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## **REVIEW**

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# **Perivascular macrophages in the CNS: From health to neurovascular diseases**

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## **Abstract**

Brain perivascular macrophages (PVMs) are attracting increasing attention as this emerging cell population in the brain has multifaced roles in supporting the central nervous system structure, brain development, and maintaining physiological functions. They also widely participate in neurological diseases such as neurodegeneration and ischemic stroke. Moreover, PVMs have been reported to have both beneficial and detrimental effects under different pathological contexts. Advanced research technologies allowed the further in-depth study of PVMs and revealed novel concepts in their origins, differentiation, and regulatory mechanisms. Deepened understanding of the roles of PVMs in different brain pathological conditions can reveal novel phenotypic changes and regulatory signaling, which might pave the way for the development of novel treatment strategies targeting PVMs.

#### **KEYWORDS**

Alzheimer's disease, neurological disease, perivascular macrophages, stroke

Li Zheng, Yunlu Guo, and Xiaozhu Zhai contributed equally to this work.

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## **1**  | **INTRODUCTION**

Innate immune cells in the brain are increasingly recognized as an important player in maintaining brain homeostasis and the development of brain diseases.<sup>1–4</sup> In addition to the widely studied microglia in the brain parenchyma, non-parenchymal border-associated macrophages (BAMs) such as PVMs, are one type of innate immune cells in the brain that have also been shown to participate in brain development, maintenance of homeostasis, neurodegenerative diseases, ischemic stroke, and other processes.<sup>[5](#page-9-1)</sup> Understanding the functions of PVMs in cerebral steady-state and disease progression can provide important insights for the development of treatment strategies for neurological diseases in the future. For this purpose, we summarize the latest research on PVMs in recent years, including the effects and modulatory mechanisms in neurodegeneration diseases, which are the novelties of this review.

## **2**  | **DISCOVERY OF PVMs IN THE BRAIN**

PVMs were first discovered in the 1980s by Mato et al<sup>[6](#page-9-2)</sup> using trypan blue and horseradish peroxidase injection into the ventricles, which were taken up by slender cells located in the perivascular space. They did a lot of research over the following several decades and found fluorescent granular perithelial (FGP) cells that could remove the metabolic waste of brain parenchyma with globular vacuolated inclusions in their cytoplasm,<sup>[7](#page-9-3)</sup> and incorpo-rate lipids in circulation.<sup>[8](#page-9-4)</sup> Notably, FGP cells were distributed in the space around cerebral arterioles and venules, which was different from the pericytes embedded in the basement membrane of capillaries.<sup>[9](#page-9-5)</sup> At the same time, in 1988, Hickey et al also described the slender and glycoprotein ED2 positive "perivascular microglia" around the blood vessels. $10$  As microglia do not express ED2 (CD163), $^{11}$  $^{11}$  $^{11}$  these perivascular cells were confirmed to be different from microglia. Thus, scientists then gradually recognized that PVMs are unique myeloid cells located in the brain perivascular Virchow-Robin space (VRS).

It is now well-accepted that BAMs are non-parenchymal macrophages in the central nervous system (CNS) and located in the boundary regions including VRS, meninges, and choroid plexus.[12](#page-9-8) As the name suggests, PVMs are macrophages located in the perivascular VRS of the CNS. Specifically, the VRS refers to invaginations surrounding cerebral vessels, and distinct interfaces connecting blood, cerebrospinal fluid (CSF), and brain parenchyma.<sup>13,14</sup> PVMs exactly reside around arterioles and venules both in cortical and subcortical regions of the mouse brain.<sup>15</sup> This special anatomical location of PVMs allows their direct contact with blood vessels and parenchyma, providing structural and functional support for the blood-brain barrier (BBB).<sup>16</sup> PVMs are also essential in maintaining brain homeostasis. In recent years, increasing evidence support that PVMs are key components of the brain resident immune system and are involved in number of pathological processes, especially in neurodegeneration and ischemic stroke.[7,8,17–24](#page-9-3)

# **3**  | **DEVELOPMENT AND DIFFERENTIATION OF PVMs**

## **3.1**  | **New views on the origin of PVMs**

The origin of PVMs has been discussed for a long time. $25-27$  For decades, it was thought that all PVMs came from circulating monocytes and were updated frequently.<sup>10,28-30</sup> However, this conclusion was questioned later, because it was based on the experiments of fullbody irradiation and bone marrow transplantation in rodents. Both of the experiments may cause the overexpression of chemokines and the destruction of the BBB, resulting in the entry of the bone marrow-derived monocytes into the CNS.

New views on the origin and renewal of PVMs have been put forward through large-scale single-cell RNA-sequencing (scRNA-seq), parabiosis, fate-mapping, and in vivo imaging. In general, PVMs and microglia are both of prenatal origin and PVMs have a closer transcriptional relationship with microglia than monocytes. It is now well accepted that PVMs arise from early erythro-myeloid progenitors in the yolk sac, which migrate into the brain in the early stage of the embryo. $31$  Using scRNAseq, two phenotypically and transcriptionally distinct macrophages, which separately differentiate into microglia and PVMs, can be found and distinguished in the developing brain and the yolk sac, indicating an early separation into two different populations. In addition, the development of PVMs is inde-pendent of TGF-β, while microglia need TGF-β for development.<sup>[32](#page-10-1)</sup> In normal conditions, PVMs are a stable cell population with a long lifespan and self-renewal ability after birth. When the PVMs are depleted by laboratory methods, they can be replenished from bone marrow-derived monocytes.

## **3.2**  | **Regulatory mechanisms of the differentiation of PVMs**

The development and normal functions of PVMs are regulated by many factors, including transcription factors and cytokines. We summarize several key influencing factors modulating PVMs.

#### 3.2.1 | Transcriptional regulation

In recent years, transcription factors have been shown as the key determinants in the orchestration of myeloid identity and differentiation fates.

PU.1 is a member of the large family of E-twenty six transcription factors and it is the product of the oncogene Spi1. PU.1 exists in almost all myeloid-specific and many lymphoid-specific gene regulatory sequences, and most PVM-specific enhancers contain binding domains for PU.1.<sup>[33](#page-10-2)</sup> The absence of Spi1 in mice can lead to fatal defects in fetal liver and/or newborn hematopoiesis, including the complete loss of macrophages.  $34,35$  The more recent analysis has demonstrated that the absence of PU.1 impairs the repopulation

capacity of the hematopoietic stem cells (HSCs), impeding their differentiation into the common myeloid progenitors and the common lymphoid progenitors.[36–38](#page-10-4)

The survival of brain PVMs also depends on transcription factor c-MAF, which is part of the large Maf family of transcription factors. Conditional knockout of c-MAF in macrophage lineages will cause ablation of PVMs in the CNS.<sup>[39](#page-10-5)</sup>

MAFB also belongs to the Maf family, with the function of controlling the proliferation rate of PVMs through the epigenetic regulation of self-renewing genes.<sup>40,41</sup> Beyond that, MAFB is also able to limit the ability of macrophage colony-stimulating factor (M-CSF) in differentiating HSCs to PVMs.<sup>[39,42](#page-10-5)</sup>

Interferon regulatory factor 8 (IRF8) is critically involved in driving the maturation and diversity of brain macrophages. The deficiency of IRF8 will cause alternations in PVMs development and function.[43,44](#page-10-7)

## 3.2.2 | The cytokines that regulate myeloid cell fate

M-CSF, also known as CSF-1 is the major cytokine modulating mac-rophages' proliferation, differentiation, and functional regulation.<sup>[45](#page-10-8)</sup> M-CSF is produced by a variety of stromal and epithelial cells. It transmits signals through M-CSF receptors (M-CSFR/CSF-1R/ CD115). It has been shown that M-CSF can guide the myeloid fate of HSCs by inducing PU.1, $46$  and it is important in establishing and maintaining tissue-resident macrophage pools.<sup>[47](#page-10-10)</sup> In addition to influencing the differentiation and maintenance of macrophages, M-CSF can also stimulate the survival and self-renewal of macrophages in steady-state and inflammation. Moreover, it is involved in the polari-zation of macrophage activation.<sup>[48,49](#page-10-11)</sup>

Interferon-gamma (IFN-γ) can interact with PVMs by upregulating major histocompatibility complex (MHCII) and B7 coreceptor expression, and shift PVMs from anti-inflammatory to proinflamma-tory cytokine profiles.<sup>[50](#page-10-12)</sup>

The above evidence suggests that  $IFN-\gamma$  has a direct effect on the phenotypic switch of PVMs, while M-CSF could also be an important determinant cytokine of PVM cell fate and phenotypic polarization. However, further studies are warranted to identify the roles of different cytokines in the regulation of PVMs.

