ORIGINAL ARTICLE



Glucose metabolism patterns: A potential index to characterize brain ageing and predict high conversion risk into cognitive impairment

Jiehui Jiang \bigcirc · Can Sheng · Guanqun Chen · Chunhua Liu · Shichen Jin · Lanlan Li · Xueyan Jiang · Ying Han · for the Alzheimer's Disease Neuroimaging Initiative

Received: 27 January 2022 / Accepted: 7 May 2022 / Published online: 18 May 2022 © The Author(s), under exclusive licence to American Aging Association 2022

Abstract Exploring individual hallmarks of brain ageing is important. Here, we propose the agerelated glucose metabolism pattern (ARGMP) as a potential index to characterize brain ageing in cognitively normal (CN) elderly people. We collected ¹⁸F-fluorodeoxyglucose (¹⁸F-FDG) PET brain images from two independent cohorts: the Alzheimer's Disease Neuroimaging Initiative (ADNI, N=127) and the Xuanwu Hospital of Capital Medical University, Beijing, China (N=84). During follow-up (mean 80.60 months), 23 participants in the ADNI cohort converted to cognitive impairment. ARGMPs were

Jiehui Jiang, Can Sheng, and Guanqun Chen contributed equally to this work.

Supplementary Information The online version contains supplementary material available at https://doi. org/10.1007/s11357-022-00588-2.

J. Jiang (⊠) · C. Liu · S. Jin · L. Li Institute of Biomedical Engineering, School of Information and Communication Engineering, Shanghai University, Shanghai 200444, China e-mail: jiangjiehui@shu.edu.cn

C. Sheng · G. Chen · Y. Han Department of Neurology, Xuanwu Hospital of Capital Medical University, Beijing 100053, China

X. Jiang · Y. Han (⊠) School of Biomedical Engineering, Hainan University, Haikou 570228, China e-mail: hanying@xwh.ccmu.edu.cn identified using the scaled subprofile model/principal component analysis method, and cross-validations were conducted in both independent cohorts. A survival analysis was further conducted to calculate the predictive effect of conversion risk by using ARGMPs. The results showed that ARGMPs were characterized by hypometabolism with increasing age primarily in the bilateral medial superior frontal gyrus, anterior cingulate and paracingulate gyri, caudate nucleus, and left supplementary motor area and hypermetabolism in part of the left inferior cerebellum. The expression network scores of ARGMPs were significantly associated with chronological age (R = 0.808, p < 0.001), which was validated in both the ADNI and Xuanwu cohorts. Individuals with higher network scores exhibited a better predictive effect (HR: 0.30, 95% CI: 0.1340~0.6904, p = 0.0068). These findings indicate that ARGMPs

X. Jiang

German Centre for Neurodegenerative Disease, Clinical Research Group, Venusberg Campus 1, 53121 Bonn, Germany

Y. Han

Centre of Alzheimer's Disease, Beijing Institute for Brain Disorders, Beijing 100053, China

Y. Han National Clinical Research Centre for Geriatric Disorders, Beijing 100053, China derived from CN participants may represent a novel index for characterizing brain ageing and predicting high conversion risk into cognitive impairment.

Keywords Pattern · Brain ageing · Positron emission tomography · Glucose metabolism

Introduction

Ageing is a complex process occurring in both normal ageing and pathological conditions. Exploring hallmarks of brain ageing in cognitively normal (CN) elderly people is important for understanding the normal ageing process [1, 2]. Currently, existing indices used to characterize individual brain ageing focus on the cellular and molecular levels, such as mitochondrial dysfunction, accumulation of oxidatively damaged molecules, aberrant neuronal network activity, and dysregulated energy metabolism [3–5]. However, the complexity of measurement methods limits the wide application of these indices in clinical practice. In addition, the myriad of indices introduced by previous studies cannot be universally used in all conditions due to different mechanisms and their variable patterns. For instance, these indices may be influenced by both healthy brain ageing and pathological disruptions, e.g. amyloid- β (A β) and tau pathologies in Alzheimer's disease (AD) [3]. Therefore, an alternative in vivo index is needed.

Glucose metabolism alterations in the brain may represent an alternative index of brain ageing. Ageing appears to deteriorate the systemic control of glucose utilization, which, in turn, may increase the risk of reducing glucose uptake in brain regions [6]. Studies have reported that brain glucose metabolism abnormalities occur with ageing and reflect cognitive decline with high sensitivity [7, 8]. Thus, mapping age-related regional glucose metabolism changes may represent a promising avenue for understanding the neurobiological foundation of normal brain ageing. ¹⁸F-fluorodeoxyglucose (¹⁸F-FDG) positron emission tomography (PET) is a well-established technique to visualize and quantify the resting-state cerebral glucose metabolic rate in vivo and has been used to elucidate the normal ageing process of human brains [9–11]. Using ¹⁸F-FDG PET, previous studies have shown that normal brain ageing is associated with specific patterns of regional cerebral glucose metabolism, such as the anterior cingulate/medial prefrontal cortex [1, 7, 12]. Therefore, it is possible to estimate brain ageing by generating a glucose metabolism pattern using ¹⁸F-FDG PET.

Currently, data-driven methods are usually applied to estimate brain ageing using chronological age as the training label [13–15]. In particular, the scaled subprofile model/principal component analysis (SSM/PCA) method is frequently used to characterize brain glucose metabolism. SSM/PCA is a multivariate statistical method that drives a covariance pattern of voxel-based differences in brain metabolism [16]. This feature extraction method can enhance the identification of significant patterns in multivariate imaging data and mirror the underlying relationships between brain regions that are not captured by univariate techniques [17, 18]. Additionally, it enables the assessment of network-level alterations [19]. Therefore, this method is an optimal tool to evaluate abnormalities in functional brain organization in neurodegenerative disorders. Currently, the SSM/PCA method has been widely used to investigate brain metabolic patterns in several disorders, such as AD, Parkinson's disease (PD), and multiple system atrophy (MSA) [20-23]. Thus, we hypothesized that the SSM/PCA method could also be used to assess agerelated glucose metabolism patterns (ARGMPs) in CN older adults.

In the present study, we extracted ARGMPs from ¹⁸F-FDG PET images and generated the ARGMP expression score in each participant as a potential index to characterize brain ageing. The primary purposes of this study were (1) to validate the robustness of ARGMPs in both Western and Chinese cohorts; (2) to assess whether ARGMPs in normal ageing are independent from the AD pathological process; and (3) to investigate whether individuals with high ARGMPs have a high risk of developing cognitive impairment during follow-up.

Materials and methods

Overall study procedures

A total of 211 CN elderly people were collected from two independent cohorts: the Alzheimer's Disease Neuroimaging Initiative (ADNI) cohort (Cohort A, N=127) from the ADNI database (http://adni.loni. usc.edu/) and the Xuanwu cohort (Cohort B, N = 84) from the Department of Neurology, Xuanwu Hospital of Capital Medical University, Beijing, China. All participants underwent ¹⁸F-FDG PET scans. Using the SSM/PCA algorithm, ARGMPs were identified based on the combined cohort. Then, network expression scores were used to quantify ARGMPs, and the association of individual network scores with chronological age in the combined cohort was assessed. To validate the robustness of the ARGMPs, metabolic brain network analysis was conducted in the ADNI and Xuanwu cohorts. We separately assessed the correlation between individual network scores and chronological age in both independent cohorts. In addition, according to brain $A\beta$ and the standardized network score, the entire ADNI cohort was divided into four stages (stage 0, A β -Network score-; stage 1, A β -Network score+; stage 2, A β +Network score-; and stage 3, $A\beta$ + Network score +), and the conversion rate of CN participants to cognitive impairment in each stage was calculated. Survival analysis was further conducted to compare the predictive effect of conversion risk among different stages. The procedures of this study are shown in Fig. 1.

Participants

For all CN participants of the two cohorts, clinical (sex, age, education, and Mini-Mental State Examination (MMSE)) and neuroimaging (18F-FDG PET and T1-MRI image) data were collected. Each participant in the ADNI cohort underwent ¹⁸F-florbetapir (AV45) PET scans at baseline. Participants in the ADNI cohort completed the Montreal Cognitive Assessment (MoCA), and those in the Xuanwu cohort completed the MoCA-Basic Version (MoCA-B). In addition, the clinical severity was also evaluated using the Clinical Dementia Rating Sum of Boxes (CDR-SB) for participants in the ADNI cohort. The combination of the two cohorts was defined as the combined cohort. The inclusion criteria for CN participants were as follows: (1) no cognitive complaints; (2) no history of stroke, hypertension, central nervous system diseases, or mental illness; and (3) an MMSE score above or equal to 27.

Specifically, according to whether the cerebralto-cerebellar AV45 standardized uptake value ratio (SUVR) was below 1.18 [24], CN participants in the ADNI cohort were classified into A β -negative CN



Fig. 1 Whole procedure of the present study. (A) Clinical and neuroimaging date collected from two cohorts: the ADNI cohort and the Xuanwu cohort. (B) The image preprocessing procedure. (C) Extracting ARGMPs using the SSM/PCA

method. (**D**) Cross-validations of ARGMPs in both the ADNI cohort and the Xuanwu cohort. (**E**) Assessing the predictive effect of ARGMPs

(CN-, N=96) and A β -positive CN (CN+, N=31) groups. Two experienced neuroradiologists at Xuanwu Hospital, who were blinded to the classification, also visually evaluated the amyloid-PET images. The detailed selection process for participants in the ADNI cohort is shown in Supplementary Fig. S1. However, due to the lack of ¹⁸F-florbetapir (AV45) PET scans, the CN participants from the Xuanwu cohort were not divided into CN- and CN+subgroups.

