# Myeloid checkpoints for cancer immunotherapy

# Yixin Qian<sup>1,2\*</sup>, Ting Yang<sup>1,2\*</sup>, Huan Liang<sup>2</sup>, Mi Deng<sup>1,2,3</sup>

<sup>1</sup>Peking University International Cancer Institute, Health Science Center, Peking University, Beijing 100191, China; <sup>2</sup>School of Basic Medical Sciences, Health Science Center, Peking University, Beijing 100191, China; <sup>3</sup>Peking University Cancer Hospital & Institute, Peking University, Beijing 100142, China

\*These authors contributed equally to this work.

Correspondence to: Mi Deng. Peking University International Cancer Institute, Health Science Center, Peking University, Beijing 100191, China; School of Basic Medical Sciences, Health Science Center, Peking University, No. 38 Xueyuan Road, Haidian District, Beijing 100191, China. Email: mideng@bjmu.edu.cn.

#### Abstract

Myeloid checkpoints are receptors on the myeloid cell surface which can mediate inhibitory signals to modulate anti-tumor immune activities. They can either inhibit cellular phagocytosis or suppress T cells and are thus involved in the pathogenesis of various diseases. In the tumor microenvironment, besides killing tumor cells by phagocytosis or activating anti-tumor immunity by tumor antigen presentation, myeloid cells could execute protumor efficacies through myeloid checkpoints by interacting with counter-receptors on other immune cells or cancer cells. In summary, myeloid checkpoints may be promising therapeutic targets for cancer immunotherapy.

Keywords: Cancer; immune checkpoint; myeloid checkpoint; MDSC; TAM; immunotherapy

Submitted Aug 26, 2022. Accepted for publication Oct 08, 2022. doi: 10.21147/j.issn.1000-9604.2022.05.07 View this article at: https://doi.org/10.21147/j.issn.1000-9604.2022.05.07

## Introduction

Anti-tumor immunotherapies, conventionally referred to as immune checkpoint inhibitors (ICIs), target T-cell immune checkpoints and have elicited impressive therapeutic responses in the treatment of human cancers (1). Despite achieving the long-term survival of 10%–30% of treated individuals, immunotherapies are not effective for most patients suffering from cancer (2). A primary challenge of this strategy for extensive anticancer application remains the cell complexity of the tumor immune microenvironment. In addition to many endeavors to seek intrinsic factors to enhance the efficacy of immune therapy in both tumor and T cells (3-5), many studies have revealed that the therapeutic efficacy could be improved by targeting immune regulatory cells, including myeloid cells in the tumor microenvironment (TME) (6,7).

Myeloid cells are the most abundant hematopoietic cells in the human body and have diverse functions. All myeloid cells arise from multipotent hematopoietic stem cells (HSCs) that develop into mature myeloid cells through sequential steps of differentiation (*Figure 1*) (6). Various tumors show abundant infiltration of myeloid cells. More specifically, tumor-infiltrating myeloid cells (TIMs) are constitutive of several myeloid lineages, including mast cells, plasmacytoid dendritic cells (pDCs) (8), conventional dendritic cells (cDCs) (9), monocytes (10), macrophages (11), and granulocytes (12). A pan-cancer single-cell transcriptional atlas of TIMs across 15 human cancer types showed that monocytes and macrophages accounted for the largest proportion of TIMs, with an average of above 50% in most tumors, and the proportion of cDCs was relatively stable (approximately 10%–20%) across tumor types, while the proportion of mast cells showed great variation across different types of tumors (13).

Single-cell RNA-seq also indicated that in the TME, myeloid cells experience continuous reprogramming (14). Following TME-specific cues, monocytes can differentiate into inflammatory macrophages (M1-like type) and monocyte-derived DCs, whose functions are anti-tumor, or



Figure 1 Stages of myelopoiesis differentiation. Myeloid cells originate from HSCs and MPPs. Myelopoiesis is amplified during chronic inflammation to assist tumor progression and dissemination. HSCs differentiate into CMP, which can further differentiate through hematopoietic system. In physiological conditions, CMP can differentiate into neutrophils or monocytes, and subsequently into DCs or macrophages. However, with chronic inflammation, pro-inflammatory cytokines can skew monocytopoiesis of CMP into M-MDSC and TAM, and granulopoiesis into PMN-MDSC and TAN. HSC, hematopoietic stem cell; MPP, multipotent progenitor cell; CMP, common myeloid progenitor; DC, dendritic cell; M-MDSC, monocytic myeloid-derived suppressor cell; TAM, tumor-associated macrophage; PMN-MDSC, polymorphonuclear myeloid-derived suppressor cell; TAN, tumor-associated neutrophil; CDP, common DC progenitor; cDC, conventional DC; pDC, plasmacytoid DC.

alternatively activated macrophages (M2-like type) with immunosuppressive capacity. The differentiation from monocytes to M2 macrophages may be the predominant path in the process of myeloid reprogramming (15).

These cells have a profound impact on anti-tumor effect and could influence neoangiogenesis, and sustain cancer cell proliferation, metastasis and immunotherapy resistance (16). On the one hand, myeloid cells can directly kill tumor cells by phagocytosis (17) or activate anti-tumor immunity by tumor antigen presentation (6). On the other hand, a group of myeloid cells, including tumor-associated macrophages (TAMs), tumor-associated neutrophils (TANs), myeloid-derived suppressor cells (MDSCs) and inhibitory DCs, support tumor development and metastasis by suppressing infiltrating cytotoxic lymphocytes (18). The activities and functions of these myeloid cells are regulated by stimulatory or inhibitory signals mediated by receptors on the cell surface, and inhibitory signals are considered myeloid checkpoints, which may contain an immunereceptor tyrosine-based inhibitory motif (ITIM)/immunereceptor tyrosine-based switch motif (ITSM) to transduce immune modulatory signals in myeloid cells or interact with counter-receptors on other immune cells to modulate their activities or on cancer cells to execute pro-tumor efficacies. A new type of immunotherapeutic strategy targeting myeloid cells has also emerged (19).

In this review, we will discuss the biological functions and molecular mechanisms of myeloid checkpoints below.

## **Checkpoints expressed by myeloid cells**

We briefly introduce CD47, signaling lymphocytic activation molecule (SLAM) family, programmed cell death 1 (PD-1), sialic acid-binding immunoglobulin-type lectin (Siglec) family, leukocyte immunoglobulin-like receptor B (LILRB) family, triggering receptor expressed on myeloid cells 2 (TREM2), neuropilin-1 (NRP1) and common lymphatic endothelial and vascular endothelial receptor-1

#### 462

#### Qian et al. Myeloid checkpoints for cancer immunotherapy

(Clever-1) in this chapter (Figure 2).

## CD47/SIRPa

CD47 (also known as inhibitor of apoptosis protein, IAP) is a red blood cell signal that serves to discriminate self and non-self (20). Upon engagement of CD47 by signal regulatory protein alpha [SIRP $\alpha$ ; also known as SH2domain bearing protein tyrosine phosphatase (SHP) substrate-1 (SHPS-1) or CD172a] on macrophages and DCs, the ITIM in the cytoplasmic tail of SIRP $\alpha$  activates the inhibitory tyrosine phosphatases SHP-1 and SHP-2, resulting in inhibition of the actin cytoskeleton rearrangement required for phagocytosis (*Figure 3*). CD47 is upregulated on the surface of several cancer cell types, enabling these cells to evade phagocytic removal by



**Figure 2** Current myeloid checkpoints in cancer immunotherapy. Ligands and elementary structures of CD47, SLAMF7, PD-1, LILRB1, LILRB2, LILRB4, Siglec-1, Siglec-9, Siglec-10, Siglec-15, TREM2, NRP1 and Clever1 are shown in this figure. PD-1, programmed cell death 1; LILRB, leukocyte immunoglobulin-like receptor B; Siglec, sialic acid-binding immunoglobulin-type lectin; TREM2, triggering receptor expressed on myeloid cells 2; NRP1, neuropilin-1; Clever1, common lymphatic endothelial and vascular endothelial receptor-1.



Figure 3 Role of CD47/SIRP $\alpha$ . On the one hand, after binding to CD47, ITIM in the cytoplasmic tail of SIRP $\alpha$  on macrophage activates inhibitory tyrosine phosphatases SHP-1 and SHP-2, resulting in inhibition of the actin cytoskeleton rearrangement required for phagocytosis. On the other hand, SIRP $\alpha$  on DC inhibits the antigen presentation of tumor cells. ITIM, tyrosine-based inhibitory motif; DC, dendritic cell.

immune cells (21). In accord with this, high CD47 expression is negatively correlated with disease prognosis and survival in acute myeloid leukemia (AML), non-small cell lung cancer (NSCLC) and high-grade serous ovarian carcinoma (21-24). Furthermore, therapeutic reagents that antagonize the CD47-SIRPa axis in macrophages inhibit the growth of several types of tumors, including AML and non-Hodgkin lymphoma, by enhancing cellular phagocytosis (25-28). Furthermore, CD47 blockade has been shown to robustly increase immune responses against tumor antigens by activating DCs but not macrophage cross-presentation of tumor antigens (29). In recent years, combined therapies of a CD47-SIRPa axis inhibitor and other anticancer therapeutics [e.g., PD-1/programmed cell death ligand-1 (PD-L1) blockade] have been investigated in the context of potentiating anti-tumor immunity in vitro (25, 30).

# SLAM family

The SLAM family of receptors is expressed on the majority of immune cells. These receptors often serve as self-ligands and play important roles in cellular communication and adhesion, thus modulating immune responses (*Figure 4*)

(31,32). SLAM family receptors (SFRs) play a key role in inhibiting macrophage phagocytosis. SFR deficiency triggers macrophage phagocytosis of hematopoietic cells, resulting in severe rejection of donor hematopoietic grafts in recipient mice (33).