## **4**  | **CHARACTERIZATION AND RECOGNITION OF PVMS IN THE BRAIN**

PVMs are characterized by their anatomical localization, phagocytosis ability, and molecular markers. Firstly, PVMs can be recognized by their specific locations. Nowadays, people believe that PVMs are located in the VRS between the vascular basement membrane on the abluminal side and the glial limitans of the brain parenchyma. $15$ And they belong to a group of distinct myeloid cells.

Secondly, the phagocytosis function of PVMs can be utilized for their identification. For example, by intravenous injection of

fluorescence-labeled dextran, PVMs can be clearly visualized due to their phagocytosis of the fluorescent dextran.<sup>16,51</sup> Interestingly, their phagocytosis ability can also be utilized to achieve PVMs depletion by intraventricular injection of clodronate (CLO)-containing liposomes.<sup>16,17,52,53</sup>

Importantly, PVMs express a number of markers that can be distinguished from microglia (Table [1](#page-3-0), Figure [1](#page-3-1)). Besides CD45 and CD11b, the brain-resident myeloid cells all express fractalkine receptor (Cx3cr1), CSF1R, and allograft inflammatory factor 1 (Iba-1).<sup>[5](#page-9-1)</sup> PVMs have higher expression levels of CD45, F4/80, Iba-1, and MHCII, $5.54$  and lower levels of Cx3cr1 compared to microglia.<sup>55</sup> PVMs are negative for microglial-specific markers such as transmembrane protein 119 (TMEM119), sialic acid-binding immunoglobulin-like lectin H (Siglec-H), P2Y purinoceptor 12 (P2RY12), Sal-like protein 1 (Sall1), Sal-like protein3 (Sall3), or ANXA3.[5,56–59](#page-9-1) Instead, brain PVMs express nonconventional macrophage markers such as Siglec1 (CD169), which is absent in microglia.<sup>[60](#page-10-14)</sup> These features can be used to distinguish microglia and PVMs.

Some conventional PVMs markers could differentiate PVMs with microglia, such as CD163, lymphatic vessel endothelial hyaluronan receptor-1 (Lyve-1), and CD206. CD163 is expressed in PVM and monocytes, but not in microglia. $61,62$  The mannose receptor CD206 is only expressed in BAMs, which can be used to distinguish PVMs from microglia and monocytes. However, transient high expression of CD206 can be found in a subpopulation of microglia and infiltrating macrophages after a brain injury such as stroke and brain trauma.<sup>[63,64](#page-10-16)</sup> The monoclonal antibody 5D3 can be used to localize the expression of mannose receptors on PVMs in normal CNS and various models of brain pathology with good specificity.<sup>[65](#page-10-17)</sup> Based on these different markers, a binary transgenic model has recently been used to dissect microglia and PVMs for further separate study.<sup>[55](#page-10-13)</sup>

In summary, so far there is no single marker of PVMs with good specificity and sensitivity. The expressions of PVM markers combined with their anatomical location and phagocytic feature may be an ideal and reliable way for identification.

## **5**  | **PVM, THE FIRST LINE "FIREWALL" MAINTAINING THE HOMEOSTASIS OF THE CNS**

Under physiological conditions, there are different types of macrophages in the CNS, performing their functions and maintaining the homeostasis of the brain. As an important part of BAMs, PVMs act as the first "firewall" in the CNS because of their special anatomical location and innate immune functions. Paragraphs outlined below discussed PVM functions in the regulation of BBB permeability, immune regulation, phagocytosis, and lymphatic clearance.

#### **5.1**  | **PVMs regulate the integrity of BBB**

The integrity of BBB is essential for the brain to maintain its homeostasis and is an important anatomical structure that  ZHENG et al. **<sup>|</sup> 1911**

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<span id="page-3-0"></span>**TABLE 1** Differentiation markers of PVM and microglia





<span id="page-3-1"></span>**FIGURE 1** Characteristic markers of PVMs in the brain. All leukocytes express CD45 and CD11b, and besides, the brain-resident myeloid cells express Cx3cr1, CSF1R, and Iba-1. Conventional PVMs markers include CD163, Lyve-1, and CD206. Compared to microglia, PVMs have higher expression levels of CD45, F4/80, Iba-1, and MHCII. A variety of markers have been confirmed to be related to the phagocytosis of PVMs such as CD36, CD68, CD206, CCR2, and IDO-1. And some markers have been confirmed to participate in the antigen presentation such as CD163, CD45, CD11b, MCHII, and F4/80. LYVE-1 is a receptor of hyaluronan, controlling the expression of collagen in vascular smooth muscle cells.

mediates the entry of the essential components into the brain parenchyma and also prevents the invasion of pathogens and blood-derived harmful substances.<sup>79-82</sup> It is well known that brain capillary endothelial cells and their tight junctions play key roles in maintaining the BBB permeability.<sup>83-85</sup> Meanwhile, the participation of PVMs has been recognized recently.<sup>86-88</sup> Under physiological conditions, the microvasculature of the area postrema (AP) has a less restrictive BBB than is found in other

CNS areas due to the lack of tight junction.[20](#page-9-13) In this case, PVMs in this area can isolate 10–70 kDa serum proteins from the blood and combine the laminin layer, further helping to restrict the entry of solutes above 10 kDa into the parenchyma.<sup>86</sup> In addition, using the cell culture model of the BBB, Zenker et al<sup>[89](#page-11-3)</sup> found that the transendothelial electrical resistance of post-confluent brain capillary endothelial cells was significantly increased by coculture with blood-derived macrophages, which could partly

indicate that PVMs can be involved in the maintenance of BBB permeability.

The effect of PVMs on BBB seems to be two-edged. In normal condition, they are necessary for the maintenance of BBB, but in the case of CNS injury and neuroinflammation, they seem to mediate the damage of BBB. Most recently, PVMs have been confirmed to participate in BBB disruption through the release of cytotoxic me-diators under malaria.<sup>[90](#page-11-7)</sup> What's more, another review analyzed literature from 2000 to 2021 and revealed that PVMs could cause BBB damage in Alzheimer's disease (AD).<sup>[91](#page-11-8)</sup>

Taken together, the emerging literature suggests that PVMs modulate the integrity of BBB. Future research should investigate the specific regulatory mechanisms in this process.

## **5.2**  | **Immune regulation and antigen presentation of PVMs**

In a steady-state, there are few circulating immune cells in the brain, but a variety of immune cell subtypes can infiltrate into CNS in the case of inflammation, trauma, autoimmune diseases, and so on. Classical immunostimulation with lipoteichoic acid from grampositive bacteria and lipopolysaccharide (LPS) from gram-negative bacteria can lead to the proliferation of PVMs. PVMs show phenotypic plasticity in many homeostatic and pathological situations. PVMs have been confirmed that they can exert proinflammatory polarization (M1) in neurological diseases including ischemic injury in mice.<sup>15</sup> We have reason to believe that PVMs may also have different types of classification as a kind of macrophages. However, PVMs activation status may also not be simply classified into M1/ M2 due to their complex nature. Thus, further research is warranted to identify different phenotypes of PVMs.

Previous studies have confirmed that PVMs can act as antigenpresenting cells (APCs) under certain pathological circumstances,<sup>21-24</sup> they are essential in the CNS as APCs both in vitro and in vivo. $8,33,92$  One of the important characteristics of APCs is the ex-pression of MHCII.<sup>[93](#page-11-9)</sup> PVMs express MHCII and can present antigen to lymphocytes in an experimental autoimmune encephalomyelitis (EAE) model.<sup>[10](#page-9-6)</sup> MHCII<sup>high</sup> PVMs can be observed under pathological conditions, such as transient middle cerebral artery occlusion (tMCAo), EAE in rodents, and multiple sclerosis (MS) from the human autopsy.<sup>[10,62,94](#page-9-6)</sup> A recent study also showed it was PVMs that presented antigens to  $CD8^+$  T cells in experimental cerebral malaria.<sup>[90](#page-11-7)</sup> Thus, PVMs also have the function of bridging innate and adaptive immunity in the CNS.

