, R., Gomperts, B., Svendsen, C.N., Betz, U.A.K., Damoiseaux, R.D., Arumugaswami, V., 2021. Antiviral drug screen identifies DNA-damage response inhibitor as potent blocker of SARS-CoV-2 replication. Cell Rep. 35, 108940<https://doi.org/10.1016/j.celrep.2021.108940>.

Gautret, P., Lagier, J.-C., Parola, P., Hoang, V.T., Meddeb, L., Mailhe, M., Doudier, B., Courjon, J., Giordanengo, V., Vieira, V.E., Tissot Dupont, H., Honoré, S., Colson, P., Chabrière, E., La Scola, B., Rolain, J.-M., Brouqui, P., Raoult, D., 2020. Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial. Int. J. Antimicrob. Agents 56, 105949. <https://doi.org/10.1016/j.ijantimicag.2020.105949>.

Gentile, I., Schiano Moriello, N., 2022. COVID-19 prophylaxis in immunosuppressed patients: beyond vaccination. PLoS Med. 19, e1003917 [https://doi.org/10.1371/](https://doi.org/10.1371/journal.pmed.1003917) [journal.pmed.1003917](https://doi.org/10.1371/journal.pmed.1003917).

Geriak, M., Haddad, F., Kullar, R., Greenwood, K.L., Habib, M., Habib, C., Willms, D., Sakoulas, G., 2021. Randomized prospective open label study shows no impact on clinical outcome of adding losartan to hospitalized COVID-19 patients with mild hypoxemia. Infect Dis. Therp. 10, 1323–1330. [https://doi.org/10.1007/s40121-021-](https://doi.org/10.1007/s40121-021-00453-3)

[00453-3](https://doi.org/10.1007/s40121-021-00453-3). Ghasemiyeh, P., Mortazavi, N., Karimzadeh, I., Vazin, A., Mahmoudi, L., Moghimi Sarani, E., MohammadSadeghi, A., Shahisavandi, M., Kheradmand, A., Mohammadi-Samani, S., 2021. Psychiatric adverse drug reactions and potential anti-COVID-19 drug interactions with psychotropic medications. IJPR 20. [https://doi.org/](https://doi.org/10.22037/ijpr.2021.114717.15007) [10.22037/ijpr.2021.114717.15007.](https://doi.org/10.22037/ijpr.2021.114717.15007)

Gimeno, A., Mestres-Truyol, J., Ojeda-Montes, M.J., Macip, G., Saldivar-Espinoza, B., Cereto-Massagué, A., Pujadas, G., Garcia-Vallvé, S., 2020. Prediction of novel

inhibitors of the main protease (M-pro) of SARS-CoV-2 through consensus docking and drug reposition. IJMS 21, 3793. https://doi.org/10.3390/ijms2111379.

- Giossi, R., Menichelli, D., Pani, A., Tratta, E., Romandini, A., Roncato, R., Nani, A., Schenardi, P., Diani, E., Fittipaldo, V.A., Farcomeni, A., Scaglione, F., Pastori, D., 2021. A systematic review and a meta-analysis comparing prophylactic and therapeutic low molecular weight heparins for mortality reduction in 32,688 COVID-19 patients. Front. Pharmacol. 12, 698008 [https://doi.org/10.3389/](https://doi.org/10.3389/fphar.2021.698008) [fphar.2021.698008](https://doi.org/10.3389/fphar.2021.698008).
- [Gobeil, S., Janowska, K., McDowell, S., Mansouri, K., Parks, R., Manne, K., Stalls, V.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref111) [Kopp, M., Henderson, R., Edwards, R.J., others, 2020. D614G Mutation Alters SARS-](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref111)[CoV-2 Spike Conformational Dynamics and Protease Cleavage Susceptibility at the](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref111) [S1/S2 Junction. bioRxiv.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref111)
- Gomaa, A.A., Mohamed, H.S., Abd-Ellatief, R.B., Gomaa, M.A., Hammam, D.S., 2022. Advancing combination treatment with glycyrrhizin and boswellic acids for hospitalized patients with moderate COVID-19 infection: a randomized clinical trial. Inflammopharmacology 30, 477–486. [https://doi.org/10.1007/s10787-022-00939-](https://doi.org/10.1007/s10787-022-00939-7) [7](https://doi.org/10.1007/s10787-022-00939-7).
- Good, S.S., Westover, J., Jung, K.H., Zhou, X.-J., Moussa, A., La Colla, P., Collu, G., Canard, B., Sommadossi, J.-P., 2021. AT-527, a double prodrug of a guanosine nucleotide analog, is a potent inhibitor of SARS-CoV-2 in vitro and a promising oral antiviral for treatment of COVID-19. Antimicrob. Agents Chemother. 65 [https://doi.](https://doi.org/10.1128/AAC.02479-20) [org/10.1128/AAC.02479-20](https://doi.org/10.1128/AAC.02479-20) e02479-20.
- Grellet, E., Goulet, A., Imbert, I., 2022. Replication of the coronavirus genome: a paradox among positive-strand RNA viruses. J. Biol. Chem., 101923 [https://doi.org/](https://doi.org/10.1016/j.jbc.2022.101923) [10.1016/j.jbc.2022.101923.](https://doi.org/10.1016/j.jbc.2022.101923)
- Gu, Y., Cao, J., Zhang, X., Gao, H., Wang, Y., Wang, J., He, J., Jiang, X., Zhang, J., Shen, G., Yang, J., Zheng, X., Hu, G., Zhu, Yuanfei, Du, S., Zhu, Yunkai, Zhang, R., Xu, J., Lan, F., Qu, D., Xu, G., Zhao, Y., Gao, D., Xie, Y., Luo, M., Lu, Z., 2022. Receptome profiling identifies KREMEN1 and ASGR1 as alternative functional receptors of SARS-CoV-2. Cell Res. 32, 24–37. [https://doi.org/10.1038/s41422-021-](https://doi.org/10.1038/s41422-021-00595-6) [00595-6](https://doi.org/10.1038/s41422-021-00595-6).
- Gupta, A., Chun, H.J., 2021. Interleukin-1-Receptor kinase 4 inhibition: achieving immunomodulatory synergy to mitigate the impact of COVID-19. Front. Immunol. 2483 [https://doi.org/10.3389/fimmu.2021.693085.](https://doi.org/10.3389/fimmu.2021.693085)
- Gupta, T., Thakkar, P., Kalra, B., Kannan, S., 2022. Hydroxychloroquine in the treatment of coronavirus disease 2019: rapid updated systematic review and meta-analysis. Rev. Med. Virol. 32, e2276 <https://doi.org/10.1002/rmv.2276>.
- Gupta, Y., Kumar, S., Zak, S.E., Jones, K.A., Upadhyay, C., Sharma, N., Azizi, S.-A., Kathayat, R.S., Herbert, A.S., Durvasula, R., others, 2021a. Antiviral evaluation of hydroxyethylamine analogs: inhibitors of SARS-CoV-2 main protease (3CLpro), a virtual screening and simulation approach. Bioorg. Med. Chem. 116393 [https://doi.](https://doi.org/10.1016/j.bmc.2021.116393) [org/10.1016/j.bmc.2021.116393](https://doi.org/10.1016/j.bmc.2021.116393).
- Gupta, Y., Maciorowski, D., Medernach, B., Becker, D.P., Durvasula, R., Libertin, C.R., Kempaiah, P., 2022. Iron dysregulation in COVID-19 and reciprocal evolution of SARS-CoV-2: natura nihil frustra facit. J. Cell. Biochem. [https://doi.org/10.1002/](https://doi.org/10.1002/jcb.30207) [jcb.30207.](https://doi.org/10.1002/jcb.30207)
- Gupta, Y., Maciorowski, D., Zak, S.E., Jones, K.A., Kathayat, R.S., Azizi, S.-A., Mathur, R., Pearce, C.M., Ilc, D.J., Husein, H., others, 2021b. Bisindolylmaleimide IX: a novel anti-SARS-CoV2 agent targeting viral main protease 3CLpro demonstrated by virtual screening pipeline and in-vitro validation assays. Methods. https://doi.org/10.1016/ [j.ymeth.2021.01.003.](https://doi.org/10.1016/j.ymeth.2021.01.003)
- Gupta, Y., Maciorowski, D., Zak, S.E., Kulkarni, C.V., Herbert, A.S., Durvasula, R., Fareed, J., Dye, J.M., Kempaiah, P., 2021c. Heparin: a simplistic repurposing to prevent SARS-CoV-2 transmission in light of its in-vitro nanomolar efficacy. Int. J. Biol. Macromol. 183, 203–212. [https://doi.org/10.1016/j.ijbiomac.2021.04.148.](https://doi.org/10.1016/j.ijbiomac.2021.04.148)
- Hamdy, R., Mostafa, A., Abo Shama, N.M., Soliman, S.S., Fayed, B., 2022. Comparative Evaluation of Flavonoids Reveals the Superiority and Promising Inhibition Activity of Silibinin against SARS-CoV-2. Phytotherapy Research. [https://doi.org/10.1002/](https://doi.org/10.1002/ptr.7486) [ptr.7486.](https://doi.org/10.1002/ptr.7486)
- Hammond, J., Leister-Tebbe, H., Gardner, A., Abreu, P., Bao, W., Wisemandle, W., Baniecki, M., Hendrick, V.M., Damle, B., Simón-Campos, A., Pypstra, R., Rusnak, J. M., 2022. Oral nirmatrelvir for high-risk, nonhospitalized adults with covid-19. N. Engl. J. Med. 386, 1397–1408. [https://doi.org/10.1056/NEJMoa2118542.](https://doi.org/10.1056/NEJMoa2118542)
- Han, Y., Duan, X., Yang, L., Nilsson-Payant, B.E., Wang, P., Duan, F., Tang, X., Yaron, T. M., Zhang, T., Uhl, S., Bram, Y., Richardson, C., Zhu, J., Zhao, Z., Redmond, D., Houghton, S., Nguyen, D.-H.T., Xu, D., Wang, X., Jessurun, J., Borczuk, A., Huang, Y., Johnson, J.L., Liu, Y., Xiang, J., Wang, H., Cantley, L.C., tenOever, B.R., Ho, D.D., Pan, F.C., Evans, T., Chen, H.J., Schwartz, R.E., Chen, S., 2021. Identification of SARS-CoV-2 inhibitors using lung and colonic organoids. Nature 589, 270–275. [https://doi.org/10.1038/s41586-020-2901-9.](https://doi.org/10.1038/s41586-020-2901-9)
- [Hariyanto, T.I., Halim, D.A., Rosalind, J., Gunawan, C., Kurniawan, A., 2022. Ivermectin](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref125) [and outcomes from Covid-19 pneumonia: a systematic review and meta-analysis of](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref125) [randomized clinical trial studies. Rev. Med. Virol. 32, e2265](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref125).
- Harrison, C., 2020. Focus shifts to antibody cocktails for COVID-19 cytokine storm. Nat. Biotechnol. 38, 905–909.<https://doi.org/10.1038/s41587-020-0634-9>.
- Harrison, S., Thumm, L., Nash, T.E., Nutman, T.B., O'Connell, E.M., 2021. The local inflammatory profile and predictors of treatment success in subarachnoid neurocysticercosis. Clin. Infect. Dis. 72, e326–e333. [https://doi.org/10.1093/cid/](https://doi.org/10.1093/cid/ciaa1128) [ciaa1128](https://doi.org/10.1093/cid/ciaa1128).
- Hempel, T., Elez, K., Krüger, N., Raich, L., Shrimp, J.H., Danov, O., Jonigk, D., Braun, A., Shen, M., Hall, M.D., Pöhlmann, S., Hoffmann, M., Noé, F., 2021. Synergistic inhibition of SARS-CoV-2 cell entry by otamixaban and covalent protease inhibitors: pre-clinical assessment of pharmacological and molecular properties. Chem. Sci. 12, 12600–12609. [https://doi.org/10.1039/D1SC01494C.](https://doi.org/10.1039/D1SC01494C)
- Hijikata, A., Shionyu-Mitsuyama, C., Nakae, S., Shionyu, M., Ota, M., Kanaya, S., Hirokawa, T., Nakajima, S., Watashi, K., Shirai, T., 2022. Evaluating cepharanthine

analogues as natural drugs against SARS-CoV-2. FEBS Open bio 12, 285–294. [https://doi.org/10.1002/2211-5463.13337.](https://doi.org/10.1002/2211-5463.13337)

- Hines, S.L., Mohammad, A.N., Jackson, J., Macklin, S., Caulfield, T.R., 2019a. Integrative data fusion for comprehensive assessment of a novel CHEK2 variant using combined genomics, imaging, and functional-structural assessments via protein informatics. Mol Omics 15, 59–66. <https://doi.org/10.1039/c8mo00137e>.
- Hines, S.L., Richter, J.E., Mohammad, A.N., Mahim, J., Atwal, P.S., Caulfield, T.R., 2019b. Protein informatics combined with multiple data sources enriches the clinical characterization of novel TRPV4 variant causing an intermediate skeletal dysplasia. Mol Genet Genomic Med 7, e566. <https://doi.org/10.1002/mgg3.566>.

