

## Research



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# Greater variability in rhesus macaque (*Macaca mulatta*) endocranial volume among males than females

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The greater male variability (GMV) hypothesis proposes that traits are more variable among males than females, and is supported by numerous empirical studies. Interestingly, GMV is also observed for human brain size and internal brain structure, a pattern which may have implications for sex-biased neurological and psychiatric conditions. A better understanding of neuroanatomical variability in non-human primates may illuminate whether certain species are appropriate models for these conditions. Here, we tested for sex differences in the variability of endocranial volume (ECV, a proxy for brain size) in a sample of 542 rhesus macaques (*Macaca mulatta*) from a large pedigreed free-ranging population. We also examined the components of phenotypic variance (additive genetic and residual variance) to tease apart the potential drivers of sex differences in variability. Our results suggest that males exhibit more variable ECVs, and that this pattern reflects either balancing/disruptive selection on male behaviour (associated with alternative male mating strategies) or sex chromosome effects (associated with mosaic patterns of X chromosome gene expression in females), rather than extended neurodevelopment among males. This represents evidence of GMV for brain size in a non-human primate species and highlights the potential of rhesus macaques as a model for sex-biased brain-based disorders.

## 1. Introduction

The ‘greater male variability (GMV) hypothesis’ posits that males tend to exhibit more physical and behavioural variability than females. This pattern has been observed in numerous mammalian species across many morphological traits, and appears to be particularly apparent in sexually selected traits [1–7]. The phenomenon is likely to reflect some combination of evolutionary and developmental mechanisms that produce and maintain greater inter-individual variability among males, including: (i) balancing or disruptive selection, (ii) sex differences in developmental schedules and (iii) sex chromosome effects.

While disruptive selection favours divergent (i.e. extreme) phenotypes, processes of balancing selection maintain phenotypic diversity within populations through various mechanisms. For instance, under negative frequency-dependent selection (one form of balancing selection), the fitness of a phenotype decreases as it becomes more common, which can lead to cyclical shifts in the frequency of different phenotypes over time, thereby preventing one phenotype from reaching fixation. When these mechanisms act on a trait within one sex only, this can lead to the exhibition of more variable phenotypes within that sex. Placental mammals tend to exhibit sex differences in reproductive potential, since female reproduction is most directly linked to longevity while male reproduction is primarily driven by fecundity [8–10]. Consequently, males often show greater reproductive

variance between individuals [8] and are more likely to exhibit alternative reproductive strategies that are subject to balancing/disruptive selection [11]. Conversely, females may be more likely to undergo stabilizing selection in traits directly or indirectly related to reproduction, thereby reducing trait variability in females relative to males [11]. Furthermore, additional mechanisms of sexual selection (specifically, direct male–male contest competition) often drive the evolution of larger adult body sizes among male mammals. In these cases, males experience longer developmental periods that may leave them more susceptible to environmental effects on trait expression, which may produce larger trait variability in males. Finally, male mammals may be expected to exhibit more variability for traits influenced by genes on the X chromosome, since males can only express their single set of X-linked alleles, while females may express either of their two alternative sets (due to mosaic X chromosome inactivation (XCI) across cells) or both sets (for genes that escape XCI within cells). This may facilitate mosaic levels of trait expression and produce lower phenotypic variability in females [5,12].

The GMV hypothesis has important implications for our understanding of human variation, since numerous studies have demonstrated that self-identified males are more variable than self-identified females across several physical (including neuroanatomical), behavioural, cognitive and personality-related traits [13–26]. Studies demonstrating GMV in brain size and structure are almost exclusively focused on humans [20–26]. Attempts to identify this pattern among non-human primate brains are particularly rare, and are thus far limited to examinations of chimpanzee sulcal morphology [27] and strepsirrhine skull length [28], with the latter analysis limited by very low sample sizes. It therefore remains uncertain whether GMV in human neuroanatomy represents an unusual characteristic for the primate order, or whether it may represent a more widespread pattern among primates.