## **5.3**  | **The phagocytic ability of PVMs**

Under physiological conditions, PVMs are considered to be the scavenger cells and surveillant cells of the brain, as they occupy the ideal position of monitoring and removing potentially harmful substances. As mentioned before, PVMs were first discovered due to their uptake of dyes.<sup>[6](#page-9-2)</sup> Researchers also discovered that this kind of cells can remove metabolic waste from brain parenchyma, and bind to lipids in the circulation to clear the lipid deposition increased in aging animals.<sup>7,8</sup> PVMs have also been shown to gobble up metabolic waste and cellular breakdown products in some brain diseases such as experimental subarachnoid hemorrhage and cerebral amyloid angiopathy[.17,95](#page-9-15)

The phagocytosis property of PVMs has been proved relevant to the markers expressed on PVMs such as CD68, CD163, CD206, and IDO-1. CD68 is a lysosomal protein that promotes intracellular lysosomal function. $^{73}$  $^{73}$  $^{73}$  CD163 is a scavenger receptor (SR) protein that recognizes and endocytoses the hemoglobin/haptoglobin complexes and participates in antigen presentation.<sup>74</sup> CD206 is a mannose receptor that may be involved in the scavenging effect.[65,96](#page-10-17) IDO-1 is an immunosuppressive enzyme that increases cellular phagocytic capacity and might suppress the overactivation of inflammatory response.<sup>97</sup>

In summary, the phagocytic ability of PVMs under physiological and also pathological conditions is of vital importance for maintaining brain homeostasis by clearing exogenous substances, endogenous metabolic waste, and cellular debris.

#### **5.4**  | **Lymphatic clearance of PVMs**

The brain lymphatic system has momentous physiological functions: excreting interstitial fluid (ISF) to the nearby lymph nodes from the brain parenchyma, maintaining water and electrolyte balance of the ISF, clearing metabolic waste, and reabsorbing macromolecular solutes $98,99$ ; and communicating with the immune system, modulating immune surveillance, and the inflammatory response. The cerebral lymphatic drainage system is composed of a basement membrane-based perivascular pathway,<sup>[100](#page-11-13)</sup> a brain-wide paravascular glymphatic pathway, $19$  and some CSF drainage routes including sinus-associated meningeal lymphatic vessels<sup>18,101-103</sup> and olfactory/cervical lymphatic routes.<sup>104,105</sup> Given their close relationship to vessels, PVMs may facilitate the first two pathways.

The "intramural perivascular drainage pathway" (IPAD) is a pathway in the vessel wall of the tunica media which is composed of vascular smooth muscle cells (VSMCs)[.100,106](#page-11-13) Injection tracers into the caudate putamen were found to enter the arterial wall and travel along the intercellular spaces among the VSMCs. PVMs can promote the clearance of the IPAD by taking up 2 nm to 1  $\mu$ m particles. Furthermore, PVMs mediate the speed of IPAD by regulating the contraction and relaxation of VSMCs, and it is found that the increase in age will lead to a significant slowdown of IPAD. $107$  The glymphatic pathway also involves "paravascular space."<sup>[19](#page-9-16)</sup> CSF enters the parenchyma along paravascular spaces which surround penetrating arteries and the brain ISF is cleared via paravenous drainage pathways.

PVMs can facilitate lymphatic drainage in the CNS in the above two ways mentioned above. The exact role of PVMs in these pathways is worth further study and discussion.

In summary, because of the special anatomical location in the brain, PVMs can directly contact blood, CSF, and brain parenchyma. PVMs exert phagocytic function and clear metabolic waste. PVMs can also act as APCs to recruit circulating immune cells into CNS. In addition, they provide structural and functional support for BBB and lymphatic clearance, which is important for the maintenance of brain homeostasis and normal functions.

## **6**  | **PVMs IN NEUROLOGICAL DISEASES**

As mentioned above, PVMs are vitally important in maintaining brain homeostasis. In the past few years, more and more evidence supported the theory that PVMs are widely involved in neurological diseases (Table [2](#page-6-0)). Here, we mainly focused on AD, hypertensionassociated neurovascular dysfunction, and stroke.

#### **6.1**  | **PVMs and Alzheimer's disease**

AD is the main cause of cognitive impairment in the elderly, pathologically characterized by extracellular deposition of the amyloid-β (Aβ) and intracellular aggregates of the microtubule-associated protein tau (neurofibrillary tangles).

The brain is highly dependent on the continuous regulation of cerebral blood flow (CBF) to transport oxygen and glucose for the brain's energy needs. Not surprisingly, alternations in cerebral perfusion can cause brain dysfunction and cognitive impairment. A large number of studies have shown that Aβ disrupted cerebral microcirculation. Aβ inhibits the increase of CBF correlated to synaptic activity and interferes with endothelial function.<sup>113-120</sup> The brain Aβ is released to extracellular space during synaptic ac-tivity<sup>[121](#page-12-1)</sup> and reaches the VRS.<sup>[122](#page-12-2)</sup> In this space, PVMs are in direct contact with Aβ, which mediates special pathophysiological processes.

Previous studies have suggested that besides microglia, the phagocytosis of PVMs is essential for Aβ clearance. Depletion of PVMs is related to the vascular accumulation of Aβ 42 and the severity of cerebral amyloid angiopathy.<sup>17</sup> Scavenger receptors (SRs) are widely expressed by microglia/macrophages and are able to bind a diverse array of endogenous and foreign molecules, thus playing critical roles in the phagocytosis of these cells. The phagocytic function of PVMs was found to be regulated by the high-density lipoprotein receptor (SR class B type 1, SR-B1) on PVMs that regulates the flow of cholesterol. The exhaustion of SR-B1 can impair PVMs response to Aβ and accelerate the formation of cerebrovascular and also parenchymal amyloid plaques in the cerebral cortex and hippo-campus of mice, thus aggravating cognitive impairment.<sup>[53](#page-10-26)</sup>

Nevertheless, in addition to the above beneficial effects, PVMs also take part in the negative side of AD development. PVMs are

involved in Aβ-induced neurovascular dysfunction through CD36 mediated oxidative stress. CD36 binds Aβ and leads to NADPH oxidase 2 (Nox2)-dependent production of reactive oxygen species  $(ROS).<sup>122,123</sup>$  $(ROS).<sup>122,123</sup>$  $(ROS).<sup>122,123</sup>$  Selective depletion of PVMs can abrogate the neurovascular dysfunction and vascular oxidative stress induced by  $AB^{16}$  $AB^{16}$  $AB^{16}$ (Figure [2](#page-7-0)).

In addition, aging reduces the activity of PVMs and causes cell dysfunction, along with the alteration of the structure and distribution of PVMs. Mato et al found the amount of lipid precipitation in the cytoplasm increased significantly with age.<sup>7</sup> In PVMs of young subjects, most inclusion bodies are round, uniform in content, and high in electron density. However, in elderly subjects, PVMs show many enlarged inclusion bodies and often display a honeycomb structure.<sup>7</sup> At the same time, in both elderly experimental animals and humans, the swollen PVMs often appear at bifurcations of cerebral arterioles and compress arterioles, which contribute to the disturbance of cerebral blood flow.<sup>[7](#page-9-3)</sup>

To sum up, the bi-directional regulation of Aβ by PVMs reminds us not to simply block or boost PVMs in AD. Instead, it is meaningful for the future study on the regulation of PVMs to take advantage and avoid the reverse effects of AD.

## **6.2**  | **PVMs and hypertension-associated neurovascular dysfunction**

The health of the cerebrovascular system is of vital importance to the functional and structural integrity of the brain.<sup>51</sup> Remarkably, hypertension can disrupt the cerebrovascular system, which is the basis of neurovascular cognitive impairment.<sup>124-127</sup> Recent studies suggested that PVMs take part in the modulation of neurovascular and cognitive dysfunction associated with hypertension.