Hinton, G., 2007. Boltzmann machine. Scholarpedia 2, 1668. [https://doi.org/10.4249/](https://doi.org/10.4249/scholarpedia.1668) [scholarpedia.1668](https://doi.org/10.4249/scholarpedia.1668).

- Hoffmann, M., Kleine-Weber, H., Schroeder, S., Krüger, N., Herrler, T., Erichsen, S., Schiergens, T.S., Herrler, G., Wu, N.-H., Nitsche, A., others, 2020. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. Cell. <https://doi.org/10.1016/j.cell.2020.02.052>.
- Hong, M., Mandala, V., McKay, M., Shcherbakov, A., Dregni, A., Kolocouris, A., 2020. Structure and Drug Binding of the SARS-CoV-2 Envelope Protein in Phospholipid Bilayers. [https://doi.org/10.21203/rs.3.rs-77124/v1.](https://doi.org/10.21203/rs.3.rs-77124/v1)
- Hong, S., Wang, H., Zhang, Z., Qiao, L., 2022. The roles of methylprednisolone treatment in patients with COVID-19: a systematic review and meta-analysis. Steroids 183, 109022. [https://doi.org/10.1016/j.steroids.2022.109022.](https://doi.org/10.1016/j.steroids.2022.109022)
- Hsu, C.-K., Chen, C.-Y., Chen, W.-C., Lai, C.-C., Hung, S.-H., Lin, W.-T., 2022. The effect of sofosbuvir-based treatment on the clinical outcomes of patients with COVID-19: a systematic review and meta-analysis of randomized controlled trials. Int. J. Antimicrob. Agents 106545. [https://doi.org/10.1016/j.ijantimicag.2022.106545.](https://doi.org/10.1016/j.ijantimicag.2022.106545)
- Hu, K., Wang, M., Zhao, Y., Zhang, Y., Wang, T., Zheng, Z., Li, X., Zeng, S., Zhao, D., Li, H., others, 2020. A small-scale medication of leflunomide as a treatment of COVID-19 in an open-label blank-controlled clinical trial. Virol. Sin. 35, 725–733. <https://doi.org/10.1007/s12250-020-00258-7>.
- [Hu, Z., van der Ploeg, K., Chakraborty, S., Arunachalam, P., Mori, D., Jacobson, K.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref138) [Bonilla, H., Parsonnet, J., Andrews, J., Hedlin, H., others, 2021. Early immune](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref138) [responses have long-term associations with clinical, virologic, and immunologic](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref138) [outcomes in patients with COVID-19. Research Square.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref138)
- [Hung, D.T., Ghula, S., Aziz, J.M.A., Makram, A.M., Tawfik, G.M., Abozaid, A.A.-F.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref139) [Pancharatnam, R.A., Ibrahim, A.M., Shabouk, M.B., Turnage, M., others, 2022. The](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref139) [efficacy and adverse effects of favipiravir on COVID-19 patients: a systematic review](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref139) [and meta-analysis of published clinical trials and observational studies. Int. J. Infect.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref139) [Dis.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref139)
- Imai, Y., Kuba, K., Rao, S., Huan, Y., Guo, F., Guan, B., Yang, P., Sarao, R., Wada, T., Leong-Poi, H., others, 2005. Angiotensin-converting enzyme 2 protects from severe acute lung failure. Nature 436, 112–116.<https://doi.org/10.1038/nature03712>.
- Imamura, K., Sakurai, Y., Enami, T., Shibukawa, R., Nishi, Y., Ohta, A., Shu, T., Kawaguchi, J., Okada, S., Hoenen, T., Yasuda, J., Inoue, H., 2021. iPSC screening for drug repurposing identifies anti-RNA virus agents modulating host cell susceptibility. FEBS Open Bio 11, 1452–1464. [https://doi.org/10.1002/2211-](https://doi.org/10.1002/2211-5463.13153) [5463.13153.](https://doi.org/10.1002/2211-5463.13153)
- Ivanova, N., Sotirova, Y., Gavrailov, G., Nikolova, K., Andonova, V., 2022. Advances in the prophylaxis of respiratory infections by the nasal and the Oromucosal route: relevance to the fight with the SARS-CoV-2 pandemic. Pharmaceutics 14, 530. <https://doi.org/10.3390/pharmaceutics14030530>.
- Ivashchenko, A.A., Dmitriev, K.A., Vostokova, N.V., Azarova, V.N., Blinow, A.A., Egorova, A.N., Gordeev, I.G., Ilin, A.P., Karapetian, R.N., Kravchenko, D.V., Lomakin, N.V., Merkulova, E.A., Papazova, N.A., Pavlikova, E.P., Savchuk, N.P., Simakina, E.N., Sitdekov, T.A., Smolyarchuk, E.A., Tikhomolova, E.G., Yakubova, E. V., Ivachtchenko, A.V., 2021. AVIFAVIR for treatment of patients with moderate coronavirus disease 2019 (COVID-19): interim results of a phase II/III multicenter randomized clinical trial. Clin. Infect. Dis. 73, 531–534. [https://doi.org/10.1093/](https://doi.org/10.1093/cid/ciaa1176) [cid/ciaa1176.](https://doi.org/10.1093/cid/ciaa1176)
- Jackson, C.B., Farzan, M., Chen, B., Choe, H., 2022. Mechanisms of SARS-CoV-2 entry into cells. Nat. Rev. Mol. Cell Biol. 23, 3–20. [https://doi.org/10.1038/s41580-021-](https://doi.org/10.1038/s41580-021-00418-x) [00418-x.](https://doi.org/10.1038/s41580-021-00418-x)
- Jang, K.-J., Jeong, S., Kang, D.Y., Sp, N., Yang, Y.M., Kim, D.-E., 2020. A high ATP concentration enhances the cooperative translocation of the SARS coronavirus helicase nsP13 in the unwinding of duplex RNA. Sci. Rep. 10, 1-13. https://doi.org/ [10.1038/s41598-020-61432-1.](https://doi.org/10.1038/s41598-020-61432-1)
- Jaroenram, W., Chatnuntawech, I., Kampeera, J., Pengpanich, S., Leaungwutiwong, P., Tondee, B., Sirithammajak, S., Suvannakad, R., Khumwan, P., Dangtip, S., Arunrut, N., Bantuchai, S., Nguitragool, W., Wongwaroran, S., Khanchaitit, P., Sattabongkot, J., Teerapittayanon, S., Kiatpathomchai, W., 2022. One-step colorimetric isothermal detection of COVID-19 with AI-assisted automated result analysis: a platform model for future emerging point-of-care RNA/DNA disease diagnosis. Talanta 249, 123375. [https://doi.org/10.1016/j.talanta.2022.123375.](https://doi.org/10.1016/j.talanta.2022.123375)
- Jayk Bernal, A., Gomes da Silva, M.M., Musungaie, D.B., Kovalchuk, E., Gonzalez, A., Delos Reyes, V., Martín-Quirós, A., Caraco, Y., Williams-Diaz, A., Brown, M.L., others, 2022. Molnupiravir for oral treatment of Covid-19 in nonhospitalized patients. N. Engl. J. Med. 386, 509–520. [https://doi.org/10.1056/NEJMoa2116044.](https://doi.org/10.1056/NEJMoa2116044)
- [Jeong, G.U., Lyu, J., Kim, K.-D., Chung, Y.C., Yoon, G.Y., Lee, S., Hwang, I., Shin, W.-H.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref148) [Ko, J., Lee, J.-Y., others, 2022. SARS-CoV-2 infection of microglia elicits](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref148) [proinflammatory activation and apoptotic cell death. Microbiol. Spectr. e01091-22.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref148)
- Jiménez-Luna, J., Grisoni, F., Weskamp, N., Schneider, G., 2021. Artificial intelligence in drug discovery: recent advances and future perspectives. Expet Opin. Drug Discov. 16, 949–959. [https://doi.org/10.1080/17460441.2021.1909567.](https://doi.org/10.1080/17460441.2021.1909567)
- Jin, Z., Du, X., Xu, Y., Deng, Y., Liu, M., Zhao, Y., Zhang, B., Li, X., Zhang, L., Peng, C., Duan, Y., Yu, J., Wang, L., Yang, K., Liu, F., Jiang, R., Yang, Xinglou, You, T., Liu, Xiaoce, Yang, Xiuna, Bai, F., Liu, H., Liu, Xiang, Guddat, L.W., Xu, W., Xiao, G.,

Qin, C., Shi, Z., Jiang, H., Rao, Z., Yang, H., 2020. Structure of Mpro from SARS-CoV-2 and discovery of its inhibitors. Nature 582, 289–293. [https://doi.org/10.1038/](https://doi.org/10.1038/s41586-020-2223-y) [s41586-020-2223-y.](https://doi.org/10.1038/s41586-020-2223-y)