Rhesus macaques are of particular interest in this regard, as they are popular models for studying neurological and psychiatric conditions that may be linked to sex differences in human neuroanatomical variability, including male-biased conditions like autism spectrum disorder (ASD) [29] and schizophrenia [30]. Specifically, individuals with ASD or schizophrenia exhibit higher brain structure variability than control cases [31,32], suggesting a possible link between GMV and vulnerability for these conditions. Additionally, ASD aetiology may be more heterogeneous in males compared with females [31] and male-specific increases in brain gene expression variability throughout development have been linked to genetic risk for schizophrenia [33]. Whether rhesus macaques also exhibit greater male neuroanatomical variability is unknown.

To investigate potential sex differences in brain size variability (Aim 1), we analysed endocranial volumes (ECV) in a large post-mortem sample of free-ranging rhesus macaques. Previous work has demonstrated that ECV is a reliable estimate of brain size across species (e.g. primates [34], birds [35,36]) and within species (e.g. budgerigars [35]) throughout development (e.g. domestic chickens [37], American alligators [37]); however, the proportion of non-neural tissue within the adult cranium may vary across individuals and increase with age [38] (including in macaque males [39] and females [40]). Nevertheless, unless this proportion varies with age and ECV in a sex-specific manner, a phenomenon not reported in previous literature, then our analysis of ECV variability is likely to reflect brain size variability.

We predicted that male rhesus macaques would exhibit greater variability for absolute ECV (Prediction 1a) and relative ECV (Prediction 1b), in accordance with the GMV hypothesis, and with previous reports of neuroanatomical GMV in humans [20–26] and chimpanzees [27]. Male rhesus macaques are subject to sex-specific selective pressures, including direct male–male contest competition, as demonstrated by intermediate body and canine size dimorphism [41,42]. This moderate level of sexual size dimorphism leads to extended development, including neurodevelopment [43], among males, which is similar to patterns observed in humans (years to peak cerebral volume: rhesus macaque  $F = 4$ ,  $M = 6$ ; human  $F = 10.5$ ,  $M = 14.5$ ; i.e. males develop about 50% slower) [43,44]. Given that brain and body size are correlated within and across species [45] (including rhesus macaques [46]), any observations of GMV in absolute ECV may simply reflect GMV in body size; however, since relative ECV (measured with ECV and body size proxies for each individual specimen) controls for inter-individual variation in body size, GMV in relative ECV may implicate mechanisms acting on this trait (or on the processes linking ECV and body size) specifically. In addition, although higher ranking males tend to have greater reproductive success than low ranking males, this correlation is relatively low [47,48], suggesting there may be multiple behavioural routes to reproductive success in males that could potentially be under balancing selection. Finally, genes expressed in the brain tend to be located on the X chromosome in rodents and primates, including macaques [49], and X chromosome genes affect the development of brain and region size in humans [50,51] and mice [52,53]. These characteristics of the rhesus macaque socio-sexual system, in addition to the fact that rhesus macaques exhibit a typical mammalian XY sex chromosome structure, suggests that any of the aforementioned mechanisms may produce GMV in this species.

To investigate the potential drivers of sex differences in brain size variability (Aim 2), we examined sex differences in the components of phenotypic variance, namely additive genetic and residual (environmental) variances. Previous work has demonstrated that brain size and structure are heritable in humans [54] and non-human primates, including rhesus macaques [55], baboons [56] and chimpanzees [27,57]. If GMV is a result of sex differences in developmental schedules, males should exhibit higher environmental variance than females (Prediction 2a). If GMV is a result of balancing or disruptive selection in males, or sex chromosome effects, males should exhibit higher additive genetic variance than females (Prediction 2b).