PVMs mediate cerebral neurovascular dysfunction in hyper-tension through the angiotensin type 1 receptor (Atr1).<sup>[51](#page-10-20)</sup> In hypertension, the elevated Ang II can reach the perivascular space through the damaged BBB, then activate Atr1 on PVMs, resulting in NOX2-dependent ROS production, finally leading to cerebral vascular dysfunction and cognitive dysfunction.<sup>51</sup> Another study demonstrated that by depleting most of the PVMs and all the microglia in Ang II-induced hypertensive mouse model, short-term memory impairment can be prevented.<sup>[128](#page-12-4)</sup> What's more, PVMs contribute to the development of hypertension, both the number and activity of the PVMs are increased by the stimulation of proinflammatory cytokine.<sup>112,129</sup> Then, prostaglandin E2 (PGE2) produced by PVMs enters the brain parenchyma, resulting in sympathetic activation and blood pressure elevation.<sup>[112](#page-12-5)</sup> Interestingly, in vitro study, it is confirmed that extracellular PGE can promote microglia to produce more PGE and COX-2.[130](#page-12-6) All of the above processes will play momentous roles in the development of CNS diseases. Based on these studies, we speculate whether PVMs and microglia have a synergistic effect in Ang II-mediated hypertensive cerebrovascular disease worth deeper study.

<span id="page-6-0"></span>

**FIGURE 2** The role of PVMs in AD PVMs has both positive and negative effects on the development of AD. AD is characterized pathologically by extracellular deposition of the amyloid-β peptide (Aβ) in amyloid plaques and intracellular aggregates of the microbubble-associated protein tau. A-β is released to extracellular in the process of synaptic activity and reaches the VRS, where the phagocytosis function of PVMs is essential for A-β clearance. Besides, PVMs are involved in A-β-induced neurovascular dysfunction through CD36 mediated oxidative stress. CD36 binds A-β and leads to NADPH oxidase 2 (Nox2) dependent production of reactive oxygen species (ROS).

<span id="page-7-1"></span>**FIGURE 3** The role of PVMs in hypertension-associated cognitive impairment. PVMs can mediate hypertension-associated neurovascular and cognitive dysfunction in many ways. Expressing Col1a1MRNA, PVMs mediate the production of type I collagen. Collagen deposits around the cerebral small vessels induces the change of atherosclerosis during hypertension. Circulating AngII can activate Atr1 on PVMs, resulting in NOX2 dependent ROS production, which then leads to cerebral vascular dysfunction and cognitive impairment. PVMs produce PGE2 through COX-2, PGE 2 enters the brain parenchyma and activates PGE2 receptors on the PVN and the rostral ventrolateral medulla (RVLM), leading to sympathetic activation and an increase in blood pressure.

<span id="page-7-0"></span>

hypertension associated cognitive impairment

PVMs are involved in the process of cerebrovascular remodeling in hypertension as well. The remodeling and progression of atherosclerosis in hypertension contain fibrosis and the production of type I collagen around the cerebral arterioles.<sup>[111](#page-12-9)</sup> PVMs around the cerebral small vessels express Col1a1 mRNA, which mediates the production of type I collagen, makes collagen deposition around the cerebral small vessels, and participates in the change of atherosclerosis during hypertension $111$  (Figure [3](#page-7-1)).

It can be seen that PVMs participate in neurovascular and cognitive dysfunction related to hypertension from many aspects. Hence, a deeper understanding of how PVMs influence the remodeling of the cerebrovascular system may help to optimize the therapies for the recovery and rehabilitation of related diseases.

## **6.3**  | **Stroke and PVMs**

Stroke remains the second leading cause of death and the third leading cause of death and disability worldwide in 2019.<sup>[131](#page-12-10)</sup> Therefore, it is important and necessary to clarify the mechanisms of the stroke to lighten the burden on families and even the whole world. It is increasingly recognized that PVMs play an important role in the acute inflammatory phase and secondary injury after stroke.

According to statistics, ischemic stroke constituted 62.4% of all strokes.<sup>[131](#page-12-10)</sup> Researchers have confirmed the functions of PVMs in ischemic stroke in many ways. For instance, PVMs participate in the neuropathological process through cell proliferation and migration to the ischemic brain parenchyma, as PVMs are found highly



<span id="page-8-0"></span>**FIGURE 4** The main modulatory mechanisms of PVMs after stroke onset. After the stroke, PVMs proliferate and migrate to the lesion core, up-regulating the expression of many inflammatory genes including COX-1, VEGF, leukocyte chemo-attractants, and promoting lymphocytes enter into the CNS. Although PVMs may still show harmful effects in the long run, they are the main cells responsible for phagocyting erythrocytes and their decomposition products in the early stage.

accumulated in peri-infarction areas and in the developing necrotic core area in the early stage after cerebral infarction, and the number continues to increase until several months after stroke.<sup>[109](#page-12-7)</sup> More than that, PVMs also up-regulate the expression of COX-1, which plays an important role in the pathophysiology of acute ischemic inflammation, tissue remodeling, and secondary injury after stroke. Aside from acting as direct proinflammatory cells, PVMs can also participate in granulocyte recruitment by upregulating the expression of leukocyte chemo-attractants.<sup>[110](#page-12-8)</sup> Moreover, PVMs have the function to elevate the expression of VEGF, increasing the permeability of pial and cortical blood vessels, and deteriorating neurological im-pairment in the acute phase of stroke.<sup>[110](#page-12-8)</sup> These results were also found in postmortem brain samples from ischemic stroke patients.<sup>[78](#page-11-6)</sup>

A current study revealed that heavy drinking of alcohol  $(>= 6$ standard drinks/day) is an independent risk factor associated with worse outcomes in ischemic stroke patients.<sup>[108](#page-11-16)</sup> PVMs are activated in mice with chronic alcohol exposure, and the inflammation significantly increased after a secondary insult (ischemic stroke or LPS challenge). Depletion of PVMs can block the alcohol-induced aggravation of ischemic lesions.

Subarachnoid hemorrhage (SAH) is a subtype of stroke and constituted 9.7% (about 1.18 million) of all strokes. $131$  As erythrocytes are damaged, there are many decomposition products such as bilirubin, heme, and free iron released into the CSF, which can cause inflammation, vasoconstriction, and direct cellular injury.<sup>132-135</sup> After SAH, erythrocytes enter the perivascular space, where they can interact with PVMs. Recent studies have found that erythrocytes are mainly removed by PVMs rather than microglia; however, the depletion of PVMs with CLO can decrease inflammation around arterioles and improve prognosis after SAH (Figure [4](#page-8-0)). This contradictory result is probably due to the reduced inflammatory burden after PVM depletion counteracting the negative effect of increased breakdown waste. It can be seen that although PVMs play a role in the phagocytosis of damaged erythrocytes and their decomposition products, PVMs may still show harmful effects in the long run.

To sum up, as a part of the brain innate immune cell population, PVMs are the important guardians of CNS homeostasis. Given the functional similarity of PVMs and microglia, whether PVMs display sex difference as in microglia is not clear.<sup>136,137</sup> The current literature is lacking on this issue. A deep understanding of how PVMs participate in the pathological mechanism of CNS diseases will be helpful to the development of treatment strategies.

## **7**  | **CURRENT KNOWLEDGE GAPS AND FUTURE PERSPECTIVES**

Growing evidence is suggesting the critical role of PVMs in maintaining brain hemostasis and regulating the progression of various neurological diseases. However, there are still a lot of unknowns and obstacles in the PVM research field. Here, we outline some of the key issues that need to be resolved.

- 1. The key regulatory genes and the underlying regulatory mechanisms of PVMs' differentiation, phenotypic switch, and cell fate under different disease contexts remain largely unknown.
- 2. The research findings from animal models are not able to fully reflect the changes of PVMs in the human body. The imaging techniques of PVMs in human is still underdeveloped.
- 3. Currently, there is no specific PVM targeted strategy that could allow precise manipulation of PVMs. Delivery techniques targeting PVMs are highly warranted for future PVM-associated treatments.

Collectively, as important brain innate immune cells, the role of PVMs in the brain is emerging. Further research is warranted to expand the knowledge of the regulatory mechanisms of PVMs and the role of PVMs in various brain pathologies.