- Kalantari, S., Fard, S.R., Maleki, D., Taher, M.T., Yassin, Z., Alimohamadi, Y., Minaeian, S., 2021. Comparing the effectiveness of Atazanavir/Ritonavir/ Dolutegravir/Hydroxychloroquine and Lopinavir/Ritonavir/Hydroxychloroquine treatment regimens in COVID-19 patients. J. Med. Virol. 93, 6557–6565. [https://doi.](https://doi.org/10.1002/jmv.27195) [org/10.1002/jmv.27195.](https://doi.org/10.1002/jmv.27195)
- Kamel, W.A., Kamel, M.I., Alhasawi, A., Elmasry, S., AlHamdan, F., Al-Hashel, J.Y., 2021. Effect of pre-exposure use of amantadine on COVID-19 infection: a hospital-based cohort study in patients with Parkinson's disease or multiple sclerosis. Front. Neurol. 12, 704186 [https://doi.org/10.3389/fneur.2021.704186.](https://doi.org/10.3389/fneur.2021.704186)
- Kandeel, M., Yamamoto, M., Tani, H., Kobayashi, A., Gohda, J., Kawaguchi, Y., Park, B. K., Kwon, H.-J., Inoue, J., Alkattan, A., 2021. Discovery of new fusion inhibitor peptides against SARS-CoV-2 by targeting the spike S2 subunit. Biomolecules & therapeutics 29, 282. [https://doi.org/10.4062/biomolther.2020.201.](https://doi.org/10.4062/biomolther.2020.201)
- Kaur, H., Sarma, P., Bhattacharyya, A., Sharma, S., Chhimpa, N., Prajapat, M., Prakash, A., Kumar, S., Singh, A., Singh, R., others, 2021. Efficacy and safety of dihydroorotate dehydrogenase (DHODH) inhibitors "leflunomide" and "teriflunomide" in Covid-19: a narrative review. Eur. J. Pharmacol. 906, 174233 <https://doi.org/10.1016/j.ejphar.2021.174233>.
- Kayode, O., Wang, R., Pendlebury, D.F., Cohen, I., Henin, R.D., Hockla, A., Soares, A.S., Papo, N., Caulfield, T.R., Radisky, E.S., 2016. An acrobatic substrate metamorphosis reveals a requirement for substrate conformational dynamics in trypsin proteolysis. J. Biol. Chem. 291, 26304–26319.<https://doi.org/10.1074/jbc.M116.758417>.
- Kern, D.M., Sorum, B., Hoel, C.M., Sridharan, S., Remis, J.P., Toso, D.B., Brohawn, S.G., n.d. Cryo-EM structure of the SARS-CoV-2 3a ion channel in lipid nanodiscs. [htt](https://doi.org/10.1101/2020.06.17.156554) [ps://doi.org/10.1101/2020.06.17.156554.](https://doi.org/10.1101/2020.06.17.156554)
- [Kinoshita, T., Shinoda, M., Nishizaki, Y., Shiraki, K., Hirai, Y., Kichikawa, Y.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref157) [Tsushima, K., Sinkai, M., Komura, N., Yoshida, K., others, 2022. Phase 3,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref157) [Multicentre, Double-Blind, Randomised, Parallel-Group, Placebo-Controlled Study of](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref157) [Camostat Mesilate \(FOY-305\) for the Treatment of COVID-19 \(CANDLE Study\).](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref157) [medRxiv](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref157).
- Ko, M., Jeon, S., Ryu, W.-S., Kim, S., 2021. Comparative analysis of antiviral efficacy of FDA-approved drugs against SARS-CoV-2 in human lung cells. J. Med. Virol. 93, 1403–1408. [https://doi.org/10.1002/jmv.26397.](https://doi.org/10.1002/jmv.26397)
- Kocabas¸, F., Uslu, M., 2021. The current state of validated small molecules inhibiting SARS-CoV-2 non-structural proteins. Turkish J. Biol. 45, 469–483. [https://doi.org/](https://doi.org/10.3906/biy-2106-42) [10.3906/biy-2106-42](https://doi.org/10.3906/biy-2106-42).
- Kosinsky, Y., Peskov, K., Stanski, D.R., Wetmore, D., Vinetz, J., 2022. Semi-Mechanistic pharmacokinetic-pharmacodynamic model of Camostat mesylate-predicted efficacy against SARS-CoV-2 in COVID-19. Microbiol. Spectr. 10 [https://doi.org/10.1128/](https://doi.org/10.1128/spectrum.02167-21) ectrum.02167-21 e02167-21.
- Kow, C.S., Javed, A., Ramachandram, D., Hasan, S.S., 2022. Clinical outcomes of sofosbuvir-based antivirals in patients with COVID-19: a systematic review and meta-analysis of randomized trials. Expert Rev. Anti-infect. Ther. 20, 567–575. <https://doi.org/10.1080/14787210.2022.2000861>.
- Kumar, Prateek, Bhardwaj, T., Garg, N., Giri, R., 2022a. Microsecond simulations and CD spectroscopy reveals the intrinsically disordered nature of SARS-CoV-2 spike-Cterminal cytoplasmic tail (residues 1242–1273) in isolation. Virology 566, 42–55.
- [https://doi.org/10.1016/j.virol.2021.11.005.](https://doi.org/10.1016/j.virol.2021.11.005) Kumar, Prateek, Bhardwaj, T., Giri, R., 2022b. Mitoxantrone dihydrochloride, an FDA approved drug, binds with SARS-CoV-2 NSP1 C-terminal. RSC Adv. 12, 5648–5655. [https://doi.org/10.1039/D1RA07434B.](https://doi.org/10.1039/D1RA07434B)
- Kumar, Prabhakaran, Mathayan, M., Smieszek, S.P., Przychodzen, B.P., Koprivica, V., Birznieks, G., Polymeropoulos, M.H., Prabhakar, B.S., 2022. Identification of potential COVID-19 treatment compounds which inhibit SARS Cov2 prototypic, Delta and Omicron variant infection. Virology. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.virol.2022.05.004) [virol.2022.05.004](https://doi.org/10.1016/j.virol.2022.05.004).
- Kumar, S., Singh, B., Kumari, P., Kumar, P.V., Agnihotri, G., Khan, S., Kant Beuria, T., Syed, G.H., Dixit, A., 2021. Identification of multipotent drugs for COVID-19 therapeutics with the evaluation of their SARS-CoV2 inhibitory activity. Comput. Struct. Biotechnol. J. 19, 1998–2017. <https://doi.org/10.1016/j.csbj.2021.04.014>.
- Kwon, P.S., Oh, H., Kwon, S.-J., Jin, W., Zhang, F., Fraser, K., Hong, J.J., Linhardt, R.J., Dordick, J.S., 2020. Sulfated polysaccharides effectively inhibit SARS-CoV-2 in vitro. Cell. Discov. 6, 1–4. <https://doi.org/10.1038/s41421-020-00192-8>.
- Kyrou, I., Randeva, H.S., Spandidos, D.A., Karteris, E., 2021. Not only ACE2—the quest for additional host cell mediators of SARS-CoV-2 infection: neuropilin-1 (NRP1) as a novel SARS-CoV-2 host cell entry mediator implicated in COVID-19. Signal Transduct. Targeted Ther. 6, 1–3.<https://doi.org/10.1038/s41392-020-00460-9>.
- Lai, M.Y., Bukhari, F.D.M., Zulkefli, N.Z., Ismail, I., Mustapa, N.I., Soh, T.S.T., Hassan, A. H., Peariasamy, K.M., Lee, Y.L., Suppiah, J., Thayan, R., Lau, Y.L., 2022. Colorimetric detection of SARS-CoV-2 by uracil-DNA glycosylase (UDG) reverse transcription loop-mediated isothermal amplification (RT-LAMP). Int. J. Infect. Dis. 120, 132–134. <https://doi.org/10.1016/j.ijid.2022.04.036>.
- [Lam, E., Sayedy, N., Dasgupta, N., Akella, J., Iqbal, J., 2022. COVID-19-Related Diffuse](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref169) [Leukoencephalopathy clinical improvement with amantadine therapy. In: B24.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref169) [REPORTING ON COVID-19 AND ITS COMPLICATIONS. American Thoracic Society.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref169) A2478–[A2478](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref169).

[Lamb, Y.N., 2022. Nirmatrelvir plus Ritonavir: first approval. Drugs 1](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref170)–7.

Lan, Q., Wang, C., Zhou, J., Wang, L., Jiao, F., Zhang, Y., Cai, Y., Lu, L., Xia, S., Jiang, S., 2021. 25-Hydroxycholesterol-Conjugated EK1 peptide with potent and broadspectrum inhibitory activity against SARS-CoV-2, its variants of concern, and other human coronaviruses. Int. J. Mol. Sci. 22, 11869 https://doi.org/10.3390 [ijms222111869.](https://doi.org/10.3390/ijms222111869)

- Laponogov, I., Gonzalez, G., Shepherd, M., Qureshi, A., Veselkov, D., Charkoftaki, G., Vasiliou, V., Youssef, J., Mirnezami, R., Bronstein, M., Veselkov, K., 2021. Network machine learning maps phytochemically rich "Hyperfoods" to fight COVID-19. Hum. Genom. 15, 1. https://doi.org/10.1186/s40246-020-00297
- Lau, E.Y., Negrete, O.A., Bennett, W.F.D., Bennion, B.J., Borucki, M., Bourguet, F., Epstein, A., Franco, M., Harmon, B., He, S., Jones, D., Kim, H., Kirshner, D., Lao, V., Lo, J., McLoughlin, K., Mosesso, R., Murugesh, D.K., Saada, E.A., Segelke, B., Stefan, M.A., Stevenson, G.A., Torres, M.W., Weilhammer, D.R., Wong, S., Yang, Y., Zemla, A., Zhang, X., Zhu, F., Allen, J.E., Lightstone, F.C., 2021. Discovery of smallmolecule inhibitors of SARS-CoV-2 proteins using a computational and experimental pipeline. Front. Mol. Biosci. 8, 678701 [https://doi.org/10.3389/](https://doi.org/10.3389/fmolb.2021.678701) [fmolb.2021.678701](https://doi.org/10.3389/fmolb.2021.678701).
- Lehrer, S., Rheinstein, P.H., 2021. Homozygosity for rs17775810 minor allele associated with reduced mortality of COVID-19 in the UK Biobank Cohort. In Vivo 35, 965–968. <https://doi.org/10.21873/invivo.12338>.
- Leung, W.F., Chorlton, S., Tyson, J., Al-Rawahi, G.N., Jassem, A.N., Prystajecky, N., Masud, S., Deans, G.D., Chapman, M.G., Mirzanejad, Y., others, 2022. COVID-19 in an immunocompromised host: persistent shedding of viable SARS-CoV-2 and emergence of multiple mutations: a case report. Int. J. Infect. Dis. 114, 178–182. [https://doi.org/10.1016/j.ijid.2021.10.045.](https://doi.org/10.1016/j.ijid.2021.10.045)
- Lewis, D.S., Ho, J., Wills, S., Kawall, A., Sharma, A., Chavada, K., Ebert, M.C., Evoli, S., Singh, A., Rayalam, S., others, 2022. Aloin isoforms (A and B) selectively inhibits proteolytic and deubiquitinating activity of papain like protease (PLpro) of SARS-CoV-2 in vitro. Sci. Rep. 12, 1–11. <https://doi.org/10.1038/s41598-022-06104-y>.
- Li, J., Chen, G., Meng, Z., Wu, Z., Gan, H., Zhu, X., Han, P., Liu, T., Wang, F., Gu, R., others, 2022. Bioavailability enhancement of cepharanthine via pulmonary administration in rats and its therapeutic potential for pulmonary fibrosis associated with COVID-19 infection. Molecules 27, 2745. [https://doi.org/10.3390/](https://doi.org/10.3390/molecules27092745) [molecules27092745](https://doi.org/10.3390/molecules27092745).
- [Li, S., Zhang, Y., Guan, Z., Li, H., Ye, M., Chen, X., Shen, J., Zhou, Y., Shi, Z.-L., Zhou, P.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref178) [others, 2020. SARS-CoV-2 triggers inflammatory responses and cell death through](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref178) [caspase-8 activation. Signal Transduct. Targeted Ther. 5, 1](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref178)–10.
- Lima, A.N., Philot, E.A., Trossini, G.H.G., Scott, L.P.B., Maltarollo, V.G., Honorio, K.M., 2016. Use of machine learning approaches for novel drug discovery. Expet Opin. Drug Discov. 11, 225–239. [https://doi.org/10.1517/17460441.2016.1146250.](https://doi.org/10.1517/17460441.2016.1146250)
- [Lin, C., Li, Yue, Zhang, Y., Liu, Z., Mu, X., Gu, C., Liu, J., Li, Yutang, Li, G., Chen, J., 2021.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref180) [Ceftazidime is a potential drug to inhibit SARS-CoV-2 infection in vitro by blocking](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref180) spike protein–[ACE2 interaction. Signal Transduct. Targeted Ther. 6, 1](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref180)–4.
- Lin, H., Cherukupalli, S., Feng, D., Gao, S., Kang, D., Zhan, P., Liu, X., 2022. SARS-CoV-2 Entry inhibitors targeting virus-ACE2 or virus-TMPRSS2 interactions. Curr. Med. Chem. 29, 682–699. [https://doi.org/10.2174/0929867328666210420103021.](https://doi.org/10.2174/0929867328666210420103021)
- Lin, Y., Zhang, Z., Mahjour, B., Wang, D., Zhang, R., Shim, E., McGrath, A., Shen, Y., Brugger, N., Turnbull, R., others, 2021. Reinforcing the supply chain of umifenovir and other antiviral drugs with retrosynthetic software. Nat. Commun. 12, 1–8. <https://doi.org/10.1038/s41467-021-27547-3>.
- Liu, X., Wang, X.-J., 2020. Potential inhibitors against 2019-nCoV coronavirus M protease from clinically approved medicines. Journal of Genetics and Genomics 47, 119. [https://doi.org/10.1016/j.jgg.2020.02.001.](https://doi.org/10.1016/j.jgg.2020.02.001)
- Lo, H.S., Hui, K.P.Y., Lai, H.-M., He, X., Khan, K.S., Kaur, S., Huang, J., Li, Z., Chan, A.K., Cheung, H.H.-Y., others, 2021. Simeprevir potently suppresses SARS-CoV-2 replication and synergizes with remdesivir. ACS Cent. Sci. 7, 792–802. [https://doi.](https://doi.org/10.1021/acscentsci.0c01186) [org/10.1021/acscentsci.0c01186.](https://doi.org/10.1021/acscentsci.0c01186)
- Loffredo, M., Lucero, H., Chen, D.-Y., O'Connell, A., Bergqvist, S., Munawar, A., Bandara, A., De Graef, S., Weeks, S.D., Douam, F., others, 2021. The in-vitro effect of famotidine on sars-cov-2 proteases and virus replication. Sci. Rep. 11, 1–9. [https://](https://doi.org/10.1038/s41598-021-84782-w) [doi.org/10.1038/s41598-021-84782-w.](https://doi.org/10.1038/s41598-021-84782-w)
- Luban, J., Sattler, R.A., Mühlberger, E., Graci, J.D., Cao, L., Weetall, M., Trotta, C., Colacino, J.M., Bavari, S., Strambio-De-Castillia, C., others, 2021. The DHODH inhibitor PTC299 arrests SARS-CoV-2 replication and suppresses induction of inflammatory cytokines. Virus Res. 292, 198246 https://doi.org/10.1016/ [virusres.2020.198246](https://doi.org/10.1016/j.virusres.2020.198246).
- Lucas, S., 2016. The pharmacology of indomethacin. Headache. The Journal of Head and Face Pain 56, 436–446. [https://doi.org/10.1111/head.12769.](https://doi.org/10.1111/head.12769)
- Ma, C., Hu, Y., Townsend, J.A., Lagarias, P.I., Marty, M.T., Kolocouris, A., Wang, J., 2020a. Ebselen, disulfiram, carmofur, PX-12, tideglusib, and shikonin are nonspecific promiscuous SARS-CoV-2 main protease inhibitors. ACS Pharmacol. Transl. Sci. 3, 1265–1277. [https://doi.org/10.1021/acsptsci.0c00130.](https://doi.org/10.1021/acsptsci.0c00130)
- Ma, C., Sacco, M.D., Hurst, B., Townsend, J.A., Hu, Y., Szeto, T., Zhang, X., Tarbet, B., Marty, M.T., Chen, Y., Wang, J., 2020b. Boceprevir, GC-376, and calpain inhibitors II, XII inhibit SARS-CoV-2 viral replication by targeting the viral main protease. Cell Res. 30, 678–692. <https://doi.org/10.1038/s41422-020-0356-z>.
- Maghzi, A.H., Houtchens, M.K., Preziosa, P., Ionete, C., Beretich, B.D., Stankiewicz, J.M., Tauhid, S., Cabot, A., Berriosmorales, I., Schwartz, T.H., others, 2020. COVID-19 in teriflunomide-treated patients with multiple sclerosis. J. Neurol. 267, 2790–2796. <https://doi.org/10.1007/s00415-020-09944-8>.
- Mahdi, M., Mótyán, J.A., Szojka, Z.I., Golda, M., Miczi, M., Tőzsér, J., 2020. Analysis of the efficacy of HIV protease inhibitors against SARS-CoV-2's main protease. Virol. J. 17, 190. [https://doi.org/10.1186/s12985-020-01457-0.](https://doi.org/10.1186/s12985-020-01457-0)
- Mahmoud, A., Mostafa, A., Al-Karmalawy, A.A., Zidan, A., Abulkhair, H.S., Mahmoud, S. H., Shehata, M., Elhefnawi, M.M., Ali, M.A., 2021. Telaprevir is a potential drug for repurposing against SARS-CoV-2: computational and in vitro studies. Heliyon 7, e07962. <https://doi.org/10.1016/j.heliyon.2021.e07962>.
- Malchair, P., Otero, A., Giol, J., Solanich, X., Carnaval, T., Fernández-Nistal, A., Sánchez-Gabriel, A., Montoto, C., Lleonart, R., Videla, S., 2022. A multicenter, open-label, randomized, proof-of-concept phase II clinical trial to assess the efficacy and safety of icatibant in patients infected with SARS-CoV-2 (COVID-19) and admitted to

