

Nanozymes in the Treatment of Diseases Caused by Excessive Reactive Oxygen Species

Shufeng Liang^{1,2}, Xin Tian³, Chunyan Wang⁴ 

¹Department of Molecular Biology, Shanxi Province Cancer Hospital/Shanxi Hospital, Affiliated to Cancer Hospital, Chinese Academy of Medical Sciences/Cancer Hospital Affiliated to Shanxi Medical University, Taiyuan, People's Republic of China; ²Institute of Environmental Sciences, Shanxi University, Taiyuan, People's Republic of China; ³State Key Laboratory of Radiation Medicine and Protection, School of Radiation Medicine and Protection, Medical College of Soochow University, Collaborative Innovation Center of Radiation Medicine of Jiangsu Higher Education Institutions, Soochow University, Suzhou, People's Republic of China; ⁴Department of Transfusion, Shanxi Province Cancer Hospital/Shanxi Hospital Affiliated to Cancer Hospital, Chinese Academy of Medical Sciences/Cancer Hospital Affiliated to Shanxi Medical University, Taiyuan, People's Republic of China

Correspondence: Xin Tian; Chunyan Wang, Department of Transfusion, Shanxi Province Cancer Hospital/Shanxi Hospital Affiliated to Cancer Hospital, Chinese Academy of Medical Sciences/Cancer Hospital Affiliated to Shanxi Medical University, No 3 Zhigongxin Street, Xinghualing, Taiyuan, Shanxi, People's Republic of China, Tel +86 15110313926, Email xtian@suda.edu.cn; chunyanwang2007@163.com

Abstract: Excessive reactive oxygen species (ROS) may generate deleterious effects on biomolecules, such as DNA damage, protein oxidation and lipid peroxidation, causing cell and tissue damage and eventually leading to the pathogenesis of diseases, such as neurodegenerative diseases, ischemia/reperfusion (I/R) injury, and inflammatory diseases. Therefore, the modulation of ROS can be an efficient means to relieve the aforementioned diseases. Several studies have verified that antioxidants such as Mitoquinone (a mitochondrial-targeted coenzyme Q10 derivative) can scavenge ROS and attenuate related diseases. Nanozymes, defined as nanomaterials with intrinsic enzyme-like properties that also possess antioxidant properties, are hence expected to be promising alternatives for the treatment of ROS-related diseases. This review introduces the types of nanozymes with inherent antioxidant activities, elaborates on various strategies (eg, controlling the size or shape of nanozymes, regulating the composition of nanozymes and environmental factors) for modulating their catalytic activities, and summarizes their performances in treating ROS-induced diseases.

Keywords: nanozymes, enzyme-mimic, oxidative stress, antioxidant, ROS-related diseases

Introduction

Reactive oxygen species (ROS) contain various reactive molecules and free radicals (species with unpaired electrons) derived from molecular oxygen, such as hydrogen peroxide (H₂O₂), superoxide (O₂⁻) and hydroxyl radicals (·OH).¹⁻³ ROS are produced by a cell converting oxygen into oxygen radicals during physiological processes in biological systems and play roles as signaling molecules in cells, including activating cell signaling cascades, inducing apoptosis, and influencing gene expression.⁴⁻⁷

However, oxidative stress occurs when ROS generation increases or antioxidant defense systems become weaker or both, which means unquenched ROS may trigger further reactions.^{5,8,9} In turn, oxidative stress destroys biomacromolecules and plays a part in a series of pathologies, including neurodegeneration, inflammation, atherosclerosis, diabetes, and aging.^{4,10-12} As massive ROS production and associated oxidative damage are also identified as important causative factors for diseases such as I/R, researchers speculated that scavenging excessive ROS might be a viable method for oxidative stress-related diseases in the future.^{13,14}

Nanozymes are inorganic nanomaterials that mimic enzyme activities.¹⁵ Since ferromagnetic nanoparticles (NPs) were detected to possess enzyme-like activity for the first time,¹⁶ various nanomaterials have been developed with peroxidase-like, oxidase-like, catalase-like and superoxide dismutase (SOD)-like activities and are widely used in biomedicine fields such as biosensing, disease diagnosis and treatment.^{17,18} Furthermore, as Gao and Yan et al concluded, nanozymes should be consistent with the enzyme kinetics and catalytic mechanism of natural enzymes, such as the

Michaelis–Menten equation. As a substitute for natural enzymes, even compared to other artificial enzymes, nanozymes are more prominent in their stability to pH and temperature, low production cost, ease of synthesis in large quantities, and reusability.^{19,20} In addition, the unique physicochemical properties of nanomaterials endow them with a variety of functions and provide various methods for rational design.²¹ By virtue of enzyme-like activities, especially SOD-like and catalase-like activities, nanozymes are expected to eliminate intracellular ROS and have therapeutic effects in ROS-related damages or diseases.

In this review, we focus on the catalytic activity of antioxidant nanozymes and their significant research progress in the treatment of excessive ROS-caused diseases. First, nanozymes that show antioxidant activity, including carbon-based, metal-based, and metal oxide-based nanozymes, are summarized. Second, strategies for modulating nanozymes to elevate their ROS-quenching capacities are outlined. Third, the applications of nanozymes in relevant treatments are introduced. We hope that this review will provide an overview of antioxidant nanozymes in the ROS-induced disease treatment field, as well as inspiration for future clinical applications.

Antioxidant Nanozymes

Here, we introduce the representative types of antioxidant nanozymes, which have been reported as antioxidants in recent years in the following section (Table 1).

Carbon-Based Nanozymes

Carbon-based nanomaterials have outstanding physicochemical properties, such as electrical, thermal, and optical properties, which make them promising materials for biomedical applications.²² They contain fullerene, graphene nanosheets, graphene oxide (GO), graphene quantum dots (GQDs), and carbon nanotubes (CNTs), which can be divided into zero-dimensional nanomaterials (fullerene, carbon dots, GQDs), one-dimensional nanomaterials (CNTs), two-dimensional nanomaterials (graphene), and three-dimensional nanomaterials (nanodiamonds) based on the number of dimensions exceeding the nanoscale (100 nm).²³

Fullerenes are carbon spheres, in which C-atoms are sp^2 -hybridized carbon atoms, and presently are used for various applications in nanomedicine.^{24,25} The existence of abundant conjugated double bonds and low lying lowest unoccupied molecular orbital endow them the ability to scavenge radical species.²⁶ With the abovementioned characteristics, fullerenes show antioxidant activity in dealing with $O_2^{\cdot-}$, $\cdot OH$, and H_2O_2 , and are called “free radical sponges”.^{27–29}

Through inhibiting cellular ROS levels, fullerenes could suppress proinflammatory cytokine release in synovial inflammation-related cells in vitro, and alleviate synovitis and joint destruction significantly in vivo.³⁰ Fullerenol belongs to fullerene derivatives and exhibits excellent water solubility. With the intrinsic capability of eliminating ROS, fullerenol/alginate hydrogels were synthesized to protect brown adipose-derived stem cells (BADSCs) against the oxidative stress damage after myocardial infarction (MI) and improve the cardiomyogenic differentiation.³¹ In MI rat model, the fullerenol/alginate hydrogel seeded with BADSCs reduced infarct size, increased wall thickness, and improved cardiac functions successfully.

CNTs, with hollow and cylindrical structure, are formed by rolled graphene sheets. Single-walled CNT consist of one graphene sheet, while multiwalled carbon nanotubes (MWCNTs) consist of multiple single graphene layers.³² For the first time, Fubini et al reported MWCNTs exhibited a remarkable ROS scavenging capacity, including $\cdot OH$ and $O_2^{\cdot-}$.³³ The excellent electron affinity of the molecular orbitals or electronic bands of the carbon atoms in the nanotube framework was conducive to the scavenging reaction.

Graphene is a flat-like layer of densely packed sp^2 carbon atoms. For graphene-based nanomaterials, Akhavan et al had summarized that the electron density, chemical composition, sp^2 -hybridized carbon content and chemical properties were responsible for their ROS-scavenging abilities.¹³ Just as Hurt et al reported that a series of graphene-based materials showed $\cdot OH$ scavenging activities.³⁴ Among them, few layer graphene exhibited the best performance, followed by reduced graphene oxide (rGO) and GO. The activity order was in inverse to their total surface area. Thus, the main antioxidant activities of graphene-based materials were ascribed to pristine sp^2 carbon domains rather than oxygen-containing functional groups.

Table 1 Representative Antioxidant Nanozymes and Their Applications

Nanozymes	Activity/Mechanism	Application	Reference
Fullerenes	·OH and O ₂ ^{·-} scavenging activity	Cardiac functional recovery	[31]
		Allergic response	[165]
		Doxorubicin-induced hepatotoxicity	[166]
Graphene based NPs	·OH and O ₂ ^{·-} scavenging activity Catalase	Neuroprotection	[34]
		Cardiac repair	[36]
		Acute kidney injury	[134]
Fe ₃ O ₄ NPs	Peroxidase Catalase	Aging	[39]
		Neurodegeneration	[39]
		Cerebral malaria	[167]
PBNPs	Peroxidase Catalase SOD	Colitis	[43]
		Acute pancreatitis	[47]
		Full-thickness skin wound	[48]
		Ischemic Stroke	[49]
CeO ₂ NPs	Catalase SOD Oxidase	Ischemic stroke	[62,63]
		AD	[64–66]
		Depression	[67]
		IBD	[69]
		Acute gout	[70]
		Drug-induced liver injury	[73]
		PD	[75]
		Rheumatoid arthritis Traumatic brain injury	[168] [169]
Mn ₃ O ₄ NPs	SOD Catalase GPx	Ear inflammation	[82]
		PD	[84]
		IBD	[170]
Cu based NPs	Peroxidase SOD Catalase GPx	Acute kidney injury, acute liver injury, wound healing	[20]
		PD	[79]
Mo based NPs	SOD Catalases Peroxidases	Acute kidney injury	[162]
		Hepatic fibrosis	[171]
Pt NPs	SOD Catalases Peroxidases	Pulmonary inflammation	[172]
		Hepatic ischemia/reperfusion injury	[173]

(Continued)

Table I (Continued).

Nanozymes	Activity/Mechanism	Application	Reference
Melanin-like NPs	·OH, O ₂ ⁻ , H ₂ O ₂ , NO, and ONOO ⁻ scavenging activity	Ischemic stroke	[153]
		Periodontal disease	[150]
		Osteoarthritis	[148]
		Acute peritonitis and acute lung injury	[174]
MOF based NPs	SOD Catalase	Endotoxemia	[97]
		IBD	[175]

In virtue of the antioxidant activities, graphene-based nanomaterials are used for medical treatment applications, such as neurotoxicity and cardiac repair.^{35,36}

GOQDS could decompose H₂O₂ to H₂O and O₂, thus verifying their catalase-like activity.³⁶ Pretreatment with GOQDS in 1-methyl-4-phenyl-pyridinium ion (MPP⁺) induced PC12 cells and the brains of larval zebrafish could prevent overproduction of ROS and provide neuroprotection.³⁵

GOs was modified with polyethylenimine and folic acid–polyethylene glycol (PEG) to form a macrophage-targeting /polarizing GO complex (MGC).³⁶ The as-prepared material was verified to eliminate ·OH which invoked cardiac failure after MI. In turn, inflammatory biomarkers, such as tumor necrosis factor- α (TNF- α), were less produced during the inflammatory phase. Additionally, DNA-functionalized MGC promoted the polarization of M1 to M2 macrophages and accelerated secretion of cardiac repair-favorable cytokines. Collectively, this compound played dual roles to attenuate inflammation and improve heart function.

Metal and Metal Oxide-Based Nanozymes

Iron Based

Under different reaction conditions, iron oxide nanozymes (including Fe₃O₄ and Fe₂O₃) exhibited either peroxidase-like or catalase-like activities.^{16,37,38} These activities conferred iron oxide nanozymes the abilities to scavenge ROS under appropriate conditions.

Intrigued by the discovery of the catalase-like activity of iron oxide NPs, researchers have explored how these nanoparticles dealt with oxidative stress in aging or neurodegeneration.³⁹ Fe₃O₄ NPs alleviated a cell model of Parkinson's disease (PD) death and reduced the levels of α -synuclein and cleaved caspase-3, which are presynaptic neuronal proteins tightly linked to PD pathogenesis and apoptosis markers, respectively. Finally, the effects of dietary Fe₃O₄ NPs on aging and the Alzheimer's disease (AD) model of *Drosophila* were tested, and Fe₃O₄ NPs were effective in increasing life span, antiaging, promoting locomotor activity and reducing apoptosis in these disease models.

Combining nanozymes and natural antioxidants has also been used as free radical scavengers, as shown in Figure 1A.⁴⁰ As Zhao et al designed, Fe₃O₄ NPs and the polyphenol tannic acid (TA), which is a natural antioxidant, were combined into Fe₃O₄@TAN nanoflowers (NFs), as shown in Figure 1B. The Fe₃O₄@TAN NFs could mimic peroxidase, catalase, and SOD. Through the complementary action of Fe₃O₄ and TA, the as-designed nanomaterial exhibited improved and broad-spectrum ROS eliminating activity. Additionally, the nanoflower morphology of the Fe₃O₄@TAN NFs made them fully exposed to oxidative stress conditions, thus contributing to the excellent activity. Owing to a remarkable multiple ROS scavenging activity, the Fe₃O₄@Tan NFs were employed to treat mouse endotoxemia with excellent results.