#### SLAMF7

The SLAMF7 [also acknowledged as CD2-like receptoractivating cvtotoxic cell (CRACC), CS1 and CD319] receptor belongs to the SLAM family of receptors (34), presenting at diverse periodicities on different types of immune cells, such as macrophages, and restrictively expressed on hematopoietic cells (32,34,35). Except for SLAMF4 (2B4), SLAM family members function as homotypic receptors, which recruit different SH2 domaincontaining proteins to their cytoplasmic immune-receptor tyrosine-based switch motifs when activated (34) so that SLAMF7 can regulate immune cell-specific functions across diverse types of immune cells (34-36). SLAM family receptors are correlated with T-cell exhaustion. SLAMF6 is a marker of the progenitor of exhausted CD8<sup>+</sup> T cells (37), and both SLAMF6 and SLAMF4 (2B4) are reported as inhibitory receptors of CD8+ T cells (34). As reported,



**Figure 4** Role of SLAM family. In T cells, the activation of self-ligand SLAMF7 immune receptor could induce phosphorylation of STAT1 and STAT3 and expression of various inhibitory receptors and transcription factors associated with T cell exhaustion. In macrophages, SLAMF7 could mediate phagocytosis interacting with integrin Mac-1 and do with signals involving immune-receptor tyrosine-based activation motifs. While most SLAM-mediated phagocytosis functions depend on SAP adaptors. SLAMF3 and SLAMF4 are identified as "don't eat me" receptors on macrophages which inhibit "eat-me" signals such as LRP1-mediated activation of mTOR and Syk through SH2 domain-containing phosphatases. STAT, signal transduction and activator of transcription; SAP, signaling lymphocyte activation molecule-associated protein; GPI, glycosyl phosphatidyl inositol.

SLAMF7 is also expressed on a CD8+ T-cell subset enriched in melanoma patients who are not responsive to checkpoint blockade immune therapy (38). Moreover, SLAMF7 is expressed on certain kinds of memoryprecursor and effector CD8+ T cells, which respond to checkpoint blockade in an indirect way (39). A recent study showed that activation of the self-ligand SLAMF7 immune receptor on T cells could induce phosphorylation of signal transduction and activator of transcription 1 (STAT1) and STAT3, and expression of various inhibitory receptors and transcription factors associated with T-cell exhaustion (31,40). High SLAMF7 expression is reported to indicate poor survival in clear cell renal cell carcinoma (ccRCC) (31). Moreover, SLAMF7<sup>high</sup>CD38<sup>high</sup> TAMs show strong relations with exhausted T cells and are an independent prognostic factor of ccRCC (31). Apart from the role of SLAMF7 in modulating T-cell function in the TME, SLAMF7 could mediate phagocytosis by interacting with integrin Mac-1 (41) and with signals involving immunereceptor tyrosine-based activation motifs (42). Most SLAM-mediated phagocytosis functions depend on molecule-associated lymphocyte activation signaling protein (SAP) adaptors (34,43). What's more, it has been shown that patients whose tumors express SLAMF7 are more likely to respond to SIRPa-CD47 blockade therapy (33).

# SLAMF3 & SLAMF4

Specifically, SLAMF3 (Ly-9, CD229) and SLAMF4 (2B4, CD244) were identified as "do not eat me" receptors on macrophages (44). These receptors inhibit "eat-me" signals, such as LRP1-mediated activation of mTOR and Syk, through SH2 domain-containing phosphatases. SFRs bind to but are independent of CD47 to alleviate macrophage phagocytosis, and combined deletion of SFRs and CD47 results in reduced hematopoietic cells in mice. This SFR-mediated tolerance is compromised in patients with hemophagocytic lymphohistiocytosis, a syndrome characterized by inappropriate phagocytosis of hematopoietic cells. Thus, SFR-mediated inhibition of macrophage phagocytosis is critical for hematopoietic homeostasis, and SFR may represent a previously unknown tumor immunotherapy target (44).

#### PD-1/PD-L1

PD-1 (also known as CD279) is one of the bestcharacterized immune checkpoint targets for cancer immunotherapy, and PD-1 blockade has proven to be the most successful immunotherapy strategy to date (45). It has been reported that increased galectin-9<sup>+</sup> M-MDSCs in the peripheral blood of NSCLC patients are involved in resistance to anti-PD-1 therapy (46). Therefore, the presence of MDSCs in the TME is detrimental for the anti-PD-1/PD-L1 response. As expected, several studies revealed the relationship between MDSC infiltration and PD-1 blockade resistance, and selective depletion of MDSCs could restore anti-PD-1 efficacy (47,48).

In addition, it has been found that PD-1 is expressed in TAMs but not in circulating monocytes or splenic macrophages and that it can negatively regulate cancer cell phagocytosis (30). In addition, as reported, blockade of PD-1 or PD-L1 enhanced the phagocytic activity of TAMs towards tumor cells in vivo and inhibited tumor growth in a macrophage-dependent fashion, suggesting that the PD-1/PD-L1 axis is an immune checkpoint for regulating both innate and adaptive immunity. They further found that PD-1 blockade also changed the nature of TAMs from immunosuppressive M2-like cells to pro-inflammatory M1like cells. The presence of TAMs in pancreatic cancer exaggerates immunosuppression within the microenvironment and leads to PD-1/PD-L1 blockade resistance. Inhibition of colony-stimulating factor 1 receptor (CSF1R) on TAMs can upregulate the expression of PD-L1 and increase CD8+ T-cell infiltration, which ablates anti-PD-1/PD-L1 resistance (49). PD-1-expressing macrophages exhibit an anti-inflammatory-like surface profile in tumor settings. PD-1 expression is negatively correlated with the phagocytic ability of macrophages, and blockade of PD-1/PD-L1 enhances antitumor responses and prolongs survival by inhibiting pro-inflammatory to anti-inflammatory macrophage polarization (50,51). PD-L1 antibody treatment promoted the proliferation of cultured bone marrow-derived macrophages, which was related to the activation of the AKT-mTOR pathway (Figure 5) (52). The transcriptomic profiles of macrophages were switched to inflammatory phenotypes following PD-L1 antibody treatment (53). The interaction between the immune checkpoint molecules PD-1 and PD-L1 may also be responsible for TAN-mediated immune suppression. In human hepatocellular carcinoma (HCC) tissues and HCCbearing mice, PD-L1 and PD-1 were expressed in peritumoral neutrophils and T lymphocytes, respectively, and PD-L1<sup>+</sup> neutrophils from patients with HCC effectively suppressed the proliferation and activation of T cells by utilizing PD-1/PD-L1 interactions (54).

In addition to PD-L1, B7 superfamily member 1 (B7S1)



Figure 5 Role of PD-1/PD-L1. PD-1-expressing macrophages exhibit an anti- inflammatory-like surface profile in tumor settings. PD-1 expression is negatively correlated to phagocytic ability of macrophages. PD-L1 antibody treatment promotes cell proliferation of cultured bone marrow-derived macrophages, which is related to the activation of the AKT-mTOR pathway. PD-1, programmed cell death 1; PD-L1, programmed cell death ligand 1.

(also known as B7-H4, B7x, or VTCN1) is expressed by tumor-infiltrating antigen-presenting cells (APCs) and triggers dysfunction and exhaustion of activated CD8+ TILs by Eomes upregulation (55). Murine experiments indicated that B7S1 inhibition suppresses the development of liver cancer and enhances antitumor CTLs and that the combination of B7S1 and PD-1 blockade could inhibit tumor growth in a synergistic way (55).

In summary, together with PD-1, B7S1 could be an intriguing target to reverse T-cell exhaustion and maximize immunotherapeutic efficacy in cancer.

# Siglec family

Sialic acid-binding immunoglobulin-type lectins (Siglecs) contain a family of cell surface proteins playing an essential role in regulating immune homeostasis (Figure 6). The dysregulation of Siglecs is reported to be correlated with various diseases, including autoimmunity diseases. infections and cancer Siglecs (56). are type Ι transmembrane proteins with one V-set immunoglobulin (Ig) domain, including a sialic acid-binding site, and one or more C2-set Ig domains in the extracellular region. Most Siglecs, such as CD22 (Siglec-2) and most CD33 (Siglec-3)-related Siglecs, have ITIMs and/or ITIM-like motifs in the intracellular domain and convey signaling mediated by

inhibitory receptors (57). Each Siglec prefers a different kind of sialic acid expressed on all mammalian cells to distinguish self and non-self (58). Mammalian cells have a relatively high density of sialoglycans on their surface in comparison to most pathogens. This high density of sialoglycans, which can be considered self-associated molecular patterns (SAMPs), can lead to the inhibition of immune responses against the self (59). Siglec is mainly expressed on hematopoietic cells, almost on myeloid cells and B cells or other types of cells, such as neurons (56,58).

## CD169

CD169, which is also called sialoadhesin or Siglec-1, is expressed mostly on macrophages conserved among mammals (60,61). The CD169 molecule consists of 17 Iglike domains, including an N-terminal V-set domain and 16 C2-set domains (62). CD169 can serve as a functional ligand, different from most Siglecs, which work as receptors (57). The functions of CD169 are involved in cell-to-cell adhesion and cell-pathogen interactions (63) including sialylated pathogen uptake, antigen presentation, and modulating self-antigen tolerance (64). CD169 could potentially contribute to both the damping and the facilitation of antitumor immunity (65). On the one hand, Siglec1 can recognize and bind surface polysaccharides,



**Figure 6** Role of Siglec family. On the one hand, Siglec-1 could associate with DAP12 to recruit and activate SHP2, which then could recruit E3 ubiquitin ligase TRIM27, inducing TBK1 degradation. Therefore, type I IFN production is inhibited and innate immune response could be suppressed. On the other hand, Syk activation occurs when Siglec-1 binds to DAP12, leading to increased production of TGF- $\beta$ 1, which could inhibit production of NF-kB, Siglec-9 and Siglec-10. Membrane-proximal ITIM of Siglec-9 and Siglec-10 could offer docking sites for SHP-1/2 once tyrosine is phosphorylated, which could inhibit the role of Toll-like-receptor, resulting in increased production of NF-kB. SHP, SH2-domain bearing protein tyrosine phosphatase; IFN, interferon; TGF, transforming growth factor; Siglec, sialic acid-binding immunoglobulin-type lectin.

such as CD43, on T cells to implement cell-to-cell communication (66). CD169+ macrophages in the marginal zone of the spleen recognize phosphatidylserine on the surface of apoptotic cells, acting as APCs to present apoptotic cell antigens so that Tregs are recruited to exhibit their immune tolerance role (67). On the other hand, CD169 in lymph nodes plays an immunoregulatory role by interacting with MUC-1 binding on the surface of breast cancer tumor cells (68). A high density of CD169+ macrophages in the regional lymph nodes of colorectal, endometrial carcinoma and malignant melanoma patients indicates a better clinical prognosis, perhaps owing to an increase in the number of tumor-infiltrating cytotoxic immune cells (69,70). CD169+ macrophages can also directly contact CD43-expressing CD8+ T cells (71). In endometrial carcinoma patients, a high density of CD169+ cells in the regional lymph nodes of the tumor is associated with a higher density of CD8+ T cells and NK cells in the tumor tissue (69,70). The number of intratumoral cells

expressing CD169 also positively correlates with the number of infiltrating CD8<sup>+</sup> T cells in the tumor and with patient survival (71). In contrast, the subcapsular sinus macrophages in regional lymph nodes of patients with advanced-stage endometrial carcinoma or metastatic carcinoma bear lower levels of CD169 or are completely dismissed from this molecule (70). Furthermore, some results suggested that upon vesicular stomatitis virus (VSV) infection, Siglec1 can associate with DAP12 and SHP2, triggering suppression of type I IFN production (72). There is also evidence that Siglec-1 can interact with and activate the DAP12-Syk pathway to increase the production of TGF- $\beta$ 1, which plays an essential role in sepsis and endotoxin tolerance (73).