Participants with CDR-SB scores ≥ 1 at the followup visit were considered as the transformers. Two experienced neurologists (CS and YH) evaluated the clinical diagnosis of conversion to cognitive impairment. In our study, during the follow-up (follow-up range: $6 \sim 168$ months, mean follow-up period: 80.60 months), 23 participants in the ADNI cohort developed cognitive impairment (conversion rate: 18.11%).

This study was approved by the Institutional Review Board of the ADNI and the Research Ethics Committee of Xuanwu Hospital, Beijing, China (ClinicalTrials.gov Identifier: NCT03370744). All participants voluntarily participated in this study and provided written informed consent.

Image acquisition protocol

In the ADNI cohort, all participants were invited to undergo optional ¹⁸F-FDG PET and ¹⁸F-AV45 PET scans in three-dimensional acquisition mode. The process of data acquisition in the ADNI cohort is described in detail in the imaging protocol column of the ADNI database (http://adni.loni.usc.edu/). In the Xuanwu cohort, PET and T1-MRI data were simultaneously obtained on an integrated 3.0 T time-of-flight (TOF) PET/MR scanner (SIGNA PET/MR, GE Healthcare, Milwaukee, Wisconsin, USA) for each participant. For FDG-PET, each participant was instructed to fast for at least 6 h and was required to have a confirmed serum glucose level below 8 mmol/L. The images were acquired approximately 40 min after an intravenous injection of 3.7 MBq/kg of ¹⁸F-FDG, and the data were recorded by using a TOF ordered subset expectation maximization algorithm with the following parameters: scan duration = 35 min, eight iterations, 32 subsets matrix = 192×192 , field of view (FOV) = 350×350 , and half-width height = 3. Three-dimensional T1-weighted magnetization-prepared rapid gradient echo scans were performed with the following parameters: spoiled gradient-recalled echo (SPGR) sequence, FOV= $256 \times 256 \text{ mm}^2$, matrix= 256×256 , slice thickness=1 mm, gap=0, slice number=192, repetition time (TR)=6.9 ms, echo time (TE)=2.98 ms, inversion time (TI)=450 ms, flip angle= 12° , and voxel size= $1 \times 1 \times 1$ mm³.

Image preprocessing

All PET images and T1-MRI images were preprocessed using statistical parametric mapping software https://www.fil.ion.ucl.ac.uk/spm/software/ (SPM12; spm12/) in MATLAB (Version R2014a; MathWorks, Natick, MA, USA). We first used the realigning method to ensure that all frames in the dynamic PET scans were motion-corrected to the first frame, processed the output single average functional image, and reduced system or head motion errors. Then, the average functional image was normalized to the standard Montreal Neurological Institute (MNI) brain space using the deformation field from the MRI image to the MNI space. Finally, normalized functional images were smoothed to reduce noise and improve image quality using an isotropic Gaussian smoothing kernel with a Gaussian filter of 8 mm full-width at half-maximum.

SSM/PCA

To explore the effect of ageing on glucose metabolism in the brains of CN participants, metabolic brain network analysis was conducted on their FDG PET images to identify ARGMPs using the SSM/ PCA voxel-based spatial covariance mapping algorithm. The SSM/PCA method was implemented using ScAnVp (Scan Analysis and Visualization Processor) software, version 7.0w (available at http://www.feinsteinneuroscience.org at the Centre for Neuroscience, the Feinstein Institute for Medical Research, Manhasset, NY). FDG PET images were first multiplied using a binary mask (determined by the standardized automated anatomical labelling (AAL) template) to eliminate artefacts associated with noise and areas unrelated to brain activity, including the ventricle and white matter, from subsequent analysis. Then, the SSM/PCA method was used to derive the group invariant subprofile (GIS) and the participant score of each principal component (PC) in the spatial covariance model of the combined CN group. Specifically, the logarithmically transformed training set image data matrix is double centred to obtain the subject residual profile (SPR), which reveals a covariance structure that has biological significance for the group. The SPR is expressed as follows:

$$SRP = logD - logD - GMP$$

where logD is the row mean of the log values and GMP (group mean profile) is the mean row image. Next, PCA is applied to the SRP covariance matrix to derive orthogonal eigenvectors, associated eigenvalues, and participant scores to obtain subject scaling factors (SSFs) and GIS using Eq. 2.

$$SRP = \sum_{n} SSF * GIS$$

We selected the series of PCs with the maximal correlation to chronological age in the multiple linear regression analysis, with the participant scores of the first series of PCs (cumulative variance accounted for 60%) and age as the independent and dependent variables, respectively. ARGMPs were identified based on the linear combination of the regression coefficients and GISs of these selected PCs. The stability of the ARGMPs is based on a bootstrapping scheme. The voxels were obtained by bootstrapping 1000 times on ARGMPs at p < 0.001 with an absolute weight threshold of 1.4 and a cluster threshold of 100 voxels (800 mm³). They were then mapped to the AAL atlas to determine anatomical brain regions that exhibited age-related changes in glucose metabolism.

Using the voxel-based topographic profile rating algorithm, network expression scores were used to quantify ARGMPs in all participants. The correlation between network scores and chronological age of CN participants in the combined cohort was used to assess the accuracy and reliability of the ARGMPs. To validate the robustness of the ARGMPs, metabolic brain network analysis was also conducted in the ADNI and Xuanwu cohorts. In addition, to validate the independence of ARGMPs in the normal ageing process and AD pathological process, the correlation between network scores and chronological age was assessed separately in the CN- and CN + subgroups of the ADNI cohort.

Staging the transformers in the ADNI cohort

All CN participants in the ADNI cohort underwent amyloid-PET scans. The $A\beta$ positivity threshold was

calculated as an SUVR of 1.18. Then, we calculated the standardized chronological age and standardized network score, with a score of 0 as the threshold. According to whether the A β was greater than 1.18 and the standardized chronological age was greater than 0, the entire ADNI cohort was divided as follows: stage 0, A β -Age-; stage 1, A β -Age+; stage 2, $A\beta$ +Age-; and stage 3, $A\beta$ +Age+. According to whether the standardized network score was greater than 0, participants in the ADNI cohort were also classified into four stages: stage 0, Aβ-Network score-; stage 1, A β -Network score+; stage 2, $A\beta$ + Network score-; and stage 3, $A\beta$ + Network score +. In each stage, the conversion rate was calculated. The standardized values (z scores) of individual chronological age and network score were calculated using the following formula:

$$Zscore = \frac{Age - M_{age}}{SD_{age}}$$

$$Zscore = \frac{NS - M_{NS}}{SD_{NS}}$$

where M_{age} denotes mean chronological age, SD_{age} denotes the standard deviation of the chronological age, M_{NS} denotes the mean network score, and SD_{NS} denotes the standard deviation of the network score.

Statistical analysis

A two-sample t-test was performed to compare differences in continuous variables, and a Chi-square test was conducted to assess categorical variables. A multiple linear regression model was used, controlling for sex and years of education as covariates. Pearson's correlation coefficient was then used to describe the correlation between the network scores and chronological age of the CN individuals in each cohort. To validate the independence of ARGMPs from ADrelated pathology, we further evaluated the correlation between the network scores and chronological age in the CN- and CN+subgroups. The Gramm toolbox (available at https://github.com/piermorel/ gramm) in MATLAB was used to plot and visualize all of the statistical data presented in this paper.

Survival was estimated using the Kaplan-Meier method, and any differences in survival were evaluated

using a log-rank test in R version 3.6.3. To assess the validity of ARGMPs in predicting high conversion risk to cognitive impairment, we first compared the survival probability between the standardized network score > 0(Network score+) group and the standardized network score < 0 (Network score-) group. Next, the survival probability between the standardized chronological age>0 (Age+) group and standardized chronological age < 0 (Age-) group was also compared. Moreover, four subgroups, the Aβ-Network score-, Aβ-Network score+, $A\beta$ +Network score-, and $A\beta$ +Network score+, were compared to evaluate the risk of conversion. Hazard ratios were used to indicate the risk of conversion among different groups. The p-value was calculated using the log-rank test. P < 0.05 was considered statistically significant.

Results

Demographic characteristics of participants at baseline

There were a total of 211 CN participants (age range 53.0~84.8 years) in the present study, including 127 CN participants (age range 59.8~84.8 years) from the ADNI cohort and 84 CN participants (age range

53.0~76.0 years) from the Xuanwu cohort. Table 1 shows the detailed demographic and clinical details of all participants at baseline. Significant differences were observed in age (p < 0.001), sex (p < 0.001), and educational years (p < 0.001) between the ADNI and Xuanwu CN cohorts, while no significant differences in the MMSE (p=0.933) or MoCA (p=0.061) were observed. In addition, the CN+subgroup was older than the CN- subgroup (p < 0.001). The follow-up period of the ADNI cohort was 80.60 ± 36.2 months, with a total of 23 participants (18.11%) converting to cognitive impairment.