hospital units without invasive mechanical ventilation: study protocol (ICAT-COVID). Trials 23, 1–15, https://doi.org/10.1186/s13063-022-06219-7 COVID). Trials 23, 1-15. https://doi.org/10.1186/s13063-022

Malone, R.W., Tisdall, P., Fremont-Smith, P., Liu, Y., Huang, X.-P., White, K.M., Miorin, L., Moreno, E., Alon, A., Delaforge, E., others, 2021. COVID-19: famotidine, histamine, mast cells, and mechanisms. Front. Pharmacol. 12, 633680 https://doi. [org/10.3389/fphar.2021.633680.](https://doi.org/10.3389/fphar.2021.633680)

Mandala, V.S., McKay, M.J., Shcherbakov, A.A., Dregni, A.J., Kolocouris, A., Hong, M., 2020. Structure and drug binding of the SARS-CoV-2 envelope protein transmembrane domain in lipid bilayers. Nat. Struct. Mol. Biol. 27, 1202–1208. <https://doi.org/10.1038/s41594-020-00536-8>.

Manna, S., Chowdhury, T., Baindara, P., Mandal, S.M., 2020. Fusion protein targeted antiviral peptides: fragment-based drug design (FBDD) guided rational design of dipeptides against SARS-CoV-2. Curr. Protein Pept. Sci. 21, 938–947. [https://doi.](https://doi.org/10.2174/1389203721666200908164641) [org/10.2174/1389203721666200908164641](https://doi.org/10.2174/1389203721666200908164641).

Martin, R., Löchel, H.F., Welzel, M., Hattab, G., Hauschild, A.-C., Heider, D., 2020. CORDITE: the curated CORona drug InTERactions database for SARS-CoV-2. iScience 23, 101297. [https://doi.org/10.1016/j.isci.2020.101297.](https://doi.org/10.1016/j.isci.2020.101297)

Mashayekhi-Sardoo, H., Hosseinjani, H., 2022. A new application of mTOR inhibitor drugs as potential therapeutic agents for COVID-19. J. Basic Clin. Physiol. Pharmacol. 33, 17–25. <https://doi.org/10.1515/jbcpp-2020-0495>.

Matsuyama, S., Kawase, M., Nao, N., Shirato, K., Ujike, M., Kamitani, W., Shimojima, M., Fukushi, S., 2020. The inhaled steroid ciclesonide blocks SARS-CoV-2 RNA replication by targeting the viral replication-transcription complex in cultured cells. J. Virol. 95 <https://doi.org/10.1128/JVI.01648-20>e01648-20.

[Mazaherpour, H., Sofian, M., Farahani, E., Abdi, A., Mazaherpour, S., Larijani, M.S., abd](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref200) [Ramezani, A., 2021. Higher rate of hyperbilirubinemia and arrythmia in COVID-19](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref200) [cases receiving combination therapy atazanavir/ritonavir vs. Lopinavir/ritonavir.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref200)

Mellott, D.M., Tseng, C.-T., Drelich, A., Fajtová, P., Chenna, B.C., Kostomiris, D.H., Hsu, J., Zhu, J., Taylor, Z.W., Kocurek, K.I., Tat, V., Katzfuss, A., Li, L., Giardini, M. A., Skinner, D., Hirata, K., Yoon, M.C., Beck, S., Carlin, A.F., Clark, A.E., Beretta, L., Maneval, D., Hook, V., Frueh, F., Hurst, B.L., Wang, H., Raushel, F.M., O'Donoghue, A.J., de Siqueira-Neto, J.L., Meek, T.D., McKerrow, J.H., 2021. A clinical-stage cysteine protease inhibitor blocks SARS-CoV-2 infection of human and monkey cells. ACS Chem. Biol. 16, 642–650. [https://doi.org/10.1021/](https://doi.org/10.1021/acschembio.0c00875) c00875.

Mendieta Zerón, H., Meneses Calderón, J., Paniagua Coria, L., Meneses Figueroa, J., Vargas Contreras, M.J., Vives Aceves, H.L., Carranza Salazar, F.M., Californias Hernández, D., Miraflores Vidaurri, E., Carrillo González, A., others, 2021. Nitazoxanide as an early treatment to reduce the intensity of COVID-19 outbreaks among health personnel. World Academy of Sciences Journal 3, 1–6. [https://doi.](https://doi.org/10.3892/wasj.2021.94) [org/10.3892/wasj.2021.94](https://doi.org/10.3892/wasj.2021.94).

Merat, S., 2020. SD1000: high sustained viral response rate in 1361 patients with hepatitis C genotypes 1, 2, 3, and 4 using a low-cost, fixed-dose combination tablet of generic sofosbuvir and daclatasvir: a multicenter, phase III clinical trial. Clin. Infect. Dis. 70, 2206–2212.<https://doi.org/10.1093/cid/ciz628>.

Messina, V., Nevola, R., Izzi, A., De Lucia Sposito, P., Marrone, A., Rega, R., Fusco, R., Lumino, P., Rinaldi, L., Gaglione, P., others, 2022. Efficacy and safety of the sofosbuvir/velpatasvir combination for the treatment of patients with early mild to moderate COVID-19. Sci. Rep. 12, 1–6. [https://doi.org/10.1038/s41598-022-09741-](https://doi.org/10.1038/s41598-022-09741-5) [5](https://doi.org/10.1038/s41598-022-09741-5).

Meyer, B., Chiaravalli, J., Gellenoncourt, S., Brownridge, P., Bryne, D.P., Daly, L.A., Grauslys, A., Walter, M., Agou, F., Chakrabarti, L.A., Craik, C.S., Eyers, C.E., Eyers, P. A., Gambin, Y., Jones, A.R., Sierecki, E., Verdin, E., Vignuzzi, M., Emmott, E., 2021. Characterising proteolysis during SARS-CoV-2 infection identifies viral cleavage sites and cellular targets with therapeutic potential. Nat. Commun. 12, 5553. [https://doi.](https://doi.org/10.1038/s41467-021-25796-w) [org/10.1038/s41467-021-25796-w.](https://doi.org/10.1038/s41467-021-25796-w)

Minasov, G., Rosas-Lemus, M., Shuvalova, L., Inniss, N.L., Brunzelle, J.S., Daczkowski, C. M., Hoover, P., Mesecar, A.D., Satchell, K.J., 2021. Mn2+ coordinates Cap-0-RNA to align substrates for efficient 2′ -O-methyl transfer by SARS-CoV-2 nsp16. Sci. Signal. 14, eabh2071 <https://doi.org/10.1126/scisignal.abh2071>.

Miorin, L., Mire, C., Ranjbar, S., Hume, A., Huang, J., Crossland, N., White, K., Laporte, M., Kehrer, T., Haridas, V., others, n.d. The Oral Drug Nitazoxanide Restricts SARS-CoV-2 Infection and Attenuates Disease Pathogenesis in Syrian Hamsters (preprint).

Monteil, V., Kwon, H., Prado, P., Hagelkrüys, A., Wimmer, R.A., Stahl, M., Leopoldi, A., Garreta, E., Del Pozo, C.H., Prosper, F., others, 2020. Inhibition of SARS-CoV-2 infections in engineered human tissues using clinical-grade soluble human ACE2.

Cell 181, 905–913. [https://doi.org/10.1016/j.cell.2020.04.004.](https://doi.org/10.1016/j.cell.2020.04.004) Moran-Lev, H., Weisman, Y., Cohen, S., Deutsch, V., Cipok, M., Bondar, E., Lubetzky, R., Mandel, D., 2018. The interrelationship between hepcidin, vitamin D, and anemia in children with acute infectious disease. Pediatr. Res. 84, 62-65. https://doi.org [10.1038/s41390-018-0005-0](https://doi.org/10.1038/s41390-018-0005-0).

Mostafa, A., Kandeil, A., A. M. M. Elshaier, Y., Kutkat, O., Moatasim, Y., Rashad, A.A., Shehata, M., Gomaa, M.R., Mahrous, N., Mahmoud, S.H., GabAllah, M., Abbas, H., Taweel, A.E., Kayed, A.E., Kamel, M.N., Sayes, M.E., Mahmoud, D.B., El-Shesheny, R., Kayali, G., Ali, M.A., 2020. FDA-approved drugs with potent in vitro antiviral activity against severe acute respiratory syndrome coronavirus 2. Pharmaceuticals 13, 443. <https://doi.org/10.3390/ph13120443>.

Mostafa, M.A., 2020. Role of Zidovudine and Candesartan in the Novel SARS-CoV-2 Treatment Trials: Theoretical Study (Preprint). [https://doi.org/10.21467/](https://doi.org/10.21467/preprints.30) [preprints.30](https://doi.org/10.21467/preprints.30).

Narayanan, A., Narwal, M., Majowicz, S.A., Varricchio, C., Toner, S.A., Ballatore, C., Brancale, A., Murakami, K.S., Jose, J., 2022. Identification of SARS-CoV-2 inhibitors targeting Mpro and PLpro using in-cell-protease assay. Commun Biol 5, 169. [https://](https://doi.org/10.1038/s42003-022-03090-9) doi.org/10.1038/s42003-022-03090-9.

Nassar, A., Ibrahim, I.M., Amin, F.G., Magdy, M., Elgharib, A.M., Azzam, E.B., Nasser, F., Yousry, K., Shamkh, I.M., Mahdy, S.M., others, 2021. A review of human

coronaviruses' receptors: the host-cell targets for the crown bearing viruses. Molecules 26, 6455. [https://doi.org/10.3390/molecules26216455.](https://doi.org/10.3390/molecules26216455)

Nicastri, E., Marinangeli, F., Pivetta, E., Torri, E., Reggiani, F., Fiorentino, G., Scorzolini, L., Vettori, S., Marsiglia, C., Gavioli, E.M., Beccari, A.R., Terpolilli, G., De Pizzol, M., Goisis, G., Mantelli, F., Vaia, F., Allegretti, M., 2022. A phase 2 randomized, double-blinded, placebo-controlled, multicenter trial evaluating the efficacy and safety of raloxifene for patients with mild to moderate COVID-19. eClinicalMedicine 48, 101450. [https://doi.org/10.1016/j.eclinm.2022.101450.](https://doi.org/10.1016/j.eclinm.2022.101450)

Ohashi, H., Watashi, K., Saso, W., Shionoya, K., Iwanami, S., Hirokawa, T., Shirai, T., Kanaya, S., Ito, Y., Kim, K.S., Nomura, T., Suzuki, Tateki, Nishioka, K., Ando, S., Ejima, K., Koizumi, Y., Tanaka, T., Aoki, S., Kuramochi, K., Suzuki, Tadaki, Hashiguchi, T., Maenaka, K., Matano, T., Muramatsu, M., Saijo, M., Aihara, K., Iwami, S., Takeda, M., McKeating, J.A., Wakita, T., 2021. Potential anti-COVID-19 agents, cepharanthine and nelfinavir, and their usage for combination treatment. iScience 24, 102367. [https://doi.org/10.1016/j.isci.2021.102367.](https://doi.org/10.1016/j.isci.2021.102367)

Ono, J., Koshimizu, U., Fukunishi, Y., Nakai, H., 2022. Multiple protonation states in ligand-free SARS-CoV-2 main protease revealed by large-scale quantum molecular dynamics simulations. Chem. Phys. Lett. 794, 139489 [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.cplett.2022.139489) [cplett.2022.139489.](https://doi.org/10.1016/j.cplett.2022.139489)

Pandey, P., Prasad, K., Prakash, A., Kumar, V., 2020. Insights into the biased activity of dextromethorphan and haloperidol towards SARS-CoV-2 NSP6: in silico binding mechanistic analysis. J. Mol. Med. 98, 1659–1673. [https://doi.org/10.1007/s00109-](https://doi.org/10.1007/s00109-020-01980-1) [020-01980-1.](https://doi.org/10.1007/s00109-020-01980-1)

Patel, L., Shukla, T., Huang, X., Ussery, D.W., Wang, S., 2020. Machine learning methods in drug discovery. Molecules 25, E5277. [https://doi.org/10.3390/](https://doi.org/10.3390/molecules25225277) [molecules25225277](https://doi.org/10.3390/molecules25225277).