## Siglec-9

Siglec-9 is expressed on various kinds of human blood leukocytes, including lymphocytes and myeloid cells such as B cells, small groups of T cells, monocytes, neutrophils,

and NK cells (74). Siglec-9 is mainly expressed on neutrophils in peripheral blood (75) and on NK cells, B cells and monocytes afterwards (76). As plentiful evidence indicates, Siglec-9 plays inhibitory roles in modulating immune homeostasis (77) by interacting with the membrane-proximal ITIM. ITIMs can offer docking sites for SHP-1/2 once tyrosine is phosphorylated (78). Siglec-9 could also link with transmembrane epithelial MUC, a kind of strongly glycosylated protein basically produced by epithelial tissues, which could trigger immune evasion (79). MUC1 overexpressed on adenocarcinomas and hematological cancers could recruit β-catenin binding to its C-terminal domain to give rise to the growth of tumor cells (80). MUC1-sialylated O-linked glycan binding to Siglec-9 could induce calcium flux, leading to the activation of MEK-ERK kinases instead of recruiting SHP-1/2 (81). Siglec-9 could also interact with MUC16 expressed on epithelial ovarian cancer cells, which protects tumor cells from immune clearance (82).

#### Siglec-E

Siglec-E is the mouse orthologue of human Siglec-9. Similar to Siglec-9, Siglec-E was first found on neutrophils, macrophages and monocytes (83). Siglec-E could stop inflammatory responses mediated by immune cells when binding to sialoglycan ligands in its extracellular region (83). A Siglec-E-deficient mouse model (84) was utilized to explore the role of Siglecs in myeloid cells (85). As reported, neutrophil-associated Siglec-E could promote extravasation and colony formation of tumor cells in the lungs. The tumoricidal effect and reactive oxygen species (ROS) production mediated by neutrophils are shown to be enhanced *in vitro* with the deficiency of Siglec-E (85,86). TAMs in mice with Siglec-E deficiency have a higher preference for polarization into M2 macrophages and could enhance subcutaneous tumor growth (85).

# Siglec-10

CD24 (also known as heat stable antigen or small-cell lung carcinoma cluster 4 antigen) is a heavily glycosylated glycosylphosphatidylinositol-anchored surface protein (87). It is reported to interact with the inhibitory receptor sialic-acid-binding Ig-like lectin 10 (Siglec-10) on innate immune cells to dampen damaging inflammatory responses to infection (88), sepsis (89), liver damage and chronic graft *vs.* host diseases (87,90). The binding of CD24 to Siglec-10 triggers an inhibitory signaling cascade, which is mediated

Src homology region 2 domain-containing bv phosphatases, SHP-1 and/or SHP-2. The phosphatases are associated with the two immune-receptor tyrosine-based inhibition motifs in the cytoplasmic tail of Siglec-10, thereby blocking Toll-like receptor-mediated inflammation and the cytoskeletal rearrangement required for cellular engulfment by macrophages (58,91,92). Moreover, the role of CD24 in modulating tumor immune responses has been reported. CD24 is a potent antiphagocytic "don't eat me" signal that is capable of directly protecting cancer cells from attack by Siglec-10-expressing macrophages. Many tumors overexpress CD24, and TAMs express high levels of Siglec-10. Genetic ablation of either CD24 or Siglec-10, as well as blockade of the CD24-Siglec-10 interaction using monoclonal antibodies, robustly augmented the phagocytosis of all CD24-expressing human tumors and led to macrophage-dependent reduction of tumor growth in vivo and an increase in survival time (93). Studies have shown that CD24 is expressed by several solid tumors, such as ovarian cancer and breast cancer (94,95), which demonstrate weaker responses to anti-PD-L1/PD-1 immunotherapies than other cancers (52,96), suggesting that an alternative strategy may be required to achieve responses across a wide range of cancers.

#### Siglec-15

Among the Siglec family members, Siglec-15 has been identified as a very unique member, selectively expressed on myeloid cells and osteoclasts (a bone-specific myeloid lineage) and generally absent in other immune cells and tissues (97,98). Siglec-15 was identified as a novel T-cellinhibitory molecule, which was originally characterized as an osteoclast modulator (99). The expression of Siglec-15 is normally limited to cells in the myeloid lineage but can be upregulated in many human cancers (97). As a recent study indicated, Siglec-15 strongly suppresses antigen-specific Tcell responses in vitro and in vivo and can mediate immune evasion in the TME (100). As an immune suppressive molecule, the Siglec-15 pathway is nonredundant to the B7-H1/PD-1 pathway (100,101). Siglec-15 can be upregulated by macrophage colony-stimulating factor (M-CSF) released by diverse cell types in response to inflammatory cytokines or by tumor cells, while under physiological conditions, it is expressed on macrophages at a low level (102).

Apart from Siglec-15's unique induction mechanism by M-CSF, IFN $\gamma$ , the major inducer of PD-L1 (101), is

significantly demonstrated to suppress the expression of Siglec-15 on macrophages (100). Unlike the majority of Siglec members, there are no typical ITIMs or ITIM-like motifs in the intracellular domain of Siglec15 (58). Instead, it was reported to be associated with the adaptors DAP12 and DAP10, which contain an immunoreceptor tyrosinebased activation motif (ITAM) (103). DAP12 can recruit PI3K (104) and then promote an inflammatory response by activating the mitogen-activated protein kinases (MAPK) pathway (105). Furthermore, Siglec-15 could inhibit T cell NF-KB/NFAT signaling in a direct way once binding and could suppress the proliferation of T cells and the production of cytokines, and the inhibitory function could be mediated by IL-10 (99). In addition, Siglec-15 could behave as a macrophage receptor and produce TGF<sup>β</sup> when binding with its sialic acid ligand Sialyl-Tn on tumor cells (101). When the levels of IL10 and TGF<sup>β</sup> rise in the TME, the immune-suppressive effect of Siglec-15 is magnified (100).

# LILRB family

The LILR family comprises a set of paired immunomodulatory receptors expressed among human myeloid and lymphocyte cell populations. The leukocyte immunoglobulin-like receptor (LILR, LIR, ILT, CD85) family can be divided into two classes: the inhibitory LILR subfamily B (LILRB1-5) and the activating LILR subfamily A (LILRA1-6). Inhibitory LILRB receptors were first identified in 1997 (106). Expression is enriched in myeloid cell populations and is primate specific, reflecting rapid gene duplication and evolution within the leukocyte receptor complex of chromosomes (106). LILRs are closely linked to the human killer cell inhibitory receptor (KIR) family, and both LILRs and KIRs share similar Ig-like structures and cytoplasmic signaling domains. Whereas KIR expression is restricted to natural killer (NK) cells, LILRs are expressed on various immune cells, including NK, T, and B lymphocytes and myelomonocytic cells (monocytes, macrophages, DCs and granulocytes). LILRB expression has also been reported in other cell types, including osteoclasts (107), leukemia, stromal and endothelial cells, and various cancers (108). LILRB expression in cancer has been associated with enhanced tumor growth and correlates with poor survival outcomes. LILRB1 is broadly expressed on myeloid cells, B cells and subsets of NK cells and T cells. LILRB2-5 are more restricted to cells of myeloid origin and DCs. LILRB receptors contain cytoplasmic (S/I/V/LxYxxI/V/L) ITIM

domains to recruit the Src homology 2 domain-containing phosphatase SHP1/SHP2/SHIP, leading to inhibited immune signaling cascades. SHP/SHIP phosphatase activity is critical in maintaining immune homeostasis (*Figure 7*) (109). LILRBs act as both immune checkpoint molecules and tumor sustaining factors but may not affect normal development. Thus, they have potential as attractive targets for cancer treatment.

# LILRB1

LILRB1, an immunoreceptor tyrosine-based inhibitory motif-containing receptor, is widely expressed on human immune cells, including B cells, monocytes and macrophages, dendritic cells and subsets of NK cells and T cells (110). The ligands of LILRB1, such as major histocompatibility complex (MHC) class I molecules, activate LILRB1 and transduce a suppressive signal, which inhibits immune responses (111). The MHC class I component  $\beta$ 2-microglobulin ( $\beta$ 2M) has been found to act as a potential antiphagocytic signal in cancer cells (110). MHC-I on the cancer cell surface is sensed by LILRB1 on TAMs, resulting in negative regulation of cancer cell phagocytosis. LILRB1 is an immunoglobulin-like receptor for the gene product of human cytomegalovirus UL18, a homolog of cellular MHC class I antigens (112). LILRB1 in NK cells inhibits Fc receptor-mediated signaling in monocytes by activating the inhibitory phosphatase SHP-1 (113). It has been reported that LILRB1 possibly plays an antitumor role in hepatocarcinoma cells by interacting with SHP1 (114). Ablation of MHC-I on cancer cells or LILRB1 on macrophages has been reported to promote the phagocytic clearance of tumor cells and inhibit tumor growth in an in vivo mouse model (110).

# LILRB2

LILRB2 is known as an immune inhibitory receptor that suppresses the immune system. CD8+CD28<sup>-</sup> alloantigenspecific T suppressor cells could trigger the expression of LILRB2 on monocytes and DCs, leading to the tolerance effect on these APCs (115). LILRB2<sup>high</sup> DCs could give rise to the anergy of CD4+CD45RO+CD25+ T cells and suppress their further differentiation into regulatory T cells (116). As recent studies have reported, LILRB2 is highly expressed on hematopoietic stem cells or leukemia stem cells and is critical for the maintenance of stemness supporting hematopoiesis or leukemogenesis (117). LILRB2 is also reported to be expressed in many types of



**Figure 7** Role of LILRB family. LILRB1 inhibits Fc receptor-mediated signaling in NK cells and monocytes and possibly plays an antitumor effect in hepatocarcinoma cells by activating SHP-1, which could then inhibit PI3K/Akt pathway. In endometrial cancer cells, LILRB2 could enhance the SHP2/CaMK1/CREB signaling pathways. LILRB4 supports tumor cell infiltration into tissues and suppresses T cell activity possibly via APOE, LILRB4, SHP-2, uPAR and ARG1 in AML cells. LILRB, leukocyte immunoglobulin-like receptor B; SHP, SH2-domain bearing protein tyrosine phosphatase; NK, natural killer; AML, acute myeloid leukemia.

solid cancers. For instance, LILRB2 is linked with fewer infiltrating lymphocytes and more lymphatic metastasis in breast cancer (117). LILRB2 can be detected on the cell membrane and in the intracellular region of NSCLC and enhances progression (118). LILRB2 plays a vital role in sustaining epithelial-mesenchymal transition (EMT) and the early metastatic behavior of pancreatic adenocarcinoma (119). Furthermore, LILRB2 is shown to be highly expressed in endometrial cancer, and the expression levels are conversely correlated with overall survival of patients (119). In vitro experiments indicate that knockdown of LILRB2 leads to a significant decrease in proliferation, colony formation and migration in several endometrial cancer cell lines. LILRB2 could enhance the SHP2/CaMK1/CREB signaling pathways, which supports the expansion and migration of endometrial cancer cells (119).

