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High-Throughput PBTK models for *In Vitro* to *In Vivo* Extrapolation

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Abstract

Introduction: Toxicity data are unavailable for many thousands of chemicals in commerce and the environment. Therefore, risk assessors need to rapidly screen these chemicals for potential risk to public health. High-throughput screening (HTS) for *in vitro* bioactivity, when used with high-throughput toxicokinetic (HTTK) data and models, allows characterization of these thousands of chemicals.

Areas covered: This review covers generic physiologically-based toxicokinetic (PBTK) models and high-throughput PBTK modeling for *in vitro-in vivo* extrapolation (IVIVE) of HTS data. We focus on “httk”, a public, open-source set of computational modeling tools and *in vitro* toxicokinetic (TK) data.

Expert opinion: HTTK benefits chemical risk assessors with its ability to support rapid chemical screening/prioritization, perform IVIVE, and provide provisional TK modeling for large numbers of chemicals using only limited chemical-specific data. Although generic TK model design can increase prediction uncertainty, these models provide offsetting benefits by increasing model implementation accuracy. Also, public distribution of the models and data enhances reproducibility. For the httk package, the modular and open-source design can enable the tool to be used and continuously improved by a broad user community in support of the critical need for high-throughput chemical prioritization and rapid dose estimation to facilitate rapid hazard assessments.

Keywords

Generic Physiologically Based Toxicokinetic Models; High-Throughput; *In Vitro* to *In Vivo* Extrapolation; Modeling Software Tools; Open Source Tools; Physiologically-based toxicokinetics; Toxicokinetics

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Introduction

There are thousands of chemicals present in our environment, many of which are routinely detected in environmental and human blood samples [1,2]. There is a need to rapidly assess potential risk to human health for these chemicals, but the necessary toxicity data are unavailable for most [3–5]. *In vivo* toxicity testing of such a large number of chemicals is not feasible [6,7]. As an alternative, *in vitro* high-throughput screening (HTS) assays are performed, in which a large number of chemicals may be examined by a battery of *in vitro* tests for biological activity across a variety of different biologically-based endpoints [8–10]. HTS typically characterizes the concentration at which a chemical causes bioactivity *in vitro*. To use *in vitro* HTS to assess *in vivo* potential risk to humans, the *in vitro* bioactive concentration, representing an internal dose or threshold, must be extrapolated to an equivalent *in vivo* external dose or exposure (*in vitro-in vivo* extrapolation, or IVIVE, Figure 1) [11]. This equivalent external dose can then be compared to estimates of exposure to assess potential risk [12].

A model of toxicokinetics (TK) quantitatively describes the body's absorption, distribution, metabolism, and excretion (ADME) of a chemical or substance [13]. (Different terms for this concept are preferred in different fields, including “toxicokinetics”, “pharmacokinetics”, “biokinetics”, and simply “kinetics”; in this review, we use “TK”). TK models are used to estimate internal dose following an external exposure. For IVIVE of toxicological bioactivity data, an administered equivalent dose (AED) is determined using reverse dosimetry by solving the TK model in reverse, deriving the external dose of chemical (that is TK model input) that produces a specified internal concentration (that is TK model output) (Figures 1 and 2) [14]. While reverse dosimetry has been applied one chemical at a time [15–17], applying reverse dosimetry to HTS data requires rapid approaches amenable to thousands of chemicals. Therefore, a *high-throughput toxicokinetics* (HTTK) approach is needed [18]. HTTK methods use generic TK models that can be parameterized rapidly for large numbers of chemicals, using *in vitro* measurements and/or *in silico* predictions of chemical-specific TK properties like metabolism [7]. Thus, HTTK relies on TK IVIVE to perform toxicological IVIVE of HTS data.

While IVIVE broadly includes any use of *in vitro* data to predict phenomena *in vivo* it is useful to distinguish between TK IVIVE (that is, the use of *in vitro* data to predict ADME) and toxicodynamic (TD) IVIVE, which includes the use of *in vitro* data to predict toxic effects *in vivo* (Figure 1). Both TK IVIVE and *in vitro* disposition modeling (for example, the Armitage model [19]) are useful for enhancing TD IVIVE [20], but the specific *in vitro* measurements and IVIVE methods used to inform TK IVIVE may be very different from those for TD IVIVE. For example, TK IVIVE often uses the clearance of hepatocyte suspensions and the binding of plasma proteins along with a TK model to predict ADME [21,22]. Those ADME predictions might in turn be used to scale concentrations for TD IVIVE in which the *in vitro* bioactivity data are statistically correlated with *in vivo* toxic endpoints observed in animals [23]. Both the data and the methods (for example, mechanistic ADME vs. statistical modeling) can vary significantly between TK IVIVE and TD IVIVE.

HTTK methods have been used by the pharmaceutical industry to determine the range of efficacious doses, to prospectively evaluate success of planned clinical trials, and to minimize side effects and drug-drug interactions. The TK IVIVE methods initially developed for pharmaceutical HTTK have also been applied to environmental and industrial chemicals, to rapidly assess potential human health risks for chemicals which (rightly) cannot ethically be tested in humans, but to which humans are nonetheless exposed. Because of the demonstrated utility of HTTK methods for environmental and industrial chemicals, the U.S. Environmental Protection Agency (U.S. EPA) provides an implementation of HTTK methods through the publicly-available software package called “httk” (<https://CRAN.R-project.org/package=httk>). Here we distinguish the scientific methodology of HTTK from the R package httk via capitalization. The httk package was developed to achieve two main goals: IVIVE of *in vitro* HTS bioactive concentrations to predict human dose context [6,28], and providing open source data and models for evaluations and applications by the broader scientific community [31]. The use of HTTK methodology and the httk package has been recently advocated by the Health Canada, as they envision using such an approach for future screening level assessments under the Canadian Environmental Protection Act [32]. This review provides a description of HTTK models, with a focus on the httk package, its capabilities, and applications. However, comparison between other HTTK implementations is also provided.

1. Generic Physiologically-Based Toxicokinetic Models

TK models describe the ADME of a chemical by representing the body as one or more connected compartments among which the substance can enter (absorption), move (distribution), and leave (metabolism and elimination) [33,34]. Mathematically, a TK model describes the time-dependent amount of a substance in each compartment using a set of mass-balance ordinary differential equations [35]. The parameters of the model describe the size of each compartment and the rates at which the substance moves into, between, and out of compartments. The structure of the model defines the number of compartments and how they are connected.

Physiologically-based toxicokinetic (PBTK) models are designed such that the structure and parameters of the model have physiologically meaningful interpretations, as opposed to “empirical” non-physiological compartmental TK models [35]. Compartments in PBTK models typically represent individual organs or tissues and/or groups of physiologically similar organs or tissues [36–38]. Instead of simply applying uncertainty factors or extrapolating from non-physiological TK models to assess potential risk, PBTK models can estimate internal dose metrics (that is the biologically effective dose) by predicting internal dose of target organs to replace the externally administered dose (or exposure), as the observed effects are expected to be more directly related to the target tissue dose than the administered dose [37,39].

PBTK model parameters are based on anatomical, physiological, and biochemical properties [37]. Parameters may represent either physiological quantities (for example, tissue blood flow, tissue volume) or chemical-specific quantities describing the interactions between the chemical and the body (for example, tissue:plasma equilibrium partition coefficients)

[36,37]. Model parameters are estimated using data obtained from known exposure scenarios. The models can be used to make predictions of hypothetical exposure scenarios (exposure conditions of interest) and simulate the chemical TK in potentially untestable situations. These extrapolations can be not only from *in vitro* to *in vivo* conditions [11,40], but also across exposure routes [41,42]; between species [41–43]; between chemicals [44,45]; across populations [46–49]; and across life-stages [50,51]. The model parameters are modified to represent differences among the extrapolation situations, with no need to collect new chemical-specific or *in vivo* data [39].