In addition, Prussian blue nanoparticles (PBNPs), as Fe-containing nanomaterials with good biosafety, were also found to possess multienzyme activities. The different redox potentials of PB, Prussian white (PW), Berlin green (BG), and Prussian yellow (PY) conferred these materials tremendous abilities to transfer electrons.^{41–44} Dong et al proposed that PBNPs represented the first example of a bioinspired nanozyme.⁴⁵ They looked for the active site of PBNPs and

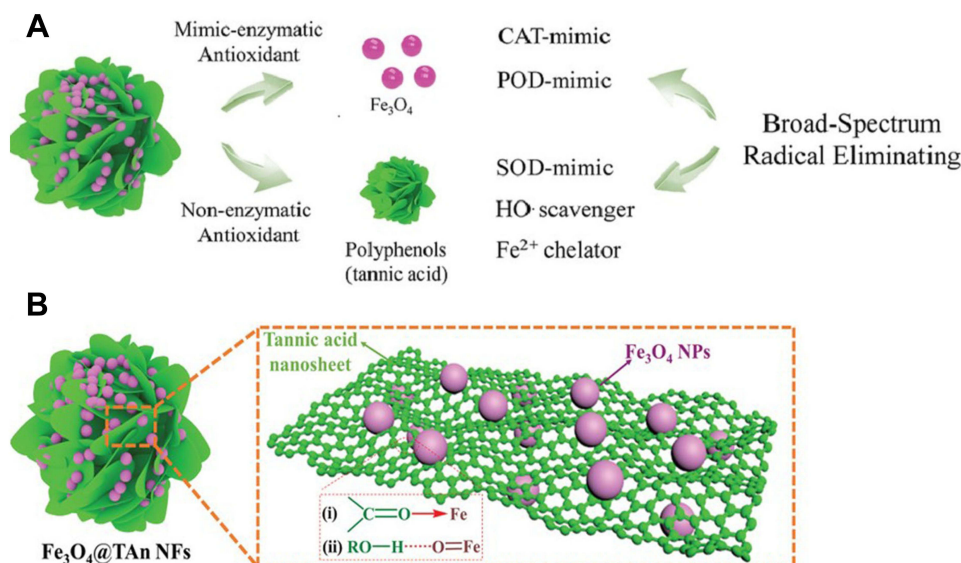


Figure 1 Iron oxide nanozyme and nonenzymatic antioxidant constructed a nanoplatform to mimic the antioxidant defense system. **(A)** The antioxidant ability of Fe_3O_4 @TAN NFs. **(B)** Schematic illustration of the petal structure of Fe_3O_4 @TA1.0 NFs. Reproduced with permission from Wei Z, Wang L, Tang C et al. Metal-phenolic networks nanoplatform to mimic antioxidant defense system for broad-spectrum radical eliminating and endotoxemia treatment. *Adv Funct Mater.* 2020;30(49). Copyright (2020) John Wiley and Sons.⁴⁰

concluded that N-coordinated Fe units (FeN_x ($x=4-6$)) with intrinsic heme-like structures endowed PBNPs with peroxidase-like activity.

Thus, PBNPs could efficiently control ROS-induced cell damage and treat diseases such as mouse liver inflammation and injury, colitis, acute pancreatitis, cutaneous wound healing and ischemic stroke.^{42,46-50}

Gold-Based

Au NPs were demonstrated to decompose H_2O_2 to O_2 at alkaline pH and scavenge superoxide at acidic pH.⁵¹ The activities made Au NPs potential catalase and SOD mimetics. In addition, Guo and Lu et al showed that Au NPs played important roles in ameliorating neurologic deficits and infarction volumes and protecting the cerebra from I/R injury in rats via the regulation of inflammatory and immune responses.⁵²

To eliminate the toxicity induced by $\cdot\text{OH}$ produced by the catalytic action of Au nanoclusters (AuNCs), polymeric 3^o-amine was added to Au NCs to simultaneously restrain the generation of $\cdot\text{OH}$ and retain their catalase-like activity. The derived AuNCs- NH_2 acted as H_2O_2 scavengers, thus protecting primary mouse neurons against oxidative stress.⁵³

Cerium Oxide (Ceria)-Based

As early as 2006, Schubert et al proposed that cerium oxide NPs could protect nerve cells from oxidative stress through the ability to limit the amount of ROS.⁵⁴

As relevant studies developed, cerium oxide NPs were proven to have excellent multiple enzyme-mimicking properties due to their $\text{Ce}^{3+}/\text{Ce}^{4+}$ oxidation state and the presence of oxygen vacancies.⁵⁵⁻⁵⁷

Numerous studies are performed on nanoceria and its biomedical application. CeO_2 NPs and related materials have been widely used to protect cells that are destroyed by excessive ROS, such as nerve cells, macrophages, cardiac progenitor cells, human breast cells, fibrosarcoma cells and human hepatic cells.^{54,57-61} Furthermore, they may serve as a novel therapy for neurodegenerative diseases, depression, ischemic stroke, autoimmune diseases, hepatic injury, sepsis, psoriasis, inflammatory bowel disease (IBD), periodontitis, and acute gout.⁶²⁻⁷⁷ Based on the excellent ROS eliminating capabilities of CeO_2 , researchers have explored various kinds of targeted methods to address the aforementioned diseases.

During the treatment of ROS-related diseases, CeO_2 also plays roles in downregulating proinflammatory cytokine levels, upregulating anti-inflammatory cytokine levels, inhibiting the ROS-NF κ B pathway, etc.^{69,73,77}

Copper Based

Cu-based nanozymes have been widely used to eliminate ROS and improve ROS-related diseases, including PD, acute liver injury, acute kidney injury, and wound healing.^{20,78,79}

Deng's group considered that large surface-to-volume ratios would confer high catalytic activity, so they made ultrasmall Cu_{5,4}O NPs (Cu_{5,4}O USNPs), the average hydrodynamic diameter of which was as small as 4.5 nm.²⁰ The as-designed nanoparticles were verified to have multiple enzyme-mimicking activities, including catalase, glutathione peroxidase (GPx), SOD and excellent broad-spectrum ROS scavenging ability. The reason for the name "Cu_{5,4}O USNPs" was because the materials were a mixture of Cu and Cu₂O NPs.

Manganese-Based

Lv et al first discovered that manganese dioxide (MnO₂) nanoparticles possessed enzyme-mimicking activities, which was further demonstrated in another study in 2017.^{80,81}

Recently, Mn₃O₄ nanozymes were also confirmed to possess antioxidant enzyme-like properties, and when compared with the most widely used CeO₂, researchers found that Mn₃O₄ nanozymes were more effective in terms of ROS elimination.^{82,83} The mixed valance states of Mn²⁺/Mn³⁺, tolerance to oxidation, large surface area and extremely large pore size of Mn₃O₄ nanozymes contributed to their outstanding multienzyme activity.⁸²

Mn₃O₄ nanozymes could provide efficient cytoprotection in SHSY-5Y, which is a human cell line model for the PD phenotype, and protect mice from phorbol 12-myristate 13-acetate (PMA)-induced ear inflammation.^{82,84} Furthermore, Mn₃O₄ nanozymes were proven to protect cells suffering from oxidative stress.⁸³ The related expression of various stress markers and antioxidant enzymes in the presence of Mn₃O₄ nanozymes was also detected. Conclusively, Mn₃O₄ nanozymes complemented the antioxidant machinery and rescued the cells under oxidative stress independent of the endogenous antioxidant machinery.

Other Metal-Based Nanozymes

Qu and his cooperators synthesized GO-Se nanocomposites and Se@PDA (polydopamine) nanozymes, which both showed GPx-like properties.^{85,86} Se@PDA nanocomposites showed excellent antioxidant activity, which was ascribed to the enzyme-like effect of the Se component and the nonenzymatic antioxidant ability of pDA.⁸⁶ By decomposing excessive intracellular H₂O₂, the nanocomposites successfully alleviated pneumonia in the mouse model. To achieve higher GPx-like activity, mesoporous selenium NPs (MSeNPs) with a high specific surface were synthesized.⁸⁷ Then, hyaluronic acid (HA), which could specifically attach to CD44 on the surface of macrophages, was assembled on the MSe NPs to facilitate the accumulation of the as-designed nanozyme at the site of inflammation. Taken together, treatment with MSe-HANPs successfully relieved local inflammation and sepsis.

Metal Organic Frameworks (MOFs) Based

MOFs are a class of supramolecular coordination complexes with metal ions/clusters bridged with organic linkers by coordination bonds.^{88,89} They also have natural enzyme-mimicking properties. For example, MOF-derived copper nanoparticle@carbon nanocomposites, iron(III)-based MOFs (MIL-68 and MIL-100) (MIL=Material Institute of Lavoisier), PA-Tb-Cu MOF, Cu²⁺ ion-modified UiO-type MOF NPs (UiO = University of Oslo), MOF-808 and zirconium-ferriporphyrin (Zr-FeP) MOF were verified to possess peroxidase-like activities.⁹⁰⁻⁹⁵

Tb³⁺ as a metal ion and a boronic acid ligand as a linker constituted a luminescent Tb-MOF.⁹⁶ The boronic acid groups of Tb-MOF reacted with ROS through the rapid nucleophilic addition reaction, which endowed the MOF with ROS quenching capability. Further investigation proved that Tb-MOF effectively scavenges exogenous or endogenous ROS in living cells, including HeLa cells and RAW 264.7 cells.

According to the size and active sites of natural SOD, Qu and Ren et al designed ultrasmall and bioinspire Cu-TCPP MOF nanodots (CTMDs), the average size of which was less than 5 nm.⁹⁷ The Cu active sites of CTMDs showed a similar coordination of N and O atoms to that of SOD, while the ordered channels were similar to substrate channels that could accumulate superoxide, thus contributing to the potent SOD-like activities of CTMDs. Additionally, CTMDs could catalyze H₂O₂ into H₂O with glutathione (GSH), which mimicked the activity of GPx. Combining SOD-like and

GPx-like activity, CTMDs could effectively destroy $O_2^{\cdot-}$ and H_2O_2 . As demonstrated in vivo, CTMDs could alleviate endotoxemia efficiently by eliminating ROS and reducing systemic inflammation.

Others

Melanin and Its Derivatives

Nanomaterials derived from natural organisms have been used in biological and medical applications due to their biocompatibility and biodegradability. As a natural biopolymer, melanin is distributed in the human body, such as skin, eye, brain, and hair.^{98,99} It showed scavenging activity toward ROS and protected skin from UV damage.¹⁰⁰ In addition, PDA NPs, as the most typical synthetic melanin, are produced by the oxidative polymerization of dopamine. The reductive functional groups of PDA NPs, including catechol and imine, endow them with robust potential in scavenging ROS.^{98,101,102}

Ultrasmall Mn^{2+} -chelated melanin NPs (MMPPs) were also verified to exhibit multiple ROS scavenging activities and used to treat murine acute kidney injury.¹⁰³ Considering the characteristics of targeted disease, MMPPs were designed with an ultrasmall hydrodynamic size, good physiological stability, and stable radiolabeling performance. Thus, they could preferentially concentrate in the kidney and show a robust antioxidative protection response with few side effects.

Single Atom Nanozymes (SAzymes)

SAzymes contain single metal atoms on various supports, which can maximize the utilization of metal atoms, thus achieving high activity and selectivity and shrinking the difference between nanozymes and natural enzymes.^{104–106}

To obtain an efficient antioxidant therapy for sepsis, Qu and Ren et al synthesized a SAzyme, Co/PMCS, characterized by atomically dispersed coordinatively unsaturated active Co porphyrin centers. The SAzymes could mimic SOD, catalase and GPx, thus obliterating multiple RONS.¹⁰⁷ In another experiment, the researchers tried to develop a clinically applicable bandage based on a single-atom Pt/CeO₂ nanozyme for brain trauma (Figure 2).¹⁰⁸ The nanozyme had high and persistent catalytic activity, which could fight against oxidative stress, decrease neuroinflammation and improve neurotrauma. The high catalytic activity was ascribed to the 2% lattice expansion induced by Pt single atoms dispersed on the CeO₂ (111) matrix, thus decreasing the binding energy of the chemical reactions. Moreover, Pt/CeO₂ avoided obvious decay due to continuous exposure to oxygen and maintained catalytic activity for up to 30 days, which was ascribed to its sustained electron-donating ability via the Mars–van Krevelen reaction.

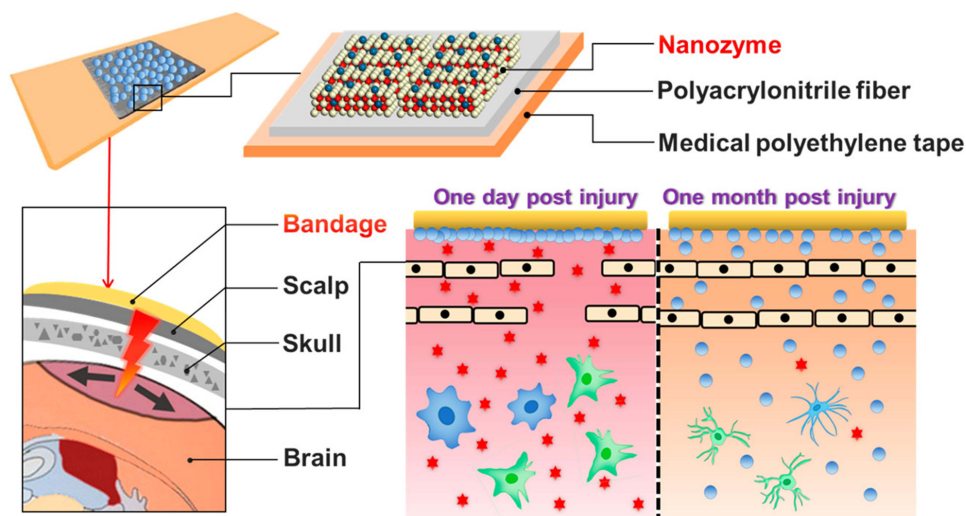


Figure 2 Schematic diagram of the nanozyme-based bandage to treat brain trauma. Reprinted with permission from Yan R, Sun S, Yang J et al. Nanozyme-based bandage with single-atom catalysis for brain trauma. *ACS nano*. 2019;13(10):11,552–11,560. Copyright (2019) American Chemical Society.¹⁰⁸

Regulatory Factors for Nanozyme Activity

Elevating the catalytic properties of nanozymes is a topic of common concern. Studies focused on activity regulation have been performed in the past few years. The following are several important factors that influence the antioxidant activity of nanozymes.

Physicochemical Properties

Size

Generally, smaller nanozymes show better catalytic activity than larger nanozymes. As the size decreases, nanozymes obtain a higher surface-to-volume ratio, expose more active sites, and have a larger surface energy.^{21,109} However, there are some special cases; for example, the catalytic performance of Pd-Ir NPs increased with increasing size.¹¹⁰ To exclude other physicochemical factors, the researchers synthesized nanomaterials with the same morphologies and surface structures except for size. Catalytic activity parameters, such as K_{cat} (catalytic rate constant) and K_{cat}/S (area-specific K_{cat}), were compared. As the size of the Pd-Ir NPs increased, K_{cat} continuously increased by nearly 12-fold. However, the K_{cat}/S values for different sizes of Pd-Ir NPs were similar. Combining the results, the conclusion was that the catalytic activity of the NPs was mainly ascribed to the catalytic surface areas, which could promote the interaction of individual nanoparticles and more substrates.