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#### **CONFLICT OF INTEREST**

The author(s) declared no potential conflicts of interest with respect to the authorship, and/or publication of this article. Peiying Li is an Editorial Board member of CNS Neuroscience and Therapeutics and a co-author of this article. To minimize bias, they were excluded from all editorial decision-making related to the acceptance of this article for publication.

#### **DATA AVAILABILITY STATEMENT**

Data sharing is not applicable to this article as no new data were created or analyzed in this study.

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#### **REFERENCES**

- <span id="page-9-0"></span>1. Wang X, Xuan W, Zhu ZY, et al. The evolving role of neuro-immune interaction in brain repair after cerebral ischemic stroke. *CNS Neurosci Ther*. 2018;24(12):1100-1114.
- 2. Prinz M, Erny D, Hagemeyer N. Ontogeny and homeostasis of CNS myeloid cells. *Nat Immunol*. 2017;18(4):385-392.
- 3. Engelhardt B, Vajkoczy P, Weller RO. The movers and shapers in immune privilege of the CNS. *Nat Immunol*. 2017;18(2):123-131.
- 4. Herz J, Filiano AJ, Smith A, Yogev N, Kipnis J. Myeloid cells in the central nervous system. *Immunity*. 2017;46(6):943-956.
- <span id="page-9-1"></span>5. Goldmann T, Wieghofer P, Jordão MJC, et al. Origin, fate and dynamics of macrophages at central nervous system interfaces. *Nat Immunol*. 2016;17(7):797-805.
- <span id="page-9-2"></span>6. Mato M, Ookawara S, Kurihara K. Uptake of exogenous substances and marked infoldings of the fluorescent granular pericyte in cerebral fine vessels. *Am J Anat*. 1980;157(3):329-332.
- <span id="page-9-3"></span>7. Mato M, Aikawa E, Mato TK, Kurihara K. Tridimensional observation of fluorescent granular perithelial (FGP) cells in rat cerebral blood vessels. *Anat Rec*. 1986;215(4):413-419.
- <span id="page-9-4"></span>8. Mato M, Ookawara S, Sakamoto A, et al. Involvement of specific macrophage-lineage cells surrounding arterioles in barrier and scavenger function in brain cortex. *Proc Natl Acad Sci USA*. 1996;93(8):3269-3274.
- <span id="page-9-5"></span>9. Ookawara S, Mitsuhashi U, Suminaga Y, Mato M. Study on distribution of pericyte and fluorescent granular perithelial (FGP) cell in the transitional region between arteriole and capillary in rat cerebral cortex. *Anat Rec*. 1996;244(2):257-264.
- <span id="page-9-6"></span>10. Hickey WF, Kimura H. Perivascular microglial cells of the CNS are bone marrow-derived and present antigen in vivo. *Science*. 1988;239(4837):290-292.
- <span id="page-9-7"></span>11. Graeber MB, Streit WJ, Kreutzberg GW. Identity of ED2-positive perivascular cells in rat brain. *J Neurosci Res*. 1989;22(1):103-106.
- <span id="page-9-8"></span>12. Koizumi T, Kerkhofs D, Mizuno T, Steinbusch HWM, Foulquier S. Vessel-associated immune cells in cerebrovascular diseases: from perivascular macrophages to vessel-associated microglia. *Front Neurosci*. 2019;13:1291.
- <span id="page-9-9"></span>13. Esiri MM, Gay D. Immunological and neuropathological significance of the Virchow-Robin space. *J Neurol Sci*. 1990;100(1–2):3-8.
- 14. Hutchings M, Weller RO. Anatomical relationships of the pia mater to cerebral blood vessels in man. *J Neurosurg*. 1986;65(3):316-325.
- <span id="page-9-10"></span>15. Yang T, Guo R, Zhang F. Brain perivascular macrophages: recent advances and implications in health and diseases. *CNS Neurosci Ther*. 2019;25(12):1318-1328.
- <span id="page-9-11"></span>16. Park L, Uekawa K, Garcia-Bonilla L, et al. Brain perivascular macrophages initiate the neurovascular dysfunction of Alzheimer Aβ peptides. *Circ Res*. 2017;121(3):258-269.
- <span id="page-9-15"></span>17. Hawkes CA, McLaurin J. Selective targeting of perivascular macrophages for clearance of beta-amyloid in cerebral amyloid angiopathy. *Proc Natl Acad Sci USA*. 2009;106(4):1261-1266.
- <span id="page-9-17"></span>18. Sun BL, Wang LH, Yang T, et al. Lymphatic drainage system of the brain: a novel target for intervention of neurological diseases. *Prog Neurobiol*. 2018;163-164:118-143.
- <span id="page-9-16"></span>19. Iliff JJ, Wang M, Liao Y, et al. A paravascular pathway facilitates CSF flow through the brain parenchyma and the clearance of interstitial solutes, including amyloid beta. *Sci Transl Med*. 2012;4(147):147ra111.
- <span id="page-9-13"></span>20. Goehler LE, Erisir A, Gaykema RP. Neural-immune interface in the rat area postrema. *Neuroscience*. 2006;140(4):1415-1434.
- <span id="page-9-14"></span>21. Galea I, Bernardes-Silva M, Forse PA, van Rooijen N, Liblau RS, Perry VH. An antigen-specific pathway for CD8 T cells across the blood-brain barrier. *J Exp Med*. 2007;204(9):2023-2030.
- 22. McGavern DB, Kang SS. Illuminating viral infections in the nervous system. *Nat Rev Immunol*. 2011;11(5):318-329.
- 23. Sallusto F, Impellizzieri D, Basso C, et al. T-cell trafficking in the central nervous system. *Immunol Rev*. 2012;248(1):216-227.
- 24. Barkauskas DS, Evans TA, Myers J, Petrosiute A, Silver J, Huang AY. Extravascular CX3CR1+ cells extend intravascular dendritic processes into intact central nervous system vessel lumen. *Microsc Microanal*. 2013;19(4):778-790.
- <span id="page-9-12"></span>25. Kierdorf K, Erny D, Goldmann T, et al. Microglia emerge from erythromyeloid precursors via Pu.1- and Irf8-dependent pathways. *Nat Neurosci*. 2013;16(3):273-280.
- 26. Schulz C, Perdiguero EG, Chorro L, et al. A lineage of myeloid cells independent of Myb and hematopoietic stem cells. *Science*. 2012;336(6077):86-90.
- 27. Ginhoux F, Greter M, Leboeuf M, et al. Fate mapping analysis reveals that adult microglia derive from primitive macrophages. *Science*. 2010;330(6005):841-845.
- 28. Prinz M, Priller J. Microglia and brain macrophages in the molecular age: from origin to neuropsychiatric disease. *Nat Rev Neurosci*. 2014;15(5):300-312.
- 29. Hickey WF, Vass K, Lassmann H. Bone marrow-derived elements in the central nervous system: an immunohistochemical and ultrastructural survey of rat chimeras. *J Neuropathol Exp Neurol*. 1992;51(3):246-256.
- 30. Aguzzi A, Barres BA, Bennett ML. Microglia: scapegoat, saboteur, or something else? *Science*. 2013;339(6116):156-161.