Supplementary Table S1 shows the demographic and clinical information for transformers (n=23)and nontransformers (n=104) in the ADNI cohort at baseline. There were significant differences in age (p=0.008) and MMSE scores (p=0.040) between transformers and nontransformers, while no differences in sex, education, or MoCA scores were observed.

SSM/PCA based on the combined cohort

Network analysis for CN participants in the combined cohort evaluated the first eight PCs, including 51.5% participant×voxel variance. ARGMPs were identified by linear combination with the first six PCs

 Table 1
 Demographic and clinical characteristics of CN participants at baseline

	The combined cohort	ADNI cohort	ADNI cohort			
		Total	CN-	CN+		
N	211	127	96	31	84	
Age (y)	70.5 ± 6.6	73.9 ± 5.4	72.88 ± 5.5	$76.88 \pm 4.1^{\#}$	$65.3 \pm 4.7 **$	
Sex (F/M)	134/77	68/59	47/49	21/10	66/18**	
Education (y)	15.1 ± 3.2	16.5 ± 2.6	16.60 ± 2.6	16.00 ± 2.5	$13.1 \pm 2.9 **$	
MMSE	29.3 ± 0.8	29.3 ± 0.8	29.32 ± 0.8	29.16 ± 1.0	29.3 ± 0.8	
CDR-SB	/	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0	/	
MoCA/MoCA-B ^a	26.2 ± 2.1	26.0 ± 2.3	26.12 ± 2.3	25.67 ± 2.1	26.6 ± 1.9	
Follow-up (m)	/	80.60 ± 36.2	84.06 ± 35.9	69.87 ± 34.9	/	
Conversion rate	1	23 (18.11%)	11 (11.46%)	12 (38.71%)	/	

Abbreviations: MMSE, Mini-Mental State Examination; MoCA, Montreal Cognitive Assessment; MoCA-B, Montreal Cognitive Assessment-Basic Version; CDR-SB, Clinical Dementia Rating Sum of Boxes; CN, cognitively normal; CN-, $A\beta$ negative cognitively normal; CN+, $A\beta$ positive cognitively normal; y, year; F, female; M, male; m, months; a, MoCA used in the ADNI cohort, and MoCA-B used in the Xuanwu cohort

* means p < 0.05 compared to the CN group in the ADNI cohort

** means p<0.001 compared to the CN group in the ADNI cohort

means p<0.001 compared to the CN- group in the ADNI cohort

(variance accounted for 22.9%, 9.2%, 6.0%, 3.9%, 3.2%, and 2.4%, and cumulative variance accounted for 49.7%) whose participant scores maximally correlated with chronological age. Brain regions that exhibited reduced glucose metabolism with ageing were primarily distributed in the left supplementary motor area (SMA.L), bilateral olfactory cortex (OLF), medial superior frontal gyrus (SFGmed), anterior cingulate and paracingulate gyri (ACG), caudate nucleus (CAU), and dorsal thalamus (THAdor). Brain regions manifesting increased glucose metabolism with ageing included the bilateral ventral thalamus (THAven) and part of the cerebellum (Cerebellum_Crus II) (Fig. 2A, Table 2). There were significant positive

correlations between network scores and chronological age for the CN participants in the combined cohort (R=0.808, p<0.001, Fig. 2B), ADNI cohort (R=0.785, p<0.001, Fig. 2C), and Xuanwu cohort (R=0.668, p<0.001, Fig. 2D).

The consistency of ARGMPs between different centres

Network analysis for CN participants in the ADNI cohort evaluated the first six PCs, including 50.0% participant×voxel variance. ARGMP_{ADNI} was identified by linear combination with PC2, PC3, PC4, and PC5 (variance accounted for 9.0%, 4.8%, 4.4%,



Fig. 2 ARGMPs in CN adults in the combined cohort during normal ageing. The red areas indicate a significant increase in glucose metabolism with ageing, and the blue areas indicate a significant decrease (A). Scatter plot and fit results, with 95% confidence limits, between network scores and CN participant age in the combined cohort (B), ADNI cohort (C), and

Xuanwu cohort (**D**); SMA, supplementary motor area; OLF, olfactory cortex; SFGmed, medial superior frontal gyrus; ACG, anterior cingulate and paracingulate gyri; CAU, caudate nucleus; THAdor, dorsal thalamus; THAven, bilateral ventral thalamus and part of Cerebellum_Crus II

Brain region		AAL number	Laterality	MNI Coordinates		
				X	Y	Z
Decreased Metabolism						
Supplementary motor area	Frontal	19	L	-5.3	4.8	61.4
Olfactory cortex	Prefrontal	21	L	-8.1	15.1	-11.5
Olfactory cortex	Prefrontal	22	R	10.4	15.9	-11.3
Superior frontal gyrus, medial	Prefrontal	23	L	-4.8	49.2	30.9
Superior frontal gyrus, medial	Prefrontal	24	R	9.1	50.8	30.2
Anterior cingulate and paracingulate gyri	Prefrontal	31	L	-4.0	35.4	14.0
Anterior cingulate and paracingulate gyri	Prefrontal	32	R	8.5	37.0	15.8
Caudate nucleus	Subcortical	71	L	-11.5	11.0	9.2
Caudate nucleus	Subcortical	72	R	14.8	12.1	9.4
Dorsal thalamus	Subcortical	77	L	-10.8	-17.6	8.0
Dorsal thalamus	Subcortical	78	R	13.0	-17.6	8.1
Increased Metabolism						
Ventral thalamus	Subcortical	77	L	-10.8	-17.6	8.0
Ventral thalamus	Subcortical	78	R	13.0	-17.6	8.1
Cerebellum_Crus II	Cerebellum	101	L	-	-	-
Cerebellum_Crus II	Cerebellum	103	L	-	-	-
Cerebellum_Crus II	Cerebellum	104	R	-	-	-

Table 2 Brain regions loading by the voxels survived from 1000 bootstrapping on ARGMP at p < 0.001, with absolute weight threshold of 1.4 and cluster threshold of 100 voxels (800 mm³)

and 3.4%, respectively, and cumulative variance accounted for 21.6%), whose participant scores maximally correlated with chronological age. Brain regions that exhibited reduced glucose metabolism with ageing were distributed in the SMA.L, bilateral OLF, SFGmed, ACG, CAU, THAdor, and part of the vermis (Vermis_3). Brain regions that manifested increased glucose metabolism with ageing included the right inferior occipital gyrus (IOG.R), THAven, and part of the cerebellum (Cerebellum Crus II) (Fig. 3A, Table 3). Significant correlations between the network scores on the ARGMP_{ADNI} and chronological age of CN participants were observed in both the ADNI (R = 0.738, p < 0.001, Fig. 3B) and Xuanwu cohorts (R=0.586, p<0.001, Fig. 3C). In addition, the network scores of the ARGMP_{ADNI} and the ARGMP_{Xuanwu} exhibited highly similar topographies (R = 0.576, p < 0.001, Fig. 3D).

Network analysis for the CN participants in the Xuanwu cohort included the evaluation of the first nine PCs, including 50.1% participant × voxel variance. ARGMP_{Xuanwu} was identified by the linear combination with PC1, PC2, PC3, PC6, and PC7

(variance accounted for 23.9%, 6.2%, 4.9%, 2.4%, and 2.3%, respectively, and cumulative variance accounted for 39.7%), whose participant scores maximally correlated with chronological age. Brain regions characterized by reduced glucose metabolism with ageing included the left orbital part of the middle frontal gyrus (ORBmid.L), SMA.L, left medial orbital of the superior frontal gyrus (ORBsupmed.L), bilateral SFGmed, ACG, median cingulate and paracingulate gyri (DCG), and CAU. Brain regions displaying increased glucose metabolism with ageing were distributed in the left paracentral lobule (PCL.L), left temporal pole: superior and middle temporal gyrus (TPOsup.L, TPOmid.L), and part of the left inferior cerebellum (Fig. 4A, Table 4). Significant correlations were observed between the ARGMP_{Xuanwu} network scores and the chronological age of the CN participants in both the Xuanwu (R = 0.734, p < 0.001, Fig. 4B) and ADNI cohorts (R = 0.608, p < 0.001, Fig. 4C). The correlation between network scores of the ARGMP_{ADNI} and the ARGMP_{Xuanwu} was significantly positive (R = 0.737, p < 0.001, Fig. 4D).



Fig. 3 ARGMP_{ADNI} in CN adults in the ADNI cohort during normal ageing. The red areas indicate a significant increase in glucose metabolism with ageing, and the blue areas indicate a significant decrease (**A**). Scatter plot and the fit results, with 95% confidence limits, between network scores and the age of the CN participants in the ADNI (**B**) and Xuanwu cohorts (**C**). Scatter plot and fit results with 95% confidence limits and between network scores on the ARGMP_{ADNI} and

ARGMP_{Xuanwu} of the CN participants in the ADNI cohort (**D**). Correlations between the network scores and chronological age in the CN- (E) and CN+(F) subgroups in the ADNI cohort. SMA, supplementary motor area; OLF, olfactory cortex; SFGmed, medial superior frontal gyrus; ACG, anterior cingulate and paracingulate gyri; CAU, caudate nucleus; THAdor, dorsal thalamus; THAven, bilateral ventral thalamus; and part of Cerebellum_CrusII

The correlation between network scores and chronological age in CN subgroups

Network analysis for the CN- and CN+subgroups in the ADNI cohort was also performed. For CN- participants, there was a significantly positive correlation between network scores and chronological age (R=0.753, p<0.001, Fig. 3E). Meanwhile, a significant association between network scores and chronological age was also observed in CN+participants (R=0.640, p<0.001, Fig. 3F).