[Patidar, V., Sharma, A., Garg, V., Tripathi, A.P., Dhaneriya, S., 2022. Methylene blue in](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref219) [management of COVID19. J. Assoc. Phys. India 70, 11](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref219)–12.

- Pearlman, B.L., 2012. Protease inhibitors for the treatment of chronic hepatitis C genotype-1 infection: the new standard of care. Lancet Infect. Dis. 12, 717–728. [https://doi.org/10.1016/S1473-3099\(12\)70060-9.](https://doi.org/10.1016/S1473-3099(12)70060-9)
- Pfefferle, S., Schöpf, J., Kögl, M., Friedel, C.C., Müller, M.A., Carbajo-Lozoya, J., Stellberger, T., von Dall'Armi, E., Herzog, P., Kallies, S., others, 2011. The SARScoronavirus-host interactome: identification of cyclophilins as target for pancoronavirus inhibitors. PLoS Pathog. 7, e1002331 [https://doi.org/10.1371/journal.](https://doi.org/10.1371/journal.ppat.1002331) [ppat.1002331](https://doi.org/10.1371/journal.ppat.1002331).
- [Pitts, J., Li, J., Perry, J.K., Du Pont, V., Riola, N., Rodriguez, L., Lu, X., Kurhade, C.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref222) [Xie, X., Camus, G., others, 2022. Remdesivir and GS-441524 retain antiviral activity](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref222) [against Delta, Omicron, and other emergent SARS-CoV-2 variants. Antimicrob.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref222) [Agents Chemother., e00222-22](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref222)

Ponti, G., Roli, L., Oliva, G., Manfredini, M., Trenti, T., Kaleci, S., Iannella, R., Balzano, B., Coppola, A., Fiorentino, G., others, 2021. Homocysteine (Hcy) assessment to predict outcomes of hospitalized Covid-19 patients: a multicenter study on 313 Covid-19 patients. Clin. Chem. Lab. Med. 59, e354–e357. [https://doi.](https://doi.org/10.1515/cclm-2021-0168) [org/10.1515/cclm-2021-0168](https://doi.org/10.1515/cclm-2021-0168).

Prajapat, M., Sarma, P., Shekhar, N., Avti, P., Sinha, S., Kaur, H., Kumar, S., Bhattacharyya, A., Kumar, H., Bansal, S., Medhi, B., 2020. Drug targets for corona virus: a systematic review. Indian J. Pharmacol. 52, 56–65. [https://doi.org/](https://doi.org/10.4103/ijp.IJP_115_20) [10.4103/ijp.IJP_115_20](https://doi.org/10.4103/ijp.IJP_115_20).

[Protein kinase inhibitors. In: LiverTox: Clinical and Research Information on Drug-](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref225)[Induced Liver Injury, 2012. National Institute of Diabetes and Digestive and Kidney](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref225) [Diseases, Bethesda \(MD\)](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref225).

Puschmann, A., Fiesel, F.C., Caulfield, T.R., Hudec, R., Ando, M., Truban, D., Hou, X., Ogaki, K., Heckman, M.G., James, E.D., Swanberg, M., Jimenez-Ferrer, I., Hansson, O., Opala, G., Siuda, J., Boczarska-Jedynak, M., Friedman, A., Koziorowski, D., Rudzińska-Bar, M., Aasly, J.O., Lynch, T., Mellick, G.D., Mohan, M., Silburn, P.A., Sanotsky, Y., Vilariño-Güell, C., Farrer, M.J., Chen, L., Dawson, V.L., Dawson, T.M., Wszolek, Z.K., Ross, O.A., Springer, W., 2017. Heterozygous PINK1 p. G411S increases risk of Parkinson's disease via a dominant-negative mechanism. Brain 140, 98–117. <https://doi.org/10.1093/brain/aww261>.

Puskarich, M.A., Cummins, N.W., Ingraham, N.E., Wacker, D.A., Reilkoff, R.A., Driver, B. E., Biros, M.H., Bellolio, F., Chipman, J.G., Nelson, A.C., others, 2021. A multi-center phase II randomized clinical trial of losartan on symptomatic outpatients with COVID-19. EClinicalMedicine 37, 100957. https://doi.org/10.1016/ [eclinm.2021.100957.](https://doi.org/10.1016/j.eclinm.2021.100957)

Puskarich, M.A., Ingraham, N.E., Merck, L.H., Driver, B.E., Wacker, D.A., Black, L.P., Jones, A.E., Fletcher, C.V., South, A.M., Murray, T.A., others, 2022. Efficacy of losartan in hospitalized patients with COVID-19–induced lung injury: a randomized clinical trial. JAMA Netw. Open 5. [https://doi.org/10.1001/](https://doi.org/10.1001/jamanetworkopen.2022.2735) amanetworkopen.2022.2735 e222735-e222735.

Rabie, A.M., 2021. Teriflunomide: a possible effective drug for the comprehensive treatment of COVID-19. Current Research in Pharmacology and Drug Discovery 2, 100055. <https://doi.org/10.1016/j.crphar.2021.100055>.

Ramachandran, R., Bhosale, V., Reddy, H., Atam, V., Faridi, M., Fatima, J., Shukla, V., Khan, Z.A., Khan, H., Singh, V., Negi, M.P.S., Srivastava, M., Srivastava, A.K., Tripathi, C.B., Ghosh, N., Majumdar, N., Tripathi, R.K., Rath, S.K., Mishra, P.R., Sharma, S., Kundu, T.K., 2022. Phase III, randomized, double-blind, placebo controlled trial of efficacy, safety and tolerability of antiviral drug umifenovir vs standard care of therapy in non-severe COVID-19 patients. Int. J. Infect. Dis. 115, 62–69. <https://doi.org/10.1016/j.ijid.2021.11.025>.

[Ramos Jr., F., Zeze, V., Velut, S., Jan, M., 1987. Cystic meningiomas. Practical value of a](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref231) [radio-surgical classification. J. Neuroradiol. 14 \(3\), 271](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref231)–286.

Rapicavoli, R.V., Alaimo, S., Ferro, A., Pulvirenti, A., 2022. Computational methods for drug repurposing. Adv. Exp. Med. Biol. 1361, 119–141. [https://doi.org/10.1007/](https://doi.org/10.1007/978-3-030-91836-1_7) [978-3-030-91836-1_7](https://doi.org/10.1007/978-3-030-91836-1_7).