- <span id="page-10-0"></span>31. Lee E, Eo JC, Lee C, Yu JW. Distinct features of brain-resident macrophages: microglia and non-parenchymal brain macrophages. *Mol Cells*. 2021;44(5):281-291.
- <span id="page-10-1"></span>32. Utz SG, See P, Mildenberger W, et al. Early fate defines microglia and non-parenchymal brain macrophage development. *Cell*. 2020;181(3):557-573 e18.
- <span id="page-10-2"></span>33. Tenen DG, Hromas R, Licht JD, Zhang DE. Transcription factors, normal myeloid development, and leukemia. *Blood*. 1997;90(2):489-519.
- <span id="page-10-3"></span>34. Waddell LA, Lefevre L, Bush SJ, et al. ADGRE1 (EMR1, F4/80) is a rapidly-evolving gene expressed in mammalian monocytemacrophages. *Front Immunol*. 2018;9:2246.
- <span id="page-10-21"></span>35. Buxade M, Huerga Encabo H, Riera-Borrull M, et al. Macrophagespecific MHCII expression is regulated by a remote Ciita enhancer controlled by NFAT5. *J Exp Med*. 2018;215(11):2901-2918.
- <span id="page-10-4"></span>36. Guiducci C, Vicari AP, Sangaletti S, Trinchieri G, Colombo MP. Redirecting in vivo elicited tumor infiltrating macrophages and dendritic cells towards tumor rejection. *Cancer Res*. 2005;65(8):3437-3446.
- 37. Scott EW, Fisher RC, Olson MC, Kehrli EW, Simon MC, Singh H. PU.1 functions in a cell-autonomous manner to control the differentiation of multipotential lymphoid-myeloid progenitors. *Immunity*. 1997;6(4):437-447.
- 38. Dakic A, Metcalf D, di Rago L, Mifsud S, Wu L, Nutt SL. PU.1 regulates the commitment of adult hematopoietic progenitors and restricts granulopoiesis. *J Exp Med*. 2005;201(9):1487-1502.
- <span id="page-10-5"></span>39. Moura Silva H, Kitoko JZ, Queiroz CP, et al. c-MAF-dependent perivascular macrophages regulate diet-induced metabolic syndrome. *Sci Immunol*. 2021;6(64):eabg7506.
- <span id="page-10-6"></span>40. Bottcher C, Schlickeiser S, Sneeboer MA, et al. Human microglia regional heterogeneity and phenotypes determined by multiplexed single-cell mass cytometry. *Nat Neurosci*. 2019;22(1):78-90.
- <span id="page-10-22"></span>41. Jordao MJC, Sankowski R, Brendecke SM, et al. Single-cell profiling identifies myeloid cell subsets with distinct fates during neuroinflammation. *Science*. 2019;363(6425):eaat7554.
- 42. Sarrazin S, Mossadegh-Keller N, Fukao T, et al. MafB restricts M-CSF-dependent myeloid commitment divisions of hematopoietic stem cells. *Cell*. 2009;138(2):300-313.
- <span id="page-10-7"></span>43. Hagemeyer N, Kierdorf K, Frenzel K, et al. Transcriptome-based profiling of yolk sac-derived macrophages reveals a role for Irf8 in macrophage maturation. *EMBO J*. 2016;35(16):1730-1744.
- 44. Tamura T, Kurotaki D, Koizumi S. Regulation of myelopoiesis by the transcription factor IRF8. *Int J Hematol*. 2015;101(4):342-351.
- <span id="page-10-8"></span>45. Hamilton JA, Achuthan A. Colony stimulating factors and myeloid cell biology in health and disease. *Trends Immunol*. 2013;34(2):81-89.
- <span id="page-10-9"></span>46. Mossadegh-Keller N, Sarrazin S, Kandalla PK, et al. M-CSF instructs myeloid lineage fate in single haematopoietic stem cells. *Nature*. 2013;497(7448):239-243.
- <span id="page-10-10"></span>47. Wiktor-Jedrzejczak W, Bartocci A, Ferrante AW Jr, et al. Total absence of colony-stimulating factor 1 in the macrophagedeficient osteopetrotic (op/op) mouse. *Proc Natl Acad Sci USA*. 1990;87(12):4828-4832.
- <span id="page-10-11"></span>48. McKercher SR, Torbett BE, Anderson KL, et al. Targeted disruption of the PU.1 gene results in multiple hematopoietic abnormalities. *EMBO J*. 1996;15(20):5647-5658.
- 49. Scott EW, Simon MC, Anastasi J, Singh H. Requirement of transcription factor PU.1 in the development of multiple hematopoietic lineages. *Science*. 1994;265(5178):1573-1577.
- <span id="page-10-12"></span>50. Mahe D, Fisson S, Montoni A, Morel A, Couez D. Identification and IFNgamma-regulation of differentially expressed mRNAs in murine microglial and CNS-associated macrophage subpopulations. *Mol Cell Neurosci*. 2001;18(4):363-380.
- <span id="page-10-20"></span>51. Faraco G, Sugiyama Y, Lane D, et al. Perivascular macrophages mediate the neurovascular and cognitive dysfunction associated with hypertension. *J Clin Invest*. 2016;126(12):4674-4689.
- 52. Steel CD, Kim WK, Sanford LD, et al. Distinct macrophage subpopulations regulate viral encephalitis but not viral clearance in the CNS. *J Neuroimmunol*. 2010;226(1–2):81-92.
- <span id="page-10-26"></span>53. Thanopoulou K, Fragkouli A, Stylianopoulou F, Georgopoulos S. Scavenger receptor class B type I (SR-BI) regulates perivascular macrophages and modifies amyloid pathology in an Alzheimer mouse model. *Proc Natl Acad Sci USA*. 2010;107(48):20816-20821.
- 54. Li Q, Barres BA. Microglia and macrophages in brain homeostasis and disease. *Nat Rev Immunol*. 2018;18(4):225-242.
- <span id="page-10-13"></span>55. Jung S, Aliberti J, Graemmel P, et al. Analysis of fractalkine receptor CX(3)CR1 function by targeted deletion and green fluorescent protein reporter gene insertion. *Mol Cell Biol*. 2000;20(11):4106-4114.
- <span id="page-10-24"></span>56. Zhang Z, Li Z, Ma Z, et al. Annexin A3 as a marker protein for microglia in the central nervous system of rats. *Neural Plast*. 2021;2021:5575090.
- <span id="page-10-25"></span>57. Konishi H, Kobayashi M, Kunisawa T, et al. Siglec-H is a microglia-specific marker that discriminates microglia from CNSassociated macrophages and CNS-infiltrating monocytes. *Glia*. 2017;65(12):1927-1943.
- <span id="page-10-23"></span>58. Buttgereit A, Lelios I, Yu X, et al. Sall1 is a transcriptional regulator defining microglia identity and function. *Nat Immunol*. 2016;17(12):1397-1406.
- 59. Butovsky O, Weiner HL. Microglial signatures and their role in health and disease. *Nat Rev Neurosci*. 2018;19(10):622-635.
- <span id="page-10-14"></span>60. Bennett ML, Bennett FC, Liddelow SA, et al. New tools for studying microglia in the mouse and human CNS. *Proc Natl Acad Sci USA*. 2016;113(12):E1738-E1746.
- <span id="page-10-15"></span>61. Kim WK, Alvarez X, Fisher J, et al. CD163 identifies perivascular macrophages in normal and viral encephalitic brains and potential precursors to perivascular macrophages in blood. *Am J Pathol*. 2006;168(3):822-834.
- 62. Fabriek BO, van Haastert ES, Galea I, et al. CD163-positive perivascular macrophages in the human CNS express molecules for antigen recognition and presentation. *Glia*. 2005;51(4):297-305.
- <span id="page-10-16"></span>63. Wang G, Zhang J, Hu X, et al. Microglia/macrophage polarization dynamics in white matter after traumatic brain injury. *J Cereb Blood Flow Metab*. 2013;33(12):1864-1874.
- 64. Hu X, Li P, Guo Y, et al. Microglia/macrophage polarization dynamics reveal novel mechanism of injury expansion after focal cerebral ischemia. *Stroke*. 2012;43(11):3063-3070.
- <span id="page-10-17"></span>65. Galea I, Palin K, Newman TA, van Rooijen N, Perry VH, Boche D. Mannose receptor expression specifically reveals perivascular macrophages in normal, injured, and diseased mouse brain. *Glia*. 2005;49(3):375-384.
- <span id="page-10-18"></span>66. Martin E, el-Behi M, Fontaine B, Delarasse C. Analysis of microglia and monocyte-derived macrophages from the central nervous system by flow cytometry. *J Vis Exp*. 2017;(124):55781.
- 67. Robinson AP, White TM, Mason DW. Macrophage heterogeneity in the rat as delineated by two monoclonal antibodies MRC OX-41 and MRC OX-42, the latter recognizing complement receptor type 3. *Immunology*. 1986;57(2):239-247.
- 68. Hammond TR, Dufort C, Dissing-Olesen L, et al. Single-cell RNA sequencing of microglia throughout the mouse lifespan and in the injured brain reveals complex cell-state changes. *Immunity*. 2019;50(1):253-271 e6.
- <span id="page-10-19"></span>69. Raivich G, Haas S, Werner A, Klein MA, Kloss C, Kreutzberg GW. Regulation of MCSF receptors on microglia in the normal and injured mouse central nervous system: a quantitative immunofluorescence study using confocal laser microscopy. *J Comp Neurol*. 1998;395(3):342-358.
- 70. Akiyama H, Nishimura T, Kondo H, Ikeda K, Hayashi Y, McGeer PL. Expression of the receptor for macrophage colony stimulating factor by brain microglia and its upregulation in brains of patients with Alzheimer's disease and amyotrophic lateral sclerosis. *Brain Res*. 1994;639(1):171-174.
- 71. Ito D, Imai Y, Ohsawa K, Nakajima K, Fukuuchi Y, Kohsaka S. Microglia-specific localisation of a novel calcium binding protein, Iba1. *Brain Res Mol Brain Res*. 1998;57(1):1-9.
- 72. Lin HH, Faunce DE, Stacey M, et al. The macrophage F4/80 receptor is required for the induction of antigen-specific efferent regulatory T cells in peripheral tolerance. *J Exp Med*. 2005;201(10):1615-1625.
- <span id="page-11-4"></span>73. Holness CL, Simmons DL. Molecular cloning of CD68, a human macrophage marker related to lysosomal glycoproteins. *Blood*. 1993;81(6):1607-1613.
- <span id="page-11-10"></span>74. Pey P, Pearce RKB, Kalaitzakis ME, Griffin WST, Gentleman SM. Phenotypic profile of alternative activation marker CD163 is different in Alzheimer's and Parkinson's disease. *Acta Neuropathol Commun*. 2014;2:21.
- <span id="page-11-5"></span>75. Lim HY, Lim SY, Tan CK, et al. Hyaluronan receptor LYVE-1-expressing macrophages maintain arterial tone through hyaluronan-mediated regulation of smooth muscle cell collagen. *Immunity*. 2018;49(2):326-341 e7.
- 76. Furube E, Kawai S, Inagaki H, Takagi S, Miyata S. Brain regiondependent heterogeneity and dose-dependent difference in transient microglia population increase during lipopolysaccharideinduced inflammation. *Sci Rep*. 2018;8(1):2203.
- 77. Mildner A, Huang H, Radke J, Stenzel W, Priller J. P2Y12 receptor is expressed on human microglia under physiological conditions throughout development and is sensitive to neuroinflammatory diseases. *Glia*. 2017;65(2):375-387.
- <span id="page-11-6"></span>78. Rajan WD, Wojtas B, Gielniewski B, et al. Defining molecular identity and fates of CNS-border associated macrophages after ischemic stroke in rodents and humans. *Neurobiol Dis*. 2020;137:104722.
- <span id="page-11-0"></span>79. Sweeney MD, Zhao Z, Montagne A, Nelson AR, Zlokovic BV. Blood-brain barrier: from physiology to disease and back. *Physiol Rev*. 2019;99(1):21-78.
- 80. Abbott NJ. Astrocyte-endothelial interactions and blood-brain barrier permeability. *J Anat*. 2002;200(6):629-638.
- 81. Brightman MW, Reese TS. Junctions between intimately apposed cell membranes in the vertebrate brain. *J Cell Biol*. 1969;40(3):648-677.
- 82. Reese TS, Karnovsky MJ. Fine structural localization of a blood-brain barrier to exogenous peroxidase. *J Cell Biol*. 1967;34(1):207-217.
- <span id="page-11-1"></span>83. Ben-Zvi A, Lacoste B, Kur E, et al. Mfsd2a is critical for the formation and function of the blood-brain barrier. *Nature*. 2014;509(7501):507-511.
- 84. Andreone BJ, Chow BW, Tata A, et al. Blood-brain barrier permeability is regulated by lipid transport-dependent suppression of caveolae-mediated transcytosis. *Neuron*. 2017;94(3):581-594 e5.
- 85. Nguyen LN, Ma D, Shui G, et al. Mfsd2a is a transporter for the essential omega-3 fatty acid docosahexaenoic acid. *Nature*. 2014;509(7501):503-506.
- <span id="page-11-2"></span>86. Willis CL, Garwood CJ, Ray DE. A size selective vascular barrier in the rat area postrema formed by perivascular macrophages and the extracellular matrix. *Neuroscience*. 2007;150(2):498-509.
- 87. Prior MJ, Brown AM, Mavroudis G, Lister T, Ray DE, et al. MRI characterisation of a novel rat model of focal astrocyte loss. *MAGMA*. 2004;17(3–6):125-132.
- 88. Willis CL, Leach L, Clarke GJ, Nolan CC, Ray DE. Reversible disruption of tight junction complexes in the rat blood-brain barrier, following transitory focal astrocyte loss. *Glia*. 2004;48(1):1-13.
- <span id="page-11-3"></span>89. Zenker D, Begley D, Bratzke H, Rubsamen-Waigmann H, von Briesen H. Human blood-derived macrophages enhance barrier function of cultured primary bovine and human brain capillary endothelial cells. *J Physiol*. 2003;551(Pt 3):1023-1032.
- <span id="page-11-7"></span>90. Qin J, Lovelace MD, Mitchell AJ, de Koning-Ward T, Grau GE, Pai S. Perivascular macrophages create an intravascular niche for CD8(+)

