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Rare variants in genes encoding subunits of the epithelial Na⁺ channel are associated with blood pressure and kidney function

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Abstract

Background: The epithelial Na⁺ channel (ENaC) is intrinsically linked to fluid volume homeostasis and blood pressure. Specific rare mutations in *SCNN1A*, *SCNN1B*, and *SCNN1G*, genes encoding the α , β , and γ subunits of ENaC, respectively, are associated with extreme blood pressure phenotypes. No associations between blood pressure and *SCNN1D*, which encodes the δ subunit of ENaC, have been reported. A small number of sequence variants in ENaC subunits have been reported to affect functional transport *in vitro* and/or blood pressure. The effects of the vast majority of rare and low frequency ENaC variants on blood pressure are not known.

Methods: We explored the association of low frequency and rare variants in the genes encoding ENaC subunits, with systolic blood pressure (SBP), diastolic blood pressure (DBP), mean arterial pressure (MAP), and pulse pressure (PP). Using whole-genome sequencing data from fourteen studies participating in the TOPMed Whole-Genome Sequencing Program and sequence kernel association tests.

Results: We found that variants in *SCNN1A* and *SCNN1B* were associated with DBP and MAP (p < 0.00625). Although *SCNN1D* is poorly expressed in human kidney tissue, *SCNN1D* variants were associated with SBP, DBP, MAP, and PP (p < 0.00625). ENaC variants in two of the four subunits (*SCNN1B*, and *SCNN1D*) were also associated with estimated glomerular filtration rate (p < 0.00625), but not with stroke.

Conclusions: Our results suggest that variants in extrarenal ENaCs, in addition to ENaCs expressed in kidneys, influence blood pressure and kidney function.

Graphical Abstract



Keywords

ENaC; blood pressure; estimated glomerular filtration rate; stroke; gene variants

Introduction

The regulated absorption of filtered Na⁺ along the nephron helps govern extracellular fluid volume and blood pressure. The aldosterone-sensitive distal nephron (ASDN) has a critical role in Na⁺ absorption and is where key volume regulatory hormones and signaling pathways affect the absorption of filtered Na⁺. The epithelial Na⁺ channel (ENaC) is expressed in the distal aspects of the ASDN where it mediates the absorption of filtered Na⁺ across the luminal membrane of principal cells^{1,2}. In addition to its role in Na⁺ absorption, ENaC function is required for K⁺ secretion in the ASDN^{1,3}. ENaCs are also found at other tissues that affect total body Na⁺ and blood pressure. It functions as a salt sensor in lingual

epithelia and influences salt intake, whereas expression in the distal colon has a role, albeit minor, in absorption of ingested Na^{+4,5}. ENaC expression in antigen presenting immune cells has been proposed to facilitate release of cytokines in response to increased salt intake, which contribute to an increase in blood pressure^{6,7}. ENaCs in vascular smooth muscle and endothelial cells may influence vascular tone^{7,8}. Finally, ENaCs are expressed at specific sites in the central nervous system where they influence autonomic tone and blood pressure⁹.

ENaCs are formed from structurally related subunits, termed α , β , γ , and δ , which are encoded by the genes SCNN1A, SCNN1B, SCNN1G, and SCNN1D, respectively. These are Na⁺ selective channels that are primarily $\alpha\beta\gamma$ or $\delta\beta\gamma$ heterotrimers, which exhibit differences in functional and regulatory properties². ENaCs in human kidney are primarily a $\alpha\beta\gamma$ heterotrimer¹⁰, while ENaCs containing the δ subunit are expressed in other tissues^{11,12}. Each ENaC subunit has two transmembrane domains. The second transmembrane domain from each subunit within a channel complex contributes to the channel pore, cation selectivity filter, and gate. The transmembrane domains are connected by a large, structurally complex extracellular domain that functions as a sensor, where specific extracellular factors, including monovalent cations and anions (Na⁺, Cl⁻, H⁺), peptides, proteases, and shear stress interact with the extracellular domain to regulate channel activity². Short cytoplasmic amino (N)- and carboxyl (C)-termini also have key regulatory sites. For example, a Pro-Tyr (PY) motif in the cytoplasmic C-termini of ENaC subunits is a binding site for the ubiquitin ligase NEDD4-2 that facilitates channel ubiquitination at the cell surface and subsequent internalization¹³. The cytoplasmic Ntermini of ENaC subunits have a His-Gly motif that affects channel gating¹⁴.

Several rare ENaC variants that have large effects on blood pressure with Mendelian inheritance have been identified in *SCNN1A*, *SCNN1B*, and *SCNN1G*. These disorders include (i) Liddle syndrome, an autosomal dominant disorder where specific gain-of-function mutations in *SCNN1B*, and *SCNN1G* are associated with hypertension and hypokalemia, and (ii) pseudohypoaldosteronism type I (PHA1), an autosomal recessive disorder where specific loss-of-function mutations in *SCNN1B*, or *SCNN1G* are associated with hypotension and hyperkalemia¹⁵. Liddle syndrome mutations primarily result in disruption of the PY motif, significantly increasing channel residency time at the cell surface¹³. PHA1 mutations have been described that result in specific deletions or amino acid substitutions that cause a profound loss of function *in vitro*^{14,15}.

Our group and others have identified a growing number of sites, including rare single nucleotide variants (SNVs) within the extracellular domains, where mutations affect channel gating activity^{2,12}. This is congruent with the extracellular domains' role in sensing extracellular factors and regulating channel gating in response to these factors. For most variants that alter ENaC function and are not associated with Liddle syndrome or PHA1, it is unclear whether they influence blood pressure, serum [K⁺], or the prevalence of blood pressure-associated disorders including stroke, myocardial infarction, and chronic kidney disease in specific populations.

Using whole-genome sequencing (WGS) and phenotype data available through the Trans-Omics in Precision Medicine (TOPMed) Whole-Genome Sequencing Project, we examined

common functional human ENaC variants and arrays of low-frequency and rare ENaC variants for association with blood pressure levels and related traits and health outcomes.