# LILRB4

LILRB4 (also known as CD85K, ILT3, LIR5, and HM18) has two extracellular Ig-like domains (D1 and D2) and three ITIMs. On the one hand, LILRB4 is a marker for

monocytic AML (120). Further demonstration is that LILRB4 is more highly expressed on monocytic AML cells than on their normal counterparts and that LILRB4 expression inversely correlates with overall survival of patients with AML (120). Experiments in mouse models and human cells show that LILRB4 supports tumor cell infiltration into tissues and suppresses T-cell activity via a signaling pathway that involves APOE, LILRB4, SHP-2, uPAR and ARG1 in AML cells (120). Furthermore, LILRB4 represents a compelling target for the treatment of monocytic AML. It has been reported that a LILRB4which targeted humanized mAb, blocks the LILRB4/APOE interaction in a competitive manner, inhibits monocytic AML cell tissue infiltration and reverses T cell suppression (121). On the other hand, LILRB4 is expressed on MDSCs and TAMs (122). Expression of LILRB4 on PMN-MDSCs and M-MDSCs in NSCLC is associated with poor patient outcomes (123). In vitro, the capacity of M-MDSCs from normal human monocytes to induce IL-10-producing Treg cells could be potentiated by prostaglandin E2 through LILRB4 on M-MDSCs (124). Anti-LILRB4 antibodies impaired the T cell suppression induced by M-MDSCs (122). Furthermore, fibronectin

expressed by stromal cells in the TME can bind and activate LILRB4 on MDSCs, which recruits SHP-1 to inhibit Syk-mediated FcyR signaling and the immunosuppressive activities of MDSCs (125). Apart from MDSCs, studies have indicated that LILRB4 is expressed on TAMs in various human cancers and mouse models, including melanoma, lung cancer, colon carcinoma and pancreatic carcinoma (108). Blocking LILRB4 or gp49b deficiency could increase the infiltration of antineoplastic immune cells into the TME and lessen the inhibitory effect of Treg cells by regulating IL-1b and iNOS production from TAMs (126). In summary, LILRB4 expressed on MDSCs and TAMs may be an interesting target for cancer immunotherapy.

# TREM2

TREM2 was originally recognized in Alzheimer's disease as a myeloid receptor that transmits intracellular signals to maintain microglial responses. TREM2 is also expressed on TAMs, acting as an activating receptor of the Ig superfamily and transmitting intracellular signals when binding to the adaptor DNAX-activating protein of 12 kDa (DAP12) (127). DAP12, which recruits the protein tyrosine kinase Syk, could conduct a tyrosine phosphorylation cascade activating downstream mediators such as PLCy2, PI-3K, mTOR and MAPK, finally triggering cell activation (Figure 8). Mouse models show that TREM2<sup>-/-</sup> mice are more resistant to the growth of diverse kinds of cancers than wild-type mice and are more responsive to anti-PD-1 immunotherapy (128). Moreover, treatment with an anti-TREM2 monoclonal antibody suppresses tumor growth and raises regression when combined with anti-PD-1 (128). Moreover, in colorectal carcinoma (CRC) and triplenegative breast cancer, TREM2 expression is reported to have a converse correlation with higher overall survival and relapse-free survival (128,129).

# NRP1

NRP1 was originally distinguished as an adhesion molecule in the frog nervous system and found as a transmembrane glycoprotein located on axons of nerve fibers (130). NRP1 located on the cell surface acts as a nontyrosine kinase transmembrane glycoprotein and plays a role as a coreceptor of secreted Semaphorin-3A (Sema-3A) (*Figure 9*). The *NRP1* gene is widely expressed in various kinds of cells, tissues and organs, such as endothelial cells, and the heart, liver, lung, kidney, pancreas, and skeletal Qian et al. Myeloid checkpoints for cancer immunotherapy



**Figure 8** Role of TREM2. TREM2 expressed on TAMs could transmit intracellular signals when binding to DAP12, which recruits the protein tyrosine kinase Syk, conducting a tyrosine phosphorylation cascade activating downstream mediators such as PLC $\gamma$ 2, PI-3K, mTOR and MAPK, and finally triggers cell activation. TREM2, triggering receptor expressed on myeloidcells 2; TAM, tumor-associated macrophage; MAPK, mitogenactivated protein kinases.

muscle (131), playing an essential role in promoting angiogenesis, neural development, cvtoskeleton remodelling, inflammation, the initial immune response, and tumor development (132,133). The expression of NRP1 in the immune system is more restricted and regulated. NRP1, which is also known as blood DC antigen 4 (BDCA4, or CD304), was identified as a human DC marker expressed in all pDCs (134). Other types of APCs, including monocytes and macrophages, also express NPR1 (135). NRP1 expressed in monocytes and macrophages generally plays proangiogenic and anti-inflammatory roles, contributing to tissue remodelling and wound healing (136). As recently reported, NPR1 is also expressed in CD8+ T cells in mice and humans and is mainly detected in intratumoral CD8+ T cells marked by high expression of PD-1 (137).

A recent study indicated the role of NRP1 in the emergence and development of human malignant tumors (138). Higher expression or mutations of NRP1 are linked with the initiation, progression, and prognosis of human malignant tumors such as hepatocellular carcinoma (139), gastric cancer (140,141), breast cancer (142), prostatic cancer, and pancreatic cancer (143). Overexpression of NRP1 significantly reduces the survival rate in NSCLC



Proliferation, Apoptosis, Cell Cycle

**Figure 9** Role of NRP1. NRP1 located on the cell surface acts as a non-tyrosine kinase transmembrane glycoprotein and plays a role as a co-receptor of secreted Semaphorin-3A (Sema-3A). *NRP1* gene widely expresses over various kinds of cells, tissues and organs such as endothelial cells, the heart, liver, lung, kidney, pancreas, and skeletal muscle, playing an essential part in promoting angiogenesis, neural development, cytoskeleton remodeling, inflammation, initial immune response, and tumors development. VEGF, vascular endothelial growth factor.

patients. The reduction of NRP1 expression by utilizing NRP1 monoclonal antibody, RNA interference or NRP1 inhibitor could inhibit tumor cell growth and tumor angiogenesis (144). Overexpression of NRP1 in human oral squamous cell carcinoma shows more spindle filaments, reduces the expression of EMT epithelial markers and increases the expression of mesenchymal markers (145). As studies report, NRP1 promotes the presence of EMT in breast cancer cells by augmenting signaling molecules such as TGF- $\beta$  (146).

# Clever-1

Clever-1, also known as stabilin-1 or FEEL-1, is a multifunctional glycoprotein expressed on a subset of antiinflammatory macrophages involved in scavenging, angiogenesis and cell adhesion, which are involved in receptor-mediated endocytosis and recycling, intracellular sorting and transcytosis (147,148). Additionally, it is the first adhesion molecule implicated in transmigration through the lymphatic arm of the immune system (147). Attenuated progression of melanoma tumor growth and metastasis has been observed in Clever-1-knockout mice and those treated with anti-Clever-1 therapy (149). Clever-1<sup>+</sup> macrophages are found in human cancers and are associated with poorer disease-free survival in colorectal cancers of advanced stage and overall survival in bladder Circulating monocytes (149). and cancer tissue macrophages show higher expression of Clever-1 in an immunosuppressive environment, such as pregnancy and cancer, which could suppress Th1 lymphocyte activation (148). Meanwhile, according to a phase 1/2 clinical trial, bexmarilimab, a novel anti-Clever-1 antibody, has shown significant efficacy in 10 patients with hard-to-treat solid tumors (150).

# **Clinical application**

While myeloid checkpoints have been increasingly recognized as a potentially hopeful target to develop novel immunotherapeutic strategies, several monoclonal antibodies (mAbs) targeting myeloid checkpoints have recently been under development, and clinical trials are ongoing. Here, we labelled the recent advanced antibodies targeting those myeloid checkpoints below (*Table 1*).

#### **Summary and prospects**

In this review, we briefly summarized the biological functions of different kinds of myeloid checkpoints expressed on various myeloid cells, which has greatly advanced cancer treatment. To overcome the immune escape mechanisms of tumors and to improve the versatility and raise the efficiency of current immunotherapies, it is necessary to understand myeloid checkpoints in more details and explore novel approaches based on these kinds of checkpoints.

#### **Acknowledgements**

This study was supported by the Imported Scholar Project and Startup from Peking University Health Science Center (BMU2021YJ063 to MD); the Biotechnology Innovation Plan from Beijing Sungen Biomedical Technology Co., Ltd (2022066 to MD); and the Excellent Young Scientists Fund Program (overseas) from National Natural Science Fund (HY2021-7 to MD).

