

Meningococcal Urethritis: Old and New

[Bethany L. Burns,](https://orcid.org/0000-0001-9391-7448)^a © [Daniel D. Rhoads](https://orcid.org/0000-0002-7636-5191)^{a,b,c}

a Department of Laboratory Medicine, Cleveland Clinic, Cleveland, Ohio, USA

bDepartment of Pathology, Cleveland Clinic Lerner College of Medicine, Case Western Reserve University, Cleveland, Ohio, USA cInfection Biology Program, Lerner Research Institute, Cleveland Clinic, Cleveland, Ohio, USA

ABSTRACT Neisseria meningitidis is a common commensal bacterium found in the respiratory tract, but it can also cause severe, invasive disease. Vaccines have been employed which have been successful in helping to prevent invasive disease caused by encapsulated N. meningitidis from the A, C, W, Y, and B serogroups. Currently, nonencapsulated N. meningitidis groups are more common commensals in the population than in the prevaccine era. One emerging nonencapsulated group of bacteria is the U.S. N. meningitidis urethritis clade (US_NmUC), which can cause meningococcal urethritis in men. US_NmUC has unique genotypic and phenotypic features that may increase its fitness in the male urethra. It is diagnostically challenging to identify and distinguish meningococcal urethritis from Neisseria gonorrhoeae, as the clinical presentation and microbiological findings are overlapping. In this review, the history of meningococcal urethritis, emergence of US_NmUC, laboratory diagnosis, and clinical treatment are all explored.

KEYWORDS meningococcal urethritis, Neisseria meningitidis, US_NmUC, urethritis

The majority of Neisseria species are considered commensal organisms found in the upper respiratory tract [\(1,](#page-7-0) [2](#page-7-1)). Although Neisseria meningitidis is typically found as a commensal, it is unique among Neisseria in that it can cause outbreaks of communityonset meningitis, which can be fatal [\(3](#page-7-2)–[6](#page-7-3)). In contrast to other Neisseria, Neisseria gonorrhoeae is an obligate pathogen and is never considered part of the healthy microbiome, regardless of symptomatology [\(7,](#page-7-4) [8](#page-7-5)). N. gonorrhoeae is a sexually transmitted infection (STI) agent that is capable of infecting the urethra, cervix, rectum, and oropharynx. The majority of N. gonorrhoeae infections present clinically as urethritis in men or cervicitis in women ([9](#page-7-6)).

Meningococcus and gonococcus are considered distinct taxa causing distinct diseases, but recent discoveries have revealed that there may be more overlap between the species than was previously recognized. For example, meningococcus is now recognized as a cause of urethritis which is clinically indistinguishable from gonococcal urethritis [\(10](#page-7-7)). In one study, 20% of urethritis caused by Neisseria spp. was attributable to meningococcus [\(11\)](#page-7-8). Genetic analyses of these N. meningitidis strains have revealed genomic features of N. gonorrhoeae [\(12\)](#page-7-9). These findings challenge our traditional understanding of the role of N. meningitidis in causing disease. Dissimilar to other STIs, meningococcal urethritis has been documented in long-term monogamous relationships in which the partner has no disease. N. meningitidis is increasingly recognized as a cause of urethritis, and in this review, we will explore this emerging disease.

MENINGOCOCCAL URETHRITIS, PRE-2010: THE EARLY DAYS

Small case series of meningococcal urethritis have been documented in the literature intermittently since the mid-1900s ([13\)](#page-8-0). Beginning in the 1970s and continuing into the 1980s, interest in meningococcal urethritis rose in parallel to the HIV/AIDS pandemic [\(14](#page-8-1)–[17](#page-8-2)). The majority of meningococcal urethritis research during this time Editor Romney M. Humphries, Vanderbilt University Medical Center

Copyright © 2022 American Society for Microbiology. [All Rights Reserved.](https://doi.org/10.1128/ASMCopyrightv2)

Address correspondence to Bethany L. Burns, pathologyburns@gmail.com.

The authors declare a conflict of interest. D.D.R. has received compensation from Luminex, Talis Biomedical, and Roche for scientific advisory. D.D.R. has sponsored agreements through his employer with the following groups: Abbott, Accelerate Diagnostics, Altona, BD, BioFire, bioMerieux, Bio-Rad, Cepheid, Cleveland Diagnostics, HelixBind, Hologic, Luminex, Qiagen, Q-Linea, Roche, Specific Diagnostics, Thermo Fisher, and Vela Diagnostics. D.D.R. is a co-investigator with NIH/NIAID R01 HS028633-01.

Published 15 August 2022

		Challenges in distinguishing	
Diagnostic approach	Testing option	N. gonorrhoeae from N. meningitidis	Practical challenge(s)
Direct exam	Clinical presentation	Cannot distinguish. Both have similar clinical presentations.	Testing is required for definitive diagnosis.
	Gram stain	Cannot distinguish. Both are Gram- negative diplococci.	N. meningitidis could be misinterpreted as N. gonorrhoeae.
Culture	Selective medium	Cannot distinguish. Both grow on agar media selective for gonococcus.	Multiple specimens would need to be collected to be able to culture for Neisseria spp. and also investigate the presence of other potential causes of urethritis (e.g., Trichomonas vaginalis, Chlamydia trachomatis, and Mycoplasma genitalium).
	Oxidase reaction	Cannot distinguish. Both are oxidase positive.	
	Carbohydrate	Cannot reliably distinguish.	
	fermentation	Overlapping carbohydrate fermentation profiles have been described (28).	
	MALDI-TOF MS	Able to distinguish. N. meningitidis and N. gonorrhoeae can be distinguished, but misidentifications within the genus have been reported.	
Molecular characterization	NAAT	Cannot reliably distinguish or detect. N. meningitidis cross-reactivity with N. gonorrhoeae NAAT has been described (65).	N. meningitidis testing of urethral specimens is not commercially available.
	WGS	Able to distinguish. N. meningitidis and N. gonorrhoeae can be distinguished using WGS.	Not routinely performed in contemporary clinical practice.

TABLE 1 Diagnostic challenges in distinguishing N. gonorrhoeae and N. meningitidis as causes of urethritis

focused on men who have sex with men (MSM). A large population study of MSM $(n = 815)$ who presented to a community clinic had infrequent carriage of N. meningitidis in the urethra (0.7%, 6/815) [\(14](#page-8-1)). In the study, participants agreed to cultures of the urethra, oropharynx, and rectal canal, and subjects were included in the study whether or not symptoms of urethritis were reported. Of the men from which N. meningitidis was recovered from urethral cultures ($n = 6$), most (83%, 5/6) had symptomatic meningococcal urethritis [\(14](#page-8-1)). In total, 2.2% (5/227) of those with urethral discharge had meningococcal urethritis [\(14](#page-8-1)). Those with positive rectal cultures (2.0%, 16/815) less frequently reported symptoms (25%, 4/16), suggesting N. meningitidis could potentially be a commensal organism or an asymptomatic infection in the rectum ([14\)](#page-8-1). Although rates of N. meningitidis recovery from the urogenital tract were low, the rate of N. meningitidis carriage in the oropharynx of this MSM cohort was much higher (42.5%, 347/815) [\(14](#page-8-1), [16](#page-8-3)).

A study published in 1980 described men with presumed gonococcal urethritis $(n = 482)$. This study concluded that N. meningitidis was more likely to cause urethritis in homosexual men (13%, 15/114) than heterosexual men (1%, 4/368) ([15\)](#page-8-4). Using these findings, the authors argued against the standard laboratory practice of presumptive identification of N. gonorrhoeae when oxidase-positive, Gram-negative diplococci were recovered by culture using Thayer-Martin medium. This study highlighted that although N. meningitidis is an uncommon cause of urethritis in the general population, it could be underrecognized and misdiagnosed as an N. gonorrhoeae infection due to the laboratory practices of the time [\(Table 1\)](#page-1-0).