## 2. Material and methods

### (a) Subjects

The data used in this study were presented in a previous study [55]. Briefly, A.C. collected morphological measurements for 542 free-ranging rhesus macaques (300 F/242 M) from the Caribbean Primate Research Center (CPRC) skeletal collections at the University of Puerto Rico (UPR) with permission granted through J.H.'s long-term memorandum of understanding (MoU) with CPRC/UPR. Individuals ranged in age at death from 6 to 31 years (mean = 12.8, s.d. = 5.2; females: mean = 13.0, s.d. = 5.0; males: mean = 12.7, s.d. = 5.4). We obtained age at death from the demographic database. Previous work on this population suggested that adult ECV is reached at approximately 4 years in females and approximately 6 years in males and does not change during the adult lifespan [43]. In line with this, linear models suggested

that age (scaled) was not correlated with ECV (scaled) in either sex (males: slope = 0.059,  $p = 0.246$ ; females: slope = 0.053,  $p = 0.221$ ) (electronic supplementary material, figure S1). However, age (scaled) did predict body size (geometric mean, cubed and scaled) in females (males: slope = 0.066,  $p = 0.095$ ; females: slope = 0.130,  $p < 0.001$ ) (electronic supplementary material, figure S1). Given that body size (geometric mean, cubed and scaled) predicted ECV (scaled) in both sexes (males: slope = 0.269,  $p < 0.001$ ; females: slope = 0.357,  $p < 0.001$ ) (electronic supplementary material, figure S1) (consistent with previous work relating brain and body weight in this species [46]), we included age at death as a covariate to account for possible age-related changes in body size and relative ECV (see below for model details).

## (b) Morphological measurements

A.C. measured ECV by pouring 2 mm glass beads into the cranium of each specimen via the foramen magnum. Subsequently, A.C. poured beads into a graduated cylinder and recorded the volume [58]. To estimate body size, we used two body size proxies [59] (femoral length (FL) and femoral mediolateral breadth (FMLB) [60]), measured by A.C., to calculate a geometric mean ( $GM = \sqrt{FL \times FMLB}$ ). We also performed an intraclass correlation analysis to assess intra-rater reliability for all morphological measurements (ICC > 0.95).

## (c) Statistical analysis

We performed all statistical analyses in R 4.0.2 [61].

To examine potential sex differences in ECV phenotypic variance (Aim 1; Predictions 1a,b) and its components (Aim 2; Predictions 2a, b), we used generalized linear mixed models (GLMMs) which incorporate relatedness information in the form of a pedigree (i.e. a quantitative genetic ‘animal’ model [62]). We obtained access to the pedigree via J.H.’s long-term MoU with CPRC/UPR. The CPRC maintains a pedigree database, which contains information on behavioural dams (identified via field observation and available for all specimens in the study) and genetic dams and sires (identified via microsatellite panel and available for individuals born after 1985) [63].

Owing to the inherently high correlations between the sex chromosome relatedness structure and autosomal chromosomes relatedness structure in natural populations, it can be difficult to separately estimate autosomal chromosomal inheritance and sex chromosomal inheritance of a single trait in quantitative genetic models [64]. Simulation modelling has shown that estimation of the additive autosomal genetic variance is a good approximation of both forms of additive genetic variance (i.e. the cumulative variance explained by autosomal and sex chromosomes) [64]. Here, we use quantitative genetic modelling estimating autosomal chromosome genetic variance which is likely to be an accurate estimator of both autosomal and sex chromosome genetic variance.

Across all models described below, we followed other studies [27,57] and used the default prior for the mean and variance of fixed effects for Gaussian-family models in MCMCglmm, and an inverse-Wishart prior for the random effects and residual variances ( $V = 1$ ,  $\nu = 1$ ). All continuous variables were scaled and centred prior to further analysis (mean = 0; s.d. = 1). Models were run for 1 000 000 iterations, sampling every 100 iterations with a burn-in of 500 000. We ensured proper mixing occurred by visually inspecting trace and density plots. Autocorrelation was below 0.1 and effective sample sizes were greater than 1000 for all variables. We implemented Heidelberger and Welch’s convergence diagnostic to test that the sampled values came from a stationary distribution, and all variables passed this stationary test. We also conducted half-width tests, which remove up to half the chain to test that the means are estimated from a chain that has converged and found that all variables passed this test. Finally, we ran each chain twice and confirmed convergence using the Gelman–Rubin statistic [65].