Table 1 C	urrent clinical trials target	ting myeloid checkpoints.				
Target	Agent	Interventions	Company	Study phase/status	Clinical trial ID	Conditions or diseases
CD47/ SIRPα	600XH	Anti-CD47/PD-1 bifunctional antibody	Waterstone Hanxbio Pty Ltd.	Phase 2/Recruiting	NCT04886271	Advanced solid tumor
	IBI188, Rituximab	Anti-CD47 monoclonal antibody	Innovent Biologics (Suzhou) Co. Ltd.	Phase 1/Active	NCT03717103	Advanced malignancies
	IBI188	Anti-CD47 monoclonal antibody	Innovent Biologics (Suzhou) Co. Ltd.	Phase 1/Completed	NCT03763149	Advanced malignancies
	Magrolimab	Anti-CD47 monoclonal antibody	University of California, San Francisco & Gilead Sciences	Phase 1/Not yet recruiting	NCT05169944	Children and adults with recurrent or progressive malignant brain tumors
	STI-6643	1	Sorrento Therapeutics, Inc.	Phase 1/Recruiting	NCT04900519	Advanced solid tumors
	CC-90002 Rituximab	1	Celgene	Phase 1/Completed	NCT02367196	Advanced solid and hematologic cancers
	ТТІ-621	Soluble recombinant fusion protein	Trillium Therapeutics Inc.	Phase 1/Recruiting	NCT02663518	Hematologic malignancies and selected solid tumors
	Hu5F9-G4	Anti-CD47 monoclonal antibody	Gilead Sciences	Phase 1/Phase 1	NCT02678338	Hematological malignancies
	Hu5F9-G4	I	Gilead Sciences	Phase 1/Phase 1	NCT02216409	Solid tumors
	PF-07257876	CD47-PD-L1 bispecific antibody	Pfizer	Phase 1/Recruiting	NCT04881045	Selected advanced tumors
	Magrolimab	Monotherapy or in combination with Azacitidine	Gilead Sciences	Phase 1/Recruiting	NCT03248479	Hematological malignancies
	SRF231	1	Surface Oncology	Phase 1/Completed	NCT03512340	Advanced solid and hematologic cancers
	AK117	Anti-CD47	Akeso	Phase 1/Recruiting	NCT04728334	Malignant neoplasms
	TQB2928	New molecular entity	Chia Tai Tianqing Pharmaceutical Group Co., Ltd.	Phase 1/Not yet recruiting	NCT04854681	Advanced solid tumors and hematological malignancies
	IMC-002	Blocking the interaction between CD47 and SIRP $\alpha$	ImmuneOncia Therapeutics Inc.	Phase 1/Recruiting	NCT04306224	Metastatic or locally advanced solid tumors and relapsed or refractory lymphomas
	IBI322	Recombinant anti-human CD47/PD-L1 bispecific antibody	Innovent Biologics (Suzhou) Co. Ltd.	Phase 1/Recruiting	NCT04328831	Advanced malignancies
	IMM2902	HER2/SIRP $\alpha$ bispecific mAb-trap antibody-receptor fusion protein	ImmuneOnco Biopharmaceuticals (Shanghai) Co., Ltd.	Phase 1/Not yet recruiting	NCT05076591	HER2-expressing advanced solid tumors
Table 1 ( $c$	ontinued)					

472

© Chinese Journal of Cancer Research. All rights reserved.

www.cjcrcn.org

Qian et al. Myeloid checkpoints for cancer immunotherapy

Table 1 ( $c$	ontinued)					
Target	Agent	Interventions	Company	Study phase/status	Clinical trial ID	Conditions or diseases
	AO-176	Humanized monoclonal antibody (mAb) targeting CD47	Arch Oncology	Phase 1/2/Recruiting	NCT03834948	Solid tumor malignancies
	IBC0966	I	SUNHO (China) BioPharmaceutical CO., Ltd.	Phase 1/2/Not yet recruiting	NCT04980690	Advanced malignant tumors
	IBI322	Recombinant anti-human CD47/PD-L1 bispecific antibody	Innovent Biologics (Suzhou) Co. Ltd.	Phase 1/Not yet recruiting	NCT04338659	Advanced malignancies
	IBI322	IBI322 monotherapy or combination therapy	Innovent Biologics (Suzhou) Co. Ltd.	Phase 1/Not yet recruiting	NCT04912466	Advanced malignant tumors
	IBI322+HMA	IBI322 monotherapy or combination therapy with HMA	Innovent Biologics (Suzhou) Co. Ltd.	Phase la/lb/Not yet recruiting	NCT05148442	Myeloid tumors
	ТП-621	Soluble recombinant fusion protein created by directly linking the sequences encoding the N-terminal CD47 binding domain of human SIRP $\alpha$ with the Fc domain of human immunoglobulin (IgG1)	Trillium Therapeutics Inc.	Phase 1/Terminated	NCT02890368	Relapsed and refractory solid tumors and mycosis fungoides
	ТП-621+ПІ-622	TTI-621 and TTI-622 in combination with daratumumab hyaluronidase-fihj	Memorial Sloan Kettering Cancer Center	Phase 1/Recruiting	NCT05139225	Multiple myeloma
	AK117	Anti-CD47 antibody	Akeso	Phase 1/2/Recruiting	NCT04980885	Acute myeloid leukemia
	AK117	Anti-CD47 antibody	Akeso	Phase 1/2/Recruiting	NCT04900350	Myelodysplastic syndrome
	ALX148 (Evorpacept)	1	ALX Oncology Inc.	Phase 1/Active, not recruiting	NCT03013218	Advanced solid tumors and lymphoma
	ALX148	Combination with trastuzumab, ramucirumab, and paclitaxel	ALX Oncology Inc.	Phase 2/3/Not yet recruiting	NCT05002127	HER2+ gastric cancer gastric cancer
	ALX148	Evaluating the combination of ALX148, cetuximab, and pembrolizumab	Criterium, Inc.	Phase 2/Not yet recruiting	NCT05167409	Microsatellite stable metastatic colorectal cancer
Table 1 ( $c$	ontinued)					

473

Table 1 ( $\epsilon$	continued)					
Target	Agent	Interventions	Company	Study phase/status	Clinical trial ID	Conditions or diseases
	AO-176	AO-176 as monotherapy and in combination with bortezomib/dexamethas one	Arch Oncology	Phase 1/2/Recruiting	NCT04445701	Relapsed/refractory multiple myeloma
	CC-90002	Monoclonal Ab to CD47	Celgene	Terminated	NCT02641002	AML and high-risk MDS
	CPO107/ JMT601	CD20-CD47 bispecific fusion protein	Conjupro Biotherapeutics, Inc.	Phase 1/2/Not yet recruiting	NCT04853329	CD20 positive non-Hodgkin lymphoma
	TG-1801	Bispecific CD47 and CD19 antibody	TG Therapeutics, Inc.	Phase 1/Recruiting	NCT04806035	B-cell lymphoma or chronic lymphocytic leukemia
	Lemzoparlimab (TJC4)	$CD47$ -SIRP $\alpha$ antibody	AbbVie	Phase 1	NCT04912063	AML; MDS
	SL-172154	Fusion protein consisting of human SIRP $\alpha$ and CD40L (SIRP $\alpha$ -Fc-CD40L) linked via a human Fc	l Shattuck Labs, Inc.	Phase 1/Recruiting	NCT04502888	Squamous cell carcinoma of the head and neck or skin
	TJ1133	Anti-CD47 antibody	The First Affiliated Hospital of Bengbu Medical College	Phase 1/2/Recruiting	NCT05148533	Advanced solid tumors
SLAMF7	SLAMF7 BATS	Combining anti-CD3 x anti-SLAMF7 bispecific antibody	University of Virginia	Phase 1/Not yet recruiting	NCT04864522	Relapsed/Refractory multiple myeloma
	Anti-SLAMF7	Anti-SLAMF7 monoclonal antibody	University of Arkansas	Phase 2/Withdrawn (No study population. No subjects enrolled.)	NCT03168100	Multiple myeloma
	Anti-SLAMF7	Anti-SLAMF7 monoclonal antibody	University of Arkansas	Phase 2/Withdrawn (No study population. No subjects enrolled.)	NCT03000634	Multiple myeloma during maintenance therapy
	Elotuzumab	mAb directed against the extracellular domain of SLAMF7	<ul> <li>Tulane University School of Medicine</li> </ul>	Phase 2/Not yet recruiting	NCT05170789	Relapsed refractory multiple myeloma
	Elotuzumab	Elotuzumab in combination with carfilzomib, lenalidomide and dexamethasone (E- KRd)	Wuerzburg University Hospital	Phase 3/Active, not recruiting	NCT03948035	Newly diagnosed multiple myeloma
	Elotuzumab	I	University of Arkansas	Phase 2/Withdrawal of study support	NCT03003728	High-risk myeloma post- autologous stem cell transplant
	Elotuzumab	Lenalidomide and dexamethasone with or without elotuzumab	Bristol-Myers Squibb	Phase 3/Terminated (Insufficient enrollment)	NCT01891643	Newly diagnosed, previously untreated multiple myeloma
Table 1 ( $c$	continued)					

# Qian et al. Myeloid checkpoints for cancer immunotherapy

Table 1 (c	ontinued)					
Target	Agent	Interventions	Company	Study phase/status	Clinical trial ID	Conditions or diseases
	Biological: Elotuzumab	Expanded access to elotuzumab (empliciti) for multiple myeloma	Bristol-Myers Squibb	No longer available	NCT03126617	Multiple myeloma
	Biological: BCMA-CS1 cCAR T cells	I	iCell Gene Therapeutics	Early phase 1/Unknown	NCT04156269	Multiple myeloma
	Biological: Elotuzumab	Anti-SLAMF7 monoclonal antibody	M.D. Anderson Cancer Center	Phase 2/Recruiting	NCT04517851	JAK2-mutated myelofibrosis
LILRB1	BND-22	Ι	<b>Biond Biologics</b>	Phase 1/2/Recruiting	NCT04717375	Advanced solid tumors
	NGM707	LILRB1/LILRB2 bispecific antibody	NGM Biopharmaceuticals, Inc.	Phase 1/2/Recruiting	NCT04913337	Advanced or metastatic solid tumor malignancies
LILRB2	JTX 8064	As monotherapy or in combination with PD-1 inhibitor	Jounce Therapeutics, Inc.	Phase 1/Recruiting	NCT04669899	Advanced refractory solid tumors
	IO-108	As monotherapy or in combination with pembrolizumab	Immune-Onc Therapeutics Inc.	Phase 1/Recruiting	NCT05054348	Solid tumors
	MK-4830	1	Merck Sharp & Dohme Corp	Phase 1/Recruiting	NCT03564691	Advanced solid tumors
LILRB4	IO-202	Monotherapy or plus azacitidine	Immune-Onc Therapeutics Inc.	Phase 1/Recruiting	NCT04372433	AML and CMML
	IO-202	Monotherapy or in combination with pembrolizumab	Immune-Onc Therapeutics Inc	Phase 1/Not yet recruiting	NCT05309187	Solid cancer
	MK-0482	In combination with pembrolizumab	Merck Sharp & Dohme LLC	Phase 2	NCT04165096	Non-small-cell lung carcinoma, acute leukemia, myeloid, neoplasms
TREM2	PY314	PY314 or in combination with pembrolizumab	Pionyr Immunotherapeutics Inc.	Phase 1/Recruiting	NCT04691375	Advanced solid tumors
Siglec15	NC318	NC318 or in combination with pembrolizumab	Yale University New Haven, Connecticut, United States	Phase 2/Recruiting	NCT04699123	Advanced non-small cell lung cancer
NRP1	ASP1948	ASP1948 or in combination with nivolumab or pembrolizumab	Astellas Pharma Global Development, Inc.	Phase 1b	NCT03565445	Advanced solid tumors
Clever1	Bexmarilimab	Dose-escalation, six dose levels	The University of Texas Health Science Center at San Antonio San Antonio, Texas, United States	Phase I/II	NCT03733990	Advanced solid tumors
LILRB, leu al and vas	kocyte immunoglobulin-li cular endothelial receptor	ke receptor B; Siglec, siali -1; AML, acute myeloid leu	c acid-binding immunoglo ikemia; MDS, myelodysplo	obulin-type lectin; NRP1, r astic syndrome; CMML, ch	neuropilin-1; Cleve1, nronic myelomonocy	, common lymphatic endotheli- rtic leukemia.