Brain region		AAL number	Laterality	MNI Coordinates		
				x	Y	Z
Decreased Metabolism						
Supplementary motor area	Frontal	19	L	-5.3	4.8	61.4
Olfactory cortex	Prefrontal	21	L	-8.1	15.1	-11.5
Olfactory cortex	Prefrontal	22	R	10.4	15.9	-11.3
Superior frontal gyrus, medial	Prefrontal	23	L	-4.8	49.2	30.9
Superior frontal gyrus, medial	Prefrontal	24	R	9.1	50.8	30.2
Anterior cingulate and paracingulate gyri	Prefrontal	31	L	-4.0	35.4	14.0
Anterior cingulate and paracingulate gyri	Prefrontal	32	R	8.5	37.0	15.8
Caudate nucleus	Subcortical	71	L	-11.5	11.0	9.2
Caudate nucleus	Subcortical	72	R	14.8	12.1	9.4
Dorsal thalamus	Subcortical	77	L	-10.8	-17.6	8.0
Dorsal thalamus	Subcortical	78	R	13.0	-17.6	8.1
Vermis	Vermis	110	R	-	-	-
Increased Metabolism						
Inferior occipital gyrus	Occipital	54	R	38.2	-82	-7.6
Ventral thalamus	Subcortical	77	L	-10.8	-17.6	8.0
Ventral thalamus	Subcortical	78	R	13.0	-17.6	8.1
Cerebellum_Crus II	Cerebellum	101	L	-	-	-
Cerebellum_Crus II	Cerebellum	103	L	-	-	-
Cerebellum_Crus II	Cerebellum	104	R	-	-	-

Table 3 Brain regions loading by the voxels survived from 1000 bootstrapping on ARGMP_{ADNI} at p < 0.001, with absolute weight threshold of 1.4 and cluster threshold of 100 voxels (800 mm³)

The predictive effect of ARGMPs on the development of cognitive impairment at follow-up

As shown in Fig. 5A, conversion stages (stage $0 \sim$ stage 3) were defined by thresholds of the A β and standardized chronological age. The rates of conversion in the four stages were 7.7%, 15.9%, 22.2%, and 45.5%, respectively. In Fig. 5B, four stages were classified by thresholds of the A β and standardized network score. The results showed that the rates of conversion in the four stages were 6.9%, 18.4%, 18.2%, and 50%. The conversion rate in stage 1 was lower in $A\beta$ -age+individuals (Fig. 5A) than in $A\beta$ -Network scores + individuals (Fig. 5B) (15.9% vs. 18.4%), while the conversion rate in stage 2 is higher in Fig. 5A than in Fig. 5B (22.2% vs. 18.2%). In stage 3, participants with $A\beta$ + Network scores + exhibited a higher conversion rate than those with $A\beta + Age + in$ the prediction models (45.5% vs. 50%).

In the survival analysis, higher network scores (standardized network score > 0, green line) indicated a better predictive effect for developing a clinical

diagnosis of cognitive impairment than lower network scores (standardized network score < 0, blue line) (HR: 0.30, 95% CI: 0.1340~0.6904, p=0.0068, Fig. 5C). However, there was no significant difference between the older group (standardized chronological age>0, orange line) and the younger group (standardized chronological age < 0, red line) (HR: 0.50, 95% CI: $0.2193 \sim 1.154$, p = 0.1250, Fig. 5C). Furthermore, the predictive effect of the conversion risk to cognitive impairment in the $A\beta$ + Network score + subgroup compared to that in the other three subgroups was as follows (Fig. 5D): the A β -Network score- subgroup (HR: 0.10, 95% CI: 0.0270~0.3704, p < 0.0001), A β -Network score + subgroup (HR: 0.21, 95% CI: $0.0668 \sim 0.6429$, p = 0.0002), and $A\beta$ + Network score- subgroups (HR: 0.25, 95% CI: $0.0810 \sim 0.7906$, p=0.035). However, compared with the other three subgroups, the predictive effect of the conversion risk in the $A\beta + Age + subgroup$ was as follows (Fig. 5D): the A β -Age- subgroup (HR: 0.14, 95% CI: 0.0428~0.4650, p<0.0001), Aβ-Age+subgroup (HR: 0.20, 95% CI: 0.0628~0.6227,



Fig. 4 ARGMP_{Xuanwu} in CN adults in the Xuanwu cohort during normal ageing. The red areas indicate a significant increase in glucose metabolism associated with advancing age, and the blue areas indicate a significant decrease (A). Scatter plot and fit results, with 95% confidence limits, between network scores and the age of the CN participants in the Xuanwu cohort (B) and ADNI cohort (C). Scatter plot and the fit results, with 95% confidence limits and between network scores in the

p=0.0002), and A β +Age- subgroups (HR: 0.38, 95% CI: 0.1162~1.260, p=0.157).

Discussion

This study proposes ARGMPs as a novel index to characterize individual brain ageing. The primary findings were that (1) ARGMPs appear to be a stable and sensitive index for characterizing brain ageing; (2) this index is independent of AD pathological biomarkers; and (3) ARGMPs display better potential for predicting the risk of cognitive impairment than chronological age in CN people.

Compared to chronological age, the novel index appeared to be more reasonable in assessing normal brain ageing. Using the SSM/PCA method, brain ageing-specific spatial metabolic profiles were generated

 $ARGMP_{Xuanwu}$ and $ARGMP_{ADNI}$ of the CN participants in the ADNI cohort (**D**). SMA, supplementary motor area; SFGmed, medial superior frontal gyrus; ACG, anterior cingulate and paracingulate gyri; CAU, caudate nucleus; DCG, median cingulate and paracingulate gyri; ORBmid, orbital part of the middle frontal gyrus; TPO, temporal pole and part of Cerebellum_Crus II

from linear combinations of several principal components [25]. This multivariate analysis method reduces the interindividual variations in FDG-PET images and provides complementary, clinically relevant information that may be superior to univariate measures [26]. In our study, ARGMPs derived from the ADNI and Xuanwu cohorts exhibited similar topographies, validating the robustness and repeatability of our findings. According to the biological definition of AD proposed in 2018 [27], individuals with evidence of A β deposition are categorized into the Alzheimer's continuum. Thus, to eliminate the latent influence of AD pathology, we further analysed ARGMPs from the CN- and CN+subgroups. Notably, significantly positive correlations between network score and chronological age were also observed in these two subgroups. Our findings validated that the novel index was independent of AD pathological biomarkers. Moreover, given the potential impact of cognitive

Brain region		AAL number	Laterality	MNI Coordinates		
				X	Y	Z
Decreased Metabolism						
Middle frontal gyrus, orbital part	Prefrontal	9	L	-30.6	50.4	-9.6
Supplementary motor area	Frontal	19	L	-5.3	4.8	61.4
Superior frontal gyrus, medial	Prefrontal	23	L	-4.8	49.2	30.9
Superior frontal gyrus, medial	Prefrontal	24	R	9.1	50.8	30.2
Superior frontal gyrus, medial orbital	Prefrontal	25	L	-5.2	54.1	-7.4
Anterior cingulate and paracingulate gyri	Prefrontal	31	L	-4.0	35.4	14.0
Anterior cingulate and paracingulate gyri	Prefrontal	32	R	8.5	37.0	15.8
Median cingulate and paracingulate gyri	Frontal	33	L	-5.5	-14.9	41.6
Median cingulate and paracingulate gyri	Frontal	34	R	8.0	-8.8	39.8
Caudate nucleus	Subcortical	71	L	-11.5	11.0	9.2
Caudate nucleus	Subcortical	72	R	14.8	12.1	9.4
Increased Metabolism						
Paracentral lobule	Parietal	69	L	-7.6	-25.4	70.1
Temporal pole: superior temporal gyrus	Temporal	83	L	-39.9	15.1	-20.2
Temporal pole: middle temporal gyrus	Temporal	87	L	-36.3	14.6	-34.1
Cerebellum_Crus II	Cerebellum	103	L	-	-	-

Table 4 Brain regions loading by the voxels survived from 1000 bootstrapping on $ARGMP_{Xuanwu}$ at P < 0.001, with absolute weight threshold of 1.4 and cluster threshold of 100 voxels (800 mm³)

reserve on age-related brain changes [28–30], other risk factors (e.g. educational level, sex) were also regressed as covariates in our study, which further ensured that the novel index was stable.