## (i) Aim 1: investigating potential sex differences in brain size variability

We first constructed the following three models to investigate sex differences in the variability of absolute ECV (Model 1; Prediction 1a), relative ECV (Model 2; Prediction 1b) and body size (Model 3):

- (1)  $ECV \sim \text{age at death} + \text{sex}$  (random =  $V_A * \text{sex}$ ;  $rcov = V_R * \text{sex}$ )
- (2)  $ECV \sim \text{body size (GM}^3) + \text{age at death} + \text{sex}$  (random =  $V_A * \text{sex}$ ;  $rcov = V_R * \text{sex}$ )
- (3)  $\text{body size (GM}^3) \sim \text{age at death} + \text{sex}$  (random =  $V_A * \text{sex}$ ;  $rcov = V_R * \text{sex}$ )

In Models 2 and 3, the linear body size proxy (GM) was raised to the third power to ensure the same dimensionality among ECV and body size measures. In all models, the additive genetic variance (estimated by the inverse relatedness matrix estimated by the pedigree) was included as a random effect. Within each model, both the additive genetic and residual terms were partitioned into their effects for males and females separately. For each model, we estimated sex-specific phenotypic variances as the sums of the sex-specific genetic and residual variances for each sample of the posterior distribution ( $V_P = V_A + V_R$ ;  $V_P$  = phenotypic variance;  $V_A$  = additive genetic variance;  $V_R$  = residual (environmental) variance), extracted mean estimates and 68% highest posterior density (HPD) intervals (i.e. the interval of values that contains 68% of the posterior probability) from the sex-specific phenotypic variance distributions, and compared them to test if mean phenotypic variance was higher in males (Predictions 1a,b).

As an additional test of sex differences in variability, we also extracted residuals from Models 1–3 and tested for significant sex differences in the variance of these residuals using permutation tests, following previous work [21,23,27]. We calculated the log male-to-female variance across the residuals (positive values = greater male variability; negative values = greater female variability), randomly permuted the sex variable among the residuals 10 000 times and calculated the proportion of permuted log male-to-female variance ratios (absolute value) greater than the observed ratio (absolute value). This proportion is referred to here as ‘pPERM’ and represents a two-sided test of sex differences in variability. A positive log male-to-female variance ratio and pPERM < 0.05 would indicate greater male variability (Predictions 1a,b).

In addition, within each sex, we calculated the average range size for ECV across 1000 random samples for every possible sample size (from  $n = 2$  to the actual subset sample size). The average range sizes were plotted against sample size to demonstrate that each distribution reached a horizontal asymptote.