T cell localisation prior to the onset of fatal experimental cerebral malaria. *Clin Transl Immunol*. 2021;10(4):e1273.

- <span id="page-11-8"></span>91. Kurz C, Walker L, Rauchmann B-S, Perneczky R. Dysfunction of the blood-brain barrier in Alzheimer's disease: evidence from human studies. *Neuropathol Appl Neurobiol*. 2021;48(3):e12782.
- 92. Bedussi B, Almasian M, de Vos J, VanBavel E, Bakker ENTP. Paravascular spaces at the brain surface: low resistance pathways for cerebrospinal fluid flow. *J Cereb Blood Flow Metab*. 2018;38(4):719-726.
- <span id="page-11-9"></span>93. Roche PA, Furuta K. The ins and outs of MHC class II-mediated antigen processing and presentation. *Nat Rev Immunol*. 2015;15(4):203-216.
- 94. Henning EC, Ruetzler CA, Gaudinski MR, et al. Feridex preloading permits tracking of CNS-resident macrophages after transient middle cerebral artery occlusion. *J Cereb Blood Flow Metab*. 2009;29(7):1229-1239.
- <span id="page-11-17"></span>95. Wan H, Brathwaite S, Ai J, Hynynen K, Macdonald RL. Role of perivascular and meningeal macrophages in outcome following experimental subarachnoid hemorrhage. *J Cereb Blood Flow Metab*. 2021;41(8):1842-1857.
- 96. Kierdorf K, Masuda T, Jordão MJC, Prinz M. Macrophages at CNS interfaces: ontogeny and function in health and disease. *Nat Rev Neurosci*. 2019;20(9):547-562.
- <span id="page-11-11"></span>97. Ji R, Ma L, Chen X, et al. Characterizing the distributions of IDO-1 expressing macrophages/microglia in human and murine brains and evaluating the immunological and physiological roles of IDO-1 in RAW264.7/BV-2 cells. *PLoS One*. 2021;16(11):e0258204.
- <span id="page-11-12"></span>98. Clapham R, O'Sullivan E, Weller RO, Carare RO. Cervical lymph nodes are found in direct relationship with the internal carotid artery: significance for the lymphatic drainage of the brain. *Clin Anat*. 2010;23(1):43-47.
- 99. Dissing-Olesen L, Hong S, Stevens B. New brain lymphatic vessels drain old concepts. *EBioMedicine*. 2015;2(8):776-777.
- <span id="page-11-13"></span>100. Carare RO, Bernardes-Silva M, Newman TA, et al. Solutes, but not cells, drain from the brain parenchyma along basement membranes of capillaries and arteries: significance for cerebral amyloid angiopathy and neuroimmunology. *Neuropathol Appl Neurobiol*. 2008;34(2):131-144.
- 101. Foldi M, Gellert A, Kozma M, Poberai M, Zoltán ÖT, Csanda E. New contributions to the anatomical connections of the brain and the lymphatic system. *Acta Anat (Basel)*. 1966;64(4):498-505.
- 102. Louveau A, Da Mesquita S, Kipnis J. Lymphatics in neurological disorders: a neuro-lympho-vascular component of multiple sclerosis and Alzheimer's disease? *Neuron*. 2016;91(5):957-973.
- 103. Aspelund A, Antila S, Proulx ST, et al. A dural lymphatic vascular system that drains brain interstitial fluid and macromolecules. *J Exp Med*. 2015;212(7):991-999.
- <span id="page-11-14"></span>104. Expression of concern: blockade of Fas signaling in breast cancer cells suppresses tumor growth and metastasis via disruption of Fas signaling-initiated cancer-related inflammation. *J Biol Chem*. 2020;295(26):8886.
- 105. Kaminski M, Bechmann I, Pohland M, Kiwit J, Nitsch R, Glumm J. Migration of monocytes after intracerebral injection at entorhinal cortex lesion site. *J Leukoc Biol*. 2012;92(1):31-39.
- 106. Kaminski M, Bechmann I, Kiwit J, Glumm J. Migration of monocytes after intracerebral injection. *Cell Adh Migr*. 2012;6(3):164-167.
- <span id="page-11-15"></span>107. Albargothy NJ, Johnston DA, MacGregor-Sharp M, et al. Convective influx/glymphatic system: tracers injected into the CSF enter and leave the brain along separate periarterial basement membrane pathways. *Acta Neuropathol*. 2018;136(1):139-152.
- <span id="page-11-16"></span>108. Drieu A, Lanquetin A, Levard D, et al. Alcohol exposure-induced neurovascular inflammatory priming impacts ischemic stroke and is linked with brain perivascular macrophages. *JCI Insight*. 2020;5(4):e129226.