- Ravichandran, R., Mohan, S.K., Sukumaran, S.K., Kamaraj, D., Daivasuga, S.S., Ravi, S.O. A.S., Vijayaraghavalu, S., Kumar, R.K., 2022. An open label randomized clinical trial of Indomethacin for mild and moderate hospitalised Covid-19 patients. Sci. Rep. 12, 1–10.<https://doi.org/10.1038/s41598-022-10370-1>.
- Reichen, F.R., Dawson, K.M., Lewis, S., Steiner, D., Amstutz, P., Engler, O., Stumpp, M.T., Stumpp, M.T., n.d. Multi-specific DARPin® Therapeutics Demonstrate Very High Potency against Mutated SARS-CoV-2 Variants in Vitro.
- [Reina, J., Iglesias, C., 2022. Nirmatrelvir Plus Ritonavir \(Paxlovid\) a Potent SARS-CoV-2](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref235) [3CLpro Protease Inhibitor Combination. Revista Espanola de Quimioterapia:](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref235) [Publicacion Oficial de la Sociedad Espanola de Quimioterapia](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref235).
- Reis, G., Silva, E.A.S.M., Silva, D.C.M., Thabane, L., Milagres, A.C., Ferreira, T.S., dos Santos, C.V.Q., Campos, V.H.S., Nogueira, A.M.R., de Almeida, A.P.F.G., Callegari, E. D., Neto, A.D.F., Savassi, L.C.M., Simplicio, M.I.C., Ribeiro, L.B., Oliveira, R., Harari, O., Forrest, J.I., Ruton, H., Sprague, S., McKay, P., Guo, C.M., Rowland-Yeo, K., Guyatt, G.H., Boulware, D.R., Rayner, C.R., Mills, E.J., 2022. Effect of early treatment with ivermectin among patients with covid-19. N. Engl. J. Med. 386, 1721–1731.<https://doi.org/10.1056/NEJMoa2115869>.
- Rejdak, K., Grieb, P., 2020. Adamantanes might be protective from COVID-19 in patients with neurological diseases: multiple sclerosis, parkinsonism and cognitive impairment. Multiple sclerosis and related disorders 42, 102163. [https://doi.org/](https://doi.org/10.1016/j.msard.2020.102163) [10.1016/j.msard.2020.102163](https://doi.org/10.1016/j.msard.2020.102163).
- Ren, Z., Luo, H., Yu, Z., Song, J., Liang, L., Wang, L., Wang, H., Cui, G., Liu, Y., Wang, J., others, 2020. A randomized, open-label, controlled clinical trial of azvudine tablets in the treatment of mild and common COVID-19, a pilot study. Adv. Sci. 7, 2001435 <https://doi.org/10.1002/advs.202001435>.
- Reznikov, L.R., Norris, M.H., Vashisht, R., Bluhm, A.P., Li, D., Liao, Y.-S.J., Brown, A., Butte, A.J., Ostrov, D.A., 2021. Identification of antiviral antihistamines for COVID-19 repurposing. Biochem. Biophys. Res. Commun. 538, 173–179. [https://doi.org/](https://doi.org/10.1016/j.bbrc.2020.11.095) [10.1016/j.bbrc.2020.11.095.](https://doi.org/10.1016/j.bbrc.2020.11.095)
- Riccio, A.A., Sullivan, E.D., Copeland, W.C., 2022. Activation of the SARS-CoV-2 NSP14 3′ –5′ exoribonuclease by NSP10 and response to antiviral inhibitors. J. Biol. Chem. 298 [https://doi.org/10.1016/j.jbc.2021.101518.](https://doi.org/10.1016/j.jbc.2021.101518)
- Robinson, P.C., Liew, D.F., Tanner, H.L., Grainger, J.R., Dwek, R.A., Reisler, R.B., Steinman, L., Feldmann, M., Ho, L.-P., Hussell, T., others, 2022. COVID-19 therapeutics: challenges and directions for the future. Proc. Natl. Acad. Sci. USA 119, e2119893119. [https://doi.org/10.1073/pnas.2119893119.](https://doi.org/10.1073/pnas.2119893119)
- Rocco, P.R., Silva, P.L., Cruz, F.F., Melo-Junior, M.A.C., Tierno, P.F., Moura, M.A., De Oliveira, L.F.G., Lima, C.C., Dos Santos, E.A., Junior, W.F., others, 2021. Early use of nitazoxanide in mild Covid-19 disease: randomised, placebo-controlled trial. Eur. Respir. J. 58 <https://doi.org/10.1183/13993003.03725-2020>.
- Rossignol, J.-F., Bardin, M.C., Fulgencio, J., Mogelnicki, D., Bréchot, C., 2022. A randomized double-blind placebo-controlled clinical trial of nitazoxanide for treatment of mild or moderate COVID-19. eClinicalMedicine 45, 101310. [https://](https://doi.org/10.1016/j.eclinm.2022.101310) doi.org/10.1016/j.eclinm.2022.101310.
- Ruggiero, V., Aquino, R.P., Del Gaudio, P., Campiglia, P., Russo, P., 2022. Post-COVID syndrome: the research progress in the treatment of pulmonary sequelae after COVID-19 infection. Pharmaceutics 14, 1135. [https://doi.org/10.3390/](https://doi.org/10.3390/pharmaceutics14061135) [pharmaceutics14061135](https://doi.org/10.3390/pharmaceutics14061135).
- Ruhm, C.J., 2022. Excess deaths in the United States during the first year of COVID-19. Prev. Med. 107174 <https://doi.org/10.1016/j.ypmed.2022.107174>.
- Russo, L.C., Tomasin, R., Matos, I.A., Manucci, A.C., Sowa, S.T., Dale, K., Caldecott, K.W., Lehtiö, L., Schechtman, D., Meotti, F.C., others, 2021. The SARS-CoV-2 Nsp3 macrodomain reverses PARP9/DTX3L-dependent ADP-ribosylation induced by interferon signaling. J. Biol. Chem. 297 [https://doi.org/10.1016/j.jbc.2021.101041.](https://doi.org/10.1016/j.jbc.2021.101041)
- Salvarani, C., Massari, M., Costantini, M., Franco Merlo, D., Lucia Mariani, G., Viale, P., Nava, S., Guaraldi, G., Dolci, G., Boni, L., Savoldi, L., Bruzzi, P., Turra, ` C., Catanoso, M., Maria Marata, A., Barbieri, C., Valcavi, A., Franzoni, F., Cavuto, S., Mazzi, G., Corsini, R., Trapani, F., Bartoloni, A., Barisione, E., Barbieri, C., Jole Burastero, G., Pan, A., Inojosa, W., Scala, R., Burattini, C., Luppi, F., Codeluppi, M., Eldin Tarek, K., Cenderello, G., Salio, M., Foti, G., Dongilli, R., Bajocchi, G., Alberto Negri, E., Ciusa, G., Fornaro, G., Bassi, I., Zammarchi, L., Aloè, T., Facciolongo, N., 2022. Intravenous methylprednisolone pulses in hospitalised patients with severe COVID-19 pneumonia, A double-blind, randomised, placebo-controlled trial. Eur. Respir. J., 2200025 [https://doi.org/10.1183/13993003.00025-2022.](https://doi.org/10.1183/13993003.00025-2022)
- Savytskyi, O.V., Yesylevskyy, S.O., Kornelyuk, A.I., 2013. Asymmetric structure and domain binding interfaces of human tyrosyl-tRNA synthetase studied by molecular dynamics simulations. J. Mol. Recogn. 26 (2), 113–120. [https://doi:10.1002/jmr.22](https://doi:10.1002/jmr.2259) [59.](https://doi:10.1002/jmr.2259)
- Savytskyi, O., Kornelyuk, A., 2022. Computational modeling of the complex between glycyrrhizin and SARS-CoV-2 protease 3CLpro as a target for the development of antiviral drugs. Reports of the National Academy of Sciences of Ukraine. [https://doi.](https://doi.org/10.15407/dopovidi2022.01.115) [org/10.15407/dopovidi2022.01.115.](https://doi.org/10.15407/dopovidi2022.01.115)
- Schloer, S., Brunotte, L., Mecate-Zambrano, A., Zheng, S., Tang, J., Ludwig, S., Rescher, U., 2021. Drug synergy of combinatory treatment with remdesivir and the repurposed drugs fluoxetine and itraconazole effectively impairs SARS-CoV-2 infection in vitro. Br. J. Pharmacol. 178, 2339–2350. [https://doi.org/10.1111/](https://doi.org/10.1111/bph.15418) [bph.15418](https://doi.org/10.1111/bph.15418).
- Schuurmans, M.M., Hage, R., 2021. Cyclosporine A and COVID-19–The COQUIMA cohort. EClinicalMedicine 31. https://doi.org/10.1016/j.eclinm.2020.10068
- Serpa Neto, A., Landoni, G., Ostermann, M., Lumlertgul, N., Forni, L., Alvarez-Belon, L., Trapani, T., Alliegro, P.V., Zacharowski, K., Wiedenbeck, C., others, 2022. Angiotensin II infusion in COVID-19: an international, multicenter, registry-based study. J. Med. Virol. 94, 2079–2088. [https://doi.org/10.1002/jmv.27592.](https://doi.org/10.1002/jmv.27592)
- Shabani, M., Sadegh Ehdaei, B., Fathi, F., Dowran, R., 2021. A mini-review on sofosbuvir and daclatasvir treatment in coronavirus disease 2019. New Microbes and New Infections 42, 100895. <https://doi.org/10.1016/j.nmni.2021.100895>.
- Shafiee, A., Teymouri Athar, M.M., Kohandel Gargari, O., Jafarabady, K., Siahvoshi, S., Mozhgani, S.-H., 2022. Ivermectin under scrutiny: a systematic review and metaanalysis of efficacy and possible sources of controversies in COVID-19 patients. Virol. J. 19, 102. https://doi.org/10.1186/s12985-022-01829-
- Shah, S.B., 2021. COVID-19 and Progesterone: Part 1. SARS-CoV-2, Progesterone and its potential clinical use. Endocrine and Metabolic Science 5, 100109. [https://doi.org/](https://doi.org/10.1016/j.endmts.2021.100109) [10.1016/j.endmts.2021.100109.](https://doi.org/10.1016/j.endmts.2021.100109)
- Shaheer, M., Singh, R., Sobhia, M.E., 2021. Protein degradation: a novel computational approach to design protein degrader probes for main protease of SARS-CoV-2. J. Biomol. Struct. Dyn. 1–13. [https://doi.org/10.1080/07391102.2021.1953601.](https://doi.org/10.1080/07391102.2021.1953601)
- Shannon, A., Fattorini, V., Sama, B., Selisko, B., Feracci, M., Falcou, C., Gauffre, P., El Kazzi, P., Delpal, A., Decroly, E., others, 2022. A dual mechanism of action of AT-527 against SARS-CoV-2 polymerase. Nat. Commun. 13, 1–9. [https://doi.org/10.1038/](https://doi.org/10.1038/s41467-022-28113-1) [s41467-022-28113-1](https://doi.org/10.1038/s41467-022-28113-1).
- Sheahan, T.P., Sims, A.C., Leist, S.R., Schäfer, A., Won, J., Brown, A.J., Montgomery, S. A., Hogg, A., Babusis, D., Clarke, M.O., others, 2020. Comparative therapeutic efficacy of remdesivir and combination lopinavir, ritonavir, and interferon beta against MERS-CoV. Nat. Commun. 11, 1–14. [https://doi.org/10.1038/s41467-019-](https://doi.org/10.1038/s41467-019-13940-6) [13940-6](https://doi.org/10.1038/s41467-019-13940-6).
- Shekhar, N., Kaur, H., Sarma, P., Prakash, A., Medhi, B., 2022. Indomethacin: an exploratory study of antiviral mechanism and host-pathogen interaction in COVID-19. Expert Rev. Anti Infect. Ther. 20, 383–390. [https://doi.org/10.1080/](https://doi.org/10.1080/14787210.2022.1990756) [14787210.2022.1990756](https://doi.org/10.1080/14787210.2022.1990756).

Shen, Z., Halberg, A., Fong, J.Y., Guo, J., Song, G., Louie, B., Luedtke, G.R., Visuthikraisee, V., Protter, A., Koh, X., Baik, T., Lum, P.Y., 2022. Elucidating host cell response pathways and repurposing therapeutics for SARS-CoV-2 and other coronaviruses using gene expression profiles of chemical and genetic perturbations (preprint). Genomics. [https://doi.org/10.1101/2022.04.18.488682.](https://doi.org/10.1101/2022.04.18.488682)

- Sherrington, D., Kirkpatrick, S., 1975. Solvable model of a spin-glass. Phys. Rev. Lett. 35, 1792–1796.<https://doi.org/10.1103/PhysRevLett.35.1792>.
- Shi, Z., Wei, J., Deng, X., Li, S., Qiu, Y., Shao, D., Li, B., Zhang, K., Xue, F., Wang, X., others, 2014. Nitazoxanide inhibits the replication of Japanese encephalitis virus in cultured cells and in a mouse model. Virol. J. 11, 1–10. [https://doi.org/10.1186/](https://doi.org/10.1186/1743-422X-11-10) [1743-422X-11-10](https://doi.org/10.1186/1743-422X-11-10).
- Shu, T., Huang, M., Wu, D., Ren, Y., Zhang, X., Han, Y., Mu, J., Wang, R., Qiu, Y., Zhang, D.-Y., others, 2020. SARS-coronavirus-2 Nsp13 possesses NTPase and RNA helicase activities that can be inhibited by bismuth salts. Virol. Sin. 35, 321–329. <https://doi.org/10.1007/s12250-020-00242-1>.
- Simeoni, M., Cavinato, T., Rodriguez, D., Gatfield, D., 2021. I (nsp1) ecting SARS-CoV-2–ribosome interactions. Communications biology 4, 1–5. [https://doi.org/10.1038/](https://doi.org/10.1038/s42003-021-02265-0) [s42003-021-02265-0](https://doi.org/10.1038/s42003-021-02265-0).
- Siragusa, L., Menna, G., Buratta, F., Baroni, M., Desantis, J., Cruciani, G., Goracci, L., 2022. CROMATIC: cro ss-Relationship Ma p of Cavi ti es from C oronaviruses. J. Chem. Inf. Model. [https://doi.org/10.1021/acs.jcim.2c00169.](https://doi.org/10.1021/acs.jcim.2c00169)
- Smieszek, S.P., Przychodzen, B.P., Polymeropoulos, M.H., 2020. Amantadine disrupts lysosomal gene expression: a hypothesis for COVID19 treatment. Int. J. Antimicrob. Agents 55, 106004. [https://doi.org/10.1016/j.ijantimicag.2020.106004.](https://doi.org/10.1016/j.ijantimicag.2020.106004)
- Song, J.-Y., Kim, Y.-S., Eom, J.-S., Kim, J.-Y., Lee, J.-S., Lee, J., Choi, W.-S., Heo, J.-Y., Sohn, J.-W., Lee, K.-D., Cho, D., Cho, I., Kim, W.-J., 2021. Oral antiviral clevudine compared with placebo in Korean COVID-19 patients with moderate severity (preprint). Infectious Diseases (except HIV/AIDS). [https://doi.org/10.1101/](https://doi.org/10.1101/2021.12.09.21267566) [2021.12.09.21267566](https://doi.org/10.1101/2021.12.09.21267566).
- Stegmann, K.M., Dickmanns, A., Gerber, S., Nikolova, V., Klemke, L., Manzini, V., Schlösser, D., Bierwirth, C., Freund, J., Sitte, M., others, 2021. The folate antagonist methotrexate diminishes replication of the coronavirus SARS-CoV-2 and enhances the antiviral efficacy of remdesivir in cell culture models. Virus Res. 302, 198469 [https://doi.org/10.1016/j.virusres.2021.198469.](https://doi.org/10.1016/j.virusres.2021.198469)
- Stevens, L.J., Pruijssers, A.J., Lee, H.W., Gordon, C.J., Tchesnokov, E.P., Gribble, J., George, A.S., Hughes, T.M., Lu, X., Li, J., others, 2022. Mutations in the SARS-CoV-2 RNA dependent RNA polymerase confer resistance to remdesivir by distinct mechanisms. Science translational medicine eabo0718. [https://doi.org/10.1126/](https://doi.org/10.1126/scitranslmed.abo0718) [scitranslmed.abo0718](https://doi.org/10.1126/scitranslmed.abo0718).
- Sugamoto, K., Tanaka, Y.L., Saito, A., Goto, Y., Nakayama, T., Okabayashi, T., Kunitake, H., Morishita, K., 2022. Highly polymerized proanthocyanidins (PAC) components from blueberry leaf and stem significantly inhibit SARS-CoV-2 infection via inhibition of ACE2 and viral 3CLpro enzymes. Biochem. Biophys. Res. Commun. 615, 56–62. <https://doi.org/10.1016/j.bbrc.2022.04.072>.
- Sun, Y.J., Velez, G., Parsons, D.E., Li, K., Ortiz, M.E., Sharma, S., McCray, P.B., Bassuk, A. G., Mahajan, V.B., others, 2021. Structure-based phylogeny identifies avoralstat as a TMPRSS2 inhibitor that prevents SARS-CoV-2 infection in mice. J. Clin. Invest. 131 <https://doi.org/10.1172/JCI147973>
- Suryamohan, K., Diwanji, D., Stawiski, E.W., Gupta, R., Miersch, S., Liu, J., Chen, C., Jiang, Y.-P., Fellouse, F.A., Sathirapongsasuti, J.F., others, 2021. Human ACE2 receptor polymorphisms and altered susceptibility to SARS-CoV-2. Communications biology 4, 1–11. [https://doi.org/10.1038/s42003-021-02030-3.](https://doi.org/10.1038/s42003-021-02030-3)
- Taccone, F.S., Gorham, J., Vincent, J.-L., 2020. Hydroxychloroquine in the management of critically ill patients with COVID-19: the need for an evidence base. Lancet Respir. Med. https://doi.org/10.1016/S2213-2600(20)30172-
- Talevi, A., Morales, J.F., Hather, G., Podichetty, J.T., Kim, Sarah, Bloomingdale, P.C., Kim, Samuel, Burton, J., Brown, J.D., Winterstein, A.G., Schmidt, S., White, J.K., Conrado, D.J., 2020. Machine learning in drug discovery and development Part 1: a primer. CPT Pharmacometrics Syst. Pharmacol. 9, 129-142. https://doi.org/ [10.1002/psp4.12491.](https://doi.org/10.1002/psp4.12491)
- Tao, X., Zhang, L., Du, L., Liao, R., Cai, H., Lu, K., Zhao, Z., Xie, Y., Wang, P.-H., Pan, J.- A., others, 2021. Allosteric inhibition of SARS-CoV-2 3CL protease by colloidal

bismuth subcitrate. Chem. Sci. 12, 14098–14102. [https://doi.org/10.1039/](https://doi.org/10.1039/D1SC03526F) [D1SC03526F](https://doi.org/10.1039/D1SC03526F).