Shape and Morphology

The catalytic activity of nanomaterials also depends on shape and morphology.¹¹¹ For instance, Ge et al speculated that antioxidant activity was structure-dependent for the carbon nanomaterial family.²⁹ They evaluated the radical scavenging activities of buckyball-shaped fullerene derivatives and sheet-shaped GQDs by electron spin resonance (ESR) measurements. After comparison, the activities were in the order C70>C60>GQD. By theoretical calculations of the Gibbs free energies of reactions (Gr), the Gr 's of C60 and C70 were much lower than that of GQD, suggesting that larger radical scavenging activity depends on the lower Gr values.

More deeply, scientists revealed the prominent effect of surface facets in determining the enzyme-like activity of Pd nanocrystals.¹¹² They discovered that Pd octahedrons enclosed by 111 facets had greater enzyme-mimic activity than Pd nanocubes enclosed by 100 facets through ESR experiments (Figure 3A). Taking advantage of theoretical simulations, the ROS clearance that reacted on Pd 111 and 100 facets was calculated. The reaction energy (Er) of the rate-limiting step could be used as the descriptor of the enzyme activity of the metal. As shown in Figure 3B, the Er values of H_2O_2 and O_2^- on the 111 facet were both stronger than those on the 100 facet, which indicated greater H_2O_2 and O_2^- scavenging activities.

Interestingly, Kuang et al prepared a complex of CuO and Cu₂O nanoparticle clusters (Cu_xO NCs) by selecting L-phenylalanine (L-Phe) as the structure-directing agent.⁷⁹ The Phe ligand was confirmed to take part in determining the Cu_xO NC structure, which had a uniform diameter and morphology, a larger volume of pores and a larger pore size than those of other materials, such as Cu_xO-aspartic acid (Asp) (Figure 3C), thus endowing the Cu_xO NCs with higher catalytic performance.

Composition

Tailoring the composition of nanomaterials, such as doping other elements or forming bimetallic or multimetallic nanocomposites, poses another way to tune the activity of nanozymes.^{113–118}

Hyeon and Lee et al introduced Zr⁴⁺ into ceria nanoparticles to increase the Ce³⁺/Ce⁴⁺ ratio and convert Ce⁴⁺ to Ce³⁺ since Ce³⁺ targets O_2^- and $\cdot OH$.⁷² Through investigation, the Ce_{0.7}Zr_{0.3}O₂ (7CZ) NPs showed the best ROS-scavenging performance. Additionally, in vivo tests also demonstrated that 7CZ NPs effectively reduced mortality and systemic inflammation in sepsis. By Li doping, the catalytic activities of ZnMn₂O₄ nanozyme developed from single antioxidant activity to multiple activities, including SOD-like, catalase-like and GPx-like activities, with enhanced performance.¹¹⁹ Through XPS analysis, the molar ratio of Mn⁴⁺/Mn³⁺ increased from 0.56 to 2.29 along with gradual Li doping.

Another doping strategy was adopted to improve the catalytic activity of Prussian blue analog (PBA)-based nanozymes (Figure 3D and E).¹²⁰ In this study, Ni-Fe bimetal PBA-based nanocages doped by molybdenum-polysulfide (Nanocages)

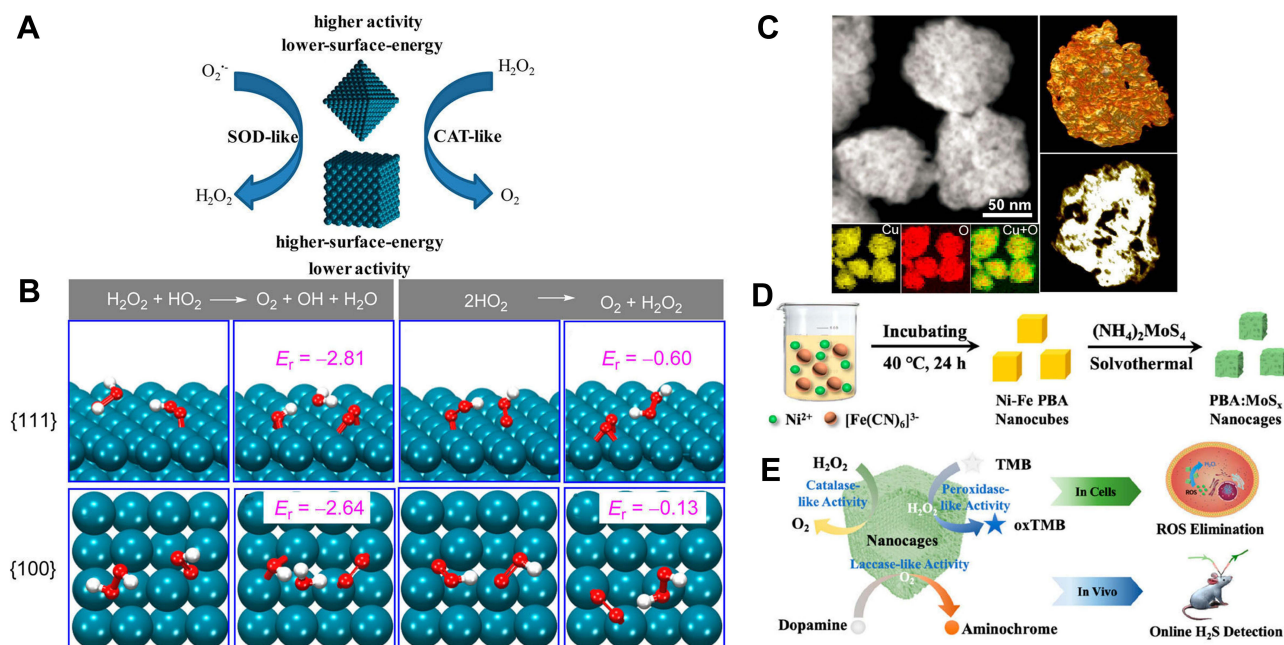


Figure 3 The regulation of nanozyme activity based on physicochemical properties. **(A)** Lower surface energy {111}-faceted Pd octahedrons and higher surface energy {100}-faceted Pd nanocubes. **(B)** Lowest-energy adsorption structures and reaction energies (in eV) for the reactions on structures having either Pd 111 or 100 facets. Reprinted with permission from Ge C, Fang G, Shen X et al. Facet energy versus enzyme-like activities: the unexpected protection of palladium nanocrystals against oxidative damage. *ACS Nano*. 2016;10(11):10,436–10,445. Copyright (2016) American Chemical Society.¹¹² **(C)** STEM, energy dispersive X-ray spectroscopy (EDS) mapping and electron tomographic reconstruction of Cu_xO NCs. Reprinted with permission from Hao C, Qu A, Xu L et al. Chiral molecule-mediated porous Cu_xO nanoparticle clusters with antioxidant activity for ameliorating Parkinson's disease. *J Am Chem Soc*. 2019;141(2):1091–1099. Copyright (2019) American Chemical Society.⁷⁹ Synthetic process **(D)** and catalytic performance **(E)** of the Nanocage. Reprinted with permission from Wang C, Ren G, Yuan B et al. Enhancing enzyme-like activities of Prussian blue analog nanocages by molybdenum doping: toward cytoprotecting and online optical hydrogen sulfide monitoring. *Anal Chem*. 2020;92(11):7822–7830. Copyright (2020) American Chemical Society.¹²⁰

were prepared and were proved to have multiple enzyme-like activities. Doping molybdenum into PBA successfully regulated the size (decreased), shape (from solid to hollow), composition, and electronic structure of the nanocage, thus bringing in more catalytic sites and binding sites.

In addition, the peroxidase-like, catalase-like and SOD-like activity of PtCu nanoalloys was related to the atomic ratio of Pt/Cu.¹²¹ At the same time, by changing the atomic ratio of Pt/Cu, the particle size, alloy composition and crystal structure of PtCu nanoalloys were controlled correspondingly.

Surface Coating and Modification

Controlling surface chemistry by surface coating and modification is also an advantageous way to improve catalytic performance because chemical reactions take place mainly on the surface of catalysts.¹²² Ions (fluoride), small molecules such as amino acids, citrate, Prussian blue, amino acids, and polymers (protein, dextran, DNA) can act as materials for modification.^{123–128}

Yan and Gao et al modified Fe₃O₄ NPs with histidine (His) residues to mimic the active site of horseradish peroxidase (HRP) and expected hydrogen bond formation between the distal imidazole group and H₂O₂.¹²⁹ The bionic design effectively altered the catalytic properties of the Fe₃O₄ nanozyme.

Specifically, abundant efforts have been made to improve the specificity of nanozymes for chiral substrates. By virtue of the surface coating of amino acids and DNA with chiral structures on nanozymes as chiral ligands, nanozymes can be designed with chiral selectivity.^{130–133}

Natural phenolic antioxidants gave Qu and Ren et al inspiration; they used NaBH₄ as the reductant to convert C=O on classical GQDs (c-GQDs) to C–OH and formed GQDs (h-GQDs) with phenol-like groups.¹³⁴ Through comparison, the antioxidant activity of h-GQDs was at least 5 times that of c-GQDs under the same conditions. By means of experimental research and theoretical calculations, the researchers discovered step by step that phenol-like groups played an important

role in the catalytic performance of h-GQDs. In addition, carbonyl groups, which suppress the H-donating ability of the phenol-like group, were removed from h-GQDs, which also promoted the antioxidative activity of h-GQDs.

Form Complex or Hybrid Nanozymes

The construction of different nanozymes may improve their catalytic performance and result in synergistic effects and even multistep cascade reaction activity, thus attracting many intentions in recent years.^{135–139}

A multinanozyme cooperative platform was assembled in which V_2O_5 nanowires acted as GP_x mimics, while MnO_2 nanoparticles mimicked SOD and catalase (Figure 4A).¹⁴⁰ $V_2O_5@pDA@MnO_2$ exhibited excellent ROS depletion performance that could mimic a natural antioxidant enzyme system (Figure 4B). Compared with Se NPs, GO-Se nanocomposites exhibited much higher GP_x-like activity.⁸⁵ The reason might be that GO endows the GO-Se hybrid with a greater surface area and rapid electron transfer capacity.

Zeolitic imidazolate framework-8 (ZIF-8) was used to encapsulate CeO_2 NPs to form $CeO_2@ZIF-8$ and overcame some limitations of CeO_2 .¹⁴¹ According to the design, ZIF-8 mimicked peroxidase to maintain antioxidant activity and controlled the size, morphology, and surface potential of CeO_2 , which was beneficial for biological applications.

Environmental Factor

The antioxidant performance of nanozymes is also sensitive to the surrounding environment, such as pH, temperature, and light. The catalytic activity of nanozymes is greatly influenced by pH. As Wei concluded, under acidic conditions, nanozymes showed peroxidase-mimicking activities, while under neutral and alkaline conditions, they exhibited SOD-like and catalase-like properties.²¹

Application of Nanozymes in Excessive ROS-Induced Diseases Treatment

In this section, we introduce the application of nanozymes in excessive ROS-induced diseases, including neurodegenerative diseases, inflammatory diseases, and stroke et al.

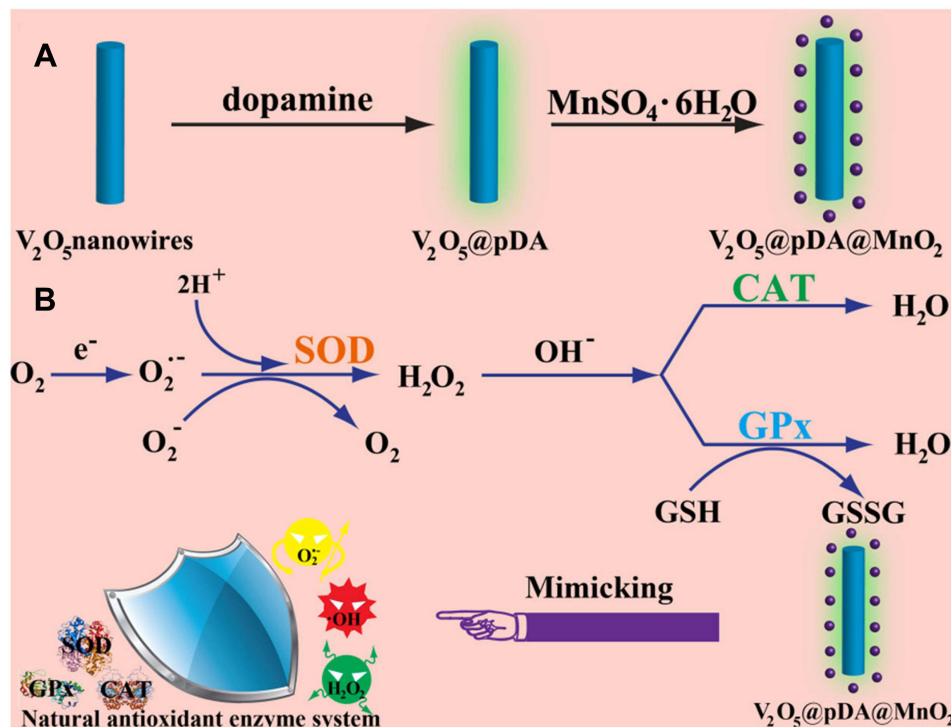


Figure 4 A multinanozyme cooperative platform to efficiently reduce ROS levels. (A) The synthesis process of $V_2O_5@pDA@MnO_2$ nanocomposites. (B) The multinanozyme system mimics the intracellular antioxidant enzyme-based defense system. Reprinted with permission from Huang Y, Ren J, Qu X. Nanozymes: classification, catalytic mechanisms, activity regulation, and applications. *Chem. Rev.* 2019, 119, 6: 4357–4412. Copyright (2019) American Chemical Society.¹⁷⁶

Treatment of Inflammatory Diseases

The accumulation of ROS may be one of the signaling pathways connecting the UPR (unfolded-protein response) and inflammation, thus initiating an inflammatory response.^{12,142} Excessive inflammation can cause chronic or systemic inflammatory diseases, including cardiovascular disease, atherosclerosis, asthma, cystic fibrosis, and rheumatoid arthritis.^{143,144}

IBD, as a troublesome colorectal inflammatory disease, is also closely related to excessive ROS.¹⁴⁵ Current treatment for IBD mainly relies on antibiotics, which will induce problems such as antibiotic resistance. Prussian blue, which has been approved as an antidote by the US Food and Drug Administration (FDA) and with good biosafety, can also act as an artificial nanozyme, effectively scavenging ROS.¹⁴⁶ To improve the solubility of PB NPs, PPB NPs were prepared with the help of poly(vinylpyrrolidone) via a one-pot strategy.⁴³ The *in vivo* experiments showed that through intravenous administration, PPBs targeted the inflamed sites and alleviated colitis in mice; in addition, proinflammatory cytokines were inhibited significantly. Furthermore, to realize oral administration, it was an ingenious design to combine clinically approved montmorillonite (MMT) and CeO₂ in targeting the inflamed colon.⁶⁹ MMT, with negative charges and high tolerance to pH, endowed CeO₂@MMT with stability to pH and specific adsorption onto the inflamed colon, which was positively charged based on electrostatic interactions. Thus, the nanozyme could be efficiently delivered through the digestive tract, which made it suitable for oral administration. MMT could simultaneously decrease the systemic absorption of CeO₂, thus reducing potential toxicity. Budesonide, a promising drug for IBD, and MnO₂ nanozyme, a nanocarrier, when taken together, became more effective and could be taken orally, as Tong et al demonstrated in mouse models.¹⁴⁷ The main reasons for selecting hollow MnO₂ NPs were as follows: first, they possessed intrinsic enzyme-like properties, which could scavenge ROS; second, they had an adjustable hollow structure, which was conducive to the loading of drugs; and thirdly, they could be modified with dextran sulfate sodium (DSS), which could target activated macrophages. In addition, hollow MnO₂ NPs were influenced by the inflammatory microenvironment and were prone to release budesonide, thus fulfilling their role as nanocarriers. Compared with free budesonide, the nanocarrier system exhibited fewer side effects and had a better curative effect on IBD.