Studies performed through the 1990s reached similar conclusions; meningococcal urethritis was an uncommon entity. One retrospective study from London, England, by Maini et al. identified low incidence of genital N. meningitidis carriage when using urethral and cervical cultures among patients being screened for STIs. Meningococcal urethritis was identified rarely in the MSM cohort (0.2%, 11/5571) ([18\)](#page-8-5), and meningococcus was not isolated from a man identifying as heterosexual (0%, 0/8,992) [\(18](#page-8-5)). N. meningitidis was not isolated in a cervical culture (0%, 0/15,976) [\(18](#page-8-5)), although rare case reports of encapsulated N. meningitidis associated with cervicitis have been published [\(19](#page-8-7)). Later, studies identified asymptomatic N. meningitidis carriage in the upper respiratory tract of sexual partners of monogamous men who developed meningococcal urethritis, which fostered the hypothesis that oral sexual contact may be the means of transmission for individuals with meningococcal urethritis [\(20](#page-8-8), [21](#page-8-9)). At the end of the 20th century and beginning of the 21st century, meningococcal urethritis seemed to be a rare STI, nearly exclusively impacting MSM and thought to be transmitted by fellatio [\(20,](#page-8-8) [22](#page-8-10)–[25](#page-8-11)).

EMERGENCE OF A CONTEMPORARY MENINGOCOCCUS CLADE: THE RECENT DECADE

In the 2010s, the U.S. Center for Disease Control and Prevention (CDC)'s Gonococcal Isolates Surveillance Program (GISP) detected several unusual clusters of meningococcal urethritis [\(26](#page-8-12)). Novel findings were published detailing the unexpected demographics and microbial genetics associated with the clusters ([11](#page-7-8), [25](#page-8-11), [27](#page-8-13)). These novel findings have challenged previous conclusions about the pathophysiology of meningococcal urethritis.

Patient demographics. The clusters of meningococcal urethritis predominately affected heterosexual men (92 to 99%) ([11,](#page-7-8) [25,](#page-8-11) [27\)](#page-8-13). Interestingly, Bazan et al. identified that significantly more men with meningococcal urethritis identified as heterosexual (99%) than men with gonococcal urethritis (78%) [\(11\)](#page-7-8). Like other STIs, meningococcal urethritis was frequently observed in younger men, often 20 to 30 years old (median, 31 or 32 years of age) but some men as old as 60 years [\(25](#page-8-11), [27\)](#page-8-13). The population of men with N. meningiti-dis had similar age, race, and ethnicity to those with N. gonorrhoeae infection [\(11\)](#page-7-8).

The clusters of cases were mostly identified in STI clinics in the midwestern United States. Specifically, large clusters in cities in Ohio, Indiana, and Michigan have been reported ([11](#page-7-8), [25](#page-8-11), [27\)](#page-8-13). However, reports of similar cases outside the United States are also published ([28\)](#page-8-6).

Clinical presentation, sexual history, and coinfections. Men with meningococcal urethritis were symptomatic (97 to 100%) ([25](#page-8-11), [27](#page-8-13)). Most had urethral discharge $(>90%)$, and many had dysuria [\(11](#page-7-8), [25](#page-8-11), [27\)](#page-8-13). These findings align with historic studies that identified N. meningitidis from male genital specimens where most men were symptomatic at presentation [\(14\)](#page-8-1). Men reported symptoms ranging from 2 to 7 days with a median of 4 days duration.

The number of days since last sexual encounter ranged from 3 to 10 days with a median of 7 days [\(11](#page-7-8)). Days of symptoms and number of days since last sexual encounter were similar to that of patients with gonococcal urethritis [\(25](#page-8-11)). Data collected from both self-reporting forms and physician interviews established that almost all men with meningococcal urethritis reported oral sex encounters (93 to 100%) ([25\)](#page-8-11). In one cohort, all men reported receiving fellatio (100%) ([25\)](#page-8-11). Men with meningococcal urethritis reported a similar number of partners over 90 days (median, 2) and 12 months (median, 3) as men with gonococcal urethritis (median, 2 per month, 3 per year) [\(11](#page-7-8)).

It is imperative to consider coinfections when an STI is identified in order to adequately manage all infections that may be present. The urethral specimens collected from the Midwest clusters were tested for N. gonorrhoeae (nucleic acid amplification test [NAAT]), Chlamydia trachomatis (NAAT), syphilis (serology), and trichomoniasis (culture) [\(11\)](#page-7-8). Curiously, coinfections with meningococcus and gonococcus have not been described; however, history of prior gonococcal urethritis was common. In one cohort of men with meningococcal urethritis, over half of the men (52%) reported previous N. gonorrhoeae infection, with one-third (33%) reporting infection within the last 12 months [\(25](#page-8-11)). Whereas coinfection with N. gonorrhoeae was not identified, a marked proportion of men with meningococcal urethritis had a concurrent infection with C. trachomatis (15 to 19%), which is similar to coinfection rates of gonococcal urethritis and C. trachomatis (24.2%) ([11](#page-7-8), [25](#page-8-11), [27,](#page-8-13) [29,](#page-8-14) [30](#page-8-15)). HIV infection was uncommon in those with meningococcal urethritis (0 to 2%) [\(11](#page-7-8), [25,](#page-8-11) [27\)](#page-8-13). Interestingly, in an MSM cohort, different types of N. meningitidis have been isolated from upper respiratory and urethral cultures from the same person [\(16](#page-8-3)).

Genomic and phenotypic features of US_NmUC. The recent meningococcal urethritis clusters have been characterized and determined to be a novel clade. This clade

TABLE 2 Summary of approaches to genetic characterization of N. meningitidis strains and genetic characterization of the United States N. meningitidis urethritis clade (US_NmUC)^a

^aMLST, multilocus sequence typing; ST, sequence type; cc, clonal complex; MLEE, multilocus enzyme electrophoresis; ET, electrophoretic profile; WGS, whole-gene sequencing; cgMLST, core genome MLST.

^bUsed in conjunction with MLST.

is often described in publications as "U.S. N. meningitidis urethritis clade" (US_NmUC) or "N. meningitidis, nongroupable" (NmNG).

Like many bacteria, the relatedness of N. meningitidis isolates is commonly characterized using multilocus sequence typing (MLST) [\(Table 2](#page-3-0)). Rather than examining the entire genome, MLST identifies genotypic variation in several specific housekeeping genes, and the isolates are classified based on this sequence type (ST). When multiple STs share a certain amount of similarity, they are grouped into a clonal complex (cc) ([31\)](#page-8-16). A cc may further be divided into sublineages based on additional genotypic characterization ([32](#page-8-17)).

The MLST approach for N. meningitidis was published in 1998 by Maiden et al., who proposed using seven housekeeping genes (abcZ, adk, aroE, FumC, gdh, pdhc, and pqm) ([33](#page-8-18)), and this MLST scheme is still used today. Adjunct classification is achieved by "fine typing" of several supplementary genes and their respective variable regions (PorA, PorB, FetA, fHbp, nadA, and nhbA) [\(34](#page-8-19), [35](#page-8-20)). Fine typing also provides information on a strain's virulence and can provide vaccine development insights. MLST remains a mainstay of classification, but classification based on whole-genome sequencing may be on the horizon ([12,](#page-7-9) [36\)](#page-8-21). Whole-genome sequencing uses 1,605 loci of the core genome with specific molecular typing loci to characterize N. meningitidis strains ([34,](#page-8-19) [37](#page-8-22)).