Materials and Methods

WGS and harmonized blood pressure phenotype data were available for analysis from 28,058 participants in fourteen studies from TOPMed (selected from the > 142,000 individuals in forty-one studies in TOPMed) (Table 1). After excluding 300 individuals < 20 years old or > 90 years old and including 641 individuals from the Samoan Soifua Manuia Study that were not included in the TOPMed cohort, 28,399 individuals from the harmonized blood pressure phenotype dataset were included in our main (blood pressure) analyses. Subsets of this sample were included in our secondary analyses: 9,090 for total strokes (4,399 cases and 4,691 controls), and 14,557 for eGFR. The individuals included in our main analyses were from six self-reported ancestry groups (58.3% European, 29.9% African, 6.3% Asian, 4.1% Samoan, 0.2% Native American, and 1.2% other) (Table S2). To account for underlying population substructure, we used principal components of ancestry (PCAs) calculated by TOPMed. Based on the spaghetti plot (Figure S1), we concluded that PCAs 1–11 accounted for the vast majority of inter-ancestry variance in our population and should be included as covariates in our analyses. Relatedness in the analysis cohorts was controlled for using a genetic relatedness matrix (GRM) calculated by TOPMed.

We conducted single variant analyses for three common (minor allele frequency (MAF) 0.05) ENaC variants, two of which are functional variants^{16,17}. We also performed sequence kernel association tests (SKAT) and burden tests for all low-frequency and rare variants (MAF < 0.05), as well as separate SKAT for rare (MAF < 0.01) and low-frequency (0.01)

MAF < 0.05) SNVs within the genomic regions of *SCNN1A*, *SCNN1B*, *SCNN1D*, and *SCNN1G*. In each analysis, we have tested for association with systolic blood pressure (SBP), diastolic blood pressure (DBP), pulse pressure (PP), mean arterial pressure (MAP), estimated glomerular filtration rate (eGFR), and stroke.

Finally, we assessed *SCNN1D* expression in monocytes from eleven healthy patients. Monocytes isolated from heparinized blood were cultured in media containing either 150 mM NaCl or 190 mM NaCl for 72 hours. Total RNA isolated from the monocytes was sequenced, and expression analysis was performed to determine whether *SCNN1D* was expressed in those cells.

Additional methods details are presented in the Supplemental Materials^{18–27}.

Results

Three common functional ENaC variants— α A334T, α T663A, and β G165R—are not associated with SBP, DBP, MAP, or PP

Two common ENaC a subunit variants, aA334T (rs11542844, MAF = 0.17) and aT663A (rs2228576, MAF = 0.27) alter channel function in heterologous expression systems^{16,17}, although previous studies largely suggest that these variants do not affect blood pressure phenotypes^{28–30}. There is also a common ENaC variant in the β subunit, β G165R

(rs2303157, MAF = 0.24), although its effect on ENaC function has not been described. To account for the impact of these common variants in our analyses of low frequency variants, we tested whether any of these common variants were associated with blood pressure phenotypes. We conducted single-variant analyses for the association of each variant with SBP, DBP, MAP and PP using age, sex, BMI, study, and the first eleven PCAs as covariates and controlled for relatedness with a GRM. No association was found between any of the variants and SBP, DBP, MAP, or PP (Table 2). These results are largely consistent with previous findings for α A334T and α T663A^{16,17,28–30}.

Low frequency and rare ENaC variation in specific subunits is, in aggregate, associated with SBP, DBP, MAP or PP

We performed SKAT to determine the impact of aggregated low frequency (MAF < 0.05) variation within the genomic regions of SCNN1A, SCNN1B, SCNN1D, and SCNN1G on four blood pressure measures (SBP, DBP, PP, and MAP) using age, sex, BMI, and PCAs 1-11 as covariates, and controlled for relatedness using a GRM. The threshold for significance, based on the number of genes tested, was 0.00625 (see Methods). There were 3,972 SCNN1A, 12,334 SCNN1B, 2,405 SCNN1D, and 4,093 SCNN1G variants with a MAF < 0.05 within the promoter regions and gene boundaries of each gene that also had genotype missingness rates < 0.15. These variants present across the 14 studies were included in the analyses. There was a significant association of SCNNIA aggregate variants with DBP (p = 0.00004) and MAP (p = 0.0002), but not with SBP or PP (Table 4). SCNNIB variants were significantly associated with DBP (p = 0.002) and MAP (p = 0.003). SCNN1D variants were significantly associated with all of four blood pressure phenotypes: SBP (p =0.0008), DBP (p = 0.002), PP (p = 0.003), and MAP (p = 0.0008). Finally, those at SCNN1G were not significantly associated with any of the four blood pressure phenotypes. As a negative control, DBP was permuted by reassignment at random in the population and no associations were observed between any of the subunit genes and the permuted phenotype values. We also performed SKAT with the combined variants from SCNN1A, SCNN1B, and SCNN1G (20,399 in total) and each of the four blood pressure measures. Because of the associations with SBP, DBP, PP, and MAP, SCNN1D variants were not included in the combined analyses of blood pressure phenotypes to avoid biasing the results. The combined analysis showed association with both DBP (p = 0.0002) and MAP (p = 0.0006, see Table 3).

We repeated the analyses with rare (MAF < 0.01) and low frequency (0.01 MAF < 0.05) variants separately to determine if one class of variants was driving the significance of our findings. In our analyses of rare variants: *SCNN1A* (3,859 variants) were significantly associated with DBP (p = 0.0004) and MAP (p = 0.002); *SCNN1B* (11,925 variants) were also associated with DBP (p = 0.003) and MAP (p = 0.006); the combined analysis of *SCNN1A*, *SCNN1B*, and *SCNN1G* (19,806 variants) was associated with DBP (p = 0.0004) and MAP (p = 0.006); the combined analysis of *SCNN1A*, *SCNN1B*, and *SCNN1G* (19,806 variants) was associated with DBP (p = 0.0004) and MAP (p = 0.001); and the combined analysis of *SCNN1A*, *SCNN1B*, *SCNN1D*, and *SCNN1G* (22,099 variants) was associated with DBP (p = 0.0002) and MAP (p = 0.0002) and MAP (p = 0.0009). Neither *SCNN1D* (2,293 variants) nor *SCNN1G* (4,022 variants) were associated with any of the four phenotypes (See Table 3).