The novel index revealed that the overlapping brain regions with age-related hypometabolism focused on the bilateral SFGmed, ACG, CAU, and left SMA based on the three cohorts (the combined cohort, the ADNI cohort, and the Xuanwu cohort), whereas age-related hypermetabolism was primarily distributed in part of the left Cerebellum Crus II. Thus, age-related regional hypometabolism was primarily observed in the frontal lobe and striatum, suggesting that these regions may be more sensitive to normal brain ageing. The involvement of the prefrontal cortex (e.g. SFGmed, ACG) in normal brain ageing has been supported by previous studies [7, 31, 32]. In contrast with AD, the prefrontal cortex is regarded as the first structure affected by normal ageing. For instance, Moeller [33] provided evidence of decreased glucose metabolism in the medial frontal region of 130 healthy volunteers aged 21-90 years. Age-related differences in working memory loadrelated activity have also been observed in the SFGmed [32]. The prefrontal cortex has multiple physiological functions, especially in working memory, attention, and mood regulation [31, 34, 35]. The decline in glucose metabolism in the prefrontal cortex with increasing age may be attributed to several potential mechanisms, such as decreased neuronal function [36] and altered synaptic efficiency [7].

Our study also demonstrated decreased glucose metabolism in the left SMA with normal brain ageing. The SMA, which is a critical component of the cortical motor network, plays a major role in planning and executing rhythmic bilateral movements [37]. Growing evidence indicates that age-related decline in motor control is due to special brain structural and functional changes, such as grey matter atrophy in the pre- and postcentral gyri [38], reduced white matter integrity in the corpus callosum [39], abnormal functional connectivity, or hypometabolism within the cortical motor network. Current studies have reported decreased cerebral glucose metabolism in the left SMA in healthy older adults [40] and reduced SMA-primary motor cortex (M1) interactions [41] with ageing. In contrast with normal brain ageing, AD pathology initially affects the entorhinal cortex, while motor areas are generally damaged during the late stage. Therefore,



Fig. 5 Staging the transformers and survival analysis estimated by the Kaplan–Meier method. According to the threshold of A β (1.18) and the standardized chronological age (0), CN participants in the ADNI cohort were classified into four stages (stage 0 to 3). In each stage, the conversion rate of CN participants to cognitive impairment was calculated (**A**). According to the threshold of A β (1.18) and the standardized network score (0), CN participants in the ADNI cohort were also classified into four stages (stage 0 to 3). In each stage, the conversion rate of CN participants in the ADNI cohort were also classified into four stages (stage 0 to 3). In each stage, the conversion rate of CN participants to cognitive impairment was calculated (**B**). In the survival analysis, we compared different predictive effects of conversion risk between higher net-

age-related hypometabolism in the SMA may represent a distinct biomarker for identifying normal brain ageing. In our study, the subcortical CAU was another region manifesting significant metabolic decline with ageing. The CAU, a key component of the striatum, is associated with navigational skills [42, 43]. Previous work has revealed age-related reductions in perfusion in the CAU [44, 45].

Finally, we investigated the ability of ARGMPs to predict the conversion risk of CN individuals to cognitive impairment. Regional hypometabolism in AD patients has been reported in many previous studies and may represent a predictor of cognitive decline from mild cognitive impairment (MCI) to AD

work scores (standardized network score > 0, green line) and lower network scores (standardized network score > 0, blue line) (C) and between the older group (standardized chronological age > 0, orange line) and the younger group (standardized chronological age < 0, red line) (C). Furthermore, we compared different predictive effects of conversion risk among the standardized network score subgroup and the standardized chronological age subgroup: the A β +Network score+(stage 3), A β -Network score- (stage 0), A β -Network score+(stage 1), and A β +Network score- (stage 2); the A β +Age+(stage 3), A β -Age- (stage 0), A β -Age+(stage 1), and A β +Age- (stage 2) (D)

dementia [46, 47]. Decreased glucose metabolism is considered a biomarker of neurodegeneration, which is associated with the clinical progression of AD [48]. In this study, participants with evidence of A β deposition and high network scores exhibited higher conversion rates than other three subgroups, indicating that the combination of A β burden and ARGMPs has the potential to identify individuals at risk of developing cognitive impairment. We also assessed the prediction effect of chronological age. To the best of our knowledge, the prevalence of AD and the accumulation of A β increase with ageing [49]. We also found that older people with A β positivity evidence (stage 3) presented a relatively higher conversion rate than those at other stages, consistent with previous studies [50]. However, compared to chronological age, individuals with higher network scores had a higher risk of conversion than those with lower network scores.

It should be noted that this study has some limitations. First, age differences existed between the ADNI and Xuanwu cohorts, with the average age of the participants from Xuanwu Hospital being 8.6 years younger than those in the ADNI cohort. Correlations between ARGMP expression network scores and chronological age were identified in both cohorts, but whether this correlation exists in CN populations with wider age ranges needs to be validated in the future. Second, the underlying mechanism of the age-related metabolic pattern in older CN adults was not elucidated in the present study. Future studies are needed to investigate the development of ARGMPs through molecular pathways, such as neurotransmitter and endocrine signalling, etc. Third, the effect of other factors, especially biological sex, on the correlation between brain regional glucose metabolism dysfunction and chronological ageing was not considered in this study. Notably, sex is a key risk factor that contributes to variability in AD manifestation [51]. Previous studies have reported that females are at greater risk of developing AD, as well as exhibit more pathological changes [52–54]. However, the effect of biological sex on brain glucose metabolism remains unclear. Thus, the impact of sex differences on ARGMPs needs to be evaluated in future studies. Finally, the sample size in this study was relatively small, and only data in the ADNI cohort were longitudinal. Future multicentre studies with a larger sample size are essential to provide more accurate evidence. In addition, the similarities and differences between AD-related ageing and chronological ageing patterns warrant further study.

Conclusions

This study revealed ARGMP topographies in the spatial covariance brain network and the distribution of regional metabolism alterations that occur during normal brain ageing using ¹⁸F-FDG PET images. Crossvalidation between the ADNI and Xuanwu cohorts confirmed highly reproducible ARGMPs across different CN populations and imaging instrumentation/protocols. The expression score of ARGMPs in individual participants objectively mirrored normal brain ageing, such as chronological age, and better predicted the conversion of CN participants to cognitive impairment. This study indicates that the glucose metabolism pattern in CN older adults may represent an effective biomarker of normal brain ageing and has the potential for future application in routine clinical practice.

Acknowledgements Data collection and dissemination for this project were funded by the Alzheimer's Disease Neuroimaging Initiative (ADNI): the National Institutes of Health (grant number U01 AG024904), and the Department of Defense (award numberW81XWH-12-2-0012). ADNI is funded by the National Institute of Aging and the National Institute of Biomedical Imaging and Bioengineering as well as through generous contributions from the following organizations: AbbVie, Alzheimer's Association, Alzheimer's Drug Discovery Foundation, Araclon Biotech, BioClinica Inc., Biogen, Bristol-Myers Squibb Company, CereSpir Inc., Eisai Inc., Elan Pharmaceuticals Inc., Eli Lilly and Company, EuroImmun, F. Hoffmann-La Roche Ltd. and its affiliated company Genentech Inc., Fujirebio, GE Healthcare, IXICO Ltd., Janssen Alzheimer Immunotherapy Research & Development LLC., Johnson & Johnson Pharmaceutical Research & Development LLC., Lumosity, Lundbeck, Merck & Co. Inc., Meso Scale Diagnostics LLC., NeuroRx Research, Neurotrack Technologies, Novartis Pharmaceuticals Corporation, Pfizer Inc., Piramal Imaging, Servier, Takeda Pharmaceutical Company, and Transition Therapeutics. The Canadian Institutes of Health Research are providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health (www.fnih.org). The grantee organization is the Northern California Institute for Research and Education, and the study is coordinated by the Alzheimer's Disease Cooperative Study at the University of California, San Diego, CA, USA. ADNI data are disseminated by the Laboratory for Neuro Imaging at the University of Southern California, CA, USA.