The researchers noticed some problems in the therapy of osteoarthritis, such as antioxidants, melatonin and N-acetylcysteine (NAC), were too small to stay long enough in the joint.¹⁴⁸ Therefore, they designed appropriately sized dopamine-melanin (DM) NPs, which were 112.5 nm in size, and made the nanomaterials suitable for retention in the joint for a long time. In a rat osteoarthritis model, DM NPs were found to combat damage caused by inflammation and protect chondro by scavenging intracellular RONS and activating antioxidant enzymes by autophagy. Sharma et al chose cartilage that suffered irreversible damage under oxidative stress as the therapeutic target.¹⁴⁹ PEG-MnO₂ NPs were designed with a size of less than 20 nm and positive surface charge. Due to the small size and zeta potential, the nanoparticles enabled cartilage penetration and chondrocyte uptake *in vitro* and *in vivo*. After one week, 75% of the initial signal remained in the joint space, which might be connected with the electrostatic interactions between nanoparticles and anionic cartilage tissue. Combined with the intrinsic ROS scavenging capability of MnO₂ NPs, the materials improved chondrocyte viability and extracellular matrix preservation by reducing inflammation-induced oxidative stress in cartilage.

Yang et al pointed out that in periodontal disease suffering from oxidative stress, PDA NPs with strong ROS removal capacity successfully decreased local periodontal inflammation.¹⁵⁰ PDA NPs that could satisfy the following conditions, such as robust antioxidative activity, degradability and low toxicity, might act as intelligent treatments for clinical application.

Treatment of Ischemic Stroke

Ischemic stroke, caused by artery thrombi in blood vessels, in turn leads to excessive ROS production and the resulting damaged neurons during the I/R process.^{151,152}

Bioinspired melanin NPs (Me NPs) were employed as a novel antioxidant to treat ischemic stroke disease, as shown in Figure 5. The infarction area of the brain in a murine model was greatly reduced compared to that in the control.¹⁵³

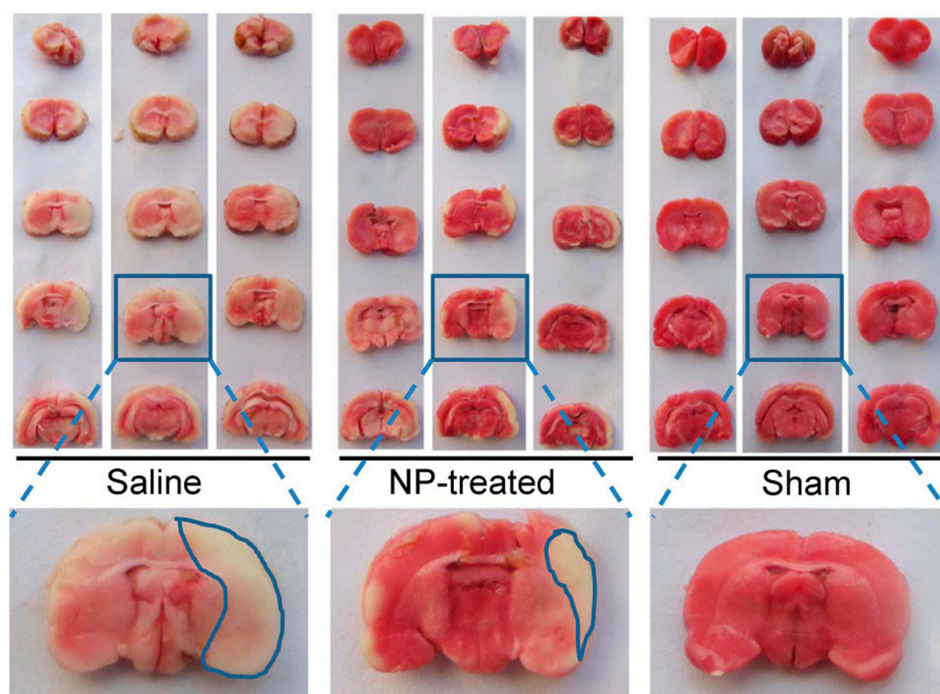


Figure 5 Comparison of the infarction area of 2,3,5-triphenyltetrazolium chloride-stained brain slices from different groups. Reprinted with permission from Liu Y, Ai K, Ji X et al. Comprehensive insights into the multi-antioxidative mechanisms of melanin nanoparticles and their application to protect brain from injury in ischemic stroke. *J Am Chem Soc.* 2017;139(2):856–862. Copyright (2017) American Chemical Society.¹⁵³

The melanin NPs not only scavenged ROS, including $O_2^{\cdot -}$, H_2O_2 , and $\cdot OH$, but also reduced inflammatory responses by inhibiting the expression of inflammatory mediators and cytokines.

A stable composite formed by human serum albumin (HSA) bound to the Mn_3O_4 nanozyme was used to alleviate nervous system injury caused by ischemic stroke reperfusion.¹⁵⁴ Owing to HSA, the as-designed nanozyme showed excellent stability, which was important for high bioavailability and prolonged blood circulation time. Moreover, Mn ions were released from HSA- Mn_3O_4 into bodily circulation and thus promoted SOD2 expression. PEG- and angiopep-2 (ANG)-modified CeO_2 nanozyme was loaded with edaravone for the treatment of ischemic stroke.⁶² PEGylation endowed CeO_2 nanozyme with superior characteristics of uniformity, small size, and prolonged blood circulation, while ANG led to effective cross blood–brain barrier (BBB) (through receptor-mediated transcytosis), which solved the problem of maintaining BBB integrity and the high accumulation need of nanozyme in damaged areas (Figure 6). The loaded edaravone and CeO_2 core both contributed to the removal of ROS, thus providing an efficient strategy for treating strokes. To solve the problems that exist in nanozyme treatment, such as less nanozyme accumulation in the ischemic brain site, a neutrophil-like cell membrane-coated mesoporous Prussian blue nanozyme (MPBzyme@NCM) was prepared.⁵⁰ The preparation of the nanomaterials translocated the entire cell membrane to the surface of the nanozyme and realized interaction between inflamed brain microvascular endothelial cells and neutrophils. The operation could help to promote nanozyme to access damaged brain and realize a noninvasive active-targeting therapy for ischemic stroke. PBNPs also polarized microglia from M1 to M2, decreased the accumulation of neutrophils, reduced neuronal apoptosis, and upregulated neurogenesis. This work proposed a new way to treat brain diseases by combining biomembranes and nanozymes.

Treatment of Neurological Diseases

The common feature of neurodegenerative diseases, such as AD and PD, is a gradual loss of neuron function. More deeply, the deposition of the peptide amyloid- β ($A\beta$) for plaques and the deposition of the protein tau for neurofibrillary tangles comprise the histopathological hallmarks of AD.^{155–157} Studies have confirmed that abnormal ROS affect the

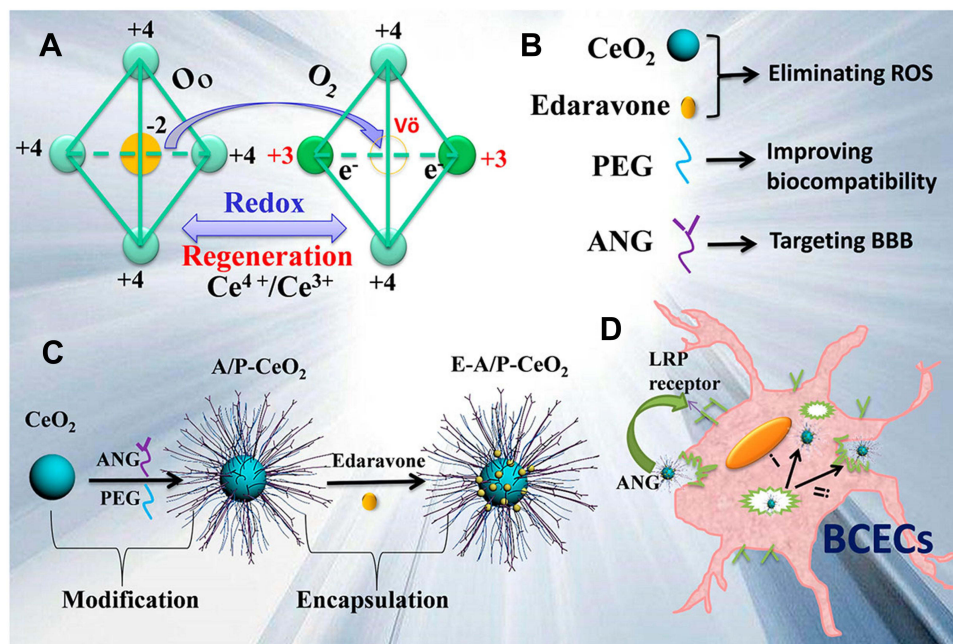


Figure 6 TPP-CeO₂ NPs acted as antioxidants for AD. **(A)** Formations of oxygen vacancies, cerium(III) and cerium(IV) species in CeO₂ NPs. **(B)** Main roles of components on edaravone-carried and PEG/ANG-conjugated CeO₂ NPs (E-A/P-CeO₂). **(C)** Synthetic process for E-A/P-CeO₂. **(D)** ANG-LRP endocytosis of E-A/P-CeO₂. Reprinted with permission from Bao Q, Hu P, Xu Y et al. Simultaneous blood-brain barrier crossing and protection for stroke treatment based on edaravone-loaded ceria nanoparticles. *ACS Nano*. 2018;12(7):6794–6805. Copyright (2018) American Chemical Society.⁶²

production and aggregation of A β peptides and metal ion homeostasis and cause mitochondrial dysfunction in AD.¹⁵⁸ ROS can also contribute to the degeneration of dopamine cells in PD.¹⁵⁹

For the treatment of AD, researchers synthesized small and positively charged triphenylphosphonium-conjugated CeO₂ NPs (TPP-CeO₂ NPs), which could enter mitochondria and destroy ROS to relieve oxidative stress in a 5XFAD transgenic AD mouse model.⁶⁴ Concretely, the derived nanoparticles mitigated reactive gliosis and maintained mitochondrial morphology. The TPP orientation verified the feasibility of mitochondrial therapeutics against neuroinflammation. Another strategy aimed at the accumulation of A β is shown in Figure 7.⁶⁵ A β antibody-conjugated magnetite/CeO₂ nanoparticle assemblies (MCNAs) with core/shell structures were synthesized to remove A β peptides from blood. When applying an external magnetic field, MCNAs isolated the captured A β peptides through magnetic force, while CeO₂ NPs in MCNAs scavenged ROS overproduced by the immune response. Collectively, the assemblies reduced the levels of blood A β and brain A β in 5XFAD transgenic mice and prevented spatial working memory deficits.

Ling and Tian et al exploited a new approach aimed at the tau pathway to deal with AD, involving tau hyperphosphorylation and aggregation of hyperphosphorylated tau.⁶⁶ As Figure 8 shows, the nanocomposite CeNC/IONC/MSN was formed by modifying CeO₂ and iron oxide nanocrystals on mesoporous silica NPs (MSN). Then, T807, which can bind tau protein, was immobilized on its surface; while methylene blue (MB), which can inhibit tau aggregation, was adsorbed into the pores of MSNs. CeO₂ scavenged ROS and suppressed tau hyperphosphorylation, while MB inhibited hyperphosphorylated tau aggregation. Through the cooperation of components, the as-nanocomposite successfully protected neuronal survival and ameliorated learning and memory impairments developed in AD.