US_NmUC is considered a novel clade of the ST-11 within the hyperinvasive lineage of N. meningitidis (cc11) [\(11\)](#page-7-8). Specifically, US_NmUC is an ST-11 cc11 and an ET-15 variant of sublineage 11.2 (fine-type PorA VR1 5-1, VR2 10-8, VR3 36-2, FETA F3-6, and PorB 2-2 [or rarely, PorB 2 to 78 and 2 to 52]) [\(38](#page-8-23), [39](#page-8-24)). The clade was first described in by Bazan and colleagues from isolates cultured in Columbus, Ohio. It has subsequently been identified in Oakland County, Michigan, and in other U.S. cities [\(11,](#page-7-8) [12](#page-7-9), [25,](#page-8-11) [27](#page-8-13)). As its name suggests, the clade has novel phenotypic characteristics that may enable it to cause urethritis more effectively than previously characterized lineage 11 isolates of N. meningitidis ([Table 3](#page-3-1)) [\(32,](#page-8-17) [37,](#page-8-22) [39](#page-8-24)).

TABLE 3 Summary of genetic alterations identified in United States N. meningitidis urethritis clade (US_NmUC) with resulting phenotypic effects and presumed mechanisms for increased urethrotropic fitness

The US_NmUC clade is nongroupable, which means it lacks a polysaccharide capsule; it is unencapsulated ([39\)](#page-8-24). This is one distinguishing feature of US_NmUC, which is in contrast to N. meningitidis, which has caused meningitis outbreaks in MSM and has been linked to serotype C ([32](#page-8-17)). US_NmUC capsular biosynthesis gene cluster is disrupted by an insertion sequence (IS1301) that replaces ccsA, cssB, cssC, and part of csc. This disruption of the capsular polysaccharide (cps) locus prevents production of functional capsule polymerase, so the bacteria are unencapsulated [\(12,](#page-7-9) [39](#page-8-24)). This genetic mutation also yields loss of wild-type lipo-oligosaccharide (LOS) sialylation production, and the loss of function may improve mucosal adherence, which has been demonstrated in studies involving Nm ST-11 ([39,](#page-8-24) [40\)](#page-8-25). Improved mucosal adherence could improve the fitness of N. meningitidis living in the urethra [\(39](#page-8-24), [41\)](#page-8-26).

US_NmUC has features that may enable it to better evade innate immunity than other meningococcal strains [\(Table 3](#page-3-1)). A unique feature of US_NmUC is its hybrid factor H binding protein gene (fHbp). The hybrid fHbp in US NmUC is thought to result in high protein expression of factor H binding protein (FHbp) on the surface of US_NmUC isolates. FHbp binds to human factor H, which is a complement regulator that interacts with C3b. When FHbp on the surface of US_NmUC binds to circulating human factor H, the alternative pathway of complement activation is hindered because the human factor H is unavailable for binding to C3b. Tzeng et al. hypothesized that high expression of FHbp may be protective in the urogenital tract where levels of human factor H are inherently low [\(39\)](#page-8-24). FHbp is an important target protein of N. meningitidis vaccines, particularly serogroup B vaccines [\(42\)](#page-8-27). Although it would seem intuitive that serogroup B vaccines targeting FHbp would be protective against US_NmUC, it is unclear if serogroup B vaccines are cross-protective in preventing meningococcal urethritis by this mechanism or others, as demonstrated in gonococcal urethritis ([12](#page-7-9), [43\)](#page-8-28). Additionally, point mutations associated with the type IV pillin biogenesis apparatus demonstrated high levels of heteroresistance to antimicrobial peptides (AMPS), which are part of the innate immune system [\(44](#page-8-29)). Overexpression of FHbp and mutations in type IV pillin may enable US_NmUC to evade segments of the host immune response [\(39\)](#page-8-24).

US_NmUC isolates were found to have genetic material normally present in N. gonorrhoeae [\(12](#page-7-9), [39\)](#page-8-24). US_NmUC isolates contain a complete norB-aniA cassette that enables denitrification. Nitrates are commonly found in urine, and interestingly, reduction of nitrogen compounds is a defining feature of Enterobacterales, which cause the majority of genitourinary tract infections. When present in N. gonorrhoeae, nitrite reducatase (AniA) and nitric ox-ide reductase (NorB) allow for denitrification and, ultimately, anaerobic growth ([45](#page-8-30)), and these pathways may also be important in biofilm formation [\(46](#page-8-31)). Compared to N. gonorrhoeae, US_NmUC grew robustly on gonococcal agar plates supplemented with sodium ni-trite [\(39\)](#page-8-24). US_NmUC's acquisition of the norB-aniA cassette from N. gonorrhoeae likely enhances its ability to live in the urogenital tract ([39](#page-8-24)). Although US_NmUC' s novel phenotypic characteristics may enable it to cause urethritis more effectively, other strains of N. meningitidis are also capable of causing urethritis in men ([47](#page-8-32)).

US_NmUC has been rarely reported to cause invasive disease. Retchless et al. reported five isolates obtained from 2013 to 2016 that were cultured from cerebrospinal fluid (CSF) ($n = 2$) and blood ($n = 3$) from male and female patients ([12\)](#page-7-9). These rare cases of invasive disease caused by unencapsulated US_NmUC occurred in immunocompromised patients ([3](#page-7-2), [48\)](#page-8-33). Invasive disease caused by unencapsulated meningococcus is often associated with immunodeficiency ([49](#page-8-34)[–](#page-9-1)[51\)](#page-9-2).

THE VACCINE ERA AND CHANGES IN MENINGOCOCCUS OROPHARYNGEAL CARRIAGE

Almost all invasive meningococcal disease is caused by groupable (encapsulated) strains of N. meningitidis, specifically serogroups A, B, C, W, X, and Y [\(52\)](#page-9-3). As of early 2022, three quadrivalent meningococcal vaccines (groups A, C, Y, and W) and two recombinant vaccines (group B) are available in the United States ([52,](#page-9-3) [53](#page-9-4)). Since the late 1990s, the incidence of invasive meningococcal disease has declined ([54](#page-9-5)). The incidence of invasive meningococcal disease declined 2-fold to 3-fold in vaccinated age groups after implementing quadrivalent vaccines ([55](#page-9-6)). In the vaccine era, oropharyngeal carriage of unencapsulated strains is

now more common than in the prevaccine era [\(56](#page-9-7)). We raise the possibility that an increase in nongroupable meningococcus carriage may be one variable playing a role in the growing number of meningococcal urethritis cases in the United States today.

Meningococcal serogroup B vaccination has been correlated with a decrease in gonococcal infection [\(12](#page-7-9), [43](#page-8-28)). Similar cross-protective immunity from the meningococcus B vaccine could provide some protection from US_NmUC urethritis, but no evidence of this cross-protection has been reported.

LABORATORY DETECTION OF MENINGOCOCCAL URETHRITIS

Traditionally, a bedside Gram stain of urethral exudate was used to identify gonococcus, but this approach cannot differentiate N. gonorrhoeae and N. meningitidis, as they both appear as Gram-negative diplococci [\(15\)](#page-8-4). In culture, both gonococcus and meningococcus are fastidious organisms that grow to various degrees on nonselective media like chocolate agar and also on selective agars like Thayer-Martin agar and New York City agar ([57](#page-9-8)). Both are oxidase positive. Historical methods of isolate identification include sugar fermentation reaction profiles to differentiate N. meningitidis (ferments glucose and maltose, but not lactose and sucrose) from N. gonorrhoeae (ferments glucose but not lactose, maltose, or sucrose), although these methods are not routinely used today [\(58](#page-9-9)). Some meningococcus isolates that cause urethritis have demonstrated loss in the ability to ferment maltose, so they could be mistaken for gonococcus if using sugar fermentation for identification [\(28\)](#page-8-6). Biochemical testing, including evaluation of enzyme production, can be used for identification, but these tests also have limitations [\(59\)](#page-9-10). In contemporary clinical bacteriology practice, matrix-assisted laser desorption ionization–time of flight mass spectrometry (MALDI-TOF MS) has proven to be a practical and accurate method to identify and discriminate N. gonorrhoeae and N. meningitidis isolates; however, lack of specificity in distinguishing these species from others within the Neisseria genus has been reported [\(60](#page-9-11)–[63\)](#page-9-12). In our own clinical microbiology practice, we have found the MALDI-TOF MS useful in the identification of N. meningitidis recovered in low quantities in routine urine cultures from men who are being evaluated for a potential urinary tract infection and nongonococcal urethritis.