In our analyses of low frequency variants: *SCNN1A* (113 variants) were significantly associated with DBP (p = 0.0007) and MAP (p = 0.002); *SCNN1D* (112 variants) were associated with SPP (p = 0.0005) DPP (p = 0.002); *DPP* (p = 0.002); *scnN1D* (112 variants) were

associated with SBP (p = 0.0005), DBP (p = 0.003), PP (p = 0.005), and MAP (p = 0.0007); the combined analysis of *SCNN1A*, *SCNN1B*, and *SCNN1G* (593 variants) were associated with DBP (p = 0.004) and MAP (p = 0.005); and the combined analysis of *SCNN1A*, *SCNN1B*, *SCNN1D*, and *SCNN1G* (705 variants) were associated with SBP (p = 0.006), DBP (p = 0.001), and MAP (p = 0.001). Neither *SCNN1B* (409 variants) nor *SCNN1G* (71 variants) were associated with any of the four phenotypes (See Table 3).

We repeated these analyses using another gene-based test, the burden test, and found no association between *SCNN1A*, *SCNN1B*, *SCNN1D*, or *SCNN1G* and SBP, DBP, MAP, or PP, suggesting that our analyses include variants that are increasing blood pressure as well as variants that are decreasing blood pressure (Table S3). We also performed SKAT analyses on *SCNN1A* (MAF < 0.05), *SCNN1B* (MAF < 0.05), and *SCNN1D* (0.01 MAF 0.05) upstream, downstream, missense, synonymous, and intronic variants, separately, with DBP to determine if any class of variant was contributing to our results. We found no association between DBP and *SCNN1A*, *SCNN1B*, or *SCNN1D* upstream, downstream, missense, synonymous, and intronic variants, missense, synonymous, and intronic variants.

Low frequency and rare ENaC variation in specific subunits is, in aggregate, associated with eGFR

High blood pressure is associated with chronic kidney disease (CKD) and stroke^{31,32}. ENaC variants associated with altered blood pressure may also be associated with health outcomes such as stroke, eGFR and CKD. We hypothesized that ENaC variants are associated with eGFR and that SBP and DBP mediate these associations. Stepwise regression indicated that DBP was contributing to the variance of the null model for eGFR and therefore, we included DBP as a covariate in our eGFR analyses. We performed SKAT analyses using data from 14,557 individuals from seven TOPMed studies (Tables S5 and S6) to test for associations between eGFR (from which CKD is diagnosed) and variants with MAF < 0.05 in SCNN1A, SCNN1B, SCNN1D, and SCNN1G. In addition to DBP, models were adjusted for age, sex, BMI, and PCs 1–11 as covariates. We identified associations between eGFR and SCNN1B (n = 8,770) and SCNN1D (n = 1,758) variants (p = 0.005, p = 0.002, respectively), but not the SCNN1A (n = 2,739) and SCNN1G (n = 2,868) variants included in the analyses (p =0.01, and p = 0.1, respectively) (Table 4). When combining ENaC variants (MAF < 0.05) from all four subunits (16,135 variants), we observed a significant association between the variants and eGFR (p=0.0003). As a negative control, eGFR phenotypes were permuted to randomize eGFR across samples and the analyses were repeated. There were no associations with permuted eGFR.

We repeated these analyses separately with rare and low frequency variants. Only low frequency variants in *SCNN1D* (110 variants) were significantly associated with eGFR (p = 0.001). There was no association between eGFR and rare variants in individual subunits, *SCNN1A* (2,626 variants), *SCNN1B* (8,365 variants), *SCNN1D* (1,648 variants), and *SCNN1G* (2,797 variants) (p = 0.008, p = 0.007, p = 0.2, and p = 0.01, respectively, see Table 4). However, when combining ENaC variants of all four subunits (15,436 variants)

we observed significant associations between rare variants and low frequency variants and eGFR (p = 0.002 and 0.003, respectively).

These analyses were repeated using burden tests. Of the conditions tested, only *SCNN1D* low frequency variants (0.01 MAF < 0.05) were significantly associated with eGFR, suggesting strong directionality of effect for these variants (Table S7).

Low-frequency and rare variation in SCNN1A, SCNN1B, SCNN1D, and SCNN1G is not associated with stroke

We also performed SKAT analyses using data from a subset of individuals from the blood pressure analyses consisting of 9,090 individuals (4,399 cases and 4,691 controls) from the Women's Health initiative (Tables S8 and S9) to test for associations between stroke and low frequency and rare variation (MAF < 0.05) in SCNN1A, SCNN1B, SCNN1D, and SCNN1G using age, sex, BMI, and PCAs 1-11 as covariates. After removing variants monomorphic in this subset, 2,330 SCNN1A variants 6,965 SCNN1B variants, 1,621 SCNN1D, and 2,208 SCNN1G variants remained in the analyses. There was no significant association of combined variants with MAF < 0.05 in SCNN1A, SCNN1B, SCNN1D, or SCNN1G with overall stroke (p = 0.08, p = 0.5, p = 0.5, and p = 0.2, respectively) (Table 5). Additionally, we performed SKAT with 13,124 variants from SCNN1A, SCNN1B, SCNN1D, and SCNN1G combined. The combined ENaC variation SKAT was not associated with stroke (p = 0.3). We repeated these analyses with rare and low frequency variants separately. There were no associations between SCNN1A (2,216 and 114 variants, respectively), SCNN1B (6,556 and 409 variants, respectively), SCNN1D (1,497 and 124 variants, respectively), SCNN1G (2,137 and 71 variants, respectively), or all four subunits combined (12,406 and 718 variants, respectively) (See Table 5). We repeated these analyses using burden tests, and found no association between SCNN1A, SCNN1B, SCNN1D, or SCNN1G and stroke (Table S10).

SCNN1D is expressed in human monocytes

The ENaC δ subunit is poorly expressed in transporting epithelia that regulate total body Na⁺ (kidney and colon)^{10,33}, in contrast to the α , β , and γ subunits. Our results showing association of *SCNN1D* low-frequency and rare variants with blood pressure phenotypes, and not *SCNN1G*, suggest that ENaC function in Na⁺ sensing cells influence blood pressure variation in the general population. *SCNN1D* is expressed in the central nervous system, where rare variants could influence blood pressure parameters. Antigen-presenting cells have recently been reported to have an ENaC-dependent response to increased Na⁺ intake, leading to an increase in blood pressure⁶. We previously showed that *SCNN1A* and *SCNN1G*, but not *SCNN1D* may have a role in Na⁺ sensing in human monocytes and monocyte-derived cells, we performed RNA sequencing on human monocytes isolated from 11 volunteers and exposed to either normal or elevated sodium. We detected *SCNN1D* expression in cells from each of the volunteers (Figure 1). We did not detect an effect of elevated sodium exposure on *SCNN1D* expression in these cells. Nonetheless, expression of *SCNN1D* suggests the δ subunit of ENaC may be involved in Na⁺ sensing in these cells.