PD is characterized by the accumulation of misfolded α -synuclein (a prion-like protein) into Lewy bodies, which plays a central role in the pathogenesis of PD.¹⁶⁰ A concept of proof that used nanozymes to fight against α -synuclein spreading was provided by Mao's group.¹²¹ PtCu nanoalloys (NAs) were prepared and possessed three enzyme-like activities, including peroxidase-like, catalase-like and SOD-like activities. The application of PtCu NAs in vitro or in vivo significantly inhibited α -synuclein preformed fibril (PFF)-induced α -synuclein pathology, cell death, and neuron-to-neuron transmission by scavenging ROS. Moreover, the PtCu NAs strongly inhibited α -synuclein spreading in the PD brain. Zheng and Cai et al considered that inflammasome-mediated pyroptosis may be a therapeutic target

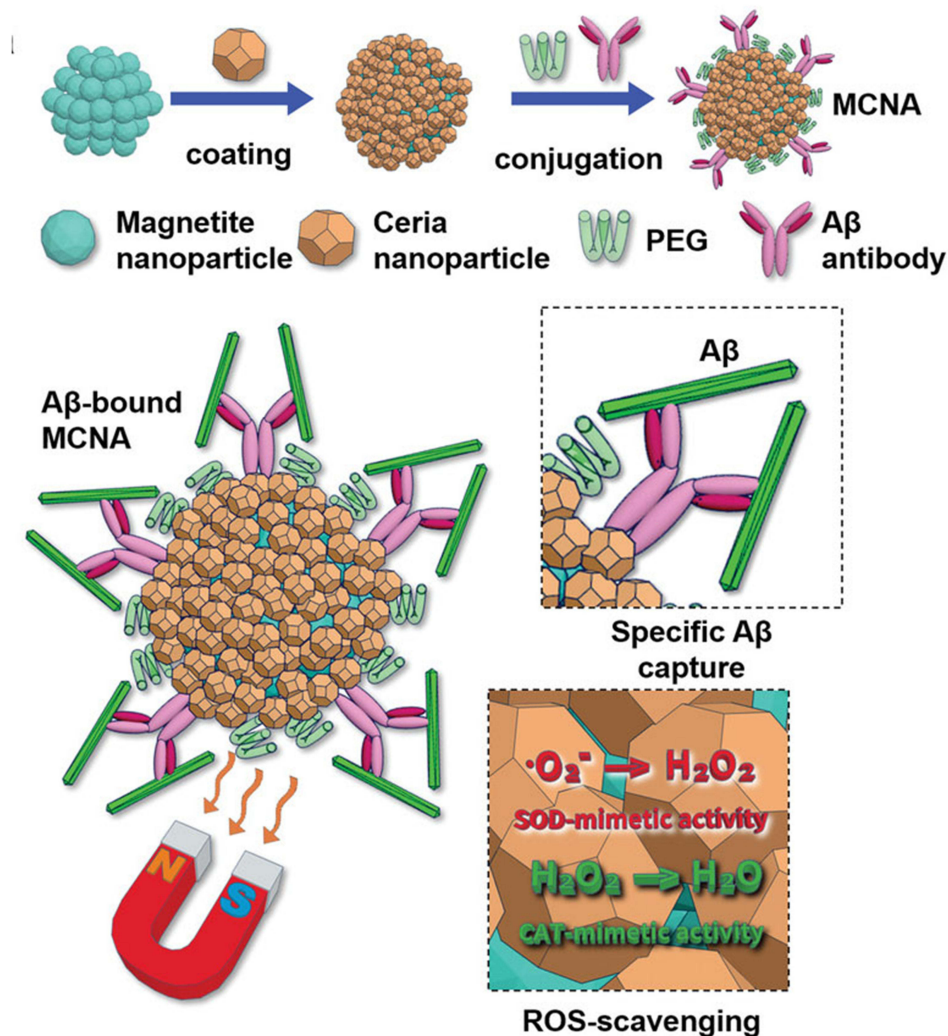


Figure 7 Schematic illustration of MCNA synthesis and mechanism. Reprinted with permission from Kim D, Kwon HJ, Hyeon T. Magnetite/ceria nanoparticle assemblies for extracorporeal cleansing of amyloid-beta in Alzheimer's disease. *Adv Mater.* 2019;31(19):e1807965. Copyright (2019) John Wiley and Sons.⁶⁵

for PD and discovered PB NPs as a pyroptosis inhibitor.¹⁶¹ Due to their potent ROS-scavenging activity, PB NPs inhibited the activation of the nucleotide-binding domain and leucine-rich repeat family pyrin domain containing 3 (NLRP3) inflammasome and caspase-1, reduced the cleavage of gasdermin D and the production of inflammatory factors, and effectively suppressed the pyroptosis of microglia in PD cell and mouse models. Although the study reveals a good effect of PB NPs on the treatment of PD, the deeper mechanism in vivo is currently unclear. The pharmacokinetics and long-term toxicity of PB NPs remain an issue of concern.

Treatment of Acute Liver/Kidney Injury

The researchers used $\text{Cu}_{5,4}\text{O}$ USNPs to treat acute kidney injury and acute liver injury, and verified that $\text{Cu}_{5,4}\text{O}$ USNPs functioned as antioxidants to eliminate ROS and induced remission of the diseases.²⁰ In the meantime, the ultrafine size of the nanoparticles accelerated renal clearance and conferred high biocompatibility. The therapeutic mechanism involved in acute kidney injury was uncovered in a further study. GSH metabolism, the MAPK signaling pathway, and the TNF signaling pathway, which exacerbated renal injury, were inhibited by $\text{Cu}_{5,4}\text{O}$ USNPs. The nanoparticles also maintained the expression of antioxidant genes, inhibited excessive proinflammatory factors such as TNF- α and interleukin-1 β (IL-1 β), and promoted the expression of kidney-associated genes. The $\text{Cu}_{5,4}\text{O}$ USNPs were reused three

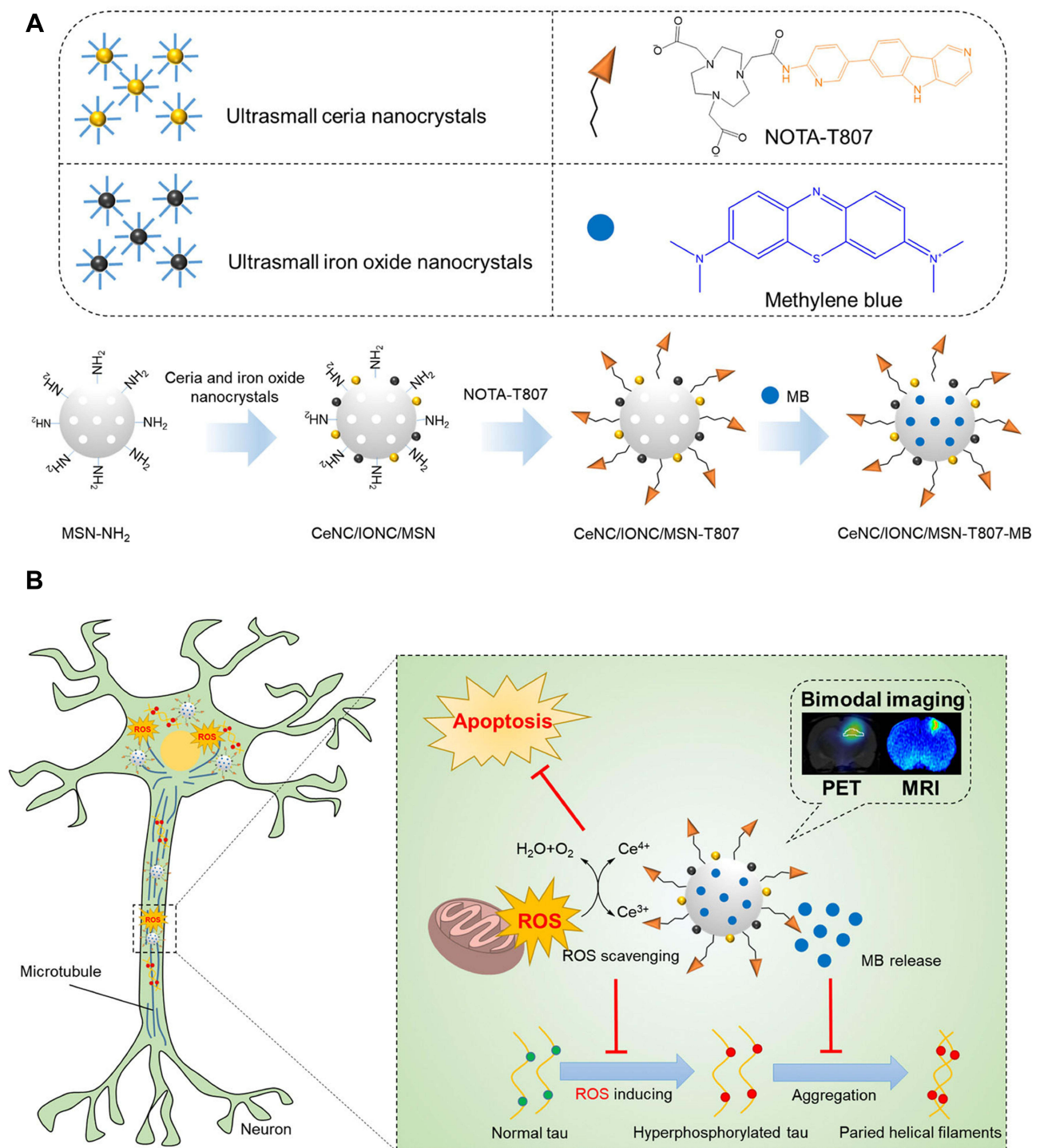


Figure 8 CeNC/IONC/MSN-T807-MB and its tau-targeted synergistic treatment for AD. **(A)** Schematic illustration of the progressive fabrication of CeNC/IONC/MSN-T807-MB. **(B)** Treatment mechanism of the nanocomposite. Reprinted with permission from Chen Q, Du Y, Zhang K et al. Tau-targeted multifunctional nanocomposite for combinational therapy of Alzheimer's disease. *ACS Nano*. 2018;12(2):1321–1338. Copyright (2018) American Chemical Society.⁶⁶

times to decompose H₂O₂. The results showed that the nanomaterials could maintain almost the same catalytic activity as the original solution, indicating that they had good stability and recyclability.

Mo-based polyoxometalate (POM) nanoclusters were synthesized with potent ROS-scavenging activity owing to the dual oxidation states of Mo⁵⁺/Mo⁶⁺.¹⁶² Furthermore, POM nanoclusters acted as new antioxidants for ameliorating acute kidney injury in a murine model. By positron emission tomography (PET) imaging, radionuclide ⁸⁹Zr-labeled POM nanoclusters

were observed to be more likely to accumulate in the injured kidneys due to their ultrasmall hydrodynamic diameter. The treatment effect demonstrated that POM improved the renal function of acute kidney injury mice with excellent safety.

The therapeutic mechanisms of PBNPs were probed in anthracycline-induced liver injury.⁴⁶ PBNPs activated the Nrf2/ARE pathway to upregulate antioxidative genes, thus reducing oxidative stress in response to liver injury. They also attenuated inflammatory reactions by decreasing the expression levels of myeloperoxidase and F4/80 (a specific indicator for macrophages) in the liver.

Others

The main pathological features of depression, a serious mental disease, are oxidative stress and excessive ROS. Thus, Zheng and Zhang et al considered using CeO₂ nanozyme as a novel drug to treat depression.⁶⁷ To overcome the disadvantage of CeO₂, such as its large size, bovine serum albumin (BSA) served as a space constraint to avoid nanoparticle aggregation. The as-designed CeO₂@BSA nanoclusters were endowed with an ultrasmall size as small as 2 nm, potent ROS scavenging activity and BBB penetration ability. When used in a depressive model induced by chronic restraint stress, CeO₂@BSA decreased depression-like behaviors, relieved neuroinflammation and provided neuroprotection with few side effects.

Conclusion and Future Perspective

An imbalance of ROS leads to oxidative stress and is linked to various diseases, including diabetes, neurodegeneration, and aging.⁴ Nanozymes with intrinsic antioxidant ability are widely employed to treat ROS-involved diseases.

In this review, we mainly introduce considerable experimental studies that showed satisfactory performances of nanozymes in disease models such as inflammation, neurodegenerative diseases and stroke. In addition, we summarize the enzyme-mimicking traits of nanozymes and show the rational design of nanozymes to improve antioxidant enzyme-like activities. For example, shapes, surface areas, surface facets, particle morphologies, porous structures, pore sizes and volumes, metal doping, and combinations of different functional enzymes may lead to the different catalytic abilities of nanozymes. Although abundant efforts have been made on nanozymes in ROS-involved diseases, there are still shortcomings and deficiencies that need further improvement.

1. The low catalytic activity and poor substrate selectivity of nanozymes are still big problems, which will be a huge stumbling block for their practical application in the future. According to the above summary, doping, control of shape and morphology, forming complex nanozymes, single atom technology, etc., are effective measures to improve the activity of nanozymes. A biomimetic strategy that simulates the active sites of natural enzymes may help to enhance the efficiency of nanozymes.
2. It is well known that living organisms are complex systems. When nanozymes are used in vivo, their pharmacokinetics, ie, the absorption, distribution, biochemical conversion and excretion of nanozymes in the body, need a detailed examination, and based on this information, the application prospects of nanozymes can be overall assessed. In addition, the lack of detailed evaluation of the biosafety and biocompatibility of nanozymes is another problem. Fears have been raised about the potential toxicity of nanozymes on biological systems. The problem in the usage of nanozymes in the central nervous system also includes BBB permeability. It is worth noting that the compatibility of nanozymes can be improved by the loading of natural proteins, which may be a feasible strategy.^{163,164}
3. At present, the use of nanozymes for ROS-related diseases involves AD, PD, colitis, acute kidney injury and acute liver injury, which are serious and incurable; however, the treatment should expand to diseases that are not life threatening but also bring pain to patients, such as oral ulcers caused by radiation therapy and difficult-to-heal wounds.

With respect to the problems above, further refinement for high-performance, excellent biosafety, and detailed research in diverse animal models, nanozymes, which are easy to synthesize and low-cost, may be further developed and hold promise as a new therapeutic strategy for ROS-related diseases in clinics.

Funding

This work is supported by the Medical Key Science Project of Shanxi Province, No. 2020XM52, the Scientific and Technological Activities for Overseas Students in Shanxi Province, No. 20200042, and Priority Academic Program Development of Jiangsu Higher Education institutions (PAPD).

Disclosure

The authors report no conflicts of interest in this work.