Michael W. Weiner⁷, Paul Aisen⁸, Ronald Petersen⁹, Clifford R. Jack⁹, William Jagust¹⁰, John Q. Trojanowski¹¹, Arthur W. Toga¹², Laurel Beckett¹³, Robert C. Green¹⁴, Andrew J. Saykin¹⁵, John Morris¹⁶, Leslie M. Shaw¹¹, Zaven Khachaturian^{13,17}, Greg Sorensen¹⁸, Lew Kuller¹⁹, Marcus Raichle¹⁶, Steven Paul²⁰, Peter Davies²¹, Howard Fillit²², Franz Hefti²³, David Holtzman¹⁶, Marek M. Mesulam²⁴, William Potter²⁵, Peter Snyder²⁶, Adam Schwartz²⁷, Tom Montine²⁸, Ronald G. Thomas²⁸, Michael Donohue²⁸, Sarah Walter²⁸, Devon Gessert²⁸, Tamie Sather²⁸, Gus Jiminez²⁸, Danielle Harvey¹³, Matthew Bernstein⁹, Paul Thompson²⁹, Norbert Schuff^{7,13}, Bret Borowski⁹, Jeff Gunter⁹, Matt Senjem⁹, Prashanthi Vemuri⁹, David Jones⁹, Kejal Kantarci⁹, Chad Ward⁹, Robert A. Koeppe³⁰, Norm Foster³¹, Eric M. Reiman³², Kewei Chen³², Chet Mathis¹⁹, Susan Landau¹⁰, Nigel J. Cairns¹⁶, Erin Householder¹⁶, Lisa Taylor-Reinwald¹⁶, Virginia Lee¹¹, Magdalena Korecka¹¹, Michal Figurski¹¹, Karen Crawford¹², Scott Neu¹², Tatiana M. Foroud¹⁵, Steven G. Potkin³³, Li Shen¹⁵, Kelley Faber¹⁵, Sungeun Kim¹⁵, Kwangsik Nho¹⁵, Leon Thal⁸, Neil Buckholtz³⁴, Marylyn Albert³⁵, Richard Frank³⁶, John Hsiao³⁴, Jeffrey Kaye³⁷, Joseph Quinn³⁷, Betty Lind³⁷, Raina Carter³⁷, Sara Dolen³⁷, Lon S. Schneider¹², Sonia Pawluczyk¹², Mauricio Beccera¹², Liberty Teodoro¹², Bryan M. Spann¹², James Brewer⁸, Helen Vanderswag⁸, Adam Fleisher^{8,32}, Judith L. Heidebrink³⁰, Joanne L. Lord³⁰, Sara S. Mason⁹, Colleen S. Albers⁹, David Knopman⁹, Kris Johnson⁹, Rachelle S. Doody³⁸, Javier Villanueva-Meyer³⁸, Munir Chowdhury³⁸, Susan Rountree³⁸, Mimi Dang³⁸, Yaakov Stern³⁸, Lawrence S. Honig³⁸, Karen L. Bell³⁸, Beau Ances¹⁶, Maria Carroll¹⁶, Sue Leon¹⁶, Mark A. Mintun¹⁶, Ances , Mara Carlon , Sue Leon , Mark A. Mintun , Stacy Schneider¹⁶, Angela Oliver¹⁶, Daniel Marson³⁹, Ran-dall Griffith³⁹, David Clark³⁹, David Geldmacher³⁹, John Brockington³⁹, Erik Roberson³⁹, Hillel Grossman⁴⁰, Effie Mitsis⁴⁰, Leyla de Toledo-Morrell⁴¹, Raj C. Shah⁴¹, Ranjan Duara⁴², Daniel Varon⁴², Maria T. Greig⁴², Peggy Roberts⁴² Chiadi Onyike³⁵, Daniel D'Agostino³⁵, Stephanie Kielb³⁵, James E. Galvin⁴³, Brittany Cerbone⁴³, Christina A. Michel⁴³, Henry Rusinek⁴³, Mony J. de Leon⁴³, Lidia Glodzik4³, Susan De Santi⁴³, P Murali Doraiswamy⁴⁴, Jeffrey R. Petrella⁴⁴, Terence Z. Wong44, Steven E. Arnold11, Jason H. Karlawish11, David Wolk¹¹, Charles D. Smith⁴⁵, Greg Jicha⁴⁵, Peter Hardy⁴⁵, Partha Sinha45, Elizabeth Oates45, Gary Conrad45, Oscar L. Lopez¹⁹, MaryAnn Oakley¹⁹, Donna M. Simpson³⁵, Anton P. Porsteinsson⁴⁶, Bonnie S. Goldstein⁴⁶, Kim Martin⁴⁶, Kelly M. Makino⁴⁶, M Saleem Ismail⁴⁶, Connie Brand⁴⁶, Ruth A. Mulnard³³, Gaby Thai³³, Catherine McAdams-Ortiz³³, Kyle Womack⁴⁷, Dana Mathews⁴⁷, Mary Quiceno⁴⁷, Ramon Diaz-Arrastia⁴⁷, Richard King⁴⁷, Myron Weiner⁴⁷, Kristen Martin-Cook⁴⁷, Michael DeVous⁴⁷, Allan I Levey⁴⁸, James J. Lah⁴⁸, Janet S. Cellar⁴⁸, Jeffrey M. Burns⁴⁸, Heather S. Anderson⁴⁹, Russell H. Swerdlow⁴⁹, Liana Apostolova²⁹, Kathleen Tingus²⁹, Ellen Woo²⁹, Daniel H.S. Silverman²⁹, Po Rathleen Tingus⁻⁷, Eilen Woo⁻⁷, Daniel H.S. Silverman⁻⁷, Po H. Lu²⁹, George Bartzokis²⁹, Neill R. Graff-Radford⁵⁰, Fran-cine Parfitt⁵⁰, Tracy Kendall⁵⁰, Heather Johnson⁵⁰, Martin R. Farlow¹⁵, Ann Marie Hake¹⁵, Brandy R. Matthews¹⁵, Scott Herring¹⁵, Cynthia Hunt¹⁵, Christopher H. van Dyck⁵¹, Richard E. Carson⁵¹, Martha G. MacAvoy⁵¹, Howard Chertkow⁵², How-ard Bergman⁵², Chris Hosein⁵², Ging-Yuek Robin Hsiung⁵³, Howard Faldmor⁵³, Bonita, Mudga⁵³, Michela Accelu⁵³, Howard Feldman⁵³, Benita Mudge⁵³, Michele Assaly⁵³, Charles Bernick⁵⁴, Donna Munic⁵⁴, Andrew Kertesz⁵⁵, John Rogers⁵⁵, Dick Trost⁵⁵, Diana Kerwin²⁴, Kristine Lipowski²⁴, Chuang-Kuo Wu²⁴, Nancy Johnson²⁴, Carl Sadowsky⁵⁶, Walter Martinez⁵⁶, Teresa Villena⁵⁶, Raymond Scott Turner⁵⁷ Kathleen Johnson⁵⁷, Brigid Reynolds⁵⁷, Reisa A. Sperling¹⁴, Keith A. Johnson¹⁴, Gad Marshall¹⁴, Meghan Frey¹⁴, Barton Lane¹⁴, Allyson Rosen¹⁴, Jared Tinklenberg¹⁴, Marwan N. Sabbagh⁵⁸, Christine M. Belden⁵⁸, Sandra A. Jacobson⁵⁸, Sherye A. Sirrel⁵⁸, Neil Kowall⁵⁸, Ronald Killiany⁵⁹, Andrew E. Budson⁵⁹, Alexander Norbash⁵⁹, Patricia Lynn Johnson⁵⁹, Joanne Allard⁵⁹, Alan Lerner⁶¹, Paula Ogrocki⁶¹, Leon Hudson⁶¹, Evan Fletcher¹³, Owen Carmichae²³, John Olichney¹³, Charles DeCarli¹³, Smita Kittur⁶², Michael Borrie⁶³, T-Y. Lee⁶³, Rob Bartha⁶³, Sterling Johnson⁶⁴, Sanjay Asthana⁶⁴, Cynthia M. Carlsson⁶⁴, Adrian Preda²⁹, Dana Nguyen²⁹, Pierre Tariot³¹, Stephanie Reeder³¹, Vernice Bates⁶⁵, Horacio Capote⁶⁵, Michelle Rainka⁶⁵, Douglas W. Scharre⁶⁶, Maria Kataki⁶⁶, Anahita Adeli⁶⁶, Earl A. Zimmerman⁶⁷, Dzintra Celmins⁶⁷, Alice D. Brown⁶⁷, Godfrey D. Pearlson⁶⁸, Karen Blank⁶⁸, Karen Anderson⁶⁸, Robert B. Santulli⁶⁹, Tamar J. Kitzmiller⁶⁹, Eben S. Schwartz⁶⁹, Kaycee M. Sink⁷⁰, Jeff D. Williamson⁷⁰, Pradeep Garg⁷⁰, Franklin Watkins⁷⁰, Brian R. Ott⁷¹, Henry Querfurth⁷¹, Geoffrey Tremont⁷¹, Stephen

Salloway⁷², Paul Malloy⁷², Stephen Correia⁷², Howard J. Rosen⁷, Bruce L. Miller⁷, Jacobo Mintzer⁷³, Kenneth Spicer⁷³, David Bachman⁷³, Stephen Pasternak⁵⁵, Irina Rachinsky⁵⁵, Dick Drost⁵⁵, Nunzio Pomara⁷⁴, Raymundo Hernando⁷⁴, Antero Sarrael⁷⁴, Susan K. Schultz⁷⁵, Laura L. Boles Ponto⁷⁵, Hyungsub Shim⁷⁵, Karen Elizabeth Smith⁷⁵, Norman Relkin²⁰, Gloria Chaing²⁰, Lisa Raudin^{17,20}, Amanda Smith⁷⁶, Kristin Fargher⁷⁶, Balebail Ashok Raj⁷⁶, Thomas Neylan⁷, Jordan Grafman²⁴, Melissa Davis⁸, Rosemary Morrison⁸, Jacqueline Hayes⁷, Shannon Finley⁷, Karl Friedl⁷⁷, Debra Fleischman⁴¹, Konstantinos Arfanakis⁴¹, Olga James⁴⁴, Dino Massoglia⁷³, J Jay Fruehling⁶⁴, Sandra Harding⁶⁴, Elaine R. Peskind²⁸, Eric C. Petrie⁶⁶, Gail Li⁶⁶, Jerome A. Yesavage⁷⁸, Joy L. Taylor⁷⁸ & Ansgar J. Furst⁷⁸