Discussion

The consequences of variants that lead to extreme increases or decreases in ENaC activity, which manifest as Liddle syndrome and PHA1, respectively, are well known monogenic disorders. There are few studies examining the effects of other ENaC variants on complex traits such as blood pressure variation or hypertension-related health outcomes. We found that, in aggregate, low frequency and rare variants in SCNN1A and SCNN1B are associated with DBP and MAP, with both rare and low frequency variants contributing to the associations of SCNN1A with DBP and MAP and rare variants driving the associations of SCNN1B with MAP and DBP. Variants in SCNN1D are associated with SBP, DBP, PP, and MAP, and are driven by low-frequency variants. Variants in SCNN1B and SCNN1D are associated with eGFR, which, for SCNN1D, is, again, driven by low frequency variants. While there has been speculation about the impact of common and rare functional ENaC variants on blood pressure^{7,34}, we found no evidence that these common functional variants (aA334T, aT663A) are associated with SBP, DBP, PP, or MAP, consistent with the evidence provided by ClinVar³⁵. These results are largely in agreement with previous findings^{16,17,28–30}. We did not find an association of low frequency and rare variants in SCNN1A, SCNN1B, SCNN1D, and SCNN1G, assessed either separately or together, with stroke, despite the associations of hypertension with stroke 31,32 . Potential reasons for the lack of associations are small effect of these combined variants on blood pressure variation or heterogeneity of effects across ancestries that may have reduce power to detect effects, since these outcomes are likely mediated through increases in blood pressure. In addition, the SKAT test combines variants that increase and decrease blood pressure and therefore does not provide estimates for associations. As analyses of a large database may result in spurious results, and it will be important to confirm our key findings using other large databases.

ENaCs in human kidney are primarily $\alpha\beta\gamma$ channels¹⁰. We expected to find associations of low frequency and rare variants in SCNN1A, SCNN1B, and SCNN1G with blood pressure traits based on the known roles of ENaC in regulating the reabsorption of filtered Na⁺ in the ASDN and whole-body Na⁺ content, which are major determinants of extracellular fluid volume and blood pressure^{1,2}. ENaC-dependent Na⁺ absorption also has an important role in regulating serum [K⁺], as it is tightly coupled to renal K⁺ secretion mediated by the K⁺ channels Kir1.1 and BK^{1,3}. A lower serum [K⁺] in the setting of increased in ENaC activity, coupled with increased K⁺ secretion, is predicted to enhance activity of the Na⁺-Cl⁻ cotransporter (NCC) in the DCT³⁶. It was surprising that variants in SCNN1G were not associated with blood pressure traits, given what is known regarding the roles of specific sites in the γ subunit in regulating channel activity. We and others have identified sites in the γ subunit where specific mutations affect ENaC activity in heterologous expression systems^{2,37,38}. Cleavage of the γ subunit at defined sites, with the release of an imbedded inhibitory tract, has a large effect on channel activity. Modification of specific γ subunit cytoplasmic residues by palmitoylation, and the interaction of specific acidic phospholipids with γ subunit residues also affect channel activity².

ENaC function in non-renal tissues may drive the blood pressure associations we observed. Among these are sites in humans that express channels with a δ subunit. We found by

RNA sequencing that human monocytes express ENaC δ subunits (Figure 2), which could contribute to the release of cytokines that affect blood pressure⁶. The δ subunit is expressed in human umbilical vein endothelial cells³⁹. Previous studies have suggested that ENaC function in endothelial cells influence cellular stiffness and NO release^{7,8}, raising the possibility that SCNN1D variants could influence blood pressure. Recently, Paudel, et al. identified SCNN1A, SCNN1B, SCNN1D, and SCNN1G mRNA and protein expression in human internal mammary artery and aorta and suggested that SCNN1D may be associated with hypertension⁴⁰. ENaC δ subunits are also expressed in human taste buds and may have a role in mediating salt taste⁴ and influence Na⁺ intake. This could influence blood pressure by a number of mechanisms, including changes in extracellular and intravascular volume, release of cytokines from circulating dendritic cells⁶ and monocytes, and through alterations in the gut microbiota and the subsequent induction of T_H17 cells⁴¹. Our observed association of low frequency and rare variants in SCNN1D with blood pressures suggest that the δ subunit has an unrecognized role in blood pressure regulation in human. This is consistent with a wider scope of SCNN1D expression as previously documented¹², based on the recently released encyclopedias of DNA elements (ENCODE 3) data¹¹. Expression of *SCNN1D* in B cells, T cells and monocytes suggests a potential role of the δ subunit in immunity and immunity-mediated diseases.

Aside from the well-defined Liddle syndrome variants, it is notable that few studies, to date, have found an association between low-frequency or rare ENaC variants and blood pressure measures. Five ENaC variants associated with increases or decreases in blood pressure salt-sensitivity were observed in the GenSalt study⁴². In addition, seven functional ENaC missense variants in the GenSalt study were identified by Ray et al.³⁷, although these were not associated with differences in salt-sensitivity. Other variants have been associated with hypertension in specific populations, including variants in *SCNN1B* and *SCNN1G*^{43–45}. Additionally, an ENaC gain-of-function variant in *SCNN1A* was associated with a modest Liddle syndrome-like phenotype and a blunting of the inhibitory effect of extracellular Na⁺ 2,46,47.

We noted that variants in *SCNN1B* and *SCNN1D* are associated with eGFR. While DBP is contributing to the variance of eGFR in our population, we controlled for DBP in our eGFR analyses and the associations of *SCNN1B* with eGFR are independent of DBP. The association with *SCNN1B* could reflect the effects of variants on ENaC function in the ASDN. Changes in extracellular fluid volume, blood pressure or volume regulatory hormones may influence glomerular filtration rate (GFR). Also, connecting tubule/collecting duct tubuloglomerular feedback, a phenomenon where ENaC dependent Na⁺ transport in the ASDN influences renal afferent arterial tone⁴⁸ may be altered by ENaC variants. As mentioned above, associations with *SCNN1D* suggest that extrarenal ENaC influences GFR. Vascular ENaC influences renal vascular tone and blood flow, factors that will affect GFR⁴⁹. ENaC-dependent release of cytokines from circulating dendritic cells⁶ and monocytes may also affect glomerular function⁵⁰. Low-frequency variation (0.01 MAF < 0.05) in *SCNN1D* was also associated with eGFR via burden test. This may be caused by many of the variants exerting an effect in the same direction on eGFR, or one or several low-frequency variants with strong and similar directional effects on eGFR. These

possibilities are not mutually exclusive and will require future study to elucidate the impact of *SCNN1D* variants on eGFR.