## (ii) Aim 2: investigating the potential drivers of sex differences in brain size variability

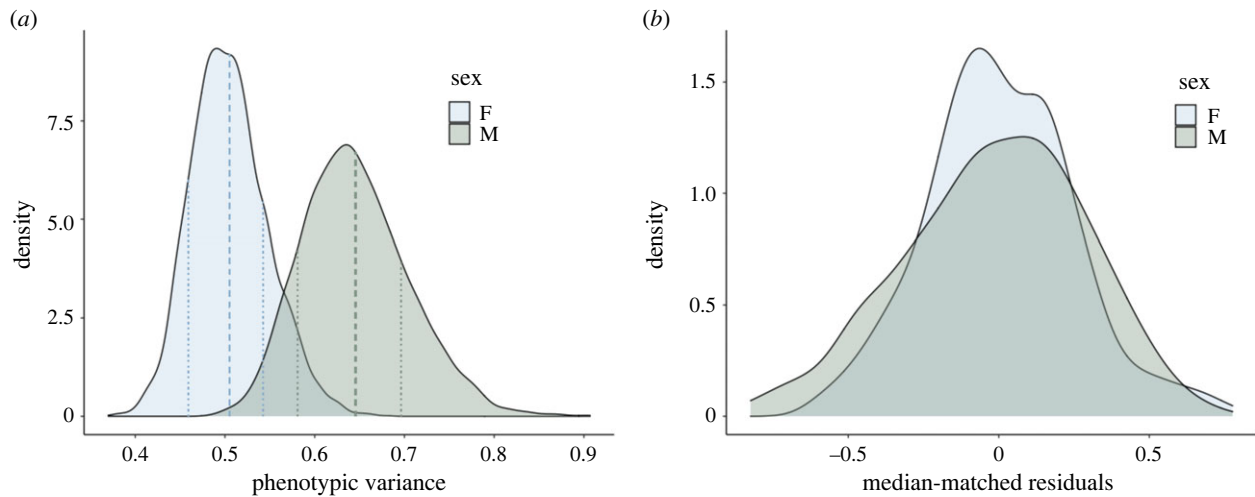
To test whether residual (Prediction 2a) and/or additive genetic variance (Prediction 2b) contributed to sex differences in phenotypic variance, we compared Models 1–3 to reduced models which either: (1) did not partition additive genetic variance by sex, (2) did not partition residual variance by sex or (3) did not partition either additive genetic or residual variance by sex. We compared model fits using the Deviance Information Criterion (DIC). We also extracted mean estimates and 68% HPD intervals from the sex-specific variance distributions for all models (where applicable) and compared them.

## 3. Results

### (a) Aim 1: investigating potential sex differences in brain size variability

Consistent with Prediction 1a, males exhibited more variable absolute ECVs, as demonstrated by their higher mean





**Figure 1.** Males exhibit more variable relative ECVs than females. (a) Density plots of the posterior distributions of phenotypic variance for males (green) and females (blue) from a GLMM of relative ECV (in which both the additive genetic and residual terms were partitioned into their effects for males and females separately). Dashed lines indicate sex-specific means and dotted lines indicate 68% HPD intervals for these means. (b) Median-matched (0 = median for each sex) density plots of male (green) and female (blue) residual distributions from a GLMM of relative ECV (in which both the additive genetic and residual terms were partitioned into their effects for males and females separately).

phenotypic variances with non-overlapping 68% HPD intervals between the sexes (phenotypic variance: males: mean = 0.670 [0.598, 0.717], females: mean = 0.533 [0.481, 0.568]), and significantly more variable residual values (log M-to-F variance ratio = 0.396; pPERM = 0.001) (electronic supplementary material, figures S2, S3 and table S1). Since males also exhibited more variable body sizes (phenotypic variance: males: mean = 0.501 [0.454, 0.544], females: mean = 0.296 [0.269, 0.318]; M-to-F variance ratio = 0.352; pPERM = 0.007) (electronic supplementary material, figures S2, S3 and table S1), GMV in absolute ECV may reflect correlated development between ECV and body size. However, consistent with Prediction 1b, males also exhibited more variable relative ECVs (phenotypic variance: males: mean = 0.646 [0.581, 0.696], females: mean = 0.505 [0.459, 0.542]; M-to-F variance ratio = 0.288; pPERM = 0.016) (figure 1; electronic supplementary material, figures S2, S3 and table S1). Across all models, males were significantly larger (pMCMC < 0.05) (electronic supplementary material, table S1). In all models of relative ECV, body size was a significant, positive predictor (pMCMC < 0.05) (electronic supplementary material, table S1), and in all models of absolute ECV and of body size, age at death was a significant, positive predictor (pMCMC < 0.05) (electronic supplementary material, table S1). ECV increasing with age is not consistent with our exploratory linear model results (see Material and methods) or with previous work on ECV variation in this population [43], which may reflect differences in statistical modelling approaches, sample sizes and age distributions.