PBTK models are traditionally developed for individual chemicals, with both the parameters and structure of the models tailored to make the most accurate predictions possible for each substance [52]. Different physiological processes may be included or omitted depending on their importance to the chemical under consideration [37], for example saturable resorption of ethylene glycol by the kidney [53] or extrahepatic metabolism of bisphenol-A and naloxone [54,55]. These “bespoke” (chemical-specific or study-specific) PBTK models require large amounts of information regarding the behavior and partitioning of a chemical in the body. Because of the challenges posed by the specificity of these PBTK models, these type of detailed models are more likely to suffer from implementation and documentation errors [56]. McLanahan, El-Masri [57] observed that peer-reviewed publications alone are often insufficient to allow proper verification and reproducibility of PBTK models – for example, model code and parameters are often outdated, even when provided as supplemental material, and scripts for generating figures and other analyses often do not meet the threshold for inclusion in supplemental material. Reproducibility of PBTK models is highly desirable [58], but currently deficient due to the lack of adherence to published criteria or standards for evaluating these models [59–61]. For this reason, many tools and approaches have been developed to better document PBTK models [62–66], or to better identify the underlying chemical space that PBTK models have traditionally captured, and where there may be gaps [67]. Moreover, because of the uniqueness of these bespoke PBTK models, they require substantial chemical-specific TK data. Among non-pharmaceutical chemicals, comprehensive TK data only exist for relatively few, well-studied chemicals such as dioxin, lead, and trichloroethylene. The level of detailed data needed to build these models is not feasible in the context of HTTK as it can take years to develop for one of the thousands of chemicals potentially of interest.

A structurally-simpler approach to developing TK models is the empirical non-physiological compartmental approach that is not physiologically based, but is instead a best fit to *in vivo* measurements of the time course of body concentrations after single or repeated dosing of a chemical [13]. If concentrations appear to decrease with a single time constant, then an empirical model with a single compartment is developed; if concentrations appear to decrease with two time constants (for example, an initial rapid decrease and a second slower decrease), then an empirical model with two compartments is developed; and so forth. These empirical TK model structures are considered generic, since they can be parameterized to describe many different chemicals, and they do not attempt to describe chemical-specific ADME mechanisms [68]. However, parameterizing empirical models requires collecting species-specific *in vivo* measurements of concentration vs. dose and time, which is not feasible for large numbers of chemicals in an HTTK context. Moreover, because the

parameters of empirical models are not physiologically based, empirical TK models do not allow the same degree of extrapolation (such as inter-species and route-to-route) as PBTK models, and therefore are less than ideal for many aspects of assessment for non-therapeutic chemicals where human studies are unlikely [39].

A third approach, “generic” PBTK models, uses a common structure across chemicals, bridging the gap between empirical and bespoke models. That is, the same physiological compartments and the same physiological processes are represented for all chemicals as opposed to customizing the physiological processes to those relevant for a specific chemical. Only the parameters representing chemical-body interaction differ between chemicals. These chemical-specific parameters may be derived from *in vitro* measurements [21,27–30], meaning that generic PBTK models are amenable to rapid parameterization for many chemicals, and therefore useful for HTTK applications. In fact, HTTK can be described as the combination of generic PBTK models with *in vitro* TK data. The consistent model structure helps improve reporting accuracy, fidelity of implementation, reproducibility, and statistical evaluation of generic PBTK models compared to bespoke PBTK models [62,69–73]. Several modeling software tools have been developed to facilitate the use of generic PBTK modeling [24,31,74–76]. Generic PBTK models vary with respect to the amount of chemical-specific data required, with more sophisticated models having “higher” data needs, such as enzyme-specific metabolism rates to address genetic polymorphisms, while those with “lower” data needs might only use a few parameters to characterize chemicals. An overview of some key generic PBTK modeling tools, including the htk package, are described in Table 1.

2. High-Throughput PBTK Modeling for IVIVE: The htk package

The htk package provides open source data and models with transparent documentation of the model design, the input parameters, and the differential equations used to calculate tissue concentrations. Since HTTK is the combination of *in vitro* data and generic TK modeling, the htk package [31] contains tables of chemical specific data (>1000 for humans and >200 for rats) as well as a suite of generic TK models that may be parameterized with those data. Released as an R package, the numerical modeling modules of htk are written in the C programming language for efficiency (that is, speed) of compiled code. The form of an R package was chosen to make htk open-source, transparent, freely available, and as platform-independent as possible. R and an integrated development environment (IDE) called RStudio are free software for statistical computing available for Windows, Mac, and Unix/Linux operating systems. Additional R packages are available to solve the required initial value problems (IVP) of ordinary differential equations (ODE). Using a command-line interface in R, the typical workflow of htk is shown in Figure 3. Rudimentary instructions for using htk is provided in Table 2 and further described in section 3.7.

While htk is designed to be accessed through the R command line and scripts of commands, two graphical user interface (GUI) tools for htk are currently available with PLETHEM [77] and Web-ICE [78]. Additional custom htk web interfaces can be developed with R-Shiny (<https://shiny.rstudio.com/>). The htk R package can also be integrated with the Konstanz Information Miner (KNIME) [79]: “R nodes” for input/output calls and third party reporting

(<https://www.knime.com/nodeguide/scripting/r>) allow the highly customizable KNIME user interface to integrate R with many other tools. Eventually, httpk-specific web-service apps might facilitate broader usability.

3.1. Generic TK Models in httpk—The current version of the httpk package, version 2.0.3, has five options for TK models: a one-compartment empirical model [13], a generic three-compartment PBTK model [24], three-compartment steady-state model [18,21], a more elaborate generic PBTK model for intravenous and oral exposure [31], and a generic gas inhalation PBTK model [80]. The generic three-compartment PBTK model (contains a systemic blood compartment with separate tissue compartments for the liver and gut) is the condensed form of the generic compartmental PBTK model (contains separate tissue compartments for the gut, liver, lungs, arteries, veins, and kidneys) [31]. The three-compartment steady-state model is the simplest model, which describes the steady-state concentration in the liver of the three-compartment PBTK model without partitioning (contain only plasma without separate compartments for blood and tissue) [31]. The user selects which model to use, and can use the selected model to perform forward dosimetry (using functions such as *solve_gas_pbtk()*) or reverse dosimetry (using functions such as *calc_mc_oral_equiv()*).

For reverse dosimetry, as in HT risk screening, either the generic compartmental PBTK model for oral exposure or the generic compartmental gas inhalation PBTK model for inhalation exposure are typically recommended, depending upon the volatility of the compound and the scenario. For instance, the oral exposure model would be suitable for the calculation of AEDs for a chemical in drinking water. The one-compartment empirical model is included mainly for use when *in vivo* TK data are available for model evaluation: it allows comparison of model predictions with *in vivo* experiments [22,81], and uncertainty quantification for regulatory decision making [56]. The three-compartment steady-state model is applicable to the largest number of chemicals, specifically those which are missing information needed to predict tissue partitioning in the other models [31].

The structure of the httpk generic compartmental PBTK models [31] — for example, the generic inhalation PBTK model in Figure 4 — includes compartments corresponding to blood and other tissues that are homogenous (well-mixed) and can be described by a volume and a single chemical concentration. Some tissues (for example arterial blood) are simple compartments, while others (for example kidney) are compound compartments consisting of separate blood and tissue sections. Some specific tissues (lung, kidney, gut, and liver) are modeled explicitly, while remaining tissues of body that are not from the organ of interest (for example fat, brain, bones) are lumped into the “Rest of Body” compartment for model simplifications. Following oral exposure, chemicals are absorbed from gut lumen, and chemicals are transformed through hepatic metabolism or excretion by passive renal clearance. The generic compartmental gas inhalation PBTK model [80] added a new component to allow modeling of inhalation exposures at a specified air concentration Figure 4. The structure of the inhalation model was developed from two previously published PBTK models from Jongeneelen and Berge [76] and Clewell III, Gentry [42], where chemicals are absorbed from an “alveolar space” compartment.