Perspectives

Low frequency and rare variants in *SCNN1A*, *SCNN1B*, and *SCNN1D* are, in aggregate, associated with key blood pressure parameters and eGFR in TOPMed data. The association of *SCNN1D* variants with BP and eGFR highlight the importance of ENaCs outside of the nephron in regulating these physiologic parameters. These observations raise the possibility that humans with hypertension and specific non–Liddle ENaC variants, particularly variants with a gain-of function phenotype, may benefit from a trial of ENaC inhibitors, such as amiloride, to lower blood pressure.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Non standard abbreviations:

ASDN	aldosterone-sensitive distal nephron
DBP	diastolic blood pressure
ENaC	epithelial Na ⁺ channel
eGFR	estimated glomerular filtration rate
GRM	genetic relatedness matrix
МАР	mean arterial pressure
MAF	minor allele frequency
PCA	principal components of ancestry
PHA1	pseudohypoaldosteronism type I
PP	pulse pressure
SKAT	sequence kernel association tests
SNVs	single nucleotide variants

SBP	systolic blood pressure
TOPMed	Trans-Omics in Precision Medicine
WGS	whole-genome sequencing

References

- Pearce D, Soundararajan R, Trimpert C, Kashlan OB, Deen PM, Kohan DE. Collecting duct principal cell transport processes and their regulation. Clin J Am Soc Nephrol. 2015;10:135–146. doi: 10.2215/CJN.05760513 [PubMed: 24875192]
- Kleyman TR, Eaton DC. Regulating ENaC's gate. Am J Physiol Cell Physiol. 2020;318:C150– C162. doi: 10.1152/ajpcell.00418.2019 [PubMed: 31721612]
- Carrisoza-Gaytan R, Ray EC, Flores D, Marciszyn AL, Wu P, Liu L, Subramanya AR, Wang W, Sheng S, Nkashama LJ, et al. Intercalated cell BKalpha subunit is required for flow-induced K+ secretion. JCI Insight. 2020;5. doi: 10.1172/jci.insight.130553
- 4. Xu JJ, Elkaddi N, Garcia-Blanco A, Spielman AI, Bachmanov AA, Chung HY, Ozdener MH. Arginyl dipeptides increase the frequency of NaCl-elicited responses via epithelial sodium channel alpha and delta subunits in cultured human fungiform taste papillae cells. Sci Rep. 2017;7:7483. doi: 10.1038/s41598-017-07756-x [PubMed: 28790369]
- Malsure S, Wang Q, Charles RP, Sergi C, Perrier R, Christensen BM, Maillard M, Rossier BC, Hummler E. Colon-specific deletion of epithelial sodium channel causes sodium loss and aldosterone resistance. J Am Soc Nephrol. 2014;25:1453–1464. doi: 10.1681/ASN.2013090936 [PubMed: 24480829]
- Barbaro NR, Foss JD, Kryshtal DO, Tsyba N, Kumaresan S, Xiao L, Mernaugh RL, Itani HA, Loperena R, Chen W, et al. Dendritic Cell Amiloride-Sensitive Channels Mediate Sodium-Induced Inflammation and Hypertension. Cell Rep. 2017;21:1009–1020. doi: 10.1016/j.celrep.2017.10.002 [PubMed: 29069584]
- Mutchler SM, Kleyman TR. New insights regarding epithelial Na+ channel regulation and its role in the kidney, immune system and vasculature. Curr Opin Nephrol Hypertens. 2019;28:113–119. doi: 10.1097/MNH.0000000000000479 [PubMed: 30585851]
- Warnock DG, Kusche-Vihrog K, Tarjus A, Sheng S, Oberleithner H, Kleyman TR, Jaisser F. Blood pressure and amiloride-sensitive sodium channels in vascular and renal cells. Nat Rev Nephrol. 2014;10:146–157. doi: 10.1038/nrneph.2013.275 [PubMed: 24419567]
- Lu J, Wang HW, Ahmad M, Keshtkar-Jahromi M, Blaustein MP, Hamlyn JM, Leenen FHH. Central and peripheral slow-pressor mechanisms contributing to Angiotensin II-salt hypertension in rats. Cardiovasc Res. 2018;114:233–246. doi: 10.1093/cvr/cvx214 [PubMed: 29126194]
- Menon R, Otto EA, Hoover P, Eddy S, Mariani L, Godfrey B, Berthier CC, Eichinger F, Subramanian L, Harder J, et al. Single cell transcriptomics identifies focal segmental glomerulosclerosis remission endothelial biomarker. JCI Insight. 2020;5. doi: 10.1172/ jci.insight.133267
- Consortium EP, Moore JE, Purcaro MJ, Pratt HE, Epstein CB, Shoresh N, Adrian J, Kawli T, Davis CA, Dobin A, et al. Expanded encyclopaedias of DNA elements in the human and mouse genomes. Nature. 2020;583:699–710. doi: 10.1038/s41586-020-2493-4 [PubMed: 32728249]
- Giraldez T, Rojas P, Jou J, Flores C, Alvarez de la Rosa D. The epithelial sodium channel delta-subunit: new notes for an old song. Am J Physiol Renal Physiol. 2012;303:F328–338. doi: 10.1152/ajprenal.00116.2012 [PubMed: 22573384]
- Kamynina E, Staub O. Concerted action of ENaC, Nedd4–2, and Sgk1 in transpithelial Na(+) transport. Am J Physiol Renal Physiol. 2002;283:F377–387. doi: 10.1152/ajprenal.00143.2002 [PubMed: 12167587]
- 14. Grunder S, Firsov D, Chang SS, Jaeger NF, Gautschi I, Schild L, Lifton RP, Rossier BC. A mutation causing pseudohypoaldosteronism type 1 identifies a conserved glycine that is involved in the gating of the epithelial sodium channel. EMBO J. 1997;16:899–907. doi: 10.1093/emboj/ 16.5.899 [PubMed: 9118951]