Thachil, J., 2020. The versatile heparin in COVID-19. J. Thromb. Haemostasis 18, 1020–1022. [https://doi.org/10.1111/jth.14821.](https://doi.org/10.1111/jth.14821)

Than, M.E., Henrich, S., Bourenkov, G.P., Bartunik, H.D., Huber, R., Bode, W., 2005. The endoproteinase furin contains two essential Ca2+ ions stabilizing its N-terminus and the unique S1 specificity pocket. Acta Crystallogr. Sect. D Biol. Crystallogr. 61, 505–512. https://doi.org/10.1107/S090744490500255

Theodorakopoulou, M.P., Alexandrou, M.-E., Boutou, A.K., Ferro, C.J., Ortiz, A., Sarafidis, P., 2022. Renin–angiotensin system blockers during the COVID-19 pandemic: an update for patients with hypertension and chronic kidney disease. Clinical kidney journal 15, 397–406. [https://doi.org/10.1093/ckj/sfab272.](https://doi.org/10.1093/ckj/sfab272)

[Toft-Bertelsen, T.L., Jeppesen, M.G., Tzortzini, E., Xue, K., Giller, K., Becker, S.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref279) [Mujezinovic, A., Bentzen, B.H., B Andreas, L., Kolocouris, A., others, 2021.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref279) [Amantadine inhibits known and novel ion channels encoded by SARS-CoV-2 in vitro.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref279) [Commun. Biol. 4, 1](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref279)–10.

Tomar, P.P.S., Krugliak, M., Arkin, I.T., 2021. Identification of sars-cov-2 e channel blockers from a repurposed drug library. Pharmaceuticals 14, 604. [https://doi.org/](https://doi.org/10.3390/ph14070604) [10.3390/ph14070604.](https://doi.org/10.3390/ph14070604)

Tornling, G., Batta, R., Porter, J.C., Williams, B., Bengtsson, T., Parmar, K., Kashiva, R., Hallberg, A., Cohrt, A.K., Westergaard, K., others, 2021. Seven days treatment with the angiotensin II type 2 receptor agonist C21 in hospitalized COVID-19 patients; a placebo-controlled randomised multi-centre double-blind phase 2 trial. EClinicalMedicine 41, 101152.<https://doi.org/10.1016/j.eclinm.2021.101152>.

Townsend, J.A., Sanders, H.M., Rolland, A.D., Park, C.K., Horton, N.C., Prell, J.S., Wang, J., Marty, M.T., 2021. Influenza AM2 channel Oligomerization is sensitive to its chemical environment. Anal. Chem. 93, 16273–16281. [https://doi.org/10.1021/](https://doi.org/10.1021/acs.analchem.1c04660) cs.analchem.1c04660.

Tran, D.P., Taira, Y., Ogawa, T., Misu, R., Miyazawa, Y., Kitao, A., 2022. Inhibition of the hexamerization of SARS-CoV-2 endoribonuclease and modeling of RNA structures bound to the hexamer. Sci. Rep. 12, 1–15. [https://doi.org/10.1038/s41598-022-](https://doi.org/10.1038/s41598-022-07792-2) [07792-2](https://doi.org/10.1038/s41598-022-07792-2).

Tu, B., Wang, H., An, X., Qu, J., Li, Q., Gao, Y., Shi, M., Qiu, H., Huang, Y., 2022. Inhaled heparin polysaccharide nanodecoy against SARS-CoV-2 and variants. Acta Pharm. Sin. B. <https://doi.org/10.1016/j.apsb.2022.01.019>.

Tu, J., Mo, X., Zhang, X., Xun, J., Chen, X., Liu, Y., Jing, W., Xie, T., 2022. Effects of different corticosteroid therapy on severe COVID-19 patients: a meta-analysis of randomized controlled trials. Expet Rev. Respir. Med. 16, 79–89. [https://doi.org/](https://doi.org/10.1080/17476348.2021.1983429) [10.1080/17476348.2021.1983429.](https://doi.org/10.1080/17476348.2021.1983429)

Ullrich, S., Ekanayake, K.B., Otting, G., Nitsche, C., 2022. Main protease mutants of SARS-CoV-2 variants remain susceptible to nirmatrelvir. Bioorg. Med. Chem. Lett 62, 128629. [https://doi.org/10.1016/j.bmcl.2022.128629.](https://doi.org/10.1016/j.bmcl.2022.128629)

Vandyck, K., Deval, J., 2021. Considerations for the discovery and development of 3 chymotrypsin-like cysteine protease inhibitors targeting SARS-CoV-2 infection.

Current Opinion in Virology. [https://doi.org/10.1016/j.coviro.2021.04.006.](https://doi.org/10.1016/j.coviro.2021.04.006) Varona, J.F., Landete, P., Lopez-Martin, J.A., Estrada, V., Paredes, R., Guisado-Vasco, P., de Orueta, L.F., Torralba, M., Fortun, J., Vates, R., others, 2022. Preclinical and Randomized Phase I Studies of Plitidepsin in Adults Hospitalized with COVID-19. Life Science Alliance, vol. 5. [https://doi.org/10.26508/lsa.202101200.](https://doi.org/10.26508/lsa.202101200)

Vela, J.M., 2020. Repurposing sigma-1 receptor ligands for COVID-19 therapy? Front. Pharmacol. 11, 582310 [https://doi.org/10.3389/fphar.2020.582310.](https://doi.org/10.3389/fphar.2020.582310)

Verma, J.S., Libertin, C.R., Gupta, Y., Khanna, G., Kumar, R., Arora, B.S., Krishna, L., Fasina, F.O., Hittner, J.B., Antoniades, A., others, 2022. Multi-cellular immunological interactions associated with COVID-19 infections. Front. Immunol. 13, 794006<https://doi.org/10.3389/fimmu.2022.794006>.

von Roemeling, C.A., Caulfield, T.R., Marlow, L., Bok, I., Wen, J., Miller, J.L., Hughes, R., Hazlehurst, L., Pinkerton, A.B., Radisky, D.C., Tun, H.W., Kim, Y.S.B., Lane, A.L., Copland, J.A., 2018. Accelerated bottom-up drug design platform enables the discovery of novel stearoyl-CoA desaturase 1 inhibitors for cancer therapy. Oncotarget 9, 3–20. [https://doi.org/10.18632/oncotarget.21545.](https://doi.org/10.18632/oncotarget.21545)

Vuong, W., Khan, M.B., Fischer, C., Arutyunova, E., Lamer, T., Shields, J., Saffran, H.A., McKay, R.T., van Belkum, M.J., Joyce, M.A., Young, H.S., Tyrrell, D.L., Vederas, J.C., Lemieux, M.J., 2020. Feline coronavirus drug inhibits the main protease of SARS-CoV-2 and blocks virus replication. Nat. Commun. 11, 4282. https://doi.or [10.1038/s41467-020-18096-2.](https://doi.org/10.1038/s41467-020-18096-2)

Wahl, A., Gralinski, L.E., Johnson, C.E., Yao, W., Kovarova, M., Dinnon, K.H., Liu, H., Madden, V.J., Krzystek, H.M., De, C., others, 2021. SARS-CoV-2 infection is effectively treated and prevented by EIDD-2801. Nature 591, 451–457. [https://doi.](https://doi.org/10.1038/s41586-021-03312-w) [org/10.1038/s41586-021-03312-w.](https://doi.org/10.1038/s41586-021-03312-w)

[Wang, C., Li, H., Xiao, S., Li, Z., Zhao, X., Xie, J., Ye, C., Xia, L., Lou, X., Zhou, X., 2022.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref294) [Abnormal dynamic ventilation function of COVID-19 survivors detected by](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref294) [pulmonary free-breathing proton MRI. Eur. Radiol. 1](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref294)–11.

Wang, Y., Fang, S., Wu, Y., Cheng, X., Zhang, L., Shen, X., Li, S., Xu, J., Shang, W., Gao, Z., others, 2022. Discovery of SARS-CoV-2-E channel inhibitors as antiviral candidates. Acta Pharmacol. Sin. 43, 781–787. [https://doi.org/10.1038/s41401-](https://doi.org/10.1038/s41401-021-00732-2) 021-00732-

Wang, Z., Joshi, A., Leopold, K., Jackson, S., Christensen, S., Nayfeh, T., Mohammed, K., Creo, A., Tebben, P., Kumar, S., 2022. Association of vitamin D deficiency with COVID-19 infection severity: systematic review and meta-analysis. Clin. Endocrinol. 96, 281–287. [https://doi.org/10.1111/cen.14540.](https://doi.org/10.1111/cen.14540)

Watson, C., 2022. Rise of the preprint: how rapid data sharing during COVID-19 has changed science forever. Nat. Med. 28, 2–5. [https://doi.org/10.1038/s41591-021-](https://doi.org/10.1038/s41591-021-01654-6) [01654-6](https://doi.org/10.1038/s41591-021-01654-6).

Whitley, R., 2022. Molnupiravir—[a step toward orally bioavailable therapies for Covid-](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref298)[19. N. Engl. J. Med.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref298)

WHO coronavirus disease (COVID-19). Dashboard [WWW Document], n.d. URL. [http](https://covid19.who.int) [s://covid19.who.int](https://covid19.who.int). accessed 1.20.21.

[Wimmer, S., Keestra, S.M., 2022. Public risk-taking and rewards during the COVID-19](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref300) [pandemic-a case study of remdesivir in the context of global health equity. Int. J.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref300) [Health Pol. Manag. 11, 567](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref300)–578.

[Wong, C.K., Au, I.C., Lau, K.T., Lau, E., Cowling, B.J., Leung, G.M., 2022. Real-world](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref301) [Effectiveness of Molnupiravir and Nirmatrelvir/ritonavir Among COVID-19](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref301) Inpatients during Hong Kong'[s Omicron BA. 2 Wave: an Observational Study.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref301) [medRxiv](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref301).

Xian, Y., Zhang, J., Bian, Z., Zhou, H., Zhang, Z., Lin, Z., Xu, H., 2020. Bioactive natural compounds against human coronaviruses: a review and perspective. Acta Pharm. Sin. B 10, 1163–1174. <https://doi.org/10.1016/j.apsb.2020.06.002>.

Xiang, R., Yu, Z., Wang, Y., Wang, L., Huo, S., Li, Y., Liang, R., Hao, Q., Ying, T., Gao, Y., Yu, F., Jiang, S., 2022. Recent advances in developing small-molecule inhibitors against SARS-CoV-2. Acta Pharm. Sin. B 12, 1591–1623. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.apsb.2021.06.016) [apsb.2021.06.016](https://doi.org/10.1016/j.apsb.2021.06.016).

Xiong, R., Zhang, L., Li, S., Sun, Y., Ding, M., Wang, Y., Zhao, Y., Wu, Y., Shang, W., Jiang, X., others, 2020. Novel and potent inhibitors targeting DHODH are broadspectrum antivirals against RNA viruses including newly-emerged coronavirus SARS-CoV-2. Protein & cell 11, 723–739. [https://doi.org/10.1007/s13238-020-](https://doi.org/10.1007/s13238-020-00768-w)

[00768-w.](https://doi.org/10.1007/s13238-020-00768-w) Yalcinkaya, M., Liu, W., Islam, M.N., Kotini, A.G., Gusarova, G.A., Fidler, T.P., Papapetrou, E.P., Bhattacharya, J., Wang, N., Tall, A.R., 2021. Modulation of the NLRP3 inflammasome by sars-CoV-2 envelope protein. Sci. Rep. 11, 1–12. [https://](https://doi.org/10.1038/s41598-021-04133-7) doi.org/10.1038/s41598-021-04133-7.