Each TK model can yield either time-course concentrations in one or more compartments (including both peak and time-integrated “area under the curve”/AUC concentrations), or a steady-state concentration in one or more compartments, assuming repeated dosing at a constant level for a long duration. Typically, in the context of reverse dosimetry for HT risk screening, the steady-state concentration is used since analytical steady state solutions to the models can substantially shorten the computation time, and the resulting *in vivo* equivalent dose represents a long-term near-constant exposure. The `httk` package contains functions that automate the calculation of AEDs under the steady-state assumption. This allows the user to simply input the chemical and *in vitro* bioactive concentration, select the TK model, and then automatically obtain the *in vivo* equivalent dose which would produce a body concentration equal to the *in vitro* bioactive concentration. It relies on the linearity of the models to calculate a scaling factor to relate *in vitro* concentrations (μM) with AED. The scaling factor is the inverse of the steady-state plasma concentration (C_{ss}) predicted for a 1 mg/kg/day exposure dose rate (Figure 2):

$$\text{AED} = \frac{[X]}{C_{ss}}$$

where *in vitro* concentration $[X]$ and C_{ss} must be in the same units. Note that it is typical for *in vitro* concentrations to be reported in units of μM and C_{ss} in units of mg/L, in which case one must be converted to the other using:

$$\mu\text{M} = 1000 \frac{1 \text{ mol mg}}{\text{MW L}}$$

where MW is the molecular weight of the compound in units of g/mole.

If non-steady-state concentrations are needed (for example, to perform reverse dosimetry for an acute or subchronic exposure period), `httk` can automatically calculate peak concentration, average concentration, and AUC in a specified compartment with the functions `calc_tkstats()` and the Monte Carlo version `calc_mc_tk()`. As with C_{ss} , the user can again employ the linearity of the models in `httk` to determine AED by dividing the *in vitro* bioactive concentration with the relevant concentration summary statistic (peak, average, or AUC) for a dose of 1 mg/kg/day. The full time course of concentration in each compartment can be calculated using the function `solve_model()`.

The generic TK models have parameters that can be divided into two main categories: chemical-specific parameters and physiological parameters. Chemical-specific parameters describe kinetics-related quantities that are different for different chemicals (for example, hepatic clearance rate, plasma protein binding, tissue partitioning, oral absorption). Physiological parameters describe kinetics-related quantities that depend on the anatomical and physiological characteristics of each person or animal and are not chemical-dependent (for example, body weight, organ and tissue volumes, blood flow rates, hepatocellularity). Below, we describe the details of these generic TK model parameters.

3.2. Chemical-specific Parameters: Experimental Data Needs/Inputs—Four chemical-specific parameters are used in htkk:

1. intrinsic hepatic clearance (that is, disappearance of compound when incubated with primary hepatocytes)
2. fraction of compound unbound in plasma
3. Caco-2 membrane permeability, and
4. relative concentration in blood and plasma (that is, ratio of the blood concentration of a chemical to the plasma concentration).

Constant $R_{\text{blood}}/2_{\text{plasma}}$ (ratio of the blood concentration of a chemical to the plasma concentration) is used throughout the body, predicted using hematocrit and the predicted partitioning between red blood cells and plasma when *in vivo* values are unavailable. These chemical-specific TK parameters for the htkk models are based on *in vitro* measurements for a total of 1043 chemicals (see, for example, Rotroff, Wetmore [27], Wetmore, Wambaugh [29], Wetmore, Wambaugh [12], Honda, Pearce [20]). Of the four *in vitro* TK parameters mentioned above, the htkk package primarily uses two: 1) intrinsic hepatic clearance [82] and 2) fraction unbound in plasma [83]. Currently only hepatic clearance estimates from primary hepatocyte suspensions [82] are used by htkk, though this clearance might also be estimated by other means including microsomes and whole-body *in vivo* data. Protein binding in plasma, as characterized by the fraction unbound, is particularly important for predicting partitioning between tissues and renal excretion, and may impact the rate of metabolism [7,84] as only unbound chemical is able to freely partition and distribute into tissues. When data are available, the two remaining parameters, Caco2 permeability [85] (used to model oral absorption) and relative blood/plasma concentrations, can also be used by the htkk package.

The htkk package includes parameters (namely, intrinsic hepatic clearance and fraction unbound in plasma) from *in vitro* measurements with human-specific data for 1016 chemicals and rat-specific data for 212 chemicals. For other chemicals, user-provided model parameters can be entered into htkk based on values obtained from other *in vitro* measurements, or from *in silico* models derived from applications such as OPERA structure-activity Relationship App (OPERA), Molecular Operating Environment (MOE) [86–89], or any cheminformatics tools that can predict molecular descriptors and derive QSAR (i.e. Schrödinger QikProp (New York, USA), Simulations Plus (Lancaster, USA) and Discovery Studio (Polouzane, France)).

Although physiological parameters (such as organ volumes and flows) are included for species beyond humans and rats, htkk contains only very limited species-specific *in vitro* measured TK parameters for mouse and rabbit. While “chemical-specific” *in vitro* measured TK parameters might be considered more like inherent properties of a chemical than purely physiological parameters, properties like hepatic clearance do vary between species [29] and individuals [30] owing to inter-species and inter-individual differences in hepatic enzyme affinity, abundance, and polymorphism. When non-human parameter values are unavailable for a particular chemical, htkk can automatically substitute human values (typically by

setting the argument `default.to.human = TRUE`). Inter-individual variability in intrinsic hepatic clearance rate and fraction unbound in plasma is accounted for in `httk`, assuming a truncated normal distribution with 30% CV [47]. For hepatic clearance rate, a group of ultra-low metabolizers (with mean clearance ten times less than measured) are assumed to make up 5% of the population [47].

3.3. Chemical-Specific Parameters: Tissue:plasma partition coefficients—

Since the chemical-specific tissue:plasma partition coefficients are more difficult to measure *in vitro*, `httk` contains a module to predict them based on physical-chemical properties and published tissue properties. It uses an empirically calibrated version of Schmitt's method [90] that has been expanded with methods from Peyret, Poulin [91] using tissue data: cellular and water fractions of total volume, lipid and protein fractions of cellular volume, lipid fractions of the total lipid volume, the pH of each tissue, and the fractional volume of protein in plasma [31]. It calculates ionization with a default plasma pH of 7.4 [90], and the partition coefficient for the mass and volume of the body unaccounted for by the tissues included in Schmitt [90], which is determined with the averages of the fractional volumes and pH of these tissues, excluding red blood cells [31]. The model predictions are calibrated based on tissue-specific regressions of experimentally observed tissue partition coefficients on the predictions of the model (this can be turned off via “`regression = FALSE`”) [38].

3.4. Physiological parameters and Monte Carlo simulations to represent inter-individual variability—

To support reverse dosimetry for a variety of species and inter-species extrapolation, `httk` contains default physiological parameter values for rat, rabbit, dog, mouse, and human. Here, physiological parameters represent values specific to the species of interest and independent of the chemical being simulated such as cardiac output, tissue volume, and respiratory rates. These default values are intended to represent an “average” or “standard” individual. However, default parameters may not be sufficient for the typical use case of prioritizing environmental chemicals based on potential human health risk in the population, because TK vary between individuals [47]. Each person's body can process a chemical differently owing to inherent variability in organ size, blood flow rates, and enzymatic abundance. Therefore, different people with the same external exposure can have different internal concentrations. The TK models account for this inter-individual variability by varying both the physiological and chemical-specific parameters between individuals. The physiological parameters are varied to account for individual physiological differences, whereas chemical-specific parameters are varied to account for individual differences in the interaction between the body and the chemical.

The `httk` package includes the capability to simulate inter-individual TK variability using a Monte Carlo approach [92,93], which assumes distributions for the TK model parameters that represent population variability. The model is solved repeatedly, typically using an analytical steady-state solution, each time randomly drawing a set of parameter values from these distributions. The resulting set of model outputs approximates the distribution of AEDs among individuals. A lower percentile AED represents a more sensitive individual – a person that can reach the *in vitro* bioactive concentration with a lower exposure. A

higher percentile AED represents a less-sensitive individual – a person that requires a higher exposure to reach the *in vitro* bioactive concentration.

The HHTK-Pop module of htk generates a “virtual population” of individual physiologies through Monte Carlo simulations that incorporate physiologies, incorporating observed correlations (for example, among age, sex, height, and weight) to produce a more realistic distribution of physiologies [47]. The htk package allows the user to specify attributes of the population to be simulated. For example, the user can specify a population for ages 6–11 to determine the range of AEDs for children, or a population of women ages 18–45 to determine the range of AEDs for reproductive-aged women. Since htk was developed primarily for risk prioritization in the US, it uses physiological data for the US population from the Centers for Disease Control and Prevention (CDC) the National Health and Nutrition Examination Survey (NHANES) database: demographics, body measurements, and certain direct measurements of physiology. For physiological parameters that are not measured by NHANES, such as tissue volumes and blood flows, htk uses allometric scaling or regression models from the literature to predict these parameters based on quantities that NHANES does measure (for example age, sex, body weight, height).