References

1. Turrens JF. Mitochondrial formation of reactive oxygen species. *J Physiol*. 2003;552(2):335–344. doi:10.1113/jphysiol.2003.049478
2. Held P. An introduction to reactive oxygen species. *Tech Resources-App Guides*. 2012;802:5–9.
3. Bayir H. Reactive oxygen species. *Crit Care Med*. 2005;33(12 Suppl):S498–501. doi:10.1097/01.ccm.0000186787.64500.12
4. Ray PD, Huang BW, Tsuji Y. Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling. *Cell Signal*. 2012;24(5):981–990. doi:10.1016/j.cellsig.2012.01.008
5. Dhiman A, Handa M, Ruwali M, Singh DP, Kesharwani P, Shukla R. Recent trends of natural based therapeutics for mitochondria targeting in Alzheimer's disease. *Mitochondrion*. 2022;64:112–124. doi:10.1016/j.mito.2022.03.006
6. Mittler R. ROS are good. *Trends in Plant Sci*. 2017;22(1):11–19. doi:10.1016/j.tplants.2016.08.002
7. Checa J, Aran JM. Reactive oxygen species: drivers of physiological and pathological processes. *J Inflamm Res*. 2020;13:1057. doi:10.2147/JIR.S275595
8. Kang DH. Oxidative stress, DNA damage, and breast cancer. *AACN Adv Crit Care*. 2002;13(4):540–549. doi:10.1097/00044067-200211000-00007
9. Monaghan P, Metcalfe NB, Torres R. Oxidative stress as a mediator of life history trade-offs: mechanisms, measurements and interpretation. *Ecol Lett*. 2009;12(1):75–92. doi:10.1111/j.1461-0248.2008.01258.x
10. Andersen JK. Oxidative stress in neurodegeneration: cause or consequence? *Nat Med*. 2004;10(S7):S18–25. doi:10.1038/nrm1434
11. Melo A, Monteiro L, Lima RM, de Oliveira DM, de Cerqueira MD, El-Bachá RS. Oxidative stress in neurodegenerative diseases: mechanisms and therapeutic perspectives. *Oxid Med Cell Longev*. 2011;2011:1–14. doi:10.1155/2011/467180
12. Mittal M, Siddiqui MR, Tran K, Reddy SP, Malik AB. Reactive oxygen species in inflammation and tissue injury. *Antioxid Redox Signal*. 2014;20(7):1126–1167. doi:10.1089/ars.2012.5149
13. Amani H, Habibe R, Hajmiresmail SJ, Latifi S, Pazoki-Toroudi H, Akhavan O. Antioxidant nanomaterials in advanced diagnoses and treatments of ischemia reperfusion injuries. *J Mater Chem B*. 2017;5(48):9452–9476. doi:10.1039/c7tb01689a
14. Droge W. Free radicals in the physiological control of cell function. *Physiol Rev*. 2002;82(1):47–95. doi:10.1152/physrev.00018.2001
15. Wei H, Wang E. Nanomaterials with enzyme-like characteristics (nanozymes): next-generation artificial enzymes. *Chem Soc Rev*. 2013;42(14):6060–6093. doi:10.1039/c3cs35486e
16. Gao L, Zhuang J, Nie L, et al. Intrinsic peroxidase-like activity of ferromagnetic nanoparticles. *Nat Nanotechnol*. 2007;2(9):577–583. doi:10.1038/nnano.2007.260
17. Yu X, Xu Z, Wang X, Xu Q, Chen J. Bactrian camel serum albumins-based nanocomposite as versatile biocargo for drug delivery, biocatalysis and detection of hydrogen peroxide. *Mater Sci Eng C*. 2020;109:110627. doi:10.1016/j.msec.2020.110627
18. Wang D, Jana D, Zhao Y. Metal–organic framework derived nanozymes in biomedicine. *Acc Chem Res*. 2020;53(7):1389–1400. doi:10.1021/acs.accounts.0c00268
19. Yan X, Gao L. Nanozymology: an Overview. In: Yan X, editor. *Nanozymology. Nanostructure Science and Technology*. Singapore: Springer; 2020:3–16. doi:10.1007/978-981-15-1490-6_1
20. Liu T, Xiao B, Xiang F, et al. Ultrasmall copper-based nanoparticles for reactive oxygen species scavenging and alleviation of inflammation related diseases. *Nat Commun*. 2020;11(1):2788. doi:10.1038/s41467-020-16544-7
21. Wu J, Wang X, Wang Q, et al. Nanomaterials with enzyme-like characteristics (nanozymes): next-generation artificial enzymes (II). *Chem Soc Rev*. 2019;48(4):1004–1076. doi:10.1039/c8cs00457a
22. Gaur M, Misra C, Yadav AB, et al. Biomedical applications of carbon nanomaterials: fullerenes, quantum dots, nanotubes, nanofibers, and graphene. *Materials*. 2021;14(20):5978. doi:10.3390/ma14205978
23. Karimi-Maleh H, Beitollahi H, Kumar PS, et al. Recent advances in carbon nanomaterials-based electrochemical sensors for food azo dyes detection. *Food Chem Toxicol*. 2022;164:112961. doi:10.1016/j.fct.2022.112961
24. Dellinger A, Zhou Z, Connor J, et al. Application of fullerenes in nanomedicine: an update. *Nanomedicine*. 2013;8(7):1191–1208. doi:10.2217/nmm.13.99
25. Speranza G. Carbon nanomaterials: synthesis, functionalization and sensing applications. *Nanomaterials*. 2021;11(4):967. doi:10.3390/nano11040967
26. Bakry R, Vallant RM, Najam-ul-Haq M, et al. Medicinal applications of fullerenes. *Int J Nanomedicine*. 2007;2(4):639–649.
27. Xiao L, Takada H, Gan X, Miwa N. The water-soluble fullerene derivative “Radical Sponge” exerts cytoprotective action against UVA irradiation but not visible-light-catalyzed cytotoxicity in human skin keratinocytes. *Bioorg Med Chem Lett*. 2006;16(6):1590–1595. doi:10.1016/j.bmcl.2005.12.011
28. Castro E, Garcia AH, Zavala G, Echegoyen L. Fullerenes in Biology and Medicine. *J Mater Chem B*. 2017;5(32):6523–6535. doi:10.1039/C7TB00855D
29. Cheng X, Ni X, Wu R, et al. Evaluation of the structure–activity relationship of carbon nanomaterials as antioxidants. *Nanomedicine*. 2018;13(7):733–747. doi:10.2217/nmm-2017-0314

30. Yudoh K, Karasawa R, Masuko K, Kato T. Water-soluble fullerene (C60) inhibits the development of arthritis in the rat model of arthritis. *Int J Nanomedicine*. 2009;4:217–225. doi:10.2147/ijn.s7653
31. Hao T, Li J, Yao F, et al. Injectable fullerene/alginic acid hydrogel for suppression of oxidative stress damage in brown adipose-derived stem cells and cardiac repair. *ACS Nano*. 2017;11(6):5474–5488. doi:10.1021/acsnano.7b00221
32. Aqel A, Abou El-Nour KM, Ammar RA, Al-Warthan A. Carbon nanotubes, science and technology part (I) structure, synthesis and characterisation. *Arab J Chem*. 2012;5(1):1–23. doi:10.1016/j.arabjc.2010.08.022
33. Fenoglio I, Tomatis M, Lison D, et al. Reactivity of carbon nanotubes: free radical generation or scavenging activity? *Free Radic Biol Med*. 2006;40(7):1227–1233. doi:10.1016/j.freeradbiomed.2005.11.010
34. Qiu Y, Wang Z, Owens AC, et al. Antioxidant chemistry of graphene-based materials and its role in oxidation protection technology. *Nanoscale*. 2014;6(20):11744–11755. doi:10.1039/c4nr03275f
35. Ren C, Hu X, Zhou Q. Graphene oxide quantum dots reduce oxidative stress and inhibit neurotoxicity in vitro and in vivo through catalase-like activity and metabolic regulation. *Adv Sci*. 2018;5(5):1700595. doi:10.1002/advs.201700595
36. Han J, Kim YS, Lim MY, et al. Dual roles of graphene oxide to attenuate inflammation and elicit timely polarization of macrophage phenotypes for cardiac repair. *ACS Nano*. 2018;12(2):1959–1977. doi:10.1021/acsnano.7b09107
37. Chen Z, Yin JJ, Zhou YT, et al. Dual enzyme-like activities of iron oxide nanoparticles and their implication for diminishing cytotoxicity. *ACS Nano*. 2012;6(5):4001–4012. doi:10.1021/nn300291r
38. Gao L, Fan K, Yan X. Iron oxide nanozyme: a multifunctional enzyme mimetic for biomedical applications. *Theranostics*. 2017;7(13):3207–3227. doi:10.7150/thno.19738
39. Zhang Y, Wang Z, Li X, et al. Dietary iron oxide nanoparticles delay aging and ameliorate neurodegeneration in *Drosophila*. *Adv Mater*. 2016;28(7):1387–1393. doi:10.1002/adma.201503893
40. Wei Z, Wang L, Tang C, et al. Metal-phenolic networks nanoplatform to mimic antioxidant defense system for broad-spectrum radical eliminating and endotoxemia treatment. *Adv Funct Mater*. 2020;30(49):2002234. doi:10.1002/adfm.202002234
41. Zhang W, Ma D, Du J. Prussian blue nanoparticles as peroxidase mimetics for sensitive colorimetric detection of hydrogen peroxide and glucose. *Talanta*. 2014;120:362–367. doi:10.1016/j.talanta.2013.12.028
42. Zhang W, Hu S, Yin JJ, et al. Prussian blue nanoparticles as multienzyme mimetics and reactive oxygen species scavengers. *J Am Chem Soc*. 2016;138(18):5860–5865. doi:10.1021/jacs.5b12070
43. Zhao J, Cai X, Gao W, et al. Prussian blue nanozyme with multienzyme activity reduces colitis in mice. *ACS Appl Mater Interfaces*. 2018;10(31):26108–26117. doi:10.1021/acsami.8b10345
44. Estelrich J, Busquets MA. Prussian blue: a nanozyme with versatile catalytic properties. *Int J Mol Sci*. 2021;22(11):5993. doi:10.3390/ijms22115993
45. Chen J, Wang Q, Huang L, et al. Prussian blue with intrinsic heme-like structure as peroxidase mimic. *Nano Res*. 2018;11(9):4905–4913. doi:10.1007/s12274-018-2079-8
46. Bai H, Kong F, Feng K, et al. Prussian blue nanozymes prevent anthracycline-induced liver injury by attenuating oxidative stress and regulating inflammation. *ACS Appl Mater Interfaces*. 2021;13(36):42382–42395. doi:10.1021/acsami.1c09838
47. Xie X, Zhao J, Gao W, et al. Prussian blue nanozyme-mediated nanoscavenger ameliorates acute pancreatitis via inhibiting TLRs/NF- κ B signaling pathway. *Theranostics*. 2021;11(7):3213–3228. doi:10.7150/thno.52010
48. Sahu A, Jeon J, Lee MS, Yang HS, Tae G. Antioxidant and anti-inflammatory activities of Prussian blue nanozyme promotes full-thickness skin wound healing. *Mater Sci Eng C Mater Biol Appl*. 2021;119:111596. doi:10.1016/j.msec.2020.111596
49. Zhang K, Tu M, Gao W, et al. Hollow Prussian blue nanozymes drive neuroprotection against ischemic stroke via attenuating oxidative stress, counteracting inflammation, and suppressing cell apoptosis. *Nano Lett*. 2019;19(5):2812–2823. doi:10.1021/acs.nanolett.8b04729
50. Feng L, Dou C, Xia Y, et al. Neutrophil-like cell-membrane-coated nanozyme therapy for ischemic brain damage and long-term neurological functional recovery. *ACS Nano*. 2021;15(2):2263–2280. doi:10.1021/acsnano.0c07973
51. He W, Zhou YT, Wamer WG, et al. Intrinsic catalytic activity of Au nanoparticles with respect to hydrogen peroxide decomposition and superoxide scavenging. *Biomaterials*. 2013;34(3):765–773. doi:10.1016/j.biomaterials.2012.10.010
52. Liu Z, Shen Y, Wu Y, et al. An intrinsic therapy of gold nanoparticles in focal cerebral ischemia-reperfusion injury in rats. *J Biomed Nanotechnol*. 2013;9(6):1017–1028. doi:10.1166/jbn.2013.1597
53. Liu CP, Wu TH, Lin YL, Liu CY, Wang S, Lin SY. Tailoring enzyme-like activities of gold nanoclusters by polymeric tertiary amines for protecting neurons against oxidative stress. *Small*. 2016;12(30):4127–4135. doi:10.1002/sml.201503919
54. Schubert D, Dargusch R, Raitano J, Chan SW. Cerium and yttrium oxide nanoparticles are neuroprotective. *Biochem Biophys Res Commun*. 2006;342(1):86–91. doi:10.1016/j.bbrc.2006.01.129
55. Pirmohamed T, Dowding JM, Singh S, et al. Nanoceria exhibit redox state-dependent catalase mimetic activity. *Chem Commun (Camb)*. 2010;46(16):2736–2738. doi:10.1039/b922024k
56. Li Y, He X, Yin JJ, et al. Acquired superoxide-scavenging ability of ceria nanoparticles. *Angew Chem Int Ed Engl*. 2015;54(6):1832–1835. doi:10.1002/anie.201410398
57. Akhtar MJ, Ahamed M, Alhadlaq HA, Khan MAM, Alrokayan SA. Glutathione replenishing potential of CeO₂ nanoparticles in human breast and fibrosarcoma cells. *J Colloid Interface Sci*. 2015;453:21–27. doi:10.1016/j.jcis.2015.04.049
58. Hirst SM, Karakoti AS, Tyler RD, Sriranganathan N, Seal S, Reilly CM. Anti-inflammatory properties of cerium oxide nanoparticles. *Small*. 2009;5(24):2848–2856. doi:10.1002/sml.200901048
59. Pagliari F, Mandoli C, Forte G, et al. Cerium oxide nanoparticles protect cardiac progenitor cells from oxidative stress. *ACS Nano*. 2012;6(5):3767–3775. doi:10.1021/nn2048069
60. Dowding JM, Song W, Bossy K, et al. Cerium oxide nanoparticles protect against Abeta-induced mitochondrial fragmentation and neuronal cell death. *Cell Death Differ*. 2014;21(10):1622–1632. doi:10.1038/cdd.2014.72
61. Singh R, Singh S. Redox-dependent catalase mimetic cerium oxide-based nanozyme protect human hepatic cells from 3-AT induced acatalasemia. *Colloids Surf B Biointerfaces*. 2019;175:625–635. doi:10.1016/j.colsurfb.2018.12.042
62. Bao Q, Hu P, Xu Y, et al. Simultaneous blood-brain barrier crossing and protection for stroke treatment based on edaravone-loaded ceria nanoparticles. *ACS Nano*. 2018;12(7):6794–6805. doi:10.1021/acsnano.8b01994