475

# Qian et al. Myeloid checkpoints for cancer immunotherapy

# 476

# Footnote

*Conflicts of Interest*: The authors have no conflicts of interest to declare. Figures are created with BioRender software (https://biorender.com/).

# References

- 1. O'Donnell JS, Teng MWL, Smyth MJ. Cancer immunoediting and resistance to T cell-based immunotherapy. Nat Rev Clin Oncol 2019;16: 151-67.
- 2. Brahmer JR, Tykodi SS, Chow LQ, et al. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. N Engl J Med 2012;366:2455-65.
- 3. Manguso RT, Pope HW, Zimmer MD, et al. In vivo CRISPR screening identifies Ptpn2 as a cancer immunotherapy target. Nature 2017;547:413-8.
- 4. Patel SJ, Sanjana NE, Kishton Rj, et al. Identification of essential genes for cancer immunotherapy. Nature 2017;548:537-42.
- Pan D, Kobayashi A, Jiang P, et al. A major chromatin regulator determines resistance of tumor cells to T cell-mediated killing. Science 2018; 359:770-5.
- Gabrilovich DI, Ostrand-Rosenberg S, Bronte V. Coordinated regulation of myeloid cells by tumours. Nat Rev Immunol 2012;12:253-68.
- Nakamura K, Smyth MJ. Myeloid immunosuppression and immune checkpoints in the tumor microenvironment. Cell Mol Immunol 2020;17:1-12.
- Jensen TO, Schmidt H, Møller HJ, et al. Intratumoral neutrophils and plasmacytoid dendritic cells indicate poor prognosis and are associated with pSTAT3 expression in AJCC stage I/II melanoma. Cancer 2012:118;2476-85.
- Goc J, Germain C, Vo-Bourgais TK, et al. Dendritic cells in tumor-associated tertiary lymphoid structures signal a Th1 cytotoxic immune contexture and license the positive prognostic value of infiltrating CD8+ T cells. Cancer Res 2014;74:705-15.
- 10. Huang SH, Waldron JN, Milosevic M, et al. Prognostic value of pretreatment circulating neutrophils, monocytes, and lymphocytes in oropharyngeal cancer stratified by human papillomavirus status. Cancer 2015:121;545-55.

- 11. Lavin Y, Winter D, Blecher-Gonen R, et al. Tissueresident macrophage enhancer landscapes are shaped by the local microenvironment. Cell 2014;159: 1312-26.
- Engblom C, Pfirschke C, Pittet MJ. The role of myeloid cells in cancer therapies. Nat Rev Cancer 2016;16:447-62.
- 13. Cheng S, Li Z, Gao R, et al. A pan-cancer single-cell transcriptional atlas of tumor infiltrating myeloid cells. Cell 2021;184:792-809.e23.
- Geissmann F, Manz MG, Jung S, et al. Development of monocytes, macrophages, and dendritic cells. Science 2010;327:656-61.
- Song Q, Hawkins GA, Wudel L, et al. Dissecting intratumoral myeloid cell plasticity by single cell RNA-seq. Cancer Med 2019:8;3072-85.
- Kiss M, Van Gassen S, Movahedi K, et al. Myeloid cell heterogeneity in cancer: not a single cell alike. Cell Immunol 2018:330;188-201.
- Park SY, Kim IS. Harnessing immune checkpoints in myeloid lineage cells for cancer immunotherapy. Cancer Lett 2019:452;51-8.
- Logtenberg MEW, Scheeren FA, Schumacher TN. The CD47-SIRPα immune checkpoint. Immunity 2020:52;742-52.
- Garg AD, Romano E, Rufo N, et al. Immunogenic versus tolerogenic phagocytosis during anticancer therapy: mechanisms and clinical translation. Cell Death Differ 2016:23;938-51.
- 20. Oldenborg PA, Zheleznyak A, Fang YF, et al. Role of CD47 as a marker of self on red blood cells. Science 2000;288:2051-4.
- 21. Majeti R, Chao MP, Alizadeh AA, et al. CD47 is an adverse prognostic factor and therapeutic antibody target on human acute myeloid leukemia stem cells. Cell 2009:138;286-99.
- 22. Barrera L, Montes-Servín E, Hernandez-Martinez JM. CD47 overexpression is associated with decreased neutrophil apoptosis/phagocytosis and poor prognosis in non-small-cell lung cancer patients. Br J Cancer 2017;117:385-97.
- 23. Li Y, Lu S, Xu Y, et al. Overexpression of CD47 predicts poor prognosis and promotes cancer cell invasion in high-grade serous ovarian carcinoma. Am J Transl Res 2017;9:2901-10.
- 24. Guo H, Li W, Qian L, et al. Clinical challenges in

477

neoadjuvant immunotherapy for non-small cell lung cancer. Chin J Cancer Res 2021;33:203-15.

- Ring NG, Herndler-Brandstetter D, Weiskopf K, et al. Anti-SIRPα antibody immunotherapy enhances neutrophil and macrophage antitumor activity. Proc Natl Acad Sci U S A 2017;114:E10578-85.
- Advani R, Flinn I, Popplewell L, et al. CD47 blockade by Hu5F9-G4 and rituximab in non-Hodgkin's lymphoma. N Engl J Med 2018; 379:1711-21.
- Sockolosky JT, Dougan M, Ingram JR, et al. Durable antitumor responses to CD47 blockade require adaptive immune stimulation. Proc Natl Acad Sci U S A 2016;113:E2646-54.
- Lin N, Song Y, Zhu J. Immune checkpoint inhibitors in malignant lymphoma: Advances and perspectives. Chin J Cancer Res 2020;32:303-18.
- Xu MM, Pu Y, Han D, et al. Dendritic cells but not macrophages sense tumor mitochondrial DNA for cross-priming through signal regulatory protein α signaling. Immunity 2017;47:363-73.e5.
- 30. Gordon SR, Maute RL, Dulken BW, et al. PD-1 expression by tumour-associated macrophages inhibits phagocytosis and tumour immunity. Nature 2017;545:495-9.
- 31. O'Connell P, Hyslop S, Blake MK, et al. SLAMF7 signaling reprograms T cells toward exhaustion in the tumor microenvironment. J Immunol 2021;206:193-205.
- O'Connell P, Amalfitano A, Aldhamen YA. SLAM family receptor signaling in viral infections: HIV and beyond. Vaccines (Basel) 2019;7:184.
- 33. Chen J, Zhong MC, Guo H, et al. SLAMF7 is critical for phagocytosis of haematopoietic tumour cells via Mac-1 integrin. Nature 2017;544:493-7.
- Cannons JL, Tangye SG, Schwartzberg PL. SLAM family receptors and SAP adaptors in immunity. Annu Rev Immunol 2011;29:665-705.
- O'Connell P, Pepelyayeva Y, Blake MK, et al. SLAMF7 is a critical negative regulator of IFN-αmediated CXCL10 production in chronic HIV infection. J Immunol 2019;202:228-38.
- 36. Guo H, Cruz-Munoz ME, Wu N, et al. Immune cell inhibition by SLAMF7 is mediated by a mechanism requiring src kinases, CD45, and SHIP-1 that is defective in multiple myeloma cells. Mol Cell Biol

2015;35:41-51.

- Miller BC, Sen DR, Al Abosy R, et al. Subsets of exhausted CD8<sup>+</sup> T cells differentially mediate tumor control and respond to checkpoint blockade. Nat Immunol 2019;20:326-36.
- Sade-Feldman M, Yizhak K, Bjorgaard SL, et al. Defining T cell states associated with response to checkpoint immunotherapy in melanoma. Cell 2018;175:998-1013.e20.
- Kurtulus S, Madi A, Escobar G, et al. Checkpoint blockade immunotherapy induces dynamic changes in PD-1<sup>-</sup>CD8<sup>+</sup> tumor-infiltrating T cells. Immunity 2019;50:181-94.e6.
- 40. Johnson DE, O'Keefe RA, Grandis JR. Targeting the IL-6/JAK/STAT3 signalling axis in cancer. Nat Rev Clin Oncol 2018;15:234-48.
- 41. ffrench-Constant C, Colognato H. Integrins: versatile integrators of extracellular signals. Trends Cell Biol 2004;14:678-86.
- 42. Hamerman JA, Ni M, Killebrew JR, et al. The expanding roles of ITAM adapters FcRgamma and DAP12 in myeloid cells. Immunol Rev 2009;232: 42-58.
- Wu N, Veillette A. SLAM family receptors in normal immunity and immune pathologies. Curr Opin Immunol 2016;38:45-51.
- 44. Li D, Xiong W, Wang Y, et al. SLAMF3 and SLAMF4 are immune checkpoints that constrain macrophage phagocytosis of hematopoietic tumors. Science Immunol 2022;7:eabj5501.
- 45. Ferris RL, Blumenschein G Jr, Fayette J, et al. Nivolumab for recurrent squamous-cell carcinoma of the head and neck. N Engl J Med 2016;375:1856-67.
- Limagne E, Richard C, Thibaudin M, et al. Tim-3/galectin-9 pathway and mMDSC control primary and secondary resistances to PD-1 blockade in lung cancer patients. Oncoimmunology 2019;8:e1564505.
- Highfill SL, Cui Y, Giles AJ, et al. Disruption of CXCR2-mediated MDSC tumor trafficking enhances anti-PD1 efficacy. Sci Transl Med 2014;6: 237ra267.
- 48. De Henau O, Rausch M, Winkler D, et al. Overcoming resistance to checkpoint blockade therapy by targeting PI3K $\gamma$  in myeloid cells. Nature 2016;539:443-7.
- 49. Zhu Y, Knolhoff BL, Meyer MA, et al.

CSF1/CSF1R blockade reprograms tumorinfiltrating macrophages and improves response to T-cell checkpoint immunotherapy in pancreatic cancer models. Cancer Res 2014:74;5057-69.