3.5. Uncertainty Propagation—Taking advantage of much of the same Monte Carlo simulation approach that is used for population variability, propagation of uncertainty was recently included in htk. This allows for a quantitative assessment of the impact of experimental uncertainty in measuring the chemical-specific parameters [94], which is critical to assess the quality and accuracy of the data. Within htk, most parameters can be assigned distributions with means and standard deviations representing both variability and uncertainty where variability represents a range of observed parameter values while uncertainty represents a range of model-predicted values. Chemical-specific *in vitro* measurements can be assigned distributions reflecting both the constraints of the measurement process (for example, fraction unbound should be between zero and one) and any chemical specific uncertainty estimated reflecting the mass spectrometry signal-to-noise ratio [94]. Particularly of note, by assigning appropriate estimates of uncertainty, these methods allow the predictions of QSARs and experimental data to be used jointly. By using the Monte Carlo method to simulate population variability [47,93] and propagate uncertainty [95,96], an upper 95th percentile steady-state concentration ($C_{ss,95}$) can be calculated for individuals who have higher plasma concentrations from the same exposure in order to obtain a scaling factor for relating *in vitro* concentrations with AED (Figure 2) [94]. Moreover, this allows the high-throughput chemical risk prioritization to rapidly prioritize large numbers of chemicals by comparing distributions of dose with potentially adverse effect and potential exposure, which are estimated by accounting for both uncertainty and variability (Figure 5) [47].

3.6. Chemical-Specific Parameters: *In Vitro* Distribution—In the context of *in vitro* high-throughput toxicity testing, the bioactive concentration is typically reported as the nominal applied concentration [19], and reverse dosimetry is typically applied to estimate an AED for this nominal concentration. However, experimental evidence has indicated the importance of considering *in vitro* chemical disposition when performing IVIVE: a nominal

applied concentration may partition into the different elements of an *in vitro* assay system differently depending on chemical properties, resulting in different free concentrations in culture media and cells that may affect *in vitro* bioactivity [97–100]. There are published models for predicting the freely dissolved cellular/tissue and membrane concentrations to characterize chemical distribution for *in vitro* assays [19,101]. For performing IVIVE with *in vitro* HTS assay data, htk contains a module that implements the Armitage model [19] of chemical disposition within an *in vitro* assay system. Although this is not a TK model, the Armitage model is important to correctly identify the *in vitro* concentration associated with bioactivity, and thereby derive the appropriate *in vivo* equivalent dose [20].

3.7. Getting Started with the R Package htk—The open-source nature of R and detailed documentation of its packages make htk readily accessible to the broader scientific community. This accessibility and the straightforward nature of R mean individuals with minimal coding experience should feel comfortable working with htk. RStudio is a particularly user-friendly platform for those with little to no coding background. Table 2 provides an overview of how to get started with htk. The provided tutorial is based on htk version 2.0.3, the most recent version as of this publication. Chemicals can be identified using name, CAS, or DTXSID (that is substance identifier for the Distributed Structure-Searchable Toxicity (DSSTox) database (<https://comptox.epa.gov>)). Available chemical-specific information includes logP, MW, pKa, intrinsic clearance, fraction unbound in plasma, and blood to plasma partitioning. Calculations can be performed to derive chemical properties, TK parameters, or IVIVE values. Functions are also available to perform forward dosimetry using the various models. As functions are typed at the RStudio command line, available arguments are displayed, with additional help available through the ‘?’ operator. Vignettes for the various available packages in htk are provided to give an overview of their respective capabilities. The aim of htk is to provide a readily accessible platform for working with HHTK models. Following the steps outlined in Table 2 should provide the user with a relatively solid introduction to using htk for their own work.

3. Evaluation of the HHTK Approach to IVIVE

Since HHTK is intended to support public health risk decision making, confidence in its predictions must be sufficiently established. Oreskes [102] wrote of models that “the goal of scientists working in a regulatory context should be not validation but evaluation, and where necessary, modification and even rejection. Evaluation implies an assessment in which both positive and negative results are possible, and where the grounds on which a model is declared good enough are clearly articulated”. By virtue of being applicable to many chemicals (therefore increasing the likelihood of evaluation data being available) and coupling to statistical software (as with the htk R package), it is possible to perform statistical evaluation to determine whether HHTK is “good enough”. Wambaugh, Wetmore [103] applied machine learning tools to develop a model for predicting the accuracy of HHTK methods for predicting C_{ss} , finding roughly half within a factor of 3 and most chemicals within a factor of ten. Thus a “domain of applicability” can be predicted, identifying for which chemicals HHTK may be suitable if an appropriate standard is identified.

The World Health Organization (WHO) indicates that PBTK models are “adequate” when predictions “are, on average, within a factor of 2 of the experimental data” [52]. However, they noted that both observations and model predictions are subject to uncertainty: “The experimental data, frequently obtained in a few experimental animals or human subjects, may constitute a biased sample...” Linakis, Sayre [80] have examined HTTK concentration versus time predictions using a generic PBTK model across roughly forty volatile, non-pharmaceutical chemicals and observed an overall (that is, all time points for all chemicals) root mean squared error (RMSE) of 1.11 (on a \log_{10} scale, therefore a factor of 13x) and a coefficient of determination (R^2) of 0.47. These results, while indicative of predictive ability, are nowhere near the WHO standard.

However, HTTK has been repeatedly shown to be close to the WHO standard for TK summary statistics (that is, dose metric predictions) such as peak concentration and time-integrated (“area under the curve” or AUC) concentration. Wang [25] found across 54 pharmaceutical clinical trials that the predicted AUC differed by 2.3x. Linakis, Sayre [80] found an RMSE = 0.46 or 2.9x for peak concentration and RMSE = 0.5 or 3.2x for AUC. Examining data for a mix of 45 chemicals of both pharmaceutical and non-pharmaceutical nature, Wambaugh, Hughes [22] found an RMSE of 2.2x for peak and 1.64x for AUC. The calibrated method for predicting tissue partitioning that is included in htk similarly predicted human volume of distribution with a RMSE of 0.48 (3x) [38].

The key predicted endpoint for the most common applications of HTTK is C_{ss} , which has been found by several investigations to be within a factor of three for some, but not all, chemicals [21,22,103,104]. This may be due to the use of *in vitro* measurements originally developed for pharmaceuticals, for which metabolism is driven by the liver, and therefore neglecting extra-hepatic metabolism that may be a larger factor for non-pharmaceuticals [22]. Error in prediction of TK may be further compounded by neglecting active transporters in the generic PBTK models [103].

The assumptions used for IVIVE with HTTK may vary between chemicals and applications. For example, it has long been recognized that the rate of metabolism for some compounds is “restricted” by the fraction unbound in plasma — that is, the off-binding rate of the chemical is slow relative to the rate of metabolism and therefore only the free fraction of chemical may be metabolized in any instant — while for other chemicals the metabolism is “unrestricted” [84]. In the absence of a model for predicting the off-binding rate for a chemical, one may choose to treat all chemicals as restricted [12,27,94,103], non-restricted [105], or try to make the determination using *in vivo* data [21]. The health protectiveness of the assumption varies on the application — restrictive clearance leads to higher estimated tissue concentrations which may be health-protective for human predictions but may underestimate toxic potency for animal experiments [6]. The htk package allows users to make choices as to how IVIVE is performed, but applies conservative default assumptions in the absence of user input (for example, clearance is assumed to be restricted unless the user specifies otherwise). Honda, Pearce [20] used htk to evaluate sixteen sets of assumptions for IVIVE, including restrictive vs. non-restrictive clearance. Regardless of the assumptions, Honda, Pearce [20] showed that applying a high-throughput PBTK model enhanced the

apparent correlation between *in vitro* bioactive concentrations and toxic doses determined *in vivo*, compared with no TK at all.