⁷UC San Francisco, San Francisco, CA 94143, USA. ⁸UC San Diego, San Diego, CA 92093, USA. 9Mayo Clinic, Rochester, NY 14603, USA. ¹⁰UC Berkeley, Berkeley, CA 94720, USA. ¹¹UPenn, Philadelphia, PA 9104, USA. ¹²USC, Los Angeles, CA 90089, USA. ¹³UC Davis, Davis, CA 95616, USA. ¹⁴Brigham and Women's Hospital/Harvard Medical School, Boston, MA 02115, USA. ¹⁵Indiana University, Bloomington, IN 47405, USA. ¹⁶Washington University in St Louis, St Louis, MI 63130, USA. ¹⁷Prevent Alzheimer's Disease 2020, Rockville, MD 20850, USA. ¹⁸Siemens, Munich 80333, Germany. ¹⁹University of Pittsburgh, Pittsburgh, PA 15260, USA. ²⁰Weill Cornell Medical College, Cornell University, New York City, NY 10065, USA. ²¹Albert Einstein College of Medicine of Yeshiva University, Bronx, NY 10461, USA. ²²AD Drug Discovery Foundation, New York City, NY 10019, USA. ²³Acumen Pharmaceuticals, Livermore, CA 94551, USA. 24Northwestern University, Evanston and Chicago, IL 60208, USA. ²⁵National Institute of Mental Health, Rockville, MD 20852, USA. ²⁶Brown University, Providence, RI 02912, USA. ²⁷Eli Lilly, Indianapolis, IN 46225, USA. 28 University of Washington, Seattle, WA 98195, USA. ²⁹UCLA, Los Angeles, CA 90095, USA. ³⁰University of Michigan, Ann Arbor, MI 48109, USA. ³¹University of Utah, Salt Lake City, UT 84112, USA. ³²Banner Alzheimer's Institute, Phoenix, AZ 85006, USA. ³³UC Irvine, Irvine, CA 92697, USA. ³⁴National Institute on Aging, Bethesda, MD 20892, USA. 35Johns Hopkins University, Baltimore, MD 21218, USA. ³⁶Richard Frank Consulting, Washington, DC 20001, USA. ³⁷Oregon Health and Science University, Portland, OR 97239, USA. ³⁸Baylor College of Medicine, Houston, TX 77030, USA. ³⁹University of Alabama, Birmingham, AL 35233, USA. ⁴⁰Mount Sinai School of Medicine, New York City, NY 10029, USA. ⁴¹Rush University Medical Center, Chicago, IL 60612, USA. 42Wien Center, Miami, FL 33140, USA. ⁴³New York University, New York City, NY 10003, USA. ⁴⁴Duke University Medical Center, Durham, NC 27710, USA. ⁴⁵University of Kentucky, Lexington, KY 0506, USA. ⁴⁶University of Rochester Medical Center, Rochester, NY 14642, USA. 47University of Texas Southwestern Medical School, Dallas, TX 75390, USA. ⁴⁸Emory University, Atlanta, GA 30322, USA. 49Medical Center, University of Kansas, Kansas City, KS 66103, USA. ⁵⁰Mayo Clinic, Jacksonville, FL 32224, USA. ⁵¹Yale University School of Medicine, New Haven, CT 06510, USA. 52McGill University/Montreal-Jewish General Hospital, Montreal, QC H3T 1E2, Canada. ⁵³University of British Columbia Clinic for AD & Related Disorders, Vancouver, BC V6T 1Z3, Canada. 54Cleveland Clinic Lou Ruvo Center for Brain Health, Las Vegas, NV 89106, USA.

⁵⁵St Joseph's Health Care, London, ON N6A 4V2, Canada. ⁵⁶Premiere Research Institute, Palm Beach Neurology, Miami, FL 33407, USA. ⁵⁷Georgetown University Medical Center, Washington, DC 20007, USA. 58 Banner Sun Health Research Institute, Sun City, AZ 85351, USA. 59 Boston University, Boston, MA 02215, USA. ⁶⁰Howard University, Washington, DC 20059, USA. 61Case Western Reserve University, Cleveland, OH 20002, USA. ⁶²Neurological Care of CNY, Liverpool, NY 13088, USA. 63Parkwood Hospital, London, ON N6C 0A7, Canada. ⁶⁴University of Wisconsin, Madison, WI 53706, USA. ⁶⁵Dent Neurologic Institute, Amherst, NY 14226, USA. ⁶⁶Ohio State University, Columbus, OH 43210, USA. ⁶⁷Albany Medical College, Albany, NY 12208, USA. 68 Hartford Hospital, Olin Neuropsychiatry Research Center, Hartford, CT 06114, USA. 69Dartmouth- Hitchcock Medical Center, Lebanon, NH 03766, USA. ⁷⁰Wake Forest University Health Sciences, Winston-Salem, NC 27157, USA. 71Rhode Island Hospital, Providence, RI 02903, USA. 72Butler Hospital, Providence, RI 02906, USA. ⁷³Medical University South Carolina, Charleston, SC 29425, USA. 74Nathan Kline Institute, Orangeburg, NY 10962, USA. ⁷⁵University of Iowa College of Medicine, Iowa City, IA 52242, USA. ⁷⁶University of South Florida: USF Health Byrd Alzheimer's Institute, Tampa, FL 33613, USA. ⁷⁷Department of Defense, Arlington, VA 22350, USA. ⁷⁸Stanford University, Stanford, CA 94305, USA

Author contributions JJ and YH conceived of the study. CL and XJ performed the statistical analysis. CS, GC, SJ drafted the initial manuscript. XJ and LL drew the pictures. All authors contributed to revision and editing of the manuscript.

Funding This study was supported by grants received from the National Natural Science Foundation of China (grant numbers 61633018, 82020108013, 61603236, 81830059, and 81801052); the National Key Research and Development Program of China (grant numbers 2016YFC1306300, 2018YFC1312000, and 2018YFC1707704); the 111 Project (grant number D20031); the Shanghai Municipal Science and Technology Major Project (grant number 2017SHZDZX01); and the Beijing Municipal Commission of Health and Family Planning (grant number PXM2020_026283_000002).

Availability of data and materials All data are available upon request from the authors.

Declarations

Conflict of interest The authors declare no conflict of interest.

References

- 1. Fjell AM, McEvoy L, Holland D, Dale AM, Walhovd KB. What is normal in normal aging? Effects of aging, amyloid and Alzheimer's disease on the cerebral cortex and the hippocampus. Prog Neurobiol. 2014;117:20–40.
- Toepper M. Dissociating Normal Aging from Alzheimer's Disease: A View from Cognitive Neuroscience. J Alzheimer's Dis. 2017;57:331–52.

- Mattson MP, Arumugam TV. Hallmarks of Brain Aging: Adaptive and Pathological Modification by Metabolic States. Cell Metab. 2018;27:1176–99.
- Grimm A, Friedland K, Eckert A. Mitochondrial dysfunction: the missing link between aging and sporadic Alzheimer's disease. Biogerontology. 2016;17:281–96.
- Błaszczyk JW. Energy metabolism decline in the aging brain-pathogenesis of neurodegenerative disorders. Metabolites. 2020;10(11):450.
- Cunnane S, Nugent S, Roy M, Courchesne-Loyer A, Croteau E, Tremblay S, et al. Brain fuel metabolism, aging, and Alzheimer's disease. Nutrition. 2011;27:3–20.
- Pardo JV, Lee JT, Sheikh SA, Surerus-Johnson C, Shah H, Munch KR, et al. Where the brain grows old: decline in anterior cingulate and medial prefrontal function with normal aging. Neuroimage. 2007;35:1231–7.
- Bi Q, Wang W, Niu N, Li H, Wang Y, Huang W, et al. Relationship between the disrupted topological efficiency of the structural brain connectome and glucose hypometabolism in normal aging. Neuroimage. 2021;226: 117591.
- Habeck C, Risacher S, Lee G, et al. Relationship between baseline brain metabolism measured using [18F]FDG PET and memory and executive function in prodromal and early Alzheimer's disease. Brain Imaging Behav. 2012;6(4):568–83.
- Chételat G, Landeau B, Salmon E, Yakushev I, Bahri MA, Mézenge F, et al. Relationships between brain metabolism decrease in normal aging and changes in structural and functional connectivity. Neuroimage. 2013;76:167–77.
- Adams JN, Lockhart SN, Li L, Jagust WJ. Relationships Between Tau and Glucose Metabolism Reflect Alzheimer's Disease Pathology in Cognitively Normal Older Adults. Cereb Cortex. 2019;29:1997–2009.
- 12. Inoue K, Ito H, Uchida S, Taki Y, Kinomura S, Tsuji I, et al. Decrease in glucose metabolism in frontal cortex associated with deterioration of microstructure of corpus callosum measured by diffusion tensor imaging in healthy elderly. Hum Brain Mapp. 2008;29:375–84.
- 13. Cole JH, Poudel RPK, Tsagkrasoulis D, Caan MWA, Steves C, Spector TD, et al. Predicting brain age with deep learning from raw imaging data results in a reliable and heritable biomarker. Neuroimage. 2017;163:115–24.
- de Lange AG, Anatürk M, Suri S, Kaufmann T, Cole JH, Griffanti L, et al. Multimodal brain-age prediction and cardiovascular risk: The Whitehall II MRI sub-study. Neuroimage. 2020;222: 117292.
- Beheshti I, Mishra S, Sone D, Khanna P, Matsuda H. T1-weighted MRI-driven Brain Age Estimation in Alzheimer's Disease and Parkinson's Disease. Aging Dis. 2020;11:618–28.
- Bergfield KL, Hanson KD, Chen K, Teipel SJ, Hampel H, Rapoport SI, et al. Age-related networks of regional covariance in MRI gray matter: Reproducible multivariate patterns in healthy aging. Neuroimage. 2010;49:1750–9.
- 17. Li TR, Dong QY, Jiang XY, Kang GX, Li X, Xie YY, et al. Exploring brain glucose metabolic patterns in cognitively normal adults at risk of Alzheimer's disease: A cross-validation study with Chinese and ADNI cohorts. Neuroimage Clin. 2022;33: 102900.