- <span id="page-12-7"></span>109. Schwab JM, Nguyen TD, Postler E, Meyermann R, Schluesener HJ. Selective accumulation of cyclooxygenase-1-expressing microglial cells/macrophages in lesions of human focal cerebral ischemia. *Acta Neuropathol*. 2000;99(6):609-614.
- <span id="page-12-8"></span>110. Pedragosa J, Salas-Perdomo A, Gallizioli M, et al. CNS-border associated macrophages respond to acute ischemic stroke attracting granulocytes and promoting vascular leakage. *Acta Neuropathol Commun*. 2018;6(1):76.
- <span id="page-12-9"></span>111. Inagaki T, Fujiwara K, Shinohara Y, et al. Perivascular macrophages produce type I collagen around cerebral small vessels under prolonged hypertension in rats. *Histochem Cell Biol*. 2021;155(4):503-512.
- <span id="page-12-5"></span>112. Iyonaga T, Shinohara K, Mastuura T, Hirooka Y, Tsutsui H. Brain perivascular macrophages contribute to the development of hypertension in stroke-prone spontaneously hypertensive rats via sympathetic activation. *Hypertens Res*. 2020;43(2):99-110.
- <span id="page-12-0"></span>113. Zhang F, Eckman C, Younkin S, Hsiao KK, Iadecola C. Increased susceptibility to ischemic brain damage in transgenic mice overexpressing the amyloid precursor protein. *J Neurosci*. 1997;17(20):7655-7661.
- 114. Niwa K, Kazama K, Younkin L, Younkin SG, Carlson GA, Iadecola C. Cerebrovascular autoregulation is profoundly impaired in mice overexpressing amyloid precursor protein. *Am J Physiol Heart Circ Physiol*. 2002;283(1):H315-H323.
- 115. Niwa K, Younkin L, Ebeling C, et al. Abeta 1-40-related reduction in functional hyperemia in mouse neocortex during somatosensory activation. *Proc Natl Acad Sci USA*. 2000;97(17):9735-9740.
- 116. Iadecola C, Zhang F, Niwa K, et al. SOD1 rescues cerebral endothelial dysfunction in mice overexpressing amyloid precursor protein. *Nat Neurosci*. 1999;2(2):157-161.
- 117. Han BH, Zhou ML, Abousaleh F, et al. Cerebrovascular dysfunction in amyloid precursor protein transgenic mice: contribution of soluble and insoluble amyloid-beta peptide, partial restoration via gamma-secretase inhibition. *J Neurosci*. 2008;28(50):13542-13550.
- 118. Koizumi K, Wang G, Park L. Endothelial dysfunction and amyloidbeta-induced neurovascular alterations. *Cell Mol Neurobiol*. 2016;36(2):155-165.
- 119. Katusic ZS, Austin SA. Endothelial nitric oxide: protector of a healthy mind. *Eur Heart J*. 2014;35(14):888-894.
- 120. Zlokovic BV. Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat Rev Neurosci*. 2011;12(12):723-738.
- <span id="page-12-1"></span>121. Cirrito JR, Yamada KA, Finn MB, et al. Synaptic activity regulates interstitial fluid amyloid-beta levels in vivo. *Neuron*. 2005;48(6):913-922.
- <span id="page-12-2"></span>122. Ransohoff RM, Engelhardt B. The anatomical and cellular basis of immune surveillance in the central nervous system. *Nat Rev Immunol*. 2012;12(9):623-635.
- 123. Adlimoghaddam A, Neuendorff M, Roy B, Albensi BC. A review of clinical treatment considerations of donepezil in severe Alzheimer's disease. *CNS Neurosci Ther*. 2018;24(10):876-888.
- <span id="page-12-3"></span>124. Iadecola C. The pathobiology of vascular dementia. *Neuron*. 2013;80(4):844-866.
- 125. Kazama K, Anrather J, Zhou P, et al. Angiotensin II impairs neurovascular coupling in neocortex through NADPH oxidase-derived radicals. *Circ Res*. 2004;95(10):1019-1026.
- 126. Capone C, Faraco G, Peterson JR, et al. Central cardiovascular circuits contribute to the neurovascular dysfunction in angiotensin II hypertension. *J Neurosci*. 2012;32(14):4878-4886.
- 127. Faraco G, Iadecola C. Hypertension: a harbinger of stroke and dementia. *Hypertension*. 2013;62(5):810-817.
- <span id="page-12-4"></span>128. Kerkhofs D, van Hagen BT, Milanova IV, et al. Pharmacological depletion of microglia and perivascular macrophages prevents Vascular Cognitive Impairment in Ang II-induced hypertension. *Theranostics*. 2020;10(21):9512-9527.
- 129. Schiltz JC, Sawchenko PE. Distinct brain vascular cell types manifest inducible cyclooxygenase expression as a function of the strength and nature of immune insults. *J Neurosci*. 2002;22(13):5606-5618.
- <span id="page-12-6"></span>130. Nagano T, Tsuda N, Fujimura K, Ikezawa Y, Higashi Y, Kimura SH. Prostaglandin E2 increases the expression of cyclooxygenase-2 in cultured rat microglia. *J Neuroimmunol*. 2021;361:577724.
- <span id="page-12-10"></span>131. GBD 2019 Stroke Collaborators. Global, regional, and national burden of stroke and its risk factors, 1990-2019: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet Neurol*. 2021;20(10):795-820.
- <span id="page-12-11"></span>132. Lee JY, Keep RF, He Y, Sagher O, Hua Y, Xi G. Hemoglobin and iron handling in brain after subarachnoid hemorrhage and the effect of deferoxamine on early brain injury. *J Cereb Blood Flow Metab*. 2010;30(11):1793-1803.
- 133. Clark JF, Sharp FR. Bilirubin oxidation products (BOXes) and their role in cerebral vasospasm after subarachnoid hemorrhage. *J Cereb Blood Flow Metab*. 2006;26(10):1223-1233.
- 134. Joerk A, Ritter M, Langguth N, et al. Propentdyopents as heme degradation intermediates constrict mouse cerebral arterioles and are present in the cerebrospinal fluid of patients with subarachnoid hemorrhage. *Circ Res*. 2019;124(12):e101-e114.
- 135. Rowland MJ, Hadjipavlou G, Kelly M, Westbrook J, Pattinson KTS. Delayed cerebral ischaemia after subarachnoid haemorrhage: looking beyond vasospasm. *Br J Anaesth*. 2012;109(3):315-329.
- <span id="page-12-12"></span>136. Villa A, Vegeto E, Poletti A, Maggi A. Estrogens, neuroinflammation, and neurodegeneration. *Endocr Rev*. 2016;37(4):372-402.
- 137. Thion MS, Low D, Silvin A, et al. Microbiome influences prenatal and adult microglia in a sex-specific manner. *Cell*. 2018;172(3):500- 516 e16.

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