Ominously, Oreskes [102] also wrote that “One may remove obvious errors in a model while more subtle ones remain.” It is hoped that modern, open-source, and modular tools may foster an environment in which subtle problems are more rapidly identified and remedied. Any sort of evaluation of HHTK requires data [6,7,56,106] and a recently-developed public database of TK concentration vs. time data now provides reference data for nearly 150 chemicals from more than 500 literature studies and provides a reporting standard and repository for more [81]. Empirical evaluation of HHTK across many chemicals and classes of chemicals allows quantification of the prediction uncertainty, potentially characterized by the RMSE or average fold error. In turn, regulators may decide whether the associated uncertainties are acceptable depending on context.

4. Conclusion

Generic, high-throughput PBTK models minimize the data requirements to generate chemical- and scenario-specific predictions. By sharing a common structure and software platform, these generic models help overcome reliability challenges with PBTK implementation and documentation [56,57] to allow greater reproducibility and potentially support regulatory decision making [57,58,62]. The htk R package is a key example of a suite of generic TK models and databases that allow users to rapidly predict ADME of chemicals within the body. Unlike many other similar tools, htk is open source software, and includes peer-reviewed chemical-specific data. The open accessibility of htk facilitates collaborations between scientists and enables continuous improvement of the tools, as researchers can identify and report potential issues within the htk package. The greatest strength of the htk tool over other similar tools is its ability to screen large numbers of chemicals at once due to a suite of generic TK models and databases. This capability makes it easier to identify outliers and other issues that would likely go unnoticed when screening a single chemical at a time. The htk tool makes important chemical prioritization information available for risk assessors and policymakers more rapidly than traditional models that focus on toxicity of single chemicals.

Inhalation is an important route of exposure, particularly for occupational settings. The latest version of the htk package includes an inhalation model [80]. Inhalation models are relatively new in the realm of HTS [107], since previous works mainly focused on the less volatile chemicals likely to enter the body orally (for example, Rotroff, Wetmore [27]). Inclusion of new routes of exposure and new classes of chemicals enhances the utility of HHTK.

There are some limitations with the htk package, many of which are addressed and supported by other available HHTK tools. One limitation is that htk focuses on generic, simple TK models, without a detailed description of ADME for a particular chemical and any physiological (or other) processes or compartments that may be especially relevant to that chemical. PK-Sim [75], for example, is a platform that readily supports development of chemical-specific models including potentially relevant physiology. In addition, metabolites are not currently considered in “htk,” as this package only reproduces metabolism as the

elimination of the parent compound, whereas some proprietary software such as Simulations Plus [74] can account for the formation of metabolites. In htk, metabolism is based solely on whole hepatocyte clearance, instead of ascribing metabolism to specific enzymes, a strength of the proprietary software SimCYP. SimCYP has the additional advantage of being able to follow drug-chemical or chemical-chemical interactions [24]. Despite these limitations, htk provides an open-source, replicable interface for conducting high-throughput toxicological prioritization to assess risk potential across thousands of chemicals currently used throughout commerce.

5. Expert opinion

Using TK to understand the dose-response relationship is a critical part of assessing chemical risk posed to public health [108]. Chemicals still in need of triage, prioritization, and potentially full assessment number in the thousands [10]. HTTK can assist chemical risk assessors with its ability to support rapid chemical screening/prioritization, perform IVIVE, and perform forward TK modeling for a large number of chemicals with only limited available chemical-specific data. It is unlikely that HTTK will give better predictions than a bespoke model developed with detailed chemical-specific *in vivo* data that has been tailored to include all physiological processes identified as key to that chemical's ADME. We expect a generic model to be less accurate with respect to reproducing *in vivo* ADME measurements, but more likely to be accurately reported, reproducible, and statistically evaluated. Therefore, generic models may, in some cases, be more suitable to decision making contexts. However, this hypothetical comparison may be irrelevant since it is unlikely that such detailed bespoke PBTK models will ever be developed for the vast majority of chemicals. For the thousands of chemicals for which detailed data and models will likely never be available, HTTK offers a rapid and scientifically defensible description of ADME.

The reproducibility of PBTK models is a necessity of public health risk assessment and a known weakness as documented in the scientific literature [57,60]. Model reporting/documentation criteria and templates are becoming available that may enhance reproducibility if a set of common standards are adopted by the community [62,66]. However, the longevity and portability of models become issues as languages change and, in some cases (notably the popular modeling language acsIX), become discontinued [109]. Many existing chemical-specific models were written in acsIX and these models now need to be translated if they are to continue to be used. To build detailed chemical-specific PBTK models, the free, open-source modeling software GNU MCSim might be used [110,111].

While many published PBTK models do not currently meet the Clark [62] criteria for sufficient documentation [57,61], generic models of HTTK offer a tantalizing alternative. The modular and open source design of tools like htk can enable the tools, models, and data to be used, evaluated, and continuously improved by a broad user community, including toxicological researchers, risk assessors, regulators, and discovery scientists. By trading specificity for reliability, the generic TK model design can increase model uncertainty. However, users can have greater confidence in the model structure (albeit simplified) and

software implementation, since the generic model can be tested for hundreds to thousands of chemicals [56].

It should not be expected that the “one-size-fits-all” approach of HHTK will actually be appropriate for all chemicals; the goal is instead to develop something akin to “one-size-roughly-fits-many”. To do this, we must identify to which chemical classes HHTK should or should not be applied and with what confidence. Wambaugh, Wetmore [103] was an early attempt to use machine learning to anticipate what sorts of chemicals might be well predicted by HHTK, and which might be poorly predicted (for example, transporter substrates). However, thanks to significant advances in models and data since 2015, such approaches are already in need of an update.

Beyond the general suitability of HHTK to a chemical, work is needed to better anticipate and remedy chemical-specific difficulties with measurement of *in vitro* TK parameters. Some chemicals (for example, volatiles) are ill-suited to cell culture, while others (for example, curcumin-related compounds) prove difficult for mass spectrometry. Basic decision tree classifier methods can identify specific chemical features or molecular properties/descriptors that are associated with *in vitro* measurement difficulties. The development of alternative and improved *in vitro* assays to measure TK parameters, and tools to recognize when they should be applied, would enhance the applicability of HHTK across chemical classes. Similarly, substantial effort has gone into developing QSARs for *in vitro* TK parameters and a recent focus has been on broadening beyond pharmaceutical space to a wider variety of chemical classes [87,112–115]. As with all such tools, confidence must be quantified, and the domain of applicability established.

Development of the htk R package has followed the availability of chemical-specific data in the public domain. For example, the publicly available whole hepatocyte metabolism data is the result of multiple metabolizing enzymes, including “cytochrome P450 enzymes” or “CYPs”, which are known to vary in expression and functionality throughout the population. SimCYP and other tools oriented towards drugs have invested substantially in making use of less widely available data on enzyme-specific metabolism. While SimCYP includes a module on population variability in expression/function of specific enzymes, such a module has not been included in htk because of the lack of these data for non-pharmaceutical chemicals [47]. If efforts like that of Wetmore, Allen [30] for environmental chemicals could be expanded, then it would make sense to model enzyme-specific metabolism and ability to simulate population variability in enzyme abundance in htk. Other modules should be added as the need arises and the data to support them are developed.

Similarly, new high-throughput (generic) PBTK models should be added as both the need arises and the data to evaluate them are developed. One clear area of need is integration of early life-stage parameters for potentially sensitive life stages, including pregnancy and gestation and infancy. However, for any new model it will be necessary to identify and curate appropriate chemical datasets. Key needs include *in silico* QSAR models for (a) transplacental permeability (b) embryonic plasma binding (AFP or alpha-fetoprotein, an early life-stage-specific albumin ortholog) binding, and (c) milk:plasma partition coefficients for gestational pharmacokinetics/dosimetry estimation and post-gestational developmental

models. Additionally, integrating gestational/embryonic tissue-specific expression profiles of phase I/II metabolizing enzymes will be vital.