- Spetsieris PG, Ma Y, Dhawan V, Eidelberg D. Differential diagnosis of parkinsonian syndromes using PCA-based functional imaging features. Neuroimage. 2009;45:1241–52.
- Eidelberg D. Metabolic brain networks in neurodegenerative disorders: a functional imaging approach. Trends Neurosci. 2009;32:548–57.
- Nan Z, Gordon ML, Yilong M, Bradley C, Gomar JJ, Shichun P, et al. The Age-Related Perfusion Pattern Measured With Arterial Spin Labeling MRI in Healthy Subjects. Front Aging Neurosci. 2018;10:214.
- Teune LK, Strijkert F, Renken RJ, Izaks GJ, de Vries JJ, Segbers M, et al. The Alzheimer's disease-related glucose metabolic brain pattern. Curr Alzheimer Res. 2014;11:725–32.
- Wu T, Ma Y, Zheng Z, Peng S, Wu X, Eidelberg D, et al. Parkinson's disease-related spatial covariance pattern identified with resting-state functional MRI. J Cereb Blood Flow Metab. 2015;35:1764–70.
- Mudali D, Teune LK, Renken RJ, Leenders KL, Roerdink JB. Classification of Parkinsonian syndromes from FDG-PET brain data using decision trees with SSM/PCA features. Comput Math Methods Med. 2015;2015: 136921.
- Chen K, Roontiva A, Thiyyagura P, Lee W, Liu X, Ayutyanont N, et al. Improved power for characterizing longitudinal amyloid-β PET changes and evaluating amyloid-modifying treatments with a cerebral white matter reference region. J Nucl Med. 2015;56:560.
- Iizuka T, Kameyama M. Spatial metabolic profiles to discriminate dementia with Lewy bodies from Alzheimer disease. J Neurol. 2020;267:1960–9.
- Habeck C, Foster NL, Perneczky R, Kurz A, Alexopoulos P, Koeppe RA, et al. Multivariate and univariate neuroimaging biomarkers of Alzheimer's disease. Neuroimage. 2008;40:1503–15.
- Jack CR Jr, Bennett DA, Blennow K, Carrillo MC, Dunn B, Haeberlein SB, et al. NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease. Alzheimer's Dementia. 2018;14:535–62.
- Wilson RS, Yu L, Lamar M, Schneider JA, Boyle PA, Bennett DA. Education and cognitive reserve in old age. Neurology. 2019;92:e1041–50.
- Stern Y, Arenaza-Urquijo EM, Bartrés-Faz D, Belleville S, Cantilon M, Chetelat G, et al. Whitepaper: Defining and investigating cognitive reserve, brain reserve, and brain maintenance. Alzheimer's Dementia. 2020;16:1305–11.
- Pettigrew C, Soldan A, Zhu Y, Cai Q, Wang MC, Moghekar A, et al. Cognitive reserve and rate of change in Alzheimer's and cerebrovascular disease biomarkers among cognitively normal individuals. Neurobiol Aging. 2020;88:33–41.
- 31. Sotoudeh N, Namavar MR, Zarifkar A, Heidarzadegan AR. Age-dependent changes in the medial prefrontal cortex and medial amygdala structure, and elevated plus-maze performance in the healthy male Wistar rats. IBRO Rep. 2020;9:183–94.
- 32. Bennett IJ, Rivera HG, Rypma B. Isolating age-group differences in working memory load-related neural activity: assessing the contribution of working memory

capacity using a partial-trial fMRI method. Neuroimage. 2013;72:20–32.

- Moeller JR, Ishikawa T, Dhawan V, Spetsieris P, Mandel F, Alexander GE, et al. The metabolic topography of normal aging. J Cereb Blood Flow Metab. 1996;16:385–98.
- 34. Liu D, Gu X, Zhu J, Zhang X, Han Z, Yan W, et al. Medial prefrontal activity during delay period contributes to learning of a working memory task. Science. 2014;346:458–63.
- Günseli E, Aly M. Preparation for upcoming attentional states in the hippocampus and medial prefrontal cortex. Elife. 2020;9:e53191.
- Kakimoto A, Ito S, Okada H, Nishizawa S, Minoshima S, Ouchi Y. Age-Related Sex-Specific Changes in Brain Metabolism and Morphology. J Nucl Med. 2016;57:221–5.
- Rojkova K, Volle E, Urbanski M, Humbert F, Dell'Acqua F, Thiebaut de Schotten M. Atlasing the frontal lobe connections and their variability due to age and education: a spherical deconvolution tractography study. Brain Struct Funct. 2016;221:1751–66.
- Salat DH, Buckner RL, Snyder AZ, Greve DN, Desikan RS, Busa E, et al. Thinning of the cerebral cortex in aging. Cereb Cortex. 2004;14:721–30.
- Sullivan EV, Rohlfing T, Pfefferbaum A. Quantitative fiber tracking of lateral and interhemispheric white matter systems in normal aging: relations to timed performance. Neurobiol Aging. 2010;31:464–81.
- 40. Sakurai R, Fujiwara Y, Yasunaga M, Suzuki H, Kanosue K, Montero-Odasso M, et al. Association between Hypometabolism in the Supplementary Motor Area and Fear of Falling in Older Adults. Front Aging Neurosci. 2017;9:251.
- Green PE, Ridding MC, Hill KD, Semmler JG, Drummond PD, Vallence AM. Supplementary motor area-primary motor cortex facilitation in younger but not older adults. Neurobiol Aging. 2018;64:85–91.
- Sodums DJ, Bohbot VD. Negative correlation between grey matter in the hippocampus and caudate nucleus in healthy aging. Hippocampus. 2020;30:892–908.
- 43. Lithfous S, Dufour A, Després O. Spatial navigation in normal aging and the prodromal stage of Alzheimer's disease: insights from imaging and behavioral studies. Ageing Res Rev. 2013;12:201–13.
- Chen JJ, Rosas HD, Salat DH. Age-associated reductions in cerebral blood flow are independent from regional atrophy. Neuroimage. 2011;55:468–78.
- 45 Galiano A, Mengual E, García de Eulate R, Galdeano I, Vidorreta M, Recio M, et al. Coupling of cerebral blood flow and functional connectivity is decreased in healthy aging. Brain Imaging Behav. 2020;14:436–50.
- An Y, Varma VR, Varma S, Casanova R, Dammer E, Pletnikova O, et al. Evidence for brain glucose dysregulation in Alzheimer's disease. Alzheimer's Dementia. 2018;14:318–29.
- Chételat G, Arbizu J, Barthel H, Garibotto V, Law I, Morbelli S, et al. Amyloid-PET and (18)F-FDG-PET in the diagnostic investigation of Alzheimer's disease and other dementias. Lancet Neurol. 2020;19:951–62.

- 48. Sperling RA, Aisen PS, Beckett LA, Bennett DA, Craft S, Fagan AM, et al. Toward defining the preclinical stages of Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimer's Dementia. 2011;7:280–92.
- Scheltens P, De Strooper B, Kivipelto M, Holstege H, Chételat G, Teunissen CE, et al. Alzheimer's disease. Lancet. 2021;397(10284):1577–90.
- 50. Guo T, Landau SM, Jagust WJ. Detecting earlier stages of amyloid deposition using PET in cognitively normal elderly adults. Neurology. 2020;94:e1512–24.
- Vila-Castelar C, Tariot PN, Sink KM, Clayton D, Langbaum JB, Thomas RG, et al. Sex differences in cognitive resilience in preclinical autosomal-dominant Alzheimer's disease carriers and non-carriers: Baseline findings from the API ADAD Colombia Trial. Alzheimer's Dement. 2022;10.1002/alz.12552.
- 52. Buckley RF, Mormino EC, Rabin JS, Hohman TJ, Landau S, Hanseeuw BJ, et al. Sex Differences in the Association

of Global Amyloid and Regional Tau Deposition Measured by Positron Emission Tomography in Clinically Normal Older Adults. JAMA Neurol. 2019;76:542–51.

- Hohman TJ, Dumitrescu L, Barnes LL, Thambisetty M, Beecham G, Kunkle B, et al. Sex-Specific Association of Apolipoprotein E With Cerebrospinal Fluid Levels of Tau. JAMA Neurol. 2018;75:989–98.
- Oveisgharan S, Arvanitakis Z, Yu L, Farfel J, Schneider JA, Bennett DA. Sex differences in Alzheimer's disease and common neuropathologies of aging. Acta Neuropathol. 2018;136:887–900.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.