Nucleic acid amplification tests (NAATs) have become the mainstay for clinical detection of the pathogens which cause urethritis. Testing panels for suspected STIs that cause urethritis target the most prevalent pathogens, Chlamydia trachomatis, Trichomonas vaginalis, Mycoplasma genitalium, and N. gonorrhoeae [\(64\)](#page-9-13). The Hologic Aptima (San Diego, CA) NAAT that targets N. gonorrhoeae has been reported to cross-react with the ST-11 N. meningitidis urethritis clade, but this cross-reactivity could not be replicated with the Cepheid GeneXpert (Sunnyvale, CA) NAAT [\(65](#page-9-0)). Cross-reactivity with other NAAT diagnostic systems has not been reported, but it is unknown if other NAAT systems have been interrogated for cross-reactivity with US_NmUC. Studies have described genomic mixing in which bacteria can have nucleic acid from both N. meningitidis and N. gonorrhoeae, and distinguishing between the species using routine nucleic acid diagnostic testing can be challenging in these situations [\(65,](#page-9-0) [66\)](#page-9-14). However, it is unclear how frequently these amalgamations are encountered and how frequently cross-reactivity using diagnostic testing might be occurring. Unfortunately, no FDA-authorized NAAT is currently available for the detection of meningococcal urethritis in clinical practice.

One group developed a SimpleProbe real-time PCR assay to detect N. meningitidis in urine specimens and evaluated its performance in a proof-of-concept study [\(67](#page-9-15)). The probe detects US_NmUC by targeting a region within the norB allele. The norB gene is found in both N. gonorrhoeae and US_NmUC, but a single nucleotide polymorphism (G431A) is specific for US_NmUC. N. gonorrhoeae and US_NmUC were able to be distinguished by melt curve analysis of the *norB* amplicon ([67](#page-9-15)).

A few studies have attempted to estimate the prevalence of meningococcal urethritis. At a large sexually transmitted disease (STD) clinic in Columbus, Ohio, Bazan and colleagues conducted an 11-month study in 2015 where all men who presented had a urethral swab and urine specimen collected. In Bazan's cohort, 20% of samples with Gram-negative diplococci had undetectable N. gonorrhoeae by NAAT, but N. meningitidis was identified in these

samples using multiple testing modalities (strip biochemical testing, real-time PCR, wholegenome sequencing [WGS], etc.), which confirmed all but one of these as cases of meningococcal urethritis ($n = 74/75$) [\(11](#page-7-8), [68\)](#page-9-16). During the same period, 297 culture-positive urethral gonorrhea (GC) cases were identified; all had urethral Gram-negative identification plate (GNID) and positive urine NAAT for N. gonorrhoeae [\(11\)](#page-7-8). Similarly, Toh et al. retrospectively identified an increase in samples with Gram-negative diplococci and negative N. gonorrhoeae by NAAT from 2013 to 2016 in Indianapolis, Indiana. These samples with biochemical testing characteristics of N. meningitidis increased from 2.8% ($n = 12/436$) to 9.8% $(n = 50/510)$ ([27](#page-8-13)). The majority of these samples were not confirmed by MALDI-TOF MS or WGS to rule out other Neisseria species, albeit the increase in Toh's presumptive meningococcal urethritis cases correlated with the increase in WGS-proven meningococcal urethritis cases identified in Bazan's cohort. Another study published that 30% of meningococcal urethritis cases had N. gonorrhoeae detected by NAAT, and this finding would benefit from additional investigation in other geographic locations [\(65](#page-9-0)).

It is difficult to accurately estimate how many cases of meningococcal urethritis are going undetected or may be misidentified as N. gonorrhoeae, but overlap in symptomatology and current laboratory detection methods of gonococcus and meningococcus may yield underreporting of the prevalence of meningococcal urethritis.

CLINICAL MANAGEMENT OF MENINGOCOCCAL URETHRITIS

According to 2021 CDC guidelines, meningococcal urethritis should be treated with the same approach as gonococcal urethritis [\(69](#page-9-17)). Currently, uncomplicated gonococcal infections in the urethra are treated with a single dose of ceftriaxone intramuscularly (i.m.; 250 mg) when chlamydial infection is excluded. The CDC guidelines also state that sexual partners exposed to meningococcal urethritis may be treated similarly to sexual partners exposed to N. gonorrhoeae; however, there are little to no data specific to N. meningitidis on the utility of treating sexual partners. In the case of meningococcal urethritis, the sexual partner's oropharyngeal carriage of US_NmUC is likely to be the source of the transmission but is unlikely to be causing disease in the partner.

Bazan and colleagues have described recurrent infections of meningococcal urethritis in men [\(70](#page-9-18)). Half of the men (57%, 73/128) in the original US_NmUC cohort returned for follow-up. Of them, five had a recurrent infection with US_NmUC (7%, 5/73). All identified as heterosexual men and reported oral-genital contact (100%, 5/5). The median duration of time between the first and second episode was less than a year (median, 274 days; range, 83 to 576 days). All were previously treated with the CDC-recommended treatment regimen (intramuscular ceftriaxone [250 mg] and oral azithromycin [1 g]). Bazan and colleagues suggest considering treatment with an antibiotic regimen that eradicates meningococcal oropharyngeal carriage in sex partners of men with meningococcal urethritis caused by US_NmUC [\(70](#page-9-18)). Currently, the CDC's 2021 treatment guidelines for meningococcal urethritis do not recommend treating people with N. meningitidis of the oropharynx [\(69](#page-9-17)). Given the potential that meningococcal urethritis may be normal microbiota in the upper respiratory tract of a partner, it is unclear if partner-directed therapy is appropriate or necessary (see "N. meningitidis carriage and vaccination" above).

Studies specific to the clinical management of meningococcal urethritis are extremely limited. In order to adequately study clinical management of meningococcal urethritis, NAAT diagnostic testing likely needs to be made available so cases of meningococcal urethritis can be easily diagnosed in routine medical practice (see "Laboratory detection of meningococcal urethritis").

ANTIBIOTIC RESISTANCE

To date, N. meningitidis isolates that cause urethritis are generally thought to be susceptible to antibiotics without resistance that can cause treatment failure. One study has documented the majority of isolates to be penicillin resistant (10/13) [\(65\)](#page-9-0), but these isolates were susceptible to ceftriaxone and ciprofloxacin (13/13) [\(65\)](#page-9-0). N. gonorrhoeae has rarely achieved resistance to third-generation cephalosporins, and this resistance has not yet been identified in N. meningitidis [\(71](#page-9-19), [72](#page-9-20)). The CDC has declared N. gonorrhoeae antimicrobial resistance as a significant threat to public health ([73\)](#page-9-21). Rare ciprofloxacin-resistant US_NmUC isolates have been reported, and the resistance mechanism is genetically the same as N. gonorrhoeae (mutated DNA gyrase gene [gyrA]) and was likely acquired by transmission of the genetic material from N. gonorrhoeae to N. meningitidis ([28\)](#page-8-6). Subpopulations of US_NmUC that are resistant to polymyxins due to MTR efflux and LptA-mediated lipid A modification have been described ([44\)](#page-8-29). Azithromycin resistance has also been reported [\(12,](#page-7-9) [65\)](#page-9-0).