Usability (that is, user interface) is a barrier to the adoption of any software tool, including those for TK [116]. Precalculated HHTK-based IVIVE predictions are available from on-line databases (such as <https://comptox.epa.gov/dashboard> [117]) in addition to graphical interfaces for PBTK models [77,78]. The goal of such tools is both to make the workings of the model more accessible as well as reduce software barriers (that is, lower need for third party software installation, command-line angst, drop down interaction as a web-service). Another enhancement to usability is the development of KNIME [79] workflows that take advantage of the R KNIME nodes, but enable a variety of manual structure (sketch) inputs, QSAR estimation and plugs to third party software such as MOE [89] or Schrodinger (<https://www.schrodinger.com/drug-discovery>), post simulation processing and reporting to third party visualization tools. Stewardship of HHTK tools is just as critical and will require the development of training material (including R “vignettes” and video examples) that provide a variety of user scenarios that are fully documented.

The longest-range goals of HHTK should include working toward “HTTD” (high-throughput toxicodynamics). Methods to link *in vitro* assays to *in vivo* health effects are needed. Currently, IVIVE of *in vitro* HTS data using HHTK involves the major assumptions that (1) a concentration that was bioactive *in vitro* will also be bioactive in the body and (2) even if the concentration is bioactive in the body, that activity in this one specific biological endpoint will have an actual effect on human health. Can those assumptions be refined, for instance by comparing *in vitro* predictions with the outcomes of legacy *in vivo* toxicity studies [20,29,118]? Better yet, can we integrate target specific information for receptors involved in signal transduction pathways and apical endpoints) — for instance adverse outcome pathway molecular initiating events [119] based on high-throughput receptor activity models [105]? HTTD, representing HHTK coupled to HTS, may eventually allow fully *in vitro* and computational prediction of public health effects of chemicals.

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Article highlights:

- *In vitro* high-throughput screening (HTS) to efficiently assess potential chemical risk to public health requires toxicokinetics for *in vitro-in vivo* extrapolation (IVIVE)
- The high-throughput toxicokinetics (HTTK) method uses generic toxicokinetic (TK) models that can be parameterized for thousands of chemicals with *in vitro* TK data
- The U.S. Environmental Protection Agency (U.S. EPA) provides HTTK methods through the publicly available software package called “httk”
- The open accessibility of the httk package facilitates collaborations between scientists, and enables continuous improvement of the tools
- The fundamental strength of the httk package is its ability to screen many chemicals for potential risk at once, using only limited data

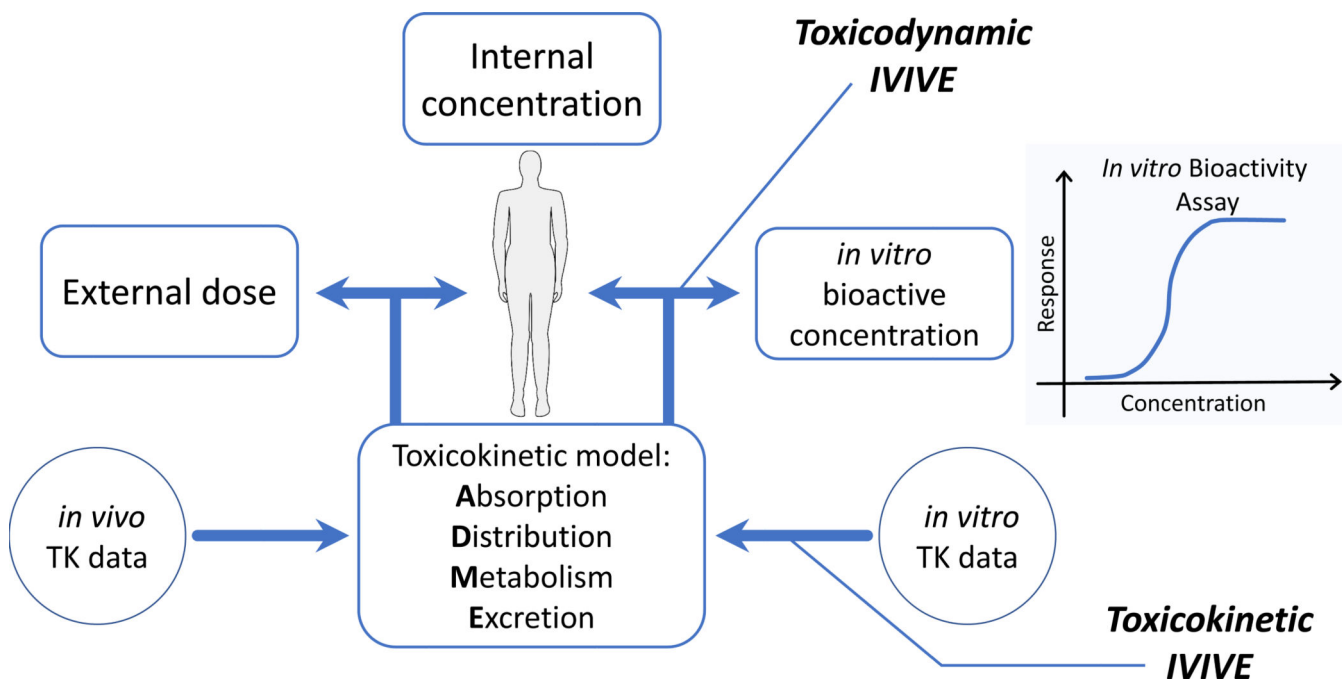


Figure 1. *In vitro* to *in vivo* extrapolation (IVIVE).

We perform that IVIVE using toxicokinetic (TK) modeling. TK models relate external dose to internal body concentration by describing “what the body does to the chemical”: absorption, distribution, metabolism, and excretion (ADME). For IVIVE of *in vitro* bioactive concentrations, we assume that bioactivity would occur in the body at a concentration equal to an *in vitro* bioactive concentration, and use TK modeling in reverse (that is, reverse dosimetry) to find the “equivalent dose” – an external dose that would produce the specified body concentration. While IVIVE broadly includes any use of *in vitro* data to predict phenomena *in vivo*, it is useful to distinguish between TK IVIVE (that is, the use of *in vitro* data to predict ADME) and toxicodynamic (TD) IVIVE, which includes the use of *in vitro* data to predict toxic effects *in vivo*.