- Hanukoglu I, Hanukoglu A. Epithelial sodium channel (ENaC) family: Phylogeny, structurefunction, tissue distribution, and associated inherited diseases. Gene. 2016;579:95–132. doi: 10.1016/j.gene.2015.12.061 [PubMed: 26772908]
- 16. Samaha FF, Rubenstein RC, Yan W, Ramkumar M, Levy DI, Ahn YJ, Sheng S, Kleyman TR. Functional polymorphism in the carboxyl terminus of the alpha-subunit of the human epithelial sodium channel. J Biol Chem. 2004;279:23900–23907. doi: 10.1074/jbc.M401941200 [PubMed: 15069064]
- Tong Q, Menon AG, Stockand JD. Functional polymorphisms in the alpha-subunit of the human epithelial Na+ channel increase activity. Am J Physiol Renal Physiol. 2006;290:F821–827. doi: 10.1152/ajprenal.00312.2005 [PubMed: 16249274]
- Sofer T, Zheng X, Gogarten SM, Laurie CA, Grinde K, Shaffer JR, Shungin D, O'Connell JR, Durazo-Arvizo RA, Raffield L, et al. A fully adjusted two-stage procedure for rank-normalization in genetic association studies. Genet Epidemiol. 2019;43:263–275. doi: 10.1002/gepi.22188 [PubMed: 30653739]
- Levey AS, Stevens LA, Schmid CH, Zhang YL, Castro AF 3rd, Feldman HI, Kusek JW, Eggers P, Van Lente F, Greene T, et al. A new equation to estimate glomerular filtration rate. Ann Intern Med. 2009;150:604–612. doi: 10.7326/0003-4819-150-9-200905050-00006 [PubMed: 19414839]
- Zheng X, Gogarten SM, Lawrence M, Stilp A, Conomos MP, Weir BS, Laurie C, Levine D. SeqArray-a storage-efficient high-performance data format for WGS variant calls. Bioinformatics. 2017;33:2251–2257. doi: 10.1093/bioinformatics/btx145 [PubMed: 28334390]
- Gogarten SM, Sofer T, Chen H, Yu C, Brody JA, Thornton TA, Rice KM, Conomos MP. Genetic association testing using the GENESIS R/Bioconductor package. Bioinformatics. 2019;35:5346– 5348. doi: 10.1093/bioinformatics/btz567 [PubMed: 31329242]
- Wu MC, Lee S, Cai T, Li Y, Boehnke M, Lin X. Rare-variant association testing for sequencing data with the sequence kernel association test. Am J Hum Genet. 2011;89:82–93. doi: 10.1016/ j.ajhg.2011.05.029 [PubMed: 21737059]
- 23. The NHLBI Trans-Omics for Precision Medicine (TOPMed) Whole Genome Sequencing Program. BRAVO variant browser: University of Michigan and NHLBI. 2018.
- 24. Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, Maller J, Sklar P, de Bakker PI, Daly MJ, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. Am J Hum Genet. 2007;81:559–575. doi: 10.1086/519795 [PubMed: 17701901]
- 25. Lee S ML, Wu M. SKAT: SNP-Set (Sequence) Kernel Association Test; 2017.
- McLaren W, Gil L, Hunt SE, Riat HS, Ritchie GR, Thormann A, Flicek P, Cunningham F. The Ensembl Variant Effect Predictor. Genome Biol. 2016;17:122. doi: 10.1186/s13059-016-0974-4 [PubMed: 27268795]
- Guo Y, Zhao S, Ye F, Sheng Q, Shyr Y. MultiRankSeq: multiperspective approach for RNAseq differential expression analysis and quality control. Biomed Res Int. 2014;2014:248090. doi: 10.1155/2014/248090 [PubMed: 24977143]
- Ambrosius WT, Bloem LJ, Zhou L, Rebhun JF, Snyder PM, Wagner MA, Guo C, Pratt JH. Genetic variants in the epithelial sodium channel in relation to aldosterone and potassium excretion and risk for hypertension. Hypertension. 1999;34:631–637. doi: 10.1161/01.hyp.34.4.631 [PubMed: 10523338]
- Wang XF, Lu XM, Lin RY, Wang SZ, Zhang LP, Qian J, Lu DR, Wen H, Jin L. Lack of association of functional variants in alpha-ENaC gene and essential hypertension in two ethnic groups in China. Kidney Blood Press Res. 2008;31:268–273. doi: 10.1159/000151286 [PubMed: 18703878]
- 30. Yang W, Zhu Z, Wang J, Ye W, Ding Y. Evaluation of the relationship between T663A polymorphism in the alpha-epithelial sodium channel gene and essential hypertension. Saudi Med J. 2015;36:1039–1045. doi: 10.15537/smj.2015.9.11822 [PubMed: 26318459]
- Rosario RF, Wesson DE. Primary hypertension and nephropathy. Curr Opin Nephrol Hypertens. 2006;15:130–134. doi: 10.1097/01.mnh.0000214771.88737.ee [PubMed: 16481878]
- Johnson RJ, Herrera-Acosta J, Schreiner GF, Rodriguez-Iturbe B. Subtle acquired renal injury as a mechanism of salt-sensitive hypertension. N Engl J Med. 2002;346:913–923. doi: 10.1056/ NEJMra011078 [PubMed: 11907292]