- 50. Lipson EJ, Forde PM, Hammers HJ, et al. Antagonists of PD-1 and PD-L1 in cancer treatment. Semin Oncol 2015;42:587-600.
- 51. Topalian SL, Hodi FS, Brahmer JR, et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. N Engl J Med 2012;366:2443-54.
- 52. Alsaab HO, Sau S, Alzhrani R, et al. PD-1 and PD-L1 Checkpoint Signaling Inhibition for Cancer Immunotherapy: Mechanism, combinations, and clinical outcome. Front Pharmacol 2017;8:561.
- 53. Mi Y, Han J, Zhu J, et al. Role of the PD-1/PD-L1 signaling in multiple sclerosis and experimental autoimmune encephalomyelitis: recent insights and future directions. Mol Neurobiol 2021;58:6249-71.
- 54. He G, Zhang H, Zhou J, et al. Peritumoural neutrophils negatively regulate adaptive immunity via the PD-L1/PD-1 signalling pathway in hepatocellular carcinoma. J Exp Clin Cancer Res 2015;34:141.
- 55. Li J, Lee Y, Li Y, et al. Co-inhibitory molecule B7 superfamily member 1 expressed by tumorinfiltrating myeloid cells induces dysfunction of antitumor CD8<sup>+</sup> T cells. Immunity 2018;48:773-86.e5.
- Macauley MS, Crocker PR, Paulson JC. Siglecmediated regulation of immune cell function in disease. Nat Rev Immunol 2014;14:653-66.
- 57. Wu C, Rauch U, Korpos E, et al. Sialoadhesinpositive macrophages bind regulatory T cells, negatively controlling their expansion and autoimmune disease progression. J Immunol 2009;182:6508-16.
- Crocker PR, Paulson JC, Varki A. Siglecs and their roles in the immune system. Nat Rev Immunol 2007;7:255-66.
- 59. Varki NM, Strobert E, Dick EJ Jr, et al. Biomedical differences between human and nonhuman hominids: potential roles for uniquely human aspects of sialic acid biology. Annu Rev Pathol 2011;6: 365-93.
- 60. O'Neill AS, van den Berg TK, Mullen GE. Sialoadhesin - a macrophage-restricted marker of immunoregulation and inflammation. Immunology

2013:138;198-207.

- 61. Duan S, Paulson JC. Siglecs as immune cell checkpoints in disease. Annu Rev Immunol 2020;38:365-95.
- 62. Nycholat CM, Rademacher C, Kawasaki N, et al. In silico-aided design of a glycan ligand of sialoadhesin for *in vivo* targeting of macrophages. J Am Chem Soc 2012;134:15696-9.
- Oetke C, Vinson MC, Jones C, et al. Sialoadhesindeficient mice exhibit subtle changes in B- and Tcell populations and reduced immunoglobulin M levels. Mol Cell Biol 2006;26:1549-57.
- 64. Gonzalez-Gil A, Schnaar RL. Siglec ligands. Cells 2021;10:1260.
- 65. Fraschilla I, Pillai S. Viewing Siglecs through the lens of tumor immunology. Immunol Rev 2017;276:178-91.
- 66. Edgar LJ, Kawasaki N, Nycholat CM. Targeted delivery of antigen to activated CD169<sup>+</sup> macrophages induces bias for expansion of CD8<sup>+</sup> T cells. Cell Chem Biol 2019;26:131-6.e4.
- Ravishankar B, Shinde R, Liu H, et al. Marginal zone CD169<sup>+</sup> macrophages coordinate apoptotic cell-driven cellular recruitment and tolerance. Proc Natl Acad Sci U S A 2014;111:4215-20.
- Shiota T, Miyasato Y, Ohnishi K, et al. The clinical significance of CD169-positive lymph node macrophage in patients with breast cancer. PLoS One 2016;11:e0166680.
- 69. Ohnishi K, Komohara Y, Saito Y, et al. CD169positive macrophages in regional lymph nodes are associated with a favorable prognosis in patients with colorectal carcinoma. Cancer Sci 2013;104:1237-44.
- Ohnishi K, Yamaguchi M, Erdenebaatar C, et al. Prognostic significance of CD169-positive lymph node sinus macrophages in patients with endometrial carcinoma. Cancer Sci 2016;107:846-52.
- 71. van Dinther D, Lopez Venegas M, Veninga H, et al. Activation of CD8<sup>+</sup> T cell responses after melanoma antigen targeting to CD169<sup>+</sup> antigen presenting cells in mice and humans. Cancers (Basel) 2019;11:183.
- 72. Zheng Q, Hou J, Zhou Y, et al. Siglec1 suppresses antiviral innate immune response by inducing TBK1 degradation via the ubiquitin ligase TRIM27. Cell Res 2015;25:1121-36.
- 73. Prenzler S, Rudrawar S, Waespy M, et al. The role

of sialic acid-binding immunoglobulin-like-lectin-1 (siglec-1) in immunology and infectious disease. Int Rev Immunol 2021;1-26. [Epub ahead of print]

- 74. Zhang JQ, Nicoll G, Jones C, et al. Siglec-9, a novel sialic acid binding member of the immunoglobulin superfamily expressed broadly on human blood leukocytes. J Biol Chem 2000;275:22121-6.
- 75. Yu H, Gonzalez-Gil A, Wei Y, et al. Siglec-8 and Siglec-9 binding specificities and endogenous airway ligand distributions and properties. Glycobiology 2017;27:657-68.
- Miyazaki K, Sakuma K, Kawamura YI, et al. Colonic epithelial cells express specific ligands for mucosal macrophage immunosuppressive receptors siglec-7 and -9. J Immunol 2012;188:4690-700.
- 77. Hudak JE, Canham SM, Bertozzi CR, et al. Glycocalyx engineering reveals a Siglec-based mechanism for NK cell immunoevasion. Nat Chem Biol 2014;10:69-75.
- Avril T, Floyd H, Lopez F, et al. The membraneproximal immunoreceptor tyrosine-based inhibitory motif is critical for the inhibitory signaling mediated by Siglecs-7 and -9, CD33-related Siglecs expressed on human monocytes and NK cells. J Immunol 2004;173:6841-9.
- 79. Ghasemi Z, Dinarvand R, Mottaghitalab F, et al. Aptamer decorated hyaluronan/chitosan nanoparticles for targeted delivery of 5-fluorouracil to MUC1 overexpressing adenocarcinomas. Carbohydr Polym 2015;121:190-8.
- Tanida S, Akita K, Ishida A, et al. Binding of the sialic acid-binding lectin, Siglec-9, to the membrane mucin, MUC1, induces recruitment of beta-catenin and subsequent cell growth. J Biol Chem 2013; 288:31842-52.
- 81. Beatson R, Tajadura-Ortega V, Achkova D, et al. The mucin MUC1 modulates the tumor immunological microenvironment through engagement of the lectin Siglec-9. Nat Immunol 2016;17:1273-81.
- Belisle JA, Horibata S, Jennifer GA, et al. Identification of Siglec-9 as the receptor for MUC16 on human NK cells, B cells, and monocytes. Mol Cancer 2010;9:118.
- 83. Adams OJ, Stanczak MA, von Gunten S, et al. Targeting sialic acid-Siglec interactions to reverse

immune suppression in cancer. Glycobiology 2018;28:640-7.

- McMillan SJ, Sharma RS, McKenzie EJ, et al. Siglec-E is a negative regulator of acute pulmonary neutrophil inflammation and suppresses CD11bβ2integrin-dependent signaling. Blood 2013;121: 2084-94.
- 85. Läubli H, Pearce OM, Schwarz F, et al. Engagement of myelomonocytic Siglecs by tumor-associated ligands modulates the innate immune response to cancer. Proc Natl Acad Sci U S A 2014;111:14211-6.
- 86. Schwarz F, Pearce OM, Wang X, et al. Siglec receptors impact mammalian lifespan by modulating oxidative stress. Elife 2015;4:e06184.
- Chen GY, Brown NK, Zheng P, et al. Siglec-G/10 in self-nonself discrimination of innate and adaptive immunity. Glycobiology 2014;24:800-6.
- Chen W, Han C, Xie B, et al. Induction of Siglec-G by RNA viruses inhibits the innate immune response by promoting RIG-I degradation. Cell 2013;152: 467-78.
- Chen GY, Chen X, King S, et al. Amelioration of sepsis by inhibiting sialidase-mediated disruption of the CD24-SiglecG interaction. Nat Biotechnol 2011;29:428-35.
- Xu H, Pan Y, Zhang N. A breakthrough in liver regeneration for treatment of liver cancer. Cancer Biol Med 2021;18:631-4.
- 91. Abram CL, Lowell CA. Shp1 function in myeloid cells. J Leukoc Biol 2017;102:657-75.
- 92. Allen RL, Raine T, Haude A, et al. Leukocyte receptor complex-encoded immunomodulatory receptors show differing specificity for alternative HLA-B27 structures. J Immunol 2001:167;5543-7.
- 93. Barkal AA, Brewer RE, Markovic M, et al. CD24 signalling through macrophage Siglec-10 is a target for cancer immunotherapy. Nature 2019;572:392-6.
- 94. Kleinmanns K, Bischof K, Anandan S, et al. CD24targeted fluorescence imaging in patient-derived xenograft models of high-grade serous ovarian carcinoma. EBioMedicine 2020;56:102782.
- 95. Taurin S, Alkhalifa H. Breast cancers, mammary stem cells, and cancer stem cells, characteristics, and hypotheses. Neoplasia 2020;22:663-78.
- 96. Ayers M, Nebozhyn M, Cristescu R, et al. Molecular profiling of cohorts of tumor samples to guide

clinical development of pembrolizumab as monotherapy. Clin Cancer Res 2019;25:1564-73.