63. Kim CK, Kim T, Choi IY, et al. Ceria nanoparticles that can protect against ischemic stroke. *Angew Chem Int Ed Engl.* 2012;51(44):11039–11043. doi:10.1002/anie.201203780
64. Kwon HJ, Cha MY, Kim D, et al. Mitochondria-targeting ceria nanoparticles as antioxidants for Alzheimer's disease. *ACS Nano.* 2016;10(2):2860–2870. doi:10.1021/acsnano.5b08045
65. Kim D, Kwon HJ, Hyeon T. Magnetite/ceria nanoparticle assemblies for extracellular cleansing of amyloid-beta in Alzheimer's disease. *Adv Mater.* 2019;31(19):e1807965. doi:10.1002/adma.201807965
66. Chen Q, Du Y, Zhang K, et al. Tau-targeted multifunctional nanocomposite for combinational therapy of Alzheimer's disease. *ACS Nano.* 2018;12(2):1321–1338. doi:10.1021/acsnano.7b07625
67. Fu S, Chen H, Yang W, et al. ROS-targeted depression therapy via BSA-incubated ceria nanoclusters. *Nano Lett.* 2022;22(11):4519–4527. doi:10.1021/acs.nanolett.2c01334
68. Kim J, Hong G, Mazaleuskaya L, et al. Ultrasmall antioxidant cerium oxide nanoparticles for regulation of acute inflammation. *ACS Appl Mater Interfaces.* 2021;13(51):60852–60864. doi:10.1021/acscami.1c16126
69. Zhao S, Li Y, Liu Q, et al. An orally administered CeO₂ montmorillonite nanozyme targets inflammation for inflammatory bowel disease therapy. *Adv Funct Mater.* 2020;30(45). doi:10.1002/adfm.202004692
70. Lin A, Sun Z, Xu X, et al. Self-cascade uricase/catalase mimics alleviate acute gout. *Nano Lett.* 2022;22(1):508–516. doi:10.1021/acs.nanolett.1c04454
71. Wu L, Liu G, Wang W, et al. Cyclodextrin-modified CeO₂ nanoparticles as a multifunctional nanozyme for combinational therapy of psoriasis. *Int J Nanomedicine.* 2020;15:2515–2527. doi:10.2147/IJN.S246783
72. Soh M, Kang DW, Jeong HG, et al. Ceria-zirconia nanoparticles as an enhanced multi-antioxidant for sepsis treatment. *Angew Chem Int Ed Engl.* 2017;56(38):11399–11403. doi:10.1002/anie.201704904
73. Li F, Qiu Y, Xia F, et al. Dual detoxification and inflammatory regulation by ceria nanozymes for drug-induced liver injury therapy. *Nano Today.* 2020;35:100925. doi:10.1016/j.nantod.2020.100925
74. Adebayo OA, Akinloye O, Adaramoye OA. Cerium oxide nanoparticles attenuate oxidative stress and inflammation in the liver of diethylnitrosamine-treated mice. *Biol Trace Elem Res.* 2020;193(1):214–225. doi:10.1007/s12011-019-01696-5
75. Kwon HJ, Kim D, Seo K, et al. Ceria nanoparticle systems for selective scavenging of mitochondrial, intracellular, and extracellular reactive oxygen species in Parkinson's disease. *Angew Chem Int Ed Engl.* 2018;57(30):9408–9412. doi:10.1002/anie.201805052
76. Heckman KL, DeCoteau W, Estevez A, et al. Custom cerium oxide nanoparticles protect against a free radical mediated autoimmune degenerative disease in the brain. *ACS Nano.* 2013;7(12):10582–10596. doi:10.1021/nn403743b
77. Yu Y, Zhao S, Gu D, et al. Cerium oxide nanozyme attenuates periodontal bone destruction by inhibiting the ROS-NFκB pathway. *Nanoscale.* 2022;14(7):2628–2637. doi:10.1039/d1nr06043k
78. Korschelt K, Ragg R, Metzger CS, et al. Glycine-functionalized copper(ii) hydroxide nanoparticles with high intrinsic superoxide dismutase activity. *Nanoscale.* 2017;9(11):3952–3960. doi:10.1039/c6nr09810j
79. Hao C, Qu A, Xu L, et al. Chiral molecule-mediated porous Cu_xO nanoparticle clusters with antioxidation activity for ameliorating Parkinson's disease. *J Am Chem Soc.* 2019;141(2):1091–1099. doi:10.1021/jacs.8b11856
80. Liu X, Wang Q, Zhao H, Zhang L, Su Y, Lv Y. BSA-templated MnO₂ nanoparticles as both peroxidase and oxidase mimics. *Analyst.* 2012;137(19):4552–4558. doi:10.1039/c2an35700c
81. Li W, Liu Z, Liu C, Guan Y, Ren J, Qu X. Manganese dioxide nanozymes as responsive cytoprotective shells for individual living cell encapsulation. *Angew Chem Int Ed Engl.* 2017;56(44):13661–13665. doi:10.1002/anie.201706910
82. Yao J, Cheng Y, Zhou M, et al. ROS scavenging Mn₃O₄ nanozymes for in vivo anti-inflammation. *Chem Sci.* 2018;9(11):2927–2933. doi:10.1039/c7sc05476a
83. Singh N, Savanur MA, Srivastava S, D'Silva P, Mughesh G. A manganese oxide nanozyme prevents the oxidative damage of biomolecules without affecting the endogenous antioxidant system. *Nanoscale.* 2019;11(9):3855–3863. doi:10.1039/c8nr09397k
84. Singh N, Savanur MA, Srivastava S, D'Silva P, Mughesh G. A redox modulatory Mn₃O₄ nanozyme with multi-enzyme activity provides efficient cytoprotection to human cells in a Parkinson's disease model. *Angew Chem Int Ed Engl.* 2017;56(45):14267–14271. doi:10.1002/anie.201708573
85. Huang Y, Liu C, Pu F, Liu Z, Ren J, Qu X. A GO-Se nanocomposite as an antioxidant nanozyme for cytoprotection. *Chem Commun (Camb).* 2017;53(21):3082–3085. doi:10.1039/c7cc00045f
86. Huang Y, Liu Z, Liu C, Zhang Y, Ren J, Qu X. Selenium-based nanozyme as biomimetic antioxidant machinery. *Chemistry.* 2018;24(40):10224–10230. doi:10.1002/chem.201801725
87. Chen X, Zhu X, Gong Y, et al. Porous selenium nanozymes targeted scavenging ROS synchronize therapy local inflammation and sepsis injury. *Appl Mater Today.* 2021;22. doi:10.1016/j.apmt.2020.100929
88. Chen B, Xiang S, Qian G. Metal-organic frameworks with functional pores for recognition of small molecules. *Acc Chem Res.* 2010;43(8):1115–1124. doi:10.1021/ar100023y
89. Kreno LE, Leong K, Farha OK, Allendorf M, Van Duyne RP, Hupp JT. Metal-organic framework materials as chemical sensors. *Chem Rev.* 2012;112(2):1105–1125. doi:10.1021/cr200324t
90. Tan H, Ma C, Gao L, et al. Metal-organic framework-derived copper nanoparticle@ carbon nanocomposites as peroxidase mimics for colorimetric sensing of ascorbic acid. *Chem Eur J.* 2014;20(49):16377–16383. doi:10.1002/chem.201404960
91. Zhang JW, Zhang HT, Du ZY, Wang X, Yu SH, Jiang HL. Water-stable metal-organic frameworks with intrinsic peroxidase-like catalytic activity as a colorimetric biosensing platform. *Chem Commun (Camb).* 2014;50(9):1092–1094. doi:10.1039/c3cc48398c
92. Qi Z, Wang L, You Q, Chen Y. PA-Tb-Cu MOF as luminescent nanoenzyme for catalytic assay of hydrogen peroxide. *Biosens Bioelectron.* 2017;96:227–232. doi:10.1016/j.bios.2017.05.013
93. Chen WH, Vazquez-Gonzalez M, Kozell A, Ceconello A, Willner I. Cu²⁺-modified metal-organic framework nanoparticles: a peroxidase-mimicking nanoenzyme. *Small.* 2018;14(5). doi:10.1002/sml.201703149
94. Zheng HQ, Liu CY, Zeng XY, et al. MOF-808: a metal-organic framework with intrinsic peroxidase-like catalytic activity at neutral pH for colorimetric biosensing. *Inorg Chem.* 2018;57(15):9096–9104. doi:10.1021/acs.inorgchem.8b01097

95. Zhang K, Meng X, Cao Y, et al. Metal-organic framework nanoshuttle for synergistic photodynamic and low-temperature photothermal therapy. *Adv Funct Mater.* 2018;28(42):1804634. doi:10.1002/adfm.201804634
96. Li H, Cao X, Fei X, Zhang S, Xian Y. Nanoscaled luminescent terbium metal-organic frameworks for measuring and scavenging reactive oxygen species in living cells. *J Mater Chem B.* 2019;7(18):3027–3033. doi:10.1039/c9tb00361d
97. Zhang L, Zhang Y, Wang Z, et al. Constructing metal-organic framework nanodots as bio-inspired artificial superoxide dismutase for alleviating endotoxemia. *Mater Horiz.* 2019;6(8):1682–1687. doi:10.1039/c9mh00339h
98. Liu H, Yang Y, Liu Y, et al. Melanin-like nanomaterials for advanced biomedical applications: a versatile platform with extraordinary promise. *Adv Sci.* 2020;7(7):1903129. doi:10.1002/advs.201903129
99. Yue Y, Zhao X. Melanin-like nanomedicine in photothermal therapy applications. *Int J Mol Sci.* 2021;22(1):399. doi:10.3390/ijms22010399
100. Tada M, Kohno M, Niwano Y. Scavenging or quenching effect of melanin on superoxide anion and singlet oxygen. *J Clin Biochem Nutr.* 2010;46:224–228. doi:10.3164/jcbn.09-84
101. Liu Y, Ai K, Lu L. Polydopamine and its derivative materials: synthesis and promising applications in energy, environmental, and biomedical fields. *Chem Rev.* 2014;114(9):5057–5115. doi:10.1021/cr400407a
102. Hu J, Yang L, Yang P, Jiang S, Liu X, Li Y. Polydopamine free radical scavengers. *Biomater Sci.* 2020;8(18):4940–4950. doi:10.1039/d0bm01070g
103. Sun T, Jiang D, Rosenkrans ZT, et al. A melanin-based natural antioxidant defense nanosystem for theranostic application in acute kidney injury. *Adv Funct Mater.* 2019;29(48):1904833. doi:10.1002/adfm.201904833
104. Yang XF, Wang A, Qiao B, Li J, Liu J, Zhang T. Single-atom catalysts: a new frontier in heterogeneous catalysis. *Acc Chem Res.* 2013;46(8):1740–1748. doi:10.1021/ar300361m
105. Jiao L, Yan H, Wu Y, et al. When nanozymes meet single-atom catalysis. *Angew Chem Int Ed Engl.* 2020;59(7):2565–2576. doi:10.1002/anie.201905645
106. Pei J, Zhao R, Mu X, Wang J, Liu C, Zhang XD. Single-atom nanozymes for biological applications. *Biomater Sci.* 2020;8(23):6428–6441. doi:10.1039/d0bm01447h
107. Cao F, Zhang L, You Y, Zheng L, Ren J, Qu X. An enzyme-mimicking single-atom catalyst as an efficient multiple reactive oxygen and nitrogen species scavenger for sepsis management. *Angewandte Chemie.* 2020;132(13):5146–5153. doi:10.1002/ange.201912182
108. Yan R, Sun S, Yang J, et al. Nanozyme-based bandage with single-atom catalysis for brain trauma. *ACS nano.* 2019;13(10):11552–11560. doi:10.1021/acsnano.9b05075
109. Korsvik C, Patil S, Seal S, Self WT. Superoxide dismutase mimetic properties exhibited by vacancy engineered ceria nanoparticles. *Chem Commun (Camb).* 2007;(10):1056–1058. doi:10.1039/b615134e
110. Xi Z, Gao W, Xia X. Size effect in Pd-Ir core-shell nanoparticles as nanozymes. *Chembiochem.* 2020;21(17):2440–2444. doi:10.1002/cbic.202000147
111. Xu R, Wang D, Zhang J, Li Y. Shape-dependent catalytic activity of silver nanoparticles for the oxidation of styrene. *Chem Asian J.* 2006;1(6):888–893. doi:10.1002/asia.200600260
112. Ge C, Fang G, Shen X, et al. Facet energy versus enzyme-like activities: the unexpected protection of palladium nanocrystals against oxidative damage. *ACS Nano.* 2016;10(11):10436–10445. doi:10.1021/acsnano.6b06297
113. He W, Wu X, Liu J, et al. Design of AgM bimetallic alloy nanostructures (M = Au, Pd, Pt) with tunable morphology and peroxidase-like activity. *Chem Mater.* 2010;22(9):2988–2994. doi:10.1021/cm100393v
114. Wu J, Qin K, Yuan D, et al. Rational design of Au@Pt multibranch nanostructures as bifunctional nanozymes. *ACS Appl Mater Interfaces.* 2018;10(15):12954–12959. doi:10.1021/acsnano.7b17945
115. Xia X, Zhang J, Lu N, et al. Pd-Ir core-shell nanocubes: a type of highly efficient and versatile peroxidase mimic. *ACS Nano.* 2015;9(10):9994–10004. doi:10.1021/acsnano.5b03525
116. Gao Z, Ye H, Tang D, et al. Platinum-decorated gold nanoparticles with dual functionalities for ultrasensitive colorimetric in vitro diagnostics. *Nano Lett.* 2017;17(9):5572–5579. doi:10.1021/acs.nanolett.7b02385
117. Lv F, Gong Y, Cao Y, et al. A convenient detection system consisting of efficient Au@PtRu nanozymes and alcohol oxidase for highly sensitive alcohol biosensing. *Nanoscale Adv.* 2020;2(4):1583–1589. doi:10.1039/d0na00002g
118. Mu X, Wang J, Li Y, et al. Redox trimetallic nanozyme with neutral environment preference for brain injury. *ACS Nano.* 2019;13(2):1870–1884. doi:10.1021/acsnano.8b08045
119. Wang Q, Cheng C, Zhao S, et al. A valence-engineered self-cascading antioxidant nanozyme for the therapy of inflammatory bowel disease. *Angew Chem Int Ed Engl.* 2022. doi:10.1002/anie.202201101
120. Wang C, Ren G, Yuan B, et al. Enhancing enzyme-like activities of Prussian blue analog nanocages by molybdenum doping: toward cytoprotecting and online optical hydrogen sulfide monitoring. *Anal Chem.* 2020;92(11):7822–7830. doi:10.1021/acs.analchem.0c01028
121. Liu Y-Q, Mao Y, Xu E, et al. Nanozyme scavenging ROS for prevention of pathologic α -synuclein transmission in Parkinson's disease. *Nano Today.* 2021;36:101027. doi:10.1016/j.nantod.2020.101027
122. Liu B, Liu J. Surface modification of nanozymes. *Nano Res.* 2017;10(4):1125–1148. doi:10.1007/s12274-017-1426-5
123. Liu B, Huang Z, Liu J. Boosting the oxidase mimicking activity of nanoceria by fluoride capping: rivaling protein enzymes and ultrasensitive F⁻ detection. *Nanoscale.* 2016;8(28):13562–13567. doi:10.1039/c6nr02730j
124. Wang S, Chen W, Liu AL, Hong L, Deng HH, Lin XH. Comparison of the peroxidase-like activity of unmodified, amino-modified, and citrate-capped gold nanoparticles. *Chemphyschem.* 2012;13(5):1199–1204. doi:10.1002/cphc.201100906
125. Zhang XQ, Gong SW, Zhang Y, Yang T, Wang CY, Gu N. Prussian blue modified iron oxide magnetic nanoparticles and their high peroxidase-like activity. *J Mater Chem.* 2010;20(24). doi:10.1039/c0jm00174k
126. Liu X, Wei W, Yuan Q, et al. Apoferritin-CeO₂ nano-truffle that has excellent artificial redox enzyme activity. *Chem Commun (Camb).* 2012;48(26):3155–3157. doi:10.1039/c1cc15815e
127. Asati A, Santra S, Kaittanis C, Nath S, Perez JM. Oxidase-like activity of polymer-coated cerium oxide nanoparticles. *Angew Chem Int Ed Engl.* 2009;48(13):2308–2312. doi:10.1002/anie.200805279
128. Pautler R, Kelly EY, Huang PJ, Cao J, Liu B, Liu J. Attaching DNA to nanoceria: regulating oxidase activity and fluorescence quenching. *ACS Appl Mater Interfaces.* 2013;5(15):6820–6825. doi:10.1021/am401886g