- Waldmann R, Champigny G, Bassilana F, Voilley N, Lazdunski M. Molecular cloning and functional expression of a novel amiloride-sensitive Na+ channel. J Biol Chem. 1995;270:27411– 27414. doi: 10.1074/jbc.270.46.27411 [PubMed: 7499195]
- Pavlov TS, Staruschenko A. Involvement of ENaC in the development of saltsensitive hypertension. Am J Physiol Renal Physiol. 2017;313:F135–F140. doi: 10.1152/ ajprenal.00427.2016 [PubMed: 28003189]
- 35. Landrum MJ, Lee JM, Benson M, Brown GR, Chao C, Chitipiralla S, Gu B, Hart J, Hoffman D, Jang W, et al. ClinVar: improving access to variant interpretations and supporting evidence. Nucleic Acids Res. 2018;46:D1062–D1067. doi: 10.1093/nar/gkx1153 [PubMed: 29165669]
- Hoorn EJ, Gritter M, Cuevas CA, Fenton RA. Regulation of the Renal NaCl Cotransporter and Its Role in Potassium Homeostasis. Physiol Rev. 2020;100:321–356. doi: 10.1152/ physrev.00044.2018 [PubMed: 31793845]
- 37. Ray EC, Chen J, Kelly TN, He J, Hamm LL, Gu D, Shimmin LC, Hixson JE, Rao DC, Sheng S, et al. Human epithelial Na+ channel missense variants identified in the GenSalt study alter channel activity. Am J Physiol Renal Physiol. 2016;311:F908–F914. doi: 10.1152/ajprenal.00426.2016 [PubMed: 27582106]
- Winarski KL, Sheng N, Chen J, Kleyman TR, Sheng S. Extracellular allosteric regulatory subdomain within the gamma subunit of the epithelial Na+ channel. J Biol Chem. 2010;285:26088–26096. doi: 10.1074/jbc.M110.149963 [PubMed: 20587418]
- Downs CA, Johnson NM, Coca C, Helms MN. Angiotensin II regulates delta-ENaC in human umbilical vein endothelial cells. Microvasc Res. 2018;116:26–33. doi: 10.1016/j.mvr.2017.10.001 [PubMed: 29051045]
- 40. Paudel P, van Hout I, Bunton RW, Parry DJ, Coffey S, McDonald FJ, Fronius M. Epithelial Sodium Channel delta Subunit Is Expressed in Human Arteries and Has Potential Association With Hypertension. Hypertension. 2022:101161HYPERTENSIONAHA12218924. doi: 10.1161/ HYPERTENSIONAHA.122.18924
- 41. Wilck N, Matus MG, Kearney SM, Olesen SW, Forslund K, Bartolomaeus H, Haase S, Mahler A, Balogh A, Marko L, et al. Salt-responsive gut commensal modulates TH17 axis and disease. Nature. 2017;551:585–589. doi: 10.1038/nature24628 [PubMed: 29143823]
- 42. Gu X, Gu D, He J, Rao DC, Hixson JE, Chen J, Li J, Huang J, Wu X, Rice TK, et al. Resequencing Epithelial Sodium Channel Genes Identifies Rare Variants Associated With Blood Pressure Salt-Sensitivity: The GenSalt Study. Am J Hypertens. 2018;31:205–211. doi: 10.1093/ajh/hpx169 [PubMed: 29036630]
- Jones ES, Owen EP, Rayner BL. The association of the R563Q genotype of the ENaC with phenotypic variation in Southern Africa. Am J Hypertens. 2012;25:1286–1291. doi: 10.1038/ ajh.2012.125 [PubMed: 22895453]
- 44. Nkeh B, Samani NJ, Badenhorst D, Libhaber E, Sareli P, Norton GR, Woodiwiss AJ. T594M variant of the epithelial sodium channel beta-subunit gene and hypertension in individuals of African ancestry in South Africa. Am J Hypertens. 2003;16:847–852. doi: 10.1016/ s0895-7061(03)01016-1 [PubMed: 14553964]
- 45. Hannila-Handelberg T, Kontula K, Tikkanen I, Tikkanen T, Fyhrquist F, Helin K, Fodstad H, Piippo K, Miettinen HE, Virtamo J, et al. Common variants of the beta and gamma subunits of the epithelial sodium channel and their relation to plasma renin and aldosterone levels in essential hypertension. BMC Med Genet. 2005;6:4. doi: 10.1186/1471-2350-6-4 [PubMed: 15661075]
- 46. Salih M, Gautschi I, van Bemmelen MX, Di Benedetto M, Brooks AS, Lugtenberg D, Schild L, Hoorn EJ. A Missense Mutation in the Extracellular Domain of alphaENaC Causes Liddle Syndrome. J Am Soc Nephrol. 2017;28:3291–3299. doi: 10.1681/ASN.2016111163 [PubMed: 28710092]
- Wang X, Chen J, Shi S, Sheng S, Kleyman TR. Analyses of epithelial Na(+) channel variants reveal that an extracellular beta-ball domain critically regulates ENaC gating. J Biol Chem. 2019;294:16765–16775. doi: 10.1074/jbc.RA119.010001 [PubMed: 31551351]
- Wang H, D'Ambrosio MA, Garvin JL, Ren Y, Carretero OA. Connecting tubule glomerular feedback in hypertension. Hypertension. 2013;62:738–745. doi: 10.1161/ HYPERTENSIONAHA.113.01846 [PubMed: 23959547]

- Drummond HA, Stec DE. betaENaC acts as a mechanosensor in renal vascular smooth muscle cells that contributes to renal myogenic blood flow regulation, protection from renal injury and hypertension. J Nephrol Res. 2015;1:1–9. doi: 10.17554/j.issn.2410-0579.2015.01.12 [PubMed: 27928552]
- Heymann F, Meyer-Schwesinger C, Hamilton-Williams EE, Hammerich L, Panzer U, Kaden S, Quaggin SE, Floege J, Grone HJ, Kurts C. Kidney dendritic cell activation is required for progression of renal disease in a mouse model of glomerular injury. J Clin Invest. 2009;119:1286– 1297. doi: 10.1172/JCI38399 [PubMed: 19381017]

Novelty and Relevance

What is New?

We found associations between low-frequency and rare variation in *SCNN1A*, *SCNN1B*, and *SCNN1D* and blood pressure, and associations between *SCNN1B* and *SCNN1D* variation and eGFR.

What is Relevant?

Variation in *SCNN1D*, encoding the ENaC δ subunit that is poorly expressed in human kidney, is associated with blood pressure and eGFR.

Clinical/Pathophysiological Implications?

Humans with hypertension and specific non-Liddle ENaC variants, particularly variants with a gain-of function phenotype, may benefit from a trial of ENaC inhibitors, such as amiloride, to lower blood pressure.





Figure 1. Expression of ENaC subunits in human monocytes.

Human monocytes were exposed to a normal (150 mM) or high (190 mM) NaCl solution for 72 h, and RNA was isolated and sequenced. Normalized fragments per kilobase of transcript per million mapped reads (FPKM) are shown. Similar levels of expression of & ENaC were seen between the two exposures (p = 0.06, unpaired Student's *t* test).

Table 1. Studies included in the blood pressure analyses.

The Genetics of Cardiometabolic Health in Amish and Women's Health Initiative (WHI) studies are subdivided according to the abbreviated study name provided in the harmonized dataset.