$$AED_{C_{ss,95}} = \frac{[X]}{C_{ss,95}}$$

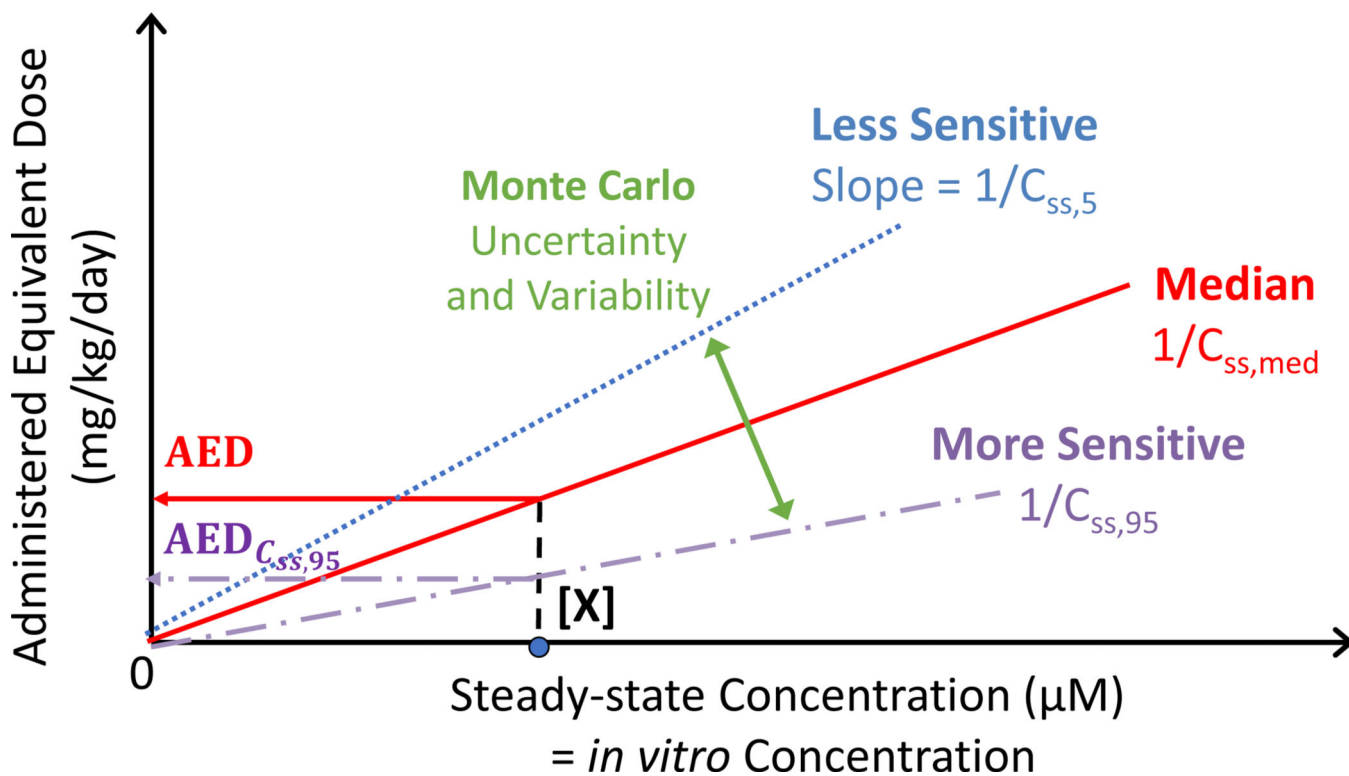


Figure 2. Reverse Dosimetry Toxicodynamic IVIVE.
 Equivalent external dose is determined by solving the TK model in reverse by deriving the external dose (that is, TK model input) that produces a specified internal concentration (that is, TK model output). Reverse dosimetry and IVIVE using HHTK relies on the linearity of the models. We calculate a scaling factor to relate *in vitro* concentrations (μM) to administered equivalent doses (AED). The scaling factor is the inverse of the steady state plasma concentration (C_{ss}) predicted for a 1 mg/kg/day exposure dose rate. We use Monte Carlo to simulate variability and propagate uncertainty to calculate an upper 95th percentile $C_{ss,95}$ for individuals who get higher plasma concentrations from the same exposure.

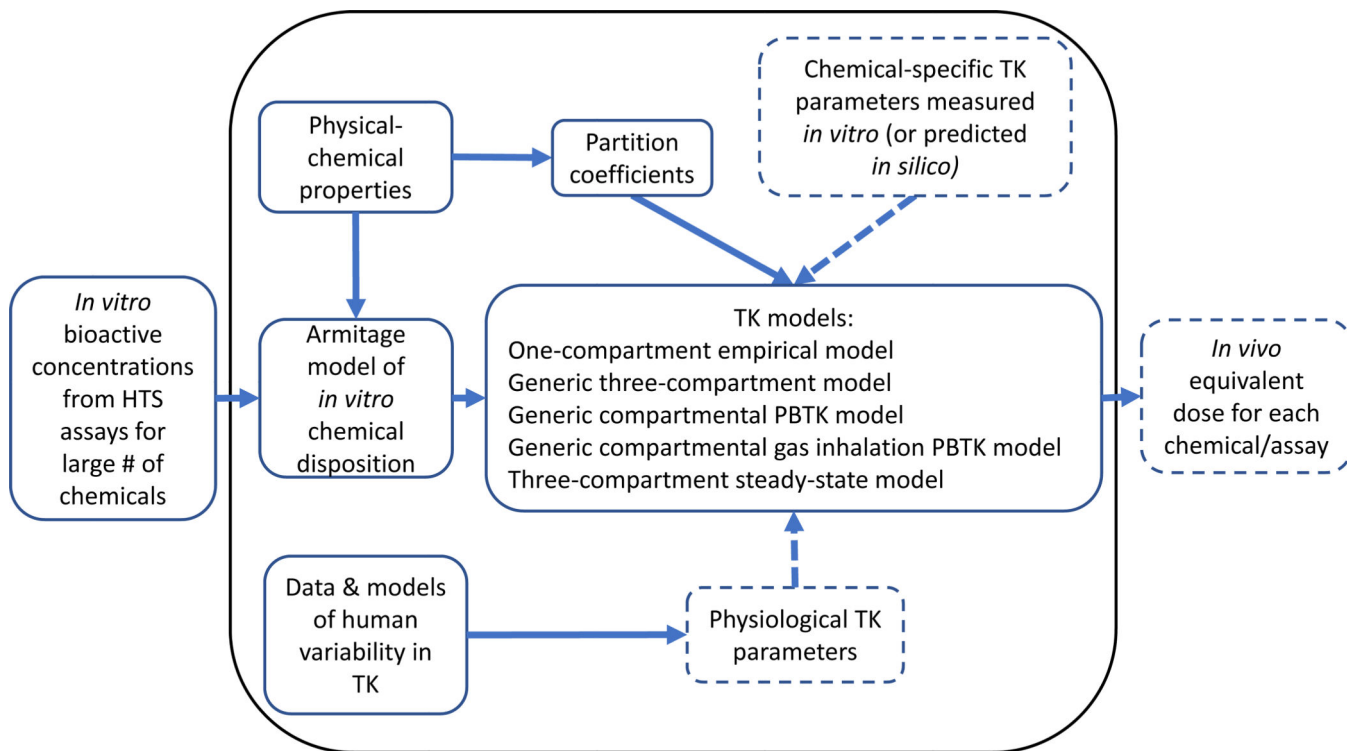


Figure 3. Schematic of httk R package.

Dashed boxes/arrows represent parameters that can be probabilistic in a Monte Carlo simulation.

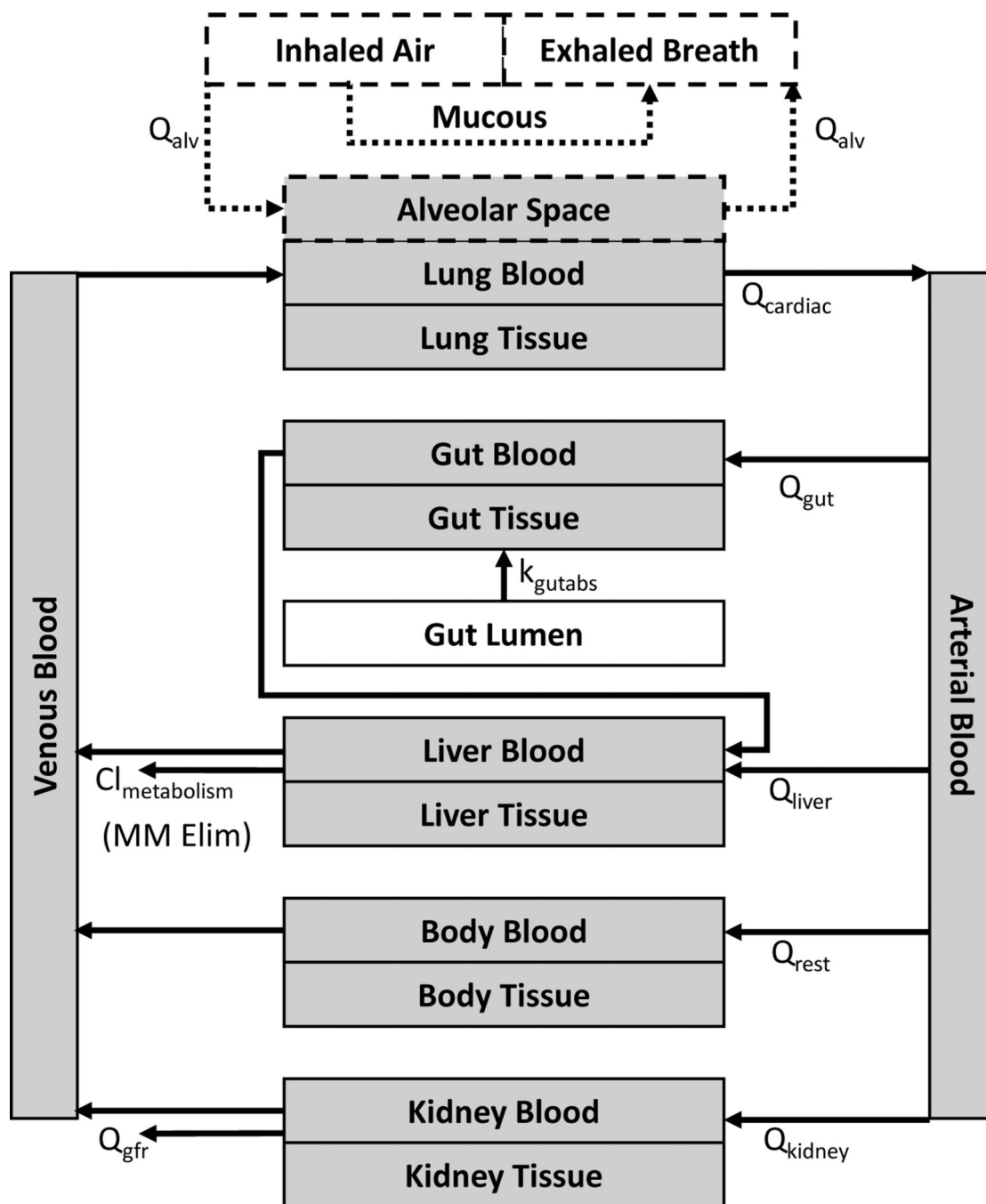


Figure 4. Example of a generic gas inhalation PBTK model. A single (generic) physiological structure is used for all appropriate chemicals.

