



HOT TOPICS



Single-cell transcriptional profiling in brain reward structures

Jeremy J. Day¹✉ and Keri Martinowich^{2,3,4,5}✉

© The Author(s), under exclusive licence to American College of Neuropsychopharmacology 2022

Neuropsychopharmacology (2023) 48:243–244; <https://doi.org/10.1038/s41386-022-01394-2>

Brain reward circuits are frequently disrupted in neuropsychiatric and substance use disorders. For example, substance use disorders are associated with prolonged molecular changes in reward-related regions including the ventral tegmental area (VTA), nucleus accumbens (NAc), prefrontal cortex, hippocampus, and amygdala. However, cellular heterogeneity has impeded progress in understanding the molecular mechanisms contributing to disease. While previous studies identified unique functions for cell types in these regions, they lacked comprehensive information on transcriptional diversity.

To overcome these challenges, recent studies used single-cell and single-nucleus RNA-sequencing (sc/snRNA-seq) approaches to generate molecular profiles of cell types in reward-related regions of the rodent, non-human primate, and human brain [1–6]. These data enabled the identification of cell types that (1) are activated by exposure to motivational cues, including psychoactive drugs; (2) harbor enrichment for genetic risk associated with substance use and neuropsychiatric disorders; and (3) converge/diverge across species. Such analyses can identify and prioritize targets at the level of specific genes and gene programs within cell populations that contribute to the reward function.

For example, while roles for VTA dopamine neurons are well studied, GABAergic and glutamatergic neurons in the VTA also contribute to reward signaling, and parallel lines of evidence suggest that some VTA neurons may synthesize and release multiple neurotransmitters. Using snRNA-seq we confirmed the identity of combinatorial neurons in rat VTA, and elucidated novel marker genes for these populations [5]. Moreover, we demonstrated cell-type-specific enrichment for gene sets associated with risk for brain disorders and phenotypes related to substance use [5]. Similarly, to understand cell-specific responses to drugs with addictive potential that alter dopamine concentrations in the NAc, we generated a molecular atlas of the rat NAc at single-cell resolution following cocaine experience. Dopaminergic medium spiny neurons (MSNs), the principal NAc cell type, are functionally segregated into D1 and D2 receptor-expressing subtypes. Cocaine elevated activity-regulated gene expression selectively in D1-MSNs, an effect that was driven by a relatively small cluster of neurons [1]. In contrast, another scRNA-seq study examining morphine response in mouse NAc identified robust transcriptional activation of glia, highlighting the utility of these approaches in identifying substance-specific molecular changes [6].

Interpreting clinical relevance and extending findings from animal models to treatments requires understanding how cell

types that contribute to the reward function differ across species. Toward this goal, we used snRNA-seq to generate a molecular taxonomy of cells across key nodes of the human brain reward circuitry (NAc, prefrontal cortex, hippocampus, and amygdala) [2]. In NAc, we identified discrete subpopulations of D1 and D2-MSNs to which we mapped cell-type-specific enrichment for genetic risk associated with both psychiatric disease and substance use. While many cell populations in the NAc were conserved, we also identified transcriptional differences in MSN subpopulations between rats and humans, indicating the presence of unique molecular features in analogous populations, or the existence of species-specific subclasses [2]. Together, these results advance our understanding of molecular mechanisms of neuropsychiatric disease and substance use disorders by revealing not only which individual cell ensembles contribute to reward processing, but also how genetic variation may influence these effects in the human brain.

REFERENCES

1. Savell KE, Tuscher JJ, Zipperly ME, Duke CG, Phillips RA, Bauman AJ, et al. A dopamine-induced gene expression signature regulates neuronal function and cocaine response. *Sci Adv.* 2020;6:eaba4221.
2. Tran MN, Maynard KR, Spangler A, Huuki LA, Montgomery KD, Sadashivaiah V, et al. Single-nucleus transcriptome analysis reveals cell-type-specific molecular signatures across reward circuitry in the human brain. *Neuron.* 2021;109:3088–103. e5
3. Chen R, Blosser TR, Djekidel MN, Hao J, Bhattacharjee A, Chen W, et al. Decoding molecular and cellular heterogeneity of mouse nucleus accumbens. *Nat Neurosci.* 2021;24:1757–71.
4. He J, Kleyman M, Chen J, Alikaya A, Rothenhoefer KM, Ozturk BE, et al. Transcriptional and anatomical diversity of medium spiny neurons in the primate striatum. *Curr Biol.* 2021;31:5473–86.e6. <https://doi.org/10.1016/j.cub.2021.10.015>.
5. Phillips RA, Tuscher JJ, Black SL, Andraka E, Fitzgerald ND, Ivanov L, et al. An atlas of transcriptionally defined cell populations in the rat ventral tegmental area. *Cell Rep.* 2022;39:110616.
6. Avey D, Sankararaman S, Yim AKY, Barve R, Milbrandt J, Mitra RD. Single-cell RNA-seq uncovers a robust transcriptional response to morphine by glia. *Cell Rep.* 2018;24:3619–29. e4

ACKNOWLEDGEMENTS

The authors thank members of the Day and Martinowich labs for suggestions.

AUTHOR CONTRIBUTIONS

JJD and KM contributed equally to the conceptualization and writing of this manuscript. Both authors approved the final version of the paper.

¹Department of Neurobiology, University of Alabama at Birmingham, Birmingham, AL 35294, USA. ²Lieber Institute for Brain Development, Johns Hopkins Medical Campus, Baltimore, MD 21205, USA. ³Department of Psychiatry and Behavioral Sciences, Johns Hopkins University School of Medicine, Baltimore, MD 21205, USA. ⁴Department of Neuroscience, Johns Hopkins University School of Medicine, Baltimore, MD 21205, USA. ⁵The Kavli Neuroscience Discovery Institute, Johns Hopkins University, Baltimore, MD 21205, USA. ✉email: jjday@uab.edu; keri.martinowich@libd.org

FUNDING

This work was supported by funding from the Lieber Institute for Brain Development as well as U01MH122849 and R01DA053581 to KM, and DP1DA039650, R01DA053743, R01MH114990, and R21DA048348 to JJD.

COMPETING INTERESTS

KM is the Social Media Editor for *Neuropsychopharmacology*.

ADDITIONAL INFORMATION

Correspondence and requests for materials should be addressed to Jeremy J. Day or Keri Martinowich.

Reprints and permission information is available at <http://www.nature.com/reprints>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.