Given that measurements are available for more females than males, we expect that our estimates of GMV are conservative. Our resampling procedure confirmed that the sex-specific distributions approached a horizontal asymptote, suggesting our sample sizes were sufficient to capture population-level variation (electronic supplementary material, figure S4).

### (b) Aim 2: investigating the potential drivers of sex differences in brain size variability

We did not find support for Prediction 2a (i.e. greater residual (environmental) variance among males), but we did find some support for Prediction 2b (i.e. greater additive genetic

variance among males). Specifically, contrary to Prediction 2a, models that estimated sex-specific residual variance performed worse (i.e. had higher DIC values) than similar models with unpartitioned residual variance (electronic supplementary material, table S1). However, for all measures, the best-fit models (i.e. with the lowest DIC values) included sex-specific additive genetic variance (electronic supplementary material, table S1) and males exhibited higher mean additive genetic variances in each of these models, with non-overlapping 68% HPD intervals for relative ECV and body size (relative ECV: males: mean = 0.483 [0.410, 0.561], females: mean = 0.347 [0.278, 0.406]; absolute ECV: males: mean = 0.514 [0.432, 0.580], females: mean = 0.382 [0.319, 0.445]; body size: males: mean = 0.367 [0.316, 0.420], females: mean = 0.168 [0.133, 0.206]).

## 4. Discussion

This work demonstrates GMV in ECV, a proxy for brain size, in a non-human primate model species, the rhesus macaque. Specifically, we found that males exhibit more variable absolute ECVs than females (in support of Prediction 1a), which may reflect that males also exhibit more variable body sizes. However, males also exhibit more variable relative ECVs (in support of Prediction 1b), suggesting there is GMV in ECV above and beyond that observed for body size. We did not find evidence that males exhibit greater environmental (residual) variance, suggesting that the patterns of GMV observed here may not reflect sex differences in developmental schedules (contra Prediction 2a). Rather, greater additive genetic variance among males suggests that GMV in rhesus macaque ECV is likely to reflect some combination of sexual selection mechanisms and sex chromosome effects (in support of Prediction 2b). Although ECV is a reliable estimate of brain size and shape across species [34–36] and within species throughout development [35,37], our analyses cannot account for the effects of inter-individual and age-related variation in the proportion of non-neural tissue. This proportion is likely to increase with age in rhesus macaques, since previous work suggests that brain

volume decreases with age in this species [39,40]. Nevertheless, unless this measure varies across this population in a manner that would produce more similar brain sizes among males compared with females (despite males having more variable ECVs), then our observation of GMV in ECV is likely to reflect GMV in brain size.

As mentioned above, development is relatively longer in male (versus female) rhesus macaques [43]. Given that longer cranial ontogeny could potentially expose males to more environmental factors that influence cranial size development, we predicted that these factors may lead male rhesus macaques to display greater ECV variability (Prediction 2a). However, we did not find support for this, as models that estimated sex-specific residual variances performed worse than similar models with unpartitioned residual variance. This suggests that sex differences in exposure to physical and/or social environments during development cannot account for observed sex differences in ECV variability. Rather, our results suggest that sexual selection mechanisms and/or sex chromosome effects are likely to explain our observation of GMV in rhesus macaque ECV (Prediction 2b). Specifically, we found that the best-fit models for all measures included sex-specific additive genetic variance, and males exhibited higher mean additive genetic variances in these models (although the HPD intervals did marginally overlap for absolute ECV).