SUMMARY OF THE CURRENT STATE OF MENINGOCOCCAL URETHRITIS

N. meningitidis is an upper respiratory tract commensal organism that also possesses the ability to cause disease. Asymptomatic oropharyngeal carriage and invasive disease caused by encapsulated strains of meningococcus are more uncommon now than in the prevaccine era, but upper respiratory carriage of unencapsulated N. meningitidis is now more common than in previous times. Over the past decade, an increase in the recognition of meningococcal urethritis cases in heterosexual males in the United States and elsewhere has occurred. A novel, unencapsulated strain of N. meningitidis, US_NmUC, has been implicated as the predominant cause of these cases of meningococcal urethritis. This urethrotropic clade has unique genetic adaptations that make it more adept at causing infection of the urethra [\(Table 3\)](#page-3-1). Symptomatology of meningococcal urethritis is indistinguishable from gonococcal urethritis, but the mechanism of transmission of N. meningitidis is unique in that it is likely nearly exclusively transmitted during oral sex from the normal upper respiratory tract microbiota of the sexual partner to the individual who eventually develops urethritis.

A number of challenges and opportunities for future study of meningococcal urethritis exist. Currently, no routine diagnostic approach is used for the clinical detection of meningococcal urethritis, and no NAAT is FDA authorized for the detection of N. meningitidis from urethral or urine specimens ([Table 1](#page-1-0)). Also, no published studies specifically describe the contemporary prevalence of US_NmUC carriage in the upper respiratory tract, and this clade may be nonspecifically characterized in N . meningitidis carriage studies as simply "unencapsulated." Meningococcal urethritis can occur in monogamous relationships, but this scenario is reportedly uncommon and not well studied. CDC's treatment guidelines for meningococcal urethritis are the same as for gonococcal urethritis, although the importance and evidence of treating sexual partners of those with meningococcal urethritis are unclear. Antimicrobial resistance genetic determinants from N. gonorrhoeae have been identified in N. meningitidis isolates that cause urethritis, which suggests the antimicrobial resistance threat identified in N. gonorrhoeae may overlap N. meningitidis ([28](#page-8-6), [74](#page-9-22)).

REFERENCES

- 1. Seifert HS. 2019. Location, location, location—commensalism, damage and evolution of the pathogenic Neisseria. J Mol Biol 431:3010–3014. <https://doi.org/10.1016/j.jmb.2019.04.007>.
- 2. Weyand NJ. 2017. Neisseria models of infection and persistence in the upper respiratory tract. Pathog Dis 75:ftx031. [https://doi.org/10.1093/femspd/ftx031.](https://doi.org/10.1093/femspd/ftx031)
- 3. Liu G, Tang CM, Exley RMY. 2015. Non-pathogenic Neisseria: members of an abundant, multi-habitat, diverse genus. Microbiology (Reading) 161: 1297–1312. <https://doi.org/10.1099/mic.0.000086>.
- 4. Castelblanco RL, Lee M, Hasbun R. 2014. Epidemiology of bacterial meningitis in the USA from 1997 to 2010: a population-based observational study. Lancet Infect Dis 14:813–819. [https://doi.org/10.1016/S1473-3099\(14\)70805-9.](https://doi.org/10.1016/S1473-3099(14)70805-9)
- 5. Brooks R, Woods CW, Benjamin DK, Jr., Rosenstein NE. 2006. Increased case-fatality rate associated with outbreaks of Neisseria meningitidis infection, compared with sporadic meningococcal disease, in the United States, 1994–2002. Clin Infect Dis 43:49–54. [https://doi.org/10.1086/504804.](https://doi.org/10.1086/504804)
- 6. Kratz MM, Weiss D, Ridpath A, Zucker JR, Geevarughese A, Rakeman J, Varma JK. 2015. Community-based outbreak of Neisseria meningitidis serogroup C infection in men who have sex with men, New York City, New York, USA, 2010–2013. Emerg Infect Dis 21:1379–1386. [https://doi](https://doi.org/10.3201/eid2108.141837) [.org/10.3201/eid2108.141837.](https://doi.org/10.3201/eid2108.141837)
- 7. Unemo M, Shafer WM. 2014. Antimicrobial resistance in Neisseria gonorrhoeae in the 21st century: past, evolution, and future. Clin Microbiol Rev 27:587–613. [https://doi.org/10.1128/CMR.00010-14.](https://doi.org/10.1128/CMR.00010-14)
- 8. St Cyr S, Barbee L, Workowski KA, Bachmann LH, Pham C, Schlanger K, Torrone E, Weinstock H, Kersh EN, Thorpe P. 2020. Update to CDC's treatment guidelines for gonococcal infection, 2020. MMWR Morb Mortal Wkly Rep 69:1911–1916. <https://doi.org/10.15585/mmwr.mm6950a6>.
- 9. Kirkcaldy RD, Weston E, Segurado AC, Hughes G. 2019. Epidemiology of gonorrhea: a global perspective. Sex Health 16:401–411. [https://doi.org/](https://doi.org/10.1071/SH19061) [10.1071/SH19061](https://doi.org/10.1071/SH19061).
- 10. Centers for Disease Control and Prevention. 2021. Urethritis and cervicitis - STI treatment guidelines. [https://www.cdc.gov/std/treatment-guidelines/](https://www.cdc.gov/std/treatment-guidelines/urethritis-and-cervicitis.htm) [urethritis-and-cervicitis.htm.](https://www.cdc.gov/std/treatment-guidelines/urethritis-and-cervicitis.htm) Retrieved 11 March 2022.
- 11. Bazan JA, Turner AN, Kirkcaldy RD, Retchless AC, Kretz CB, Briere E, Tzeng Y-L, Stephens DS, Maierhofer C, Del Rio C, Abrams AJ, Trees DL, Ervin M, Licon DB, Fields KS, Roberts MW, Dennison A, Wang X. 2017. Large cluster of Neisseria meningitidis urethritis in Columbus, Ohio, 2015. Clin Infect Dis 65:92–99. [https://doi.org/10.1093/cid/cix215.](https://doi.org/10.1093/cid/cix215)
- 12. Retchless AC, Kretz CB, Chang H-Y, Bazan JA, Abrams AJ, Norris Turner A, Jenkins LT, Trees DL, Tzeng Y-L, Stephens DS, MacNeil JR, Wang X. 2018.

Expansion of a urethritis-associated Neisseria meningitidis clade in the United States with concurrent acquisition of N. gonorrhoeae alleles. BMC Genomics 19:176. <https://doi.org/10.1186/s12864-018-4560-x>.