- 97. Wang J, Sun J, Liu LN, et al. Siglec-15 as an immune suppressor and potential target for normalization cancer immunotherapy. Nat Med 2019;25:656-66.
- Stuible M, Moraitis A, Fortin A, et al. Mechanism and function of monoclonal antibodies targeting siglec-15 for therapeutic inhibition of osteoclastic bone resorption. J Biol Chem 2014;289:6498-512.
- Hiruma Y, Hirai T, Tsuda E. Siglec-15, a member of the sialic acid-binding lectin, is a novel regulator for osteoclast differentiation. Biochem Biophys Res Commun 2011;409:424-9.
- 100. Sun J, Lu Q, Sanmamed MF, et al. Siglec-15 as an emerging target for next-generation cancer immunotherapy. Clin Cancer Res 2021;27:680-8.
- 101. Dong H, Strome SE, Salomao DR, et al. Tumorassociated B7-H1 promotes T-cell apoptosis: a potential mechanism of immune evasion. Nat Med 2002;8:793-800.
- 102. Zhang YH, He M, Wang Y, et al. Modulators of the balance between M1 and M2 macrophages during pregnancy. Front Immunol 2017;8:120.
- 103. Ishida-Kitagawa N, Tanaka K, Bao X, et al. Siglec-15 protein regulates formation of functional osteoclasts in concert with DNAX-activating protein of 12 kDa (DAP12). J Biol Chem 2012;287: 17493-502.
- 104. Cao H, Lakner U, de Bono B, et al. SIGLEC16 encodes a DAP12-associated receptor expressed in macrophages that evolved from its inhibitory counterpart SIGLEC11 and has functional and nonfunctional alleles in humans. Eur J Immunol 2008;38:2303-15.
- 105. Ali SR, Fong JJ, Carlin AF, et al. Siglec-5 and Siglec-14 are polymorphic paired receptors that modulate neutrophil and amnion signaling responses to group B Streptococcus. J Exp Med 2014;211:1231-42.
- 106. Samaridis J, Colonna M. Cloning of novel immunoglobulin superfamily receptors expressed on human myeloid and lymphoid cells: structural evidence for new stimulatory and inhibitory pathways. Eur J Immunol 1997;27:660-5.
- 107. Mori Y, Tsuji S, Inui M, et al. Inhibitory immunoglobulin-like receptors LILRB and PIR-B

negatively regulate osteoclast development. J Immunol 2008;181:4742-51.

- 108. Yang T, Qian Y, Liang X, et al. LILRB4, an immune checkpoint on myeloid cells. Blood Sci 2022;4:49-56.
- 109. Ravetch JV, Lanier LL. Immune inhibitory receptors. Science 2000;290:84-9.
- 110. Barkal AA, Weiskopf K, Kao KS, et al. Engagement of MHC class I by the inhibitory receptor LILRB1 suppresses macrophages and is a target of cancer immunotherapy. Nat Immunol 2018;19:76-84.
- 111. Chen H, Chen Y, Deng M, et al. Antagonistic anti-LILRB1 monoclonal antibody regulates antitumor functions of natural killer cells. J Immunother Cancer 2020;8:e000515.
- 112. Cosman D, Fanger N, Borges L, et al. A novel immunoglobulin superfamily receptor for cellular and viral MHC class I molecules. Immunity 1997;7:273-82.
- 113. Fanger NA, Cosman D, Peterson L, et al. The MHC class I binding proteins LIR-1 and LIR-2 inhibit Fc receptor-mediated signaling in monocytes. Eur J Immunol 1998;28:3423-34.
- 114. Cheng J, Luan J, Chen P, et al. Immunosuppressive receptor LILRB1 acts as a potential regulator in hepatocellular carcinoma by integrating with SHP1. Cancer Biomark 2020;28:309-19.
- 115. Chang CC, Ciubotariu R, Manavalan JS, et al. Tolerization of dendritic cells by T(S) cells: the crucial role of inhibitory receptors ILT3 and ILT4. Nat Immunol 2002;3:237-43.
- 116. Stallone G, Pontrelli P, Infante B, et al. Rapamycin induces ILT3(high)ILT4(high) dendritic cells promoting a new immunoregulatory pathway. Kidney Int 2014;85:888-97.
- 117. Zheng J, Umikawa M, Cui C, et al. Inhibitory receptors bind ANGPTLs and support blood stem cells and leukaemia development. Nature 2012; 485:656-60.
- 118. Grosskopf S, Eckert C, Arkona C, et al. Selective inhibitors of the protein tyrosine phosphatase SHP2 block cellular motility and growth of cancer cells *in vitro* and *in vivo*. ChemMedChem 2015;10:815-26.
- Shao H, Ma L, Jin F, et al. Immune inhibitory receptor LILRB2 is critical for the endometrial cancer progression. Biochem Biophys Res Commun 2018;506:243-50.

- Deng M, Gui X, Kim J, et al. LILRB4 signalling in leukaemia cells mediates T cell suppression and tumour infiltration. Nature 2018;562:605-9.
- 121. Gui X, Deng M, Song H, et al. Disrupting LILRB4/APOE interaction by an efficacious humanized antibody reverses T-cell suppression and blocks AML development. Cancer Immunol Res 2019;7:1244-57.
- 122. Singh L, Muise ES, Bhattacharya A, et al. ILT3 (LILRB4) promotes the immunosuppressive function of tumor-educated human monocytic myeloid-derived suppressor cells. Mol Cancer Res 2021;19:702-16.
- 123. de Goeje PL, Bezemer K, Heuvers ME, et al. Immunoglobulin-like transcript 3 is expressed by myeloid-derived suppressor cells and correlates with survival in patients with non-small cell lung cancer. Oncoimmunology 2015;4:e1014242.
- 124. Tomić S, Joksimović B, Bekić M, et al. Prostaglanin-E2 potentiates the suppressive functions of human mononuclear myeloid-derived suppressor cells and increases their capacity to expand IL-10-producing regulatory T cell subsets. Front Immunol 2019; 10:475.
- 125. Paavola KJ, Roda JM, Lin VY, et al. The fibronectin-ILT3 interaction functions as a stromal checkpoint that suppresses myeloid cells. Cancer Immunol Res 2021;9:1283-97.
- 126. Sharma N, Atolagbe OT, Ge Z, et al. LILRB4 suppresses immunity in solid tumors and is a potential target for immunotherapy. J Exp Med 2021;218:e20201811.
- 127. Peng Q, Malhotra S, Torchia JA, et al. TREM2- and DAP12-dependent activation of PI3K requires DAP10 and is inhibited by SHIP1. Sci Signal 2010;3:ra38.
- 128. Molgora M, Esaulova E, Vermi W, et al. TREM2 modulation remodels the tumor myeloid landscape enhancing anti-PD-1 immunotherapy. Cell 2020;182:886-900.e17.
- 129. Mantovani A, Marchesi F, Malesci A, et al. Tumourassociated macrophages as treatment targets in oncology. Nat Rev Clin Oncol 2017;14:399-416.
- 130. Mercurio AM. VEGF/Neuropilin signaling in cancer stem cells. Int J Mol Sci 2019;20:490.
- 131. Wild JR, Staton CA, Chapple K, et al. Neuropilins:

expression and roles in the epithelium. Int J Exp Pathol 2012;93:81-103.

- 132. Roy S, Bag AK, Singh RK, et al. Multifaceted role of neuropilins in the immune system: Potential targets for immunotherapy. Front Immunol 2017;8:1228.
- Schellenburg S, Schulz A, Poitz DM, et al. Role of neuropilin-2 in the immune system. Mol Immunol 2017:90;239-44.
- 134. Dzionek A, Fuchs A, Schmidt P, et al. BDCA-2, BDCA-3, and BDCA-4: three markers for distinct subsets of dendritic cells in human peripheral blood. J Immunol 2000;165:6037-46.
- 135. Wallerius M, Wallmann T, Bartish M, et al. Guidance molecule SEMA3A restricts tumor growth by differentially regulating the proliferation of tumor-associated macrophages. Cancer Res 2016; 76:3166-78.
- 136. DeFalco T, Bhattacharya I, Williams AV, et al. Yolk-sac-derived macrophages regulate fetal testis vascularization and morphogenesis. Proc Natl Acad Sci U S A 2014;111:E2384-93.
- Leclerc M, Voilin E, Gros G, et al. Regulation of antitumour CD8 T-cell immunity and checkpoint blockade immunotherapy by Neuropilin-1. Nat Commun 2019:10;3345.
- Torres-Salido MT, Sanchis M, Solé C, et al. Urinary neuropilin-1: A predictive biomarker for renal outcome in lupus nephritis. Int J Mol Sci 2019; 20:4610.
- 139. Bergé M, Allanic D, Bonnin P, et al. Neuropilin-1 is upregulated in hepatocellular carcinoma and contributes to tumour growth and vascular remodelling. J Hepatol 2011;55:866-75.
- 140. Peng Y, Liu YM, Li LC, et al. MicroRNA-338 inhibits growth, invasion and metastasis of gastric cancer by targeting NRP1 expression. PLoS One 2014;9:e94422.
- 141. Zhang Z, Xie T, Zhang X, et al. Immune checkpoint inhibitors for treatment of advanced gastric or gastroesophageal junction cancer: Current evidence and future perspectives. Chin J Cancer Res 2020; 32:287-302.
- 142. Naik A, Al-Zeheimi N, Bakheit CS, et al. Neuropilin-1 associated molecules in the blood distinguish poor prognosis breast cancer: A crosssectional study. Sci Rep 2017;7:3301.

#### Qian et al. Myeloid checkpoints for cancer immunotherapy

- 143. Tse BWC, Volpert M, Ratther E, et al. Neuropilin-1 is upregulated in the adaptive response of prostate tumors to androgen-targeted therapies and is prognostic of metastatic progression and patient mortality. Oncogene 2017;36:3417-27.
- 144. Dong JC, Gao H, Zuo SY, et al. Neuropilin 1 expression correlates with the radio-resistance of human non-small-cell lung cancer cells. J Cell Mol Med 2015;19:2286-95.
- 145. Chu W, Song X, Yang X, et al. Neuropilin-1 promotes epithelial-to-mesenchymal transition by stimulating nuclear factor-kappa B and is associated with poor prognosis in human oral squamous cell carcinoma. PLoS One 2014;9:e101931.
- 146. Pires BR, Mencalha AL, Ferreira GM, et al. NFkappaB is involved in the regulation of EMT genes in breast cancer cells. PLoS One 2017;12:e0169622.

**Cite this article as:** Qian Y, Yang T, Liang H, Deng M. Myeloid checkpoints for cancer immunotherapy. Chin J Cancer Res 2022;34(5):460-482. doi: 10.21147/j.issn.1000-9604.2022.05.07

- 147. Salmi M, Koskinen K, Henttinen T, et al. CLEVER-1 mediates lymphocyte transmigration through vascular and lymphatic endothelium. Blood;104:3849-57.
- 148. Tadayon S, Dunkel J, Takeda A, et al. Lymphatic endothelial cell activation and dendritic cell transmigration is modified by genetic deletion of Clever-1. Front Immunol 2021;12:602122.
- 149. Karikoski M, Marttila-Ichihara F, Elima K, et al. Clever-1/stabilin-1 controls cancer growth and metastasis. Clin Cancer Res 2014;20:6452-64.
- 150. Virtakoivu R, Rannikko JH, Viitala M, et al. Systemic blockade of Clever-1 elicits lymphocyte activation alongside checkpoint molecule downregulation in patients with solid tumors: Results from a Phase I/II clinical trial. Clin Cancer Res 2021;27:4205-20.