129. Fan K, Wang H, Xi J, et al. Optimization of Fe₃O₄ nanozyme activity via single amino acid modification mimicking an enzyme active site. *Chem Commun (Camb)*. 2017;53(2):424–427. doi:10.1039/C6CC08542C
130. Sun Y, Zhao C, Gao N, Ren J, Qu X. Stereoselective nanozyme based on ceria nanoparticles engineered with amino acids. *Chem Eur J*. 2017;23(72):18146–18150. doi:10.1002/chem.201704579
131. Zhang R, Zhou Y, Yan X, Fan K. Advances in chiral nanozymes: a review. *Mikrochim Acta*. 2019;186(12):782. doi:10.1007/s00604-019-3922-7
132. Chen JLY, Pezzato C, Scrimin P, Prins LJ. Chiral nanozymes—gold nanoparticle-based transphosphorylation catalysts capable of enantiomeric discrimination. *Chem Eur J*. 2016;22(21):7028–7032. doi:10.1002/chem.201600853
133. Zhan P, Wang ZG, Li N, Ding B. Engineering gold nanoparticles with DNA ligands for selective catalytic oxidation of chiral substrates. *ACS Catal*. 2015;5(3):1489–1498. doi:10.1021/cs5015805
134. Wang H, Yu D, Fang J, et al. Phenol-like group functionalized graphene quantum dots structurally mimicking natural antioxidants for highly efficient acute kidney injury treatment. *Chem Sci*. 2020;11(47):12721–12730. doi:10.1039/d0sc03246h
135. Sun X, Guo S, Chung CS, Zhu W, Sun S. A sensitive H₂O₂ assay based on dumbbell-like PtPd-Fe₃O₄ nanoparticles. *Adv Mater*. 2013;25(1):132–136. doi:10.1002/adma.201203218
136. Liu M, Li Z, Li Y, Chen J, Yuan Q. Self-assembled nanozyme complexes with enhanced cascade activity and high stability for colorimetric detection of glucose. *Chin Chem Lett*. 2019;30(5):1009–1012. doi:10.1016/j.ccllet.2018.12.021
137. Wang Z, Dong K, Liu Z, et al. Activation of biologically relevant levels of reactive oxygen species by Au/g-C₃N₄ hybrid nanozyme for bacteria killing and wound disinfection. *Biomaterials*. 2017;113:145–157. doi:10.1016/j.biomaterials.2016.10.041
138. Zhang L, Pan J, Long Y, et al. CeO₂-encapsulated hollow Ag-Au nanocage hybrid nanostructures as high-performance catalysts for cascade reactions. *Small*. 2019;15(43):e1903182. doi:10.1002/sml.201903182
139. Zhang S, Li H, Wang Z, et al. A strongly coupled Au/Fe₃O₄/GO hybrid material with enhanced nanozyme activity for highly sensitive colorimetric detection, and rapid and efficient removal of Hg²⁺ in aqueous solutions. *Nanoscale*. 2015;7(18):8495–8502. doi:10.1039/c5nr00527b
140. Huang Y, Liu Z, Liu C, et al. Self-assembly of multi-nanozymes to mimic an intracellular antioxidant defense system. *Angew Chem Int Ed Engl*. 2016;128(23):6758–6762. doi:10.1002/ange.201600868
141. He L, Huang G, Liu H, Sang C, Liu X, Chen T. Highly bioactive zeolitic imidazolate framework-8-capped nanotherapeutics for efficient reversal of reperfusion-induced injury in ischemic stroke. *Sci Adv*. 2020;6(12):eaay9751. doi:10.1126/sciadv.aay9751
142. Zhang K, Kaufman RJ. From endoplasmic-reticulum stress to the inflammatory response. *Nature*. 2008;454(7203):455–462. doi:10.1038/nature07203
143. Guo H, Callaway JB, Ting JP. Inflammasomes: mechanism of action, role in disease, and therapeutics. *Nat Med*. 2015;21(7):677–687. doi:10.1038/nm.3893
144. Ferrucci L, Fabbri E. Inflammageing: chronic inflammation in ageing, cardiovascular disease, and frailty. *Nat Rev Cardiol*. 2018;15(9):505–522. doi:10.1038/s41569-018-0064-2
145. Simmonds N, Rampton D. Inflammatory bowel disease—a radical view. *Gut*. 1993;34(7):865. doi:10.1136/gut.34.7.865
146. Qin Z, Li Y, Gu N. Progress in applications of Prussian blue nanoparticles in biomedicine. *Adv Healthc Mater*. 2018;7(20):1800347. doi:10.1002/adhm.201800347
147. Qiu H, Gong H, Bao Y, Jiang H, Tong W. Reactive oxygen species-scavenging hollow MnO₂ nanozymes as carriers to deliver budesonide for synergistic inflammatory bowel disease therapy. *Biomater Sci*. 2022;10(2):457–466. doi:10.1039/d1bm01525g
148. Zhong G, Yang X, Jiang X, et al. Dopamine-melanin nanoparticles scavenge reactive oxygen and nitrogen species and activate autophagy for osteoarthritis therapy. *Nanoscale*. 2019;11(24):11605–11616. doi:10.1039/c9nr03060c
149. Kumar S, Adjei IM, Brown SB, Liseth O, Sharma B. Manganese dioxide nanoparticles protect cartilage from inflammation-induced oxidative stress. *Biomaterials*. 2019;224:119467. doi:10.1016/j.biomaterials.2019.119467
150. Bao X, Zhao J, Sun J, Hu M, Yang X. Polydopamine nanoparticles as efficient scavengers for reactive oxygen species in periodontal disease. *ACS Nano*. 2018;12(9):8882–8892. doi:10.1021/acsnano.8b04022
151. Zerna C, Thomalla G, Campbell BC, Rha J-H, Hill MD. Current practice and future directions in the diagnosis and acute treatment of ischaemic stroke. *The Lancet*. 2018;392(10154):1247–1256. doi:10.1016/S0140-6736(18)31874-9
152. Rzigalinski BA, Meehan K, Davis RM, Xu Y, Miles WC, Cohen CA. Radical nanomedicine. *Nanomedicine*. 2006;1(4):399–412. doi:10.2217/17435889.1.4.399
153. Liu Y, Ai K, Ji X, et al. Comprehensive insights into the multi-antioxidative mechanisms of melanin nanoparticles and their application to protect brain from injury in ischemic stroke. *J Am Chem Soc*. 2017;139(2):856–862. doi:10.1021/jacs.6b11013
154. Huang G, Zang J, He L, et al. Bioactive nanoenzyme reverses oxidative damage and endoplasmic reticulum stress in neurons under ischemic stroke. *ACS Nano*. 2021. doi:10.1021/acsnano.1c07205
155. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science*. 2002;297(5580):353–356. doi:10.1126/science.1072994
156. Polanco JC, Li C, Bodea LG, Martinez-Marmol R, Meunier FA, Götz J. Amyloid-β and tau complexity—towards improved biomarkers and targeted therapies. *Nat Rev Neurol*. 2018;14(1):22–39. doi:10.1038/nrneurol.2017.162
157. Cheignon C, Tomas M, Bonnefont-Rousselot D, Faller P, Hureau C, Collin F. Oxidative stress and the amyloid beta peptide in Alzheimer's disease. *Redox Biol*. 2018;14:450–464. doi:10.1016/j.redox.2017.10.014
158. Savelieff MG, Nam G, Kang J, Lee HJ, Lee M, Lim MH. Development of multifunctional molecules as potential therapeutic candidates for Alzheimer's disease, Parkinson's disease, and amyotrophic lateral sclerosis in the last decade. *Chem Rev*. 2019;119(2):1221–1322. doi:10.1021/acs.chemrev.8b00138
159. Jenner P. Oxidative stress in Parkinson's disease. *Ann Neurol*. 2003;53(S3):S26–S38. doi:10.1002/ana.10483
160. Spillantini MG, Schmidt ML, Lee VM-Y, Trojanowski JQ, Jakes R, Goedert M. α-Synuclein in Lewy bodies. *Nature*. 1997;388(6645):839–840. doi:10.1038/42166
161. Ma X, Hao J, Wu J, Li Y, Cai X, Zheng Y. Prussian blue nanozyme as a pyroptosis inhibitor alleviates neurodegeneration. *Adv Mater*. 2022;34(15):2106723. doi:10.1002/adma.202106723

162. Ni D, Jiang D, Kutylreff CJ, et al. Molybdenum-based nanoclusters act as antioxidants and ameliorate acute kidney injury in mice. *Nat Commun.* 2018;9(1):5421. doi:10.1038/s41467-018-07890-8
163. Zhang J, Pei W, Xu Q, Jiang H, Chen J. Desolvation-induced formation of recombinant camel serum albumin-based nanocomposite for glutathione colorimetric determination. *Sens Actuators B Chem.* 2022;1:357.
164. Zhang J, Xu Q, Pei W, et al. Self-assembled recombinant camel serum albumin nanoparticles-encapsulated hemin with peroxidase-like activity for colorimetric detection of hydrogen peroxide and glucose. *Int J Biol Macromol.* 2021;193:2103–2112. doi:10.1016/j.ijbiomac.2021.11.042
165. Ryan JJ, Bateman HR, Stover A, et al. Fullerene nanomaterials inhibit the allergic response. *J Immunol.* 2007;179(1):665–672. doi:10.4049/jimmunol.179.1.665
166. Jacevic V, Djordjevic A, Srdjenovic B, et al. Fullerenol nanoparticles prevents doxorubicin-induced acute hepatotoxicity in rats. *Exp Mol Pathol.* 2017;102(2):360–369. doi:10.1016/j.yexmp.2017.03.005
167. Zhao S, Duan H, Yang Y, Yan X, Fan K. Fenozyme protects the integrity of the blood-brain barrier against experimental cerebral malaria. *Nano Lett.* 2019;19(12):8887–8895. doi:10.1021/acs.nanolett.9b03774
168. Kalashnikova I, Chung SJ, Nafuijjaman M, et al. Ceria-based nanotheranostic agent for rheumatoid arthritis. *Theranostics.* 2020;10(26):11863–11880. doi:10.7150/thno.49069
169. Bailey ZS, Nilson E, Bates JA, et al. Cerium oxide nanoparticles improve outcome after in vitro and in vivo mild traumatic brain injury. *J Neurotrauma.* 2020;37(12):1452–1462. doi:10.1089/neu.2016.4644
170. Cheng Y, Cheng C, Yao J, et al. Mn₃O₄ nanozyme for inflammatory bowel disease therapy. *Adv Ther.* 2021;4(9). doi:10.1002/adtp.202100081
171. Zhang X, Zhang S, Yang Z, Wang Z, Tian X, Zhou R. Self-cascade MoS₂ nanozymes for efficient intracellular antioxidation and hepatic fibrosis therapy. *Nanoscale.* 2021;13(29):12613–12622. doi:10.1039/d1nr02366g
172. Onizawa S, Aoshiba K, Kajita M, Miyamoto Y, Nagai A. Platinum nanoparticle antioxidants inhibit pulmonary inflammation in mice exposed to cigarette smoke. *Pulm Pharmacol Ther.* 2009;22(4):340–349. doi:10.1016/j.pupt.2008.12.015
173. Katsumi H, Fukui K, Sato K, et al. Pharmacokinetics and preventive effects of platinum nanoparticles as reactive oxygen species scavengers on hepatic ischemia/reperfusion injury in mice. *Metallomics.* 2014;6(5):1050–1056. doi:10.1039/c4mt00018h
174. Zhao H, Zeng Z, Liu L, et al. Polydopamine nanoparticles for the treatment of acute inflammation-induced injury. *Nanoscale.* 2018;10(15):6981–6991. doi:10.1039/c8nr00838h
175. Liu Y, Cheng Y, Zhang H, et al. Integrated cascade nanozyme catalyzes in vivo ROS scavenging for anti-inflammatory therapy. *Sci Adv.* 2020;6(29):eabb2695. doi:10.1126/sciadv.abb2695
176. Huang Y, Ren J, Qu X. Nanozymes: classification, catalytic mechanisms, activity regulation, and applications. *Chem Rev.* 2019;119(6):4357–4412. doi:10.1021/acs.chemrev.8b00672

Journal of Inflammation Research

Dovepress

Publish your work in this journal

The Journal of Inflammation Research is an international, peer-reviewed open-access journal that welcomes laboratory and clinical findings on the molecular basis, cell biology and pharmacology of inflammation including original research, reviews, symposium reports, hypothesis formation and commentaries on: acute/chronic inflammation; mediators of inflammation; cellular processes; molecular mechanisms; pharmacology and novel anti-inflammatory drugs; clinical conditions involving inflammation. The manuscript management system is completely online and includes a very quick and fair peer-review system. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/journal-of-inflammation-research-journal>