- 13. Carpenter CM, Charles R. 1942. Isolation of meningococcus from the genitourinary tract of seven patients. Am J Public Health Nations Health 32: 640–643. <https://doi.org/10.2105/ajph.32.6.640>.
- 14. Janda WM, Bohnoff M, Morello JA, Lerner SA. 1980. Prevalence and site-pathogen studies of Neisseria meningitidis and N gonorrhoeae in homosexual men. JAMA 244:2060–2064. [https://doi.org/10.1001/jama.1980.03310180026026.](https://doi.org/10.1001/jama.1980.03310180026026)
- 15. Carlson BL, Fiumara NJ, Kelly JR, McCormack WM. 1980. Isolation of Neisseria meningitidis from anogenital specimens from homosexual men. Sex Transm Dis 7:71–73. [https://doi.org/10.1097/00007435-198004000-00008.](https://doi.org/10.1097/00007435-198004000-00008)
- 16. Janda WM, Morello JA, Lerner SA, Bohnhoff M. 1983. Characteristics of pathogenic Neisseria spp. isolated from homosexual men. J Clin Microbiol 17:85–91. [https://doi.org/10.1128/jcm.17.1.85-91.1983.](https://doi.org/10.1128/jcm.17.1.85-91.1983)
- 17. Faur YC, Weisburd MH, Wilson ME. 1975. Isolation of Neisseria meningitidis from the genito-urinary tract and anal canal. J Clin Microbiol 2: 178–182. [https://doi.org/10.1128/jcm.2.3.178-182.1975.](https://doi.org/10.1128/jcm.2.3.178-182.1975)
- 18. Maini M, French P, Prince M, Bingham JS. 1992. Urethritis due to Neisseria meningitidis in a London genitourinary medicine clinic population. Int J STD AIDS 3:423–425. <https://doi.org/10.1177/095646249200300604>.
- 19. Hagman M, Forslin L, Moi H, Danielsson D. 1991. Neisseria meningitidis in specimens from urogenital sites. Is increased awareness necessary? Sex Transm Dis 18:228–232. [https://doi.org/10.1097/00007435-199110000-00006.](https://doi.org/10.1097/00007435-199110000-00006)
- 20. Katz AR, Chasnoff R, Komeya A, Lee MVC. 2011. Neisseria meningitidis urethritis: a case report highlighting clinical similarities to and epidemiological differences from gonococcal urethritis. Sex Transm Dis 38: 439–441. <https://doi.org/10.1097/OLQ.0b013e3181ffa7dc>.
- 21. Jannic A, Mammeri H, Larcher L, Descamps V, Tosini W, Phung B, Yazdanpanah Y, Bouscarat F. 2019. Orogenital transmission of Neisseria meningitidis causing acute urethritis in men who have sex with men. Emerg Infect Dis 25:175–176. <https://doi.org/10.3201/eid2501.171102>.
- 22. Nebreda T, Campos A, Merino FJ. 1999. Urethritis caused by Neisseria meningitidis serogroup C. Clin Microbiol Infect 5:57-59. [https://doi.org/](https://doi.org/10.1111/j.1469-0691.1999.tb00101.x) [10.1111/j.1469-0691.1999.tb00101.x.](https://doi.org/10.1111/j.1469-0691.1999.tb00101.x)
- 23. Orden B, Martínez-Ruíz R, González-Manjavacas C, Mombiela T, Millán R. 2004. Meningococcal urethritis in a heterosexual man. Eur J Clin Microbiol Infect Dis 23:646–647. [https://doi.org/10.1007/s10096-004-1178-5.](https://doi.org/10.1007/s10096-004-1178-5)
- 24. Hayakawa K, Itoda I, Shimuta K, Takahashi H, Ohnishi M. 2014. Urethritis caused by novel Neisseria meningitidis serogroup W in man who has sex with men, Japan. Emerg Infect Dis 20:1585–1587. [https://doi.org/10.3201/eid2009.140349.](https://doi.org/10.3201/eid2009.140349)
- 25. Bazan JA, Peterson AS, Kirkcaldy RD, Briere EC, Maierhofer C, Turner AN, Licon DB, Parker N, Dennison A, Ervin M, Johnson L, Weberman B, Hackert P, Wang X, Kretz CB, Abrams AJ, Trees DL, Del Rio C, Stephens DS, Tzeng Y-L, DiOrio M, Roberts MW. 2016. Notes from the field: increase in Neisseria meningitidis– associated urethritis among men at two sentinel clinics — Columbus, Ohio, and Oakland County, Michigan, 2015. MMWR Morb Mortal Wkly Rep 65: 550–552. <https://doi.org/10.15585/mmwr.mm6521a5>.
- 26. Centers for Disease Control and Prevention. Gonococcal isolate surveillance program (GISP) protocol. <https://stacks.cdc.gov/view/cdc/40081>. Retrieved 24 October 2021.
- 27. Toh E, Gangaiah D, Batteiger BE, Williams JA, Arno JN, Tai A, Batteiger TA, Nelson DE. 2017. Neisseria meningitidis ST11 complex isolates associated with nongonococcal urethritis, Indiana, USA, 2015–2016. Emerg Infect Dis 23:336–339. [https://doi.org/10.3201/eid2302.161434.](https://doi.org/10.3201/eid2302.161434)
- 28. Brooks A, Lucidarme J, Campbell H, Campbell L, Fifer H, Gray S, Hughes G, Lekshmi A, Schembri G, Rayment M, Ladhani SN, Ramsay ME, Borrow R. 2020. Detection of the United States Neisseria meningitidis urethritis clade in the United Kingdom, August and December 2019 - emergence of multiple antibiotic resistance calls for vigilance. Euro Surveill 25:2000375. [https://doi.org/10.2807/1560-7917.ES.2020.25.15.2000375.](https://doi.org/10.2807/1560-7917.ES.2020.25.15.2000375)
- 29. Creighton S, Tenant-Flowers M, Taylor CB, Miller R, Low N. 2003. Co-infection with gonorrhoea and chlamydia: how much is there and what does it mean? Int J STD AIDS 14:109–113. [https://doi.org/10.1258/095646203321156872.](https://doi.org/10.1258/095646203321156872)
- 30. Nsuami M, Cammarata CL, Brooks BN, Taylor SN, Martin DH. 2004. Chlamydia and gonorrhea co-occurrence in a high school population. Sex Transm Dis 31:424–427. <https://doi.org/10.1097/01.olq.0000130535.96576.d3>.
- 31. Jolley KA, Bray JE, Maiden MCJ. 2018. Open-access bacterial population genomics: BIGSdb software, the PubMLST.org website and their applications. Wellcome Open Res 3:124. [https://pubmed.ncbi.nlm.nih.gov/30345391/.](https://pubmed.ncbi.nlm.nih.gov/30345391/)
- 32. Lucidarme J, Hill DMC, Bratcher HB, Gray SJ, du Plessis M, Tsang RSW, Vazquez JA, Taha M-K, Ceyhan M, Efron AM, Gorla MC, Findlow J, Jolley KA, Maiden MCJ, Borrow R. 2015. Genomic resolution of an aggressive,
- 33. Urwin R, Maiden MCJ. 2003. Multi-locus sequence typing: a tool for global epidemiology. Trends Microbiol 11:479–487. [https://doi.org/10.1016/j.tim.2003.08.006.](https://doi.org/10.1016/j.tim.2003.08.006)
- 34. Jolley KA, Brehony C, Maiden MCJ. 2007. Molecular typing of meningococci: recommendations for target choice and nomenclature. FEMS Microbiol Rev 31:89–96. [https://doi.org/10.1111/j.1574-6976.2006.00057.x.](https://doi.org/10.1111/j.1574-6976.2006.00057.x)
- 35. Lucidarme J, Comanducci M, Findlow J, Gray SJ, Kaczmarski EB, Guiver M, Vallely PJ, Oster P, Pizza M, Bambini S, Muzzi A, Borrow R. 2010. Characterization of fHbp, nhba (gna2132), nadA, porA, and sequence type in group B meningococcal case isolates collected in England and Wales during January 2008 and potential coverage of an investigational group B meningococcal vaccine. Clin Vaccine Immunol 17:919–929. <https://doi.org/10.1128/CVI.00027-10>.