Yamaguchi, T., Hoshizaki, M., Minato, T., Nirasawa, S., Asaka, M.N., Niiyama, M., Imai, M., Uda, A., Chan, J.F.-W., Takahashi, S., 2021. ACE2-like carboxypeptidase B38-CAP protects from SARS-CoV-2-induced lung injury. Nat. Commun. 12, 1–13. <https://doi.org/10.1038/s41467-021-27097-8>.

Yamamoto, M., Kiso, M., Sakai-Tagawa, Y., Iwatsuki-Horimoto, K., Imai, M., Takeda, M., Kinoshita, N., Ohmagari, N., Gohda, J., Semba, K., Matsuda, Z., Kawaguchi, Y., Kawaoka, Y., Inoue, J., 2020. The anticoagulant nafamostat potently inhibits SARS-CoV-2 S protein-mediated fusion in a cell fusion assay system and viral infection in vitro in a cell-type-dependent manner. Viruses 12, 629. [https://doi.org/10.3390/](https://doi.org/10.3390/v12060629) [v12060629.](https://doi.org/10.3390/v12060629)

Yan, W., Zheng, Y., Zeng, X., He, B., Cheng, W., 2022. Structural biology of SARS-CoV-2: open the door for novel therapies. Signal Transduct. Targeted Ther. 7, 1–28. [https://](https://doi.org/10.1038/s41392-022-00884-5) doi.org/10.1038/s41392-022-00884-5.

Yang, C., Pan, X., Huang, Y., Cheng, C., Xu, X., Wu, Y., Xu, Y., Shang, W., Niu, X., Wan, Y., others, 2021. Drug repurposing of itraconazole and Estradiol benzoate against COVID-19 by blocking SARS-CoV-2 spike protein-mediated membrane fusion. Advanced therapeutics 4, 2000224. [https://doi.org/10.1002/](https://doi.org/10.1002/adtp.202000224) [adtp.202000224](https://doi.org/10.1002/adtp.202000224).

Yang, C.-W., Peng, T.-T., Hsu, H.-Y., Lee, Y.-Z., Wu, S.-H., Lin, W.-H., Ke, Y.-Y., Hsu, T.- A., Yeh, T.-K., Huang, W.-Z., Lin, J.-H., Sytwu, H.-K., Chen, C.-T., Lee, S.-J., 2020. Repurposing old drugs as antiviral agents for coronaviruses. Biomed. J. 43, 368–374. <https://doi.org/10.1016/j.bj.2020.05.003>.

Ye, Q., West, A.M., Silletti, S., Corbett, K.D., 2020. Architecture and self-assembly of the SARS-CoV-2 nucleocapsid protein. Protein Sci. 29, 1890–1901. [https://doi.org/](https://doi.org/10.1002/pro.3909) [10.1002/pro.3909.](https://doi.org/10.1002/pro.3909)

Yim, S.-K., Kim, K., Kim, I., Chun, S., Oh, T., Kim, J.-U., Kim, J., Jung, W., Moon, H., Ku, B., Jung, K., 2021. Inhibition of SARS-CoV-2 virus entry by the crude polysaccharides of seaweeds and abalone viscera in vitro. Mar. Drugs 19, 219. <https://doi.org/10.3390/md19040219>.

Yu, F., Pan, T., Huang, F., Ying, R., Liu, J., Fan, H., Zhang, J., Liu, W., Lin, Y., Yuan, Y., others, 2022. Glycopeptide antibiotic teicoplanin inhibits cell entry of SARS-CoV-2 by suppressing the proteolytic activity of cathepsin L. Front. Microbiol. 13 [https://](https://doi.org/10.3389/fmicb.2022.884034) [doi.org/10.3389/fmicb.2022.884034.](https://doi.org/10.3389/fmicb.2022.884034)

[Yu, J., Granberg, T., Shams, R., Petersson, S., Skold, M., Nyren, S., Lundberg, J., 2022.](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref314) [Lung Perfusion Disturbances Detected with MRI in Non-hospitalized Post-COVID-19](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref314) [Individuals with Dyspnea 3-13 Months after the Acute Disease. medRxiv](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref314).

Yuan, S., Wang, R., Chan, J.F.-W., Zhang, A.J., Cheng, T., Chik, K.K.-H., Ye, Z.-W., Wang, S., Lee, A.C.-Y., Jin, L., others, 2020. Metallodrug ranitidine bismuth citrate suppresses SARS-CoV-2 replication and relieves virus-associated pneumonia in Syrian hamsters. Nat. Microbiol. 5, 1439–1448. [https://doi.org/10.1038/s41564-](https://doi.org/10.1038/s41564-020-00802-x) $020 - 00802$

Zamai, L., 2021. Upregulation of the renin–angiotensin system pathways and SARS-CoV-2 infection: the rationale for the administration of zinc-chelating agents in COVID-19 patients. Cells 10, 506. <https://doi.org/10.3390/cells10030506>.

Zanella, I., Zizioli, D., Castelli, F., Quiros-Roldan, E., 2021. Tenofovir, another inexpensive, well-known and widely available old drug repurposed for SARS-COV-2 infection. Pharmaceuticals 14, 454. <https://doi.org/10.3390/ph14050454>

Zapata-Cardona, M.I., Flórez-Álvarez, L., Zapata-Builes, W., Guerra-Sandoval, A.L., Guerra-Almonacid, C.M., Hincapié-García, J., Rugeles, M.T., Hernandez, J.C., 2021. Atorvastatin effectively inhibits late replicative cycle steps of SARS-CoV-2 in vitro (preprint). Microbiology. https://doi.org/10.1101/2021.03.01.43349

Zein, A.F.M.Z., Sulistiyana, C.S., Raffaello, W.M., Wibowo, A., Pranata, R., 2022. Sofosbuvir with daclatasvir and the outcomes of patients with COVID-19: a systematic review and meta-analysis with GRADE assessment. Postgrad. Med. 98, 509–514. <https://doi.org/10.1136/postgradmedj-2021-140287>.

Zein, J.G., Strauss, R., Attaway, A.H., Hu, B., Milinovich, A., Jawhari, N., Chamat, S.S., Ortega, V.E., 2022. Eosinophilia is associated with improved COVID-19 outcomes in inhaled corticosteroid-treated patients. J. Allergy Clin. Immunol. Pract. 10, 742–750. <https://doi.org/10.1016/j.jaip.2021.12.034> e14.

Zendehdel, A., Bidkhori, M., Ansari, M., Jamalimoghaddamsiyahkali, S., Asoodeh, A., 2022. Efficacy of oseltamivir in the treatment of patients infected with Covid-19.

Annals of Medicine and Surgery 77, 103679. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.amsu.2022.103679) [amsu.2022.103679.](https://doi.org/10.1016/j.amsu.2022.103679)

- Zeng, X., Song, X., Ma, T., Pan, X., Zhou, Y., Hou, Y., Zhang, Z., Li, K., Karypis, G., Cheng, F., 2020. Repurpose open data to discover therapeutics for COVID-19 using deep learning. J. Proteome Res. 19, 4624–4636. [https://doi.org/10.1021/acs.](https://doi.org/10.1021/acs.jproteome.0c00316) bme.0c00316.
- Zhang, H., Yang, Y., Li, J., Wang, M., Saravanan, K.M., Wei, J., et al., 2020. A novel virtual screening procedure identifies Pralatrexate as inhibitor of SARS-CoV-2 RdRp and it reduces viral replication in vitro. PLoS Comput. Biol. 16 (12), e1008489 <https://doi.org/10.1371/journal.pcbi.1008489>.
- Zhang, H., Zhang, T., Saravanan, K.M., Liao, L., Wu, H., Zhang, H., et al., 2022. DeepBindBC: a practical deep learning method for identifying native-like proteinligand complexes in virtual screening. Methods 205, 247–262. [https://doi.org/](https://doi.org/10.1016/j.ymeth.2022.07.009) [10.1016/j.ymeth.2022.07.009](https://doi.org/10.1016/j.ymeth.2022.07.009).
- [Zhang, J., Ejikemeuwa, A., Gerzanich, V., Nasr, M., Tang, Q., Simard, J.M., Zhao, R.Y.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref325) [2022. Understanding the role of SARS-CoV-2 ORF3a in viral pathogenesis and](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref325) [COVID-19. Front. Microbiol. 13, 854567](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref325).
- Zhang, L., Lin, D., Sun, X., Curth, U., Drosten, C., Sauerhering, L., Becker, S., Rox, K., Hilgenfeld, R., 2020. Crystal structure of SARS-CoV-2 main protease provides a basis for design of improved α-ketoamide inhibitors. Science 368, 409–412. [https://doi.](https://doi.org/10.1126/science.abb3405) [org/10.1126/science.abb3405](https://doi.org/10.1126/science.abb3405).
- [Zhang, Q., Radvak, P., Lee, J., Xu, Y., Cao-Dao, V., Xu, M., Zheng, W., Chen, C.Z., Xie, H.,](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref327) [Ye, Y., 2022. Mitoxantrone modulates a heparan sulfate-spike complex to inhibit](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref327) [SARS-CoV-2 infection. Sci. Rep. 12, 1](http://refhub.elsevier.com/S0098-2997(22)00096-6/sref327)–12.
- Zhang, W., Zhang, Y., Min, Z., Mo, J., Ju, Z., Guan, W., Zeng, B., Liu, Y., Chen, J., Zhang, Q., others, 2022. COVID19db: a comprehensive database platform to
- discover potential drugs and targets of COVID-19 at whole transcriptomic scale. Nucleic Acids Res. 50, D747–D757. <https://doi.org/10.1093/nar/gkab850>.
- Zhao, Y., Du, X., Duan, Y., Pan, X., Sun, Y., You, T., Han, L., Jin, Z., Shang, W., Yu, J., others, 2021. High-throughput screening identifies established drugs as SARS-CoV-2 PLpro inhibitors. Protein & cell 12, 877–888. [https://doi.org/10.1007/s13238-021-](https://doi.org/10.1007/s13238-021-00836-9) [00836-9](https://doi.org/10.1007/s13238-021-00836-9).
- Zheng, M., Karki, R., Williams, E.P., Yang, D., Fitzpatrick, E., Vogel, P., Jonsson, C.B., Kanneganti, T.-D., 2021. TLR2 senses the SARS-CoV-2 envelope protein to produce inflammatory cytokines. Nat. Immunol. 22, 829–838. [https://doi.org/10.1038/](https://doi.org/10.1038/s41590-021-00937-x) [s41590-021-00937-x](https://doi.org/10.1038/s41590-021-00937-x).
- Zheng, Y.-X., Wang, L., Kong, W.-S., Chen, H., Wang, X.-N., Meng, Q., Zhang, H.-N., Guo, S.-J., Jiang, H.-W., Tao, S.-C., 2021. Nsp2 has the potential to be a drug target revealed by global identification of SARS-CoV-2 Nsp2-interacting proteins. Acta Biochim. Biophys. Sin. 53, 1134–1141. [https://doi.org/10.1093/abbs/gmab088.](https://doi.org/10.1093/abbs/gmab088)
- Zhou, Y., Vedantham, P., Lu, K., Agudelo, J., Carrion Jr., R., Nunneley, J.W., Barnard, D., Pöhlmann, S., McKerrow, J.H., Renslo, A.R., others, 2015. Protease inhibitors targeting coronavirus and filovirus entry. Antivir. Res. 116, 76–84. [https://doi.org/](https://doi.org/10.1016/j.antiviral.2015.01.011) [10.1016/j.antiviral.2015.01.011](https://doi.org/10.1016/j.antiviral.2015.01.011).
- Zhuravel, S.V., Khmelnitskiy, O.K., Burlaka, O.O., Gritsan, A.I., Goloshchekin, B.M., Kim, S., Hong, K.Y., 2021. Nafamostat in hospitalized patients with moderate to severe COVID-19 pneumonia: a randomised Phase II clinical trial. EClinicalMedicine 41, 101169. [https://doi.org/10.1016/j.eclinm.2021.101169.](https://doi.org/10.1016/j.eclinm.2021.101169)
- Zimniak, M., Kirschner, L., Hilpert, H., Geiger, N., Danov, O., Oberwinkler, H., Steinke, M., Sewald, K., Seibel, J., Bodem, J., 2021. The serotonin reuptake inhibitor Fluoxetine inhibits SARS-CoV-2 in human lung tissue. Sci. Rep. 11, 5890. [https://](https://doi.org/10.1038/s41598-021-85049-0) doi.org/10.1038/s41598-021-85049-0.