Given that both absolute and relative ECV are heritable [55,66], greater variability in ECV among males may, in theory, reflect selection on cognition and behaviour—specifically, disruptive or balancing selection on male reproductive strategies. Male rhesus macaques are subject to an interesting mix of sexual selection pressures, including direct male–male competition, reflected by intermediate body and canine size dimorphism [67], indirect male–male competition, reflected by large relative testis volume [68] and mechanisms of indirect female mate choice, such as female preference for males with darker red faces [69]. Accordingly, although dominance rank predicts male rhesus macaque reproductive success, male reproductive skew is relatively low and may reflect that males tend to queue for dominance rank instead of fighting directly [48]. Together, these mechanisms are expected to create several routes to reproductive success in males, which would produce and maintain variation in male behavioural phenotypes instead of generating one male phenotype that is under directional selection. Previous comparative work suggests that either of these alternative routes (e.g. greater investment in direct or indirect male–male competition) may be facilitated by increased brain size [70,71] and/or introduce constraints on brain size (in the form of, e.g. tissue/energetic trade-offs) [72,73]. Accordingly, if alternative male reproductive tactics are differentially coupled to brain size in this species, this may lead to more variable ECVs among males. Alternatively, the female fitness optimum for ECV may be narrower than the fitness optimum in males, which would suggest that our results reflect stronger stabilizing selection in females. Colby *et al.* [55] used linear and quadratic selection gradients to investigate selection in this sample and did not find evidence of selection on absolute or relative ECV in either sex, which suggests that stabilizing selection on females may not underlie the sex differences in ECV variability observed in this study. However, a lack of evidence for selection may reflect that: (1) this population is not currently under selection for ECV or (2) selection is occurring

in this population, but larger sample sizes would be required to detect it. Furthermore, if balancing or disruptive selection are occurring in this population, they would not be detectable using linear or quadratic models. Accordingly, we cannot rule out selection as a potential explanation for greater ECV variability among male rhesus macaques.

Finally, our results may reflect sex chromosome effects. While males can only express a single set of X-linked alleles throughout all of their somatic cells, females may exhibit variable X chromosome gene expression across cells (due to mosaic XCI) or within cells (due to XCI escapees), leading to more intermediate levels of trait expression and lower population variability in females [5,12]. These effects may be particularly strong on neurodevelopment since brain-expressed genes and genes associated with brain size tend to be located on the X chromosome [49–53]. This may reflect evolutionary dynamics unique to the X chromosome, including the ‘faster-X effect’ (i.e. more rapid evolution of X chromosome genes due to a relatively lower effective population size than autosomes) and the accumulation of sex-biased genes on the X chromosome (potentially reflecting resolved sexual antagonism) [74,75]. Owing to limited data availability, the current dataset did not provide the power to separately estimate autosomal versus sex chromosomal additive genetic effects (see Material and methods) [64]. Nevertheless, previous work has also provided indirect evidence for sex chromosome effects on sex differences in trait variability, including greater size correlations between brain areas in human and chimpanzee males (versus females) [21,23,27] and higher phenotypic variability among females in species with homogametic (e.g. ZZ) males [5].

While sex differences in phenotypic variability have been demonstrated in many taxa and across numerous traits, almost all studies demonstrating sex differences in neuroanatomical or behavioural variability have focused on humans. Here, we show that greater neuroanatomical variability among males is not only present in humans [26], but also another primate species, the rhesus macaque. Accordingly, this work supports the use of rhesus macaques as an animal model for sex-biased human neurological and psychiatric conditions. To tease apart which factors predominantly account for observed sex differences in neuroanatomical variability, future studies should focus on taxa that exhibit a diverse array of mating systems, sex-specific developmental schedules and sex chromosome compositions.

**Data accessibility.** All data used in the analyses presented in this study are available from the Dryad Digital Repository: <https://doi.org/10.5061/dryad.ffbg79cz0> [76].

Other supplementary material can be found at [77].

**Authors' contributions.** A.C.: conceptualization, data curation, formal analysis, methodology, writing—original draft; A.D.: conceptualization, formal analysis, methodology, writing—original draft; E.C.: conceptualization, formal analysis, writing—review and editing; J.H.: conceptualization, supervision, writing—review and editing.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

**Conflict of interest declaration.** We declare we have no competing interests.

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