- 36. Itsko M, Retchless AC, Joseph SJ, Norris Turner A, Bazan JA, Sadji AY, Ouédraogo-Traoré R, Wang X. 2020. Full molecular typing of Neisseria meningitidis directly from clinical specimens for outbreak investigation. J Clin Microbiol 58:e01780-20. <https://doi.org/10.1128/JCM.01780-20>.
- 37. Bratcher HB, Corton C, Jolley KA, Parkhill J, Maiden MCJ. 2014. A geneby-gene population genomics platform: de novo assembly, annotation and genealogical analysis of 108 representative Neisseria meningitidis genomes. BMC Genomics 15:1138. <https://doi.org/10.1186/1471-2164-15-1138>.
- 38. Vogel U, Claus H, Frosch M, Caugant DA. 2000. Molecular basis for distinction of the ET-15 clone within the ET-37 complex of Neisseria meningitidis. J Clin Microbiol 38:941–942. [https://doi.org/10.1128/JCM.38.2.941-942.2000.](https://doi.org/10.1128/JCM.38.2.941-942.2000)
- 39. Tzeng Y-L, Bazan JA, Turner AN, Wang X, Retchless AC, Read TD, Toh E, Nelson DE, Del Rio C, Stephens DS. 2017. Emergence of a new Neisseria meningitidis clonal complex 11 lineage 11.2 clade as an effective urogenital pathogen. Proc Natl Acad Sci U S A 114:4237–4242. [https://doi.org/10](https://doi.org/10.1073/pnas.1620971114) [.1073/pnas.1620971114.](https://doi.org/10.1073/pnas.1620971114)
- 40. Bartley SN, Tzeng Y-L, Heel K, Lee CW, Mowlaboccus S, Seemann T, Lu W, Lin Y-H, Ryan CS, Peacock C, Stephens DS, Davies JK, Kahler CM. 2013. Attachment and invasion of Neisseria meningitidis to host cells is related to surface hydrophobicity, bacterial cell size and capsule. PLoS One 8: e55798. <https://doi.org/10.1371/journal.pone.0055798>.
- 41. Ribet D, Cossart P. 2015. How bacterial pathogens colonize their hosts and invade deeper tissues. Microbes Infect 17:173–183. [https://doi.org/10](https://doi.org/10.1016/j.micinf.2015.01.004) [.1016/j.micinf.2015.01.004](https://doi.org/10.1016/j.micinf.2015.01.004).
- 42. McNeil LK, Zagursky RJ, Lin SL, Murphy E, Zlotnick GW, Hoiseth SK, Jansen KU, Anderson AS. 2013. Role of factor H binding protein in Neisseria meningitidis virulence and its potential as a vaccine candidate to broadly protect against meningococcal disease. Microbiol Mol Biol Rev 77:234–252. [https://doi.org/10.1128/MMBR.00056-12.](https://doi.org/10.1128/MMBR.00056-12)
- 43. Abara WE, Bernstein KT, Lewis FMT, Schillinger JA, Feemster K, Pathela P, Susan H, Islam A, Eberhart M, Cheng I, Ternier A, Slutsker JSS, Mbaeyi S, Madera R, Kirkcaldy RD. 2022. Effectiveness of a serogroup B outer membrane vesicle meningococcal vaccine against gonorrhoea: a retrospective observational study. Lancet Infect Dis 22:P1021–P1029. [https://doi.org/10](https://doi.org/10.1016/S1473-3099(21)00812-4) [.1016/S1473-3099\(21\)00812-4](https://doi.org/10.1016/S1473-3099(21)00812-4).
- 44. Tzeng Y-L, Berman Z, Toh E, Bazan JA, Turner AN, Retchless AC, Wang X, Nelson DE, Stephens DS. 2019. Heteroresistance to the model antimicrobial peptide polymyxin B in the emerging Neisseria meningitidis lineage 11.2 urethritis clade: mutations in the pilMNOPQ operon. Mol Microbiol 111:254–268. [https://doi.org/10.1111/mmi.14153.](https://doi.org/10.1111/mmi.14153)
- 45. Householder TC, Fozo EM, Cardinale JA, Clark VL. 2000. Gonococcal nitric oxide reductase is encoded by a single gene, norB, which is required for anaerobic growth and is induced by nitric oxide. Infect Immun 68: 5241–5246. [https://doi.org/10.1128/IAI.68.9.5241-5246.2000.](https://doi.org/10.1128/IAI.68.9.5241-5246.2000)
- 46. Falsetta ML, McEwan AG, Jennings MP, Apicella MA. 2010. Anaerobic metabolism occurs in the substratum of gonococcal biofilms and may be sustained in part by nitric oxide. Infect Immun 78:2320–2328. [https://doi](https://doi.org/10.1128/IAI.01312-09) [.org/10.1128/IAI.01312-09](https://doi.org/10.1128/IAI.01312-09).
- 47. Ma KC, Unemo M, Jeverica S, Kirkcaldy RD, Takahashi H, Ohnishi M, Grad YH. 2017. Genomic characterization of urethritis-associated Neisseria meningitidis shows that a wide range of N. meningitidis strains can cause urethritis. J Clin Microbiol 55:3374–3383. [https://doi.org/10.1128/JCM.01018-17.](https://doi.org/10.1128/JCM.01018-17)
- 48. McNamara LA. 2017. High risk for invasive meningococcal disease among patients receiving eculizumab (soliris) despite receipt of meningococcal vaccine. MMWR Morb Mortal Wkly Rep 66:734–737. [https://doi.org/10](https://doi.org/10.15585/mmwr.mm6627e1) [.15585/mmwr.mm6627e1](https://doi.org/10.15585/mmwr.mm6627e1).
- 49. Oliver SE, Retchless AC, Blain AE, McNamara LA, Ahrabifard S, Farley M, Weiss D, Zaremski E, Wang X, Hariri S. 2022. Risk factors for invasive meningococcal disease belonging to a novel urethritis clade of Neisseria meningitidis-United States, 2013–2017. Open Forum Infect Dis 9:ofac035. [https://doi.org/10.1093/o](https://doi.org/10.1093/ofid/ofac035)fid/ofac035.
- 50. McNamara LA, Potts CC, Blain A, Topaz N, Apostol M, Alden NB, Petit S, Farley MM, Harrison LH, Triden L, Muse A, Poissant T, Wang X, MacNeil JR. 2019. Invasive meningococcal disease due to nongroupable Neisseria meningitidis-active bacterial core surveillance sites, 2011–2016. Open Forum Infect Dis 6:ofz190. [https://doi.org/10.1093/o](https://doi.org/10.1093/ofid/ofz190)fid/ofz190.
- 51. Fijen CA, Kuijper EJ, Tjia HG, Daha MR, Dankert J. 1994. Complement deficiency predisposes for meningitis due to nongroupable meningococci and Neisseria-related bacteria. Clin Infect Dis 18:780-784. [https://doi.org/](https://doi.org/10.1093/clinids/18.5.780) [10.1093/clinids/18.5.780](https://doi.org/10.1093/clinids/18.5.780).
- 52. Mbaeyi S, Duffy J, McNamara LA. 2021. Pinkbook: meningococcal disease. [https://www.cdc.gov/vaccines/pubs/pinkbook/mening.html.](https://www.cdc.gov/vaccines/pubs/pinkbook/mening.html) Retrieved 11 March 2022.
- 53. U.S. Food and Drug Administration. 2022. Research C for BE and vaccines licensed for use in the United States. [https://www.fda.gov/vaccines-blood-biologics/](https://www.fda.gov/vaccines-blood-biologics/vaccines/vaccines-licensed-use-united-states) [vaccines/vaccines-licensed-use-united-states.](https://www.fda.gov/vaccines-blood-biologics/vaccines/vaccines-licensed-use-united-states) Retrieved 11 March 2022.