Chemical-specific parameters can be predicted from a combination of in vitro measurements and QSARs. Q_{rest} is defined as the difference between $Q_{cardiac}$ and the flow to the liver, kidney, and gut to preserve mass-balance.

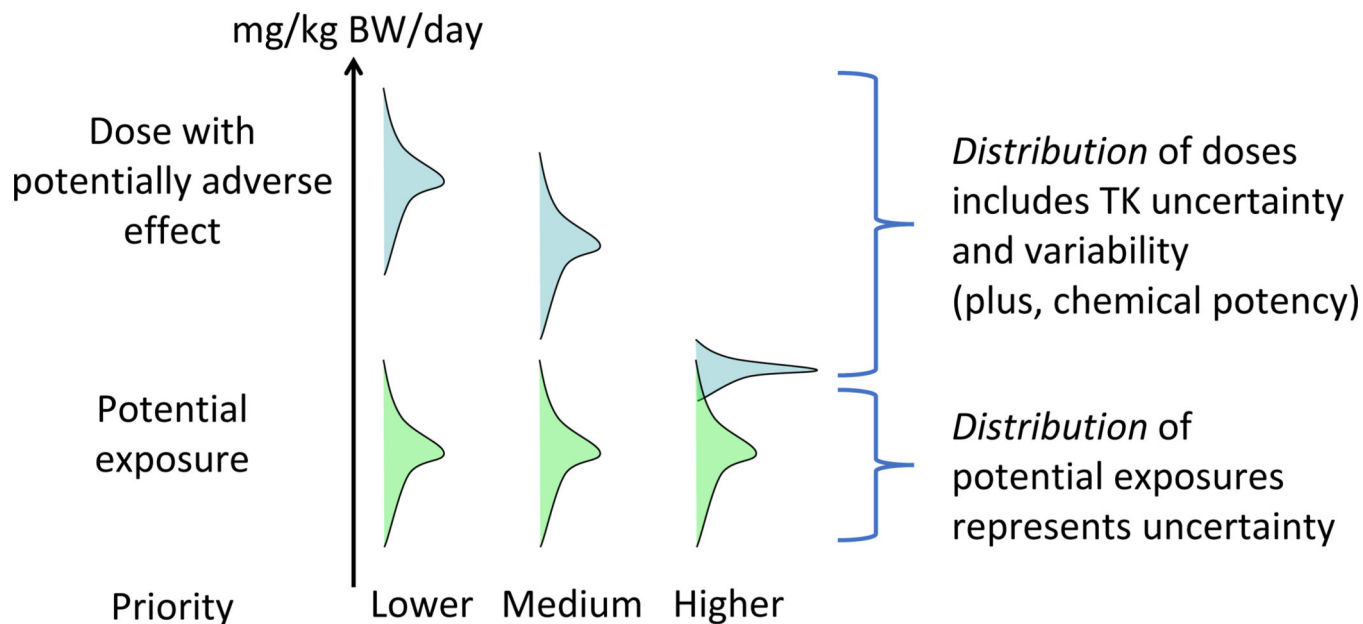


Figure 5. High-throughput chemical risk prioritization to rapidly prioritize large numbers of chemicals.

Evaluation of potential risk by comparing distributions of dose with potentially adverse effect and potential exposure. For TK we can account for both uncertainty and variability. If the hazard and exposure distributions are far apart, as shown on the left, then potential risk is lower – it means exposure probably doesn't reach a level where there would be an adverse effect. If the lower tail of the hazard distribution starts to overlap the upper tail of the exposure distribution, as shown in the middle, then potential risk is medium. And if the hazard and exposure distributions totally overlap, as shown on the right, then potential risk is higher – it means that exposure is more likely to reach a level where there might be an adverse effect.

Table 1:

Overview of Generic PBTK Modeling Tools.

	SimCYP	ADMET Predictor / GastroPlus	PK-Sim	IndusChemFate	httk
References	Jamei, Marciniak [19]	Lukacova, Woltoz [69]	Eissing, Kuepfer [70]	Jongeneelen and Berge [71]	Pearce, Setzer [27]
Availability	License, but inexpensive for research	License, but inexpensive for research	Free: http://www.open-systems-pharmacology.org/	Free: http://cefic-lri.org/lri_toolbox/induschemfate/	Free: https://CRAN.R-project.org/package=httk
Open Source	No	No	GitHub	No	CRAN and GitHub
Default PBTK Structure	Yes	Yes	Yes	Yes	Yes
Population Variability	Yes	Yes	Yes	No	Yes
Data Needs	High/Low	High/Low	High	High	Low
Typical Use Case	Drug Discovery	Drug Discovery	Drug Discovery	Environmental Assessment	Screening
Batch Mode	Yes	Yes	Yes	No	Yes
Graphical User Interface	Yes	Yes	Yes	Excel	No*
Built-in Chemical-Specific Library	Many Clinical Drugs	No	Many pharmaceutical-specific models available	15 Environmental Compounds	980 Pharmaceutical and ToxCast Compounds
Ionizable Compounds	Yes	Yes	Yes	No	Yes
Export Function	No	No	Matlab and R	No	SBML and Jarnac
R Integration	No	No	Yes (2017)	No	Yes
Reverse Dosimetry	Yes	Yes	Yes	No	Yes

Table 2:

Getting Started with httk.

Where Do I Get R?	R is freely available from the Comprehensive R Archive Network (CRAN): https://cloud.r-project.org/ Graphical user interface (GUI), RStudio, is freely available: https://rstudio.com/
Getting Started with R Package httk	<code>install.packages(httk)</code> RStudio provides a menu "Install Packages" under "Tools" tab
Load the HTTK data, models, and functions	<code>library(httk)</code>
Check what version you are using	<code>packageVersion(httk)</code>
Getting help with R Package httk	<code>help(httk)</code> You can go straight to the index: <code>help(package=httk)</code>
List all CAS numbers for all chemicals with sufficient data to run httk	<code>get_cheminfo()</code>
List all information:	<code>get_cheminfo(info="all")</code>
Is a chemical with a specified CAS number available?	<code>"80-05-7" %in% get_cheminfo()</code>
All data on chemicals A, B, C	<code>subset(get_cheminfo(info="all"),Compound%in%c("A","B","C"))</code>
Administrated equivalent dose (mg/kg BW/day) to produce 0.1 uM plasma concentration, 0.95 quantile, for a specified CAS number and species	<code>calc_mc_oral_equiv(0.1,chem.cas="34256-82-1", species="human")</code>
Calculate the mean, AUC, and peak concentrations for a simulated study (28-day daily dose, by default) for a specified CAS number and species	<code>calc_tkstats(chem.cas="34256-82-1", species="rat")</code>
Using the PBTK solver for a specified chem name	<code>solve_pbt(chem.name="bisphenol a", plots=TRUE)</code>
List all vignettes for httk	<code>vignette(package=httk)</code>
Displays the vignette for a specified vignette	<code>vignette("Frank2018")</code>
Create data set, my_data, for all data on chemicals A, B, C, in R	<code>my_data <- subset(get_cheminfo(info="all"),Compound%in%c("A","B","C"))</code>
Export data set, my_data, from R to csv file called my_data.csv in the current working directory	<code>write.csv(my_data, file = "my_data.csv")</code>