Study	n	Abbreviation
Consticts of Cardiometebolic Health in the Amish		Amish_FC13
Genetics of Cardiometabolic Health in the Amish	269	Amish_FC2
Atherosclerosis Risk in Communities Study VTE cohort	3,039	ARIC
Cleveland Family Study	744	CFS
Framingham Heart Study	3,270	FHS
Genetic Studies of Atherosclerosis Risk	1,626	GeneSTAR
Genetic Epidemiology Network of Arteriopathy	1,138	GENOA
Genetic Epidemiology Network of Salt Sensitivity	1,616	GenSalt
Hypertension Genetic Epidemiology Network	1,635	HyperGen
Jackson Heart Study	3,124	JHS
Samoan	1,167	Samoan
	4,692	WHI_ctr
Women's Health Initiative	4,403	WHI_stroke_case
	944	WHI_VTE_case
Total	28,399	

Table 2.

Single-variant association tests of common ENaC variants with SBP, DBP, and PP.

Results of single-variant analyses of two common functional ENaC α subunit variants and one common ENaC β subunit variant for associations with SBP, DBP, and PP.

Variant	rs ID	MAF	p value			
			SBP	DBP	MAP	PP
aA334T	rs11542844	0.18	0.9	0.9	0.9	0.7
aT663A	rs2228576	0.75	0.1	0.6	0.9	0.06
βG165R	rs2303157	0.24	0.3	0.9	0.6	0.07

Table 3.

Sequence kernel association tests of low frequency and rare ENaC variants with blood pressure measures.

SKAT was performed on variants with a MAF < 0.05 in the genomic regions of *SCNN1A*, *SCNN1B*, *SCNN1D*, and *SCNN1G* with SBP, DBP, PP, and MAP, separately, and then combined (*SCNN1A*, *SCNN1B* and *SCNN1G*). SKAT was separately performed on rare (MAF < 0.01) and low frequency (0.01 MAF < 0.05) variants. Significant *p* values are in bold.

Gene. Chromosome:Start-End		Variants with MAF < 0.05		Variants wit	h MAF < 0.01	Variants with 0.01 MAF < 0.05	
	Measure	п	p value	п	p value	п	p value
SCNN1A.	SBP	3,972	0.02	3,859	0.03	113	0.07
12:0340824-0379751	DBP	3,972	0.00004	3,859	0.0004	113	0.0007
	PP	3,972	0.7	3,859	0.3	113	0.9
	MAP	3,972	0.0002	3,859	0.002	113	0.002
SCNN1B.	SBP	12,334	0.02	11,925	0.04	409	0.04
16:2329812-23381320	DBP	12,334	0.002	11,925	0.003	409	0.01
	PP	12,334	0.3	11,925	0.3	409	0.04
	MAP	12,334	0.003	11,925	0.006	409	0.01
SCNN1D.	SBP	2,405	0.0008	2,293	0.2	112	0.0005
1:1280417-1292050	DBP	2,405	0.002	2,293	0.03	112	0.003
	PP	2,405	0.003	2,293	0.04	112	0.005
	MAP	2,405	0.0008	2,293	0.1	112	0.0007
SCNN1G.	SBP	4,093	0.5	4,022	0.3	71	0.6
10.23184090-23210904	DBP	4,093	0.2	4,022	0.1	71	0.4
	PP	4,093	0.06	4,022	0.1	71	0.09
	MAP	4,093	0.5	4,022	0.2	71	0.8
Combined	SBP	20,399	0.01	19,806	0.02	593	0.03
(SCNNIA, SCNNIB, SCNNIG)	DBP	20,399	0.0002	19,806	0.0004	593	0.004
	PP	20,399	0.3	19,806	0.1	593	0.4
	MAP	20,399	0.0006	19,806	0.001	593	0.005
Combined	SBP			22,099	0.01	705	0.006
(SCNN1A, SCNN1B, SCNN1D, SCNN1G)	DBP			22,099	0.0002	705	0.001
	PP			22,099	0.08	705	0.14
	MAP			22,099	0.0009	705	0.001

p values < 0.00625 are significant

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 Table 4.

 Sequence kernel association tests of low frequency and rare ENaC variants with eGFR.

SKAT was performed on variants with a MAF < 0.05 in the genomic regions of *SCNN1A*, *SCNN1B*, *SCNN1D*, and *SCNN1G* with eGFR, separately, and then combined. SKAT was separately performed on rare (MAF < 0.01) and low frequency (0.01 MAF < 0.05) variants. Significant p values are in bold.

Gene. Chromosome:Start–End	Variants with $MAF < 0.05$		Variants with MAF < 0.01		Variants with 0.01 MAF < 0.05	
	п	p value	п	p value	п	p value
SCNNIA. 12:6346824-6379751	2,739	0.01	2,626	0.008	113	0.07
SCNN1B. 16:2329812-23381320	8,770	0.005	8,365	0.007	405	0.03
SCNN1D. 1:1280417-1292050	1,758	0.002	1,648	0.2	110	0.001
SCNN1G. 16:23184696-23216904	2,868	0.1	2,797	0.01	71	0.1
Combined (SCNN1A, SCNN1B, SCNN1D, SCNN1G)	16,135	0.0003	15,436	0.002	699	0.003

p values < 0.00625 are significant

Table 5. Sequence kernel association tests of low frequency and rare ENaC variants with stroke.

SKAT was performed on variants with a MAF < 0.05 in the genomic regions of *SCNN1A*, *SCNN1B*, *SCNN1D*, and *SCNN1G* with stroke, separately, and then combined. SKAT was separately performed on rare (MAF < 0.01) and low frequency (0.01 MAF < 0.05) variants.

Gene. Chromosome:Start-End	Variants with MAF < 0.05		Variants with MAF < 0.01		Variants with 0.01 MAF < 0.05	
	п	p value	п	p value	п	p value
SCNNIA. 12:6346824-6379751	2,330	0.08	2,216	0.05	114	0.2
SCNN1B. 16:2329812-23381320	6,965	0.5	6,556	0.7	409	0.4
SCNN1D. 1:1280417-1292050	1,621	0.5	1,497	0.6	124	0.4
SCNN1G. 16:23184696-23216904	2,208	0.2	2,137	0.1	71	0.3
Combined (SCNN1A, SCNN1B, SCNN1D, SCNN1G)	13,124	0.3	12,406	0.3	718	0.3