- 54. Centers for Disease Control and Prevention. 2022. Meningococcal disease surveillance. <https://www.cdc.gov/meningococcal/surveillance/index.html>. Retrieved 11 March 2022.
- 55. Mbaeyi S, Pondo T, Blain A, Yankey D, Potts C, Cohn A, Hariri S, Shang N, MacNeil JR. 2020. Incidence of meningococcal disease before and after implementation of quadrivalent meningococcal conjugate vaccine in the United States. JAMA Pediatr 174:843-851. [https://doi.org/10.1001/jamapediatrics](https://doi.org/10.1001/jamapediatrics.2020.1990) [.2020.1990](https://doi.org/10.1001/jamapediatrics.2020.1990).
- 56. Kellerman SE, McCombs K, Ray M, Baughman W, Reeves MW, Popovic T, Rosenstein NE, Farley MM, Blake P, Stephens DS, Georgia Emerging Infections Program. 2002. Genotype-specific carriage of Neisseria meningitidis in Georgia counties with hyper- and hyposporadic rates of meningococcal disease. J Infect Dis 186:40–48. [https://doi.org/10.1086/341067.](https://doi.org/10.1086/341067)
- 57. Procop G, Church D, Hall G, Janda WM, Koneman EW, Schreckenberger PC, Woods GL. 2017. Koneman's color atlas and textbook of diagnostic microbiology, 7th ed. Wolters Kluwer Science, Philadelphia, PA.
- 58. Centers for Disease Control and Prevention. 2022. Meningitis lab manual: ID and characterization of Neisseria. [https://www.cdc.gov/meningitis/lab](https://www.cdc.gov/meningitis/lab-manual/chpt07-id-characterization-nm.html) [-manual/chpt07-id-characterization-nm.html.](https://www.cdc.gov/meningitis/lab-manual/chpt07-id-characterization-nm.html) Retrieved 4 December 2021.
- 59. Alexander S, Ison C. 2005. Evaluation of commercial kits for the identification of Neisseria gonorrhoeae. J Med Microbiol 54:827–831. [https://doi](https://doi.org/10.1099/jmm.0.46108-0) [.org/10.1099/jmm.0.46108-0](https://doi.org/10.1099/jmm.0.46108-0).
- 60. Deak E, Green N, Humphries RM. 2014. Microbiology test reliability in differentiation of Neisseria meningitidis and Neisseria polysaccharea. J Clin Microbiol 52:3496. [https://doi.org/10.1128/JCM.01703-14.](https://doi.org/10.1128/JCM.01703-14)
- 61. Cunningham SA, Mainella JM, Patel R. 2014. Misidentification of Neisseria polysaccharea as Neisseria meningitidis with the use of matrix-assisted laser desorption ionization-time of flight mass spectrometry. J Clin Microbiol 52:2270–2271. <https://doi.org/10.1128/JCM.00664-14>.
- 62. Pandey U, Naccache SN, Dien Bard J. 2019. Back to the Basics: biochemical testing for pathogen identification in the era of matrix-assisted laser desorption ionization-time of flight mass spectrometry (MALDI-TOF MS). J Clin Microbiol 57:e00498-19. <https://doi.org/10.1128/JCM.00498-19>.
- 63. Unalan-Altintop T, Karagoz A, Hazirolan G. 2020. A diagnostic challenge in clinical laboratory: misidentification of Neisseria subflava as Neisseria meningitidis by MALDI-TOF MS. Acta Microbiol Immunol Hung 67: 258–260. [https://doi.org/10.1556/030.2020.01039.](https://doi.org/10.1556/030.2020.01039)
- 64. Centers for Disease Control and Prevention. 2014. Recommendations for the laboratory-based detection of Chlamydia trachomatis and Neisseria gonorrhoeae–2014. MMWR Recomm Rep 63:1–19.
- 65. Sukhum KV, Jean S, Wallace M, Anderson N, Burnham CA, Dantas G. 2021. Genomic characterization of emerging bacterial uropathogen Neisseria meningitidis, which was misidentified as Neisseria gonorrhoeae by nucleic acid amplification testing. J Clin Microbiol 59:e01699-20. [https://](https://doi.org/10.1128/JCM.01699-20) [doi.org/10.1128/JCM.01699-20.](https://doi.org/10.1128/JCM.01699-20)
- 66. Walcher M, Skvoretz R, Montgomery-Fullerton M, Jonas V, Brentano S. 2013. Description of an unusual Neisseria meningitidis isolate containing and expressing Neisseria gonorrhoeae-specific 16S rRNA gene sequences. J Clin Microbiol 51:3199–3206. [https://doi.org/10.1128/JCM.00309-13.](https://doi.org/10.1128/JCM.00309-13)
- 67. Toh E, Williams JA, Qadadri B, Ermel A, Nelson DE. 2020. Development of a SimpleProbe real-time PCR assay for rapid detection and identification of the US novel urethrotropic clade of Neisseria meningitidis ST-11 (US_NmUC). PLoS One 15:e0228467. [https://doi.org/10.1371/journal.pone.0228467.](https://doi.org/10.1371/journal.pone.0228467)
- 68. Bazan JA, Stephens DS, Turner AN. 2021. Emergence of a novel urogenital-tropic Neisseria meningitidis. Curr Opin Infect Dis 34:34–39. [https://](https://doi.org/10.1097/QCO.0000000000000697) doi.org/10.1097/QCO.0000000000000697.
- 69. Workowski KA, Bachmann LH, Chan PA, Johnston CM, Muzny CA, Park I, Reno H, Zenilman JM, Bolan GA. 2021. Sexually transmitted infections treatment guidelines, 2021. MMWR Recomm Rep 70:192. [https://doi.org/](https://doi.org/10.15585/mmwr.rr7004a1) [10.15585/mmwr.rr7004a1](https://doi.org/10.15585/mmwr.rr7004a1).
- 70. Bazan JA, Tzeng Y-L, Stephens DS, Carter AM, Brown MA, Snyder B, Prince DJ, Turner AN. 2020. Repeat episodes of symptomatic urethritis due to a uropathogenic meningococcal clade. Sex Transm Dis 47:e1–e4. [https://](https://doi.org/10.1097/OLQ.0000000000001079) [doi.org/10.1097/OLQ.0000000000001079.](https://doi.org/10.1097/OLQ.0000000000001079)
- 71. McNamara LA. 2020. Detection of ciprofloxacin-resistant, β -lactamaseproducing Neisseria meningitidis serogroup Y isolates - United States, 2019–2020. MMWR Morb Mortal Wkly Rep 69:735–739. [https://doi.org/10](https://doi.org/10.15585/mmwr.mm6924a2) [.15585/mmwr.mm6924a2](https://doi.org/10.15585/mmwr.mm6924a2).
- 72. Kirkcaldy RD, Harvey A, Papp JR, Del Rio C, Soge OO, Holmes KK, Hook EW, Kubin G, Riedel S, Zenilman J, Pettus K, Sanders T, Sharpe S, Torrone E. 2016. Neisseria gonorrhoeae antimicrobial susceptibility surveillance - The Gonococcal Isolate Surveillance Project, 27 sites, United States, 2014. MMWR Surveill Summ 65:1–19. <https://doi.org/10.15585/mmwr.ss6507a1>.
- 73. Centers for Disease Control and Prevention. 2021. Combating the threat of antibiotic-resistant gonorrhea. [https://www.cdc.gov/std/gonorrhea/](https://www.cdc.gov/std/gonorrhea/arg/carb.htm) [arg/carb.htm](https://www.cdc.gov/std/gonorrhea/arg/carb.htm). Accessed 27 March 2022.
- 74. Centers for Disease Control and Prevention. 2019. Antibiotic resistance threats in the United States, 2019. Centers for Disease Control and Prevention, Atlanta, GA. <https://doi.org/10.15620/cdc:82532>.